

Global ecological drivers of transpiration regulation in trees

Program: Doctorat en Ecologia Terretre

Centre de Recerca Ecològica i Aplicacions Forestals

Autonomous University of Barcelona

Author:

Víctor Flo Sierra

Advisors:

Rafael Poyatos López
Jordi Martínez Vilalta

January 2021



Acknowledgements

Mucha gente es corresponsable de este trabajo, de haberme empujado y acompañado hasta aquí. Me gustaría empezar agradeciendo a mis directores de tesis, Rafa y Jordi, sin los que esta tesis no hubiera sido posible, gracias por la oportunidad de trabajar con vosotros. He sido muy afortunado de haber contado con dos mentores excelentes en lo profesional y en lo personal. Rafa, tu dedicación es enviable, gracias por todas las buenas ideas, los comentarios y críticas, las correcciones perfectas, por los viajes a congresos y por sufrir mi insomnio y jet-lag. Jordi, ha sido un privilegio tenerte como tutor, por tu conocimiento transversal y profundo, por tu ágil e innata capacidad de entenderlo todo, por mejorar siempre todas las ideas y textos, y por tu calidad humana.

Esta tesis tampoco hubiera sido igual sin haber compartido estos cuatro años con tantos buenos compañeros. Gracias a los compañeros de despacho, a Anna, Javi, Judit, Carlos, Manu, Aina, Marta, Pere y Pol. Creo que habeis sido los mejores compañeros posibles y que tuve mucha suerte de acabar en el -150. Sé que me llevo grandes amistades para toda la vida. No quiero olvidarme tampoco de muchos otros compañeros como Jordi, Sara, Kevin, Xavi, Vicenç, Joan, Marina, con los que he compartido café, discusiones y charlas. Gracias también a Luciana por acompañarnos en ese genial viaje relámpago por el desierto americano.

Gracias también en especial a los compañeros que forman o han formado parte del grupo. Maurizio, Tere, Miquel, Lucía, Víctor, Raúl, Rosella, Pipo, Mireia, Lies, David, Jacob, Aude, Antonine, Pablo, Luca y Brenda, sin duda un grupo enviable. Gracias por tantos buenos momentos, por la dimensión académica, pero especialmente por las salidas de grupo, por las barbacoas, cenas y cervezas. Me gustaría agradecer especialmente a Víctor Granda, por su excelente trabajo, sin el que hubiera sido imposible llevar a cabo SAPFLUXNET. Gracias por enseñarme a querer a R, pero sobre todo por las risas y los enfados jugando a basquet. A Jordi Bosch, Anselm Rodrigo, Joan Oca e Ingrid Masaló por permitirme iniciarme en el mundo de la investigación.

Durante estos años he tenido la suerte de vivir y conocer a mucha gente lejos de casa. Muchas personas que me han enseñado otras maneras de ver el mundo. Me gustaría mencionar especialmente a Martín y a Elena, gracias por acogerme y hacerme sentir como en casa en Salt Lake City. Gracias por darnos cobijo cuando nos quedamos en la calle por unos días, dejarme vuestro coche, y gracias por un viaje inolvidable a Glacier. No os lo podré agradecer suficiente. Gracias a Bill Anderegg por acogerme en su laboratorio donde he tenido la suerte de conocer a gente genial como Kelly, Nicole, Jaycee y Grayson. Me gustaría agradecer también a Kathy Steppe por darme la oportunidad de visitar su laboratorio en Ghent y por sus buenos consejos. A Roberto por enseñarme a intalar mi primera sonda de flujo de savia.

Gracias también a los que llevan tiempo compartiendo su camino conmigo o han caminado alguna etapa, me han marcado y ayudado, a los amigos de toda la vida y a mis compañeros de andanzas musicales. Gracias Trufa, Pablo, Marc, Laura, Óscar, Carles, Jordi, Larry, Cristian, Pol, Kuni, Julio, Lorenzo, Bárbara, Sara, Edgar, Jan, Arnau. También a Marta, Rosa, Florian, Daniel, Lola, Magdalena y Esther, por haber creído tanto en mi y haber sido parte de mi vida durante muchos años. A Tuti, Tona y Flor porque os echo de menos.

Quiero agradecer también a mi familia. A mis abuelos, a mis tíos y primos, por haber sido una referencia.

Gracias sobre todo a quien desde hace cuatro años es mi compañera de viaje. Gracias Mari Ángeles por tu sonrisa, tu apoyo incondicional y tu amor. Esta tesis no hubiera sido sin ti. Gracias por ser brillante, porque es un placer escucharte y por intentar “salvar al mundo”. Gracias por darme una familia murciana.

Gracias finalmente a mis padres, por dejarme decidir mi camino, por apoyarme en cada decisión que he tomado. Me habéis hecho libre y feliz. Mil gracias Ana por haber creído en mi, por ser un ejemplo desde que éramos pequeños, por ser tan buena y trabajadora. Si alguien es responsable directo de que me lanzara a recorrer este camino esa eres tú. Gracias Clara por ser la alegría de nuestra casa y ser una hermana maravillosa. Gracias Marc y Jose por formar parte de nuestra vida y por esas charlas sobre tecnología y negocios. Para acabar, gracias también a Arnau, el último fichaje de la familia. ¡Os quiero!

Table of Contents

| | |
|--|-----------|
| 1: Introduction | 1 |
| 1.1 Transpiration and earth system functioning | 2 |
| 1.2 Transpiration across scales | 4 |
| 1.2.1 Water transport in plants | 4 |
| 1.2.2 Upscaling to the stand and ecosystem levels | 5 |
| 1.3 Transpiration quantification | 6 |
| 1.4 Transpiration in a changing world | 7 |
| 1.5 Research aims and outline | 8 |
| | |
| 2: Bias and uncertainty in sap flow methods | 11 |
| 2.1 Introduction | 13 |
| 2.2 Material and methods | 15 |
| 2.2.1 Sap flow calibration datasets | 15 |
| 2.2.2 Calibration assessment | 16 |
| 2.2.3 Statistical analyses | 19 |
| 2.3 Results | 20 |
| 2.3.1 Calibration performance compared among methods and families of methods | 20 |
| 2.3.2 Influence of wood traits | 23 |
| 2.4 Discussion | 24 |
| 2.4.1 Sap flow measurement errors across methods and methodological families . | 25 |
| 2.4.2 The performance of sap flow calibrations is largely unrelated to species wood traits | 27 |
| 2.4.3 Implications and recommendations | 30 |
| 2.5 Conclusions | 31 |
| | |
| 3: The SAPFLUXNET database | 35 |
| 3.1 Introduction | 37 |
| 3.2 The SAPFLUXNET data workflow | 38 |
| 3.2.1 An overview of sap flow measurements | 38 |
| 3.2.2 Data compilation | 40 |
| 3.2.3 Data harmonisation and quality control: QC1 | 40 |
| 3.2.4 Data harmonisation and quality control: QC2 | 42 |
| 3.2.5 Data structure | 42 |
| 3.3 The SAPFLUXNET database | 43 |
| 3.3.1 Data coverage | 43 |

| | | |
|-----------|--|-----------|
| 3.3.2 | Methodological aspects | 45 |
| 3.3.3 | Plant characteristics | 48 |
| 3.3.4 | Stand characteristics | 48 |
| 3.3.5 | Temporal characteristics | 50 |
| 3.3.6 | Availability of environmental data | 51 |
| 3.4 | Potential applications | 52 |
| 3.4.1 | Applications in plant ecophysiology and functional ecology | 52 |
| 3.4.2 | Applications in ecosystem ecology and ecohydrology | 56 |
| 3.5 | Limitations and future developments | 58 |
| 3.5.1 | Limitations | 58 |
| 3.5.2 | Outlook | 59 |
| 3.6 | Data availability, access and feedback | 59 |
| 3.7 | Conclusion | 60 |
| 4: | Analysis of hydrometeorological drivers. | 63 |
| 4.1 | Introduction | 65 |
| 4.2 | Material and methods | 66 |
| 4.2.1 | Sapflow and environmental data | 66 |
| 4.2.2 | Data filtering | 66 |
| 4.2.3 | Whole-tree canopy conductance calculation | 67 |
| 4.2.4 | Hydrometeorological coupling quantification | 67 |
| 4.2.5 | Biome classification and site-level bioclimatic data | 68 |
| 4.2.6 | Statistical analyses | 68 |
| 4.3 | Results | 69 |
| 4.4 | Discussion | 72 |
| 5: | Climate and functional traits mediate tree water use strategies | 77 |
| 5.1 | Introduction | 79 |
| 5.2 | Material and methods | 81 |
| 5.2.1 | Sap flow data | 81 |
| 5.2.2 | Evaporative demand and soil water availability data | 81 |
| 5.2.3 | Data filtering | 83 |
| 5.2.4 | Whole-tree canopy conductance calculation | 83 |
| 5.2.5 | Traits and climatic data | 84 |
| 5.2.6 | G_{Asw} sensitivity to soil water availability and water demand | 84 |
| 5.2.7 | Statistical analyses | 85 |
| 5.3 | Results | 87 |
| 5.3.1 | Water use parameters and differences between angiosperms and gymnosperms | 87 |
| 5.3.2 | Coordination with hydraulic and allocation traits | 88 |
| 5.4 | Discussion | 91 |
| 5.4.1 | Climate influence on water use strategies across species | 92 |
| 5.4.2 | Water use parameters | 93 |
| 5.4.3 | Coordination between water use parameters and water relations traits | 93 |
| 5.4.4 | Conclusions | 95 |

| | |
|---|------------|
| 6: General discussion and conclusions | 97 |
| 6.1 Improving sap flow techniques to obtain reliable estimates of tree level transpiration | 98 |
| 6.2 The SAPLUXNET database. What next? | 99 |
| 6.3 The hydroclimatic drivers of transpiration and their complexity | 100 |
| 6.4 On the use of plant traits to characterize water use strategies at the global level . . | 101 |
| 6.5 Towards a global characterization of plant water use strategies. | 102 |
| 6.6 Conclusions | 103 |
| Appendix A: Appendix Chapter 2 | 105 |
| A.1 Figures A | 106 |
| A.2 Tables A | 109 |
| Appendix B: Appendix Chapter 3 | 113 |
| B.1 Figures B | 114 |
| B.2 Tables B | 119 |
| Appendix C: Appendix Chapter 4 | 135 |
| C.1 Figures C | 136 |
| C.2 Tables C | 140 |
| Appendix D: Appendix Chapter 5 | 149 |
| D.1 Notes D | 150 |
| D.2 Figures D | 151 |
| D.3 Tables D | 172 |
| References | 179 |

List of Tables

| | | |
|-----|---|-----|
| 2.1 | Analysis summary for the different methods and families. | 20 |
| 2.2 | Error analysis of different sap flow methods. | 23 |
| 2.3 | Least-squares means and 95% CI calculated from the LMM models testing the effect of different wood porosity types | 25 |
| 2.4 | Synthesis of the potential sources of error and use adequacy for each method. | 26 |
| 3.1 | Number of sap flow times series in SAPFLUXNET depending on whether they were calibrated for the different sap flow methods. | 45 |
| 3.2 | Number of plants in the SAPFLUXNET database using different radial and azimuthal integration. | 46 |
| 3.3 | Number of datasets, plants and species by stand-level treatment in the SAPFLUXNET database. | 54 |
| 4.1 | Analysis of variance testing differences among biomes in the coupling of tree-level water conductance to each of the main hydrometeorological drivers. | 70 |
| 4.2 | Parameters of the models explaining G_{Asw} coupling to VPD, SWC, PPF and to all three hydrometeorological drivers. | 70 |
| 5.1 | Description of variables and units in this study. | 82 |
| 5.2 | Results of the bi-variate linear models relating water use parameters and water relations traits | 88 |
| 5.3 | Results of the bi-variate linear models relating water use parameters, water relations traits, climate and tree height. | 90 |
| A.1 | Summary table of the studies used in the analyses. | 109 |
| A.2 | Anova summary of the LMM models. | 111 |
| A.3 | Summary of the LMM models of Ln-Ratio (accuracy), Slope (proportional bias), Slope (ln-ln) (linearity) and Z-Cor (precision) as a function of Methods and Calibration material. | 112 |
| B.1 | Data checks implemented in the first level of data quality control (QC1). | 119 |
| B.2 | Description of site metadata variables. | 120 |
| B.3 | Description of stand metadata variables. | 121 |
| B.4 | Description of species metadata variables. | 122 |
| B.5 | Description of plant metadata variables. | 122 |
| B.6 | Description of environmental metadata variables. | 123 |

| | | |
|------|--|-----|
| B.7 | Data checks implemented in the second level of data quality control (QC2). | 124 |
| B.8 | Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. | 125 |
| B.9 | Number of plants and number of datasets for each species present in the SAPFLUXNET database. | 130 |
| B.10 | Number of plants per genus present in the SAPFLUXNET database. | 133 |
| C.1 | SAPFLUXNET sites included in the study. | 140 |
| C.2 | SAPFLUXNET stand treatments included in the this study. | 143 |
| C.3 | Summary table of plot level. | 144 |
| C.4 | Table of equivalence between Whittaker biomes and the groups of biomes used in the study. | 148 |
| D.1 | SAPFLUXNET plot treatments included in this study. | 172 |
| D.2 | Species resume table. | 173 |
| D.3 | Results of the linear models relating water use parameters calculated using G'_{Asw} to water relations traits. | 175 |
| D.4 | Results of the linear models relating water use parameters to water relations traits using REW instead of SWC. | 176 |
| D.5 | Results of the linear models relating water use parameters calculated using G'_{Asw} , water relations traits, climate and tree height. | 177 |

List of Figures

| | | |
|-----|--|-----|
| 2.1 | Graphical representation of the calibration performance metrics. | 18 |
| 2.2 | Distribution of the calibration performance metrics for each method. | 21 |
| 2.3 | Predictions of the LMM models of the four calibration metrics. | 22 |
| 2.4 | Relationship between the four calibration performance and wood density, for different sap flow methods. | 24 |
| 2.5 | Relationship between measured and reference sap-flux density (SFD) for different sap flow methods, studies and calibrations. | 28 |
| 2.6 | Relationship between measured and reference sap flow (SF) for different sap flow methods, studies and calibrations. | 29 |
| 3.1 | Overview of the SAPFLUXNET data workflow. | 41 |
| 3.2 | Geographic, bioclimatic and vegetation type distribution of SAPFLUXNET datasets. . | 43 |
| 3.3 | Taxonomic distribution of genera and species in SAPFLUXNET. | 44 |
| 3.4 | Distribution of plants according to major taxonomic group and sap flow method. . | 47 |
| 3.5 | Characteristics of trees and stands in the SAPFLUXNET database. | 49 |
| 3.6 | Duration and number of plants mesured in each dataset. | 50 |
| 3.7 | Fingerprint plots showing hourly sap flow per unit sapwood area. | 52 |
| 3.8 | Summary of the availability of different environmental variables in SAPFLUXNET datasets. | 53 |
| 3.9 | Potential for upscaling species-specific plant sap flow to stand-level. | 55 |
| 4.1 | Bi-variate and uni-variate distributions of the coupling of G_{Asw} to the hydrometeorological drivers. | 71 |
| 4.2 | Global projection of G_{Asw} coupling to VPD, SWC and PPFD. | 72 |
| 4.3 | Relative importance (partial R^2) of the three hydrometeorological drivers of transpiration regulation. | 73 |
| 5.1 | Distribution of the plots from the SAPFLUXNET database included in this study. . | 82 |
| 5.2 | Boxplots of water use parameters for both Angiosperms and Gymnosperms. | 86 |
| 5.3 | Bi-variate relationships between G_{REF} water relations traits. | 87 |
| 5.4 | Bi-variate relationships between β_{VPD} water relations traits. | 89 |
| 5.5 | Bi-variate relationships between β_{SWC} water relations traits. | 91 |
| 5.6 | Path analyses of species-specific water use parameters. | 92 |
| A.1 | Representation of the cumulative number of studies using different sap flow methods between 1957 and 2017 | 106 |

| | | |
|-----|--|-----|
| A.2 | Relationship between root mean square error (RMSE) and sap flux density. | 107 |
| A.3 | Predictions of the LMM models calculated from least-squares means of the intercept (β_0) of the linear model. | 108 |
| B.1 | Overview of the data QC process. | 114 |
| B.2 | Structure of sfn_data objects. | 115 |
| B.3 | Example screenshot of the app used for handling outliers and out of range values in time series. | 116 |
| B.4 | Detailed geographic distribution of SAPFLUXNET datasets | 117 |
| C.1 | SAPFLUXNET global scaling relationship between basal area and sapwood area. Shaded areas are 95% model confidence interval. | 136 |
| C.2 | Bioclimatic distribution of the SAPFLUXNET datasets used in the study. | 137 |
| C.3 | Global projection of climatic, soil and stand structure variables. | 138 |
| C.4 | Log relationships of the three environmental variables estimated with the TOTAL model (VPD + SWC + PPFD) and grouped by biome. | 139 |
| D.1 | PCA plot of water relations traits using imputations. | 151 |
| D.2 | Water use parameters at the species level. | 153 |
| D.3 | Species G_{Asw} responses to VPD. Each species have different curves representing distinct levels of SWC (from dark to light, 0.1, 0.2, 0.3 [$\text{m}^3 \text{ m}^{-3}$]). | 160 |
| D.4 | Species G_{Asw} responses to SWC. Curves are fitted using a VPD reference level of 1 kPa. | 167 |
| D.5 | Scatterplot between G_{REF} and β_{VPD} parameters. | 168 |
| D.6 | Species' climatic variables and water relations traits relationships. | 169 |
| D.7 | Species' climatic variables and water use parameters relationships. | 170 |
| D.8 | Boxplots of water relations traits for angiosperms and gymnosperms. | 171 |

Abstract

Understanding how plants regulate transpiration is a central issue in ecophysiology since its origins. Plant transpiration links physiological responses of vegetation to water supply and demand with hydrological, energy and carbon budgets at the land-atmosphere interface. Although transpiration represents the main terrestrial water flux, its response to environmental drivers is currently poorly defined by observations globally.

In this thesis, I aim to give a global perspective on the ecohydrological variables driving the regulation of transpiration using sap flow data at the whole-tree level. To achieve this goal I have contributed to the development of the first global compilation of whole-plant transpiration data from sap flow measurements (SAPFLUXNET). In order to provide a harmonized sap flow database, compatibility between the different sap flow methods has to be ensured. To this end, the second chapter of this thesis deals with the uncertainty of different sap flow techniques by carrying out a meta-analysis of 290 individual calibration experiments gathered from the literature. Results suggest that Dissipation methods may be more appropriate to assess relative sap flow and Pulse methods may be more suitable to quantify absolute flows. All sap flow methods showed high precision, allowing potential correction of the measurements when a study-specific calibration is performed. In the third chapter, I present the SAPFLUXNET database, which contains 202 globally distributed datasets with sap flow time series for 2714 trees of 174 species. Datasets include sub-daily time series of sap flow and hydrometeorological drivers for one or more growing seasons, as well as metadata on the stand characteristics, plant attributes, and technical details of the measurements. In the fourth chapter, I carried out a quantification of the importance of hydroclimatic drivers controlling tree transpiration globally. I found that transpiration regulation dynamics are better explained by vapour pressure deficit (VPD) than by soil water content (SWC) or radiation in most areas. I also found that whole-tree canopy conductance (G) of trees in dryland biomes are less coupled to all three hydrometeorological drivers compared to those in other biomes. Climate, soil, and vegetation structure were common controls of all three hydrometeorological couplings with G , with wetter climates, fine-textured soils, and tall vegetation being associated with tighter coupling. Finally, in the fifth chapter, I characterized tree water use strategies across species emerging from the covariation between water use regulation and hydraulic traits, controlling also for the climatic effects produced by differences in precipitation. I found that reference canopy conductance and its sensitivity to VPD is coordinated with hydraulic and allocation traits (i.e. Ψ_{P50} , maximum sapwood hydraulic conductivity, Huber value, water potential at turgor loss point, root depth, leaf surface and tree height) rather than being directly controlled by mean annual precipitation. Species with efficient xylem transport (higher hydraulic conductivity) had higher canopy conductance but also higher sensitivity to VPD. Moreover, I found that angiosperms had higher reference canopy

conductance and higher sensitivity to VPD than gymnosperms. In conclusion, this approach allowed for a simplified global mapping of hydrometeorological drivers importance and species trait-based water use strategies. In addition, the use of simple measurable traits altogether with functional grouping can lead to a better approximation of species reference whole-tree water conductance and its sensitivity to VPD.

A reproducible copy of the thesis can be found online at
<https://github.com/vflo/PhD>

1

Introduction

1.1 Transpiration and earth system functioning

Transpiration is a dynamic process in which water from plants evaporates and diffuses into the atmosphere. Transpiration occurs as a result of the need to incorporate carbon dioxide for assimilation in photosynthesis. This is because the carbon dioxide molecule is larger than the water molecule, and when it enters the plant tissues, water escapes through the same pathway. Given that plants require proper hydration to maintain physiological processes, water loss by transpiration determines the need to absorb and transport large quantities of water from the soil (Prasad *et al.*, 2008). Yet, transpiration also contributes to regulate leaf temperature, keeping it within an tolerable range for leaf functioning. Thus, carbon uptake and transpiration water loss are inexorably linked and have to be dynamically controlled to avoid desiccation and maintain plant function. This control occurs on the long-term by changes in plant structure and anatomy, and in the short-term through the regulation of stomatal aperture in the leaves (Jones, 1997). Stomata are small epidermal valves with a pore aperture that is usually regulated by plant water status and hydraulic characteristics (Buckley, 2005, 2019). Stomata close to protect plants from excessive water loss when atmospheric humidity or water availability drops, but this closure increases physiological stress due to reduced carbon gain (Bréda *et al.*, 2006; McDowell *et al.*, 2008). Transpiration, which is therefore determined in just a few micrometres in the leaves, but is the outcome of the plant integrated hydraulics and chemical signalling upstream, has crucial implications for biological processes at the whole-plant level such as growth, competition and survival. These influences scale up to the population and species level and determine the water, carbon and energy cycles from the local to the global level (Bonan *et al.*, 2003), which are essential cycles for the maintenance of life on Earth. Therefore, due to the relevance of transpiration patterns on climate and Earth system function it is crucial to correctly understand and model stomatal regulation and vegetation water use.

Within the global hydrological cycle, vegetation transpiration accounts for about 45,000 $\text{km}^3 \text{ yr}^{-1}$ of water flowing from the soil to the atmosphere (Schlesinger & Jasechko, 2014), which represents around a 40% of global terrestrial precipitation (Oki & Kanae, 2006). Vegetation is thus the major user of the precipitation and directly affects surface runoff and subsurface recharge. Transpiration accounts for about 61% of total terrestrial surface evapotranspiration (which is the sum of transpiration and the abiotic evaporation from canopy and soil surfaces). However, this proportion is highly variable within and among regions (Schlesinger & Jasechko, 2014), which is largely explained by the different vegetation structures (e.g. leaf area index; Wei *et al.*, 2017) and water use strategies. Total land surface evapotranspiration and its partition between transpiration and evaporation drives other biosphere processes, such as vegetation dynamics or energy balance, that in turn produce feedbacks on evapotranspiration. For instance, transpiration is determinant of energy balance impacting in land surface temperature through changes in latent heat, and mediates land-atmosphere feedbacks via soil moisture depletion (Miralles *et al.*, 2019). It also promotes clouding formation (Aemisegger *et al.*, 2014) which alters regional albedo and influences large scale precipitation regimes (Spracklen *et al.*, 2012).

At the same time, climate affects vegetation physiology and how it regulates transpiration. Different tree species have a specific control of their stomata in response to environmental changes (Klein, 2014), which means that the distribution of these species and their physiological traits will

have consequences for ecosystem functioning and, ultimately, for global cycles. Environments with high regional water availability and resources generally support highly diverse and productive vegetation, whereas unfavourable environments support slow-growing vegetation and restrict species richness. Fast-growing plants tend to have higher transpiration rates (Smith & Sperry, 2014), and therefore accelerate the hydrological cycle. There is a large global heterogeneity both in space and time in water availability distribution, which drives heterogeneity in vegetation distribution and feedbacks on the spatio-temporal variability in transpiration patterns.

In addition to the water and energy cycles, transpiration is intimately linked to the carbon cycle through stomatal gas exchange. The carbon assimilated by plants in photosynthesis depends mainly on the internal leaves [CO₂], which enters the leaves by diffusion at a rate controlled by stomatal conductance (Ball *et al.*, 1987). At the plant level, carbon is used for growth and metabolism but drought-driven reductions in stomatal conductance limit the amount of carbon fixed by vegetation (Bréda *et al.*, 2006). Drought stress constrains ecosystem transpiration and photosynthesis, reducing the potential CO₂ uptake by terrestrial ecosystems (van der Molen *et al.*, 2011). Transpiration could also be affected by atmospheric [CO₂], because as atmospheric [CO₂] increases, the ratio between carbon intake and water loss increases so that plants would theoretically maintain similar photosynthetic rates using less water, leading to more efficient water use (Keenan *et al.*, 2013). However, CO₂ effects on transpiration seem to be complex and ‘water savings’ not as clear (Walker *et al.*, 2020). In addition, current [CO₂] are approaching saturation levels of the Rubisco enzyme, at least for the C₃ plants, which implies that future increases in atmospheric [CO₂] will have progressively less impact on water use efficiency, and therefore on CO₂ transpiration constrain (Sage, 2002; Flexas *et al.*, 2016).

Stomatal conductance (g_s) is therefore a central component of models characterizing land surface processes, since it is the main regulator of water, energy, carbon and nutrient fluxes between the terrestrial biosphere and atmosphere (Knauer *et al.*, 2015; Mencuccini *et al.*, 2019a). Thus, g_s is used in many ecosystem and terrestrial biosphere models at multiple spatial and temporal scales. For example, it is included in Land Surface Models to resolve the effects of vegetation (Matheny *et al.*, 2017) on global climate models (Wang & Dickinson, 2012; Fisher & Koven, 2020), or in Dynamic Global Vegetation Models to predict plant dynamics and vegetation distribution and changes under future climates (Smith *et al.*, 1992). Most ecosystem and terrestrial biosphere models are parametrized to separately estimate transpiration –calculated using diverse formulations of stomatal or canopy conductance (G) (Knauer *et al.*, 2015)–, soil evaporation and evaporation of water intercepted by the canopy or the litter layer. The mechanisms that regulate stomatal behaviour are still not completely understood, which is reflected in the different ways these processes are implemented in models. There are several approaches for modelling g_s or G , from semi-empirical (Damour *et al.*, 2010), mechanistic (Chuang *et al.*, 2006) or models employing optimality criteria (Wang *et al.*, 2020). Early models, represented vegetation g_s and G responses with a static set of parameters across plant functional classes or climates (Manzoni *et al.*, 2011). This oversimplification of diverse species into functional classes leads these models to great uncertainties (Poulter *et al.*, 2011). Therefore, new modelling approaches are moving towards a more versatile characterization of vegetation which could represent the stomatal responses to environment emerging from an integration of hydraulic traits (which are increasingly available at the species level; Mencuccini *et al.*, 2019b; Sanchez-Martinez *et al.*, 2020) at the whole-plant

level. There is an increasing recognition that water transport from roots to leaves play a key role defining plant water use strategy and stomatal conductance regulation.

1.2 Transpiration across scales

1.2.1 Water transport in plants

The water in soil and plants moves under high tensions following the framework of the cohesion-tension theory (Tyree, 1997). Water flow (J [$\text{cm}^3 \text{ s}^{-1}$]) between any two points of the soil-plant-atmosphere continuum is given by its difference in Ψ ($\Delta\Psi$) and the resistance –or its inverse, the hydraulic conductance (K_H [$\text{cm}^3 \text{ s}^{-1} \text{ MPa}^{-1}$])– that the system exerts to the flow between these two points. This relationship can be expressed as $J = K_H\Delta\Psi$ which is an expression of Darcy's law (Scheidegger, 1974). Within the plant, water moves along a gradient of water potential (Ψ ; normally negative in plant tissues) from high Ψ in the soil (i.e. less negative) to lower Ψ (i.e. more negative) in the leaves. Similarly, water diffuses out of the leaf and into the atmosphere following a gradient of water vapour concentration ($\Delta\Psi$, the difference in water mole fraction in the air, which could also be expressed as water potential, see e.g. Tyree & Ewers, 1991). Water potential –measured as water pressure in Pascals (Pa)– is the energy of water per unit volume compared to pure water and, along the soil-plant-atmosphere system, is the sum of hydrostatic potential, osmotic potential and gravitational potential. Hydrostatic potential is the physical pressure that the system exerts to the water and is positive inside turgid living cells but negative inside xylem conduits, a tissue conformed by dead cells and where water is under tension. The osmotic potential is related to a difference in solute concentration across a semi-permeable membrane, where the solution with more solutes has a lower Ψ . The gravitational potential arises because water ascent from soils to leaves must overcome gravity, but it is only important for tall trees (Franks, 2004).

Transport of water in unsaturated soil also follows mass conservation and Darcy's law. Water enters the roots mainly due to a gradient in hydrostatic pressure maintained largely by the transpiration stream (Passioura, 1988). As soils dry out, the Ψ of soils decreases because the matrix potential, i.e. the negative hydrostatic potential produced by the force by which water molecules are held on surfaces, becomes more negative as the water layer covering soil particles becomes thinner. This decrease in soil Ψ produces a parallel decrease in plant Ψ in order to maintain water movement into the plant. Water absorbed by roots reaches xylem vessels, which by the hydrostatic pressure gradient generated by water evaporation in the leaves transported downstream. Water transport in plant tissues is therefore controlled by the combination of water potentials and plant-specific hydraulic conductivity (k), which is a measure of conductance per unit path length. Xylem k is in turn dependent on water potential $k(\Psi)$, since cavitation and subsequent embolism formation occur at low Ψ , when the plant is under drought stress. Xylem k declines non-linearly with Ψ typically following a sigmoidal shape (vulnerability curve), which is characteristic of each species due to specific anatomical traits of the xylem (Venturas *et al.*, 2017). Once in the leaves, water from the mesophyll, epidermis and guard cells evaporate into the stomatal cavity. The difference in Ψ between the nearly water saturated stomatal cavity and the atmosphere represents the steepest potential gradient in all the soil-plant-atmosphere continuum. It is in the plant-atmosphere interface, therefore, where the control of water loss is critical to avoid

dangerous tensions in the plant tissues. Stomatal control and its link with plant hydraulics –but also anatomy, size, density and location of stomata– determine plant water status (McCulloh *et al.*, 2019). The mechanistic control of stomata in response to environmental drivers is still not fully understood, although it appears quite evident that there are feedback processes (and probably feedforward processes) between stomatal conductance and leaf water status (Jones, 1997; Franks, 2004), and hormonal or chemical signalling from the roots to the leaves (Tardieu *et al.*, 1993).

Quantification of transpiration and stomatal control by leaves and canopies also require a careful consideration of aerodynamic processes involved in the vapour transfer between the leaves and the atmosphere. Leaf boundary layer conductance also controls the vapour transport between the leaf and the surrounding air, as this conductance occurs in series with g_s . Leaf boundary layer conductance depends on atmospheric turbulent flux inside the canopy and it is generally much higher than g_s , especially for trees and forests under well-ventilated conditions. Under these conditions, canopies are considered well-coupled to the atmosphere and transpiration is largely controlled by stomata. However, for large-leaved species under poor ventilation boundary layer conductance can be similar to stomatal conductance and this can affect estimations of the degree of stomatal control based on transpiration measurements. When dealing with whole plants and canopies, not with individual leaves, the sum of all the plant stomatal conductances and the boundary layer conductance can be considered as the whole-plant canopy conductance (Jarvis & McNaughton, 1986).

1.2.2 Upscaling to the stand and ecosystem levels

The quantification of water transport in the soil-plant-atmosphere continuum from a micro-scale perspective (i.e. at the stomata-level) is a complex challenge due to the heterogeneity of the transporting medium and the different temporal scales of the water flux drivers (Katul *et al.*, 2007). Therefore, most studies on plant water transport have been undertaken at the organ (leaves) and the plant levels, observing emergent macroscopic responses of transpiration to environmental changes. Upscaling from the leaf to the ecosystem level requires consideration of processes that occur at the level of individual plants such as hydraulics. In addition, by using a whole-plant level perspective, the micro-environmental and physiological gradients within the canopy are integrated, so we can focus on compositional, size-related patterns that influence the upscaling to the ecosystem. However, there is further challenge when trying to upscale both in space and time, due to the heterogeneity of the distribution of processes and the non-linearities of whole-plant responses shaped by the link between physiology and ecosystem functioning (Jarvis, 1995). Thus, to scale up to ecosystem, regional or global level, we should make simplifications to allow a proper estimation of those processes distributions, either by a representation of the vegetation by functional types, or a description by hydraulic or functional traits. Both vegetation classes and traits are suitable for scaling up, but the latter is a more flexible approach since they are continuous variables, which overcome the rigidness of vegetation classes (i.e. functional type or species), and also allows to include intra-species environmental gradients and variability, and diversity within the ecosystem (Anderegg *et al.*, 2018).

1.3 Transpiration quantification

There are several methods to quantify evaporative fluxes from the leaf to the ecosystem level (Shuttleworth, 2007), which differ in their spatial and temporal domain and in their ability to separately measure transpiration and evapotranspiration. At the ecosystem level, continuous sub-daily estimations of evapotranspiration are usually performed using micrometeorological methods such as Bowen ratio energy balance (Bowen, 1926) and, mainly, Eddy covariance methods (Brutsaert, 1982), which are based on the measure of turbulence fluxes of latent heat and moisture (see Wang & Dickinson, 2012; Kool *et al.*, 2014; Stoy *et al.*, 2019 for further details). These methods have been standardized, and currently there is a global network of measuring stations (FLUXNET) that includes more than 900 sites worldwide (Pastorello, 2020). The FLUXNET network initiative provides a valuable tool to study global land hydrological fluxes. However, micrometeorological methods do not allow to easily and directly separate transpiration from evaporation fluxes, thus requiring partitioning isotopic methods (Kool *et al.*, 2014) or complex algorithms to isolate the contribution of transpiration (Nelson *et al.*, 2020). Since some seminal works such as Sellers (1985), several attempts have been made to estimate regional transpiration and evapotranspiration patterns using remote sensing (Miralles *et al.*, 2011, 2016). Although remote sensing is the best method for containing continuous estimates of transpiration at large spatial scales, these estimations (e.g., GLEAM, Martens *et al.*, 2017) are indirect and based on the combination of remotely-sensed variables and models of vegetation responses and surface energy balance.

Since the 1970s, several methods have been developed to independently measure plant level transpiration, however most of them are impractical in the field, either because of their low temporal resolution and low ecological representativeness such as leaf gas exchange methods (Evans & Santiago, 2014), or because of their limited use on a broad scale due to their elevated cost such as whole-tree gas exchange chambers (Corelli-Grappadelli & Magnanini, 1993) or lysimeters (Howell, 2005). Yet one method that can be applied continuously, unsupervised, for extended periods and at relatively low cost, is the measurement of sap flow rate (from now on Sap Flow Methods, see **Chapter 2 and 3** for details). Sap flow methods are thermal-based techniques that were first developed by Huber (1932), which have diversified into different methodologies since then (e.g. Čermák *et al.*, 1973; Swanson & Whitfield, 1981; Granier, 1985). Sap flow methods basically apply to woody plants and estimate transpiration by measuring the water flowing through stems (Schulze *et al.*, 1985). To do that, semi-invasive probes are installed in the trunk and heat is applied as a tracer of the sap flux. The quantification of the flux can be largely characterized using 4 major variants: heat balance, heat dissipation, heat pulse or heat field deformation methods, which follow different operation principles, probe configuration, and method specific flux quantification (see **Chapter 2**; Vandegehuchte & Steppe, 2013). Despite their advantages, all sap flow methods suffer from potential methodological issues, they assume that the sensors installation does not alter the sap flow, and they rely on a correct determination of sapwood cross-sectional area to scale punctual measurements to the whole-tree level (see **Chapter 2**; Vandegehuchte & Steppe, 2013). However, most of these issues can be addressed using specific calibration and applying suitable corrections (e.g. Clearwater *et al.*, 1999; Peters *et al.*, 2020). Also, sap flow methods error have been associated with wood properties but not consistent patterns have been found (Wullschleger *et al.*, 2011).

Despite the potential of sap flow measures, the lack of a global compilation of sap flow measurements has precluded their use to scale up plant water use strategies and generalize actual transpiration responses to environmental drivers, at the regional or global scale. Similarly, a global sap flow database would allow a much better characterization of species' water use strategies and the underlying functional traits. Seminal works of sap flow data synthesis have only been able to focus on answering partial questions, such as the characterization of transpiration responses along climatic gradients within a species (Poyatos *et al.*, 2007), or across a few species, typically addressing responses to single hydroclimatic factors (Oren *et al.*, 1999b), or the various studies focused on maximum water use (Manzoni *et al.*, 2013). However, increased data availability and the growing trend of data sharing for reuse, has paved the way to the creation of a global collaborative database of sap flow measurements (Poyatos *et al.*, 2016). A preliminary survey conducted in December 2015 showed that potential contributors could provide data sets for >160 species and >120 globally distributed sites (Poyatos *et al.*, 2016). Thus, in 2016 the design of the first global database of sap flow measurements (SAPFLUXNET) was initiated, with the intention of gathering, harmonizing and homogenizing the largest possible number of sap flow data sets at the whole-plant level together with hydrometeorological variables at the stand-level, and making them freely available to the scientific community.

1.4 Transpiration in a changing world

Forests are changing rapidly in response to climate change and all models predict that these changes will accelerate in the future (Anderegg *et al.*, 2020; McDowell *et al.*, 2020). Importantly an improved understanding of the regulation of tree water use is key to assess both species vulnerability and the functional changes expected under new climate regimes (Choat *et al.*, 2018; Brodribb *et al.*, 2020). The SAPFLUXNET database will allow us to study, for the first time, the regulation of whole-plant transpiration to environmental changing conditions from a global perspective. It will enable the observation of broad patterns of the water use strategies of woody plants and generate the knowledge to improve predictions of vegetation responses to future climate scenarios. A first approach to understand global dynamics of regulation of transpiration at the whole-plant level can be achieved by using semi-empirical models of sap flow or canopy conductance (G) responses to key environmental drivers, including atmospheric vapour pressure deficit, soil moisture and solar radiation. This approach would allow us to parameterize the plant's transpiration response, obtaining species-specific maximum values and sensitivities to hydroclimatic variables, and thus characterizing the corresponding water use strategies, their spatial distribution and their physiological and ecological determinants. In addition, this approach may help to study which hydrometeorological variables are the main drivers of the regulation of transpiration globally, as well as its bio-geographical patterns. In this way, modelling efforts, such as LSMs, could be optimized and focused on the responses to the environmental variables with greater predictive power.

It is expected that water transport through the soil-plant-atmosphere and its regulation require of a coordination between stomatal behaviour and hydraulic traits (Meinzer, 2002). This coordination is expected because vegetation has evolved to be adapted to the conditions of its typical climate (Sanchez-Martinez *et al.*, 2020), achieving a balance in its traits in such a way

that they optimize the transport of water and nutrients while maximizing CO₂ uptake and fitness (Manzoni *et al.*, 2013). This coordination of traits at the organ level would be a good basis to study the water use strategy that entire plant use to cope with drought, and would pave the way to trait-based models of vegetation water use at large spatial scales (Sperry *et al.*, 2019; Eller *et al.*, 2020). Thus, if there was a clear correlation between hydraulic traits (some of which are relatively easily to measure) and water use strategies, they could be used for parameterization of G on a continuous basis.

Therefore, a better understanding of transpiration and its regulation is key under the Earth's system uncertainty added by global change and the expected increase in drought conditions (Dai, 2013). It is expected that evaporation and precipitation will be globally intensified but unevenly distributed, which could trigger large forest mortality events, even in cold or wet places (Allen *et al.*, 2015). This motivates further work to improve the predictability of global transpiration estimates and vegetation drought responses, in order to obtain better predictions of vegetation dynamics and climate, so as to assist in decision-making for water resource management, climate change mitigation and forest management.

1.5 Research aims and outline

In this thesis I address the question of whether we can find global patterns in daily transpiration regulation at the plant level in response to hydrometeorological drivers using a global database of sap flow measurements. The ultimate goal is to characterize the global variation in plant water use strategies and to relate this variation to ecosystem eco-hydrological conditions and to species traits, emphasizing plant water relations and hydraulics (**Chapter 4 and 5**). Prior to this synthesis work, I first assessed the suitability of sap flow methods as reliable estimators of the variation in plant water use (**Chapter 2**). And, last, but not least, together with the rest of the members of the SAPFLUXNET core team, I worked on the compilation and harmonization of the datasets in SAPFLUXNET database, which is also presented as part of this thesis (**Chapter 3**). The specific objectives for each research chapter are listed below:

Chapter 2: To test whether there is systematic variability in sap flow measurements associated with the sap flow method employed. Here, (i) I compile all sap flow methodological calibrations obtained under laboratory or field conditions and perform a meta-analysis to obtain the mean systematic bias, the proportional bias, the linearity, and the precision of each method. In addition, I test whether (ii) the performance of each method is associated to wood anatomy or wood density.

Chapter 3: To introduce SAPFLUXNET and explain how the database was built from individual data sets contributed by the scientific community. I (i) explain the data structure design, the data harmonization and quality control process and the overall workflow, (ii) summarise the main hydrometeorological drivers and metadata documented in each dataset, and (iii) discuss the potential applications of the database, its limitations, and future perspectives.

Chapter 4: To test the absolute and relative importance of vapour pressure deficit, soil moisture and solar radiation as drivers of tree transpiration at the global scale. I quantify the predictive capacity of each hydrometeorological driver of plant-level canopy conductance (G) using

empirical models. I explain the differences in hydrometeorological coupling of G (i) across biomes and (ii) their biogeographical patterns as a function of climate, soil properties and vegetation structure.

Chapter 5: To explore water use strategies across tree species using a trait-based approach, by parameterizing the response of G to vapour pressure deficit and soil water content at the species level and for major taxonomic groups (angiosperms vs. gymnosperms). I aim to understand how water use strategies emerge from the covariation between traits, accounting for the influence of tree size and climate. I also characterize the relationships between transpiration regulation parameters and key hydraulic and allocation traits among species, controlling also for the effect produced by differences in precipitation.

2

A synthesis of bias and uncertainty in sap flow methods

Victor Flo, Jordi Martinez-Vilalta, Kathy Steppe, Bernhard Schuldt, Rafael Poyatos. (2019)
Agricultural and Forest Meteorology, 271: 362–374

Abstract

Sap flow measurements with thermometric methods are widely used to measure transpiration in plants. Different method families exist depending on how they apply heat and track sapwood temperature (heat pulse, heat dissipation, heat field deformation or heat balance). These methods have been calibrated for many species, but a global assessment of their uncertainty and reliability has not yet been conducted. Here we perform a meta-analysis of 290 individual calibration experiments assembled from the literature to assess calibration performance and how this varies across methods, experimental conditions and wood properties (density and porosity types). We used different metrics to characterize mean accuracy (closeness of the measurements to the true, reference value), proportional bias (resulting from an effect of measured flow on the magnitude of the error), linearity in the relationship between measurements and reference values, and precision (reproducibility and repeatability). We found a large intra- and inter-method variability in calibration performance, with a low proportion of this variability explained by species. Calibration performance was best when using stem segments. We did not find evidence of strong effects of wood density or porosity type in calibration performance. Dissipation methods showed lower accuracy and higher proportional bias than the other methods but they showed relatively high linearity and precision. Pulse methods also showed significant proportional bias, driven by their overestimation of low flows. These results suggest that Dissipation methods may be more appropriate to assess relative sap flow (e.g., treatment effects within a study) and Pulse methods may be more suitable to quantify absolute flows. Nevertheless, all sap flow methods showed high precision, allowing potential correction of the measurements when a study-specific calibration is performed. Our understanding of how sap flow methods perform across species would be greatly improved if experimental conditions and wood properties, including changes in wood moisture, were better reported.

