

\* Report All plant growth regulators in micromolar ( $\mu\text{M}$ )

1 **IN VITRO PROPAGATION OF NANJING LINDEN (*Tilia miqueliana* Maxim)**

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6 Short running title: *In vitro* propagation of *Tilia miqueliana* Maxim

7 **Abstract:** *Tilia miqueliana* Maxim is one of the most important tree species

8 used in landscape architecture, especially in larger cities. <sup>? in China?</sup> ~~In the light of the~~ <sup>Because</sup> ✓ where?

9 high demand for this species, the development of an efficient

10 micropropagation system can help to ensure an adequate supply of plants

11 needed for reforestation and city landscaping. ~~In this study~~ <sup>Several types of</sup> ✓

12 treatments and methods for micropropagation were compared to establish an

13 optimized protocol for the propagation of *T. miqueliana* through *in vitro*

14 axillary shoot induction. Axillary buds were obtained from <sup>in vitro</sup> seedlings grown ~~in~~ ✓

15 ~~from~~ <sup>from</sup> seeds cultured on WPM1. These buds (explants) were then cultured ✓

16 on initiation medium WPM8, and the resulting elongated shoots were cut into ✓

17 nodal segments and transferred to shoot multiplication medium. <sup>Overall best</sup> ✓

18 results were obtained using the following protocol: highest average number of

19 nodes on explants <sup>11.3</sup> ~~(11.25)~~ per explant) was induced on ~~medium WPM18;~~ ✓

20 highest rooting success <sup>91.7</sup> ~~(91.67%)~~ was achieved after first elongating nodal

21 segments on WPM10 and then transferring them to medium M84. ✓

22 **Key words:** Nanjing linden, *Tilia miqueliana*, *in vitro* propagation,

23 multiplication <sup>Half-strength Murashige and Skoog medium with 14.76  $\mu\text{M}$  IBA</sup>

woody plant medium (WPM) without glycine, but supplemented with 8.66  $\mu\text{M}$  gibberellic acid

WPM with 2.28  $\mu\text{M}$  zeatin and 0.2  $\mu\text{M}$  indole-3-butyric acid (IBA)

WPM with 2.22  $\mu\text{M}$  benzyladenine, 2.28  $\mu\text{M}$  zeatin, and 0.1  $\mu\text{M}$  IBA

WPM without glycine and plant growth regulators

\* Use the word disinfestation or disinfested instead of disinfected or disinfection.

"S" instead of "C" ✓

24 Nanjing linden (*Tilia miqueliana* Maxim), a member of the ~~family~~ Tiliaceae ✓  
25 (linden), is a native tree species of Jiangsu province, China. The linden is highly  
26 prized as an ornamental tree, especially when abundant foliage and good shading is  
27 desired. <sup>In addition</sup> ~~Furthermore~~, it can be a very important nectar source for bees, and hence is ✓  
28 welcomed by beekeepers. Propagation of linden is ~~done~~ <sup>conducted</sup> primarily via cuttings and ✓  
29 seeding. However, these methods ~~can~~ suffer from disadvantages such as poor ✓  
30 establishment rates or low germination rates (Spaeth 1932, Rose 1919, Yang et al.  
31 2011). At ~~present~~ <sup>present</sup>, micropropagation is regarded as an effective method of obtaining ✓  
32 seedlings of linden. However, ~~in previous studies, only Vladimir Chalupa (1984, 1990)~~  
33 investigated the tissue culture of small-leaved linden (*T. cordata*). In ~~1984~~ <sup>1984</sup>, he ✓  
34 ~~established~~ <sup>was established</sup> a shoot proliferation method by using nodal stem segments from ~~one~~ ✓  
35 old seedlings as explants. In 1990, he ~~established~~ <sup>protocol was established</sup> the plant regeneration by somatic ✓  
36 embryogenesis from cultured immature embryos. However, ~~to our knowledge, no~~ <sup>T</sup> ✓  
37 study has ~~previously~~ investigated the tissue culture of Nanjing linden (*T. miqueliana*). ✓  
38 To satisfy an ever increasing horticultural demand for this species it was therefore  
39 considered useful to develop an effective in vitro multiplication and propagation  
40 protocol for Nanjing linden.

