

# Getting started with the SPLITT library

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**SPLITT** is a one-header C++ library.

```
knitr::include_graphics("figures/UmlDiagram8.pdf")
```

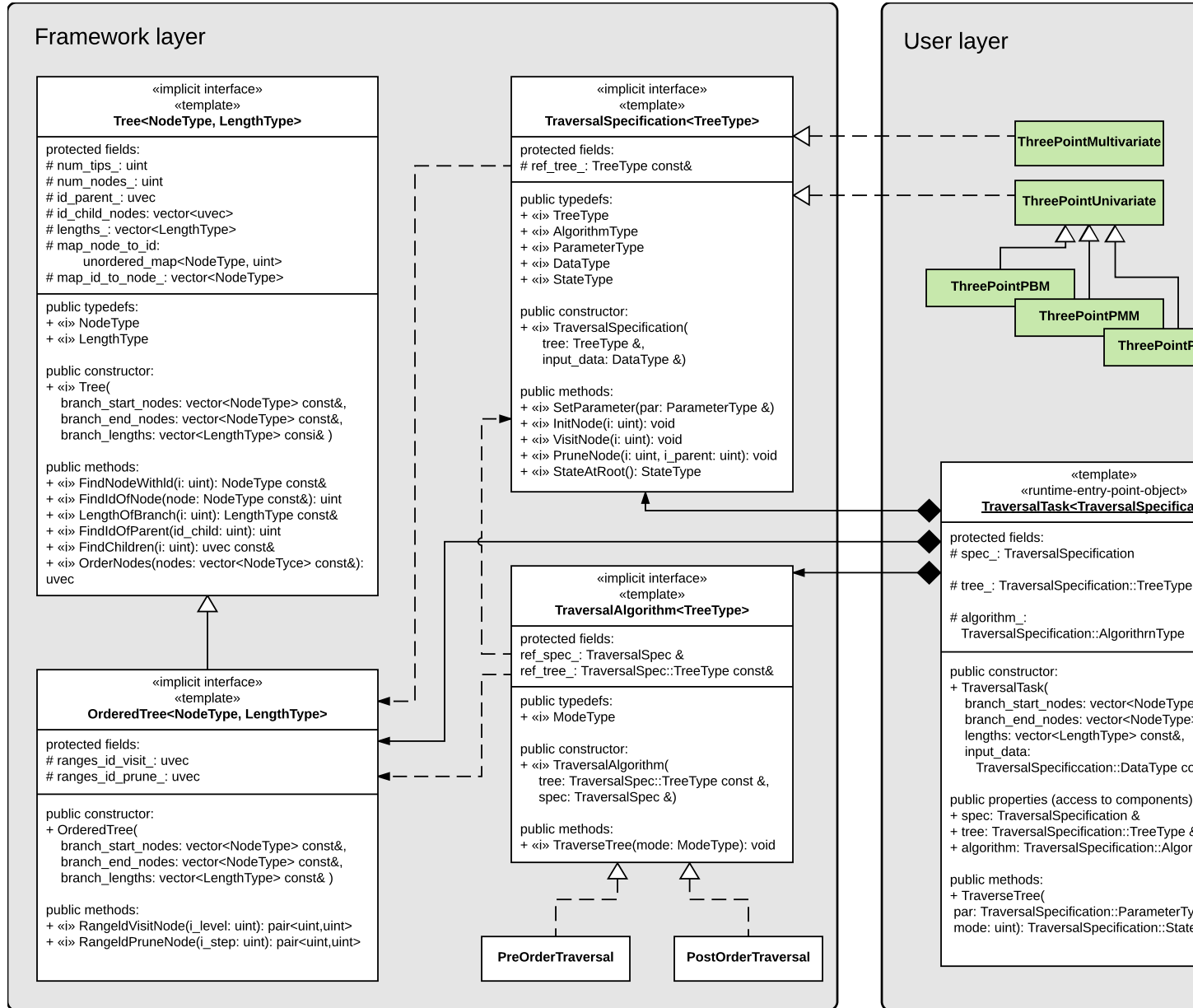
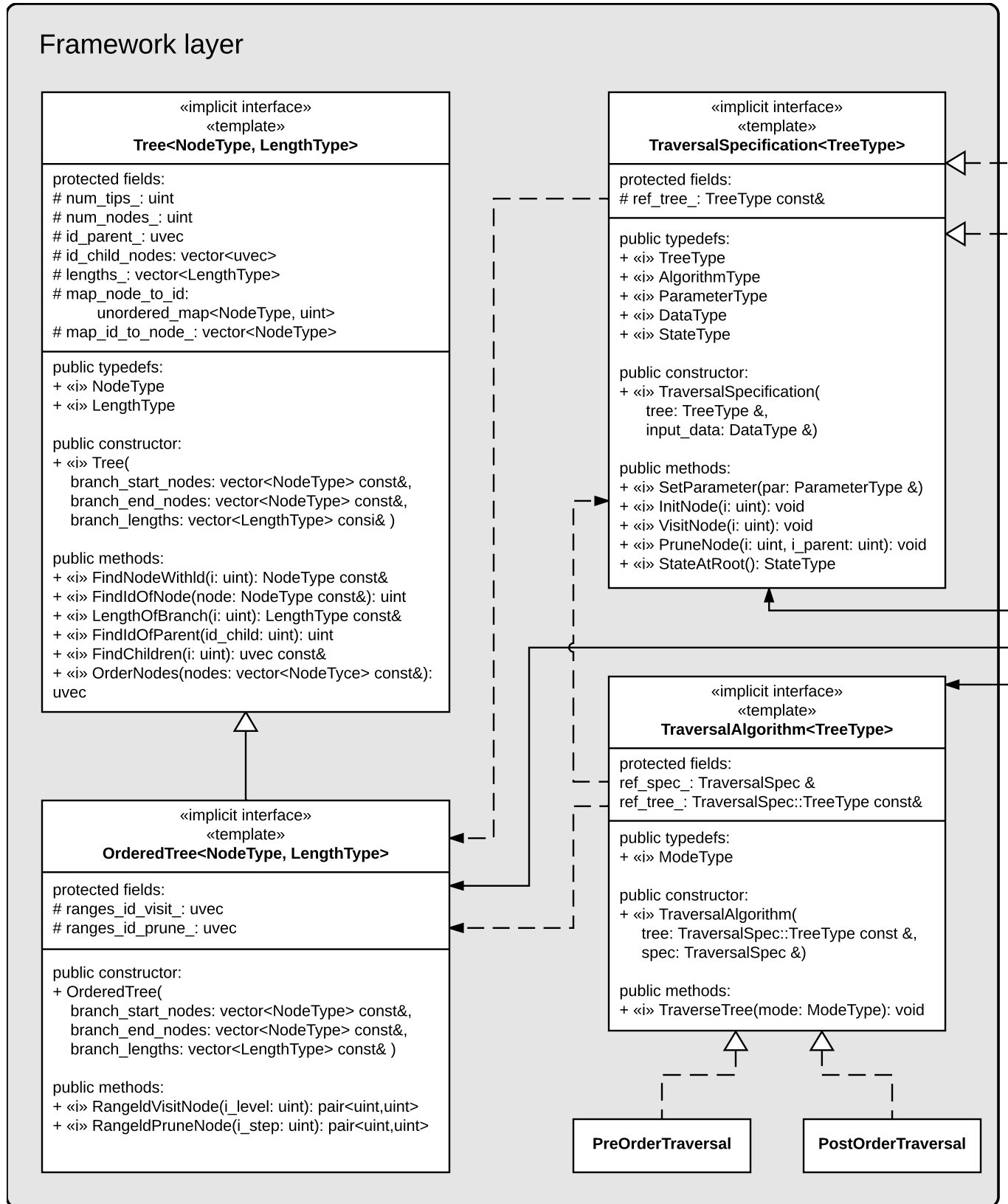