2.1 Introduction

Quantifying transpiration of vegetation is of major importance for hydrological, ecological, and agricultural sciences, since it represents 60–80% of the water that returns from the land surface to the atmosphere (Jasechko *et al.*, 2013; Schlesinger & Jasechko, 2014; Wei *et al.*, 2017). The study of transpiration and its environmental sensitivity is essential to understand vegetation water cycling (Frank *et al.*, 2015; Novick *et al.*, 2016; Konings *et al.*, 2017) and to forecast changes in vegetation functioning and composition under climate change (Allen *et al.*, 2015). Addressing these questions requires non-destructive measurements of whole-plant transpiration at multiple timescales (Wullschleger *et al.*, 1998). Thermal methods of sap flow measurement show a number of advantages over other methods such as those based on isotopes tracing or leaf gas exchange (Smith, 1995), and have become the most widely used approach to estimate tree-level transpiration (Poyatos *et al.*, 2016) (Fig. A1). When compared against independent estimates of evapotranspiration components, sap flow methods have provided reasonable qualitative and quantitative results (Diawara *et al.*, 1991; Hogg *et al.*, 1997; but see Wilson *et al.*, 2001; Oishi *et al.*, 2008; Kool *et al.*, 2014; Schlesinger & Jasechko, 2014; Zhang *et al.*, 2014; Shimizu *et al.*, 2015). However, sap flow measurements may be subject to various potential sources of error. Some of these errors are related to scaling sap flow variability both within trees and from tree to stand level (Hatton *et al.*, 1995; Mitchell *et al.*, 2009; Hernandez-Santana *et al.*, 2015), while others are related to intrinsic limitations of the methods or to how these methods are applied (see Vandegehuchte & Steppe, 2013). Although these biases have been studied, they have not yet been quantified globally and there is no conclusive assessment of how they differ across methods or species characteristics, including wood properties (Poyatos *et al.*, 2016).

Sap flow methods (Vandegehuchte & Steppe, 2013) can measure sap flow rate (SF, g h^{-1} or equivalent units) or sap flux density (i.e., sap flow rate per unit sapwood area, SFD, $\text{cm}^3 \text{cm}^{-2} \text{h}^{-1}$ or equivalent units) in a plant's conductive tissue and can be classified in four families depending on how they heat the sapwood and how they measure sapwood temperatures: (1) the Dissipation family, including thermal dissipation (TD; Granier, 1985) and transient thermal dissipation (Do & Rocheteau, 2002a, TTD Do & Rocheteau (2002b)) methods, which measure the dissipation of heat from a heated probe inserted in the sapwood with reference to a reference, non-heated probe; (2) the Pulse family, including the compensation heat pulse (CHP; Swanson & Whitfield, 1981), heat ratio (HR; Burgess *et al.*, 2001), T-max (Cohen *et al.*, 1981), calibrated average gradient (CAG; Testi & Villalobos, 2009), sapflow+ (SF+; Vandegehuchte & Steppe, 2012a; Vandegehuchte *et al.*, 2012), single probe heat pulse (SPHP; López-Bernal *et al.*, 2017) and dual heat pulse methods (Dual; Pearsall *et al.*, 2014), which all apply heat in pulses and track sapwood temperature changes caused by thermal convection and conduction; (3) the Field family, including the heat field deformation (Nadezhina *et al.*, 1998; HFD; Nadezhina, 2018) and its derivatives, which measure the shape changes of a continuous heat field in the sapwood, using axial and tangential probes; and (4) the Balance family, represented by stem heat balance (Vieweg & Ziegler, 1960; SHB; Daum, 1967; Sakuratani, 1981) and trunk heat balance (Čermák *et al.*, 1973; THB; Čermák *et al.*, 2004) methods, which measure the energy balance across a heated wood section. This latter family is the only one directly measuring sap flow rate, while all the others measure sap flux density.

Methodological errors in sap flux density measurements may be caused by wounding following probe insertion into the sapwood (except for the miniaturized non-invasive ones; see Clearwater *et al.*, 2009; Hanssens *et al.*, 2013; Schreel & Steppe, 2018), biological variation in wood parameters and diverse raw data processing approaches (Vergeynst *et al.*, 2014; Oishi *et al.*, 2016; Peters *et al.*, 2018). Wounding affects heat and water transport and thus may disrupt sap flow measurements (Green & Clothier, 1988; Barrett *et al.*, 1995; Burgess *et al.*, 2001; Green *et al.*, 2003, 2009; Steppe *et al.*, 2015), especially during long-term installations (Wiedemann *et al.*, 2013; Marañón-Jiménez *et al.*, 2018). While wound corrections have been available for a long time for some Pulse family methods (Swanson & Whitfield, 1981; Green *et al.*, 2003), they have only become recently available for other methods such as TD (Wiedemann *et al.*, 2016). Sap flux density methods are also affected by changes in radial patterns, which are not constant over time, so these methods have to measure the entire sapwood depth by sufficiently large probes, or by individual measurement points at different depths (Hatton, 1990). Although some methods have a more solid theoretical background based on the physics of thermal transport (Pulse and Balance methods), all of them rely on a certain degree of empiricism, which may introduce errors caused by biological variability and/or variation in signal processing approaches: species-specific empirical calibrations in Dissipation methods (Fuchs *et al.*, 2017), zero-flow determination or baselining (Lu *et al.*, 2004; Peters *et al.*, 2018), and different parameterization of thermal sapwood properties in Pulse and Field methods (Chen *et al.*, 2012). These thermal sapwood parameters could change over time, introducing further errors in the measurements (e.g. changes in stem water content; Vergeynst *et al.*, 2014). Within the Balance family, those using external heating do not suffer from potential errors due to wounding, but they all require zero-flow determination (Smith & Allen, 1996). Balance methods have often been considered to better integrate spatial variability in sap flow (Čermák *et al.*, 2004), but whether they perform generally better than sap flux density methods remains unknown.

Other errors in sap flow measurement may result from not accounting properly for method-specific assumptions. For many sap flow methods, natural temperature gradients (NTG) need to be minimized and/or accounted for to obtain unbiased estimates of sap flow (Reyes-Acosta *et al.*, 2012; Vandegehuchte *et al.*, 2015). Incorrect sensor geometry (misalignment) affects the accuracy of the measurements (Swanson & Whitfield, 1981; Swanson, 1983; Cabibel *et al.*, 1991; Burgess *et al.*, 2001; Ren *et al.*, 2017). Other application errors, such as those arising from the incomplete contact of TD probes with the sapwood (Clearwater *et al.*, 1999) are difficult to prevent, though they can be reasonably corrected *a posteriori* (e.g. Clearwater correction; Hultine *et al.*, 2010; Paudel *et al.*, 2013). Despite that these application errors have been well described in individual studies, a general quantification of these errors for the most employed sap flow methods is currently lacking.

Comparisons of sap flow measurements with respect to a reference method (hereafter, for simplicity, ‘sap flow calibrations’) are usually aimed at obtaining species-specific calibrations (Vandegehuchte & Steppe, 2013) to assess different parameterizations of wood thermal properties (Vandegehuchte & Steppe, 2012b) or to validate empirical corrections (e.g., wounding, NTG, changes in water content, misalignment; Burgess *et al.*, 2001; Vergeynst *et al.*, 2014). Although few studies calibrate multiple sap flow methods for different species (Fuchs *et al.*, 2017), collectively these calibration studies have shown the inherent limitations of different sap flow methods to deal with low (Green *et al.*, 2003) or high flows (Green *et al.*, 2009). Variability and quality in calibration

performance may also be related to specific wood properties such as wood density (Suleiman *et al.*, 1999; Wullschleger *et al.*, 2011), especially given the fact that wood density enters the calculation of sap flux density for some methods (Vandegehuchte & Steppe, 2012b) and co-varies with wood moisture content (Looker *et al.*, 2016). Because thermal properties of wood are dependent on both, i.e. wood density and moisture content (MacLean, 1941), they might additionally be influenced by wood anatomical traits such as wood porosity type, i.e. coniferous, diffuse-porous or ring-porous wood. In conifers, however, no clear effects of wood density on calibration variability have been reported (Peters *et al.*, 2018). Therefore, a quantitative synthesis of sap flow calibrations, accounting for variation caused by different flow ranges and wood properties is needed to generalize and understand the patterns observed in individual calibration studies.

Here, we compile a global database of published sap flow calibrations to quantify the measurement errors associated with different sap flow methods and to assess the factors underlying variability across methods. In assessing calibrations, we distinguished between mean systematic bias (accuracy), a measure of the average degree of closeness of the measurements to the value obtained with a reference method; proportional bias, which occurs when the magnitude of the error is a function of the flow; linearity in the relationship between measurements and values obtained with a reference method; and precision, a measure of reproducibility and repeatability. Our main objective is to assess the differences in accuracy, proportional bias, linearity and precision among methodological families and individual sap flow methods; in addition, we will determine whether calibration performance across methods is associated with species wood traits (wood density and porosity type).

2.2 Material and methods

2.2.1 Sap flow calibration datasets

We retrieved sap flow calibration studies of the seven most common methods (CHP, T-max, HR, HFD, SHB, TD, TTD) applied on trees, palms or lianas, using standard database searching tools (i.e. Scopus, Web of Science and Google Scholar). The search was conducted in June 2017 applying the following keywords: sap fl*, sap flux density, calibration, potomet*, gravimet*, thermal dissipation, heat pulse, heat balance, heat field deformation, compensation heat pulse, T-max, and their combinations. Other sources of data were obtained from the references of previously collected studies. For each calibration experiment, we obtained paired observations of sap flow, measured with a thermal method and with an independent reference method (typically gravimetric or volumetric). Data was digitized from published figures (using GetData Graph Digitalizer version 2.26.0.20). We asked the authors to supply the raw data when these were unavailable from the original publication. We obtained data from 60 studies (see Table A1) reporting 374 individual calibration experiments performed on 81 different shrub and trees species (10,186 data points in total). In the analysis, we only used calibrations that were properly applied according to our definition below (i.e. 290 calibrations out of 374) to restrict the variability to the intrinsic characteristics of the methods.

We always considered sap flow observations obtained with the original parameters of the methods (e.g. Granier's original calibration for TD), without applying the coefficients derived

from the calibrations themselves. Some calibrations with TD gave measured K values (i.e. sap flow index, calculated from raw sapwood temperature differences) instead of measured SFD and, in these cases, K values were transformed to SFD using Granier's original equation and calibration coefficients (Eq. 2.1, $a = 42.84 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$, $b = 1.231$) (Granier, 1985).

$$SFD = a \times K^b \quad (2.1)$$

For each calibration, we recorded the type of calibration material: whole plants, whole plants without roots or cut stem segments. We also assessed whether the sap flow method was properly applied using the best available protocol specified for each method. We considered a proper application of Dissipation methods when the probe was shorter than the sapwood depth and radial profile correction was applied, when the probe was approximately equal to the sapwood depth, or when the probe was longer than the sapwood depth and this effect was corrected for following Clearwater *et al.* (1999). To test whether our results could have been affected by this correction, we performed a preliminary analysis with the same structure as the main statistical model (cf. section 2.3.3) comparing TD calibrations with or without the Clearwater correction and we did not find significant effects on any of the metrics of calibration performance (cf. section 2.3.2). We considered a proper application of Pulse methods when wound correction was applied and either the probe had multiple measuring points along the sapwood or a radial sap flow profile correction was applied. We always considered Balance methods and Field methods as properly applied, because they always integrate (or account for) spatial variability of sap flow.

Finally, to analyze the influence of wood traits on calibration performance we used wood density, defined as fresh volume over oven-dry mass, and wood porosity type of the species employed in each study. Wood density was supplied in only a few studies (7 species \sim 80 calibration experiments \sim 4 studies). Assuming that for wood density between-species variability is typically larger than within-species variability (Siefert *et al.*, 2015; Vilà-Cabrera *et al.*, 2015), we retrieved wood density of each species from the TRY database (Kattge *et al.*, 2011). Wood densities of *Carica papaya*, *Phoenix dactylifera* and *Vitis vinifera* were obtained from Kempe (2014), Fathi (2014) and Castelan-Estrada (2002), respectively, as they were not recorded in TRY. When wood density could not be found for a given species, we used the phylogenetically nearest species of the same genus if available in TRY (10 of 81 species; e.g. *Citrus sinensis* for *Citrus reticulata*). We could not estimate wood density for three taxa (*Humulus lupulus*, *Musa* sp. and *Siagrus romanzoffiana*). A correlation between calibration-specific and species-level wood density extracted from the TRY database ($r = 0.78$, $P < 0.01$, $n = 12$ calibrations, 6 species) indicates that species-level wood density values indeed are applicable for our purpose, but the results should be interpreted with caution. Finally, wood porosity was obtained from the InsideWood database (Wheeler, 2011), using four categories: ring-porous, diffuse-porous (i.e. diffuse and semi-diffuse porous), conifer and monocots.

2.2.2 Calibration assessment

Although the reference methods always provide an estimate of sap flow through plants or stem segments, sap flow measurements can be reported as SF or SFD. It was not possible for us to interconvert between SF and SFD in all cases because sapwood areas were not always reported.

This precluded a joint analysis of all the paired observations in the same linear model because units differ between SF and SFD. To overcome this problem and to maximize the amount of data considered in the analyses, we first evaluated calibration performance using four complementary dimensionless metrics at the calibration level, which allowed us to analyze globally all calibrations regardless of the magnitude they reported (SF or SFD). In a second stage, we quantified the variability in the absolute errors in sap flow measurements across methods and flow ranges separately for SF and SFD methods. We did not expect differences between calibrations reported with SF or SFD because the inter-conversion between them only involves a scalar transformation. In addition, preliminary analyses confirmed that there was no significant difference between SF and SFD for any of the calibration performance metrics reported in this study (Table A2).

For the global analysis, the following metrics were calculated for each calibration (SF and SFD): the average ln ratio (Ln-Ratio) between measured and reference values as a measure of overall accuracy; the slope of the relationship between measured and reference sap flow to characterize proportional bias (Slope); the slope of the ln-ln relationship between measured and reference sap flow as a measure of linearity (Slope (ln-ln)); and Z Pearson's Correlation to describe precision (Z-Cor) (Fig. 2.1). To calculate these metrics, we filtered out data points with measured or reference flows less or equal to 0. All calibration metrics and subsequent statistical models were performed in R 3.4.2 (R Core Team, 2017). For model-based metrics, we always checked residuals to ensure they satisfied normality and homoscedasticity assumptions. Accuracy was evaluated as the mean of the natural logarithm of the ratio between paired measurements (j) of each calibration (i):

$$Ln - Ratio_i = \frac{\sum_{j=1}^n \ln(\frac{measured_j}{reference_j})_i}{n_i} \quad (2.2)$$

where $measured_j$ and $reference_j$ are the paired measurements of sensor-estimated and reference flow, respectively, and n the number of paired measurements for each calibration i (see Fig. 2.1). The Ln-Ratio varies between $-\infty$ and $+\infty$, and equals 0 for a calibration with perfect mean accuracy (i.e. lack of systematic bias). We also expressed this metric as the exponential of Ln-Ratio minus one multiplied by 100, as an indicator of accuracy deviation (in %).

The slope of the linear relationship (Eq. 2.3) describes how the magnitude of the error changes (linearly) as a function of the reference flow. The slope of the ln-ln relationship (Eq. 2.4) captures the linearity between the measured and the reference flow. Both slope estimates were calculated for each calibration using a simple linear regression (lm - package stats):

$$measured_{ij} \sim \beta_{0i} + \beta_{1i} reference_{ij} + e_{ij} \quad (2.3)$$

$$\ln(measured_{ij}) \sim \beta'_{0i} + \beta'_{1i} \ln(reference_{ij}) + e_{ij} \quad (2.4)$$

where β_{0i} and β'_{0i} are the intercepts and β_{1i} and β'_{1i} are the slopes for each calibration (i), and j indicates individual calibration points. Hereafter, we will refer to β_{1i} as *Slope* and to β'_{1i} as *Slope(ln - ln)*; slope values equal to 1 characterize measurements without proportional bias (Eq. 2.3) and with high linearity (Eq. 2.4), respectively.

We used Pearson's correlation coefficients r between measured and reference flow of each calibration experiment (i) as a metric to describe the precision of the methods. The distribution

of the resulting variable was skewed due to the large amount of correlation coefficients close to 1, so we used Fisher's Z transformation (Eq. 2.5) to achieve normality:

$$Z - Cor_i = \frac{1}{2} \ln\left(\frac{1+r_i}{1-r_i}\right) \quad (2.5)$$

Low values of $Z - Cor$ correspond to low r correlations, and high values of $Z - Cor$ correspond to high r correlations and thus high precision (data set range $r = [0.0491 - 0.9999]$; $r = 0.0491 \sim z = 0.0491$; $r = 0.9999 \sim z = 5.1594$).

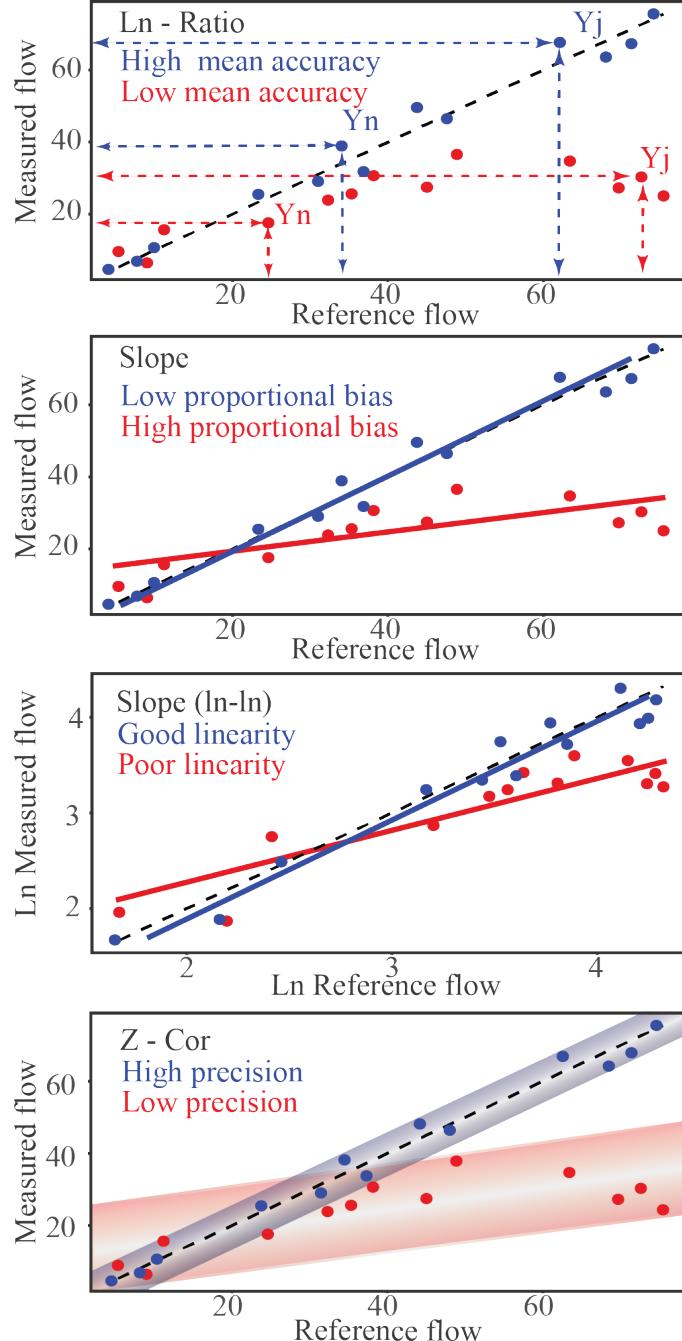


Figure 2.1: Graphical representation of the calibration performance metrics used in the analyses. Each panel presents the same simulated calibration points, representing plausible data. Blue dots represent an accurate, unbiased, linear and precise calibration, while red dots represent an inaccurate, biased, non-linear and imprecise calibration.

In the analysis of the absolute errors of sap flow measurements, we calculated the Normalized Root Mean Square Error (NRMSE) for each calibration (i) (Eq. 2.6), separately for SFD and SF methods, in order to obtain the percentage of absolute error at the mean range of each calibration (i).

$$NRMSE_i = \frac{(\sqrt{\frac{\sum_{j=1}^n (measured_j - reference_j)^2}{n}})_i \times 100}{range\ mean_i} \quad (2.6)$$

Subsequently, from the NRMSE and the mean range of each calibration, we fitted a linear model for each method allowing to quantify the absolute error (RMSE) at a given sap flow and also to obtain a RMSE at a reference flow (cf. section 2.3).

2.2.3 Statistical analyses

All the analyses were performed using linear mixed-effects models (LMM) with the package lmer (Bates *et al.*, 2015). Least-square means were estimated with package lsmeans (Lenth, 2016) and used to summarise the effects of fixed factors and to test contrasts among predictions. In all models, we used the variables Study and Species as partially crossed random effects (Schielzeth & Nakagawa, 2013), as we are interested in taking into account the variability associated with study and species, and also to analyze within- and between-group variability. We used Study as we expect experimental variability between researchers or laboratories, and Species because calibration performance has been reported to vary across species (Smith & Allen, 1996; Steppe *et al.*, 2015; Fuchs *et al.*, 2017). For each model, R_m^2 and R_c^2 (marginal and conditional coefficients of determination, respectively) based on Nakagawa & Schielzeth (2013) were calculated using the function r.squaredGLMM of the package MuMIn (Bartoń, 2017) in R. Intraclass Correlation Coefficients (ICC) were also calculated for the random factors to quantify the proportion of variance within and among groups (low ICC implies high intra-group variability).

In a first analysis, we were interested in assessing the differences in calibration metrics (Ln-Ratio, Slope, Slope (ln-ln), Z-Cor) between different families of methods (Family: Pulse, Dissipation, Balance and Field methods), because methods within a family share similar physical principles. We also analyzed differences between individual methods with a sufficient sample size (Method: CHP, T-max, HR, HFD, SHB, TD, TTD). As the calibration material determines, to a large extent, the experimental conditions, we also included this variable in our models (Material: whole plants, whole plants without roots or cut stem segments). For the analysis of absolute errors of sap flow measurements, we modelled NRMSE as a function of Method and the Mean Range of SFD (or SF for Balance methods) in each calibration, as well as their interaction. We used the same random structure as in previous models.

Finally, we assessed how each calibration metric depended on Wood Density and Wood Porosity. A first model included all methods available, with Method interacting with Wood Density as predictors. In order to test Wood Porosity effects, we fitted separate models for CHP and TD calibrations, as these two methods were the only ones that had enough data (> 5 calibrations) for more than one type of porosity. Separate models were needed because not all wood porosity types were represented for all methods. In both models we also included Material as an explanatory cofactor, and the same random structure as in the first analysis explained above.

Table 2.1: Analysis summary for the different methods and families of methods obtained from the LMM models (least-squares means). We provide the four dimensionless metrics: the Ln-Ratio as a measure of accuracy, the accuracy deviation calculated as the exponential of the Ln-Ratio minus one multiplied by 100, the Slope to characterize the proportional bias, the slope of the ln-ln-relationship, Slope (ln-ln), as a measure of linearity, and Z Pearson's correlation (Z-Cor) to describe overall precision; n: number of calibrations; studies: number of studies of each method; species: number of different species; r is the correlation calculated as the tanh of Z-Cor.

| Method | Family | n | studies | species | Ln-Ratio | Accuracy deviation % | Slope | Slope (ln-ln) | Z-Cor | r |
|--------|-------------|-----|---------|---------|----------|----------------------|-------|---------------|-------|-------|
| CHP | Pulse | 63 | 16 | 21 | 0.133 | 14.225 | 0.887 | 0.783 | 1.837 | 0.950 |
| T-max | Pulse | 11 | 5 | 6 | -0.053 | -5.162 | 0.614 | 0.697 | 1.755 | 0.942 |
| HR | Pulse | 23 | 6 | 7 | -0.145 | -13.498 | 0.845 | 0.841 | 2.000 | 0.964 |
| HFD | Field | 57 | 3 | 4 | -0.073 | -7.040 | 0.901 | 0.782 | 2.378 | 0.983 |
| SHB | Balance | 8 | 5 | 6 | -0.242 | -21.494 | 0.847 | 0.967 | 2.287 | 0.980 |
| TD | Dissipation | 115 | 18 | 35 | -0.519 | -40.488 | 0.683 | 1.066 | 1.711 | 0.937 |
| TTD | Dissipation | 14 | 2 | 6 | -0.493 | -38.921 | 0.669 | 0.985 | 1.464 | 0.899 |
| all | Pulse | 97 | NA | 30 | 0.012 | 1.167 | 0.844 | 0.787 | 1.874 | 0.954 |
| all | Field | 57 | NA | 4 | -0.008 | -0.820 | 0.896 | 0.762 | 2.322 | 0.981 |
| all | Balance | 8 | NA | 6 | -0.244 | -21.650 | 0.854 | 0.972 | 2.294 | 0.980 |
| all | Dissipation | 129 | NA | 37 | -0.464 | -37.153 | 0.681 | 1.052 | 1.666 | 0.931 |

2.3 Results

Most of the published calibrations were performed with Pulse and Dissipation methods (Table 2.1). In particular, 61% of the total number of the properly applied calibrations were conducted using TD or CHP, followed by HFD and HR. SHB, T-max and TTD methods were less represented, with 8 – 14 calibrations each. The metrics extracted from the raw calibrations were highly variable within methods (Fig. 2.2). Calibration metrics often followed a quasi-normal distribution, but in most cases distributions were truncated or skewed, particularly for methods with fewer calibrations (Fig. 2.2).

2.3.1 Calibration performance compared among methods and families of methods

The average accuracy deviation across sap flow methods (properly applied) ranged between 14.2% for CHP and -40.5% for TD (Table 2.1). There were significant differences in accuracy (Ln-Ratio) among families of methods and for methods but not for calibration materials (Fig. 2.3). The Dissipation family in general and the TD and TTD methods in particular were the only cases for which the Ln-Ratio was significantly different from 0 ($p < 0.001$) (Fig. 2.3), indicating systematic bias (underestimation).

Proportional bias, estimated by Slope, varied among methods and families of methods ($p < 0.01$). Among families, Dissipation methods showed a significantly smaller Slope than Pulse and Field methods (Fig. 2.3(a)), which was largely driven by the low value of TD (Fig. 2.3(b)). Also, both Pulse and Dissipation families had slopes significantly different from 1 ($p < 0.01$ and $p < 0.001$, respectively), but only the slope of the TD method was significantly lower than 1 ($p < 0.001$) (Fig. 2.3). As for the effects of calibration material, calibrations made with whole plants had a significant proportional bias (Slope < 1 , $p < 0.001$) (Fig. 2.3). Calibration linearity, as denoted by Slope (ln-ln), varied across methods and families of methods ($p < 0.001$). We observed higher

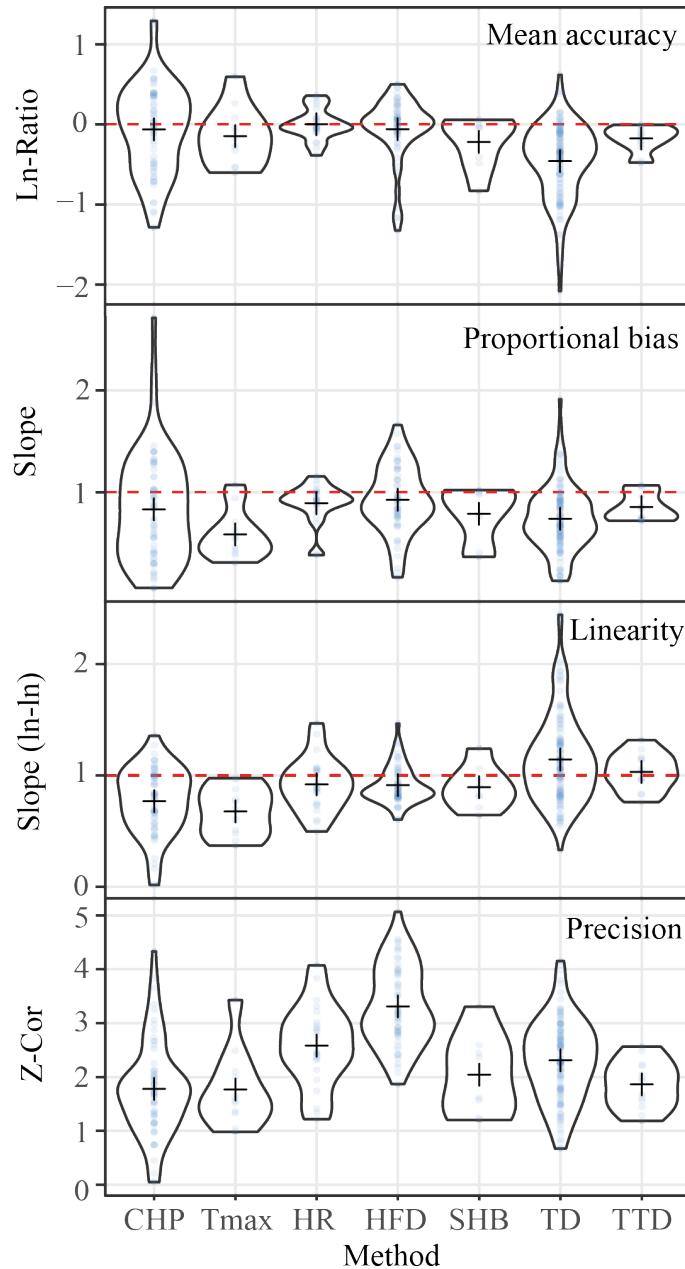


Figure 2.2: Distribution of the calibration performance metrics for each method. Dots represent the value of each individual calibration metric. Crosses represent the average of the metric for each method. Horizontal, dashed lines specify reference, perfect calibration values for a given metric.

values of Slope (ln-ln) for the TD method compared to CHP, T-max, HR and HFD. Consistently, the Dissipation family in general also had a higher Slope (ln-ln) than the Pulse and Field families (Fig. 2.3). CHP, T-max and HFD (and Pulse and Field methods in general) had a Slope (ln-ln) significantly lower than 1 (Table 2.1 and Fig. 2.3(b)), indicating a convex relationship between reference and measured flow. Calibrations performed with whole plants suffered from lack of linearity, indicated by Slope (ln-ln) significantly lower than 1 (Fig. 2.3).

Precision (Z-Cor) was explained by both method and calibration material. The HFD method (and Field methods in general) provided significantly higher precision than either Pulse or Dissipation methods (particularly CHP, TD and TTD) (Table 2.1 and Fig. 2.3(b)). Calibrations performed on stem segments provided higher precision than those conducted on whole plants

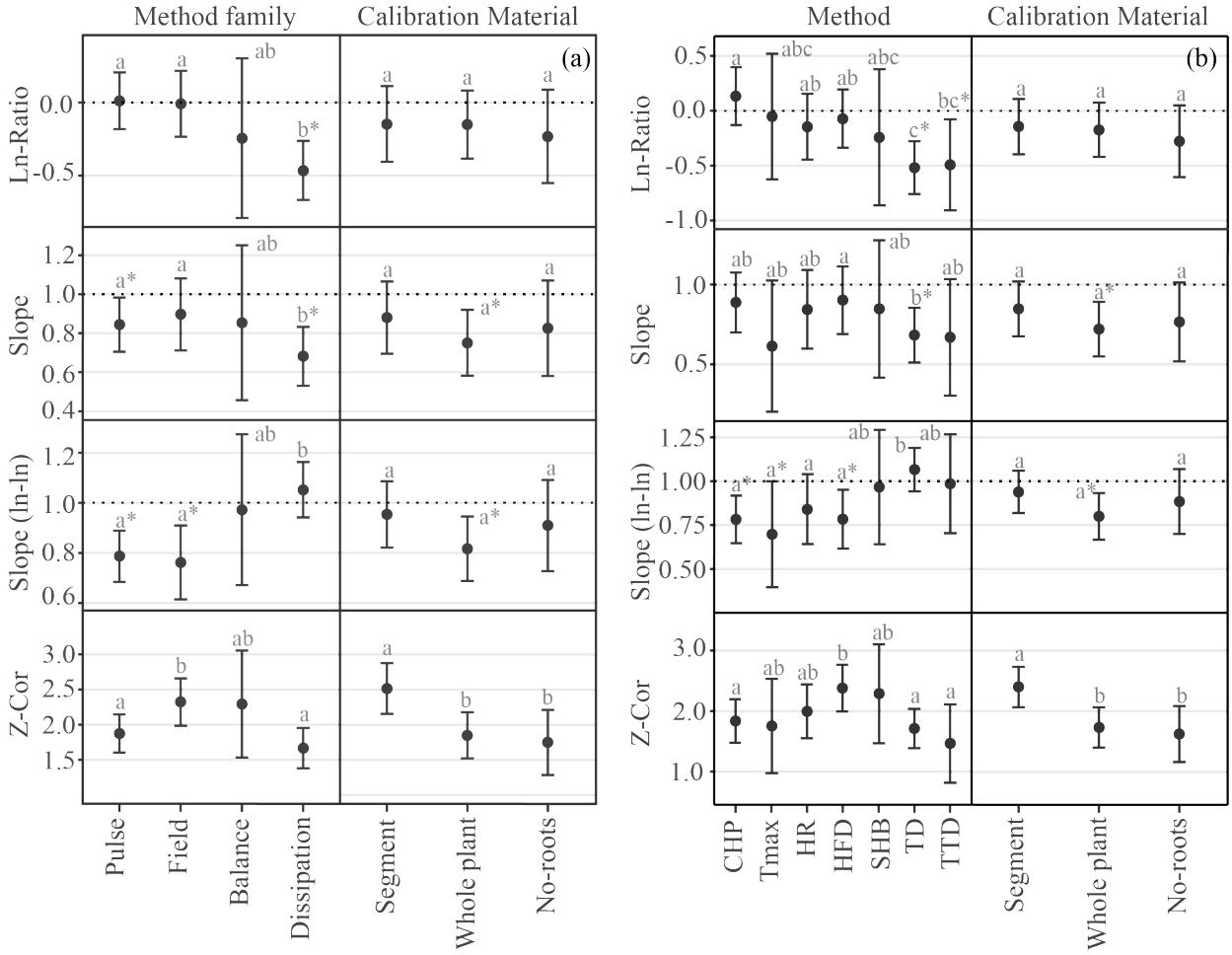


Figure 2.3: Predictions of the LMM models calculated from least-squares means of the four calibration metrics (Ln-Ratio as a proxy for mean accuracy, Slope for proportional bias, Slope (ln-ln) for linearity and Z-Cor for precision) for (a) different families of sap flow methods or for (b) different sap flow methods and for different calibration materials (Segment: stem segment; Whole plant: whole plant on a container or lysimeter; No-roots: whole plant without roots). 95% confidence intervals of the estimates are also shown. Different letters indicate significant differences between factors levels evaluated with Tukey's test. Horizontal, dotted lines indicate reference, perfect calibration values for a given metric. Asterisks (*) indicate significant departure from those reference values.

Table 2.2: Error analysis of different sap flow methods. The normalized root mean square error (NRMSE) is modelled as a function of method and the mean flow range for each calibration (and their interaction) using a LMM model with the same random structure as the main models (cf. section 2.3). β_0 and β_1 are the corresponding intercepts and slopes, respectively (β_0 expressed as % NRMSE; β_1 expressed as % NRMSE per change in $\text{cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$ for SFD or as % NRMSE per change in $\text{cm}^3 \text{ h}^{-1}$ for SF). This linear model, was also used to calculate a reference NRMSE at a sap flux equivalent to the percentile 50 of the range of the data in the calibrations (SFD: $25 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$; SF: $1300 \text{ cm}^3 \text{ h}^{-1}$). The expected NRMSE and RMSE (in brackets, in $\text{cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$, except for SHB that is in $\text{cm}^3 \text{ h}^{-1}$) at a typical flow are also given.

| Method | NRMSE | | | |
|--------|-----------|----------|-----------|----------------------------|
| | β_0 | % | β_1 | reference NRMSE (and RMSE) |
| CHP | 27.03*** | -0.08*** | | 25.04% (6.26) |
| T-max | 31.56. | 0.25*** | | 37.83% (9.46) |
| HR | 9.38 | 0.81 | | 29.59% (7.40) |
| HFD | 30.33*** | -0.12*** | | 27.45% (6.86) |
| SHB | 14.85 | 0.02*** | | 42.95% (558.36) |
| TD | 34.93*** | 0.10*** | | 37.31% (9.33) |
| TTD | 44.04*** | -0.04*** | | 42.94% (10.73) |

Statistical significant levels: " ." p<0.1 ; "*" p<0.05; "***" p<0.01; "****" p<0.001.

(with or without roots) (Fig. 2.3).

In all the previous models, little variability was explained by species (τ_{00} , species), relative to the higher variability associated to Study, particularly for the Ln-Ratio and Z-Cor models (τ_{00} , study, Table A3). This is consistent with the low ICC values observed for the species factor, indicating that there is more variability within than among species (Table A3).

In addition, the analysis of the normalized absolute error for the different methods showed that NRMSE decreased linearly with increasing measured sap flow in CHP, HFD and TTD methods and increased for T-max, SHB and TD (Table 2.2 and Fig. A2). For HR the increase in NRMSE with measured sap flow was not significant. For all the methods that measure SFD, the absolute error at a typical flow of $25 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$ ranged between $6.3 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$ for CHP and $10.7 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$ for the TTD method (Table 2.2).

2.3.2 Influence of wood traits

We did not find any significant influence of wood density on accuracy and linearity metrics (Fig. 2.4). Nonetheless, we observed a negative effect of wood density on proportional bias of HFD and TD calibrations ($p < 0.05$ and $p < 0.1$, respectively). In addition, a significant positive effect of wood density on the precision of HFD measurements was observed ($p < 0.001$), indicating that the higher the wood density, the higher the precision (Fig. 2.4).

We did not find any significant difference among wood porosity types in calibration metrics for studies using the TD or CHP methods. Nevertheless, the non-linearity (Slope (ln-ln) < 1) observed in general for the CHP method (Fig. 2.3(b)) was only significant for species with diffuse-porous wood, not for conifer species (Table 2.3).

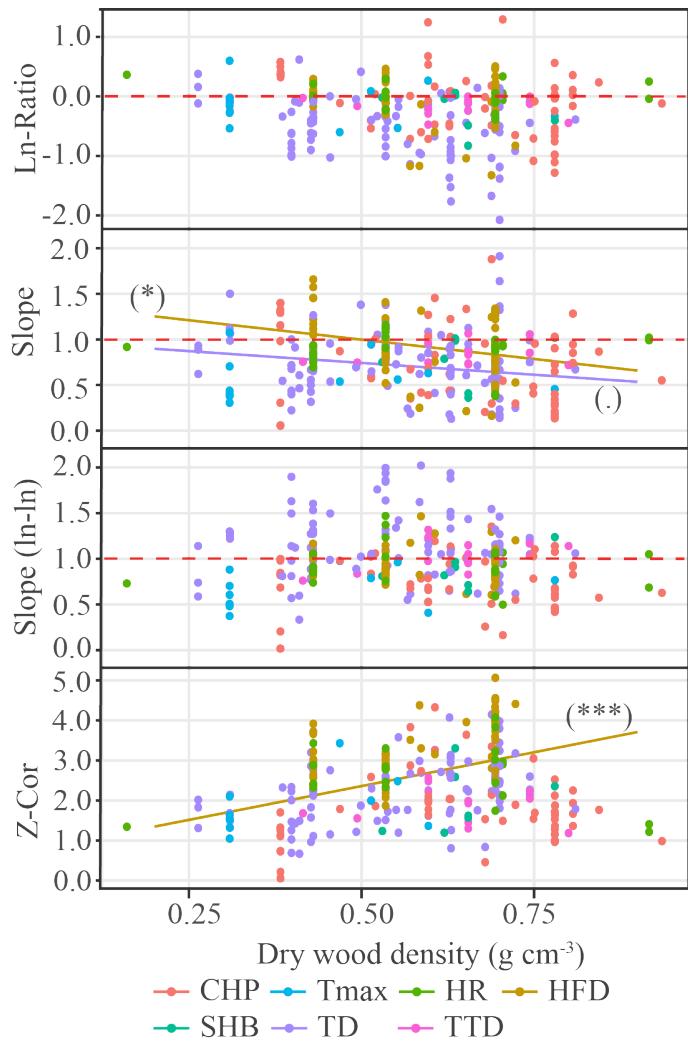


Figure 2.4: Relationship between the four calibration performance metrics (Ln-Ratio as a proxy for accuracy, Slope for proportional bias, Slope (ln-ln) for linearity, and Z-Cor for precision) and wood density, for different sap flow methods. Horizontal, dashed red lines indicate reference, perfect calibration values for a given metric. Regression lines are shown for significant effects only, and the corresponding level of significance (p -value: <0.1 : (.), <0.05 : (*), <0.01 : (**), <0.001 : (***)) is also reported

2.4 Discussion

Our results show a large variability in the quality of sap flow calibrations, even within the same sap flow method (Fig. 2.2), highlighting the large variability among and even within studies. This implies that, even if methods are properly applied (as defined in section 2.1), sap flow measurements can still produce biased estimates of water transport rates in plants, and these errors will need to be considered in quantitative analyses based on this type of measurements. On average, however, all sap flow methods assessed here produced results that may be acceptable for qualitative use in most applications, as shown by the typical high correlation between measured and reference values ($r > 0.89$ for all methods and method families, Table 2.1). For quantitative use, no method appears to be suitable for all experimental contexts, and researchers need to consider both the inherent limitations of the methods and the need to perform study-specific calibrations (see Implications and recommendations, Table 2.4).

Table 2.3: Least-squares means and 95% CI calculated from the LMM models testing the effect of different wood porosity types (Wood porosity) on sap flow calibration performance metrics (Ln-Ratio as a proxy for accuracy, Slope for proportional bias, Slope (ln-ln) for linearity, and Z-Cor for precision) for CHP and TD methods. No differences were detected among wood anatomies. Significance levels indicate departure from an ideal calibration (Ln-Ratio = 0; Slope = 1; Slope (ln-ln) = 1)

| Method | Wood porosity | n | Accuracy | | Proportional bias | | Linearity | | Precision | |
|--------|---------------|----|-------------------------|-----------------------|-----------------------|-----------------------|-----------|--|-----------|--|
| | | | Ln-Ratio | Slope | Slope (ln-ln) | Z-Cor | | | | |
| CHP | diffuse | 48 | -0.055 [-0.370 , 0.259] | 0.795 [0.557 , 1.033] | 0.799 [0.671, 0.927] | 1.980 [1.729 , 2.231] | | | | |
| CHP | conifers | 15 | 0.066 [-0.603 , 0.734] | 1.043 [0.459 , 1.627] | 0.777 [0.469 , 1.085] | 1.381 [0.775 , 1.988] | | | | |
| TD | diffuse | 81 | -0.273 [-0.658 , 0.111] | 0.752 [0.491 , 1.014] | 1.126 [0.917 , 1.336] | 1.672 [1.085 , 2.260] | | | | |
| TD | ring | 16 | -0.405 [-0.866 , 0.056] | 0.743 [0.410 , 1.077] | 0.984 [0.681 , 1.286] | 2.260 [1.572 , 2.947] | | | | |
| TD | conifers | 15 | -0.396 [-0.873 , 0.080] | 0.808 [0.468 , 1.147] | 1.142 [0.842 , 1.441] | 1.606 [0.892 , 2.321] | | | | |

Statistical significant levels: * p<0.1 ; ** p<0.05; *** p<0.01; **** p<0.001.

2.4.1 Sap flow measurement errors across methods and methodological families

A relatively small part of the total variability in the quality of calibrations is related to methods and families of methods and, to a lesser extent, to the calibration material (fixed effects explain 8 – 28% of the variability in calibration metrics; see R_m^2 values in Table A3). Despite the high variability within methods, we detected significant differences between methods. Dissipation methods were the only methods for which accuracy was significantly lower than expected for an ideal calibration. This is consistent with previous reports (Lu & Chacko, 1998; Braun & Schmid, 1999; de Oliveira Reis *et al.*, 2006; Montague & Kjelgren, 2006; McCulloh *et al.*, 2007; Taneda & Sperry, 2008; Uddling *et al.*, 2009; Bush *et al.*, 2010; Steppe *et al.*, 2010; Caterina *et al.*, 2013; Chan, 2015; Fuchs *et al.*, 2017; Rubilar *et al.*, 2017) and our synthesis confirms that most of the individual TD and all TTD calibrations underestimate sap flow systematically (Fig. 2.5). Interestingly, however, other studies have found the opposite result (Lu, 2002; Cain, 2009; Hultine *et al.*, 2010; Sperling *et al.*, 2012; Sun *et al.*, 2012) and simulation models (Wullschleger *et al.*, 2011; Hölttä *et al.*, 2015) suggest that it is difficult to state a priori whether TD will over- or underestimate flow, as the measurements obtained are highly dependent on wood properties and on flux conditions. Our results show that, globally, the conditions leading to underestimation are more frequent and support the existence of a proportional bias underlying this systematic underestimation by TD (Fig. 2.3). It must be also noted, however, that Dissipation methods have been tested against a much wider range of flow conditions compared to the rest of the methods (Fig. 2.5, Fig. 2.6).