41 Nodal segments that already have a developed meristem are suitable for  
42 micropropagation, as ~~they~~ <sup>these</sup> are more easily manipulated, and additionally have a high ✓  
43 proliferation rate while retaining clonal fidelity (Vijaya and Padmaja, 1999). It has  
44 proven difficult to establish a tissue culture protocol for Nanjing linden (*T. miqueliana*)  
45 using ~~adult~~ <sup>mature</sup> plant material, obtained directly ~~in~~ <sup>from</sup> the field. This seems to be mostly ✓  
46 attributable to high levels of contamination and vitrification (Lu et al. 2004). With  
47 regards <sup>to</sup> the selection of nodal segments, we found that the axillary buds of ✓  
48 seedlings germinated from fertile seed were the best method to obtain explants.

49 However, it <sup>has</sup> ~~had~~ been reported in other studies that rigorous seed-surface <sup>disinfestation</sup> ~~sterilization~~ ✓  
50 procedures can have a negative effect on the viability of axillary buds of seedlings  
51 (Obeidy and Smith 1990, <sup>Renukadas et al.</sup> ~~Nilima et al.~~ 2010). In the present study, we present an ✓  
52 efficient <sup>disinfestation</sup> ~~disinfection~~ method to minimize negative effects on seedlings <sup>AND produce</sup> ~~while still~~ ✓  
53 <sup>plant material</sup> ~~providing good surface sterilization.~~ ✓

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## 54 MATERIALS AND METHODS

### 55 Plant material

56 Mature fruits of Nanjing linden were collected from one 20-year-old tree in  
57 mid-November. The seeds were then extracted by <sup>placing</sup> ~~fixing~~ the fruits in a vise and ✓  
58 removing the peel with a knife. Following ~~the~~ extraction, the seeds were washed in ✓  
59 sterile water for 20 min and then sterilized by incubation in carbendazim solution for <sup>500 mg L<sup>-1</sup></sup> ✓  
60 2 h (<sup>500 mg L<sup>-1</sup></sup> ~~500 mg L<sup>-1</sup>~~, Sigma Chemical Co., St. Louis, MO, USA). After this sterilization the ✓  
61 seeds were immediately washed three times with water, and <sup>? sterile?</sup> ~~to remove any remaining~~ <sup>then</sup> ✓  
62 <sup>placed</sup> ~~carbendazim were then put~~ into 10% (v/v) liquid anionic detergent (Amway, China), ✓  
63 and <sup>agitated</sup> ~~shaken~~ on a rotary shaker for 20 min. Afterwards, <sup>to remove the detergent</sup> ~~to remove the detergent~~, The ✓  
64 seeds were then placed into a conical flask and washed for 60 min under running  
65 water. Finally, <sup>then</sup> ~~the~~ seeds were <sup>ON</sup> ~~disinfected~~ by immersing <sup>them</sup> ~~them~~ for 40 s in 75% (v/v) ✓  
66 ethanol. Following this pre-treatment, the seeds were then subjected to one of the  
67 following <sup>what does this m stand for?</sup> ~~sterilization~~ treatments: 1) 5.65% (m/v) sodium hypochlorite for 12 min; 2) <sup>should this be</sup> ✓  
68 0.1% (m/v) mercuric chloride ( $\text{HgCl}_2$ ) for 12 min; 3) 5.65% (m/v) sodium ✓  
69 hypochlorite for 6 min, and subsequent treatment with 0.1% (m/v) mercuric chloride  
70 (OR  $\text{HgCl}_2$ ) for 6 min; 4) 2.83% (m/v) sodium hypochlorite for 6 min, and subsequent ✓  
71 treatment with 0.2% (m/v) mercuric chloride ( $\text{HgCl}_2$ ) for 6 min. After treatment with  
72 surface <sup>S</sup> ~~disinfectants~~, the seeds were <sup>RINSED</sup> ~~washed by swirling them~~ in sterile deionized ✓

73 water for 5 min; ~~this washing step was~~ repeated four times. Each of the disinfected  
74 seeds ~~was~~ <sup>were</sup> then transferred to a glass bottle (230 ml, 5×12 cm) containing 30 ml  
75 WPM1. One hundred seeds per treatment with three replicates were evaluated. After  
76 ten days, the seeds without visible bacterial or fungal growth on the surface were  
77 scored, and the contamination rate was then calculated as the ratio of seeds with  
78 visible growth to the total number of seeds.

