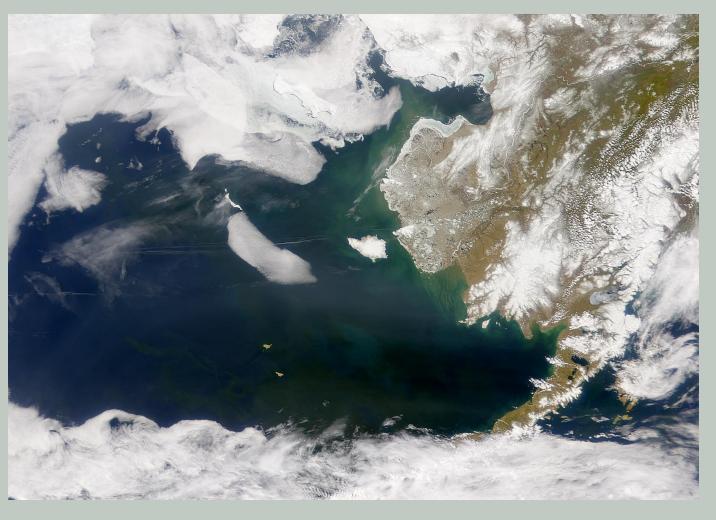
Progress Update

Laura Vary

November 4, 2021

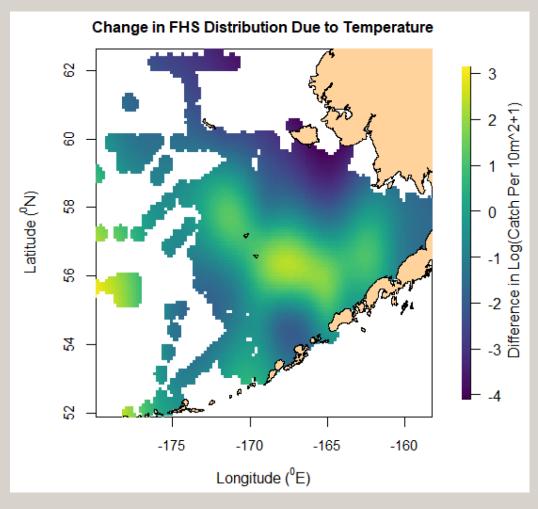
Analytical Workflows



NASA Earth Observatory, 2002

Background of Research

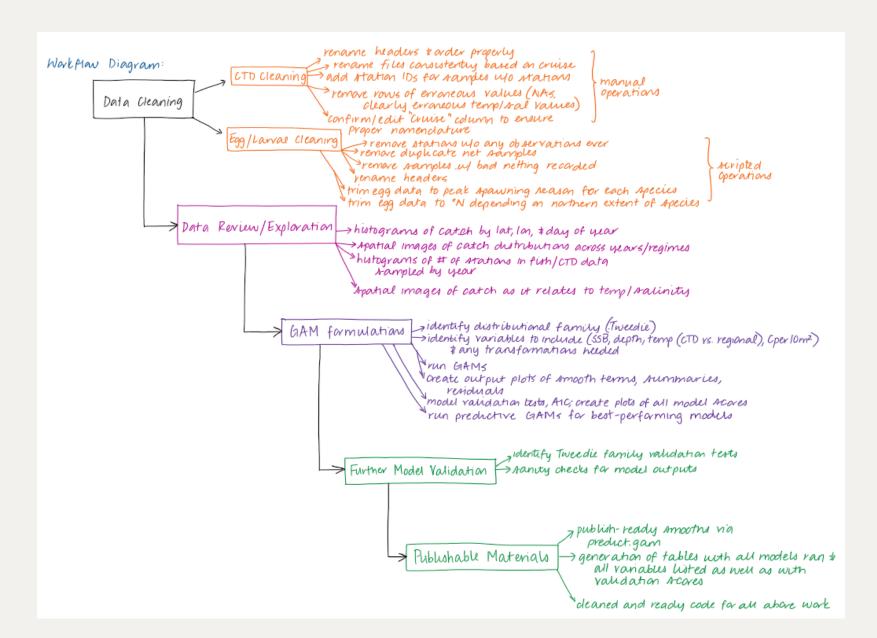
- Research Questions:
 - Spawning Behavior:
 - How flexible is spawning geography and phenology?
 - In practice: what GAM best explains variation in spawning location and time and how well does it perform?
 - Larval Biogeography:
 - What types of water mass characteristics do larvae tend to be associated with?
 - In practice: what GAM best explains larval biogeography, and where are those best explaining environmental condiitons located?