Calibration parameters of the TD method were originally considered to be universal but subsequent studies have claimed that species-specific calibrations are necessary to obtain correct sap flow measurements (Lu *et al.*, 2004; Steppe *et al.*, 2010; Fuchs *et al.*, 2017). For a set of diffuse-porous species, using a pooled calibration also substantially improved TD (but not HFD) performance compared to measurements obtained with the original calibration (Fuchs *et al.*, 2017). However, our results show that species in general and wood porosity type in particular explain a small or even no proportion of the variability in the calibrations (Table 2.3 and A3). This implies that factors related to the experimental context and, possibly, to intraspecific variability in wood properties (cf. section 2.5.2) may have a large contribution to overall uncertainty. Therefore, our results suggest that calibration parameters for TD or HFD, obtained under different experimental

Table 2.4: Synthesis of the potential sources of error and use adequacy for each method. Crosses indicates that the method is sensitive to the respective source of error (updated from Vandegehuchte and Steppe, 2013). Methods are classified according to their use effectiveness under different flow conditions: dark grey, light grey and white indicate highly, partially and no recommended use, respectively. When assessing use adequacy for high/low flows, dark and light gray indicate a Normalized Root Mean Square Error (NRMSE) less than a 22% and a 44%, respectively, calculated with the NRMSE model (Table 2.2). In Absolute flows use recommendation, dark grey shows methods with both accuracy (Ln-Ratio) and proportional bias (Slope) not significantly different from a perfect calibration. In Relative flows use recommendation, dark grey shows methods with linearity (Slope (ln-ln)) not significantly different from a perfect calibration and with reasonable precision. Potentiality of measuring small stems diameters (< 125mm) is also reported.

| Method | Potential source of measure error | | | | | | | | | Effectiveness in measuring | | | | |
|--------|-----------------------------------|-------------------------|-----------------|---------------------------|---------------------|---------------|------------|-------------|--------------|----------------------------|------------|-------------|----------------|----------------|
| | Wounding | Radial velocity profile | Wood properties | Natural thermal gradients | Sensor installation | Sensor design | Baselining | Power input | Pulse length | Reverse flows | Low flows* | High flows* | Absolute flows | Relative flows |
| CHP | x | x | x | x | x | | | | x | | | | | |
| T-max | x | x | x | | x | | x | | | | | | | |
| HR | x | x | x | | x | | | | | | | | | |
| HFD | x | x | | x | x | x | x | x | | | | | | |
| SHB | | | | x | | | x | x | | | | | | |
| TD | x | x | x | x | | x | x | x | | | | | | |
| TTD | x | x | x | x | | x | x | x | | | | | | |

* (Low/High: SFD methods: <5 / >80 cm³ cm⁻² h⁻¹; SF methods: <260 / >3900 cm³ h⁻¹)

contexts, may not be generalizable to species level, as also suggested by Fuchs *et al.* (2017).

In addition to Dissipation methods, Pulse methods also suffer proportional bias, probably driven by overestimation at low flows, although this was significant for T-max only (i.e. positive intercepts in linear models fitted to calibration data; Fig. 2.5 and 2.6 and A3). It is well known that the equations of CHP and T-max cannot be solved at sap flows close to 0, and the calibration intercepts observed here (Fig. A3) are consistent with the detection thresholds reported for T-max (~10 cm³ cm⁻² h⁻¹; Green *et al.*, 2003) and CHP (2-4 cm³ cm⁻² h⁻¹; Green *et al.*, 2003; Bleby *et al.*, 2004). Our results confirm and generalize a previously reported low-flow detectability problem for T-max (Green *et al.*, 2003, 2009; Vandegehuchte & Steppe, 2012a), but we could not confirm it for CHP as described before (Barrett *et al.*, 1995; Becker, 1998; Bleby *et al.*, 2004; Vandegehuchte & Steppe, 2012a). Despite overestimation at low flows, the average accuracy of CHP and T-max is good, which implies that low-flow overestimations may be compensated with underestimations at high flows. This is also shown by the lack of linearity observed in both methods (Slope (ln-ln) < 1; Fig. 2.3(b)).

Our analysis did not detect the saturation effect for the HR method at high flows that has been reported elsewhere (Bleby *et al.*, 2008; Green *et al.*, 2009; Steppe *et al.*, 2015). This is likely due to the fact that HR calibrations considered here include few observations in the region where this overestimation occurs ($>\sim 45 \text{ cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$; Figs 2.5 and 2.6). Moreover, the high variability in the calibrations probably precluded detection of the saturation effect (Fig. 2.3 and 2.5) and of the apparent trend of increasing NRMSE with sap flow range for HR (Table 2.3 and Fig. A2). A lack of linearity can also be observed for HFD, consistent with the suggested tendency of this method to underestimate at high flows (Vandegehuchte & Steppe, 2012c).

Despite the large variability in precision within methods, our results show that calibrations performed with HFD give more precise results than those conducted using the CHP, TD and TTD methods. Although this result should be interpreted with care as it is based on 57 calibrations but only from 3 studies, the higher precision observed with HFD could lie in the second dimension included in the method, which could better capture the effect of anisotropy of the wood structure. This would also be consistent with the fact that SHB, a method that is assumed to integrate sap flow variability within the stem, was the method with the second highest precision on average, albeit precision was very variable for this method (Fig. 2.3(b)).

We did not detect differences in accuracy, proportional bias or linearity of the calibrations across calibration materials. However, compared to an ideal calibration, we did find proportional bias and lack of linearity in calibrations performed on whole plants, probably because these calibrations use large scales whose sensitivity and resolution are usually low, potentially affecting low-flow measurements and leading to artefactual overestimation at low flows. Poor linearity may also be due to non-linear changes in belowground hydraulic resistance as the sap flow increases (Martínez-Vilalta *et al.*, 2007). In cut plants, we may have two opposite effects, as cutting could eliminate belowground resistance (favoring flow) but add resistance due to putative embolism formation after cutting. Similarly, the higher precision of calibrations conducted on cut stems relative to those conducted on whole plants (with and without roots), likely reflects that cut stem calibrations are normally conducted in laboratories with precision scales and under controlled conditions that minimize experimental random errors.

2.4.2 The performance of sap flow calibrations is largely unrelated to species wood traits

Species-specific wood density and wood porosity type explained little variability in overall calibration performance, although we detected some effects of wood density for HFD and TD calibrations. Wood density affected HFD measurements by increasing precision, which could be related to the response time of the sensors. If we assume that maximum sapwood water content is reduced as wood density increases (Simpson, 1993), associated changes in thermal diffusivity could lead to a faster sensor response (Hölttä *et al.*, 2015), higher correlation between actual and measured flows. Wood density also showed a negative relationship with proportional bias for HFD and TD, a pattern that could be caused by the combined effects of wood density and water content on wood thermal diffusivity (Vandegehuchte & Steppe, 2012c; Vergeynst *et al.*, 2014). The fact that we did not find clear effects of wood density on calibration accuracy and linearity, despite that wood density affects thermal diffusivity and hence heat transport (Wullschleger *et al.*, 2011), could be

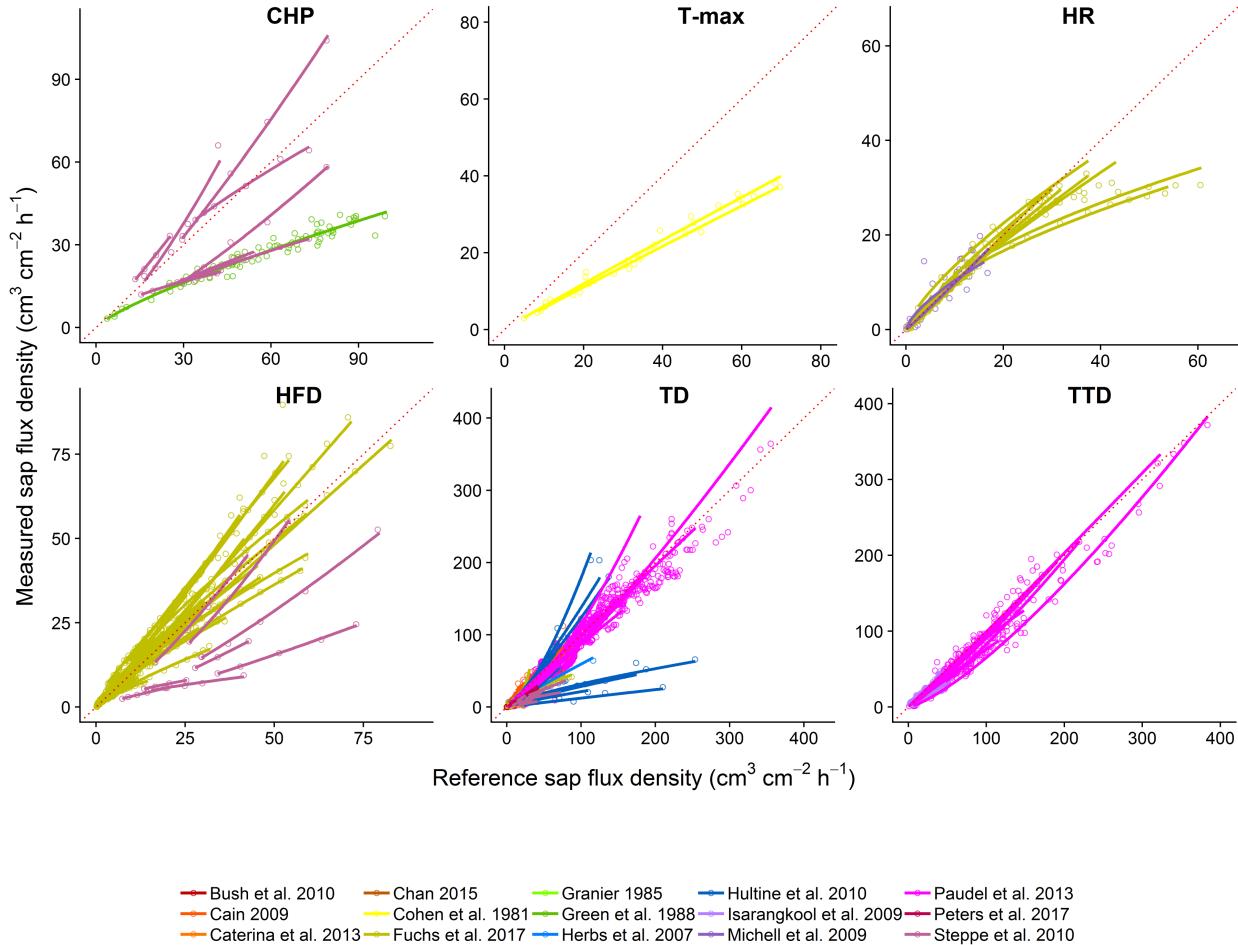


Figure 2.5: Relationship between measured and reference sap-flux density (SFD) for different sap flow methods, studies and calibrations. The fits of ln–ln regressions (Eq. 2.4) for each calibration are also depicted. Different colors represent different studies that report results in sap-flux density units. Scales vary across panels to facilitate intra method comparison. The red dotted line indicates the 1:1 relationship.

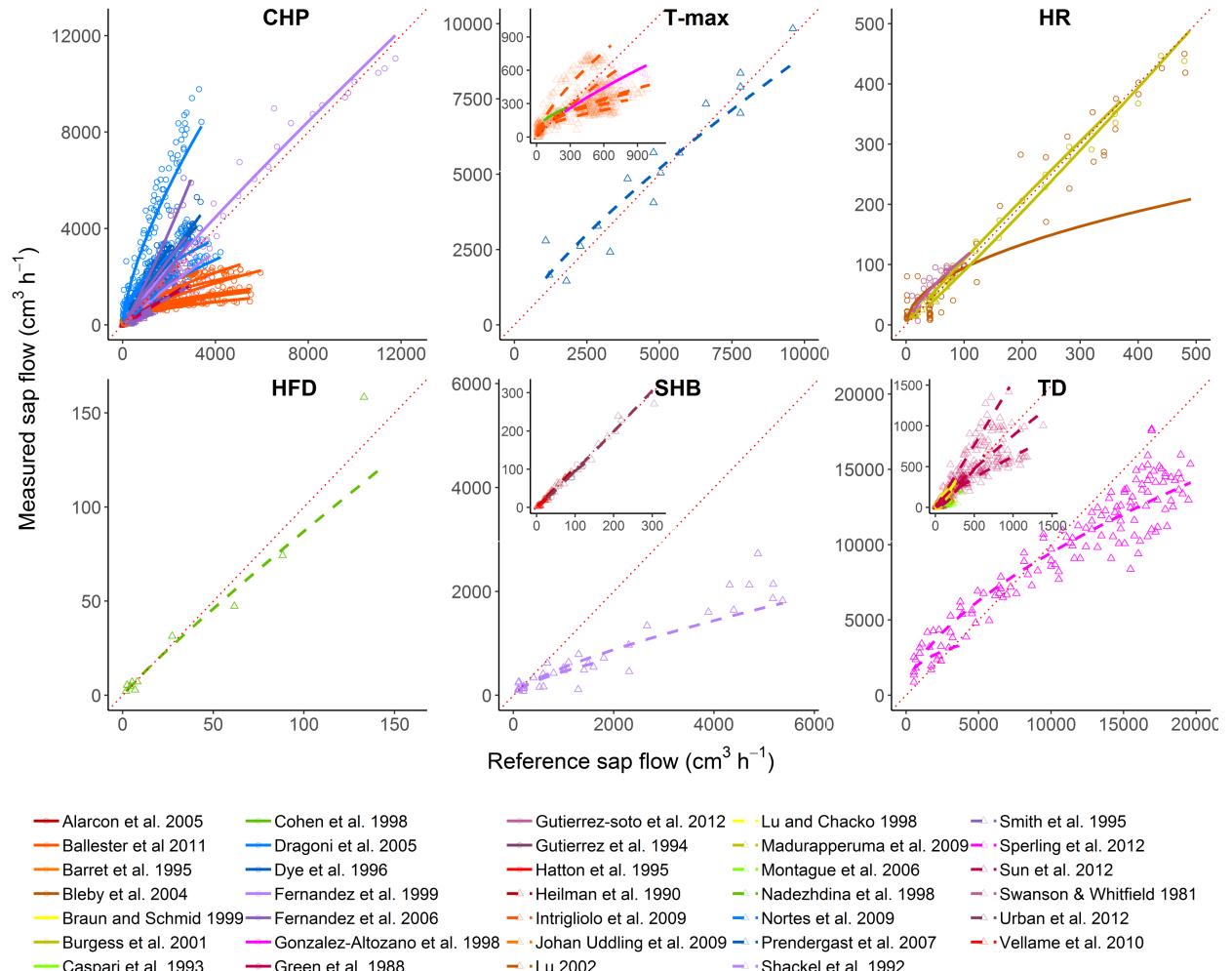


Figure 2.6: Relationship between measured and reference sap flow (SF) for different sap flow methods, studies and calibrations. The fits of ln-ln regressions (Eq. 2.4) for each calibration are also depicted. Different color symbols and line types represent different studies. Scales varies across panels to facilitate intra method comparison. Insets are shown in some panels (T-max, SHB, TD) to facilitate visualization when the flow ranges differed markedly among calibrations for the same method. The red dotted line indicates 1:1 relationship.

explained by two reasons. Firstly, we could not use the actual wood density for most of the calibrations, because it was not reported in the corresponding studies, and using species-level averages instead of the wood density of the plant material specifically used on the calibrations may mask the effect of wood density on calibration performance. Secondly, wood density in angiosperms appears to be only weakly correlated to some wood properties that could be important for sap flow calibrations, such as lumen fraction (Zanne *et al.*, 2010).

Our global analysis did not show clear and consistent differences in calibration quality between different wood porosity types (Table 2.2) as previously suggested by several studies for both CHP (Green & Clothier, 1988) and TD methods (Bush *et al.*, 2010; Sun *et al.*, 2012). According to heat transport theory, we should expect declining performance from conifer to ring-porous species (i.e. from most homogenous to most heterogeneous wood). For CHP, we found that proportional bias (only marginally) and nonlinearity departed from an ideal calibration for diffuse-porous species, but these patterns did not differ significantly from those observed for conifers. Our results did not clearly support either an inferior performance of TD in ring-porous species compared to diffuse-porous or conifers, as could be expected from the reported underestimation driven by large sap flow gradients along sensor length or by the imperfect probe contact with hydroactive xylem in species with narrow sapwood (Clearwater *et al.*, 1999; but see Wullschleger *et al.*, 2011). Wood porosity effects on sap flow calibrations have been inferred in individual studies from measurements in few species representative of each wood porosity type (Bush *et al.*, 2010; Sun *et al.*, 2012; Xie & Wan, 2018) and our inability to detect these effects here may be caused by the high variability in experimental context within our dataset. Furthermore, the effect of the different anatomies may be masked by high structural variability within wood porosity types, as for example the variation in latewood to earlywood in conifers (Fan *et al.*, 2018) and we cannot discard that calibration performance could be related to quantitative anatomical traits not assessed in this study (cf. Xie & Wan, 2018). Although the low variability we observed at the species level suggests that quantitative anatomical traits might not explain much of the variability in sap flow calibrations, we encourage that quantitative wood traits are measured in the same plant material used to calibrate sap flow sensors to better understand the influence of wood properties on the variability of sap flow calibrations.

2.4.3 Implications and recommendations

Our global analysis shows that even when the methods are applied following standard recommendations the quality of individual calibrations can be very low (Fig. 2.3). This result reflects, on one hand, systematic bias in TD and lack of linearity in CHP, two of the most widely used methods (Fig. A1) and, on the other hand, unknown sources of error related to experimental conditions and/or sample characteristics (Table 2.4). In our study, we could not account for all the experimental conditions to evaluate these sources of variability, except for the effect of the calibration material. Examples of factors that may affect calibrations when using the same type of calibration material include sensor design (Fuchs *et al.*, 2017), sensor installation (Lu & Chacko, 1998; Bleby *et al.*, 2004; Ren *et al.*, 2017), variation in calculations of wood thermal properties (Looker *et al.*, 2016), zero flow determination (Looker *et al.*, 2016; Peters *et al.*, 2018) or the mechanism of flow generation in cut stem calibrations (negative vs positive pressures) (Fuchs *et al.*, 2017) (Table 2.4). Previous reports, however, usually focus on only one of the sources of experimental error.

Importantly, relevant methodological information that could be used to assess (and account for) these sources of error is frequently not reported (Steppe *et al.*, 2015; Peters *et al.*, 2018). Clearly, further research into the effects of experimental conditions on the quality of different sap flow methods should be a priority, as well as more complete, standardized reporting of experimental conditions, including information on the sources of potential methodological errors listed in Table 2.4.

Our results show that calibrations may be needed to obtain correct absolute values of sap flow, even when Pulse methods are used (Steppe *et al.*, 2010; see also Fuchs *et al.*, 2017). However, sap flow calibrations provide a snapshot of the performance of a given sap flow method under relatively stable conditions, which may greatly differ from those experienced by plants in the field. Moreover, our analysis could not address the methodological variability related to more dynamic effects such as errors caused by changes in sapwood water content (Vergeynst *et al.*, 2014), long-term wounding or signal dampening (Wiedemann *et al.*, 2013; Marañón-Jiménez *et al.*, 2018; Peters *et al.*, 2018). In this sense, more studies should assess calibration applicability to mid- or long-term measurements (e.g., Oliveras & Llorens, 2001), possibly combined with independent estimates of sapwood water content (Vandegehuchte & Steppe, 2012a) and whether calibrations obtained from excised segments are valid for whole-plants.

Considering only their performance in calibration tests (i.e. no other logistic or technical issues, such as sensor, datalogging, or power constraints, which will be study-specific) we can provide some general recommendations on the use of sap flow methods (Table 2.4). The most widely used method, TD, appears to be consistently inaccurate, shows proportional bias and generally underestimates sap flow, by 40% on average (if used with its original calibration coefficients). However, it presents good linearity, which implies that this method can be used when sap flow responses to environmental variables and/or treatments are the primary focus of the study (i.e., good estimates of absolute sap flow values are not critical). In comparison, CHP, T-max and HFD all present a certain nonlinearity which may affect the estimation of these environmental responses. At least for CHP and T-max (specially for the latter) this pattern seems to be driven by overestimation at low flows and underestimation at high flows canceling out each other. This implies that both Pulse methods could be suitable for studies interested in absolute values of transpiration. For the HFD method, the nonlinearity could be influencing the estimations of radial sap flow patterns, as these measurements would need to correctly measure both high and low flows simultaneously. We also confirm that the HR method may not be suitable to measure high flows but it is probably the best method for detailed physiological studies involving low flows.

2.5 Conclusions

In conclusion, our global assessment contributes towards a proper incorporation of measurement errors in the interpretation of individual case studies and in modelling studies aimed at upscaling sap flow data (Hatton *et al.*, 1995; Hernandez-Santana *et al.*, 2015). Perhaps even more importantly, it paves the way towards improved intercomparison of sap flow datasets obtained with different methods to assess regional or global patterns in plant water use (e.g., the SAPFLUXNET initiative; Poyatos *et al.*, 2016). Although providing explicit correction factors for each method is beyond the scope of this paper, the typical accuracy deviations provided in Table 2.1 can be used

as a first order correction when combining sap flow data from different methods (and no additional information on study-specific uncertainty sources is available).

3

Global transpiration data from sap flow measurements: the SAPFLUXNET database

Rafael Poyatos, Víctor Granda, Víctor Flo, Jordi Martínez-Vilalta *et al.* (2020) Earth System Data Science, Pre-print.

Abstract

Plant transpiration links physiological responses of vegetation to water supply and demand with hydrological, energy and carbon budgets at the land-atmosphere interface. However, despite being the main land evaporative flux at the global scale, transpiration and its response to environmental drivers are currently not well constrained by observations. Here we introduce the first global compilation of whole-plant transpiration data from sap flow measurements (SAPFLUXNET, <https://sapfluxnet.creaf.cat/>). We harmonised and quality-controlled individual datasets supplied by contributors worldwide in a semi-automatic data workflow implemented in the R programming language. Datasets include sub-daily time series of sap flow and hydrometeorological drivers for one or more growing seasons, as well as metadata on the stand characteristics, plant attributes and technical details of the measurements. SAPFLUXNET contains 202 globally distributed datasets with sap flow time series for 2714 plants, mostly trees, of 174 species. SAPFLUXNET has a broad bioclimatic coverage, with woodland/shrubland and temperate forest biomes especially well-represented (80% of the datasets). The measurements cover a wide variety of stand structural characteristics and plant sizes. The datasets encompass the period between 1995 and 2018, with 50% of the datasets being at least 3 years long. Accompanying radiation and vapour pressure deficit data are available for most of the datasets, while on-site soil water content is available for 56% of the datasets. Many datasets contain data for species that make up 90% or more of the total stand basal area, allowing the estimation of stand transpiration in diverse ecological settings. SAPFLUXNET adds to existing plant trait datasets, ecosystem flux networks and remote sensing products to help increase our understanding of plant water use, plant responses to drought and ecohydrological processes. SAPFLUXNET version 0.1.5 is freely available from the Zenodo repository (<https://doi.org/10.5281/zenodo.3971689>, Poyatos *et al.*, 2020c). The ‘sapfluxnetr’ R package, designed to access, visualise and process SAPFLUXNET data is available from CRAN.

3.1 Introduction

Terrestrial vegetation transpires ca. 45000 km³ of water per year (Schlesinger & Jasechko, 2014; Wang-Erlandsson *et al.*, 2014; Wei *et al.*, 2017), a flux that represents 40% of global land precipitation, 70% of total land evapotranspiration (Oki & Kanae, 2006), and is comparable in magnitude to global annual river discharge (Rodell *et al.*, 2015). For most terrestrial plants, transpiration is an inevitable water loss to the atmosphere because they need to open stomata to allow CO₂ diffusion into the leaves for photosynthesis. Latent heat from transpiration represents 30–40% of surface net radiation globally (Schlesinger & Jasechko, 2014; Wild *et al.*, 2015). Transpiration is therefore a key process coupling land-atmosphere exchange of water, carbon and energy, determining several vegetation-atmosphere feedbacks, such as land evaporative cooling or moisture recycling. Regulation of transpiration in response to fluctuating water availability and/or evaporative demand is a key component of plant functioning and one of the main determinants of a plant's response to drought (Whitehead, 1998; Martin-StPaul *et al.*, 2017). Despite its relevance for earth functioning, transpiration and its spatiotemporal dynamics are poorly constrained by available observations (Schlesinger & Jasechko, 2014) and not well represented in models (Fatichi *et al.*, 2016; Mencuccini *et al.*, 2019a). An improved understanding on how plants regulate transpiration is thus needed to better predict future trajectories of land evaporative fluxes and vegetation functioning under increased drought conditions driven by global change.

Conceptually, transpiration can be quantified at different organisational scales: leaves, branches and whole plants, ecosystems and watersheds. In practice, transpiration is relatively easy to isolate from the bulk evaporative flux, evapotranspiration, only from the leaf to the plant levels. In terrestrial ecosystems, evapotranspiration includes evaporation from the soil and from water-covered surfaces, including plants. Transpiration measurements on individual leaves or branches with gas exchange systems are difficult to upscale to the plant level (Jarvis, 1995). Likewise, transpiration measurements using whole-plant chambers (e.g. Pérez-Priego *et al.*, 2010) or gravimetric methods (e.g. weighing lysimeters) in the field are still challenging. At the ecosystem scale and beyond, evapotranspiration is generally determined using micrometeorological methods, catchment water budgets or remote sensing approaches (Shuttleworth, 2007; Wang & Dickinson, 2012). In some cases, isotopic methods and different algorithms applied to measured ecosystem fluxes can provide an estimation of transpiration at the ecosystem scale (Kool *et al.*, 2014; Stoy *et al.*, 2019).

Transpiration drives water transport from roots to leaves in the form of sap flow through the plant's xylem pathway (Tyree & Zimmermann, 2002), and this sap flow affects heat transport in the xylem. Taking advantage of this, thermometric sap flow methods were first developed in the 1930s (Huber, 1932) and further refined over the following decades (Marshall, 1958; Čermák *et al.*, 1973) to provide operational measurements of plant water use. These methods have become widely used in plant ecophysiology, agronomy and hydrology (Poyatos *et al.*, 2016), especially after the development of simple, easily replicable methods (e.g. Granier, 1985, 1987). Whole-plant measurements of water use using thermometric sap flow methods provide estimates of water flow through plants from sub-daily to interannual timescales, and have been mostly applied in woody plants (but see Baker & Van Bavel, 1987 for measurements on herbaceous species). Xylem sap flow is measured semi-invasively (Brodersen *et al.*, 2019) and can be upscaled to the whole plant, obtaining a near-continuous quantification of plant water use. Multiple sap flow sensors can be

deployed, in almost any terrestrial ecosystem, to determine the magnitude and temporal dynamics of transpiration across species, environmental conditions or experimental treatments. All sap flow methods are subject to methodological and scaling issues, which may affect the quantification of absolute water use in some circumstances (Smith & Allen, 1996; Köstner *et al.*, 1998; Čermák *et al.*, 2004; Vandegehuchte & Steppe, 2013). Nevertheless, all methods are suitable for the assessment of the temporal dynamics of transpiration and of its responses to environmental changes or to experimental treatments (Flo *et al.*, 2019).

The generalised application of sap flow methods in ecological and hydrological research in the last 30 years has thus generated a large volume of data, with an enormous potential to advance our understanding of the spatiotemporal patterns and the ecological drivers of plant transpiration and its regulation (Poyatos *et al.*, 2016). However, this large volume of data needs to be compiled and harmonised to enable global syntheses and comparative studies across species and regions. Across-species data syntheses using sap flow data have mostly focused on maximum values extracted from publications (Wullschleger *et al.*, 1998; Kallarackal *et al.*, 2013; Manzoni *et al.*, 2013). Multi-site syntheses have focused on the environmental sensitivity of sap flow, using site means of plant-level sap flow or sap flow-derived stand transpiration (Poyatos *et al.*, 2007; Tor-ngern *et al.*, 2017). Since data sharing is only incipient in plant ecophysiology, sap flow datasets have not been traditionally available in open data repositories. Open data practices are now being implemented in databases, which fosters collaboration across monitoring networks in research areas relevant to plant functional ecology (Falster *et al.*, 2015; Kattge *et al.*, 2020; Gallagher *et al.*, 2020) and ecosystem ecology (Bond-Lamberty & Thomson, 2010). The success of the data sharing and data re-use policies within the FLUXNET global network of ecosystem level fluxes has shown how these practices can contribute to scientific progress (Bond-Lamberty, 2018).

Here we introduce SAPFLUXNET, the first global database of sap flow measurements built from individual community-contributed datasets. We implemented this compilation in a data structure designed to accommodate time series of sap flow and the main hydrometeorological drivers of transpiration, together with metadata documenting different aspects of each dataset. We harmonised all datasets and performed basic semi-automated quality assurance and quality control procedures. We also created a software package that provides access to the database, allows easy visualisation of the datasets and performs basic temporal aggregations. We present the ecological and geographic coverage of SAPFLUXNET version 0.1.5, (Poyatos *et al.*, 2020c) followed by a discussion of potential applications of the database, its limitations and a perspective of future developments.

3.2 The SAPFLUXNET data workflow

3.2.1 An overview of sap flow measurements

The main characteristics of sap flow methods have been reviewed elsewhere (Swanson, 1994; Smith & Allen, 1996; Čermák *et al.*, 2004; Vandegehuchte & Steppe, 2013). Given the already broad scope of the paper, here we only provide a brief methodological overview, without delving into the details of the individual methods. Sap flow sensors track the fate of heat applied to the plant's conducting tissue, or sapwood, using temperature sensors (thermocouples or thermistors), usually deployed

in the plant's main stem. Both heating and temperature sensing can be done either internally, by inserting needle-like probes containing electrical resistors (or electrodes for some methods) and temperature sensors into the sapwood, or externally; these latter systems being especially designed for small stems. Depending on how the heat is applied and the principles underlying sap flow calculations, sap flow sensors can be classified into three major groups: heat dissipation methods, heat pulse methods and heat balance methods (Flo *et al.*, 2019). Heat dissipation and heat pulse methods estimate sap flow per unit sapwood area and they have been called 'sap flux density methods' (Vandegehuchte & Steppe, 2013); heat balance methods directly yield sap flow for the entire stem or for a sapwood section. Heat dissipation methods include the constant heat dissipation (HD; Granier, 1985, 1987), the transient (or cyclic) heat dissipation (CHD; Do & Rocheteau, 2002a) and the heat deformation (HFD; Nadezhina, 2018) methods. Heat pulse methods include the compensation heat pulse (CHP; Swanson & Whitfield, 1981), heat ratio (HR; Burgess *et al.*, 2001), T-max (HPTM; Cohen *et al.*, 1981) and Sapflow+ (Vandegehuchte & Steppe, 2012c) methods. Heat balance methods include the trunk sector heat balance (TSHB; Čermák *et al.*, 1973) and the stem heat balance (SHB; Sakuratani, 1981) methods. The suitability of a certain method in a given application largely depends on plant size and the flow range of interest (Flo *et al.*, 2019), but HD and CHP are the most widely used (Poyatos *et al.*, 2016; Peters *et al.*, 2018; Flo *et al.*, 2019). Apart from these different methodologies, within each sap flow method variants exist in sensor design and in data processing approaches, resulting in relatively high levels of methodological uncertainty comparable to those in other areas of plant ecophysiology.

The output from sap flow sensors is automatically recorded by dataloggers, at hourly or even higher temporal resolution. This output relates to heat transport in the stem and needs to be converted to meaningful quantities of water transport, such as sap flow per plant or per unit sapwood area. How this conversion is achieved varies greatly across methods, with some relying on empirical calibrations and others being more physically-based and requiring the estimation of wood thermal properties and other parameters (Smith & Allen, 1996; Čermák *et al.*, 2004; Vandegehuchte & Steppe, 2013). Depending on the method and the specific sensor design, sap flow measurements can be representative of single points, linear segments along the sapwood, sapwood area sections or entire stems. Except for stem heat balance methods, these measurements need to be spatially integrated to account for radial (Phillips *et al.*, 1996; Nadezhina *et al.*, 2002; Cohen *et al.*, 2008; Berdanier *et al.*, 2016) and azimuthal (Oren *et al.*, 1999a; Lu *et al.*, 2000; Cohen *et al.*, 2008) variation of sap flow within the stem to obtain an estimate of whole-plant water use (Čermák *et al.*, 2004). At a minimum, an estimate of sapwood area is needed to upscale the measurements to whole-plant sap flow rates. Sap flow rates can thus be expressed per individual (i.e. plant or tree), per unit sapwood area (normalising by water-conducting area), and per unit leaf area (normalising by transpiring area).

Here we will use the term 'sap flow' when referring, in general, to the rate at which water moves through the sapwood of a plant and, more specifically, when we refer to sap flow per plant (i.e. water volume per unit time, Edwards *et al.*, 1997). We acknowledge that the term 'sap flux' has also been proposed for this quantity (Lemeur *et al.*, 2009), but more generally, 'sap flux density' (e.g. Vandegehuchte & Steppe, 2013) or just 'sap flux' are used to refer to 'sap flow per unit sapwood area'. Since here we include methods natively measuring sap flow per plant or per sapwood area, throughout this paper we will use the more general term 'sap flow', and, when

necessary, we will indicate explicitly the reference area used: ‘sap flow per (unit) sapwood area’, ‘sap flow per (unit) leaf area’ or ‘sap flow per (unit) ground area’.

3.2.2 Data compilation

SAPFLUXNET was conceived as a compilation of published and unpublished sap flow datasets (Table B8) and thus the ultimate success of the initiative critically depended on the contribution of datasets by the sap flow community. An expression of interest showed that a critical mass of datasets with a wide geographic distribution could potentially be contributed and the results of this survey were used to raise the interest of the sap flow community (Poyatos *et al.*, 2016). The data contribution stage was open between July 2016 and December 2017 although a few additional datasets were updated during the data quality control process and contain more recent data.

All contributed datasets had to meet some minimum criteria before they were accepted, both in terms of content and format. We required that all datasets contained sub-daily, processed sap flow data, representative of whole-plant water use under different hydrometeorological conditions. This meant that both the processing from raw temperature data to sap flow quantities and the scaling from single-point measurements to whole-plant data had been performed by the data contributor responsible for each dataset. Time-series of sap flow data and hydrometeorological drivers were required to be representative of one growing-season, setting, as broad reference, a minimum duration of 3 months. Sap flow could be either expressed as total flow rate per plant or per unit sapwood area. Contributors also needed to provide metadata on relevant ecological information of the site, stand, species and measured plants as well as on basic technical details of the sap flow and hydrometeorological time-series. Datasets had to be formatted using a documented spreadsheet template (cf. ‘sapfluxnet_metadata_template.xlsx’ in <https://github.com/vflo/PhD/tree/master/Chapter3/Supplementary>) and uploaded to a dedicated server at CREAF, Spain, using an online form.

3.2.3 Data harmonisation and quality control: QC1

Once datasets were received, they were stored and entered a process of data harmonisation and quality control (Fig. 3.1, Fig. B1). This process combined automatic data checks with human supervision, and the entire workflow was governed by functions and scripts in the R language (R Core Team, 2017), including other related tools, such as R markdown documents and Shiny applications. All R code involved in this QC process was implemented in the sapfluxnetQC1 package (Granda *et al.*, 2016). To aid in the detection of potential data issues throughout the entire process (Fig. 3.1, Fig. B1), we implemented several elements of control: (1) automatic log files tracking the output of each QC function applied, (2) automatic creation and update of status files, tracking the QC level reached by each dataset, (3) automatic QC summary reports in the form of R markdown documents, (4) interactive Shiny applications for data visualisation, (5) documentation of manual changes applied to the datasets using manually-edited text files, (6) storage of manual data cleaning operations in text files, and (7) automatic data quality flagging associated with each dataset. All these items ensure a robust, transparent, reproducible and scalable data workflow.

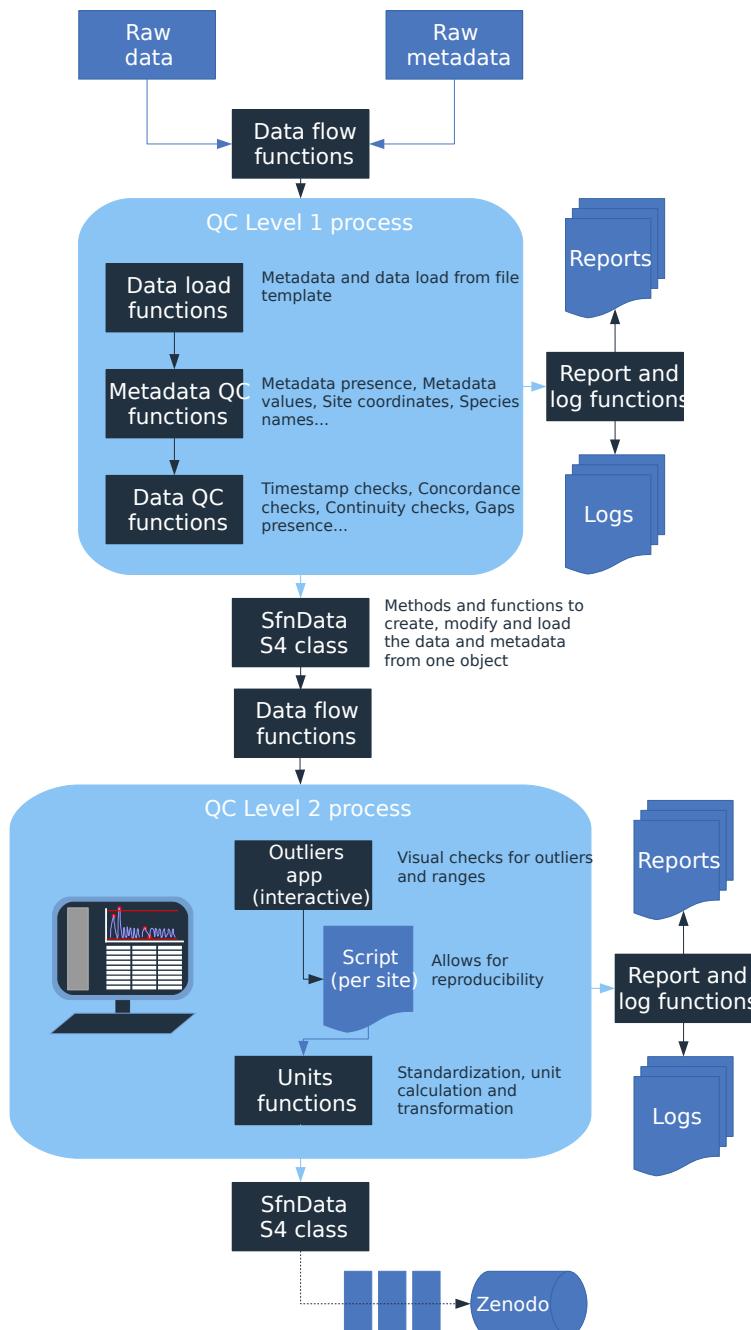


Figure 3.1: Overview of the SAPFLUXNET data workflow. Data files are received from data contributors, and undergo several quality-control processes (QC1 and QC2). Both, QC1 and QC2 produce an .RData object of the custom-designed `sfn-data` S4 class storing all data, metadata and data flags for each dataset. The progress and results of the QC processes are monitored through individual reports and log files. The final outcome, is stored in a folder structure with a either single .RData file for each dataset or a set of seven csv files for each dataset.

The first stage of the data QC (QC1) performed several data checks (Table B1) on received spreadsheet files and produced an interactive report in an R markdown document, which signalled possible inconsistencies in the data and warned of potential errors. These data issues were addressed, with the help of data contributors, if needed. Once no errors remained, the dataset was converted into an object of the custom-designed ‘`sfn_data`’ class (Fig. B2, see also section 3.2.5), which contained all data and metadata for a given dataset (Tables B2–B6 list all variable names). Data and metadata belonging to all Level 1 datasets were further visually inspected using an in-

teractive R Shiny application, and, if no major issues were detected, they were subjected to the second QC process, QC2.

3.2.4 Data harmonisation and quality control: QC2

Datasets entering QC2 underwent several data cleaning and data harmonisation processes (Table B7). We first ran outlier detection and out of range checks; these checks did not delete or modify the data, only warned about any suspicious observation ('outlier' and 'range' warnings). The outlier detection algorithm was based on a Hampel filter, which also estimates a replacement value for a candidate outlier (Hampel, 1974). For the range checks, we defined minimum and maximum allowed values for all the time series variables, based on published values of extreme weather records and maximum transpiration rates (Cerveny *et al.*, 2007; Manzoni *et al.*, 2013). The outcome of outlier and range checks were visually inspected on the actual time series being evaluated using an interactive R Shiny application (Fig. B3). Following expert knowledge, visually confirmed outliers were replaced by the values estimated by the Hampel filter. Similarly, we replaced out of range values by NA if the variable was out of its physically allowed range (Fig. B3). Outlier and out of range 'warnings' for each observation (e.g. for each variable and timestep) were documented in two data flags tables, with the same dimensions as the corresponding data tables (Fig. B2). Likewise, those observations with confirmed problematic values, which were removed or replaced, were also flagged; further information can be found in the 'data flags' vignettes in the 'sapfluxnetr' package (Granda *et al.*, 2020).

Final data harmonisation processes in QC2 involved unit transformations and the calculation of derived variables (Table B7). When plant sapwood area was provided by data contributors, we interconverted between sap flow rate per plant and per unit sapwood area. If leaf area was supplied, we also calculated sap flow per unit leaf area, but note that this transformation does not take into account the seasonal variation in leaf area. In QC2 we estimated missing environmental variables which could be derived from related variables in the dataset (Table B7). We also estimated the apparent solar time and extraterrestrial global radiation from the provided timestamp and geographic coordinates using the R package 'solaR' (Lamigueiro, 2012). All estimated or interconverted observations were flagged as 'CALCULATED' in the 'env_flags' or 'sap_flags' table (Fig. B2).

3.2.5 Data structure

One of the major benefits of the SAPFLUXNET data workflow is the encapsulation of datasets in self-contained R objects of the S4 class with a predefined structure. These objects belong to the custom-designed 'sfm_data' class, which display different slots to store time series of sap flow and environmental data, their associated data flags, and all the metadata (Fig. B2). For further information please see the 'sfm_data classes' vignette in the 'sapfluxnetr' package (Granda *et al.*, 2020). The code identifying each dataset was created by the combination of a 'country' code, a 'site' code and, if applicable, a 'stand' code and a 'treatment' code. This means that several 'stands' and/or 'treatments' can be present within one 'site' (Table B3).

At the end of the QC process, we generated a folder structure with a first-level storing datasets as either 'sfm_data' objects or as a set of comma-separated (csv) text files. Within each

of these formats, a second-level folder groups datasets according to how sap flow is normalized (per plant, sapwood or leaf area); note that the same dataset, expressing different sap flow quantities, can be present in more than one folder (e.g. ‘plant’ and ‘sapwood’). Finally, the third level contains the data files for each dataset: either a single ‘sfn_data’ object storing all data and metadata, or all the individual csv files. More details on the data structure can be found in the ‘sapfluxnetr-quick-guide’ vignette in the ‘sapfluxnetr’ package (Granda *et al.*, 2020).