79 Medium ~~WPM1~~ <sup>(WPM)</sup> was woody plants medium without glycine (Lloyd and McCown  
80 1980; McCown and Lloyd 1981) containing 3 <sup>(mg l<sup>-1</sup>)</sup> gibberellic acid (GA<sub>3</sub>, Sigma). <sup>(WPM1)</sup>  
81 ~~Dormancy breaking~~ <sup>Breaking dormancy</sup> of the seeds <sup>Report in NM</sup>

82 For breaking the dormancy of the seeds, four methods were employed. In the first  
83 method, the seeds were stored at 4°C for 2 d ~~without hot treatment~~, and then cultured  
84 on WPM1 after disinfection <sup>station</sup>. In the second method, the seeds were ~~dipped into~~ <sup>placed in</sup> 40°C  
85 water for 2 h and ~~subsequently~~ <sup>then</sup> stored at 4°C for 2 d, and ~~then cultured~~ <sup>followed by culture</sup> on WPM1 after  
86 disinfection <sup>station</sup>. In the third method, the seeds were cultured on <sup>(WPM0)</sup> after incubation  
87 in 500 <sup>(mg l<sup>-1</sup>)</sup> GA<sub>3</sub> for 3 d at 4°C. In the fourth method, the seeds were cultured on  
88 WPM1 immediately after the thick peel was ~~pared off~~ <sup>Removed</sup>. One hundred seeds per  
89 treatment with three replicates were evaluated.

90 ~~WPM0~~ <sup>(WPM)</sup> was woody plants medium (Lloyd and McCown 1980) without glycine  
91 and plant ~~hormone~~ <sup>growth regulators</sup>.

#### 92 Culture establishment and multiplication of the axillary shoots

93 After culturing in the ~~modified~~ <sup>in vitro</sup> WPM with 3 <sup>(mg l<sup>-1</sup>)</sup> GA<sub>3</sub> for ~~two~~ <sup>2</sup> months, nodal  
94 explants were prepared from the seedlings and transferred to initiation medium. All  
95 initiation media we tested <sup>was</sup> ~~were based on~~ <sup>was</sup> modified WPM, but additionally contained  
96 one of the following: 0.01, 0.2, 0.5, 1.0 <sup>(mg l<sup>-1</sup>)</sup> zeatin (ZT, Sigma) or 0.01, 0.2, 0.5, 1.0  
97 <sup>(mg l<sup>-1</sup>)</sup> N<sup>6</sup>-benzyladenine (BA, Sigma), and 0.04 <sup>(mg l<sup>-1</sup>)</sup> indole-3-butyric acid (IBA,

98 Sigma, ~~St. Louis~~<sup>culture</sup>). After 30 days of ~~incubation~~<sup>culture</sup> the elongated shoots (longer than 6 cm,  
99 3-4 ~~branches~~<sup>?</sup>) were divided into nodal segments, each bearing a single axillary bud.  
100 These segments were then ~~transplanted~~<sup>cultured</sup> onto multiplication medium consisting of  
101 modified WPM supplemented with ZT (0.01, 0.1, 0.5, 1.0  $\text{mg l}^{-1}$ ) or BA (0.01, 0.1, 0.5,  
102 1.0  $\text{mg l}^{-1}$ ) and 0.02  $\text{mg l}^{-1}$  IBA, or supplemented with 1.0  $\text{mg l}^{-1}$  BA, 1.0  $\text{mg l}^{-1}$  ZT  
103 and 0.2  $\text{mg l}^{-1}$  IBA. After ~~four~~<sup>4</sup> weeks of culture the average number of shoots was  
104 assessed, and all ~~obtained~~<sup>cultured</sup> shoots were separated and ~~incubated~~<sup>cultured</sup> further on WPM0 to  
105 increase elongation.

106 Subsequently, ~~the~~<sup>then</sup> elongated shoots were tested for rooting in ~~media~~<sup>MS1, MS2,</sup>  
107 ~~MS3, MS4 and MS5~~<sup>(size?)</sup>. The rooted Nanjing linden plantlets were transferred to small  
108 pots containing a mixture of sterilized peat and perlite (1:1 v/v), and after an  
109 acclimatization period of ~~four~~<sup>4</sup> weeks were transplanted into the field.

110 ~~Medium~~ MS1, MS2, MS3, MS4 and MS5 contained half-strength ~~MS~~<sup>medium (MS)</sup>  
111 (Murashige and Skoog (1962) with different concentrations of IBA (0.5, 1.0, 2.0, 3.0, or  
112 5.0  $\text{mg l}^{-1}$ ) (~~MS1, MS2, MS3, MS4, or MS5, respectively~~).