Figure 1:



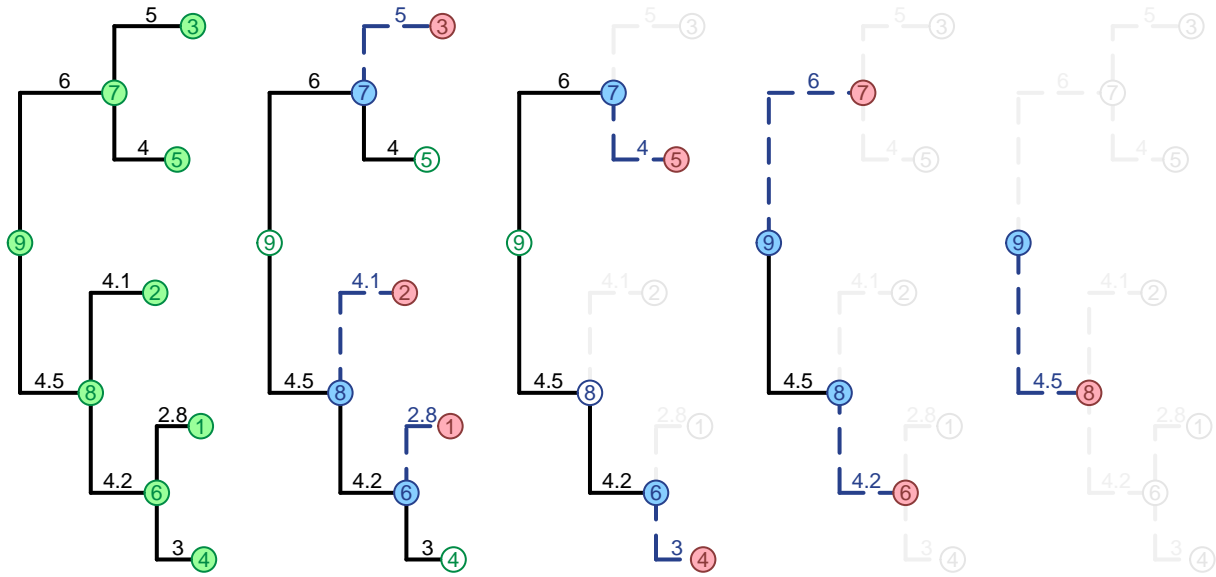
Here I show how to use **SPLITT** to estimate the broad-sense heritability,  $H^2$ , of a numeric pathogen trait.  $H^2$  is defined as the proportion of the trait variance attributable to genotypic variation in a population of

hosts (Mitov and Stadler 2016; Mitov and Stadler 2018):

## Packages used

```
newick <- '(((1:3,2:2.8)6:4.2,3:4.1)7:4.5,(4:4,5:5)8:6)0;'  
tree <- read.tree(text=newick)  
  
x0 <- 0.1  
sigma2 <- 0.25  
sigmae2 <- 1  
  
set.seed(1)  
g <- rTraitCont(tree, model = "BM", root.value = x0,  
               sigma = sqrt(sigma2),  
               ancestor = FALSE)  
  
x <- g + rnorm(n = length(tree$tip.label), mean = 0, sd = sqrt(sigmase2))  
  
PMMLogLikCpp(x, tree, x0, sigma2, sigmae2, mode = 1)  
  
## [1] -8.2  
  
# output  
# InitNode(1): x[i] t[i] a[i] b[i] c[i] : 1.9 2.8 -0.5 1.9 -2.72  
# InitNode(2): x[i] t[i] a[i] b[i] c[i] : 0.96 4.1 -0.5 0.96 -1.38  
# InitNode(3): x[i] t[i] a[i] b[i] c[i] : -0.74 5 -0.5 -0.74 -1.2  
# InitNode(4): x[i] t[i] a[i] b[i] c[i] : 1.79 3 -0.5 1.79 -2.53  
# InitNode(5): x[i] t[i] a[i] b[i] c[i] : 0.46 4 -0.5 0.46 -1.03  
# InitNode(6): x[i] t[i] a[i] b[i] c[i] : nan 4.2 0 0 0  
# InitNode(7): x[i] t[i] a[i] b[i] c[i] : nan 6 0 0 0  
# InitNode(8): x[i] t[i] a[i] b[i] c[i] : nan 4.5 0 0 0  
# InitNode(9): x[i] t[i] a[i] b[i] c[i] : nan 1.4822e-323 0 0 0  
# VisitNode(1): x[i] t[i] a[i] b[i] c[i] : 1.9 2.8 -0.29 1.12 -2.25  
# PruneNode(1, 6): x[i] t[i] a[j] b[j] c[j] : 1.9 2.8 -0.29 1.12 -2.25  
# VisitNode(2): x[i] t[i] a[i] b[i] c[i] : 0.96 4.1 -0.25 0.47 -1.5  
# PruneNode(2, 8): x[i] t[i] a[j] b[j] c[j] : 0.96 4.1 -0.25 0.47 -1.5  
# VisitNode(3): x[i] t[i] a[i] b[i] c[i] : -0.74 5 -0.22 -0.33 -1.45  
# PruneNode(3, 7): x[i] t[i] a[j] b[j] c[j] : -0.74 5 -0.22 -0.33 -1.45  
# VisitNode(4): x[i] t[i] a[i] b[i] c[i] : 1.79 3 -0.29 1.03 -2.12  
# PruneNode(4, 6): x[i] t[i] a[j] b[j] c[j] : 1.79 3 -0.58 2.14 -4.36  
# VisitNode(5): x[i] t[i] a[i] b[i] c[i] : 0.46 4 -0.25 0.23 -1.32  
# PruneNode(5, 7): x[i] t[i] a[j] b[j] c[j] : 0.46 4 -0.47 -0.1 -2.77  
# VisitNode(6): x[i] t[i] a[i] b[i] c[i] : nan 4.2 -0.26 0.97 -3.68  
# PruneNode(6, 8): x[i] t[i] a[j] b[j] c[j] : nan 4.2 -0.51 1.44 -5.18  
# VisitNode(7): x[i] t[i] a[i] b[i] c[i] : nan 6 -0.2 -0.04 -3.2  
# PruneNode(7, 9): x[i] t[i] a[j] b[j] c[j] : nan 6 -0.2 -0.04 -3.2  
# VisitNode(8): x[i] t[i] a[i] b[i] c[i] : nan 4.5 -0.24 0.67 -5.01  
# PruneNode(8, 9): x[i] t[i] a[j] b[j] c[j] : nan 4.5 -0.43 0.63 -8.22  
  
par(mfrow=c(1,5))  
par(mar=c(0,0,0,0))  
par(oma=c(0, 0,0, 0))
```

```
PlotParallelTraversal(tree)
```



```
## C++ object <0x10a930ef0> of class 'SPLITT__RCPP__OrderedTreeStringNodes' <0x101f3a7f0>
```

```
#newick <- '(((1:4,2:4.5)11:4.3,3:4)10:1.4,(4:10.25,((5:4,6:3.8)14:4.2,7:4.1)13:4.5,(8:6,9:6)15:6)12:2.0);'
```

```
newick <- '(((1:4,2:4.5)11:4.3,3:4)10:3.8,(((5:3,6:2.8)14:4.2,7:4.1)13:4.5,(8:5,9:5)15:6)12:4.2)0;'
```

```
tree <- read.tree(text=newick)
```

```
par(mfrow=c(1,5))
```

```
par(mar=c(0,0,0,0))
```

```
plotParallelPruningOrder(tree, TRUE)
```

```
fig.a <- recordPlot()
```

```
newick <- "((((1:3.2,2:3.2)10:3.2,3:3.2)11:3.2,4:3.2)12:3.2,5:3.2)13:3.2,6:3.2)14;"
```

```
tree <- read.tree(text=newick)
```

```
par(mfrow=c(1,6))
```

```
par(mar=c(0,0,0,0))
```

```
plotParallelPruningOrder(tree, TRUE)
```

```
fig.b <- recordPlot()
```

```
cowplot::plot_grid(fig.a, fig.b, nrow=2, labels = c("a", "b"))
```