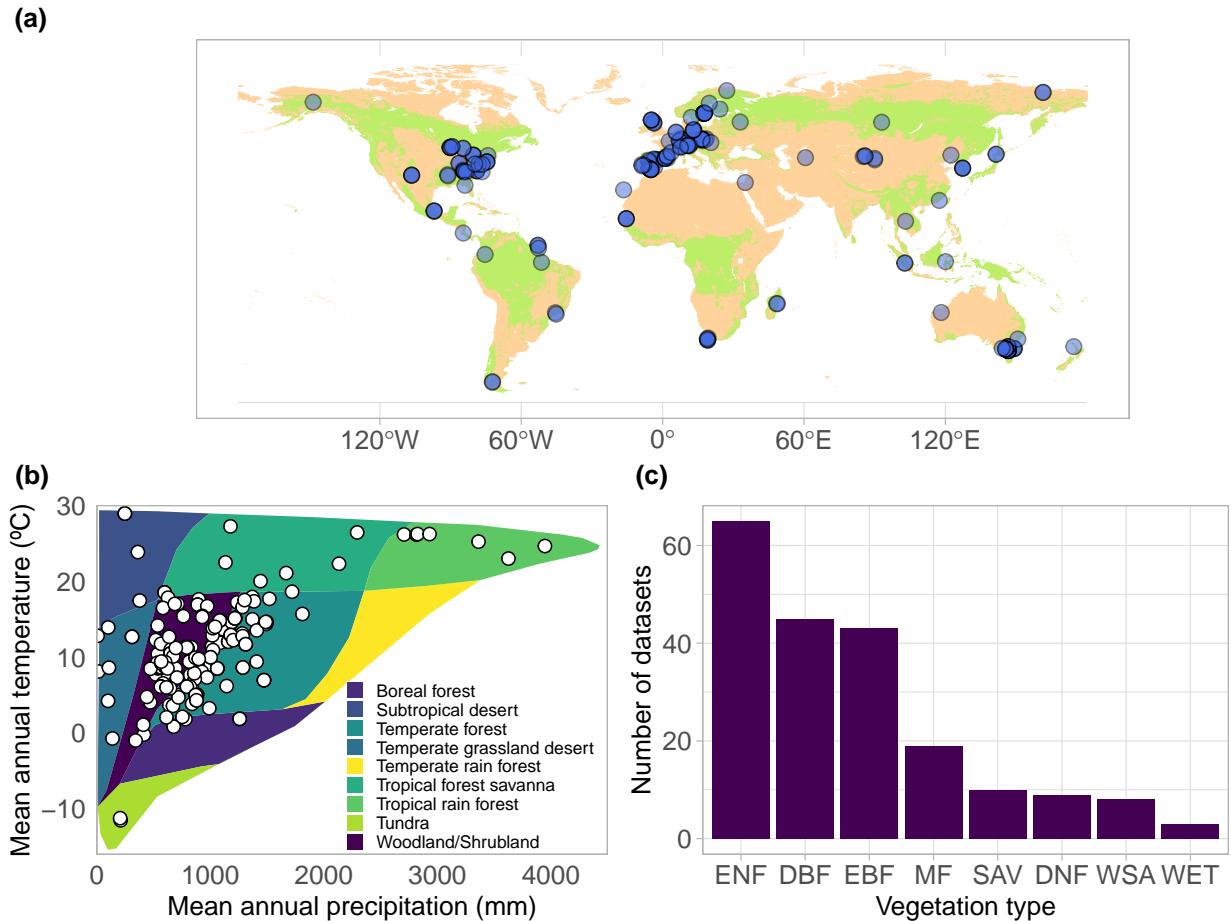


Figure 3.2: (a) Geographic, (b) bioclimatic and (c) vegetation type distribution of SAPFLUXNET datasets. In (a) woodland area from Crowther *et al.* (2015) is shown in green. In (b) we represent the different datasets according to their mean annual temperature and precipitation in a Whittaker diagram showing the classification of the main terrestrial biomes. In (c) vegetation types are defined according to the International Geosphere-Biosphere Programme (IGBP) classification (ENF: Evergreen Needleleaf Forest; DBF: Deciduous Broadleaf Forest; EBF: Evergreen Broadleaf Forest; MF: Mixed Forest; DNF: Deciduous Needleleaf forest; SAV: Savannas; WSA: Woody Savannas; WET: Permanent Wetlands).

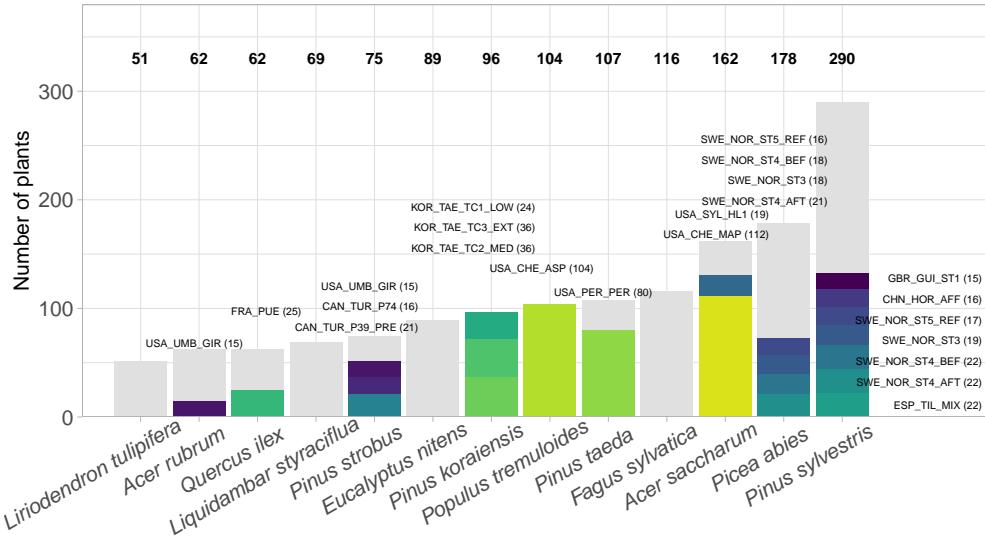
3.3 The SAPFLUXNET database

3.3.1 Data coverage

The SAPFLUXNET version 0.1.5 database harbours 202 globally distributed datasets (Fig. 3.2(a), Fig. B4 and Table B8), from 121 geographical locations, with Europe, Eastern USA and Australia especially well represented. These datasets were represented in the bioclimatic space using the terrestrial biomes delimited by Whittaker (Fig. 3.2(b)), but note that, as any bioclimatic classification, it has its limitations. Datasets have been compiled from all terrestrial biomes,

except for temperate rainforests, although some tropical montane sites have been included. Woodland/shrubland and temperate forest biomes are the most represented in the database adding up to 80% of the datasets (Fig. 3.2(b)). However, large forested areas in the tropics and in boreal regions are still not well represented (Fig. 3.2(a,b)). Looking at the distribution by vegetation type (Fig. 3.2(c)), evergreen needleleaf forest is the most represented vegetation type (65 datasets), followed by deciduous broadleaf forest (47 datasets) and evergreen broadleaf forest (43 datasets).

(a) Species with >50 plants



(b) Genera with >50 plants

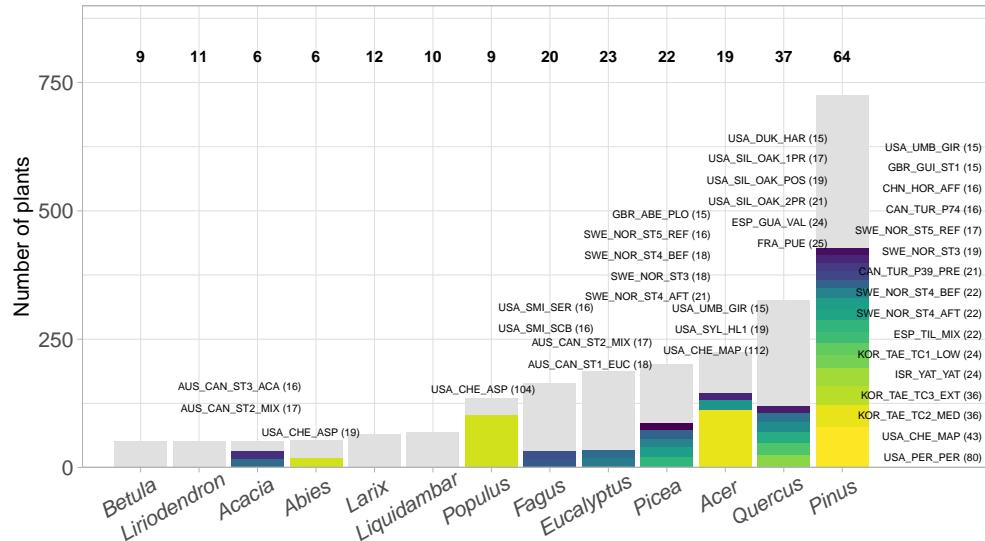


Figure 3.3: Taxonomic distribution of genera and species in SAPFLUXNET, showing (a) species and (b) genera with > 50 plants in the database. Total bar height depicts number of plants per species (a) or genera (b). Numbers on top of each bar show the number of datasets where each species (a) or genus (b) is present. Colours other than grey highlight datasets with 15 or more plants of a given species (a) or genus (b). Bar height for a given colour is proportional to the number of plants in the corresponding dataset, which is also shown in parentheses next to the dataset code.

SAPFLUXNET contains sap flow data for 2714 individual plants (1584 angiosperms and 1130 gymnosperms), belonging to 174 species (141 angiosperms and 33 gymnosperms), 95 different

genera and 45 different families (Table B9-B10). All species but one, *Elaeis guineensis*, a palm, are tree species. *Pinus* and *Quercus* are the most represented genera (Fig. 3.3(b)). Amongst the gymnosperms, *Pinus sylvestris*, *Picea abies* and *Pinus taeda* are the three most represented species with data provided on 290, 178 and 107 trees, respectively (Fig. 3.3(a)). For the angiosperms, *Acer saccharum*, *Fagus sylvatica* and *Populus tremuloides* are the most represented species, with 162, 116 and 104 trees, respectively, although most *Acer saccharum* data come from a single study with a very large sample size (Fig. 3.3(a)). Some species are present in more than 10 datasets: *Pinus sylvestris*, *Picea abies*, *Fagus sylvatica*, *Acer rubrum*, *Liriodendron tulipifera* and *Liquidambar styraciflua* (Fig. 3.3(a), Table B4).

Table 3.1: Number of sap flow times series in SAPFLUXNET depending on whether they were calibrated (species-specific), non-calibrated or this information was not provided, for the different sap flow methods: cyclic (or transient) heat dissipation (CHD), compensation heat pulse (CHP), heat dissipation (HD), heat field deformation (HFD), heat pulse T-max (HPTM), heat ratio (HR), stem heat balance (SHB) and trunk sector heat balance (TSHB). The percentage of calibrated time series was expressed with respect to the total number of sap flow time series for each method.

| Method | Calibrated | Non-calibrated | Not provided | % calibrated |
|--------|------------|----------------|--------------|--------------|
| CHD | 6 | 13 | 0 | 31.6 |
| CHP | 29 | 42 | 157 | 12.7 |
| HD | 214 | 1491 | 98 | 11.9 |
| HR | 3 | 55 | 47 | 2.9 |
| TSHB | 7 | 433 | 4 | 1.6 |
| HFD | 0 | 8 | 0 | 0.0 |
| HPTM | 0 | 80 | 0 | 0.0 |
| SHB | 0 | 27 | 0 | 0.0 |

3.3.2 Methodological aspects

For more than 90% of the plants, sap flow at the whole-plant level is available (either directly provided by contributors or calculated in the QC process); this is important for upscaling SAPFLUXNET data to the stand level (cf. section 3.4.2). Because the leaf area of the measured plants is often not available as metadata, sap flow per unit leaf area was estimated for only 18.6% of the individuals (Fig. 3.4). The heat dissipation method is the most frequent method in the database (HD, 66.4% of the plants), followed by the trunk sector heat balance (TSHB, 16.4%) and the compensation heat pulse method (CHP, 8.4%) (Fig. 3.4). This distribution is broadly similar to the use of each method documented in the literature, although the TSHB method is overrepresented here, compared to the current use of this method by the sap flow community (Poyatos *et al.*, 2016; Flo *et al.*, 2019). Some methods, especially those belonging to the heat pulse family and the cyclic (or transient) heat dissipation (CHD) method are mostly used in angiosperms, while the TSHB and the heat field deformation (HFD) methods are more frequently used in gymnosperms (Fig. 3.4).

Calibration of sap flow sensors and scaling from point measurements to the whole-plant can be critical steps towards accurate estimates of absolute sap flow rates. In SAPFLUXNET, most of the sap flow time series have not undergone a species-specific calibration, with the CHD method showing the highest percentage of calibrated time series (Table 3.1). This lack of calibrations

Table 3.2: Number of plants in the SAPFLUXNET database using different radial and azimuthal integration approaches for the different sap flow methods: cyclic (or transient) heat dissipation (CHD), compensation heat pulse (CHP), heat dissipation (HD), heat field deformation (HFD), heat pulse T-max (HPTM), heat ratio (HR), stem heat balance (SHB) and trunk sector heat balance (TSHB).

| Azimuthal integration | | | | | |
|------------------------------|----------|-------------------|---|-------------------------|--------------|
| Method | Measured | Sensor-integrated | Corrected, measured azimuthal variation | No azimuthal correction | Not provided |
| CHD | 15 | 0 | 0 | 0 | 4 |
| CHP | 61 | 0 | 0 | 167 | 0 |
| HD | 216 | 0 | 520 | 1021 | 46 |
| HFD | 0 | 0 | 0 | 8 | 0 |
| HPTM | 0 | 0 | 0 | 80 | 0 |
| HR | 7 | 0 | 2 | 88 | 8 |
| SHB | 0 | 0 | 0 | 27 | 0 |
| TSHB | 0 | 25 | 191 | 219 | 9 |

| Radial integration | | | | | |
|---------------------------|----------|-------------------|--------------------------------------|----------------------|--------------|
| Method | Measured | Sensor-integrated | Corrected, measured radial variation | No radial correction | Not provided |
| CHD | 0 | 0 | 6 | 13 | 0 |
| CHP | 222 | 0 | 6 | 0 | 0 |
| HD | 77 | 3 | 645 | 703 | 142 |
| HFD | 2 | 0 | 0 | 6 | 0 |
| HPTM | 0 | 0 | 0 | 80 | 0 |
| HR | 57 | 1 | 42 | 3 | 2 |
| SHB | 0 | 27 | 0 | 0 | 0 |
| TSHB | 0 | 338 | 8 | 89 | 9 |

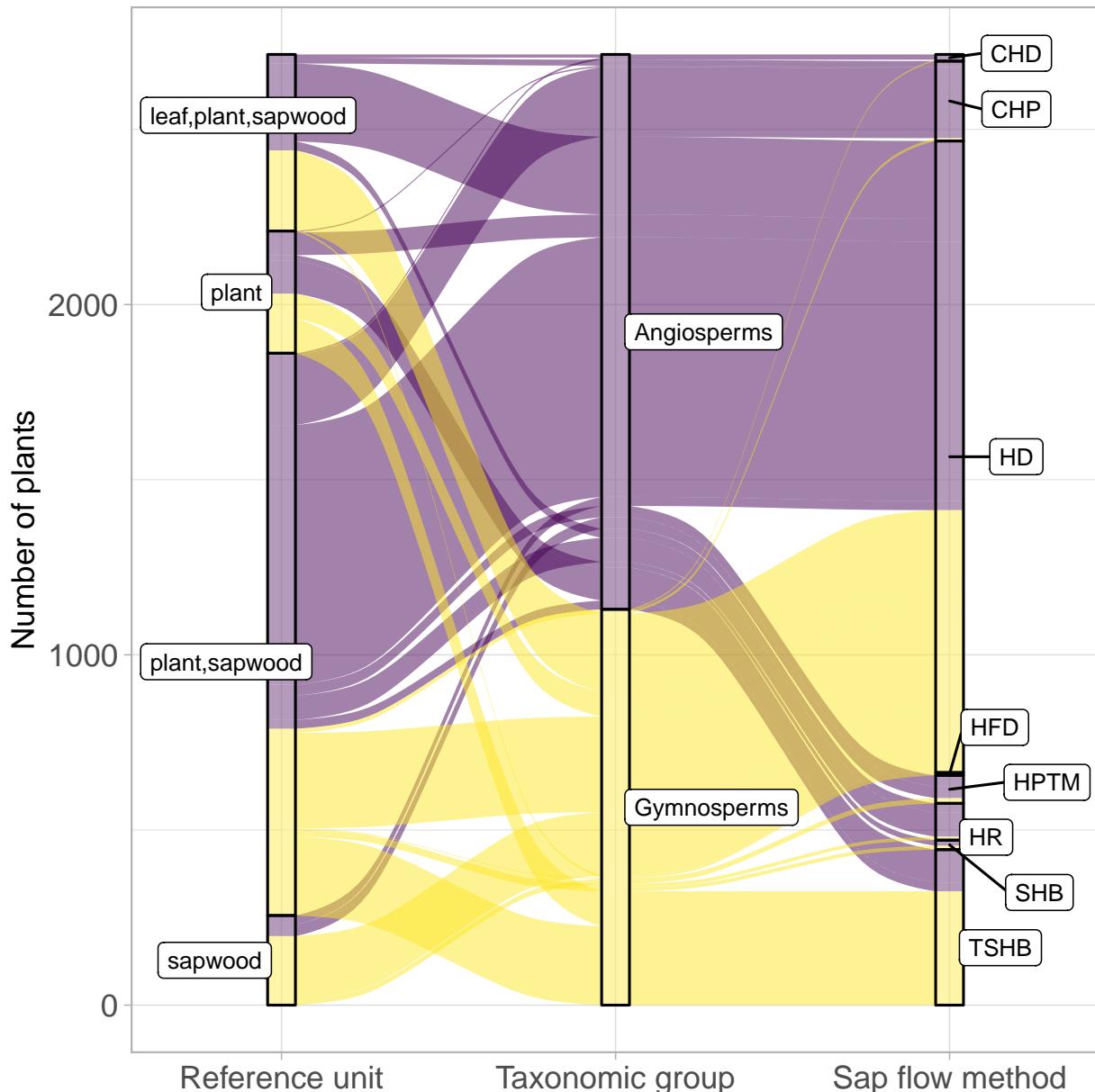


Figure 3.4: Distribution of plants in SAPFLUXNET according to major taxonomic group (angiosperms, gymnosperms), sap flow method (CHD: cycling heat dissipation; CHP: compensation heat pulse; HD: heat dissipation; HFD: heat field deformation; HPTM: heat pulse T-max (HPTM); HR: heat ratio (HR); SHB: stem heat balance; TSHB: trunk sector heat balance) and reference unit for the expression of sap flow (plant, sapwood area, leaf area). Combinations of reference units imply that data are present in multiple units.

may be relevant for the more empirical heat dissipation methods (HD and CHD), which have been shown to consistently underestimate sap flow rates (Steppe *et al.*, 2010; Peters *et al.*, 2018; Flo *et al.*, 2019). Radial integration of single-point sap flow measurements is more frequent than azimuthal integration (Table 3.2), except for the CHD method. A large number of plants using the HD method, and all plants measured using the HPTM method, do not employ any radial integration procedure. In contrast, the CHP, HR, SHB, and TSHB methods are those which more frequently addressed radial variation in one way or another (Table 3.2). Azimuthal integration procedures are also more frequent when the TSHB method is used (Table 3.2).

3.3.3 Plant characteristics

Plant-level metadata is almost complete (99.5% of the individuals) for diameter at breast height (DBH), while sapwood area and sapwood depth, important variables for sap flow upscaling, are not available, or could not be estimated, for 23% and 47% of the plants, respectively. Plant height and plant age are missing for 42% and 62% of the individuals, respectively. Sap flow data in SAPFLUXNET are representative of a broad range of plant sizes (Fig. 3.5(a)). The distribution of DBH showed a median of 25.0 cm and 20.4 cm for gymnosperms and angiosperms, respectively, with a long tail towards the largest plants, two *Mortoniodendron anisophyllum* trees from a tropical forest in Costa Rica that measured > 200 cm (Fig. 3.5(a)). The largest gymnosperm tree in SAPFLUXNET (176 cm in DBH) is a kauri tree (*Agathis australis*) from New Zealand. The distribution of plant heights is less skewed, with similar medians for angiosperms (17.6 m) and gymnosperms (17.5 m). The tallest plants are located in a tropical forest in Indonesia, where a *Pouteria firma* tree reached 44.7 m. Remarkably, of the 16 plants taller than 40 m, over 60% are *Eucalyptus* species. The tallest gymnosperm (36.2 m) is a *Pinus strobus* from NE USA.

Plant size metadata in SAPFLUXNET is complemented with plant-level data of sapwood and leaf area, that provide information on the functional areas for water transport and loss (Fig. 3.5(a)). Distributions of sapwood and leaf area show highly skewed distributions, with long tails towards the largest values and slightly higher median values for gymnosperms (262 cm² and 33.0 m² for sapwood and leaf areas, respectively), compared to angiosperms (168 cm² and 29.9 m²). Accordingly, median sapwood depth is also higher for gymnosperms (5.1 cm) compared to angiosperms (3.7 cm). The largest trees (*Mortoniodendron*, *Pouteria*, *Agathis*) with deep sapwood (17–24 cm) are also those with largest sapwood areas. Many large angiosperm trees from tropical (CRI_TAM_TOW, IDN_PON_STE, GUF_GUY_ST2; see Table B3 for dataset codes) and temperate forests (*Fagus grandifolia*, USA_SMIC_SCB) also show large sapwood areas (> 5000 cm²), but the plant with the deepest sapwood is a gymnosperm, an *Abies pinsapo* in Spain with 30.7 cm of sapwood depth.

3.3.4 Stand characteristics

Stand-level metadata include several variables associated with management, vegetation structure and soil properties. Half of the datasets originate from naturally regenerated, unmanaged stands, and 13.9% come from naturally regenerated but managed stands. Plantations add up to 32.2% and orchards only represent 4% of the datasets. Reporting of structural variables is mixed, with stand height, age, density and basal area showing relatively low missingness (6.4%, 11.4%, 12.9% and 13.4%, respectively); in contrast, soil depth and LAI are missing from 26.7% and 33.7% of the datasets.

SAPFLUXNET datasets originate from stands with diverse structural characteristics. Median stand age is 54 years and there are several datasets coming from >100 year-old forests (Fig. 3.5(b)). Stand height shows a similar range and distribution of values compared to individual plant height (Fig. 3.5(a,b)). The denser stands correspond to coppiced evergreen oak stands from Mediterranean forests (FRA_PUE, ESP_TIL_OAK), species-rich tropical forests (MDG_SEM_TAL) or relatively young temperate forests (e.g. FRA_HES_HE1_NON, USA_CHE_MAP). The sparsest stands (< 200 stems ha⁻¹) correspond to tree-grass savanna

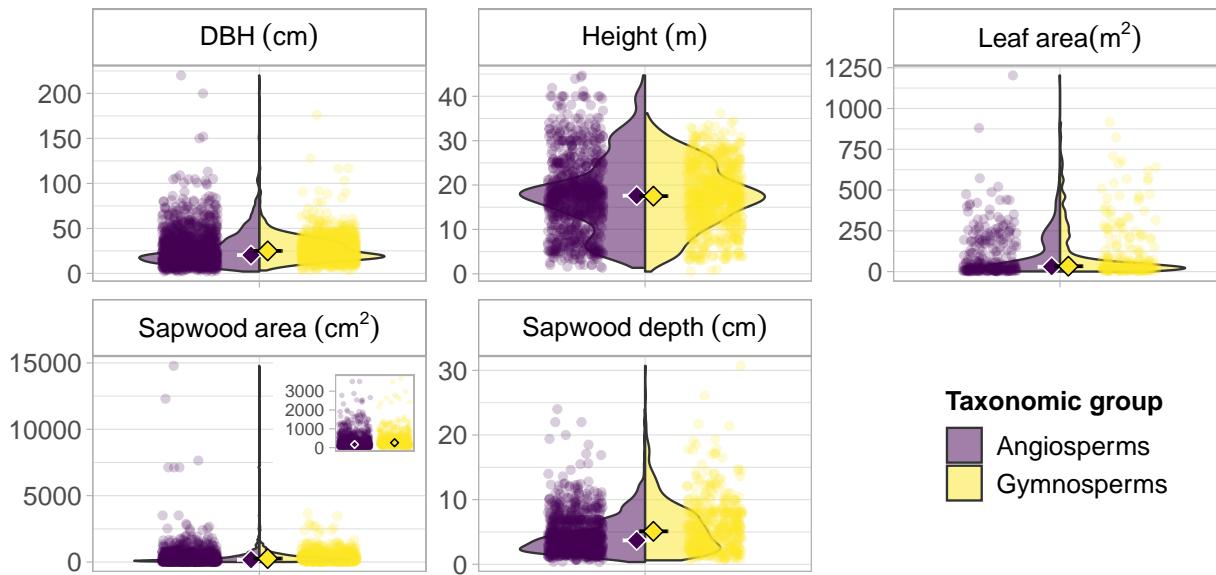
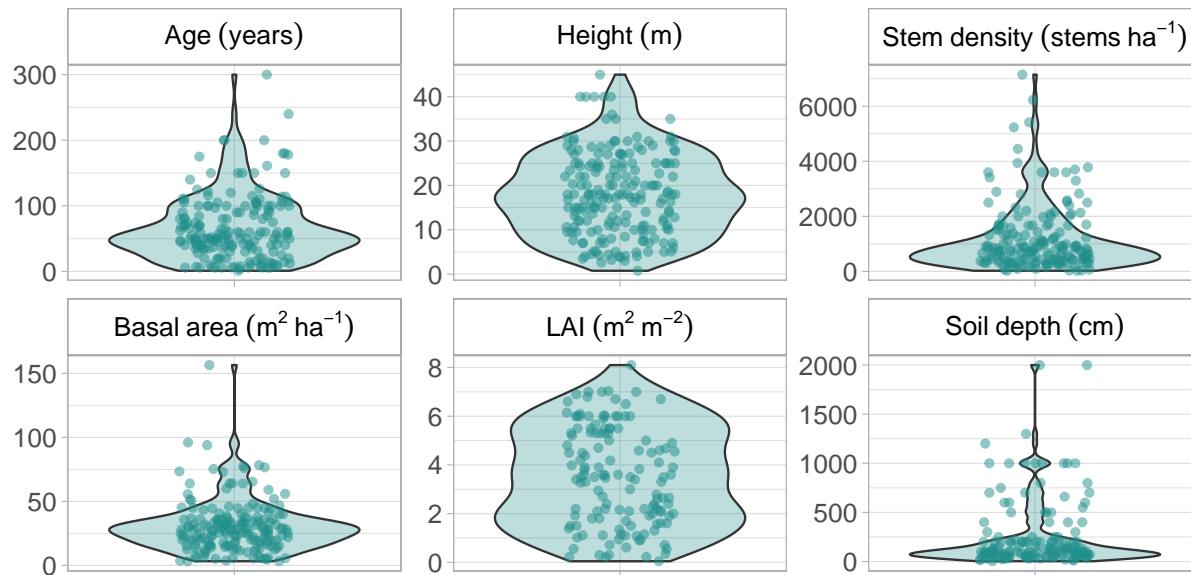
(a) Plant attributes**(b) Stand attributes**

Figure 3.5: Characteristics of trees and stands in the SAPFLUXNET database. Panel (a) shows plant data and kernel density plots of the main plant attributes, coloured by taxonomic group (angiosperms and gymnosperms): diameter at breast height (DBH), plant height, sapwood area, sapwood depth and leaf area. The inset in the sapwood area panel zooms in values lower than 5000 cm^2 . Panel (b) shows stand data and kernel density plots of the main stand attributes: stand age, stand height, stem density, stand basal area, leaf area index (LAI) and soil depth.

systems (Spain, Portugal, Australia, Senegal), dry woodlands (China), or oil palm plantations in Indonesia (IDN_JAM_OIL). Stands with the largest basal areas ($> 70 \text{ m}^2 \text{ ha}^{-1}$) are mostly dominated by broadleaf species, except for a *Picea abies* plantation in Sweden (SWE_SKO_MIN).

The distribution of leaf area index (LAI) shows a median of $3.5 \text{ m}^2 \text{ m}^{-2}$, with the largest values observed in temperate (CZE_BIK, USA_DUK_HAR, HUN_SIK) and tropical (GUF_GUY_GUY, COL_MAC_SAF_RAD) forests. The stands with the lowest LAI correspond to the sparse woodlands from Mediterranean and semi-arid locations and also those from forests near altitudinal or latitudinal tree-lines (FIN_PET, AUT_TSC). SAPFLUXNET datasets

show a median soil depth of 100 cm, with only a dozen datasets originated from sites with soils deeper than 10 m (Fig. 3.5(b)).

The number of plants per dataset is highly variable, with most of the datasets (86%) containing data for at least 4 trees and 46% of the datasets having data for at least 10 trees (Fig. 3.6(a), see also Fig. 3.9).

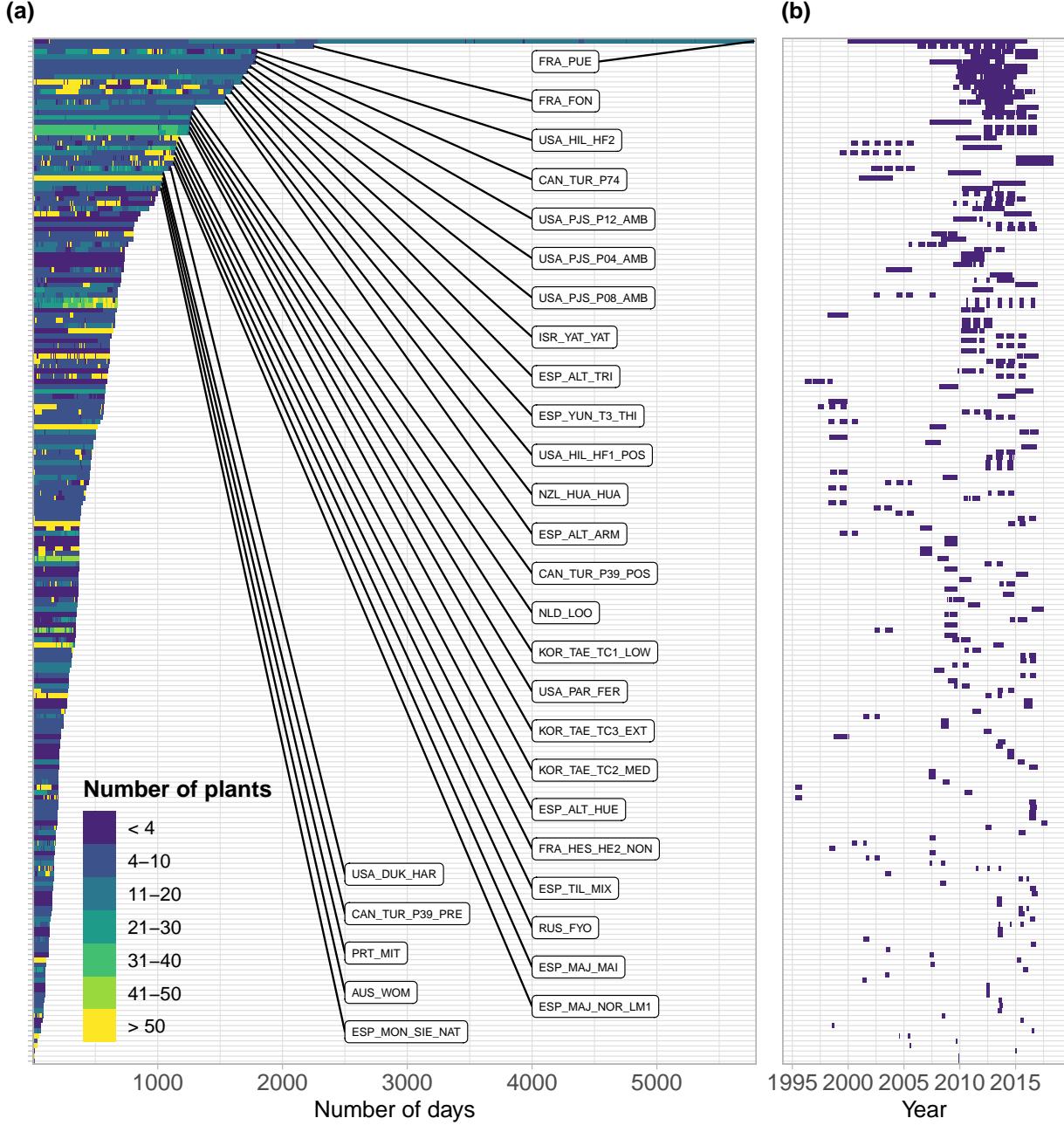


Figure 3.6: (a) Measurement duration of SAPFLUXNET datasets expressed in number of days with sap flow data and coloured by the number of plants measured on each day . The 30 longest datasets are labelled. For each dataset in panel (a), panel (b) shows its corresponding measurement period.

3.3.5 Temporal characteristics

The oldest datasets in SAPFLUXNET go back to 1995 (GBR_DEV_CON, GBR_DEV_DRO) while the most recent data reach up to 2018 (datasets from the ESP_MAJ cluster of sites). Several multi-year datasets are present in SAPFLUXNET (Fig. 3.6), with 50% of the datasets spanning a

period of at least 3 years, and some datasets being extraordinarily long (16 years in FRA_PUE). Frequently, the datasets only cover the ‘growing season’ periods, or even shorter periods for some sites which were eventually included because they improved the ecological and geographic coverage of the database (e.g. ARG_MAZ, ARG_TRE as representative of deciduous *Nothofagus* forest in South Patagonia). In contrast, a few datasets show continuous records over multiple years (Fig. 3.6(b)). Amongst the longest datasets, most of them come from European or North American sites (Fig. 3.6), except some datasets from Israel (ISR_YAT_YAT, 7 years), Russia (RUS_FYO, 7 years), South Korea (KOR_TAE cluster of sites, 6 years) or New Zealand (NZL_HUA_HUA, 5 years).

SAPFLUXNET provides an unprecedented database to study the detailed temporal dynamics of plant transpiration across species and sites globally. Sub-daily records of sap flow (e.g. at least at hourly timesteps) are available for extended periods (Fig. 3.6(b)), allowing to address both seasonal and diel patterns in water use regulation by trees and how these temporal patterns change across species or years across terrestrial biomes, reflecting different phenologies and water-use strategies. For instance, in Mediterranean forests, evergreen species such as *Quercus ilex*, *Arbutus unedo* and *Pinus halepensis* show moderate sap flow the whole year round, while the deciduous *Quercus pubescens* shows higher sap flow density during a shorter period and its water use is heavily reduced during a dry year (2012) (Fig. 3.7(a)). Temperate forests without water availability limitations show relatively high flows during the growing season and similar diel sap flow patterns among species (Fig. 3.7(b)). In contrast, tropical forests show moderate to high sap flow rates during the entire year, with different dynamics in the intradaily water use regulation across species. For example, *Inga* sp. in a highly diverse wet tropical forest in Costa Rica, reduced sap flow during mid-day hours compared to co-existing species (Fig. 3.7(c)).

3.3.6 Availability of environmental data

All SAPFLUXNET datasets contain ancillary time series of the main hydrometeorological drivers of transpiration, accompanied by information on where these variables had been measured (Fig. 3.8(a)). Air temperature is available for all datasets. Although vapour pressure deficit (VPD) was originally absent in 38% of the datasets (Fig. 3.8(a,b)), we could estimate it for those sites providing air temperature and relative humidity data (QC Level 2, see section 3.2.3), and finally only 2 out of the 202 datasets have missing VPD information. For radiation variables, shortwave radiation was most often provided, compared to photosynthetically active and net radiation; only 8 out of 202 datasets do not have any accompanying radiation data. Most of these environmental variables were measured on-site, with precipitation being the variable most frequently retrieved from nearby meteorological stations (48% of the datasets) (Fig. 3.8(a)). Soil water content measured at shallow depth, typically between 0 and 30 cm below the soil surface, is provided for 56% of the datasets, while soil moisture from deep soil layers is available for only 27% of the datasets.

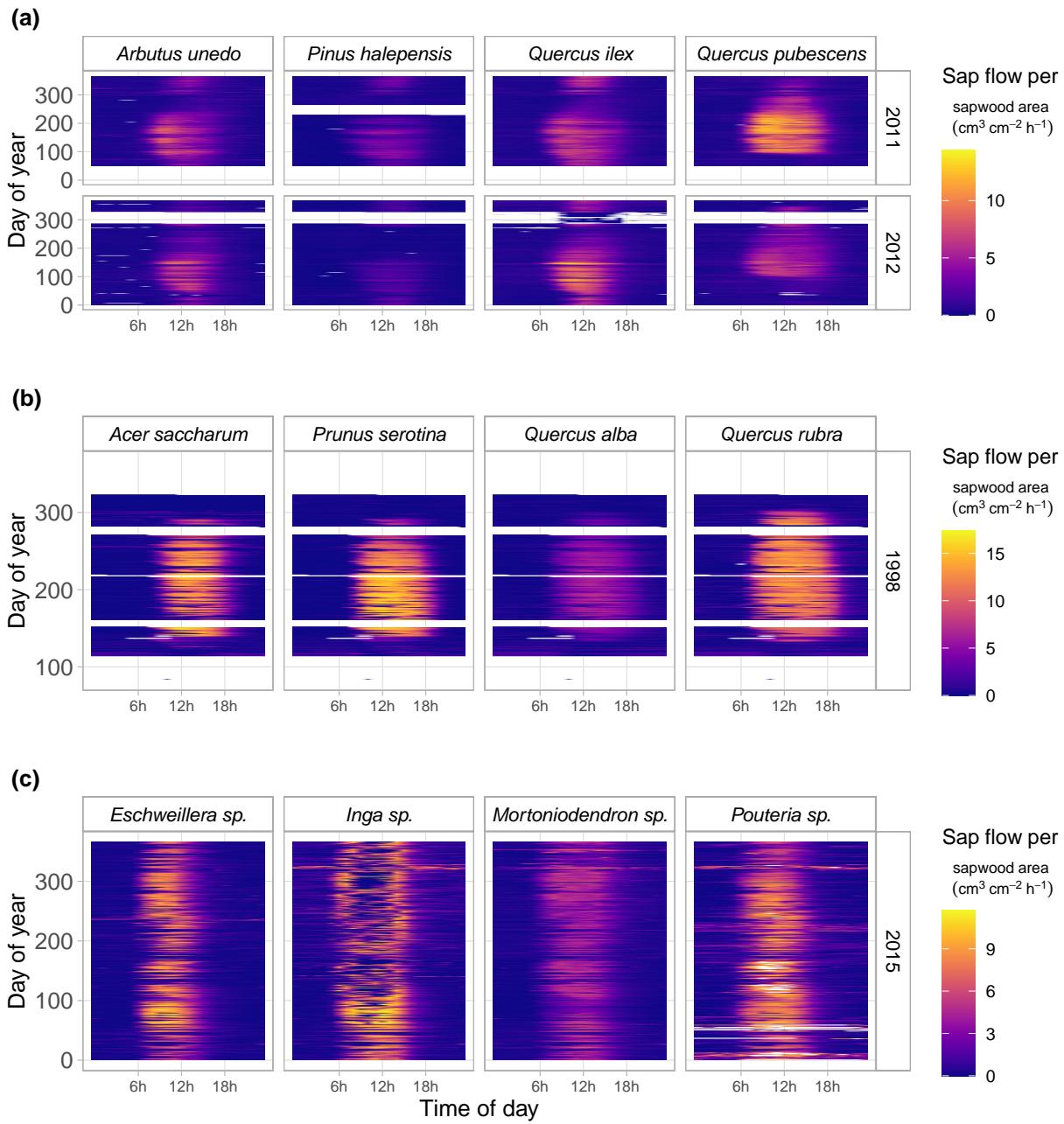


Figure 3.7: Fingerprint plots showing hourly sap flow per unit sapwood area (colour scale) as a function of hour of day (x-axis) and day of year (y-axis) for a selection of SAPFLUXNET sites with at least four co-occurring species. Panel (a) shows data from a Woodland/Shrubland forest in NE Spain (ESP_CAN), for an average (2011) and a dry (2012) year. Panel (b) shows data for a mesic Temperate forest (USA_WVF) and panel (c) shows data for a Tropical forest (CRI_TAM_TOW). For this latter site, only 4 of the 17 measured species are shown and some of them were only identified at the genus level.

3.4 Potential applications

3.4.1 Applications in plant ecophysiology and functional ecology

There are multiple potential applications of the SAPFLUXNET database to assess whole-plant water use rates and their environmental sensitivity, both across species (e.g. Oren *et al.*, 1999b) and at the intraspecific level (Poyatos *et al.*, 2007). SAPFLUXNET will allow disentangling the roles of evaporative demand and soil water content in controlling transpiration at the plant level,

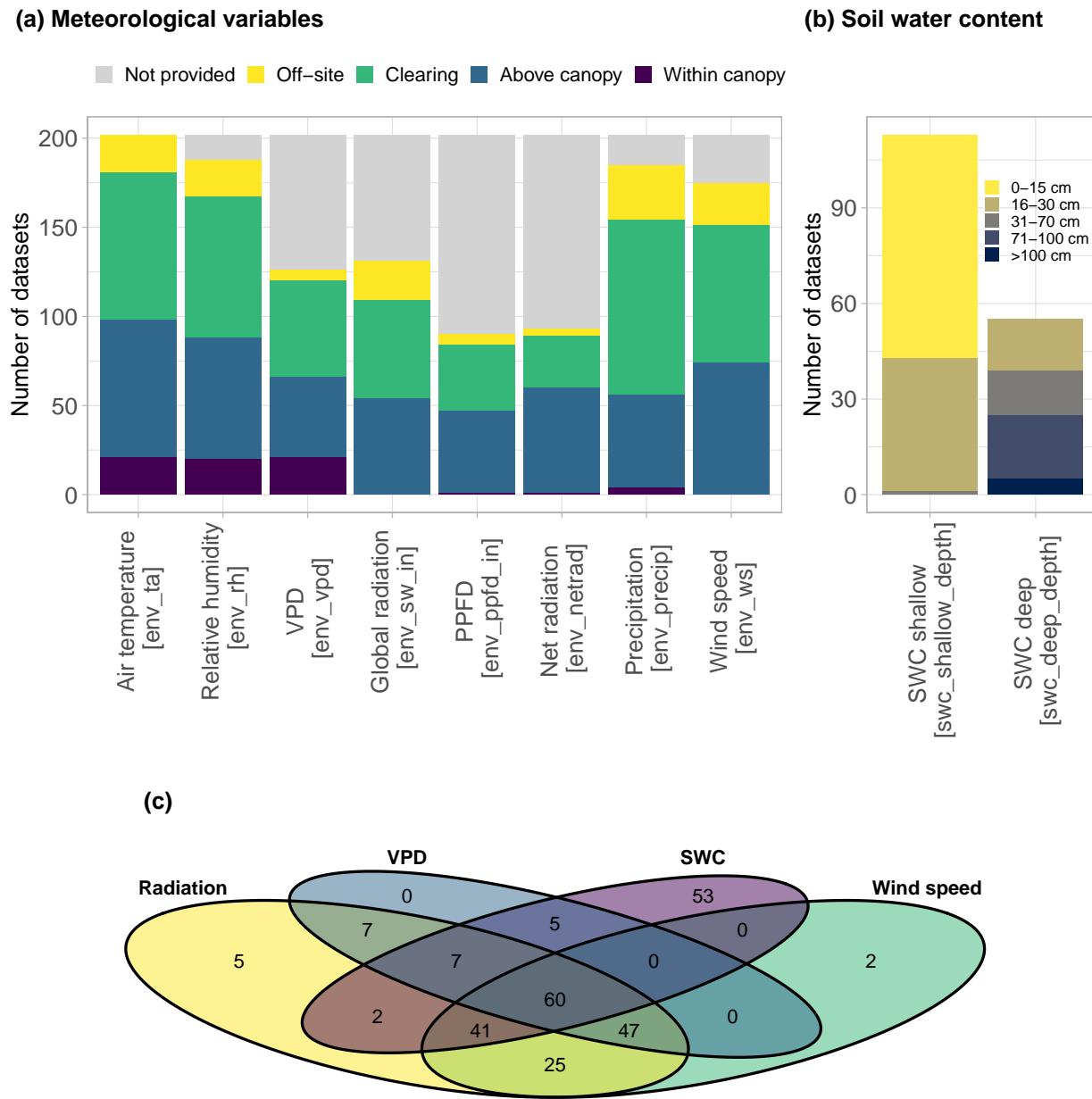


Figure 3.8: Summary of the availability of different environmental variables in SAPFLUXNET datasets. (a) Distribution of meteorological variables according to sensor location (in brackets, names of the variables in the database), (b) Distribution of soil moisture variables according to the measurement depth (in brackets, names of the variables in the database). (c) Venn diagram showing the number of datasets where each combination of different environmental variables are present, grouping shortwave, PPFD and net radiation under ‘Radiation’ variables.

complementing recent studies looking at how water supply and demand affect evapotranspiration at the ecosystem level (Novick *et al.*, 2016; Anderegg *et al.*, 2018). The availability of global sap flow data at sub-daily time resolution and spanning entire growing seasons will allow focusing on how maximum water use and its environmental sensitivity varies with plant-level attributes such as stem diameter (Meinzer *et al.*, 2005; Dierick & Hölscher, 2009), tree height (Schäfer *et al.*, 2000; Novick *et al.*, 2009), hydraulic (Poyatos *et al.*, 2007; Manzoni *et al.*, 2013) and other plant traits (Kallarackal *et al.*, 2013; Grossiord *et al.*, 2019). SAPFLUXNET thus provides an unprecedented tool to understand how structural and physiological traits scale-up to whole-plant regulation of

water fluxes (McCulloh *et al.*, 2019), and how this integration determines drought responses (Choat *et al.*, 2018) and post-drought recovery patterns (Yin & Bauerle, 2017). Analyses of the temporal dynamics of plant water use in response to specific drought events, as recently assessed for gross primary productivity (e.g. Schwalm *et al.*, 2017), can also help to quantify drought legacy effects, including the reversibility of drought-induced losses of hydraulic conductivity at the plant level.