### 113 Culture conditions

114 All media in this study, excepting ~~the~~<sup>rooting</sup> medium, were supplemented with 30 g  
115  $\text{l}^{-1}$  sucrose, and solidified with 7.5 g  $\text{l}^{-1}$  agar (Sanland Chemical Co. Ltd., USA). ~~Then~~<sup>the</sup>  
116 the pH of the ~~media~~<sup>medium</sup> was adjusted to 5.8 before autoclaving at 121°C, 151 kPa for  
117 15min. All cultures were ~~incubated~~<sup>OK</sup> at 25±1°C with 14h light at a photosynthetic  
118 photon flux density of 50  $\text{mol m}^{-2} \text{s}^{-1}$ .

### 119 Statistical analysis

120 Each treatment comprised either 100 seeds or 20 explants, and each was repeated  
121 three times. Data on the number of shoots obtained per explant, germination rate of  
122 seeds, response <sup>of</sup> in explants, and number of nodes per shoot were analyzed using SPSS

123 13.0 for Windows (SPSS, Chicago, IL, USA). The mean differences were investigated  
124 at  $p \leq 0.05$  level, according to  <sup>$\sigma$</sup> One-way ANOVA analysis and LSD's multiple range  
125 rest. For paired-comparisons between treatments, post-hoc analysis and  
126 independent-samples t test were used.

## 127 RESULTS AND DISCUSSION

### Disinfection

#### 128 Disinfection of seeds

Report results to one decimal place. ✓

129 Treating the seeds with 5.65% sodium hypochlorite for 6 min and 0.1%  $\text{HgCl}_2$  for

130 6 min resulted in <sup>95.7</sup>~~95.67~~% of contamination-free seeds, while ~~through~~ <sup>produced 36.3%</sup> the treatment ✓

131 with 2.83% sodium hypochlorite for 6 min and 0.2%  $\text{HgCl}_2$  for 6 min <sup>36.33%</sup> of ✓

132 contamination-free seeds were ~~obtained~~. The treatments employing 5.65% (m/v) ✓

133 sodium hypochlorite for 12 min, and 0.1% (m/v) mercuric chloride ( $\text{HgCl}_2$ ) for 12

134 min resulted in <sup>96.3%</sup>~~96.33~~% and <sup>91.3%</sup>~~91.33~~% bacterial contamination, respectively. We found ✓

135 that the ~~disinfectant effect~~ of the combination of sodium hypochlorite and  $\text{HgCl}_2$  was ✓

136 significantly better than sodium hypochlorite or  $\text{HgCl}_2$  alone ( $F = 229.40$ ,  $p < 0.001$ ).

137 The ~~disinfectant~~ effect of the combination of sodium hypochlorite and  $\text{HgCl}_2$  was ✓

138 larger using high concentration sodium hypochlorite and low concentration  $\text{HgCl}_2$

139 than using low concentration sodium hypochlorite and high concentration  $\text{HgCl}_2$  ( $t =$

140 12.59,  $p < 0.001$ ), which agrees with the results of other studies (He et al. 2011).

### Breaking dormancy

#### 141 Dormancy breaking of seeds

142 In this study, we identified the dormancy <sup>breaking of dormancy was</sup> breaking of seeds as the most important ✓

143 limiting factor for successful, large scale Nanjing linden *in vitro* propagation. As the ✓

144 existence of a long dormancy period in linden seed had been reported previously ✓

145 (<sup>Chalupa</sup>~~Vladimir~~ 1990, <sup>Chalupa</sup>~~Vladimir~~ 1984, Yang et al. 2011), a preliminary study was carried out ✓

146 to <sup>determine</sup> find a procedure that would allow an effective <sup>by "break dormancy"</sup> dormancy breaking. The germination ✓

147 rates of seeds treated with the four treatments to break dormancy showed considerable

Table 1 can be deleted.

