

# Analysis of Well-Mixed Fed-Batch Cultures using Unstructured Models

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## Introduction

Fed-batch cultures are the most complex of the three modes of operating a bioreactor. Fed-batch cultures are dynamic and have volume change. Like a continuous culture, fed-batch cultures have an input feed stream into the reactor. However, unlike chemostats, there is no outflow from the reaction vessel in a fed-batch reactor. Thus, the working volume in the reactor increases over time diluting the contents of the vessel. In this lecture, we'll develop a mathematical description of fed-batch cultures using the general material balances, and then explore how the performance of the fed-batch culture can be optimized.

**General model equations for fed-batch cultures.** Let's start with the general material balances we derived previously:

$$\frac{dC_j}{dt} = \sum_{s=1}^S v_s D_s C_{j,s} + \left( \sum_{r=1}^R \sigma_{jr} \hat{r}_r \right) + \left( \sum_{k=1}^T \tau_{j,k} q_k \right) X - \frac{C_j}{V} \frac{dV}{dt} \quad j = 1, 2, \dots, \mathcal{M} \quad (1)$$

$$\frac{dX}{dt} = \sum_{s=1}^S v_s D_s X_s + (\mu - k_d) X - \frac{X}{V} \frac{dV}{dt} \quad (2)$$

$$\frac{dV}{dt} = \sum_{s=1}^S v_s \frac{\rho_s}{\rho} F_s - \frac{V}{\rho} \frac{d\rho}{dt} \quad (3)$$

where the quantity  $D_s$ , called a *dilution rate* ( $\text{hr}^{-1}$ ), is given as:

$$D_s \equiv \frac{F_s}{V} \quad s = 1, 2, \dots, S \quad (4)$$

The quantity  $C_j$  denotes the concentration of the  $j$ th extracellular metabolite,  $V$  denotes the working volume of the culture and  $X$  denotes the cellmass. In a fed-batch culture, there are in-flows, but no out-flow from the culture vessel, thus  $D_{s^-} = 0$  where  $s^-$  denote the set of outflow streams. Because there is no out-flow, there is a volume change ( $dV/dt > 0$ ), and all the dilution terms in

the material balances are non-negative:

$$\frac{dC_j}{dt} = \sum_{s=1}^{S^+} v_s D_s C_{j,s} + \left( \sum_{r=1}^{\mathcal{R}} \sigma_{jr} \hat{r}_r \right) + \left( \sum_{k=1}^{\mathcal{T}} \tau_{j,k} q_k \right) X - \frac{C_j}{V} \frac{dV}{dt} \quad j = 1, 2, \dots, \mathcal{M} \quad (5)$$

$$\frac{dX}{dt} = \sum_{s=1}^{S^+} v_s D_s X_s + (\mu - k_d) X - \frac{X}{V} \frac{dV}{dt} \quad (6)$$

$$\frac{dV}{dt} = \sum_{s=1}^{S^+} v_s \frac{\rho_s}{\rho} F_s - \frac{V}{\rho} \frac{d\rho}{dt} \quad (7)$$

**Analysis of a simple fed-batch culture.** To better understand the dynamics of a fed-batch culture, let's simplify the general equations by assuming a Monod growth model (1), a single limiting nutrient  $S$ , a single sterile input feed stream ( $s=1$ ) with similar density to the working volume, no density change as a function of time, stable substrate and product, and no maintenance utilization of substrate. With these assumptions the general fed-batch balances reduce to:

$$\frac{dS}{dt} = D_1 S_1 - \frac{1}{Y_{X/S}^*} \mu X - \frac{1}{Y_{P/S}} q_p X - \frac{S}{V} \frac{dV}{dt} \quad (8)$$

$$\frac{dP}{dt} = D_1 P_1 + q_p X - \frac{P}{V} \frac{dV}{dt} \quad (9)$$

$$\frac{dX}{dt} = (\mu - k_d) X - \frac{X}{V} \frac{dV}{dt} \quad (10)$$

$$\frac{dV}{dt} = F_1(t) \quad (11)$$

If we substitute the volume balance into the substrate, product and cellmass balances (and drop the stream index on the dilution rate), we arrive at the simplified system of equations:

$$\frac{dS}{dt} = D(S_1 - S) - \frac{1}{Y_{X/S}^*} \mu X - \frac{1}{Y_{P/S}} q_p X \quad (12)$$

$$\frac{dP}{dt} = D(P_1 - P) + q_p X \quad (13)$$

$$\frac{dX}{dt} = (\mu - k_d) X - DX \quad (14)$$

$$\frac{dV}{dt} = F(t) \quad (15)$$

where  $\mu$  is given by:

$$\mu = \mu_g^{max} \left( \frac{S}{K_g + S} \right) \quad (16)$$

and we assume the Luedeking and Piret model for product formation (2):

$$q_p = \alpha \mu + \beta \quad (17)$$

The cellmass, substrate, product and volume balances are coupled nonlinear differential equations which can be solved numerically using common packages such as MATLAB or JULIA (3).

*Growth associated production formation.*

*Non-growth associated production formation.*

*Mixed product formation.*

**How do we measure the performance of a bacterial culture?**

## References

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