SAPFLUXNET will allow new insights into within-day patterns and controls in whole-plant water use, which can disclose the fine details of its physiological regulation. Circadian rhythms can modulate stomatal responses to the environment, potentially affecting sap flow dynamics (e.g. Resco de Dios *et al.*, 2015). Hysteresis in diel sap flow relationships with evaporative demand and time-lags between evaporative demand and sap flow, are two linked phenomena likely arising from plant capacitance and other mechanisms (Schulze *et al.*, 1985; O'Brien *et al.*, 2004), that also influence diel evapotranspiration dynamics (Matheny *et al.*, 2014; Zhang *et al.*, 2014). A major driver of time-lags is the use of stored water to meet the transpiration demand (Phillips *et al.*, 2009), which can now be analysed across species, plant sizes or drought conditions using time series analyses, simplified electric analogies (Phillips *et al.*, 1997, 2004; Ward *et al.*, 2013) or detailed water transport models (Bohrer *et al.*, 2005; Mirfenderesgi *et al.*, 2016). Night-time water use can be substantial for some species (Forster, 2014; Resco de Dios *et al.*, 2019). However, available syntheses rely on study-specific quantification of what constitutes nocturnal sap flow and do not address possible methodological influences (Zeppel *et al.*, 2014). SAPFLUXNET will allow applying a consistent estimation of nocturnal sap flow and control for datasets that are less suitable for the quantification of night-time fluxes, as information on zero-flow determination is included in the metadata ('pl_sens_cor_zero', Table B5).

Table 3.3: Number of datasets, plants and species by stand-level treatment in the SAPFLUXNET database.

| Treatment | N sites | N plants | N species |
|--|---------|----------|-----------|
| None/control | 155 | 2198 | 170 |
| Thinning | 18 | 332 | 18 |
| Irrigation | 9 | 36 | 4 |
| Post-fire | 6 | 18 | 4 |
| CO2 fertilisation | 3 | 28 | 2 |
| Drought | 3 | 9 | 2 |
| Soil fertilisation | 2 | 16 | 2 |
| Post-mortality | 1 | 22 | 5 |
| Soil fertilisation and pruning | 1 | 12 | 1 |
| Soil fertilisation and thinning | 1 | 12 | 1 |
| Pruning and thinning | 1 | 11 | 1 |
| Soil fertilisation, pruning and thinning | 1 | 11 | 1 |
| Pruning | 1 | 9 | 1 |

Percentage of stand basal area with sap flow measurements

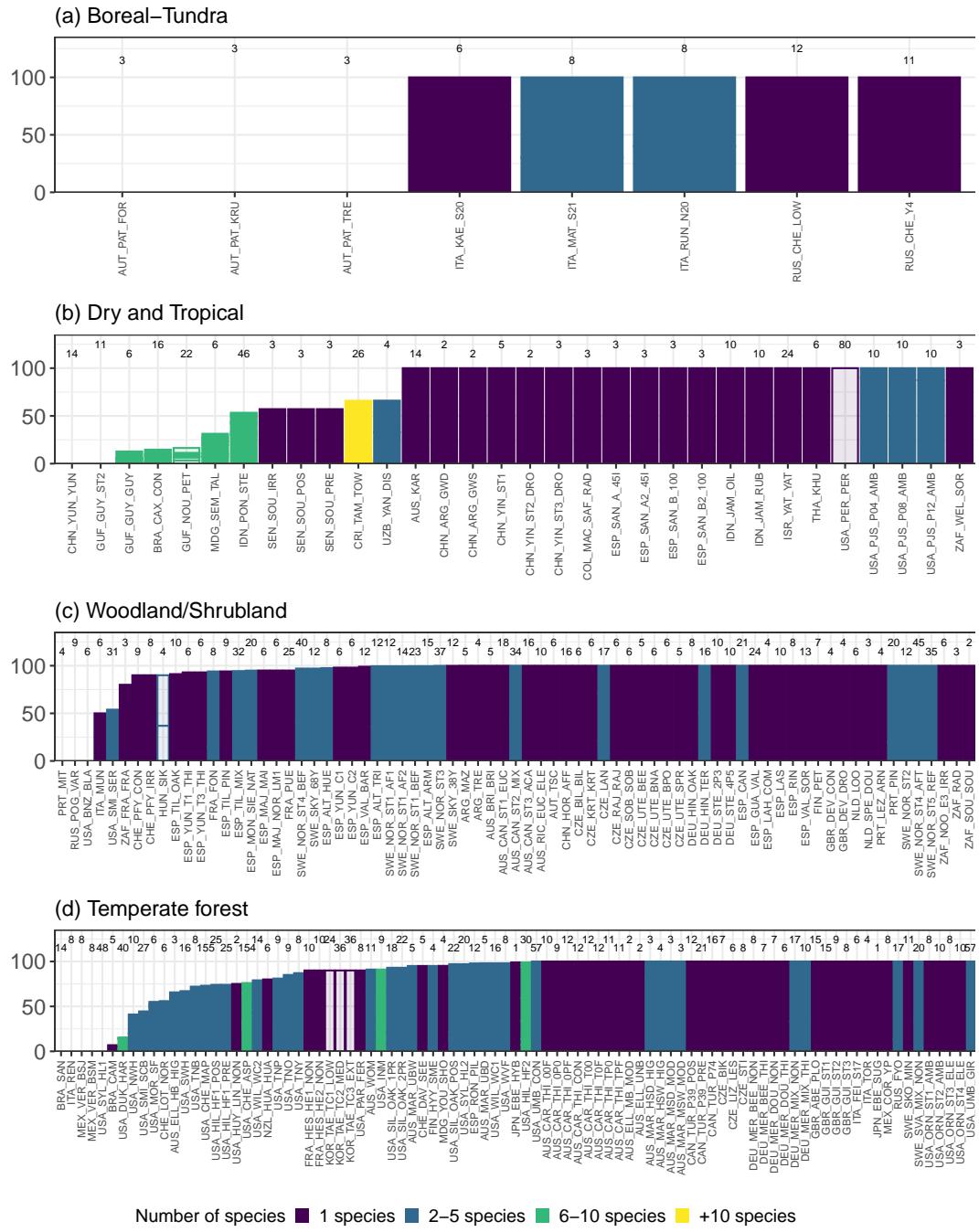


Figure 3.9: Potential for upscaling species-specific plant sap flow to stand-level sap flow using SAPFLUXNET datasets. Datasets are shown using an aggregated biome classification; ‘Dry and Tropical’ include: ‘Subtropical desert’, ‘Temperate grassland desert’, ‘Tropical forest savanna’ and ‘Tropical rain forest’. Each panel shows the percentage of total stand basal area that is covered by sap flow measurements for each species in the dataset. Datasets are also coloured by the number of species present. Numbers on top of each bar depict the total number of plants for a given dataset. Empty bars show datasets for which sap flow data expressed at the plant level were not available.

Sap flow data have been widely employed to assess changes in tree water use after biotic (e.g. Hultine *et al.*, 2010) or abiotic (Oren *et al.*, 1999a) disturbances. Likewise, sap flow data have been used to report changes in species and stand water use following experimental treatments involving resource availability modifications (e.g. Ewers *et al.*, 1999) or density changes (i.e. thinning, Simonin *et al.*, 2007). The SAPFLUXNET database includes datasets with experimental manipulations, applied either at the stand or at the individual level (Table 3.3). The main treatments

present are related to thinning, water availability changes (irrigation, throughfall exclusion) and wildfire impact (Table 3.3), potentially facilitating new data syntheses and meta-analyses using these datasets (e.g. Grossiord *et al.*, 2018).

The combination of SAPFLUXNET with other ecophysiological databases can inform on the relative sensitivity of different physiological processes in response to drought, for example those related to growth and carbon assimilation (Steppe *et al.*, 2015). Within-day fluctuations of stem diameter can be jointly analysed with co-located sap flow measurements to study the dynamics of stored water use under drought and its contribution to transpiration (e.g. Brinkmann *et al.*, 2016), and to infer parameters on tree hydraulic functioning using mechanistic models of tree hydrodynamics (Steppe *et al.*, 2006; Zweifel *et al.*, 2007; Salomón *et al.*, 2017). These analyses could be carried out for a large number of species by combining SAPFLUXNET with data from the Dendroglobal database (<http://78.90.202.92/streess/databases/dendroglobal>); there are at least 18 SAPFLUXNET datasets with dendrometer data in Dendroglobal. This database and the International Tree-Ring Data Bank (Zhao *et al.*, 2019b) could also be used with SAPFLUXNET to investigate, at the species level, the link between radial growth and water use, including their environmental sensitivity (Morán-López *et al.*, 2014), and how these two processes comparatively respond to drought (Sánchez-Costa *et al.*, 2015). Moreover, given the tight link between water use and carbon assimilation, combining SAPFLUXNET with water-use efficiency from plant $\delta^{13}C$ data could potentially be used to estimate whole-plant carbon assimilation (Hu *et al.*, 2010; Rascher *et al.*, 2010; Klein *et al.*, 2016; Vernay *et al.*, 2020), a quantity that is difficult to measure directly, especially in field-grown, mature trees.

3.4.2 Applications in ecosystem ecology and ecohydrology

SAPFLUXNET will provide a global look at plant water flows to bridge the scales between plant traits and ecosystem fluxes and properties (Reichstein *et al.*, 2014). Vegetation structure, species composition and differential water use strategies among and within species scale-up to different seasonal patterns of ecosystem transpiration, with a strong influence on ecosystem evapotranspiration and its partitioning. Global controls on evaporative fluxes from vegetation have been mostly addressed using ecosystem (Williams *et al.*, 2012) or catchment evapotranspiration data (Peel *et al.*, 2010). These studies have described global patterns in evapotranspiration driven by different plant functional types or climates, but they cannot be used to quantify and to explain the enormous variation in the regulation of transpiration across and within taxa.

The SAPFLUXNET database will provide a long-demanded data source to be used in eco-hydrological research (Asbjørnsen *et al.*, 2011). Upscaling individual measurements to the stand level (Granier *et al.*, 1996; Köstner *et al.*, 1998; Čermák *et al.*, 2004) is necessary to quantitatively compare sap-flow based transpiration with evapotranspiration and transpiration estimates at the ecosystem scale and beyond. Even though SAPFLUXNET was designed to accommodate sap flow data at the plant level, scaling to the ecosystem level is possible for many datasets. For a basic upscaling exercise using SAPFLUXNET data (Poyatos *et al.*, 2020a), whole-plant sap flow can be normalised by individual basal area (as DBH is usually available in the metadata, cf. section 3.3.3), averaged for a given species and then scaled to stand level transpiration using total stand basal area and the fraction of basal area occupied by each measured species (see stand metadata,

Table B3). For many datasets, sap flow data are available for the species comprising most of the stand basal area (often even 100%, Fig. 3.9), but species-based upscaling may be unfeasible in many tropical sites (Fig. 3.9(b)), where size-based scaling could be applied instead (e.g. Costa *et al.*, 2018). Further refinements of the upscaling procedure could be achieved by using trunk diameter distributions of the sap flow plots (Berry *et al.*, 2018). This information, however, is not readily available in SAPFLUXNET, and other data sources (e.g. forest inventories, LIDAR data) or additional simplifying assumptions (i.e. applying the size distribution of measured individuals in the dataset) would be needed.

Stand-level transpiration estimates from a large number of SAPFLUXNET sites can contribute to improve our understanding of the role of forest transpiration in the context of stand water balance and its components at the ecosystem (e.g. Tor-ngern *et al.*, 2018) and catchment levels (Wilson *et al.*, 2001; Oishi *et al.*, 2010). Importantly, SAPFLUXNET can contribute to better understand the global controls on vegetation water use (Good *et al.*, 2017), including the biological and climatic controls on evapotranspiration partitioning into transpiration and evaporation components (Schlesinger & Jasechko, 2014; Stoy *et al.*, 2019). There is some overlap between the FLUXNET network and SAPFLUXNET (47 datasets from FLUXNET sites). Hence, transpiration from SAPFLUXNET can also be used as a ‘ground-truth’ reference for transpiration estimates from remote sensing approaches (Talsma *et al.*, 2018) and from eddy covariance data (Nelson *et al.*, 2020). Extrapolating sap flow-derived stand transpiration to large spatial scales can be challenging due to landscape-scale variation in forest structure (Ford *et al.*, 2007) or topography (Hassler *et al.*, 2018), and to the low spatial representativeness of sap flow measurements (Mackay *et al.*, 2010). A promising research avenue to help elucidate the role of vegetation in driving hydrological changes across environmental gradients (Vose *et al.*, 2016) would be to combine species-specific stand transpiration data from SAPFLUXNET with stand structural and compositional data from forest inventories (e.g. sapwood area index, Benyon *et al.*, 2015).

Understanding the patterns and mechanisms underlying species interactions with respect to water use within a community is necessary to predict tree species vulnerability to drought (Grossiord *et al.*, 2019). Multispecies datasets from SAPFLUXNET (Table B4) can be used to assess competition for water resources among species, for example by identifying changes in seasonal water use across co-existing species and hence characterizing the spatiotemporal segregation of their hydrological niches (Silvertown *et al.*, 2015). By providing a detailed seasonal quantification of tree water use, SAPFLUXNET could also complement isotope-based studies and contribute to interpret the large diversity in root water uptake patterns observed worldwide (Evaristo & McDonnell, 2017; Barbeta & Peñuelas, 2017) and to explain the different seasonal origin of root-absorbed water across species and environmental gradients (Allen *et al.*, 2019).

Plant water fluxes and hydrodynamics are amongst the most uncertain components of ecosystem and terrestrial biosphere models (Fatichi *et al.*, 2016; Fisher *et al.*, 2018). These models are now incorporating hydraulic traits and processes in their transpiration regulation algorithms (Menuccini *et al.*, 2019a), but multi-site assessments of these algorithms are usually performed against evapotranspiration from eddy flux data (Matheny *et al.*, 2014; Knauer *et al.*, 2015). Model validation against sap flow data has been carried out typically in only one (Williams *et al.*, 2001; Kennedy *et al.*, 2019) or few (Buckley *et al.*, 2012) sites. SAPFLUXNET can thus contribute to

assess the performance of models simulating transpiration of stands or species within stands (e.g. de Cáceres *et al.*, 2021), for a large number of species and under diverse climatic conditions.

3.5 Limitations and future developments

3.5.1 Limitations

Sap flow data processing differs within and among methods, because different algorithms, calibrations or parameters involved in sap flow calculations may be applied. All of these methods contribute to methodological uncertainty (Looker *et al.*, 2016; Peters *et al.*, 2018) and this challenging methodological variability precludes the implementation of a complete, standardised data workflow from raw to processed data within SAPFLUXNET, as it is done for eddy flux data (Wutzler *et al.*, 2018; Vitale *et al.*, 2020). Commercial software for sap flow data processing from multiple methods is available (i.e. <http://www.sapflowtool.com/SapFlowToolSensors.html>) but it has not yet been widely adopted. Freely available data-processing software is only available for the HD method (Oishi *et al.*, 2016; Ward *et al.*, 2017; Speckman *et al.*, 2020).

Sap flow measured with thermometric methods provides a precise estimate of the temporal dynamics of water flow through plants (Flo *et al.*, 2019). However, their performance in measuring absolute flows is mixed. While some well-represented methods in SAPFLUXNET such as the CHP yield accurate estimates (at least for moderate-to-high flows), the HD method, the most represented method by far, can significantly underestimate water flows (Flo *et al.*, 2019). Because plant-level metadata contain information that document the conversion from raw to processed data (Table B5), a first-order correction for uncalibrated HD measurements based on available methodological assessments can be applied to allow intercomparability across methods. Nevertheless, given the high unexplained variability (i.e. by species and wood traits) in the performance of sap flow calibrations (Flo *et al.*, 2019), these corrections should be applied with caution. The determination of zero flow conditions (baselining) can also have significant impacts on the quantification of absolute flow for several methods (Smith & Allen, 1996; Steppe *et al.*, 2010; Peters *et al.*, 2018). The different baselining approaches are also documented in the metadata to inform data syntheses and/or to selectively apply correction factors.

SAPFLUXNET has been designed to store whole-plant sap flow data, and therefore, sap flow measured at multiple points within an individual is not available in the database. Even though this spatial variation could be useful to describe detailed aspects of plant water transport (Nadezhina *et al.*, 2009), focusing on plant-level data greatly simplifies the data structure. Hence, SAPFLUXNET only includes data already upscaled to the plant level by the data contributors. The main details of how this upscaling process was done for each dataset are provided together with other plant metadata (Table B5), but these metadata show that within-plant variation in sap flow is often not considered (Table 3.2). The impact of not accounting for radial and circumferential variability when scaling single-point measurements of sap flow to the whole-plant level can be important (Merlin *et al.*, 2020), but the estimation of sapwood area can also cause large errors (Looker *et al.*, 2016). SAPFLUXNET does not provide information on the method employed to quantify sapwood area (e.g. visual estimation with or without the application of dyes, indirect estimation through allometries at species or site levels) or on the accuracy of sapwood

area data. This precludes uncertainty estimation at the individual level. Future developments in the SAPFLUXNET data structure could include this information as metadata to better document the sensor-to-plant scaling process.

While SAPFLUXNET makes global sap flow data available for the first time, we note that spatial coverage is still sparse and some forested regions are underrepresented in the database (Fig. 3.2(a)). We note especially the relatively small number of datasets for boreal and tropical forests, two important biomes in terms of global water and carbon fluxes (Beer *et al.*, 2010; Schlesinger & Jasechko, 2014). While many geographic gaps are caused by the absence of sap flow studies from such areas, some regions where sap flow studies have been conducted are still not represented in SAPFLUXNET. For example, the recent proliferation of Asian sap flow studies (Peters *et al.*, 2018) has not translated into a high representativity of Asian datasets in SAPFLUXNET yet. Similarly, while the coverage of taxonomic and biometric diversity is unprecedented, SAPFLUXNET lacks data for the extremely tall trees (Ambrose *et al.*, 2010) or for other growth forms such as shrubs (Liu *et al.*, 2011), lianas (Chen *et al.*, 2015) and other non-woody species (Lu, 2002).

3.5.2 Outlook

The public release of SAPFLUXNET has set the stage for a first generation of sap flow-based data syntheses. The work on these syntheses will fuel new ideas and tools for future improvements of the database, as for example new computing approaches for the processing and analysis of sap flow datasets. One example would be the development of robust imputation algorithms to gap-fill time series of sap flow and environmental data, which can take advantage of tools and datasets already developed by the ecosystem flux community (Moffat *et al.*, 2007; Vuichard & Papale, 2015). The dissemination of SAPFLUXNET will encourage the use of machine-learning algorithms, only occasionally used to analyse sap flow datasets so far (e.g. Whitley *et al.*, 2013). These approaches can also be used to identify the relative importance of different hydrometeorological drivers of transpiration (Zhao *et al.*, 2019b), or to produce global transpiration maps, by combining SAPFLUXNET with other data (Jung *et al.*, 2019). This upscaling of stand transpiration to large areas will also allow addressing broader questions at the regional and continental scale, such as the role of transpiration in moisture recycling (Staal *et al.*, 2018).

The eventual success of this initiative, in terms of enabling data reuse, contributing towards the understanding and modelling of tree water use at local to global scales will likely encourage the sap flow community to contribute new datasets to future updates of the database. We expect that the development of open-source software for the processing of sap flow raw data (Speckman *et al.*, 2020), its eventual widespread use by the sap flow community and the adoption of standardized calibration practices will increase the quality and intercomparability of future sap flow datasets. These new datasets will hopefully expand the temporal, geographical and ecological representativity of SAPFLUXNET when new data contribution periods can be opened in the future.

3.6 Data availability, access and feedback

In this paper we present SAPFLUXNET version 0.1.5 (Poyatos *et al.*, 2020c), which contains some small metadata improvements on version 0.1.4, the first one to be made publicly available, in March

2020. Both versions supersede version 0.1.3 which was initially released to data contributors in March 2019. The entire database can be downloaded from its hosting webpage in the Zenodo repository (<https://doi.org/10.5281/zenodo.3971689>, Poyatos *et al.*, 2020c). In this repository, we provide the database as separate .csv files and as .RData objects; see section 3.2.4. for details on data structure. Together with the initial publication of SAPFLUXNET in March 2019, we also released the sapfluxnet R package, available on CRAN, to enable easy access, selection, temporal aggregation and visualisation of SAPFLUXNET data. Feedback on data quality issues can be forwarded to the SAPFLUXNET initiative email address: `sapfluxnet@creaf.uab.cat`. All the information about SAPFLUXNET, including the publication of new calls for data contribution, can be found in the project website: <http://sapfluxnet.creaf.cat/.par>

3.7 Conclusion

The SAPFLUXNET database provides the first global perspective of water use by individual plants at multiple timescales, with important applications in multiple fields, ranging from plant ecophysiology to Earth-system science. This database has been built from community-contributed datasets and is complemented with a software package to facilitate data access. Both the database and the software have been implemented following open science practices, ensuring public access and reproducibility. Data sharing has been a key component of the success of the FLUXNET network of ecosystem fluxes (Bond-Lamberty, 2018), and many databases in plant and ecosystem ecology now offer open data (Bond-Lamberty & Thomson, 2010; Falster *et al.*, 2015; Kattge *et al.*, 2020; Gallagher *et al.*, 2020). SAPFLUXNET fully aligns with this philosophy. We expect that this initial data infrastructure will promote data sharing among the sap flow community in the future (Dai *et al.*, 2018) and will allow the continued growth of the SAPFLUXNET database.

4

Analysis of hydrometeorological drivers of tree transpiration shows a dominant role of vapour pressure deficit across biomes.

Víctor Flo, Jordi Martínez-Vilalta, Víctor Granda, Maurizio Mencuccini, Rafael Poyatos

Abstract

We aim to identify the relative importance of vapour pressure deficit (VPD), soil water content (SWC) and photosynthetic photon flux density (PPFD) as drivers of tree transpiration, which will allow to improve mathematical models describing vegetation responses under climate change. We use sap flow time series of 1858 trees in 122 sites from the SAPFLUXNET global database to obtain whole-tree canopy conductance (G). The coupling, defined as the percentage of variance (R^2) of G explained by the three main hydrometeorological drivers (VPD, SWC and PPFD), was evaluated using linear mixed models. For each hydrometeorological driver we assess differences in coupling among biomes, and use multivariate models to explain R^2 by climate, soil and vegetation structure. We found that in most areas transpiration is better explained by VPD than by SWC or PPFD. We also found that sites in dryland biomes are less coupled to all three hydrometeorological drivers than those in other biomes. Climate, soil and vegetation structure were common controls of all three hydrometeorological couplings with G , with wetter climates, fine textured soils and tall vegetation being associated to tighter coupling. Differences across sites in the hydrometeorological coupling of tree transpiration may affect predictions of ecosystem and vegetation dynamics under future climates, and should be accounted for explicitly in models.

4.1 Introduction

Plants regulate transpiration in response to variation in hydrometeorological conditions. However, despite decades of ecophysiological research measuring responses of leaf, plant or ecosystem evaporative fluxes to atmospheric dryness, soil moisture and radiation (Beerling, 2015), the relative importance of these drivers in determining plant controls on transpiration at the global scale is still poorly known. It is important to disentangle the biogeographical patterns of the individual dominant drivers of transpiration control, as such drivers are expected to show spatially heterogeneous dynamics with global change (Zhou *et al.*, 2019b). Thus, understanding their separate roles may help improve models to anticipate climate change impacts on vegetation function and on global water and carbon cycles, and to disentangle land-atmosphere feedbacks (Massmann *et al.*, 2019).

Conductance to water vapour (G) derived from leaf, plant or ecosystem evaporative fluxes has been frequently used to describe the dynamic control of transpiration by plants at different organisational and temporal scales (Jarvis & McNaughton, 1986). At short timescales, this control is usually attributed to the regulation of stomatal aperture. Under high atmospheric water demand, which is often assessed using atmospheric vapour pressure deficit (VPD), or low soil water content (SWC), plants respond constraining G to avoid dangerous declines in water potentials preventing physiological damage and severe dehydration (Oren *et al.*, 1999b). In contrast, G responses to light (i.e. photosynthetic photon flux density, PPF) are linked to plant water use efficiency (WUE). Thus, plants would increase G following PPF in order to optimize photosynthesis in relation to water loss (Sperry *et al.*, 2016). In addition, PPF effects on G may respond to the need to regulate leaf temperature under high radiation levels (Fauset *et al.*, 2018). These responses have been assessed in multiple, single-site studies (Jarvis, 1976; Oren *et al.*, 1999b; Wang *et al.*, 2020). However, the fact that these studies frequently used different phenomenological models and model-fitting approaches complicates synthesis efforts aimed at building a common understanding on the controls of G at broad spatial scales. In addition, most of these studies focused on overall G sensitivity (e.g. Hoshika *et al.*, 2018), not on the importance of the individual drivers (but see for instance Bretfeld *et al.*, 2018), hampering our understanding of which hydrometeorological drivers dominate G regulation globally.

Large-scale syntheses of the relative importance of hydrometeorological drivers regulating transpiration have been conducted using ecosystem evapotranspiration data. Novick *et al.* (2016) compared the limiting effect of SWC and VPD across vegetation types and climates, and found that limitation on ecosystem surface conductance to water vapour caused by SWC increased with climatic dryness, but that VPD limitation was higher than SWC across most mesic biomes. Conversely, Zhao *et al.* (2019a) identified that, globally, ecosystem evapotranspiration was not primarily limited by hydrometeorological drivers, but by vegetation height, followed by SWC and PPF. However, some of the assumptions underlying these studies, that assimilate evapotranspiration to transpiration after data filtering, may not always hold (Nelson *et al.*, 2020). Here, we overcome the limitations of ecosystem-scale approaches by taking advantage of the first global database of plant-level transpiration from sap flow measurements (Poyatos *et al.*, 2020b).

In this study, we investigate the hydrometeorological coupling of tree-level canopy conductance by quantifying the explanatory power (R^2) of individual hydrometeorological drivers of G

(VPD, SWC, PPFD). We also estimate the total predictive ability of a model including all three drivers. We then examine the biogeographical patterns of this hydrometeorological coupling of G , as a function of climate, soil properties and vegetation structure. We hypothesize differences in absolute and relative G coupling to the hydrometeorological drivers across biomes as a result of specific environmental constraints, with tighter coupling with VPD and SWC in drier biomes. We also expect that climate, soil and vegetation structure determine VPD, SWC and PPFD coupling of G , with greater coupling in sites with drier conditions and marked climatic seasonality, in fine textured soils associated with higher water tensions, and in tall stands with low leaf areas that are expected to have tighter canopy-atmosphere coupling.

4.2 Material and methods

4.2.1 Sapflow and environmental data

We extracted 1858 time series of tree sap flow from the SAPFLUXNET database (Poyatos *et al.*, 2020b). These time series met our requirements for data quality (see filtering section below) and corresponded to 130 species on 122 sites without any experimental treatment (Table C1 and C2). Sub-daily sap flow time series were obtained directly in sap flux density units (SFD; $\text{cm}^3 \text{ cm}_{\text{ASW}}^{-2} \text{ h}^{-1}$) or, when sapwood area was not available, in whole-tree sap flow units (SF; $\text{cm}^3 \text{ h}^{-1}$; 24 out of 122 data-sets). In those latter cases, SF time series were converted to SFD units by dividing SF data by an estimation of tree sapwood area (ASW) using a global allometric relationship as a function of tree basal area and functional type (i.e. angiosperm vs gymnosperm) as predictors ($R^2 = 0.78$; $n = 2262$) (Fig. C1). Using the ‘sapfluxnetr’ R package (Granda *et al.*, 2020), sub-daily SFD time series were aggregated to daytime SFD values (i.e., 6 am to 6 pm solar time). Following Flo *et al.* (2019), sap flow time series measured with non-calibrated heat dissipation sensors were corrected for bias in absolute SFD multiplying by a constant factor (1.405).

Similarly to SFD, we obtained VPD [kPa] and PPFD [$\mu\text{mol m}_{\text{ASW}}^{-2} \text{ s}^{-1}$] time series for each site from SAPFLUXNET on-site measurements, which were subsequently averaged to daytime values. When PPFD data were not available in the datasets (12 out of 122 sites), PPFD was calculated using the mean short-wave radiation between 6 am and 6 pm extracted from the ERA5 re-analyses data base ((C3S), 2017) and then multiplying by 2.3 to transform it into PPFD. Soil water content (SWC; v/v) data were missing in 43% of the SAPFLUXNET datasets included in this study. To ensure homogeneity across sites, we used SWC from the 15–30 cm soil depth layer obtained from the ERA5-land reanalysis dataset (2019) at 9x9km resolution (see database validation in Chapter 5).

4.2.2 Data filtering

In order to minimize seasonal phenological changes in leaf area, we excluded all periods between 15 days before the first daytime average temperature under 0°C and 30 days after the last day with temperatures under 0°C , during the cold season of each site (similar to Novick *et al.*, 2016). To prevent artefacts in whole-tree canopy conductance calculation (Ewers & Oren, 2000), we filtered out rainy days –days when SWC increased– and days when average daytime VPD was under 0.3 kPa (Anderegg *et al.*, 2018). We also ensured a sufficient range in hydrometeorological conditions

by discarding sites with a total VPD range below 0.5 kPa or SWC range below 0.05 m³ m⁻³, and with PPFD maximum values below 400 μmol m_{Asw}⁻² s⁻¹.

4.2.3 Whole-tree canopy conductance calculation

To obtain G_s , we firstly transformed SFD units from [cm³ cm_{Asw}⁻² h⁻¹] to [Kg m_{Asw}⁻² s⁻¹] and then we converted it to daytime tree canopy conductance per unit of sapwood area G_{Asw} [mol m_{Asw}⁻² s⁻¹] following Phillips & Oren (1998) and a unit transformation (eq.1).

$$G_{Asw,j,i,k} = \frac{115.8 + 0.4236 T_{j,i} \cdot SFD_{j,i,k}}{VPD_{j,i}} \cdot \eta \cdot \frac{T_0}{(T_0 + T_{j,i})} \cdot \frac{P_0 e^{0.00012 \cdot h_i}}{P_0} \quad (4.1)$$

Where $SFD_{j,i,k}$ is the sap flux density value of each site (j), day (i), and tree (k); $T_{j,i}$ [°C] is the temperature, $VPD_{j,i}$ [kPa] is the daytime vapour pressure deficit, η equals 44.6 mol m⁻³, T_0 is 273 K, P_0 is 101,325 Pa and h [m] is the altitude of each site. For two sites where h values were not available, it was extracted from The Shuttle Radar Topography Mission (STRM) (Center, 2017).

4.2.4 Hydrometeorological coupling quantification

We define hydrometeorological coupling as the coefficient of determination (R^2) of simple and multiple linear mixed models of VPD, SWC and PPFD explaining G_{Asw} at the site-tree level. High R^2 levels imply high predictive power of hydrometeorological drivers over G_{Asw} . To do that, we started by binning the data to avoid issues related to unbalanced distributions of G_{Asw} throughout the range of VPD, SWC or PPFD. Specifically, we calculated the average of G_{Asw} measurements comprised in 0.2 kPa VPD intervals, five site-specific SWC intervals and 250 μmol m_{Asw}⁻² s⁻¹ PPFD intervals. For each summarized G_{Asw} we defined a specific VPD, SWC and PPFD value as the average values of VPD, SWC and PPFD of the data inside each bin. Then, we fitted uni-variate models for each site using G_{Asw} as response variable and the neperian logarithm of each driver as predictor (Fig. C4). Similarly, we also fitted an additive, multiple regression models of site-level G_{Asw} as a function of the logarithm of all three hydrometeorological drivers (TOTAL model). The hierarchical structure of species and trees within sites was taken into account using linear mixed models, implemented with the lmer function of the ‘lme4’ R package (Bates *et al.*, 2015). When sites had more than one tree per species and more than one species (54 out of 122 sites), random intercept and slopes parameters were fitted for species, and random intercept parameters for trees nested into species. When models did not converge, the random structure was simplified and only trees random intercept parameters were fitted (33 out of 54 sites). When sites had just one species and multiple trees (67 out of 122 sites), we fitted a random intercept for trees. When a site had multiple species and just one tree per species (1 out of 122), random intercept and slopes were fitted for species.

Since we were interested in total coupling of the individuals of the site, hydrometeorological coupling was set as the conditional R^2 of the models (i.e. R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} , R^2_{TOTAL}) (Table C3), calculated with the ‘MuMIn’ R package (Bartoń, 2020). We fitted simple and multiple regression models instead of more sophisticated non-linear models to reduce complexity and gain generalizability across the data sets. An alternative analysis fitting generalized additive models (GAM) as implemented in the bam function of the ‘mgcv’ R package (Wood, 2011), and specifying

the same random structure, resulted in coupling values highly correlated to those obtained with the simpler linear modelling approach presented here ($r_{VPD} = 0.88$, $r_{SWC} = 0.89$, $r_{PPFD} = 0.88$, $r_{TOTAL} = 0.74$).

4.2.5 Biome classification and site-level bioclimatic data

The estimates of G_{Asw} hydrometeorological coupling were complemented with site-level data on climate, soil properties and vegetation structure. These data were either directly obtained from the metadata associated to each SAPFLUXNET dataset or from additional data sources. We took from SAPFLUXNET the biome corresponding to each site –obtained from Whittaker diagrams using Chelsa Climate databases (Karger *et al.*, 2017) (Fig. C2)– and carried out an exhaustive quality control to reassign site biomes when necessary (Table C2) in consultation with SAPFLUXNET datasets contributors. Biomes were simplified into 5 groups; drylands (DRY), woodlands (WOOD), temperate forest (TEMP), boreal forests (BOR) and tropical forests (TROP) (Table C4 and Fig. C2).

For each site, we extracted climate information from global rasters (Fig. C3). We used monthly mean precipitation, monthly maximum temperature and monthly minimum temperature rasters for the period 1979 to 2013 from the Chelsa Climate databases (Karger *et al.*, 2017), to estimate monthly potential evapotranspiration (mPET), annual potential evapotranspiration (PET) and mean annual precipitation (MAP) using the ‘envirem’ R package [Title & Bemmels, 2018]. Then, we calculated MAP over PET (PPET) as a water availability index, and the standard deviation of the monthly differences between mean precipitation and mPET ($P-PET_{sd}$) as an index of seasonality in water availability. Relevant soil parameters were obtained from in situ data in SAPFLUXNET and complemented with SoilGrids 2.0 (Hengl *et al.*, 2017) when data were not available in SAPFLUXNET (Table C3). We used the proportion of sand and clay particles in the fine earth fraction [%], the total nitrogen [g kg^{-1}] and the depth to bedrock (up to 200 cm) to characterize soils. We used bedrock depth because of its ecological relevance, but results for this variable should be considered with caution due to its particularly high variability at fine spatial scales. Stand height was available in SAPFLUXNET for most sites. When this was not the case, information was completed using the average tree height of the corresponding site (again from SAPFLUXNET, 3 out of 122 sites) or when both were absent it was extracted from the Global 1km Forest Canopy Height raster (Simard *et al.*, 2011) (3 out of 122 sites) (Table C3). When site LAI was not available from SAPFLUXNET (37 out of 122 sites), it was estimated as the average of the 95th percentile of the period 2010 to 2016 of the MCD15A3H.006 MODIS Leaf Area Index product (0.5x0.5 km grid) (Myneni, 2015), calculated using Google Earth Engine (Gorelick *et al.*, 2017) (Table C3). All raster manipulation was performed with ‘raster’ and ‘stars’ R packages (Hijmans, 2020; Pebesma, 2020) using a common coordinate reference system.

4.2.6 Statistical analyses

In order to test whether the hydrometeorological coupling of G_{Asw} varies across biomes, we fitted weighted regressions using the modelled R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} and R^2_{TOTAL} as response variables and biome as explanatory variable (fixed factor). The number of tree-days with SFD measurements

in each site was used as a weighting variable. Similarly, we also tested the significance of cross-biome differences between paired hydrometeorological couplings (e.g. difference between VPD and SWC coupling, $R^2_{VPD} - R^2_{SWC}$) using the same model structure.

We further explained the biogeographical patterns in the hydrometeorological coupling across sites as a function of climate, soil properties and vegetation structure. We fitted four multiple weighted regression models with R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} and R^2_{TOTAL} as response variables and $\log(PPET)$, $\log(P-PET_{sd})$, soil % clay, soil total nitrogen, soil bedrock depth, stand height and LAI as bioclimatic predictors (Fig. C3). We also used the number of tree-days of each site as weighting variable. Sand percentage was not included due to a high correlation with soil % clay ($r = -0.73$). A stepwise model selection process based on minimising AIC was applied. We checked for normality and homoscedasticity of residuals in all models, and we also checked for multicollinearity by quantifying Variance Inflation Factors (VIF) using the ‘performance’ R package (Lüdecke *et al.*, 2020). These models were combined with the rasters of bioclimatic data (at a uniform resolution of 9x9 km), to predict and map global patterns of G hydrometeorological coupling to VPD, SWC and PPFD.

We also assessed the relative importance of each hydrometeorological driver by extracting the marginal partial R^2 of each hydrometeorological variable of the complete (TOTAL) model. These partial R^2 were calculated using ‘r2beta’ function of the R r2glmm package (Jaeger, 2017) and relativized by the sum of the three partial R^2 (relative R^2). These relative R^2 values can be interpreted as the relative importance of each hydrometeorological variable in limiting transpiration. Then, similarly as above, we fitted three multiple weighted regression models using the estimated relative R^2 as response variables and the bioclimatic variables as predictors. The resulting models were used to project the relative importance of each hydrometeorological driver globally. All statistical analyses were performed in R 3.6.2 (R Core Team, 2017).

4.3 Results

We found large differences in the coupling of G_{Asw} (R^2 coupling metric) to each of the individual hydrometeorological drivers globally and among biomes (Fig. 4.1 and Table 4.1). We observed that G_{Asw} was predominantly coupled to VPD across biomes (Fig. 4.1), while G_{Asw} coupling to SWC and PPFD was comparatively less important. Although coupling to SWC and PPFD were relatively similar, the effect of SWC was higher for DRY, TEMP and particularly WOOD biomes whereas PPFD tended to dominate for BOR and TROP biomes (Table 4.1). The outcomes of the linear models show a significantly higher VPD coupling for TEMP and TROP biomes than for DRY (Table. 1). Somewhat surprisingly, SWC coupling was significantly higher for TEMP, TROP and also WOOD biomes than for the DRY biome. The PPFD coupling was also lowest for the DRY biome and was significantly higher for TEMP, BOR and TROP biomes; the G_{Asw} coupling to PPFD was also significantly lower in the WOOD biome compared to TEMP and TROP biomes (Table 4.1). The DRY biome was the one in which all three drivers collectively explained less variability in G_{Asw} .

In the models explaining the biogeographical patterns of G_{Asw} hydrometeorological coupling (which explained 26-35% of the variance), climate, soil and vegetation structure variables were

Table 4.1: Analysis of variance testing differences among biomes in the coupling (conditional R^2 's from mixed models) of tree-level water conductance (G_{Asw}) to each of the main hydrometeorological drivers: vapour pressure deficit (R^2_{VPD}), soil water content (R^2_{SWC}), radiation (R^2_{PPFD}) and the complete model including all drivers (R^2_{TOTAL}). The table shows the mean coupling obtained across all sites in each biome. We also show the means of the paired differences between individual hydrometeorological couplings and the corresponding statistical significance. DRY: dry and desert biomes; WOOD: woodlands and shrublands; TEMP: temperate biomes; BOR: boreal and tundra; TROP: tropical and subtropical biomes. Different superscript letters indicate significant ($p < 0.05$) Tukey tests of paired differences between biomes. Asterisks indicate statistically significant differences from zero for the paired differences between hydrometeorological couplings.

| Biome | R^2_{VPD} | R^2_{SWC} | R^2_{PPFD} | R^2_{TOTAL} | $R^2_{VPD} - R^2_{SWC}$ | $R^2_{VPD} - R^2_{PPFD}$ | $R^2_{SWC} - R^2_{PPFD}$ | Number of sites |
|-------|---------------------|---------------------|---------------------|---------------------|-------------------------|--------------------------|--------------------------|-----------------|
| DRY | 0.439 ^A | 0.225 ^A | 0.210 ^A | 0.502 ^A | 0.215 ^{A***} | 0.229 ^{AB***} | 0.015 ^{AB} | 7 |
| WOOD | 0.514 ^{AB} | 0.390 ^B | 0.295 ^{AB} | 0.652 ^B | 0.124 ^{A***} | 0.219 ^{B***} | 0.095 ^{B***} | 29 |
| TEMP | 0.567 ^B | 0.452 ^B | 0.417 ^C | 0.666 ^B | 0.116 ^{A***} | 0.150 ^{A***} | 0.034 ^{A*} | 70 |
| BOR | 0.622 ^{AB} | 0.453 ^{AB} | 0.499 ^{BC} | 0.650 ^{AB} | 0.168 ^A | 0.122 ^{AB} | -0.046 ^{AB} | 8 |
| TROP | 0.651 ^B | 0.413 ^B | 0.444 ^C | 0.696 ^{AB} | 0.238 ^{A***} | 0.203 ^{AB***} | -0.031 ^A | 8 |

Statistical significant levels: " ." $p < 0.1$; " **" $p < 0.05$; " ***" $p < 0.01$; " ****" $p < 0.001$.

identified as common controls on the G_{Asw} hydrometeorological coupling (Table 4.2). In particular, $\log(\text{PPET})$, soil clay %, and stand height were selected for all three hydrometeorological drivers (i.e. VPD, SWC and PPFD) and the TOTAL model, with tighter coupling always associated to higher climatic water availability, fine textured soils and taller vegetation. In addition, seasonality in water availability and LAI were associated with lower coupling to VPD and PPFD, whereas soil nitrogen had the exact opposite effect. Bedrock depth was only selected for the R^2_{SWC} model, in which deeper soils were associated with looser coupling, although the effect was not statistically significant (Table 4.2).

Table 4.2: Parameters of the models explaining G_{Asw} coupling to VPD, SWC, PPFD and to all three hydrometeorological drivers (R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} and R^2_{TOTAL} , respectively) as a function of climatic, soil and stand structure variables. $\log(\text{PPET})$: logarithm of precipitation over potential evapotranspiration [% $\log(\text{mm mm}^{-1})^{-1}$]; $\log(\text{P-PET}_{sd})$: logarithm of the standard deviation of the difference between precipitation and potential evapotranspiration [% $\log(\text{mm})^{-1}$]; Clay percentage [% $\%_{\text{clay}}$]; Total Nitrogen [% (Kg g^{-1}) $^{-1}$]; Bedrock depth [% cm^{-1}]; Stand Height [% m^{-1}]; LAI: leaf area index [% $(\text{m}^2 \text{m}^{-2})^{-1}$]. NI means that the variable was not included in the model after model selection. The R^2 of each multiple regression is also shown.

| G_{Asw} coupling | Intercept [%] | Climate | | Soil | | | Vegetation structure | | R^2 |
|--------------------|---------------|---------------------|---------------------------|-----------|----------------|---------------|----------------------|------------|-------|
| | | $\log(\text{PPET})$ | $\log(\text{P-PET}_{sd})$ | Clay | Total Nitrogen | Bedrock depth | Stand Height | LAI | |
| R^2_{VPD} | 80.242 *** | 4.862 . | -8.642 * | 0.313 ** | 1.930 * | NI | 0.603 *** | -3.322 *** | 0.255 |
| R^2_{SWC} | 53.978 *** | 10.850 *** | NI | 0.314 *** | NI | -0.120 ns | 0.348 ** | NI | 0.353 |
| R^2_{PPFD} | 47.591 ** | 7.531 * | -6.501 . | 0.323 ** | 1.603 . | NI | 0.788 *** | -2.202 * | 0.352 |
| R^2_{TOTAL} | 54.464 *** | 4.698 * | NI | 0.248 ** | 1.416 . | NI | 0.228 . | NI | 0.191 |

Statistical significant levels: " ." $p < 0.1$; " **" $p < 0.05$; " ***" $p < 0.01$; " ****" $p < 0.001$; ns not significant..