148 variation (Table 1). Seeds ~~treated with refrigeration~~ stored at 4°C for 2 d without ✓  
149 hot-treatment<sup>g</sup> showed <sup>28.3%</sup> 28.33% germination rate after <sup>2</sup> two weeks and 45.00% after ✓  
150 <sup>2</sup> two months. Seeds treated with a combination of hot-treatment<sup>g</sup> ~~(dipped into 40°C~~ <sup>incubated at</sup> ✓  
151 water for 2 h) and refrigeration<sup>g</sup> (stored at 4°C for 2 d) showed <sup>30%</sup> 30.00% germination ✓  
152 after <sup>2</sup> two weeks and <sup>49.7%</sup> 49.67% after <sup>2</sup> two months of culture, which <sup>was</sup> is in both cases not ✓  
153 significantly different from the ~~refrigeration-only~~ <sup>cold only</sup> treatment ( $t = 0.791$ ,  $p = 0.437$  and  $t$  ✓  
154  $= 1.306$ ,  $p = 0.262$ ; respectively). Seeds that were incubated in 500 mg l<sup>-1</sup> GA<sub>3</sub> for 3 d ✓ <sup>pm</sup>  
155 and cultured on basal medium without GA<sub>3</sub> showed only <sup>10.3%</sup> 10.33% and <sup>16.3%</sup> 16.33% ✓  
156 germination after <sup>2</sup> two weeks and <sup>2</sup> two months, respectively. However, completely ✓  
157 untreated seeds that were directly <sup>cultured</sup> incubated on WPM1 after their extraction from ✓  
158 fruits, showed a germination rate of <sup>33.3%</sup> 33.33% after <sup>2</sup> two weeks, and <sup>65.3%</sup> 65.33% after <sup>2</sup> two ✓  
159 months of culture, which <sup>was</sup> is in both cases higher than all other three treatments ( $t =$  ✓  
160 11.663,  $p < 0.001$ ;  $t = 32.870$ ,  $p < 0.001$ ). During germination the seeds started to ✓  
161 develop roots after <sup>4</sup> four weeks and a shoot was observable after <sup>5</sup> five weeks. When the ✓  
162 seedlings had developed into small plants after <sup>2</sup> two months (Fig. 1A), the nodal ✓  
163 explants were prepared from the shoots (Fig. 1B).

164

#### 165 Initiation of explants

166 Overall ZT performed better than BA with regards to the effectiveness of ✓  
167 initiating axillary buds <sup>on</sup> as explants (66-98% vs. 50-<sup>73%</sup> 70% responded buds), with ✓  
168 differences becoming significant for concentrations at 0.5 mg l<sup>-1</sup> <sup>ZT or BA?</sup> ( $t = 8.50$ ,  $p = 0.001$ ) ✓  
169 and 1.0 <sup>ZT or BA?</sup> (mg l<sup>-1</sup>) ( $t = 3.13$ ,  $p = 0.035$ ). This <sup>was</sup> is in accordance with results reported for ✓  
170 other woody species (e.g. Tatsuhiro et al. 1975, Kyong et al. 2001, Simon et al. 1993, ✓  
171 Gubis et al. 2004). Of the eight media formulations tested, WPM8 supplemented with ✓

172 0.5 mg l<sup>-1</sup> ZT and 0.04 mg l<sup>-1</sup> IBA yielded the best results as initiation medium for  
173 Nanjing linden explants (Table 2, Fig. 1C). ~~Not only~~ <sup>exhibited</sup> explants the highest  
174 induction response (98%), <sup>and</sup> ~~but~~ shoots also showed significantly faster growth,  
175 resulting in an average length of 6.8 cm and 4.3 nodes per shoot.

#### 176 *Shoot multiplication and elongation*

177 Shoot multiplication is an important factor for tissue culture, determining the  
178 suitability of the protocol in question for mass propagation (Quraishi et al. 1996).  
179 Zeatin <sup>was</sup> ~~is~~ the most widely used <sup>plant growth Regulator</sup> ~~growth hormone~~ in tissue culture techniques  
180 <sup>for</sup> ~~concerning~~ the propagation of woody species, as it <sup>was</sup> ~~has~~ proven to be an effective agent  
181 for shoot multiplication (Nour ~~et al.~~ 1993, Al-Juboory et al. 1998, Lucchesini and  
182 Mensuali-Sodi 2004, Sghir et al. 2005, Reed ~~et al.~~ 1991). In the present study, the  
183 average number of branches per explant after <sup>4</sup> ~~four~~ weeks of <sup>culture</sup> ~~incubation~~ was  
184 significantly higher ~~when grown~~ on medium containing both ZT and BA, as opposed  
185 to containing only one of the substances, ZT or BA (F = 4.471, p = 0.003; see Table 3).  
186 <sup>In addition</sup> ~~Additionally~~, the observed number of nodes correlated in the same way <sup>the</sup> ~~the~~ plants  
187 grown on medium containing ZT and BA <sup>had</sup> ~~having~~ a significantly higher average  
188 number of nodes per branch as plants grown on medium with one of the two  
189 substances missing (F = 3.524, p = 0.009; Table 3). The highest average number of  
190 branches per explant, <sup>(2.5)</sup> ~~as well as the highest average number of nodes per branch,~~ <sup>(4.5)</sup> ~~was~~  
191 obtained on WPM18 with 0.5 mg l<sup>-1</sup> ZT and 0.5 mg l<sup>-1</sup> BA combined with 0.02 mg l<sup>-1</sup>  
192 IBA <sup>(2.5 and 4.5, respectively)</sup> ~~respectively~~; Table 3).  
193 We conclude <sup>was</sup> ~~is~~ that the combination of both cytokinins, ZT and BA, <sup>was</sup> ~~is~~ a successful  
194 strategy to enhance shoot multiplication, as long as certain concentrations <sup>were</sup> ~~are~~ not  
195 exceeded.