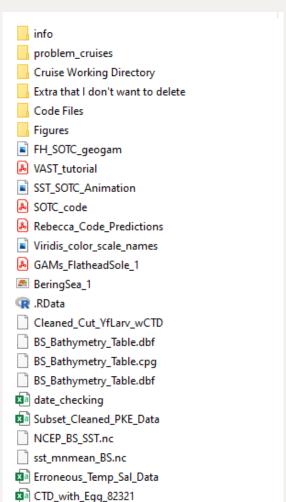
Example of Model Output – Threshold Geography GAM for Flathead Sole

11/4/2021

Created a Workflow to Stick To:



Created and Improved GitHub Repo and Project Organization



Spawning Behavior and Larval Biogeography of Bering Sea Marine Fishes		
E README.md		0
README.md	Update README.md	9 days ago
.gitignore	Initial commit	last montl
Organized Thesis Analyses (Code)	Improving code, rerunning GAMs	4 days ago
Ichthyo Data	Updating GAM models	16 days ago
GAM Models	File organization, rewrote some scripts, added a new script for varia	7 days ago
Figures	Added in a figure	9 days ago
	Improving code, rerunning GAMs	4 days ago

This repository includes all code and data for my M.Sc. thesis and associated manuscripts (expected thesis

completion date: 6/15/2022). These analyses were conducted at Oregon State University within the Marine

Resource Management program and under the oversight of my advisor, Dr. Lorenzo Ciannelli.

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Improved Readability of Code

2 # EGGS: Spawning Behavior Flathead Sole -----

```
#the following code creates generalized additive models for eggs and larvae of flathead sole.
    #these analyses form the basis of my MS thesis.
    #egg data uses an averaged sea surface temperature for the month of March in the Southeastern Bering Sea
           #March index was chosen because it is two months before the peak of flathead egg CPUE, and thus March
           #conditions are likely more relevant to spawning behavior than temperatures in later months.
   # Load in data -----
    fhsub<-read.csv(file='../Ichthyo Data/Cleaned_Cut_FhEggs.csv',header=TRUE,check.names=TRUE) #for egg GAMs
    fhlarv.ctd<-read.csv(file='../Ichthyo Data/Cleaned Cut FhLarv wCTD.csv',header=TRUE,check.names=TRUE) #for larval GAMs
    ##Load in regional temperature index for March (2 mos before peak egg CPUE in May), for egg GAMs
    reg.sst<-read.csv('../Environmental Data/Mar_SST_RegionalIndex_NCEP_BS.csv',header=TRUE,check.names=TRUE)
    head(reg.sst) #range of regional average: lon: -180 to -151, lat: 50.5 to 67.5
    for(i in 1:nrow(fhsub)){
      fhsub$reg.SST[i]<-reg.sst$SST[reg.sst$year==fhsub$year[i]]}
20
   # Spawning Behavior - Egg GAMs -----
    #the following code generations GAMs that quantify variation in space and time of spawning behavior
        #(as proxied by egg CPUE) in relation to regional temperature index close to typical time of spawning
24
25
                                                                                   # Threshold Phenology Egg GAM -----
27
                                                                               47
    eg.base<-gam((Cper10m2+1)~factor(year)+s(lon,lat)+s(doy)+s(bottom depth,k=5),
                                                                                   temps<-sort(unique(reg.sst$SST)) #order by unique values of regional SST (March temps here)
                data=fhsub.familv=tw(link='log').method='REML')
                                                                               49
30
                                                                               50
                                                                                   bd<-4 #dictates essentially how many unique temperatures we check. can vary this depending on your system, how long you want the model to take,
                                                                                      #and probably some other factors that I don't know as well.. relatively arbitrary. smaller values for bd check more unique temperature values
                                                                               51
                                                                               52
                                                                                    temps.in<-temps[bd:(length(temps)-bd)] #vector whose length is dependent on bd; this vector is what the for loop uses to test many models at different thresholds
                                                                               53
                                                                               54
                                                                                    aic.pheno<-NA*(temps.in) #create an empty vector with the proper length to fill in later
                                                                               55
                                                                                    thr.pheno<-as.list(1:(length(temps.in))) #another list with the same dimensions to fill in later
                                                                               58
                                                                                   #this function below takes the unique temperature values and creates a model for each temperature as a threshold.
                                                                                   #then, we use AIC to ask which model and at what threshold best explains variation in phenology?
```

11/4/2021

Continuous Goals

Add comments and introductions to all code files

Add environmental data sources to .readme file and update with any new tools

Create streamlined figure creation workflow (once results are finalized)