Apart from base R functionality, the patherit package uses a number of 3rd party R-packages:

- For tree processing: ape v5.1 (Paradis et al. 2016);
- For reporting: data.table v1.11.2 (Dowle and Srinivasan 2016);
- For the POUMM and PP estimates: POUMM v2.1.0 (Mitov and Stadler 2017), boot v1.3.20 (Canty and Ripley 2016);
- For testing: testthat v2.0.0 (Wickham 2016).

## References

- Canty, Angelo, and Brian Ripley. 2016. *Boot: Bootstrap R (S-Plus) Functions*. *R-Package*. <https://cran.r-project.org/web/packages/boot/index.html>.
- Dowle, Matt, and Arun Srinivasan. 2016. *Data.table: Extension of 'Data.frame'*. <https://CRAN.R-project.org/package=data.table>.
- Mitov, Venelin, and Tanja Stadler. 2016. "The heritability of pathogen traits - definitions and estimators." *Unpublished Data* <https://www.biorxiv.org/content/early/2016/06/12/058503>.
- . 2017. *Fast and Robust Inference of Phylogenetic Ornstein-Uhlenbeck Models Using Parallel Likelihood Calculation*. *BiorXiv*. <http://biorxiv.org/content/early/2017/05/30/115089>.
- . 2018. "A Practical Guide to Estimating the Heritability of Pathogen Traits." *Molecular Biology and Evolution*, msx328. doi:10.1093/molbev/msx328.
- Paradis, Emmanuel, Simon Blomberg, Ben Bolker, Julien Claude, Hoa Sien Cuong, Richard Desper, Gilles Didier, et al. 2016. *Ape: Analyses of Phylogenetics and Evolution*. <https://CRAN.R-project.org/package=ape>.
- Wickham, Hadley. 2016. *Testthat: Unit Testing for R*. <https://CRAN.R-project.org/package=testthat>.