196 The shoots obtained after the first multiplication step were cut into short nodal  
197 segments and were placed on WPM0, and <sup>cultured</sup> ~~incubated~~ for ~~one~~ month, for further  
198 elongation.

<sup>Rooting of shoots</sup>  
199 ~~Shoot~~ ~~rooting~~ and acclimatization

200 For rooting experiments, shoots 4-6 cm <sup>in length</sup> ~~long~~ were selected from healthy cultures.

201 Low salt medium containing low concentration of auxin (IBA, <sup>spell out the abbreviate</sup> ~~IAA~~ or NAA) has been

202 widely used as the basic rooting medium for several woody plants (<sup>Beasley</sup> ~~Rochelle~~ and

203 <sup>Pilot</sup> ~~Paula~~ 2013, Azad et al. 2005, Brissette et al. 1990, Rugini 1984). ~~Furthermore~~, <sup>earlier</sup>

204 studies on *in vitro* rooting of linden indicated that half-strength basal medium had a

205 higher instance of rooting success than full-strength basal medium (<sup>Chalupa</sup> ~~Vladimir~~ 1984,

206 Liu et al. 2009). Rooting induction was observed in rooting medium after <sup>2</sup> ~~two~~ weeks

207 of culture (Figure 1E). Over <sup>91.7%</sup> ~~91.67%~~ rooting was observed at 3 mg l<sup>-1</sup> IBA

208 ~~concentration~~, and <sup>30.7%, 50%, 70.3%, 88%</sup> ~~30.67%, 50.00%, 70.33% and 88.00%~~ rooting were observed at 0.5,

209 1.0, 2.0 <sup>or</sup> and 5.0 mg l<sup>-1</sup> IBA <sup>concentration</sup> (Figure 2). The rooting effect of IBA was

210 better in MS4 containing 3 mg l<sup>-1</sup> and MS5 containing 5 mg l<sup>-1</sup> than in MS1

211 containing 0.5 mg l<sup>-1</sup> (t=32.25, p<0.001, t=34.40, p<0.001). This result of a higher

212 instance of rooting at high concentrations of auxins contrasts with results obtained for

213 *T. cordata*, where a high proportion of rooting was induced at much lower

214 concentrations (<sup>Chalupa</sup> ~~Vladimir~~ 1984). Plants <sup>with</sup> ~~in which~~ roots had ~~successfully~~ been induced

215 were transplanted to pots containing a mixture of sterilized peat and perlite (1:1 v/v)

216 and were then grown for 3-4 weeks in a greenhouse under high humidity conditions.

217 The survival rate for ~~this late step in the protocol~~ was always very high, with 95-100%

218 of the root-induced <sup>shoots</sup> ~~explants~~ surviving. The transfer to the field <sup>was</sup> ~~would~~ be initiated by

219 slowly reducing humidity levels, and thereby acclimatizing the plants to outside

220 conditions, and finally transplantation to the field.

Were plants actually  
transplanted to the field?  
If so, please show a photo.

221 The present study introduces<sup>d</sup> a new customized protocol for the *in vitro* ✓  
222 propagation of Nanjing linden (*Tilia miqueliana*). By fine tuning culture media used for  
223 ~~the~~<sup>y</sup> several steps, we established an overall improved method for generating clonal ✓  
224 plants from explants obtained from axial<sup>lary</sup> buds<sup>in vitro</sup> of seedlings. It may also be possible to ✓  
225 use this protocol to obtain clones from mature tissues of *T. miqueliana* through  
226 micropropagation, and we hope this will contribute to a better supply of this highly  
227 valued species for landscaping purposes.

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\* Check ALL References for Accuracy AS \*  
MANY have the first name instead of last name,  
WRONG  
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NAME, etc.