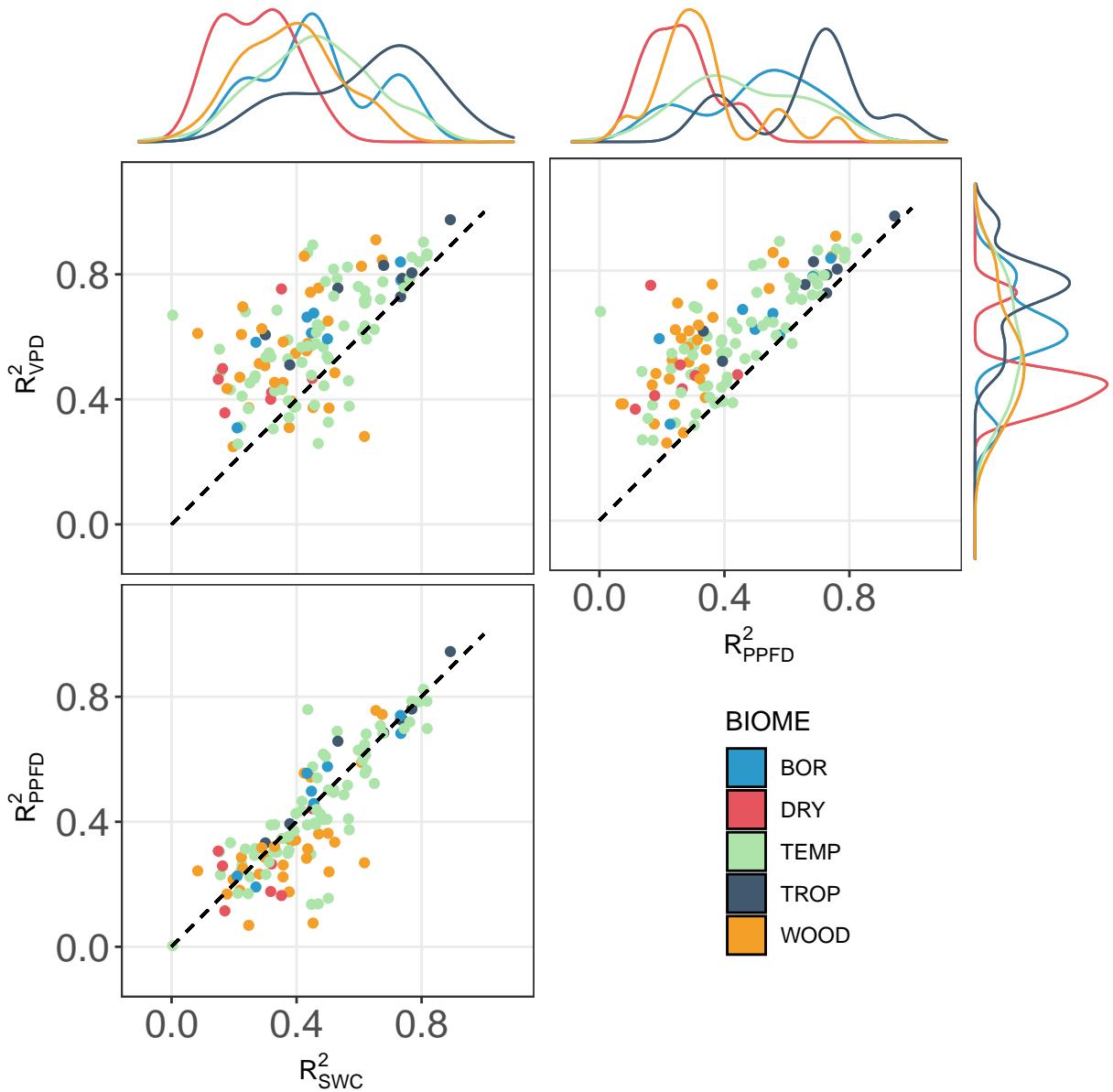


Figure 4.1: Bi-variate and uni-variate distributions of the coupling of $G_{A\text{sw}}$ to the hydrometeorological drivers studied: vapour pressure deficit (R^2_{VPD}), soil water content (R^2_{SWC}) and radiation (R^2_{PPFD}) for different biomes. Points correspond to site-level modelled conditional R^2 values. Colours represent different biomes, DRY: dry and desert biomes; WOOD: woodlands and shrublands; TEMP: temperate biomes; BOR: boreal and tundra; TROP: tropical and subtropical biomes. Dashed black line shows 1:1 relation.

When predictions of $G_{A\text{sw}}$ coupling to each of the hydrometeorological drivers were mapped at the global scale, a different spatial pattern was observed for all three drivers (Fig. 4.2). $G_{A\text{sw}}$ coupling to VPD was higher than ca. 50% almost everywhere except for some sub-tropical regions. The regulation of tree water fluxes at high northern latitudes (above 50° N) and in tropical regions was highly coupled to VPD, SWC and PPFD (Fig. 4.2). In contrast, trees living in subtropical regions dominated by the DRY and WOOD biomes tended to be poorly coupled to SWC and PPFD, and relatively more coupled to VPD (Fig. 4.2). When considering the relative importance (partial R^2) of each of the three variables in driving transpiration (Fig. 4.3), northern areas, temperate regions, drylands and savannas were typically limited by VPD, whereas tropical regions

tended to be co-limited by VPD and PPFD (Fig. 4.3).

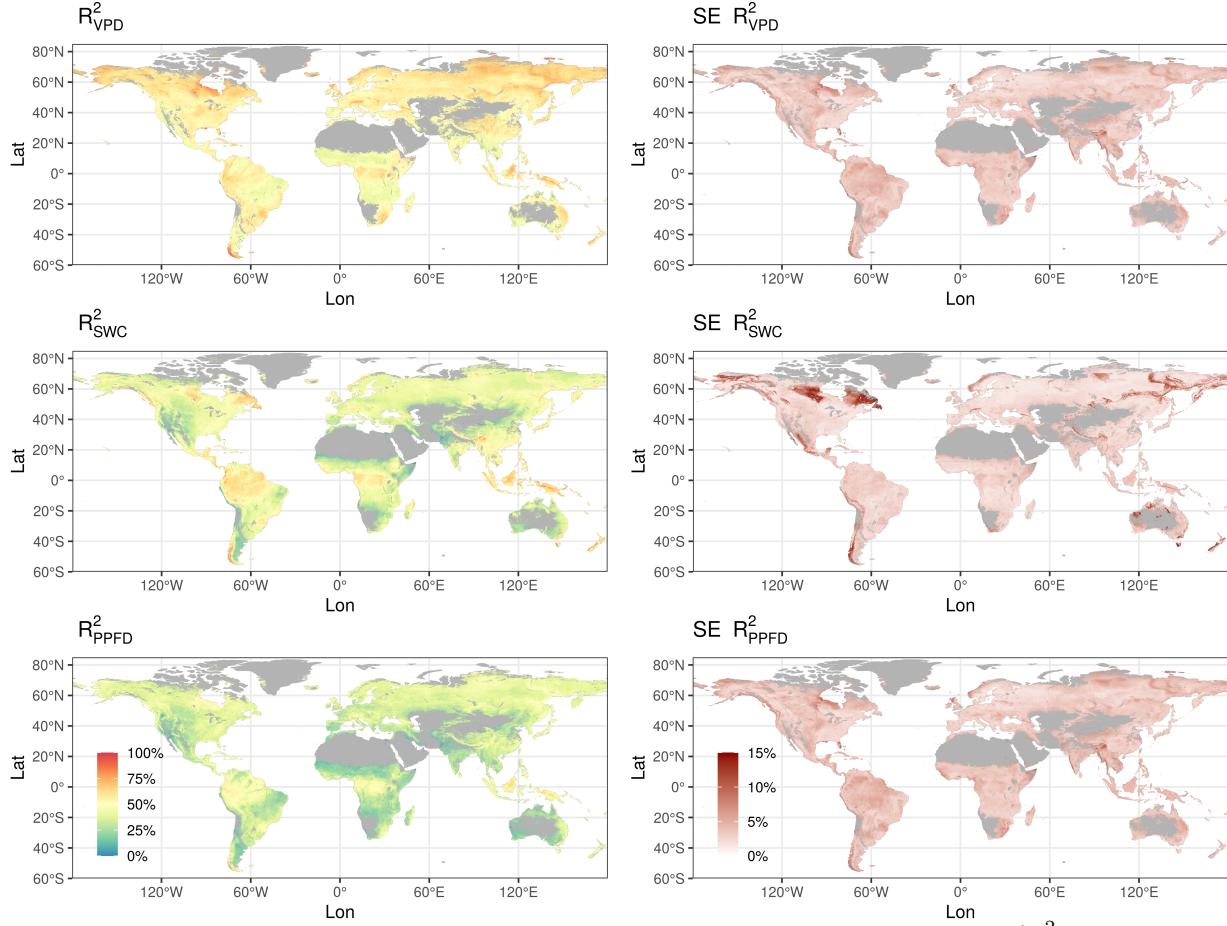


Figure 4.2: Global projection of G_{Asw} coupling to VPD, SWC and PPFD (R^2_{VPD} , R^2_{SWC} and R^2_{PPFD} , respectively), obtained from regression models of each coupling as a function of climatic, soil and stand structure variables (left panels). Right panels show projected Standard Error of the corresponding model.

4.4 Discussion

This study provides the first attempt to examine the absolute and relative importance of VPD, SWC and PPFD as the main hydrometeorological drivers of the regulation of tree transpiration at the global level. All sites presented some degree of transpiration coupling to the hydrometeorological drivers considered, although there was substantial variability in the magnitude of this coupling (i.e. R^2_{VPD} , R^2_{SWC} , R^2_{PPFD}). We demonstrate that the regulation of transpiration is predominantly coupled to VPD in all biomes, while transpiration limitation caused by SWC and PPFD is generally comparable and lower. Although our sample size is much smaller and possibly biased (see Fig. 4.3 and Table 4.1) relative to global studies based on remote sensing approaches, our results clearly identify atmospheric dryness as the major regulator of transpiration globally, which is consistent with recent reports showing that VPD limits vegetation growth at the global scale (Babst *et al.*, 2019; Yuan *et al.*, 2019). Yet, contrary to other studies focusing on the controls of primary productivity (Jung *et al.*, 2017; Liu *et al.*, 2020), we do not find a dominant role of SWC on transpiration regulation. However, unlike these studies, our approach focuses at the plant-level and uses actual transpiration data, also considering the effect of light, which has been rarely assessed in this type of studies.

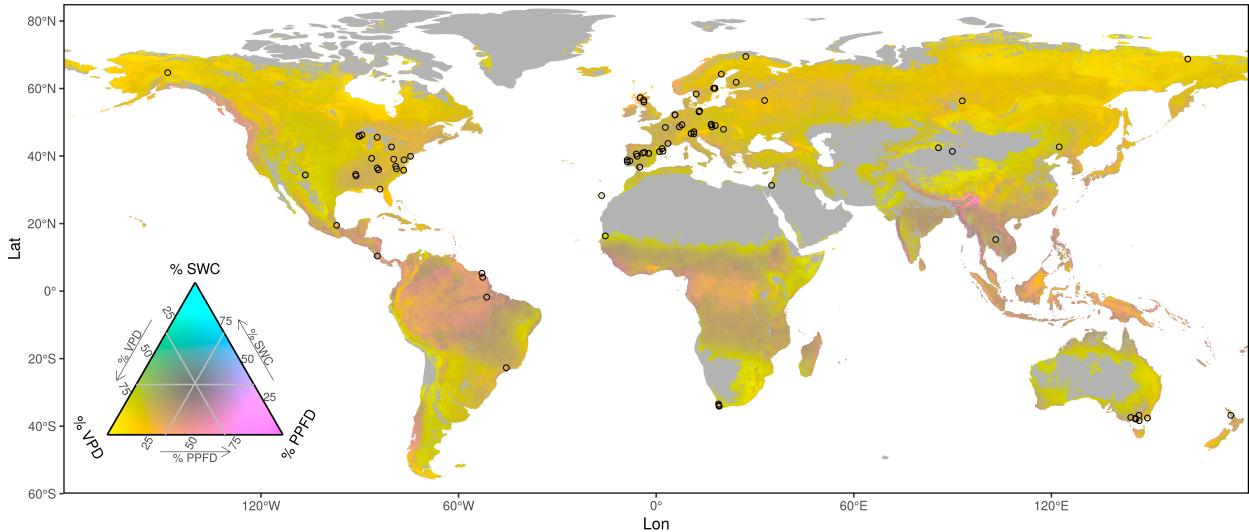


Figure 4.3: Relative importance (partial R^2) of the three hydrometeorological drivers of transpiration regulation calculated from the complete (TOTAL) model, and projected at the global scale using linear models with climate, soil and vegetation structural variables as explanatory variables. Grid values were calculated using the ‘tricolore’ package (Schöley & Kashnitsky, 2020) for each cell as the relative value of the projections of the relative importance of each hydrometeorological variable. Colour gradient indicate the relative importance of the three hydrometeorological constraints. Light grey colour are deserts or non-forested areas. % VPD: vapour pressure deficit relative importance. % SWC: soil water content relative importance. % PPFD: photosynthetic photon flux density relative importance. Points indicate locations of study sites.

Soil moisture coupling of transpiration regulation was only tighter than coupling to radiation in WOOD and TEMP biomes, indicating the importance of soil water limitations on transpiration in these biomes. Interestingly, the importance of SWC decreased in DRY biomes, even if actual sensitivity to SWC was high (Fig. C4). This result contrasts to those found at the ecosystem level, where drier sites present larger SWC limitations than wetter ones (Novick *et al.*, 2016). This opposite result between transpiration and evapotranspiration limitations suggests that the soil contribution to ecosystem surface conductance may be large (Li *et al.*, 2019) and strongly limited by SWC in DRY biomes. The lack of coupling in drylands is intriguing and may be related to the diversity of water use strategies in water limited systems, which range from drought-deciduousness to deep rooting or high hydraulic safety (e.g., Ackerly, 2004; Jacobsen *et al.*, 2007). Deep roots, for instance, could allow sufficient water supply to uncouple transpiration from hydrometeorological drivers and specifically from shallow SWC (Barbeta & Peñuelas, 2017). At the other extreme, exposure to low water potentials results in early stomatal closure (Martin-StPaul *et al.*, 2017), effectively disconnecting transpiration from hydrometeorological drivers for long periods of the year. Memory effects (Ogle *et al.*, 2015) are also likely to be more common in water-limited systems, which may result in more complex responses of transpiration to individual hydrometeorological drivers. Finally, we did not consider the co-variation between VPD, SWC and PPFD. VPD and SWC co-variation is relatively low at the daily level at large spatial scales (Novick *et al.*, 2016), but in water-limited regions SWC shows strong interactions with VPD (Zhou *et al.*, 2019a) and PPFD (Boese *et al.*, 2018), which could produce compound drought effects that would complicate

disentangling the coupling of transpiration to individual drivers. It should also be noted that we focus here on relatively tall woody vegetation, as this is the one likely to be measured with sap flow sensors (Poyatos *et al.*, 2020b), and hence our analysis excludes extremely arid sites likely to be totally driven by water availability (compare the grey areas in our Fig. 4.3 with Fig. 4.1 in Running *et al.* (2004)).

Beyond general biome effects, differences in coupling among sites were partially explained by differences in climate, soil and stand structure. Besides the generally lower coupling in areas with less climatic water availability, consistent with the results discussed in the previous paragraph, we have found that regions with high climatic seasonality (i.e. high $\log(P-PET_{sd})$) are less coupled to VPD (and to a lower extent radiation) than regions with more seasonally stable climate. Low R^2_{VPD} under high climatic seasonality may be related to changes in transpiration regulation responses to VPD between the dry and wet seasons (Renninger *et al.*, 2010), which could be the case of, for example, seasonal tropical forest biomes with semi-deciduous species (Monasterio & Sarmiento, 1976). Alternatively, low VPD coupling could be related to transpiration regulation driven by other bioclimatic variables or their combination, as partially observed in the co-limitation by all three hydrometeorological drivers in many areas where climatic seasonality is large (Fig. 4.3, Fig. C3). Coupling to radiation was higher in wetter sites (i.e. high PPET), which are mainly located in tropical and boreal regions (Fig. C3). This would reflect the key role of radiation as a driver of transpiration in energy-limited areas with shallow clouds and fogs such as the tropics (Fig. 4.2) (Gentine *et al.*, 2019), and in areas where radiation is a strong limiting factor such as boreal regions (Kasurinen *et al.*, 2014). Our analysis of the relative importance of the three hydrometeorological drivers indicates a dominant role of radiation as a limiting factor in the tropics particularly in South-East Asia (Fig. 4.3), consistent with previous reports (Running *et al.*, 2004).

Our results show the key importance of soil characteristics, particularly texture, in explaining variability in transpiration coupling. Trees increase coupling to all three hydrometeorological drivers under high clay content (finer texture), which results in more negative water potentials at a given water content (Hillel, 1998). This result is to be expected considering that we assessed transpiration responses to soil water content, which implies that plants in fine textured soils would effectively experience lower water availability than plants in more coarsely textured soils with the same water content. In addition, our results indicate deeper soils (higher bedrock depth) were associated to lower coupling to soil water availability, consistent with the notion that access to deep water may uncouple transpiration from shallow SWC (Barbeta & Peñuelas, 2017).

Vegetation height and LAI are also important drivers of vegetation transpiration coupling. VPD, SWC and PPFD limitations to G_{Asw} are typically higher in taller trees, consistent with previous studies (Boese *et al.*, 2018; Zhao *et al.*, 2019a). Tree height is associated with productive areas with high resource availability, including water. In general, taller trees live in wetter regions and have higher water transport efficiencies and lower resistance to embolism (Liu *et al.*, 2019, Chapter 5). These traits are associated with acquisitive water use strategies and a tighter stomatal control of transpiration (Klein, 2014). A similar argument can be used to explain tighter coupling to VPD and radiation in areas with high soil nitrogen concentrations, as the latter have been related to increased transpiration rates in wet soil and greater degree of stomatal control under drought (Shimshi, 1970; Ewers *et al.*, 2001). Taller canopies are also more aerodynamically rough

and, therefore, show higher VPD coupling due to higher levels of leaf surface VPD. Interestingly, once accounting for the effect of vegetation height LAI had a negative effect on transpiration coupling to VPD and PPFD. We associate this result to the fact that higher LAI is related to lush canopy structures that would have a significant proportion of the leaves decoupled from the atmosphere and from direct radiation income (Zhang *et al.*, 2016).

Differences in coupling among sites should also reflect different water use strategies in the corresponding communities. These differences are reflected in part in the climatic, soil, and structural differences we studied, but also underlie the relatively large unexplained variance, which could be related to contrasting water use strategies coexisting in the same biomes and even in the same sites (Anderegg *et al.*, 2018). This implies that species traits should be included if we aim to understand the fine-scale distribution of transpiration responses (Chapter 5) and their coupling to hidrometeorological drivers to predict G responses to environmental variation.

In conclusion, we found that VPD is the main hidrometeorological driver of transpiration regulation globally but we also showed that VPD coupling did not increase in warmer sites, as found in ecosystem-level studies (Novick *et al.*, 2016). Nevertheless, the role of VPD in driving transpiration regulation will likely be dominant in a warmer world, given the generalised increases in projected VPD (Ficklin & Novick, 2017). Using machine learning methods (Zhao *et al.*, 2019a) and sub-daily data (when SWC is effectively constant) could help to better disentangle the effect of each hidrometeorological driver (Lin *et al.*, 2018), and would allow a more explicit consideration of interactions between drivers (Zhou *et al.*, 2019b). Our results indicate clear differences among hidrometeorological couplings and contribute to disentangle their relative effects on transpiration regulation, which determines vegetation water use, tree growth and ecosystem production. Consequently, global models simulating vegetation-atmosphere fluxes should account for the limiting effects produced by VPD, SWC and PPFD and their variability in space and time, which should be facilitated by an explicit description of water transport in plants (Anderegg & Venturas, 2020).

5

Climate and functional traits jointly mediate tree water use strategies

Victor Flo, Jordi Martínez-Vilalta, Maurizio Mencuccini, Victor Granda, William R. L. Anderegg,
Rafael Poyatos. *New Phytologist*, Under revision.

Abstract

Tree water use is central to plant function and ecosystem fluxes. However, it is still unknown how organ-level water relations traits are coordinated to determine whole-tree water use strategies in response to drought, and if this coordination depends on climate. Here we used a global sap flow data base (SAPFLUXNET) to study the response of water use, in terms of whole-tree canopy conductance (G), to vapour pressure deficit (VPD) and to soil water content (SWC) for 142 tree species. We investigated the individual and coordinated effect of six water relations traits (vulnerability to embolism, Huber value, hydraulic conductivity, turgor-loss point, rooting depth and leaf size) on water use parameters, also accounting for the effect of tree height and climate (mean annual precipitation, MAP). Reference G and its sensitivity to VPD were tightly coordinated with water relations traits rather than with MAP. Species with efficient xylem transport had higher canopy conductance but also higher sensitivity to VPD. Moreover, we found that angiosperms had higher reference G and higher sensitivity to VPD than gymnosperms. Our results highlight the importance of trait coordination and the complications of defining a single, whole-plant resource use spectrum ranging from ‘acquisitive’ to ‘conservative’.

5.1 Introduction

Plant water use is a key component of the global water cycle (Katul *et al.*, 2012). Plants regulate water use across a broad range of timescales to maintain a favourable water status under varying water availability (Feng *et al.*, 2017). This regulation is the result of evolutionary processes together with environmental and biophysical constraints that have determined a huge diversity of species-specific water use strategies mediated by a particular suite of traits (Bacelar *et al.*, 2012; Lu *et al.*, 2020). These specific strategies determine plant survival under drought (Mitchell *et al.*, 2013), species coexistence (Ehleringer *et al.*, 1991; Jackson *et al.*, 1995) and ecosystem CO₂ and water fluxes (Mencuccini *et al.*, 2019a). Thus, a comprehensive understanding of how regulation of whole-plant water use relates to organ-level water relations traits would allow for a better characterization of plant responses to drought and improved prediction of climate change impacts on vegetation (Anderegg, 2015).

Among many other traits, the sensitivity of stomata to drought stress is a major regulator of plant water use over relatively short timescales (Martin-StPaul *et al.*, 2017). In the absence of effective stomatal control under drought, water uptake by roots might not compensate water loss, resulting in high tensions in plants' vascular system, which can trigger the entrance of air bubbles leading to xylem embolism (Tyree & Zimmermann, 2002). If embolism spreads to most of the xylem conduits, water transport becomes restricted and plants may eventually die from hydraulic failure (Tyree & Sperry, 1988; Choat *et al.*, 2018). Through stomatal closure, plants reduce whole-plant canopy conductance (G) and therefore water loss and embolism risk, and maintain water status within tolerable limits, at the expense, however, of reducing gas exchange. Plants close stomata in response to drops in leaf and/or soil water potentials (see Martínez-Vilalta & Garcia-Forner, 2017) produced by increased atmospheric water demand (i.e., vapor pressure deficit; VPD) and reduced soil water availability (i.e., soil water content; SWC) (Jarvis, 1976; Grossiord *et al.*, 2017). Stomatal responses have been largely described using semi-empirical models (Jarvis, 1976; Oren *et al.*, 1999b; Damour *et al.*, 2010) and optimality approaches relying on the coupling of photosynthesis and transpiration (Wang *et al.*, 2020), with a recent focus on plant hydraulics (Sperry *et al.*, 2016). Global syntheses of these two approaches exist (Lin *et al.*, 2015; Hoshika *et al.*, 2018) but they are only at the leaf level, and they do not consider the coordination of stomatal and hydraulic traits.

Coordination among stomatal sensitivity and other hydraulic and allocation traits is thought to underlie differences in water use strategies among species (Meinzer, 2002; Sperry *et al.*, 2016; McCulloh *et al.*, 2019). Because in woody plants water has to be transported from the soil through the xylem to supply leaf transpiration, the hydraulic properties of the xylem are key determinants of plant water relations and water use strategies. Particularly, maximum sapwood hydraulic conductivity (K_s ; Table 5.1) and vulnerability to xylem embolism (usually quantified as Ψ_{P50} ; i.e. the water potential at which half of K_s is lost) are key determinants of maximum transpiration rates (Manzoni *et al.*, 2013). In addition, a vulnerable xylem (i.e. high $|\Psi_{P50}|$) has been related to higher canopy-level stomatal sensitivity to VPD across (Litvak *et al.*, 2012) and within some species (Aspinwall *et al.*, 2011). K_s and Ψ_{P50} have been hypothesized to define a safety-efficiency trade-off at the tissue level, by which species with high K_s (i.e., high water transport efficiency) are also more vulnerable to embolism (low $|\Psi_{P50}|$ and low safety) and vice

versa (Venturas *et al.*, 2017). However, this trade-off appears to be weak at the global scale across species, at least as captured with current measurement techniques (Gleason *et al.*, 2016; Sanchez-Martinez *et al.*, 2020). At the leaf level, another key component is the water potential at turgor-loss point (Ψ_{TLP}), which is tightly associated with drought tolerance and habitat water availability (Bartlett *et al.*, 2012). Ψ_{TLP} is also closely related to water potential at complete stomatal closure (Brodribb & Holbrook, 2003; Martin-StPaul *et al.*, 2017), and thus can be used as a proxy for quantifying the sensitivity of plant water use to changes in water availability.

Allocation ratios between the organs involved in water loss, transport and uptake are also important determinants of water use strategies. The ratio of cross-sectional sapwood area to leaf area or Huber value (Hv) is a major trait defining water use strategies because it expresses the water conducting area per unit transpiring area. Increased Hv can contribute to reduce water potential gradients within the plant and therefore potentially compensate for a species vulnerable xylem (Mencuccini *et al.*, 2019b). In addition, a high Hv has also been associated to strict stomatal control in conifers (Martínez-Vilalta *et al.*, 2004; Poyatos *et al.*, 2007) and to higher reference canopy conductance (Novick *et al.*, 2009). Similarly, increasing rooting depth (R_{depth}) gives access to deeper water sources, which potentially allows maintaining less negative and stable water potentials as well as supporting high water use even under climates with high evaporative demand (Martínez-Vilalta & Garcia-Forner, 2017). At the leaf level, the importance of individual leaf size (L_s) for light penetration and crown architecture has been thoroughly studied (Sellers, 1985), but its influence on water use strategies has been little explored, despite the role of leaf evaporative cooling for thermoregulation (Wright *et al.*, 2017). Large leaves with thick leaf boundary layers and an ineffective stomatal control of water loss (Jarvis & McNaughton, 1986), may need to sustain higher transpiration rates to maintain leaf temperature within operative limits under intense radiative heating and/or heat waves (e.g. Drake *et al.*, 2018). This may lead to a trade-off between leaf thermoregulation and the conservation of water and/or hydraulic function (Fauset *et al.*, 2018), especially in hotter sites (Aparecido *et al.*, 2020).

Water use strategies are also mediated by plant height, community composition, and environmental conditions, particularly climate, topography and soil properties; as well as spatial and temporal variability in environmental conditions (Feng *et al.*, 2018). Plant height increases hydraulic path length and hydraulic resistance, and thus plays a major role in the global coordination of several water use traits such as Ψ_{P50} or K_s (Liu *et al.*, 2019), Hv (Mencuccini *et al.*, 2019b) and reference canopy conductance (Novick *et al.*, 2009). Likewise, increasing tree height has been related to enhanced sensitivity of canopy conductance to VPD for some species (Schäfer *et al.*, 2000). Because soil water is a common resource belowground that is influenced by water uptake from many individuals and species, the community composition and diversity of water use strategies in an ecological community can also affect ecosystem fluxes and drought progression (Anderegg *et al.*, 2018, Anderegg *et al.* (2019)). In addition, phylogeny may constrain flexibility in water use strategies independently of the environment, as it has been shown for example by the consistent differences in hydraulic traits between angiosperms and gymnosperms (Johnson *et al.*, 2012; Bartlett *et al.*, 2016) and by the strong phylogenetic conservatism reported for some hydraulic traits such as Ψ_{P50} and K_s at the global scale (Sanchez-Martinez *et al.*, 2020).

In this study we explore water use strategies across tree species using a trait-based approach, which provides a simplified framework to understand species responses to drought at the global

scale (Feng *et al.*, 2018). Our ultimate aim is to better understand how water use strategies emerge from the covariation between traits and the influence of climate. To that end, we use a global database of sap flow measurements (SAPFLUXNET) to calculate G for 142 tree species growing on 126 sites. We then parameterize the response of G to VPD and SWC at the species level and for major taxonomic groups (i.e. angiosperms and gymnosperms). Finally, we characterize the relationships between water use traits and key hydraulic and allocation traits among species (i.e. Ψ_{P50} , K_s , Hv , Ψ_{TLP} , R_{depth} , L_s and tree height), controlling also for the climatic effects produced by differences in precipitation. We hypothesize that (i) water use and water relations traits are coordinated to determine water use strategies at the species level, and (ii) species occupying drier habitats will tend to be more ‘conservative’ in their water use and also tend to have drought tolerance traits. (iii) After controlling for climatic effects, a safety-efficiency trade-off is visible at the scale of whole-plant water use, as opposed to the scale of individual xylem conduits.

5.2 Material and methods

We took a two-step approach to test the previous hypotheses. First, we modelled species-level whole-tree canopy conductance responses to evaporative demand and soil water availability to obtain species’ water use parameters, taking advantage of a recently-compiled global sap flow dataset. Next, we modelled the variability of those water use parameters as a function of species’ water relations traits, mean annual precipitation (MAP), tree height and broad functional types (i.e., angiosperms and gymnosperms).

5.2.1 Sap flow data

Data from 1929 trees belonging to 142 species on 126 plots without experimental treatments (Table D1) and meeting data quality criteria (see Data filtering section below) were obtained from the global SAPFLUXNET database (Poyatos *et al.*, 2020b) (Fig. 5.1 and Table D2). For each species-site combination, we extracted sub-daily sap flux density (SFD; Table 5.1) or whole-tree sap flow (SF; 24 out of 126 data-sets) when tree sapwood areas (ASW) were not available. For these datasets, SF data were then transformed into SFD units by dividing SF values by tree ASW estimated with an allometric relationship. This relationship was obtained using all the SAPFLUXNET trees for which ASW data were available and taking diameter at breast height (DBH) and functional type (i.e., angiosperm or gymnosperm) as predictors ($R^2 = 0.78$; $n = 2262$). Sub-daily SFD time-series were aggregated to daytime SFD averages (i.e., 6am to 6pm solar time) using the `sfn_metrics` function of the `sapfluxnetr` R package (Granda *et al.*, 2020). Time-series obtained from non-calibrated thermal dissipation sensors were corrected for potential bias in absolute SFD by applying a multiplier of 1.405, according to the global synthesis of sap flow calibrations by Flo *et al.* (2019).

5.2.2 Evaporative demand and soil water availability data

We used vapour pressure deficit (VPD) and soil water content (SWC) as proxies of evaporative demand and soil water availability, respectively. Similar to SFD, VPD was obtained from on-

Table 5.1: Description of variables and units in this study.

| Variable | Description | Units |
|--------------------|--|---|
| VPD | Vapour pressure deficit | kPa |
| SWC | Soil water content | $\text{m}_\text{water}^3 \text{m}_\text{soil}^{-3}$ |
| REW | Relative extractable water | |
| Asw | Sapwood area | m^2 |
| SF | Sap flow | $\text{cm}^3 \text{h}^{-1}$ |
| SFD | Sap flux density | $\text{cm}^3 \text{cm}_\text{Asw}^{-2} \text{h}^{-1}$ |
| G | Whole-tree canopy conductance | mol s^{-1} |
| G_Asw | Whole-tree canopy conductance per unit of sapwood area | $\text{mol m}_\text{Asw}^{-2} \text{s}^{-1}$ |
| G'_Asw | Whole-tree stomatal conductance (i.e., G_Asw without aerodynamic conductance) | $\text{mol m}_\text{Asw}^{-2} \text{s}^{-1}$ |
| MAP | Mean annual precipitation | mm |
| MAT | Mean annual temperature | $^\circ\text{C}$ |
| PPET | Mean annual precipitation over potential evapotranspiration | mm mm^{-1} |
| H | Tree height | m |
| Ψ_{P50} | water potential at which half of Ks is lost | MPa |
| K_s | Maximum sapwood hydraulic conductivity | $\text{kg m}^{-1} \text{MPa}^{-1} \text{s}^{-1}$ |
| Hv | Huber value: ratio of cross-sectional sapwood area to leaf area | $\text{cm}_\text{Asw}^2 \text{m}_\text{leaf}^{-2}$ |
| Ψ_TLP | Water potential at leaf turgor-lost point | MPa |
| R_depth | Rooting depth | m |
| L_s | Individual leaf size | cm^2 |
| T | Temperature | $^\circ\text{C}$ |
| h | Plot altitude | m |
| G_REF | Reference G_Asw at VPD = 1 kPa and SWC = 0.5 $\text{m}^3 \text{m}^{-3}$ | $\text{mol m}_\text{Asw}^{-2} \text{s}^{-1}$ |
| β_VPD | G_Asw sensitivity to ln(VPD) | $\text{mol m}_\text{Asw}^{-2} \text{s}^{-1}$ |
| β_SWC | G_Asw sensitivity to ln(SWC) | $\text{mol m}_\text{Asw}^{-2} \text{s}^{-1}$ |

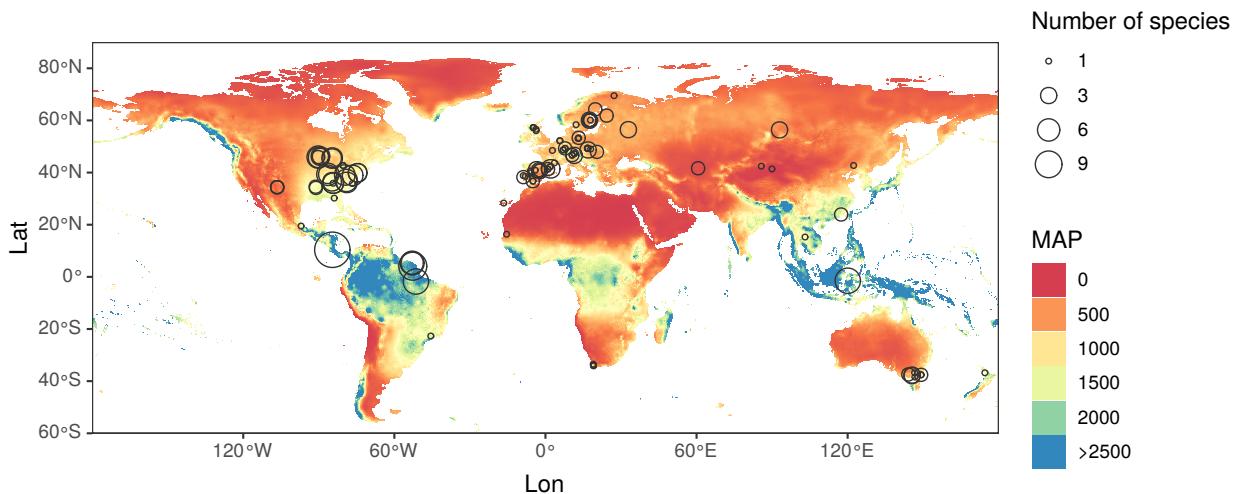


Figure 5.1: Distribution of the plots from the SAPFLUXNET database included in this study. Size of the dots represent the number of different species in the plot. Color gradient show mean annual precipitation (MAP).

site sub-daily measurements from SAPFLUXNET averaged to daytime values. Soil water content (SWC; v/v) was obtained from the 15-30 cm depth layer at 12 am from the ERA5-land re-analysis

product ((C3S), 2019) at 9x9 km resolution. We used ERA5-land re-analyses instead of on-site SWC measures in order to maximize the number of plots and species included in the study, since SWC data were missing in 44% of the SAPFLUXNET data-sets included in this study. In addition, ERA5-land had longer time series (1980 to 2019). We validated the use of ERA5-land data using a linear mixed-model (LMM) regression between ERA5-land and on-site shallow SWC measurements by letting random intercepts and slopes of the response vary by site (n observations = 32815; n plots = 71; R^2 conditional = 0.97, R^2 marginal = 0.26).

To complement SWC, we also calculated relative extractable water (REW), as a normalized measure of soil water availability, as follows:

$$REW_{j,i} = \frac{SWC_{j,i} - SWC_{min}}{SWC_{max} - SWC_{min}} \quad (5.1)$$

where $REW_{j,i}$ and $SWC_{j,i}$ are plot (j) daily (i) values, and SWC_{max} and SWC_{min} , the overall maximum and minimum SWC measured at a plot, respectively. REW takes values between 0 and 1, being 0 the absolute plot lowest SWC and 1 being the highest.

5.2.3 Data filtering

We restricted the analysis to periods without potential phenological changes in leaf area to minimize variations in conductance unrelated to VPD and SWC changes. In the absence of detailed plot-specific observations, we excluded all data for periods comprised between 15 days prior to the first day with temperatures below 0°C and 30 days following the last day under 0°C, respectively, during the cold seasons of each plot site (similarly to Novick *et al.*, 2016). To prevent potential artefacts due to unstable weather conditions in the calculation of whole-tree canopy conductance (Ewers & Oren, 2000) or in the estimation of model parameters, we filtered out days when SWC increased (rainy days), as well as days when daytime-averaged VPD was below 0.3 kPa (Anderegg *et al.*, 2018). To ensure sufficiently contrasting conditions of evaporative demand and soil water availability, we also discarded species with both VPD ranges below 0.5 kPa and SWC ranges below 0.05 (n = 8 species).

After data filtering, the study covers a large geographic area –being Europe and the east of North America especially well represented (Fig. 5.1)– and a wide range of climate conditions, with MAP values ranging from 14 mm to 3626 mm (mean \pm SD = 953 mm \pm 545 mm). Out of the 142 species used in the analyses, 116 were angiosperms and 26 gymnosperms. The number of trees per species ranges from 215 trees (*Pinus sylvestris*) to 1 (this being the case for 23 species) (Table D2). Tree species-level heights (H) range from 2 m (*Coprosma quadrifida*) to 40 m (*Carya glabra*) (mean \pm SD = 21 m \pm 9.75 m).

5.2.4 Whole-tree canopy conductance calculation

Daytime SFD was transformed from [$\text{cm}^3 \text{ cm}_{\text{Asw}}^{-2} \text{ h}^{-1}$] to [$\text{kg m}_{\text{Asw}}^{-2} \text{ s}^{-1}$] and converted to daily whole-tree canopy conductance normalized per unit of sapwood area G_{Asw} using Phillips & Oren (1998) and unit transformations (eq. 5.2).

$$G_{\text{Asw},j,i,k} = \frac{115.8 + 0.4236 T_{j,i} \cdot SFD_{j,i,k}}{VPD_{j,i}} \cdot \eta \cdot \frac{T_0}{(T_0 + T_{j,i})} \cdot \frac{P_0 e^{0.00012 \cdot h_i}}{P_0} \quad (5.2)$$

Where $SFD_{j,i,k}$ is the sap flux density value of each site (j), day (i), and tree (k); $T_{j,i}$ [$^{\circ}\text{C}$] is the temperature, $VPD_{j,i}$ [kPa] is the daytime vapour pressure deficit, η equals 44.6 mol m^{-3} , T_0 is 273 K, P_0 is 101,325 Pa and h [m] is the altitude of each site. When h was not available it was obtained from The Shuttle Radar Topography Mission (SRTM) (Center, 2017) ($n = 2$ plots).

The conductance obtained using eq. 5.2 is considered a good proxy of the tree-level stomatal conductance under the assumption that the canopy and the atmosphere are well-coupled, i.e., when the aerodynamic conductance is much larger than the stomatal conductance. Although this is generally assumed in sap flow studies for both needleleaf and even broadleaf species, there is evidence that coupling may only be partial in some cases (Magnani *et al.*, 1998; Kauwe *et al.*, 2017). Therefore, we also calculated whole-tree stomatal conductance (G'_{Asw}) by removing the contribution of aerodynamic conductance in a subset of plots-species (n plots = 64; n species = 47) where wind speed data were available in SAPFLUXNET (see appendix D notes for details, Chu *et al.*, 2018; Tan *et al.*, 2019). We also related the environmental sensitivity of G'_{Asw} to the hydraulic and allocation traits (see Statistical analyses section below) to assess the potential impact of partial canopy-atmosphere coupling on our results.

5.2.5 Traits and climatic data

Species-level traits ($|\Psi_{P50}|$, Hv , K_s , $|\Psi_{TLP}|$, R_{depth} and L_s) were taken from HydraTry (Mencuccini *et al.*, 2019b; Sanchez-Martinez *et al.*, 2020) and the Global Leaf Size Dataset (Wright *et al.*, 2017) (Table D2). $|\Psi_{P50}|$, Hv , K_s and L_s were log-transformed to achieve normality. In addition, we obtained tree species-level height (H) as the average of SAPFLUXNET actual tree heights, with the number of tree-days with available sap flow values as weighting factor. The height of the stand was used when the actual height of a tree was not available (792 out of 1929 trees).

To account for climatic effects on the species' water use parameters and on water relation traits, we used mean annual precipitation (MAP), mean annual temperature (MAT) and an aridity index defined as precipitation over potential evapotranspiration (PPET) (Fig. 5.1, Fig. D6, Fig. D7). However, for simplicity, we only included MAP in the analyses since for the species in the study MAP was strongly correlated with PPET ($r = 0.94$) and MAT ($r = 0.76$), whereas PPET and MAT correlation was lower ($r = 0.56$). MAP and MAT were obtained for all study plots from the CHELSA data set (1x1 km resolution) (Karger *et al.*, 2017) and averaged at the species level weighting by the number of tree-days. PPET were obtained from CGIAR-CSI Global Aridity index (Trabucco & Zomer, 2019).

5.2.6 G_{Asw} sensitivity to soil water availability and water demand

In some species, G_{Asw} measurements were distributed very heterogeneously throughout the range of VPD or SWC. To avoid the issues associated with such unbalanced distributions we used binned data. Specifically, we calculated the average of G_{Asw} measurements comprised into 0.2 kPa VPD intervals and five bins spanning the plot-species specific SWC range. For each summarized G_{Asw} we defined a characteristic VPD and SWC as the average values of VPD and SWC of the data in the bin. The summarized values of G_{Asw} were fitted using LMM as a function of the logarithm of VPD ($\ln(\text{VPD})$) and the logarithm of SWC ($\ln(\text{SWC})$) as additive explanatory variables using uncorrelated random slopes for each species and a random intercept for each tree nested in each

species. We log-transformed the independent variables to linearise the relationships and ensure normal residuals. LMMs were fitted using the lmer function from the ‘lme4’ R package (Bates *et al.*, 2015). Using the coef function from ‘lme4’ we obtained species parameters β_{VPD} and β_{SWC} (i.e. species G_{Asw} sensitivity to VPD and to SWC, respectively), with higher values of β_{VPD} or β_{SWC} meaning stronger G reductions with increasing VPD or decreasing SWC, respectively. In addition, a reference G_{Asw} (G_{REF}) characterizing water use under standard conditions for each species, was predicted setting VPD = 1 kPa and SWC = 0.5 m³ m⁻³ (which is close to the average maximum of all sites). Complementary models were also fitted following the same procedure but with REW instead of SWC, so that soil water content variability was normalized across sites. Additional models were also fitted using canopy stomatal conductance (G'_{Asw}) as response variable. Finally, to remove parameter outliers, species with G_{REF} , β_{VPD} or β_{SWC} outside the 99.9 percentile of their normal distribution were excluded from subsequent analyses (2 out of 142 species excluded).

5.2.7 Statistical analyses

We tested whether water use regulation traits (G_{REF} , β_{VPD} and β_{SWC}) differ between angiosperms and gymnosperms using a simple linear model. Next, we constructed bi-variate linear relationships between species' fitted parameters and species' water relations traits. Furthermore, we repeated these linear relationships by adding MAP and H as predictors and applying a stepwise model selection, to discern whether the effect of the traits remained significant once these new variables were added to the model. In all the analyses we used the number of species' tree-days as a weighting factor.

