



Timely Accurate Diagonostics for a TB-Free Africa

# Training on *Mycobacterium tuberculosis*drug susceptibility testing (first and second line LJ DST)

Module 9: Setting of LJ DST isolates

Venue:

Presenter:

Date:

## **Introduction and Objective**

#### Introduction

•This module details the steps involved in setting LJ DST from clinical isolates.

#### **Objectives:**

At the end participates should be able;

- •To prepare the bacterial suspension of MacFarland 1.0 turbidity and make the required serial dilutions.
- •To set LJ DST from bacterial suspensions of clinical isolates.

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#### **Module Outline**

- Overview of LJ DST
- Materials required for LJ DST.
- Preparation of 1.0 McFarland bacteria suspension
- Preparation of bacteria dilutions
- LJ DST Inoculation





## Over view of LJ DST

- This is a proportional DST technique based on determining the 1% MTBc resistant population that is clinically significant.
- It is based on the principle of comparing the growth on drug free media (growth control) to the growth on drug containing media.
- The strain is presumed to be resistant when growth of more than a certain proportion of the inoculum (critical proportion) occurs on culture media containing a defined drug concentration(critical concentration)





#### Over view of LJ DST

#### critical concentration

The lowest concentration of an anti-TB drug in the culture medium at which growth of tubercle bacilli indicates resistance of clinical significance.

#### critical proportion

The percentage of tubercle bacilli in the inoculum whose growth on culture media containing the critical concentration of an anti-TB drug signifies the clinical ineffectiveness of that drug.





#### Over view of LJ DST

#### **Growth control:**

\*Culture yielded after inoculation of tubercle bacilli on a culture medium without any test drug in order to exhibit unrestricted growth.





## Materials/Equipment

#### Reagents/ materials

- Sterile distilled water
- Calibrated pasture pippetes(1ml and 3 ml)
- Khan tubes.
- Culture isolates positive for MTBc.
- Sterile Inoculation loops
- Universal bottles
- Plain LJ media

#### **Materials**

PNB growth control (500µg/ml)

MacFarland 1.0

Reference strains(H37Rv)

#### Equipment

- Biosafety cabinet
- Incubator
- Votexer





## Materials required for LJ **DST** setting

1st and 2nd Line LJ DST media of the following critical concentrations:

DRUG	Critical concentration (µg/ml)
Isoniazid	0.2
Rifampicin	40
Ethambutol	2.0
Moxifloxacin	1.0
Levofloxacin	2.0
Amikacin	30





## Work procedure(1)

- Prepare 1.0 Macfarland bacteria suspension as below:
  - With a loop, scrape colonies from all over the culture (try to pick up portions from all colonies and not media because it gives you false turbidity).
  - \*Use a sterile, small, thick-walled screw-capped glass tube containing 5-7 sterile glass beads (approximately 3 mm in diameter).
  - Gently shake the loop over the beads.





## Work procedure(2)

## Preparation of 1.0 Macfarland suspension(neat suspension):

- Add some drops of sterile saline or distilled water, shake, add 2 further drops and vortex.
- \*Let stand for 30 minutes to allow the larger aggregates of bacteria to settle.
- Transfer the homogenous upper part of the supernatant into another sterile tube without disturbing the sediment. It should not contain any visible clumps.

## Work procedure(3)

Preparation of 1.0 McFarland suspension (neat suspension):

Adjust the turbidity of the bacterial suspension to match the McFarland standard No.1. it approximates to 1mg of wet bacterial mass/ml

#### NB;

- If the suspension is too turbid, add some drops of sterile distilled water.
- If the suspension is insufficiently turbid, do not add more cells to the suspension (because mycobacterial cells are difficult to homogenize);
- Let the suspension settle, discard some of the supernatant to concentrate cells, and adjust the turbidity by adding few drops of distilled water.



A Wrong Mcfarland concentration can result into false resistance or false susceptible results?

## Work procedure(4)

#### Preparation of bacteria dilutions:

- Make serial 10-fold dilutions of the standard suspension by diluting sequentially 1.0 ml of the culture suspension in tubes containing 9 ml of sterile distilled water or normal saline (0.85% sodium chloride).
- Make sure to mix each dilution thoroughly.





## Work procedure(4)

#### Preparation of bacteria dilutions:

- Dilutions of 10-2 (for Control 1) and 10-4 (for Control 2) should be inoculated as the growth controls (that is, LJ medium without any anti-TB agents).
- Some laboratories inoculate duplicate LJ tubes for both Control 1 and Control 2.
- One tube with medium containing each anti-TB agent is inoculated with only the 10-2 dilution.





## Work procedure (5)

#### LJ DST DST Inoculation;

- The volume of the inoculum should be 0.1 ml.
- The inoculum should be placed onto the middle two thirds of the slant avoiding the edges.
- Ensure that the inoculum is spread evenly on the middle part of the slide by slighty tilting the inoculated slant.
- Ensure that all tubes are properly labelled.



## Work procedure (6)

#### Incubation

- •After inoculation, the tubes may be placed at an angle and are incubated at 37 OC ± 1 OC.
- •Some laboratories incubate the tubes with the screw caps slightly loosened to allow for evaporation of the inoculum.
- •Then, after 24-48 hours, the caps are tightened and the tubes are incubated for about 6 weeks.
- •Extreme caution should be taken if the caps are left loose since strains of MDR-TB and XDR-TB pose a potentially major biosafety risk.





## Work procedure(7)

- Document your work on the LJ DST work sheet/register.
- Read at 4<sup>th</sup> week, final results for ethambutol and PNB and preliminary results for the rest of the drugs.
- Incubate for more 2 weeks and read final results at 6<sup>th</sup> week for other drugs.

#### NB:

- 1. The inoculated media should always be examined for contamination after 1 week of incubation.
- 2.Interpretable results for all drugs can be concluded at weeks

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Reference Laboratory

#### **Assesment**

- Describe the principle behind LJ DST?
- Define Critical concentration and critical proportion as applied in LJ DST?
- What materials are required for LJ DST setting?
- What is the relevance of Macfarland 1.0 in LJ DST?





## **Summary**

- LJ DST technique is based on the principle of comparing growth on drug containing media to the growth on drug free media to ascertain the 1% clinically significant MTBc population.
- A Wrong McFarland concentration can result into false resistance or false susceptible results?





#### References

- GLI TB training package http://www.stoptb.org/wg/gli/trainingpackages.asp
- https://www.who.int/tb/publications/2018/WHO\_technical\_d rug\_susceptibility\_testing/en/





## **Acknowledgments**



















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