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\* Report All results to one decimal place \*

\* Report all plant growth regulators in micromolar ( $\mu\text{M}$ )

265 Table1. Germination rate of Nanjing linden seeds after application of different dormancy breaking treatments.

Treatment	% of seed germinated	
	After 2 weeks	After 2 months
stored at 4°C for 2 d without hot-treatment	28.33±0.58 b	45.00±1.00 b
incubated in 40°C water for 2 h	30.00±3.61 bc	49.67±6.11 b
incubated in 500 mg l <sup>-1</sup> gibberellic acid for 3 d	10.33±1.53 a	16.33±2.08 a
Cultured on modified WPM with 3 mg l <sup>-1</sup> gibberellic acid	33.33±3.06 c	65.33±1.53 c

266 One hundred seeds per treatment with three replicates were evaluated. Data for germination of seeds were scored  
267 after two weeks and two months of culture.

268  
269 Table2. Effects of ZT and BA on axillary bud initiation in Nanjing linden

Medium No.	Plant growth regulators (mg l <sup>-1</sup> )			Responded explants (%) (mean ± SD)	Length of shoots (cm) (mean ± SD)	No. of nodes per shoot (mean ± SD)
	BA/ZT	IBA				
WPM2	BA 0.01	0.04		50±8.7 a	4.3±0.41 a	2.4±0.30 a
WPM3	BA 0.20	0.04		73±2.9 b	4.5±0.31 a	2.8±0.10 ab
WPM4	BA 0.50	0.04		70±5.0 b	5.1±0.39 ab	2.1±0.28 a
WPM5	BA 1.00	0.04		68±7.6 b	4.3±0.24 a	2.8±0.20 ab
WPM6	ZT 0.01	0.04		66±15.2 b	5.5±0.26 bc	3.3±0.33 bc
WPM7	ZT 0.20	0.04		80±5.0 bc	5.8±0.44 bc	3.6±0.21 c
WPM8	ZT 0.50	0.04		98±2.9 d	6.8±0.12 d	4.3±0.44 d
WPM9	ZT 1.00	0.04		92±10.4 cd	6.0±0.54 c	3.8±0.60 cd

270 Data recorded after three weeks of culture in initiation medium. Twenty explants were cultured for each treatment  
271 with three replicates. Means followed by the same letters are significantly different at  $p \leq 0.05$ , according to LSD's  
272 multiple range test.

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275 Table 3. Effects of ZT and BA on shoot multiplication in Nanjing linden

Medium No.	Plant growth regulators (mg l <sup>-1</sup> )			Responded shoots (%) (mean ± SD)	No. of branches (mean ± SD)	No. of nodes per branch (mean ± SD)
	BA	ZT	IBA			
WPM10	0.01	0.00	0.02	80±8.6 a	1.0±0.00 a	4.4±0.55 b
WPM11	0.10	0.00	0.02	83±7.6 ab	1.0±0.00 a	3.8±0.44 ab
WPM12	0.50	0.00	0.02	88±6.2 ab	1.5±0.46 ab	4.0±0.60 ab
WPM13	1.00	0.00	0.02	77±15.2 a	2.0±0.41 ab	4.8±0.15 b
WPM14	0.00	0.01	0.02	78±12.6 a	1.1±0.13 a	3.0±0.35 a
WPM15	0.00	0.10	0.02	85±10.0 ab	1.9±0.36 ab	4.3±0.83 ab
WPM16	0.00	0.50	0.02	90±10.0 ab	1.5±0.50 ab	3.4±0.20 ab
WPM17	0.00	1.00	0.02	80±0.0 a	2.0±0.88 ab	3.6±0.55 ab
WPM18	0.50	0.50	0.02	98±2.8 b	2.5±0.53 b	4.5±0.30 b
WPM19	1.00	1.00	0.20	92±14.4 ab	1.1±0.08 a	3.8±0.74 ab

276 Plants were measured after three weeks of culture in initiation medium. Twenty explants were cultured for  
277 each treatment with three replicates. Means followed by the same letters are significantly different at  $p \leq 0.05$ ,  
278 according to LSD's multiple range test.