Finally, we performed a path analysis using the SEM function of the lavaan R package (Rosseel, 2012). Path analysis accounts for direct and indirect dependencies among variables. To account for the coordinated effect of the species' relations traits and to maximize the number of species (106 species), we imputed species' trait missing values (Table D2) using the imputePCA function of the package missMDA (Josse & Husson, 2016) and then performed a Principal Component Analysis (PCA) to extract the two main principal components (Fig. D1). A single path model was built including the three parameters describing G_{Asw} behaviour (G_{REF} , β_{VPD} and β_{SWC}) as response variables and using MAP, H and the two dimensions of the traits' PCA as explanatory variables. In addition to direct relationships, indirect effects of MAP and H on the fitted parameters were also included through their effect on the PCA dimensions (Liu *et al.*, 2019). We also accounted for the effect of MAP on H . We included the number of tree-days per species as a weighting factor in the model. We performed a model selection procedure to include only paths with at least moderately strong support ($P < 0.1$). Finally, we also checked that the fit of the final model was not significantly different from the saturated model using the lavTestLRT function of the lavaan R package (Rosseel, 2012) with Satorra & Bentler (2001) approximation. All variables were standardized before fitting the path models. All the analyses of the study were performed in R3.6.1 (R Core Team, 2017).

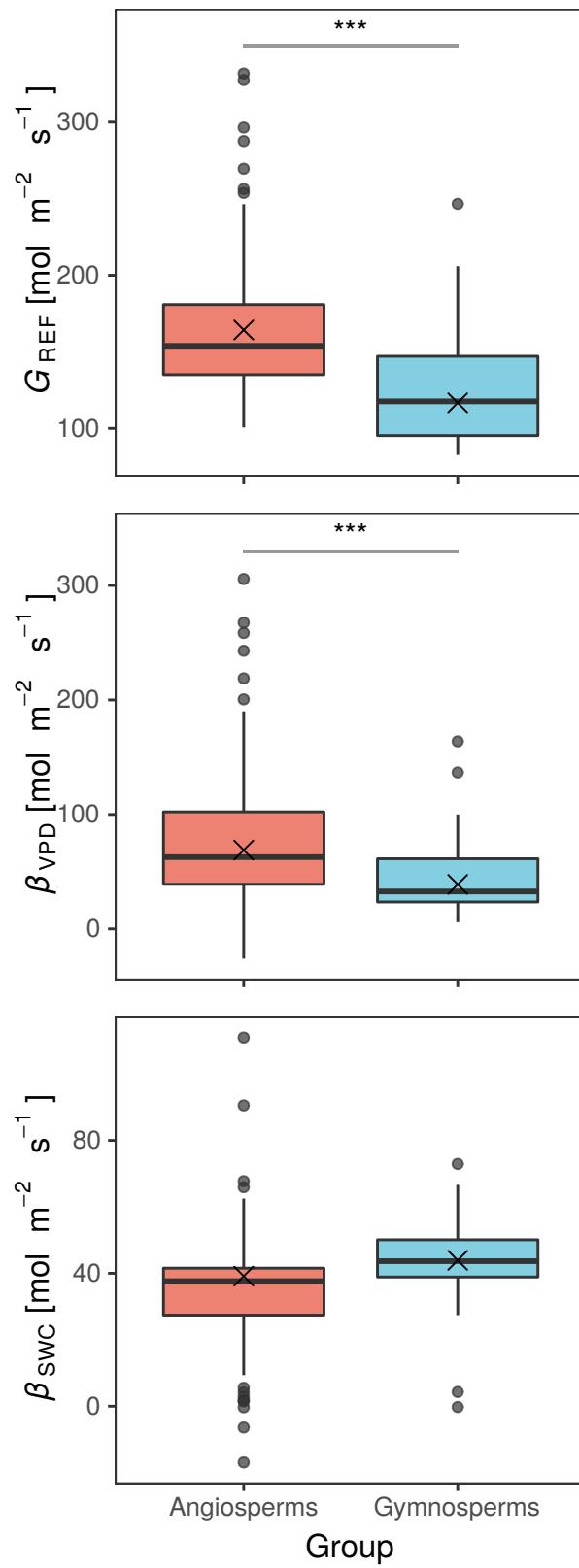


Figure 5.2: Boxplots of water use parameters for both Angiosperms and Gymnosperms. Crosses are weighted means of the parameters.

5.3 Results

5.3.1 Water use parameters and differences between angiosperms and gymnosperms

The model used to obtain water use parameters explained a 59.8% of the total conditional variance in the original data. Reference conductance (G_{REF}) across species ranged between 82.4 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Juniperus monosperma*) and 333 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Acacia longifolia*), β_{VPD} between -26 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Avicennia marina*) and 306 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Ulmus americana*) and β_{SWC} between -17.3 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Acer saccharum*) and 112 mol m $_{\text{Asw}}^{-2}$ s $^{-1}$ (*Acacia longifolia*) (Fig. D2, D3, D4). Most species showed declining G with increasing VPD (positive β_{VPD}) and increasing G with increasing SWC (positive β_{SWC}).

Species showing opposite responses were two temperate (*Acer saccharum*, *Quercus petraea*), one tropical (*Ampelocera macrocarpa*) and a mangrove (*Avicennia marina*) species that showed negative sensitivities to one of the variables (Fig. D2). Mean G_{REF} and β_{VPD} were significantly higher for angiosperms than for gymnosperms (Fig. 5.2). However, there were no differences in β_{SWC} between angiosperms and gymnosperms. Across all species (including angiosperms and gymnosperms), G_{REF} and β_{VPD} were strongly and positively correlated ($r = 0.75$; with a weighted slope of 0.67; Fig. D5), being the species with high G_{REF} more sensitive to VPD, but strong correlations were not found between G_{REF} and β_{SWC} or between β_{VPD} and β_{SWC} ($|r| < 0.34$ in both cases). When comparing water use parameters calculated considering and not considering aerodynamic conductance, we found that G_{REF} was strongly correlated to G'_{REF} ($r = 0.8$); however, β_{VPD} and β'_{VPD} were poorly correlated ($r = 0.26$) while β_{SWC} and β'_{SWC} showed no significant correlation.

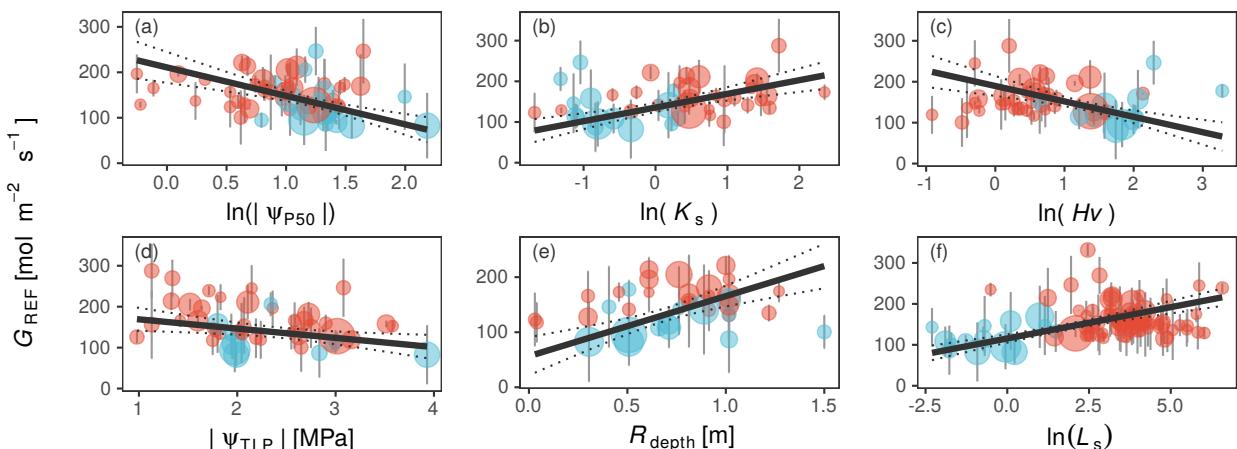


Figure 5.3: Bi-variate relationships between G_{REF} water relations traits. Individual water relation traits are shown in different panels: (a) logarithm of absolute water potential at 50% water conductivity loss, (b) logarithm of maximum sapwood water conductivity, (c) logarithm of Huber value, (d) absolute water potential at turgor-loss point, (e) rooting depth and (f) logarithm of individual leaf area. Black continuous lines depict significant linear relationships (Table 5.2) and the dashed lines represent the 95% confidence intervals of the models. Red dots are angiosperms and blue dots are gymnosperms. Size of the points is equivalent to the number of tree-days of the species. Vertical lines are the posterior standard deviation of the parameters calculated using REsim function of merTools package (Knowles & Frederick, 2016).

Table 5.2: Results of the bi-variate linear models relating water use parameters (G_{REF} , β_{VPD} , β_{SWC}) and water relations traits. Parameters are explained by individual traits using simple linear models using number of species-days as weighting factor.

| Water use parameter | Water relations Traits | N species | Intercept | Slope | R^2 |
|----------------------|------------------------|-----------|------------|-------------|-------|
| G_{REF} | $\ln(\Psi_{P50})$ | 55 | 210.701*** | -62.724 *** | 0.282 |
| | $\ln(K_s)$ | 43 | 135.527*** | 33.614 *** | 0.338 |
| | $\ln(Hv)$ | 49 | 189.439*** | -37.602 *** | 0.285 |
| | $ \Psi_{\text{TLP}} $ | 48 | 191.764*** | -22.702 * | 0.112 |
| | R_{depth} | 37 | 56.204** | 109.296 *** | 0.366 |
| | $\ln(L_s)$ | 86 | 115.537*** | 15.28 *** | 0.391 |
| β_{VPD} | $\ln(\Psi_{P50})$ | 55 | 109.201*** | -47.716 *** | 0.240 |
| | $\ln(K_s)$ | 43 | 50.199*** | 20.427 ** | 0.180 |
| | $\ln(Hv)$ | 49 | 96.503*** | -33.817 *** | 0.363 |
| | $ \Psi_{\text{TLP}} $ | 48 | 80.321*** | -12.209 . | 0.040 |
| | R_{depth} | 37 | -2.190 ns | 80.383 *** | 0.260 |
| | $\ln(L_s)$ | 86 | 34.577*** | 11.219 *** | 0.327 |
| β_{SWC} | $\ln(\Psi_{P50})$ | 55 | 16.103 ns | 19.327 * | 0.059 |
| | $\ln(K_s)$ | 43 | 42.113*** | -5.021 ns | 0.000 |
| | $\ln(Hv)$ | 49 | 23.952** | 13.586 * | 0.088 |
| | $ \Psi_{\text{TLP}} $ | 48 | 40.468** | 1.052 ns | 0.000 |
| | R_{depth} | 37 | 53.851*** | -28.471 . | 0.051 |
| | $\ln(L_s)$ | 86 | 51.112*** | -4.095 ** | 0.087 |

Statistical significant levels: " ." p<0.1 ; " * " p<0.05; " ** " p<0.01; " *** " p<0.001; ns not significant.

5.3.2 Coordination with hydraulic and allocation traits

In the bi-variate models relating G_{REF} with hydraulic and morphological traits, we found that G_{REF} showed a negative relationship with $|\Psi_{P50}|$, Hv and $|\Psi_{\text{TLP}}|$ (Table 5.2 and Fig. 5.3(a,c,d)), whereby species more resistant to embolism, with higher allocation to the sapwood relative to leaves and with more negative turgor-loss pressures showed lower G_{REF} . Furthermore, G_{REF} was positively related to K_s , R_{depth} and L_s (Table 5.2 and Fig. 5.3(b,e,f)), with species with efficient xylem, deeper roots and bigger leaves showing higher G_{REF} . For these traits, L_s was the one explaining the largest fraction of G_{REF} variability ($R^2 = 0.39$). With the exception of Ψ_{TLP} , these relationships remained significant also for G'_{Asw} (Table D3).

β_{VPD} was negatively related with $|\Psi_{P50}|$ and Hv and positively with K_s (Table 5.2 and Fig. 5.4(a,b,c)), i.e., species with less safe and more efficient xylem present higher sensitivity to VPD, and hence more strict stomatal control as atmospheric water demand increases. β_{VPD} was also positively related to R_{depth} and L_s (Table 5.2 and Fig. 5.4(e,f)), indicating that deeper roots and larger leaves were associated to higher stomatal sensitivity to VPD. Absolute turgor-loss point ($|\Psi_{\text{TLP}}|$) was weakly negatively related with β_{VPD} (p value = 0.093; Table 5.2). Hv and L_s were the traits explaining most of β_{VPD} variability ($R^2 = 0.36$ and $R^2 = 0.33$, respectively). With the exception of Ψ_{TLP} , these relationships remained significant also for G'_{Asw} (Table D3).

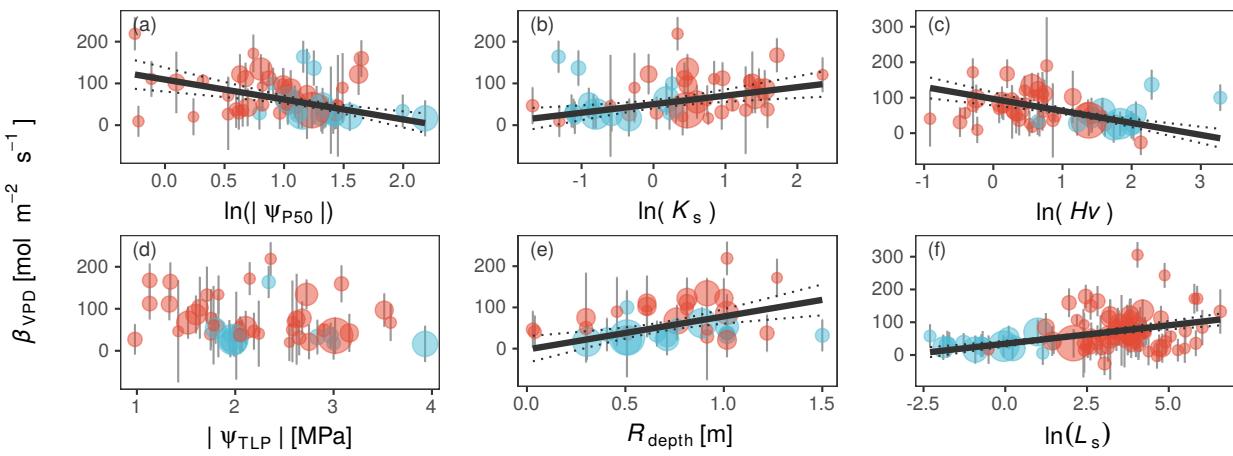


Figure 5.4: Bi-variate relationships between β_{VPD} water relations traits. Individual water relation traits are shown in different panels: (a) logarithm of absolute water potential at 50% water conductivity loss, (b) logarithm of maximum sapwood water conductivity, (c) logarithm of Huber value, (d) absolute water potential at turgor-loss point, (e) rooting depth and (f) logarithm of individual leaf area. Black continuous lines depict significant linear relationships (Table 5.2) and the dashed lines represent the 95% confidence intervals of the models. Red dots are angiosperms and blue dots are gymnosperms. Size of the points is equivalent to the number of tree-days of the species. Vertical lines are the posterior standard deviation of the parameters calculated using REsim function of merTools package (Knowles & Frederick, 2016).

β_{SWC} was positively related to $|\Psi_{\text{P}50}|$ and Hv (Table 5.2 and Fig. 5.5(a,c)), i.e., species with higher resistance to embolism and larger ratios of sapwood to leaf area were more sensitive to soil water depletion. In addition, species with larger leaves (L_s) and, marginally (p value = 0.095), with deeper roots (R_{depth}) were less sensitive to soil water stress (Table 5.2 and Fig. 5.5(e,f)). In general, water relations traits explained a lower proportion (at most 9%) of the variability in β_{SWC} than of G_{REF} or β_{VPD} . However, we should treat relationships between β_{SWC} and $|\Psi_{\text{P}50}|$, Hv and R_{depth} with caution, since they all become non-significant when aerodynamic conductance was taken into account (Table D3) or when REW was used instead of SWC (Table D3). Furthermore, when REW was used instead of SWC, all soil moisture sensitivity-trait relationships became non-significant (Table D4).

Ecological factors associated with water use parameters and coordination When the coordination between water use parameters and water relations traits was assessed while also accounting for the effects of MAP (mean annual precipitation) and H (tree species-level mean height), most of the relationships described in the previous section remained significant, with only three exceptions. The relationships that were no longer observed corresponded to the effect of Ψ_{TLP} on G_{REF} and the effects of $|\Psi_{\text{P}50}|$ and Hv on β_{SWC} (Table 5.3). In these models, MAP was the variable that explained most of the variability in stomatal responses to soil water (β_{SWC}), with generally lower sensitivity to SWC in locations with high MAP (Table 5.3), although this effect reversed in the L_s model. However, when models were calculated using β'_{SWC} , MAP effects were all negative (Table D5). On the other hand, MAP was largely unrelated to G_{REF} and β_{VPD} in most of the G_{REF} and β_{VPD} models (Table 5.3). Finally, our results show that taller trees tend to have higher G_{REF} and higher sensitivity to VPD (β_{VPD}), however, when G_{REF} was obtained from G'_{Asw} (i.e. G'_{REF}), H was not selected in the final models (Table D5). The relationship between H and soil drought sensitivity (β_{SWC}) was less clear, since it was significantly negative in only two of the

Table 5.3: Results of the bi-variate linear models relating water use parameters (G_{REF} , β_{VPD} , β_{SWC}), water relations traits, climate and tree height. Water use parameters are explained using simple linear models with number of species-days as weighting factor. β column values are the slopes for each explanatory variable. N species = number of species included. NI = not included variable after model selection.

| Water use parameter | Water relations Traits | N species | Intercept | β_{trait} | β_{MAP} | β_{H} | R^2 |
|----------------------|------------------------|-----------|-------------|------------------------|----------------------|--------------------|-------|
| G_{REF} | $\ln(\Psi_{P50})$ | 54 | 173.6 *** | -48.549 ** | NI | 1.418 * | 0.325 |
| | $\ln(K_s)$ | 42 | 109.773 *** | 30.731 *** | NI | 1.822 * | 0.423 |
| | $\ln(Hv)$ | 43 | 162.178 *** | -32.21 ** | NI | 1.448 . | 0.332 |
| | $ \Psi_{\text{TLP}} $ | 47 | 66.861 *** | NI | 0.057 * | 1.962 * | 0.346 |
| | R_{depth} | 36 | 41.932 * | 88.301 *** | NI | 1.802 * | 0.448 |
| | $\ln(L_s)$ | 80 | 98.531 *** | 13.389 *** | NI | 1.384 ** | 0.450 |
| β_{VPD} | $\ln(\Psi_{P50})$ | 54 | 70.665 *** | -32.97 ** | NI | 1.468 * | 0.317 |
| | $\ln(K_s)$ | 42 | 25.16 * | 17.444 ** | NI | 1.768 ** | 0.303 |
| | $\ln(Hv)$ | 43 | 73.255 *** | -28.931 *** | NI | 1.197 * | 0.408 |
| | $ \Psi_{\text{TLP}} $ | 47 | 16.363 . | NI | NI | 2.502 *** | 0.291 |
| | R_{depth} | 36 | -18.282 ns | 56.571 * | NI | 2.034 ** | 0.412 |
| | $\ln(L_s)$ | 80 | 16.099 * | 8.955 *** | NI | 1.512 *** | 0.429 |
| β_{SWC} | $\ln(\Psi_{P50})$ | 54 | 62.606 *** | NI | -0.029 * | NI | 0.088 |
| | $\ln(K_s)$ | 42 | 73.319 *** | NI | -0.038 ** | NI | 0.142 |
| | $\ln(Hv)$ | 43 | 50.25 ** | 10.221 ns | -0.026 . | NI | 0.149 |
| | $ \Psi_{\text{TLP}} $ | 47 | 55.33 *** | NI | NI | -0.903 * | 0.067 |
| | R_{depth} | 36 | 62.889 *** | NI | -0.037 ** | NI | 0.213 |
| | $\ln(L_s)$ | 80 | 44.671 *** | -5.002 ** | 0.025 * | -0.749 * | 0.142 |

Statistical significant levels: " ." p<0.1 ; ** p<0.05; *** p<0.01; **** p<0.001; ns not significant.

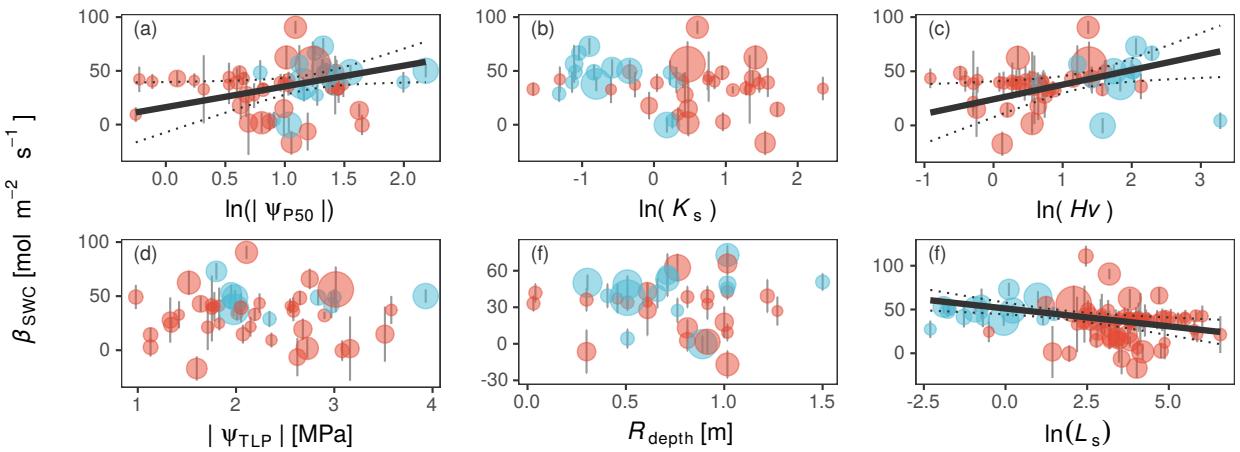


Figure 5.5: Bi-variate relationships between β_{SWC} water relations traits. Individual water relation traits are shown in different panels: (a) logarithm of absolute water potential at 50% water conductivity loss, (b) logarithm of maximum sapwood water conductivity, (c) logarithm of Huber value, (d) absolute water potential at turgor-loss point, (e) rooting depth and (f) logarithm of individual leaf area. Black continuous lines depict significant linear relationships (Table 5.2) and the dashed lines represent the 95% confidence intervals of the models. Red dots are angiosperms and blue dots are gymnosperms. Size of the points is equivalent to the number of tree-days of the species. Vertical lines are the posterior standard deviation of the parameters calculated using REsim function of merTools package (Knowles & Frederick, 2016).

models (Table 5.3), suggesting higher sensitivity to soil drought in shorter trees. However when aerodynamic coupling was taken into account, the relationship was inverted and taller trees had higher sensitivity to SWC (Table D5).

The two main dimensions of the PCA analyses describing water relations trait coordination explained a 69.8% of the total variance (Fig. D1). The primary PCA dimension (Dim1; 56.8% of variance) could be interpreted as a safety-efficiency trade-off axis, whereby positive values are related to elevated K_s , large L_s , deep roots, low Hv and low $|\Psi_{P50}|$. The second PCA dimension (Dim2; 13% of variance) was associated to leaf turgor-loss pressure (Ψ_{TLP}), with positive values related to high $|\Psi_{TLP}|$ levels and, to a lower extent, deeper roots.

In the path analyses, efficiency traits (positive PCA Dim1 values) were significantly related to high annual precipitation (high MAP) and to taller trees (large H) (Fig. 5.6). Also, H increased with MAP. G_{REF} was positively associated with Dim1 (Fig. 5.6) and taller trees also had marginally higher G_{REF} (Fig. 5.6). Similarly, higher β_{VPD} (i.e., higher VPD sensitivity) was positively related to efficient water transport (Dim1) and to H (Fig. 5.6). Sensitivity to SWC (β_{SWC}) showed a marginal, negative relationship with Dim1, so that sensitivity increased with xylem resistance to embolism (Fig. 5.6). Finally, G_{REF} co-varied positively with β_{VPD} and β_{SWC} , implying that as G_{REF} increases so do β_{VPD} and β_{SWC} (Fig. 5.6, Fig. D5). The second dimension of the PCA was not included in the final path model, suggesting a lack of coordination between Ψ_{TLP} and water use parameters.

5.4 Discussion

In this study, we present a novel analysis linking organ-level traits with whole-plant water use strategies at the global scale, made possible by the compilation of the first global sap flow database.

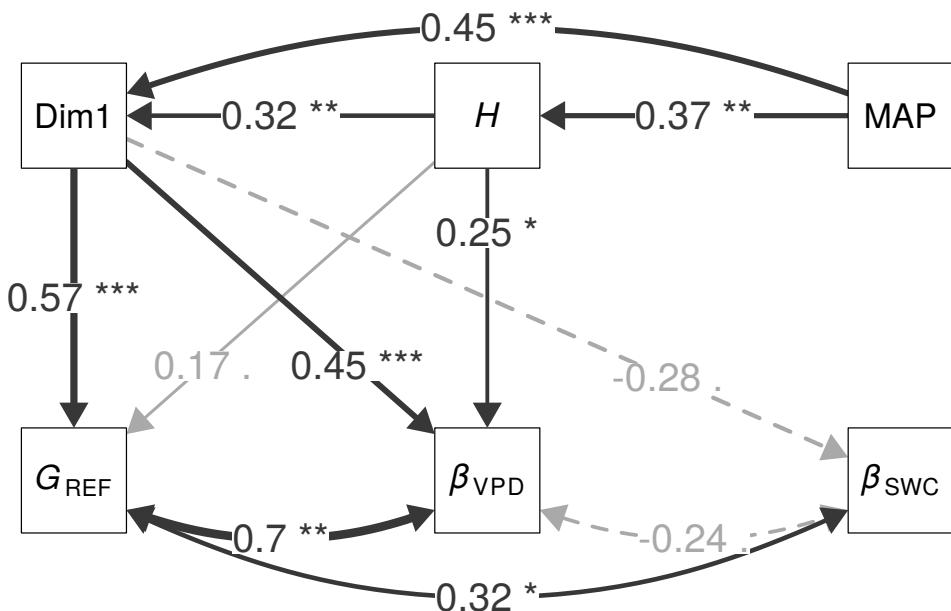


Figure 5.6: Path analyses of species-specific water use parameters explained by mean annual precipitation (MAP), tree height (H) and coordinated hydraulic traits Dim1. Dim1 is the hydraulic traits' PCA dimension 1 (Fig. D1). Positive Dim1 values are mainly related to efficient water use strategies, while negative to safety strategies. Dim2 was not selected in the final model. Arrow labels are standardized parameters. Continuous lines are positive relationships while dashed lines are negative relationships. Black and grey lines are significant and marginally significant relationships, respectively. Statistical significant levels: ., $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$

We provide evidence of a coordination between water relations traits and water use parameters (G_{REF} , β_{VPD} and β_{SWC}), while accounting for the effects of climate and tree size. Some water use and trait associations were explained by climate affiliations of species, but most relationships between water use and water relations traits remained after accounting for climate and tree size effects. As any synthesis effort of this magnitude based on diverse data sources, our study presents several limitations. First, sap flow data used to estimate G carry some uncertainty issues, although these may be less relevant for assessing environmental responses as we do here than for characterizing absolute values (Flo *et al.*, 2019). Second, SAPFLUXNET may have an incomplete coverage of global forest ecosystems (Fig. 5.1, Poyatos *et al.*, 2020b). Third, highly non-linear or threshold-based SWC responses may be difficult to capture, especially when having to resort to reanalysis data. Fourth, other ecological processes such as partial canopy-atmosphere coupling or intra-specific variability in traits and/or water use regulation may influence our results.

5.4.1 Climate influence on water use strategies across species

Our results showed no direct effect of mean annual precipitation (MAP) on G_{REF} and β_{VPD} , but instead indirect MAP effects on water use strategies mediated through hydraulic and allocation traits (Fig. 5.6 and Fig. D6), suggesting that MAP constrains feasible water relations traits (Bourne *et al.*, 2017; Liu *et al.*, 2019), which then directly determine water use rate and β_{VPD} . The direction of these effects is consistent with previous studies showing that β_{VPD} increased with aridity in rainforest species (Cunningham, 2004; but see Grossiord *et al.*, 2019) and at continental scales across ecosystems and functional types (Novick *et al.*, 2016), but these studies did not

disentangle direct from indirect effects. Although the global controls on β_{SWC} were less clear in our analyses, in part due to the influence of aerodynamic coupling (cf. Table 5.3 and D5), climate effects on this variable also appeared to be largely indirect (Fig. 5.6). Therefore, our results underscore the importance of using water relations traits, rather than climate when addressing species whole-tree water use strategies and ecosystem flux sensitivities to VPD.

5.4.2 Water use parameters

Water use parameters differed widely among species (Fig. D2) and defined a gradient of water use sensitivities to drought stress. Within the gradient of parameters, angiosperms and gymnosperms showed distinct whole-plant water use strategies (Fig. 5.2). We found that gymnosperms have generally more ‘conservative’ water use strategies in terms of lower G_{REF} but not in terms of enhanced sensitivity to VPD or SWC. Similarly, previous studies also showed lower sensitivity to VPD in gymnosperms compared to angiosperms (Johnson *et al.*, 2012; Lin *et al.*, 2015), which could be associated to higher safety margins (Choat *et al.*, 2012; Anderegg *et al.*, 2016). The ‘conservative’ G_{REF} strategy of gymnosperms could be explained by group-specific trait syndromes associated to water relations traits (Fig. D8), wood anatomy (Venturas *et al.*, 2017), and lower photosynthetic rates, stomatal conductance or leaf N concentrations (Lusk *et al.*, 2003).

G_{REF} and β_{VPD} showed a strong positive correlation across species (Fig. D5) similar to the one found by Oren *et al.* (1999b). This implies stronger VPD control on transpiration in species with higher water use under optimal conditions (higher G_{REF}). However, our global cross-species analyses might mask finer variations, as β_{VPD} (or $\beta_{\text{VPD}} / G_{\text{REF}}$) is expected to be lower across species from dry sites (Oren *et al.*, 1999b) or along a decreasing gradient of SWC within species (but see Poyatos *et al.*, 2007; Domec & Johnson, 2012; Zhang *et al.*, 2012). Nevertheless, this result suggests that G_{REF} can be a suitable proxy of whole-tree canopy conductance sensitivity to VPD, as it is at the leaf (Oren *et al.*, 1999b) or ecosystem (Grossiord *et al.*, 2020) levels.

The lack of correlation found between the sensitivity to SWC calculated with and without aerodynamic conductance, indicates that canopy coupling could be important in calculating β_{SWC} and that in order to predict and model plant water responses to soil water dynamics, we likely have to explicitly consider aerodynamic and boundary layer conductances. In addition, these land-atmosphere interactions might be crucial in diagnosing and modelling the soil moisture controls over other ecosystem processes such as the carbon cycle (Green *et al.*, 2019; Kannenberg *et al.*, 2020).

5.4.3 Coordination between water use parameters and water relations traits

Our results support the hypothesis of a strong coordination between G_{REF} and individual hydraulic and allocation traits. G_{REF} aligns with ‘efficiency’ traits (K_s) in the hydraulic safety-efficiency axis, and is thus negatively related to ‘safety’ traits (particularly Ψ_{P50}). These results are consistent with the overall proposed coordination between plant hydraulics and gas exchange (Meinzer, 2002; Sperry *et al.*, 2002; Mencuccini, 2003; Maherali *et al.*, 2006; Henry *et al.*, 2019) and with the notion that species operate close to their maximum transport capacity sustained by their hydraulic system (Manzoni *et al.*, 2013). Large individual leaf areas were also related to higher G_{REF} , probably due to higher leaf hydraulic conductance mediated by wider conduits (Schreiber *et al.*, 2016; Ding *et*

al., 2020). The positive association of elevated G_{REF} with deeper roots points out the requirement of deep rooting to supply water for keeping high transpiration rates, and is also found in the coordination between R_{depth} , $\Psi_{\text{P}50}$ and K_s (Mursinna *et al.*, 2018).

Coordination between whole-tree water use sensitivity to VPD (β_{VPD}) and to SWC (β_{SWC}) with organ-level water relations traits had not been assessed before at a global scale. All the studied water relations traits (except Ψ_{TLP}) appear to be related to β_{VPD} , whereby the species with more ‘efficient’ or less “safe” traits tend to be those which show higher β_{VPD} . These results are consistent with previous studies relating stomatal responses and water relation traits (Lu *et al.*, 2020) and with the stomatal gas exchange optimization theory (see Tyree & Sperry, 1988; Wang *et al.*, 2020). By contrast, after controlling for climate and tree height, conductance sensitivity to SWC (β_{SWC}) was only (negatively) related to L_s . Notably, β_{SWC} was unrelated to Ψ_{TLP} , in contrast to Maréchaux *et al.* (2018), that evidenced more negative Ψ_{TLP} related to lower reductions in sap flow with decreasing SWC. This absence of relationship, including the weak $\beta_{\text{VPD}} - \Psi_{\text{TLP}}$ correlation, could be attributed to noise and uncertainty in Ψ_{TLP} measures (Meinzer *et al.*, 2014) or with Ψ_{TLP} plasticity (Bartlett *et al.*, 2012; Rosas *et al.*, 2019). In addition, the lack of relationship between β_{SWC} and R_{depth} could be explained by the complexity of rooting depth dependency on soil water infiltration, tree height and climate (Fan *et al.*, 2017).

We also explored the coordinated effect of water relations traits on water use parameters and accounted for direct and indirect climate and tree height effects through the PCA and the path model. Based on our results, water use strategies would be, in terms of G_{REF} , consistent with the Reich (2014) notion of a whole-plant resource use spectrum, ranging from ‘conservative’ to ‘acquisitive’ species. However, in terms of absolute sensitivity to VPD, our results go against this idea, since acquisitive species (with high G_{REF}) are also more sensitive to VPD (more ‘conservative’). Therefore, our study would support a more physiological interpretation of water use strategies that stresses trait coordination. According to this interpretation, plants with ‘safer’ hydraulic systems (high resistance to embolism) are able to function at higher water tensions without requiring a strict water use regulation, implying that they can show a more ‘acquisitive’ regulation of water use so that they can benefit from having a wider range of conditions to operate safely. In other words, high transport capacity in the xylem (K_s) is associated with high canopy conductance (G_{REF}) and a vulnerable (sensitive) xylem is also associated with a stricter regulation of gas exchange. However, a vulnerable xylem reduces safety, and is usually interpreted as part of an ‘acquisitive’ strategy, whereas a strict regulation of water use prevents hydraulic failure and hence corresponds to a ‘conservative’ strategy. This view is also consistent with the positive relationship between $\Psi_{\text{P}50}$ and Ψ_{TLP} , even if it saturates at relatively low water potentials (cf. Martin-StPaul *et al.*, 2017). These results highlight the complications of defining a single, whole-plant resource use spectrum ranging from ‘acquisitive’ to ‘conservative’ species (*sensu* Reich, 2014), and points to the need of considering different organs and functional axes when assessing whole-plant functional integration.

Regarding tree’s height, it was coordinated directly and indirectly –through water relations traits– with water use parameters (Table 5.3 and Fig. 5.6), in a way that taller trees displayed more ‘efficient’ water use strategies. Alignment of water relations traits and H was consistent with results found by Liu *et al.* (2019), relating maximum plant size with $\Psi_{\text{P}50}$, K_s or H_v at the global

scale across species and life forms. These complex direct and indirect H relationships might be driven by ecosystem water availability (Fig. 5.6) and low freezing risk (Olson *et al.*, 2018), which allows for increased water use through efficient water transport (e.g. high K_s or Hv), compensating the increase of resistance due to the enlarged water path of taller trees (Barnard & Ryan, 2003; Liu *et al.*, 2019; Mencuccini *et al.*, 2019b). Furthermore, H could also affect differential sensitivity to VPD and SWC in tall trees (Giardina *et al.*, 2018), as their canopy would be more exposed to VPD, requiring higher β_{VPD} , and would potentially have more developed root systems, which would decrease β_{SWC} .

5.4.4 Conclusions

Understanding tree water use strategies at the global scale is crucial to better predict ecosystem water cycles and drought vulnerability of species and ecosystems. Here we demonstrate that there is a global spectrum of water use strategies determined by the coordination of hydraulic and allocation traits, rather than by climate. In particular, species-specific G_{REF} and β_{VPD} (but not β_{SWC}) are closely related to the species-specific water relations traits. We have also shown significant differences between angiosperm and gymnosperm water use strategies, showing greater water use and sensitivity to VPD in angiosperms than gymnosperms, a finding that could be related to distinct water relations traits syndromes (Fig. D8). Our trait-based approach allowed for a simplified global mapping of water use strategies. The use of simple measurable traits (e.g. leaf size) altogether with functional grouping can lead to a better approximation of species reference water conductance and its sensitivity to VPD. Recently developed global maps of traits (Moreno-Martínez *et al.*, 2018; Trugman *et al.*, 2020) would permit the inclusion of such water use strategies in Land Surface and Earth System Models potentially improving ecosystem carbon and water fluxes predictions.

6

General discussion and conclusions

Despite the fact that forest transpiration is the main terrestrial evaporative flux and a key component of the global hydrological cycle and Earth System functioning, its quantification is still replete with uncertainty (Schlesinger & Jasechko, 2014). Our knowledge of how trees control transpiration in response to hydrometeorological conditions worldwide remains incomplete. Transpiration regulation and water use strategy under drought conditions determine individual survival (Choat *et al.*, 2018), which extends to forest function and dynamics across the globe, impacting the provision of forest ecosystem services such as carbon sequestration, hydroclimatic regulation and biodiversity conservation (Trumbore *et al.*, 2015). Thus, a better understanding of transpiration and its spatiotemporal variability will improve assessments of vegetation dynamics and climate feedbacks at regional to global scales, especially in the face of global change (Choat *et al.*, 2018), which is increasing drought episodes and their severity in several regions (Dai, 2013). Along this thesis I have tried to give a global perspective on the ecohydrological variables driving the regulation of transpiration using sap flow data at the whole-tree level. Due to the lack of a global and harmonized database of sap flow measurements in the past, this sort of synthesis had not been attempted before. However, the launch of the SAPFLUXNET database, to which I have contributed, has made this possible. Because a global, harmonized sap flow database requires comparability across methods, the second chapter of this thesis deals with calibration uncertainty of different sap flow techniques.

6.1 Improving sap flow techniques to obtain reliable estimates of tree level transpiration

To reduce the uncertainty in our knowledge of plant water transport processes, it is essential to first reduce the uncertainty in the measurements of plant water flow. All sap flow methods have specific limitations (Smith & Allen, 1996; Vandegehuchte & Steppe, 2013) and can yield biased estimates, which implies that this bias needs to be taken into account when sap flow data are used in quantitative analyses (Chapter 2). In particular, I show that heat dissipation techniques, which are by far the most widely used method, consistently underestimate sap flow by 40% on average (if used with its original calibration coefficients; Chapter 2). At the same time, all methods provide high average correlations between sap flow measurements and actual values, suggesting that they are suitable for studies based on qualitative or relative values, including research on the relative importance of the hydrometeorological drivers explaining canopy conductance (Chapter 4), or on the sensitivity of transpiration to hydrometeorological drivers (Chapter 5).

To improve the quality of plant water flow estimates obtained using sap flow methods, further efforts are needed to develop and follow transparent protocols for each method, including protocols for installation; homogenization of raw data processing, and quantification of uncertainty (see Peters *et al.*, 2020). Also, calibration of probes is essential whenever possible; otherwise, applying generic or species-specific correction factors for each method may be an alternative solution, although less accurate. To improve method-specific correction factors, it would be important to identify sapwood traits that might explain measurement errors, such as wood density, or anatomical and vessel distribution traits (Suleiman *et al.*, 1999; Wullschleger *et al.*, 2011). However, so far attempts to identify the potential effect of those wood traits on sap flow measurement errors have not been entirely successful (Peters *et al.*, 2018, Chapter 2), possibly due to the fact that the

actual traits are rarely measured on the same plant material used in the calibrations. Studying these wood traits in the context of sap flow calibrations for a range of species with different sapwood anatomies would allow us to understand their potential influence on sap flow measurement uncertainty, and to obtain correction factors for all species without the need for calibration. Besides, scaling of single-point measurements to the whole-tree level constitutes another major source of uncertainty (Hernandez-Santana *et al.*, 2015) that cannot be easily addressed with correction factors. Sap flow integration requires to consider the tree sapwood area and its radial profile, whose quantification carries a lot of uncertainty even with destructive tree sampling (Hernandez-Santana *et al.*, 2015). Nonetheless, the scaling error could be minimized by taking more than several sap flow measurements along the perimeter and radial profile of the plant, and estimating sapwood area more accurately by using on-site robust species allometrical relationships or multiple core samples of the measured individuals.

Improving the accuracy of sap flow methods has been a major research goal for decades. For this purpose, specific corrections have been implemented, for example, for dealing with wounding effects or for fixing signal dampening due to installation time (Swanson & Whitfield, 1981; Green *et al.*, 2003; Peters *et al.*, 2018). In addition, new probes configurations and sap flow techniques have been developed. Some of the technologies currently being designed are promising, such as those based on dual techniques applying different methods throughout the flow range (Forster, 2019), on single-probe methods to minimize wound (López-Bernal *et al.*, 2017), or on developing integrated, compact and low cost systems (Jones *et al.*, 2020). Besides the traditional thermometric methods, some attempts have been made using other approaches such as acoustic emissions (Cerný *et al.*, 2011; Mazal *et al.*, 2012), or thermal imaging (Anfodillo *et al.*, 1993), but they have not been entirely successful. Despite the uncertainties associated to all thermometric sap flow methods, they remain the best tool we have at hand to estimate transpiration dynamics at the tree-level, and the only solid basis for the compilation of a global sap flow database (Poyatos *et al.*, 2016).

6.2 The SAPLUXNET database. What next?

Over the course of the thesis, the SAPFLUXNET database has become a reality (Poyatos *et al.*, 2020b), and it is currently available and open access at ZENODO (Poyatos *et al.*, 2020c) since 2019. Additionally, we have built a data infrastructure capable of processing and analysing large amounts of sap flow related data, which is prepared for the inclusion of new data sets if further requests for data contributions are reopened in the future. SAPFLUXNET has gained increased attention from ecophysicists and ecohydrologists, and by the end of 2020 there have been around 1500 downloads of the data base. As discussed in Chapter 3, SAPFLUXNET and its combination with existing datasets, such as FLUXNET (Pastorello, 2020) or HydraTRY (Mencuccini *et al.*, 2019b; Sanchez-Martinez *et al.*, 2020) or remote sensing products such as GLEAM (Martens *et al.*, 2017), will help us to better characterize plant drought responses and forest transpiration across the globe. Also, in conjunction with, for example, the Tree-Ring Data Bank (Grissino-Mayer & Fritts, 1997) or remote sensing data (Simard *et al.*, 2011), SAPFLUXNET will improve the characterization of drought legacy effects and recovery of water use after drought, potentially bridging the gap between water use, tree growth and ecosystem productivity. This thesis is an example of the kind of studies that can be accomplished taking advantage of SAPFLUXNET. In particular, I carried out the first

quantification of the importance of hydroclimatic drivers controlling tree transpiration globally, and also characterized tree water use strategies across species that emerge from the covariation between water use regulation and hydraulic traits.

6.3 The hydroclimatic drivers of transpiration and their complexity

The closure of stomata to prevent water loss through transpiration responds to hydroclimatic changes at temporal scales that range from minutes to seasons (Buckley, 2005). As a result, plant transpiration is coupled to some extent to hydroclimatic variables (i.e. vapour pressure deficit, soil water content and radiation). This coupling allows plants to cope with drought, and may differ depending on environmental conditions, water use strategy and legacy effects. I have shown in this thesis that vapour pressure deficit (VPD) is the main driver of transpiration globally. This result may be explained by the different temporal scale at which changes in VPD, soil water content (SWC) and radiation occur. Stomatal closure might be tightly and promptly coupled to VPD to protect plant hydraulic integrity under abrupt dry atmospheric conditions, while the dynamics of soil moisture supply are slower, allowing to maintain transpiration rates by, for example, exploring and extracting water from deeper layers of the soil as drought progresses (Barbeta & Peñuelas, 2017). By contrast, although radiation (PPFD) has similar temporal scale dynamics as VPD, it is less coupled to plant transpiration. This may be explained by the direct relationship of the VPD with water flows and plant water status, especially in well-coupled canopies, while the function of PPFD as a G driver would be a more complex process produced via photosynthesis. This direct effect can be found in most ecosystem-level G models which show better results using only VPD (Lin *et al.*, 2018), although VPD and solar radiation are strongly coupled at most temporal scales.