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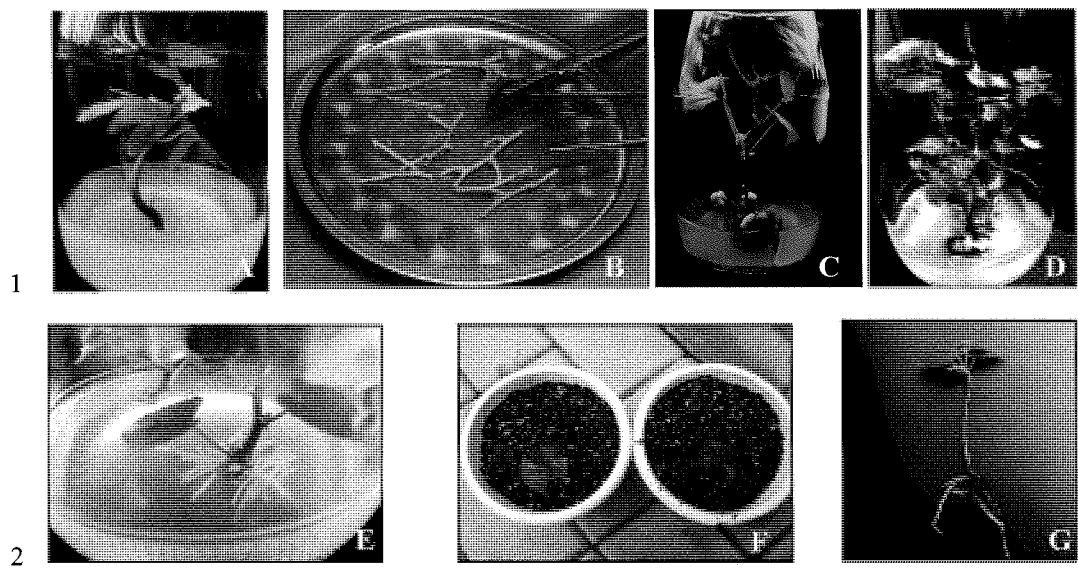
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All tables must stand alone from the text. Spell out abbreviations in a footnote for each table. e.g. WPM, BA, ZT, IBA.

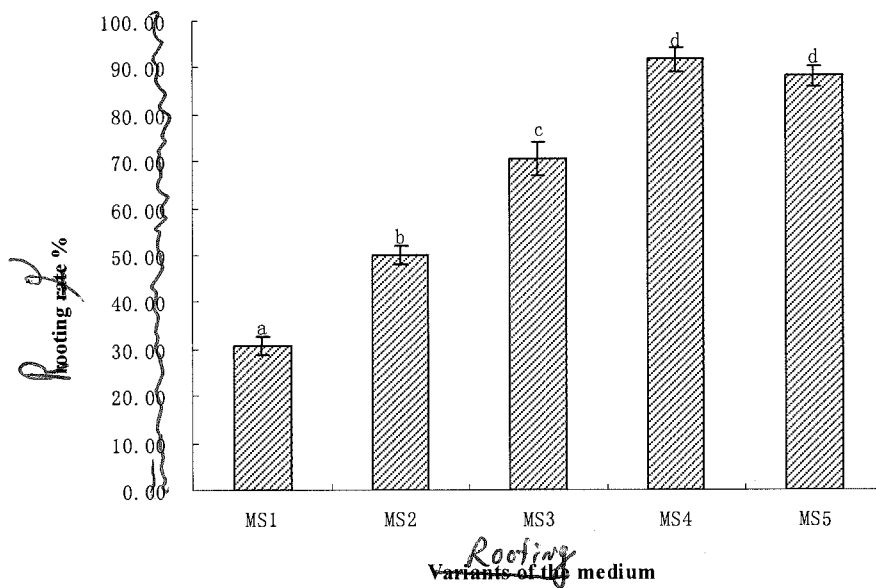


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3 Fig.1. Micropropagation of Nanjing linden (*T. miqueliana*) A) Seedling developed from disinfected seed, B) Nodal  
4 explants used for culture, C) Elongated shoots on elongation medium supplemented with 0.04 mg l<sup>-1</sup> IBA, 0.5 mg l<sup>-1</sup>  
5 ZT, D) Multiplication of axillary shoots, E) Roots induced on rooting medium, F) Plantlets transferred into small  
6 pots, G) Fully established pot plants of Nanjing linden.

Rooted

disinfested

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 2 **Fig. 2.** Effects of <sup>of IBA</sup> the plant growth regulators on the rooting rate of shoots. MS1: 0.5  $\text{mg l}^{-1}$  IBA, MS2: 1.0  $\text{mg l}^{-1}$  IBA, MS3: 2.0  $\text{mg l}^{-1}$  IBA, MS4: 3.0  $\text{mg l}^{-1}$  IBA, MS5: 5.0  $\text{mg l}^{-1}$  IBA. Treatments with different letters ~~are~~ <sup>were</sup>  
 3 significantly different at  $p \leq 0.05$  according to LSD's multiple range tests; Vertical bars represent mean values of  
 4 three replications  $\pm$  SD.