The lack of coupling between transpiration dynamics and hydrometeorological variables in drylands suggested by our results might be related to complex responses of transpiration regulation to the interaction of the hydrometeorological drivers (Zhou *et al.*, 2019b), which we have not considered explicitly. Alternatively, it might be associated to the diversity of water use strategies in water limited systems that allow plants living in dry biomes to escape from drought dynamics and better compete for resources and maximize growth. These water use strategies may be related to multiple anatomical and hydraulic traits conferring drought tolerance and safety towards hydraulic failure, which is in concordance with results in Chapter 5.

The low variance explained by VPD, SWC and PPFD in some regions limits our ability to explain (and model) transpiration patterns at the global scale. An additional reason for this low explanatory power may be the simple shape of the log models of canopy conductance used in Chapter 4 -although similar explained variance was obtained by more flexible GAM models-, which may fail for some species or sites, especially for the SWC models. Thus, it would be important to further test whether the empirical couplings reported in this thesis are similar to those predicted by the main stomatal regulation algorithms used by land surface models (Mencuccini *et al.*, 2019a). In addition, other ecohydrological variables not considered in Chapter 4 could also explain transpiration regulation, such as wind speed or [CO₂]. Finally, this thesis evidences large differences in transpiration responses to hydroclimatic drivers across species and sites. The possibility to explain

these differences using measurable plant traits opens new ground towards the characterization and modelling of plant water use strategies at the global scale, a topic that I address in the final empirical chapter of the thesis (Chapter 5).

6.4 On the use of plant traits to characterize water use strategies at the global level

A better understanding of the spatial distribution of tree water use strategies will allow the improvement of climate and ecosystem models, particularly in drought-prone situations (Matheny *et al.*, 2017). Static parameters defining water use strategies for plant functional types (PFT) used in current terrestrial biosphere and dynamic vegetation models have been identified as a potential source of uncertainty due to PFT over-aggregation (Wullschleger *et al.*, 2014; Yang *et al.*, 2015). Moving to a trait-based approach would therefore improve our capacity to predict ecosystem responses, since for each parameter used in the models there would be a continuum set of trait values, one corresponding to each species (or even, potentially, to each population). Results from Chapter 5 show that plant traits related to water transport (hydraulics) and water relations explain water use regulation by plants, particularly regarding responses to VPD. These results pave the ground for the inclusion of these traits in land surface models (as already done in some cases: Kennedy *et al.*, 2019; Eller *et al.*, 2020) and for the use of these traits in up-scaling water use from the tree to the ecosystem in diverse communities. However, as shown in Chapter 4, our predictive capacity of transpiration regulation at the daily level is still relatively low for some regions, and it would be interesting to explore different temporal scales and also consider more complex models.

Following results shown in Chapter 5 and in concordance to the postulated trade-off between hydraulic safety and efficiency, trees with high water transport capacity (e.g., large xylem conduits) would tend to be more vulnerable to hydraulic dysfunction (e.g., high vulnerability to xylem embolism) and therefore would show a tight stomatal regulation to protect their hydraulic system (Taneda & Sperry, 2008). In contrast, species less conductive species with safer conduits would tend to have a less strict stomatal regulation. However, there are several examples of trees with high conductive capacity in the xylem and relatively loose stomatal regulation (Martínez-Vilalta *et al.*, 2014), which implies that other traits rather than xylem properties might be determining the water use strategy (Matheny *et al.*, 2015). These considerations suggest, as we have explored in Chapter 5, that water-use strategies at the tree level emerge from the coordination of different traits, and can only be understood if this coordination is explicitly taken into account.

Other traits would potentially be useful for describing water use strategies in addition to the hydraulic and anatomical traits explored in this thesis, in addition to the effects of climate, soil and stand structure properties (Chapter 4). Many functional traits have been recognized as important determinants of plant ecological strategies (Yang *et al.*, 2015), including specific leaf area or seed mass, which may link the plant economic spectrum hypothesis to plant hydraulics and water use strategies (Rosas Torrent *et al.*, 2019). Another important consideration is that, in order to implement this trait-based approach to define water use strategies that can be applied to land surface models, it is essential that these traits are mapped at a global scale. Global to regional maps of plant traits are currently limited to a small group of leaf (Moreno-Martínez *et al.*, 2018) or hydraulic (Trugman *et al.*, 2020) traits. However, recent studies highlighting the strong phylogenetic

conservatism of key hydraulic traits (Sanchez-Martinez *et al.*, 2020), together with the increased availability of tree distribution data (Serra-Diaz *et al.*, 2017), suggest that global maps for these traits will be available soon.

6.5 Towards a global characterization of plant water use strategies.

In the present thesis we have explored plant water use from the perspective of whole-tree transpiration regulation in response to concomitant variations in hydrometeorological forcing. However, we have left out important aspects to understand and define the water use strategies of trees. For instance, in Chapter 5 we focus on xylem hydraulics without explicitly considering the hydraulics of leaves and roots, which are important determinants of plants' water status and the drought sensitivity of whole-plant hydraulic conductance. Accounting for these different organs will help integrate the internal and external conditions that regulate transpiration and define water use strategies, and to better understand the link between plant economic spectrum, hydraulic safety and efficiency and the regulation of plant water status (Reich, 2014, Martínez-Vilalta & Garcia-Forner (2017)). This would be facilitated by building an additional database for SAPFLUXNET sites with available measured midday and pre-down leaf water potentials, which combined with midday sap flow data would allow to estimate whole-tree hydraulic conductance (Eller *et al.*, 2018). Most current models estimate whole-tree hydraulic conductance from branch-level xylem hydraulic conductance, either directly or applying a simple upscaling exercise based on conduit tapering (e.g., Christoffersen *et al.*, 2016; Eller *et al.*, 2020), which is likely to be inaccurate and biased across species. Besides, to complement the study of transpiration responses, high resolution dendrometric measures would allow to estimate the role of steam capacitance in water use strategies (Steppe *et al.*, 2015). There is still much to be understood about tree water use strategies and the drivers of transpiration regulation. This is in part due to the lack of high quality data for an adequate parametrization of transpiration dynamics at relevant spatiotemporal scales. The use of the SAPFLUXNET global database represents a major step forward and a powerful tool to overcome this challenge to a significant extent.

6.6 Conclusions

- 2.1. Calibrations of sap flow methods shows large intra- and inter-method variability in performance, with a low proportion of this variability explained by species and no consistent effects of wood density or porosity type.
- 2.2. Dissipation methods showed lower accuracy and higher proportional bias than other methods, but relatively high linearity and precision, while Pulse methods also showed significant proportional bias driven by their overestimation of low flows. Therefore, Dissipation methods may be more appropriate to assess relative sap flow and Pulse methods may be more suitable to quantify medium and high absolute flows.
- 2.3. All sap flow methods showed high precision, justifying their use to quantify environmental responses of transpiration and allowing potential correction of the measurements using study-specific calibrations.
- 3.1. We present the global SAPFLUXNET database, including harmonised and quality-controlled individual datasets supplied by contributors worldwide. Datasets include sub-daily time series of sap flow and hydrometeorological drivers for 202 globally distributed sites, including 2714 trees belonging to 174 species.
- 3.2. The SAPFLUXNET database provides the first global perspective of water use by individual plants at multiple timescales, with important applications in fields ranging from plant ecophysiology to Earth-system science.
- 4.1. Transpiration regulation is better explained by vapour pressure deficit (VPD) than by soil water content (SWC) or radiation (PPFD). Trees in dryland biomes are less coupled to all three hydrometeorological drivers than those in other biomes.
- 4.2. Climate, soil and vegetation structure were common controls of all three hydrometeorological couplings with canopy conductance, with wetter climates, fine textured soils and tall vegetation being associated to tighter coupling.
- 5.1. Reference canopy conductance and its sensitivity to VPD is coordinated with hydraulic and allocation traits (i.e. Ψ_{P50} , K_s , H_v , Ψ_{TLP} , R_{depth} , L_s and tree height) rather than being directly controlled by climate (e.g. mean annual precipitation).
- 5.2. Species with efficient xylem transport (higher hydraulic conductance) had higher canopy conductance but also higher sensitivity to VPD. Moreover, we found that angiosperms had higher reference canopy conductance and higher sensitivity to VPD than gymnosperms.

A

Appendix Chapter 2

A.1 Figures A

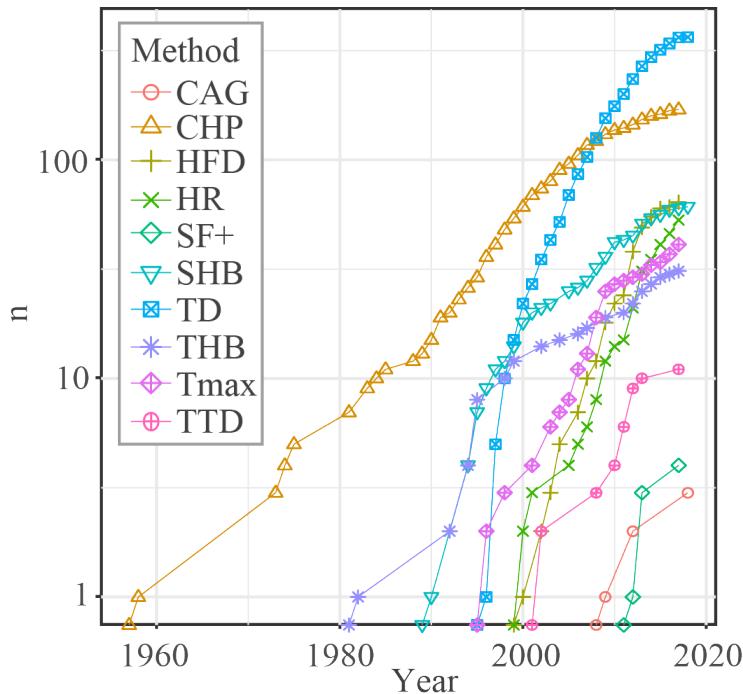


Figure A.1: Representation of the cumulative number of studies using different sap flow methods between 1957 and 2017 (adapted and updated from Potatos *et al.* (2016)). CAG: calibrated average gradient; CHP: compensation heat pulse (early heat pulse methods have been considered CHP, (Edwards *et al.*, 1997)); HFD: head field deformation; HR: heat ratio, SF+: sapflow+; SHB: stem heat balance; TD: thermal dissipation; THB: trunk heat balance; Tmax: T-max heat pulse; TTD: transient thermal dissipation. Notice the logarithmic scale on the y-axis.

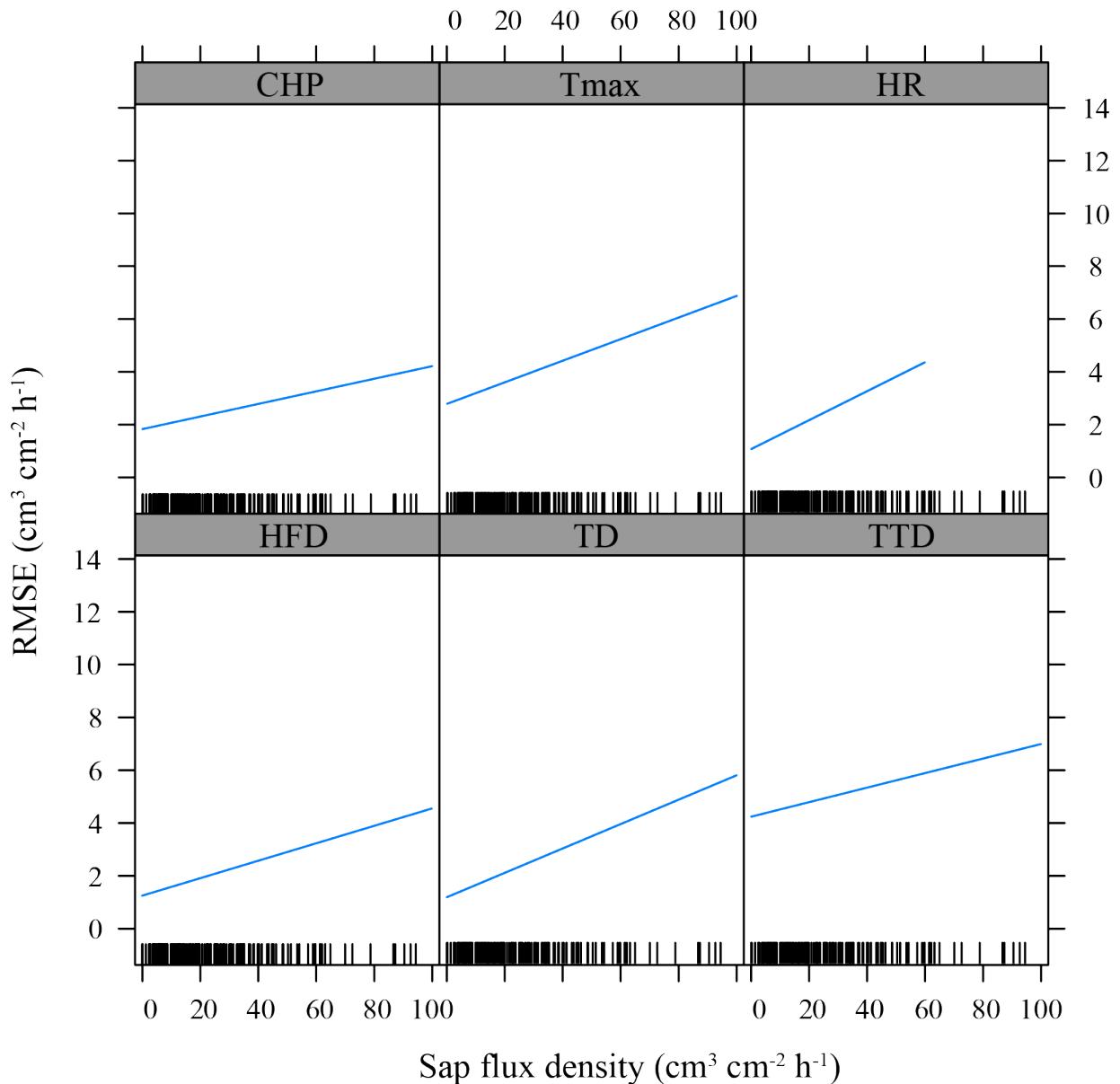


Figure A.2: Relationship between root mean square error (RMSE) and sap flux density (mean calibration range) and for different sap flux density methods, as predicted by the LMM model presented in Table 3.

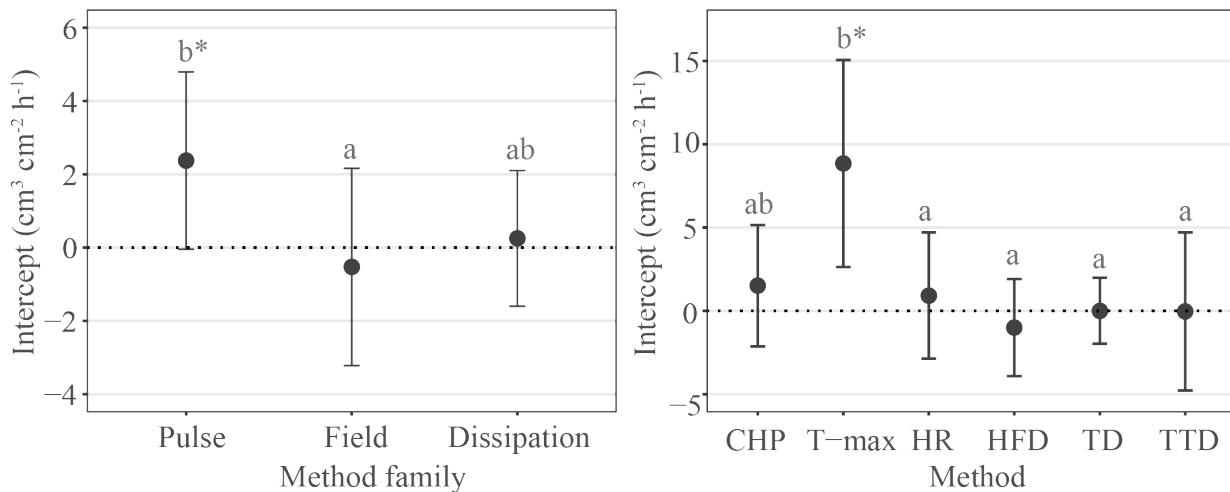


Figure A.3: Predictions of the LMM models calculated from least-squares means of the intercept (β_0) of the linear model (Eq. 3). Different letters indicate significant differences between factors levels evaluated with Tukey's test. Horizontal, dotted lines indicate reference, perfect calibration values for a given metric. Asterisks (*) indicate significant ($p < 0.05$) departure from those reference values.

A.2 Tables A

Table A.1: Summary table of the studies used in the analyses presented in the paper. Sap flow method, species, calibration material, porosity and average stem/tree diameter are reported.

| Study | Method | Species | Calibration material | Wood porosity | Diameter (cm) |
|--------------------------------------|--------|------------------------------|----------------------|----------------|---------------|
| Alarcon <i>et al.</i> 2005 | CHP | <i>Citrus limon</i> | whole plant | Diffuse porous | 2.50 |
| Ballester <i>et al</i> 2011 | CHP | <i>Citrus clementina</i> | whole plant | Diffuse porous | |
| Barret <i>et al.</i> 1995 | CHP | <i>Corymbia maculata</i> | without roots | Diffuse porous | |
| Bleby <i>et al.</i> 2004 | CHP | <i>Eucalyptus marginata</i> | whole plant | Diffuse porous | 10.00 |
| Bleby <i>et al.</i> 2004 | HR | <i>Eucalyptus marginata</i> | whole plant | Diffuse porous | 10.00 |
| Braun and Schmid 1999 | TD | <i>Vitis vinifera</i> | whole plant | Ring porous | 3.75 |
| Burgess <i>et al.</i> 2001 | HR | <i>Eucalyptus marginata</i> | whole plant | Diffuse porous | |
| Bush <i>et al.</i> 2010 | TD | <i>Populus fremontii</i> | stem segment | Diffuse porous | 5.08 |
| Bush <i>et al.</i> 2010 | TD | <i>Tilia cordata</i> | stem segment | Diffuse porous | 4.83 |
| Cain 2009 | TD | <i>Macaranga hypoleuca</i> | stem segment | Diffuse porous | 82.00 |
| Cain 2009 | TD | <i>Macaranga pearsonii</i> | stem segment | Diffuse porous | 67.00 |
| Caspari <i>et al.</i> 1993 | CHP | <i>Pyrus serotina</i> | whole plant | Diffuse porous | 6.62 |
| Caterina <i>et al.</i> 2013 | TD | <i>Juniperus virginiana</i> | stem segment | Tracheids | 8.00 |
| Chan 2015 | TD | <i>Abies concolor</i> | stem segment | Tracheids | 6.00 |
| Cohen <i>et al.</i> 1981 | T-max | <i>Platanus orientalis</i> | stem segment | Diffuse porous | 6.70 |
| Cohen <i>et al.</i> 1981 | T-max | <i>Populus alba</i> | stem segment | Diffuse porous | 6.70 |
| Cohen <i>et al.</i> 1998 | T-max | <i>Malus domestica</i> | whole plant | Diffuse porous | |
| Dragoni <i>et al.</i> 2005 | CHP | <i>Malus domestica</i> | whole plant | Diffuse porous | 6.50 |
| Dye <i>et al.</i> 1996 | CHP | <i>Pinus patula</i> | without roots | Tracheids | |
| Fernandez <i>et al.</i> 1999 | CHP | <i>Olea europaea</i> | stem segment | Diffuse porous | 8.80 |
| Fernandez <i>et al.</i> 1999 | CHP | <i>Olea europaea</i> | without roots | Diffuse porous | |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Citrus sinensis</i> | stem segment | Diffuse porous | 7.80 |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Citrus sinensis</i> | without roots | Diffuse porous | 10.40 |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Olea europaea</i> | stem segment | Diffuse porous | 8.20 |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Olea europaea</i> | without roots | Diffuse porous | 9.80 |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Prunus domestica</i> | stem segment | Diffuse porous | 8.00 |
| Fernandez <i>et al.</i> 2006 | CHP | <i>Prunus domestica</i> | without roots | Diffuse porous | 7.00 |
| Fuchs <i>et al.</i> 2017 | HFD | <i>Acer pseudoplatanus</i> | stem segment | Diffuse porous | 10.56 |
| Fuchs <i>et al.</i> 2017 | HFD | <i>Fagus sylvatica</i> | stem segment | Diffuse porous | 9.42 |
| Fuchs <i>et al.</i> 2017 | HFD | <i>Tilia cordata</i> | stem segment | Diffuse porous | 9.29 |
| Fuchs <i>et al.</i> 2017 | HR | <i>Acer pseudoplatanus</i> | stem segment | Diffuse porous | |
| Fuchs <i>et al.</i> 2017 | HR | <i>Fagus sylvatica</i> | stem segment | Diffuse porous | |
| Fuchs <i>et al.</i> 2017 | HR | <i>Tilia cordata</i> | stem segment | Diffuse porous | |
| Fuchs <i>et al.</i> 2017 | TD | <i>Acer campestre</i> | stem segment | Diffuse porous | 11.81 |
| Fuchs <i>et al.</i> 2017 | TD | <i>Acer pseudoplatanus</i> | stem segment | Diffuse porous | 10.80 |
| Fuchs <i>et al.</i> 2017 | TD | <i>Fagus sylvatica</i> | stem segment | Diffuse porous | 9.83 |
| Fuchs <i>et al.</i> 2017 | TD | <i>Populus nigra</i> | stem segment | Diffuse porous | 10.77 |
| Fuchs <i>et al.</i> 2017 | TD | <i>Tilia cordata</i> | stem segment | Diffuse porous | 9.41 |
| Gonzalez-Altozano <i>et al.</i> 1998 | CHP | <i>Citrus reticulata</i> | whole plant | Diffuse porous | 11.50 |
| Gonzalez-Altozano <i>et al.</i> 1998 | T-max | <i>Citrus reticulata</i> | whole plant | Diffuse porous | 11.50 |
| Granier 1985 | TD | <i>Pinus nigra</i> | stem segment | Tracheids | 4.50 |
| Granier 1985 | TD | <i>Pseudotzuga menziesii</i> | stem segment | Tracheids | 4.50 |
| Granier 1985 | TD | <i>Quercus pedunculata</i> | stem segment | Ring porous | 4.50 |
| Green <i>et al.</i> 1988 | CHP | <i>Actinidia chinensis</i> | stem segment | Diffuse porous | 5.25 |
| Green <i>et al.</i> 1988 | CHP | <i>Actinidia chinensis</i> | whole plant | Diffuse porous | 5.40 |
| Green <i>et al.</i> 1988 | CHP | <i>Malus sylvestris</i> | whole plant | Diffuse porous | 5.60 |

Table A.1: Summary table of the studies used in the analyses presented in the paper. Sap flow method, species, calibration material, porosity and average stem/tree diameter are reported. (*continued*)

| Study | Method | Species | Calibration material | Wood porosity | Diameter (cm) |
|-----------------------------------|--------|-----------------------------------|----------------------|----------------|---------------|
| Gutierrez <i>et al.</i> 1994 | SHB | <i>Acacia koa</i> | whole plant | Diffuse porous | |
| Gutierrez <i>et al.</i> 1994 | SHB | <i>Coffea arabica</i> | whole plant | Diffuse porous | |
| Gutierrez Soto <i>et al.</i> 2012 | HR | <i>Carica papaya</i> | whole plant | Monocot | |
| Hatton <i>et al.</i> 1995 | CHP | <i>Eucalyptus populnea</i> | without roots | Diffuse porous | 5.40 |
| Heilman <i>et al.</i> 1990 | SHB | <i>Ligustrum japonicum</i> | whole plant | Diffuse porous | 1.00 |
| Herbs <i>et al.</i> 2007 | TD | <i>Acer campestre</i> | stem segment | Diffuse porous | |
| Herbs <i>et al.</i> 2007 | TD | <i>Crataegus monogyna</i> | stem segment | Diffuse porous | |
| Hultine <i>et al.</i> 2010 | TD | <i>Tamarix ramosissima</i> | stem segment | Ring porous | 4.16 |
| Intrigliolo <i>et al.</i> 2009 | T-max | <i>Vitis vinifera</i> | whole plant | Ring porous | |
| Isarangkool <i>et al.</i> 2009 | TTD | <i>Abies concolor</i> | stem segment | Diffuse porous | 5.14 |
| Isarangkool <i>et al.</i> 2009 | TTD | <i>Hevea brasiliensis</i> | stem segment | Diffuse porous | 4.69 |
| Isarangkool <i>et al.</i> 2009 | TTD | <i>Mangifera indica</i> | stem segment | Diffuse porous | 4.35 |
| Johan Uddling <i>et al.</i> 2009 | TD | <i>Betula papyrifera</i> | stem segment | Diffuse porous | |
| Lu 2002 | TD | <i>Garcinia mangostana</i> | whole plant | Diffuse porous | 4.00 |
| Lu 2002 | TD | <i>Mangifera indica</i> | whole plant | Diffuse porous | 2.30 |
| Lu 2002 | TD | <i>Musa spp.</i> | whole plant | Monocot | 12.00 |
| Lu and Chacko 1998 | TD | <i>Mangifera indica</i> | whole plant | Diffuse porous | 2.30 |
| Madurapperuma <i>et al.</i> 2009 | HR | <i>Syagrus romanzoffiana</i> | whole plant | Monocot | |
| Michell <i>et al.</i> 2009 | HR | <i>Eucalyptus capillosa</i> | without roots | Diffuse porous | 6.50 |
| Montague <i>et al.</i> 2006 | TD | <i>Liquidambar styraciflua</i> | whole plant | Diffuse porous | 5.30 |
| Montague <i>et al.</i> 2006 | TD | <i>Populus deltoides</i> | whole plant | Diffuse porous | 5.60 |
| Montague <i>et al.</i> 2006 | TD | <i>Pyrus calleryana</i> | whole plant | Diffuse porous | 6.60 |
| Montague <i>et al.</i> 2006 | TD | <i>Quercus robur x Q. Bicolor</i> | whole plant | Ring porous | 5.70 |
| Nadezhina <i>et al.</i> 1998 | HFD | <i>Tilia cordata</i> | without roots | Diffuse porous | 12.00 |
| Nortes <i>et al.</i> 2009 | CHP | <i>Prunus dulcis</i> | whole plant | Diffuse porous | 15.00 |
| Paudel <i>et al.</i> 2013 | TD | <i>Malus domestica</i> | stem segment | Diffuse porous | 4.01 |
| Paudel <i>et al.</i> 2013 | TD | <i>Peltophorum dubium</i> | stem segment | Diffuse porous | 3.70 |
| Paudel <i>et al.</i> 2013 | TD | <i>Prunus persica</i> | stem segment | Diffuse porous | 4.00 |
| Paudel <i>et al.</i> 2013 | TTD | <i>Malus domestica</i> | stem segment | Diffuse porous | 4.01 |
| Paudel <i>et al.</i> 2013 | TTD | <i>Peltophorum dubium</i> | stem segment | Diffuse porous | 3.70 |
| Paudel <i>et al.</i> 2013 | TTD | <i>Prunus persica</i> | stem segment | Diffuse porous | 4.00 |
| Peters <i>et al.</i> 2017 | TD | <i>Larix decidua</i> | stem segment | Tracheids | 16.50 |
| Peters <i>et al.</i> 2017 | TD | <i>Picea abies</i> | stem segment | Tracheids | 15.90 |
| Prendergast <i>et al.</i> 2007 | T-max | <i>Actinidia chinensis</i> | stem segment | Diffuse porous | 9.50 |
| Shackel <i>et al.</i> 1992 | SHB | <i>Prunus persica</i> | whole plant | Diffuse porous | 6.25 |
| Smith <i>et al.</i> 1995 | CHP | <i>Acacia holosericea</i> | stem segment | Diffuse porous | |
| Smith <i>et al.</i> 1995 | CHP | <i>Acacia holosericea</i> | without roots | Diffuse porous | |
| Smith <i>et al.</i> 1995 | CHP | <i>Acacia nilotica</i> | stem segment | Diffuse porous | |
| Smith <i>et al.</i> 1995 | CHP | <i>Azadirachta indica</i> | stem segment | Diffuse porous | |
| Smith <i>et al.</i> 1995 | CHP | <i>Azadirachta indica</i> | without roots | Diffuse porous | |
| Sperling <i>et al.</i> 2012 | TD | <i>Phoenix dactylifera</i> | whole plant | Monocot | 60.00 |
| Steppe <i>et al.</i> 2010 | CHP | <i>Fagus grandifolia</i> | stem segment | Diffuse porous | 18.00 |
| Steppe <i>et al.</i> 2010 | HFD | <i>Fagus grandifolia</i> | stem segment | Diffuse porous | 18.12 |
| Steppe <i>et al.</i> 2010 | TD | <i>Fagus grandifolia</i> | stem segment | Diffuse porous | 18.00 |
| Sun <i>et al.</i> 2012 | TD | <i>Liquidambar styraciflua</i> | without roots | Diffuse porous | 7.50 |
| Sun <i>et al.</i> 2012 | TD | <i>Pinus echinata</i> | without roots | Tracheids | 7.50 |
| Sun <i>et al.</i> 2012 | TD | <i>Pinus taeda</i> | without roots | Tracheids | 7.50 |
| Sun <i>et al.</i> 2012 | TD | <i>Populus deltoides</i> | without roots | Diffuse porous | 7.50 |
| Sun <i>et al.</i> 2012 | TD | <i>Quercus alba</i> | without roots | Ring porous | 7.50 |
| Sun <i>et al.</i> 2012 | TD | <i>Ulmus americana</i> | without roots | Ring porous | 7.50 |

Table A.1: Summary table of the studies used in the analyses presented in the paper. Sap flow method, species, calibration material, porosity and average stem/tree diameter are reported. (*continued*)

| Study | Method | Species | Calibration material | Wood porosity | Diameter (cm) |
|----------------------------|--------|----------------------------|----------------------|----------------|---------------|
| Swanson and Whitfield 1981 | CHP | <i>Nothofagus solandri</i> | whole plant | Diffuse porous | 11.00 |
| Swanson and Whitfield 1981 | CHP | <i>Pinus radiata</i> | whole plant | Tracheids | 5.00 |
| Urban <i>et al.</i> 2012 | SHB | <i>Humulus lupulus</i> | without roots | Ring porous | |
| Vellame <i>et al.</i> 2010 | SHB | <i>Citrus sinensis</i> | whole plant | Diffuse porous | 1.40 |

Table A.2: Anova summary of the LMM models, using the same structure as objective 1, comparing calibrations reported in SFD and SF units (Units) for CHP and TD. CM (Calibration material).

| Method | Calibration metric | Variable | Sum sq | Mean Sq | NumDF | DenDF | F.value | Pr(>F) |
|--------|--------------------|----------|--------|---------|-------|-------|---------|-----------|
| CHP | Ln-Ratio | Units | 0.09 | 0.09 | 1 | 52.05 | 1.09 | 0.302 |
| | | CM | 0.20 | 0.10 | 2 | 44.43 | 1.24 | 0.299 |
| | Slope | Units | 0.07 | 0.07 | 1 | 25.27 | 0.37 | 0.55 |
| | | CM | 0.24 | 0.12 | 2 | 40.07 | 0.59 | 0.558 |
| | Slope (ln-ln) | Units | 0.00 | 0.00 | 1 | 12.35 | 0.02 | 0.89 |
| | | CM | 0.14 | 0.07 | 2 | 22.13 | 0.91 | 0.416 |
| TD | Z-Cor | Units | 1.07 | 1.07 | 1 | 20.77 | 4.30 | 0.051 . |
| | | CM | 4.63 | 2.32 | 2 | 34.98 | 9.28 | 0.001 *** |
| | Ln-Ratio | Units | 0.05 | 0.05 | 1 | 26.72 | 0.41 | 0.529 |
| | | CM | 0.14 | 0.07 | 2 | 16.64 | 0.58 | 0.571 |
| | Slope | Units | 0.16 | 0.16 | 1 | 44.72 | 2.28 | 0.138 |
| | | CM | 0.17 | 0.08 | 2 | 14.84 | 1.17 | 0.338 |
| TD | Slope (ln-ln) | Units | 0.22 | 0.22 | 1 | 72.67 | 2.18 | 0.144 |
| | | CM | 0.14 | 0.07 | 2 | 29.27 | 0.71 | 0.501 |
| | Z-Cor | Units | 0.08 | 0.08 | 1 | 26.07 | 0.28 | 0.598 |
| | | CM | 0.02 | 0.01 | 2 | 17.41 | 0.03 | 0.969 |

Table A.3: Summary of the LMM models of Ln-Ratio (accuracy), Slope (proportional bias), Slope (ln-ln) (linearity) and Z-Cor (precision) as a function of Methods and Calibration material (CM; Whole plant: whole plant on a container or lysimeter; No-roots: whole plant without roots). CHP is the reference level for the variable Method and Stem segment is the reference level for CM, corresponding to the model intercept. All other coefficient estimates indicate the difference relative to the intercept. σ^2 is the within-groups random variability (residuals of the model). τ_{00} is the between-group random variability. N is the number of levels within random groups. ICC is the Intraclass Correlation Coefficients of each random group. R^2m and R^2c are the variability explained by the fixed and the random factors, respectively.

| Coefficients | Estimate | Conf. Int. | p-value |
|-----------------------|----------|-----------------|---------|----------|-----------------|---------|----------|-----------------|---------|----------|-----------------|---------|
| Fixed effects | | | | | | | | | | | | |
| (Intercept) | 0.19 | -0.052 , 0.425 | 0.129 | 0.96 | 0.779 , 1.135 | <0.001 | 0.85 | 0.719 , 0.976 | <0.001 | 2.32 | 1.982 , 2.653 | 2.32 |
| Method (T-max) | -0.19 | -0.611 , 0.240 | 0.395 | -0.27 | -0.584 , 0.038 | 0.089 | -0.08 | -0.312 , 0.142 | 0.463 | -0.08 | -0.669 , 0.506 | -0.08 |
| Method (HR) | -0.28 | -0.510 , -0.047 | 0.019 | -0.04 | -0.247 , 0.162 | 0.683 | 0.06 | -0.106 , 0.221 | 0.488 | 0.16 | -0.205 , 0.525 | 0.16 |
| Method (HFD) | -0.21 | -0.409 , -0.003 | 0.048 | 0.01 | -0.167 , 0.193 | 0.886 | 0.00 | -0.143 , 0.143 | 1 | 0.54 | 0.220 , 0.862 | 0.54 |
| Method (SHB) | -0.37 | -0.843 , 0.095 | 0.124 | -0.04 | -0.365 , 0.285 | 0.809 | 0.18 | -0.058 , 0.427 | 0.139 | 0.45 | -0.170 , 1.070 | 0.45 |
| Method (TD) | -0.65 | -0.847 , -0.457 | <0.001 | -0.20 | -0.368 , -0.041 | 0.015 | 0.28 | 0.160 , 0.406 | <0.001 | -0.12 | -0.423 , 0.173 | -0.12 |
| Method (TTD) | -0.62 | -0.941 , -0.310 | <0.001 | -0.22 | -0.503 , 0.066 | 0.134 | 0.20 | -0.016 , 0.421 | 0.072 | -0.37 | -0.877 , 0.133 | -0.37 |
| CM (Whole plant) | -0.03 | -0.300 , 0.245 | 0.841 | -0.13 | -0.317 , 0.064 | 0.197 | -0.14 | -0.276 , -0.003 | 0.05 | -0.67 | -1.034 , -0.301 | -0.67 |
| CM (No-roots) | -0.13 | -0.394 , 0.128 | 0.319 | -0.08 | -0.298 , 0.135 | 0.463 | -0.06 | -0.213 , 0.104 | 0.503 | -0.78 | -1.172 , -0.379 | -0.78 |
| Random effects | | | | | | | | | | | | |
| σ^2 | | 0.091 | | | 0.1 | | | 0.076 | | | 0.278 | |
| τ_{00} Species | | 0.013 | | | 0.004 | | | 0.013 | | | 0.04 | |
| τ_{00} Study | | 0.159 | | | 0.041 | | | 0.004 | | | 0.184 | |
| $N_{Species}$ | | 65 | | | 65 | | | 65 | | | 65 | |
| N_{Study} | | 48 | | | 48 | | | 48 | | | 48 | |
| $ICC_{Species}$ | | 0.049 | | | 0.029 | | | 0.137 | | | 0.08 | |
| ICC_{Study} | | 0.604 | | | 0.284 | | | 0.048 | | | 0.366 | |
| Observations | | 290 | | | 290 | | | 290 | | | 290 | |

B

Appendix Chapter 3

B.1 Figures B

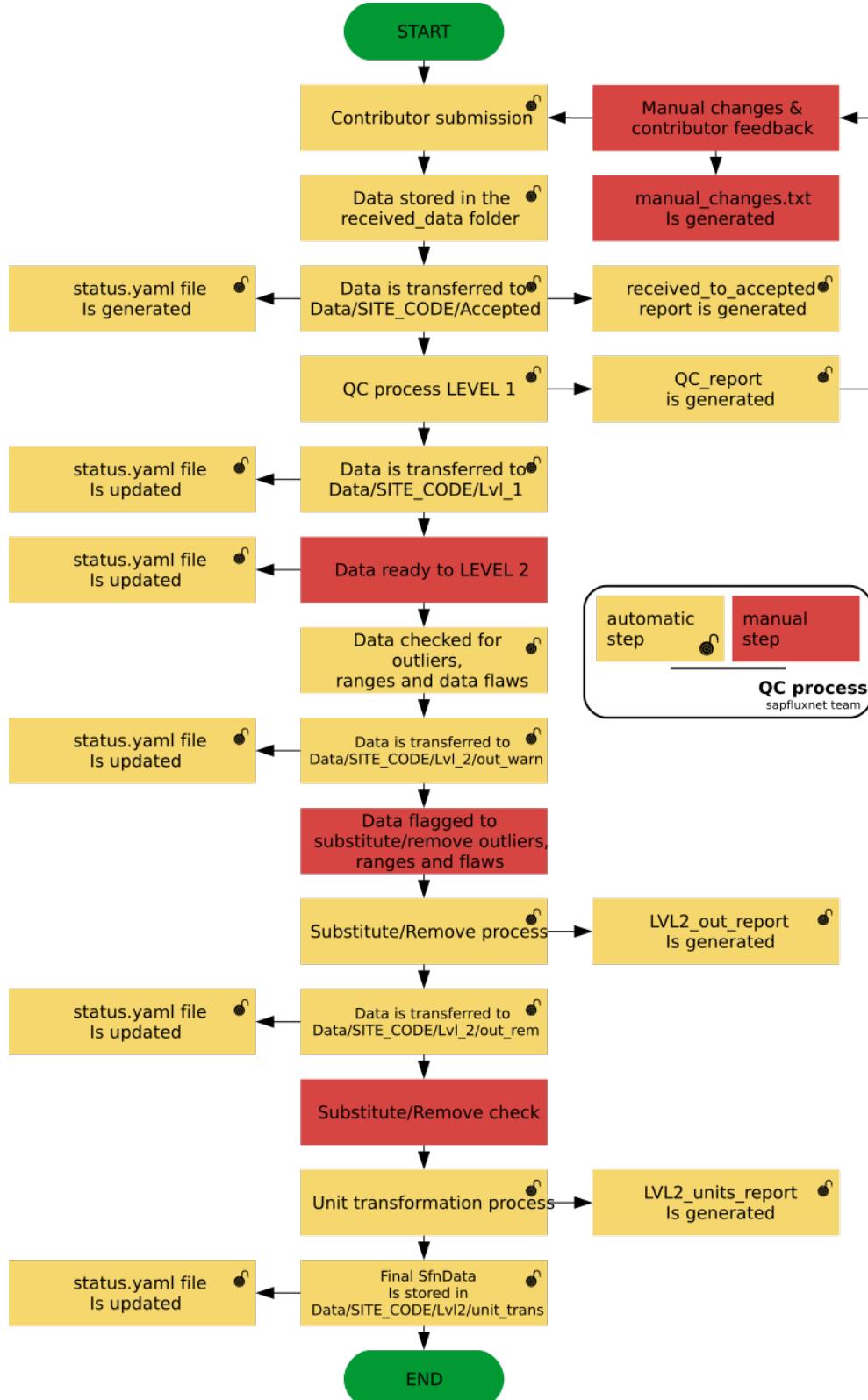


Figure B.1: Overview of the data QC process, showing file management and identifying automatic (in yellow) and manual steps (in red). The column on the left shows the different updates of the status file for each dataset and the column on the right shows generated data reports and steps requiring feedback or manual changes.

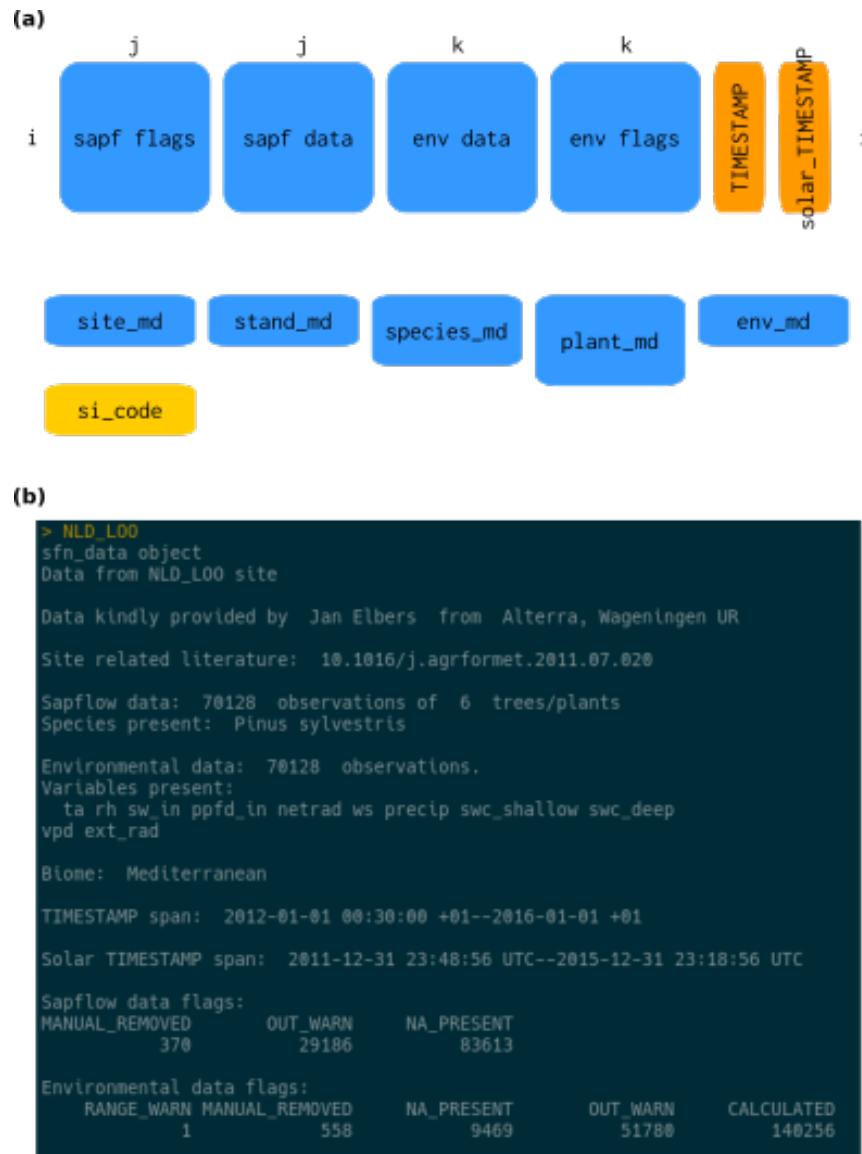


Figure B.2: (a) Structure of sfn_data objects, which are based on the S4 class. Boxes in the figure represent different slots where data are stored. Each object is identified by the 'si_code', stored as a slot in the object, with the format of a character vector. Slots storing time series of data and the associated data flags are of class 'tibble' and have all the same number of rows (i), corresponding to the the number of timesteps in the dataset and labelled with two POSIXct timestamp vectors TIMESTAMP, solar_TIMESTAMP).The slot storing sap flow data, 'sapf_data' contains(j) columns and environmental data ('env_data') contains k columns, corresponding to the number of environmental variables present. Slots with the suffix 'md' refer to the different metadata and all are objects of class 'tibble' with different dimensions. For example, the number of rows in plant_md' depends on the number of plants in the dataset (and this is depicted by the different length of the box). More information on the 'sfm_data-classes' of the package *sapfluxnetr* (Granda et al. 2020). (b) Summary of an sfn_data object, showing highlights of site metadata, data dimensions, timestamp span and flags present on the data.

Outliers, ranges and flaw values flagging

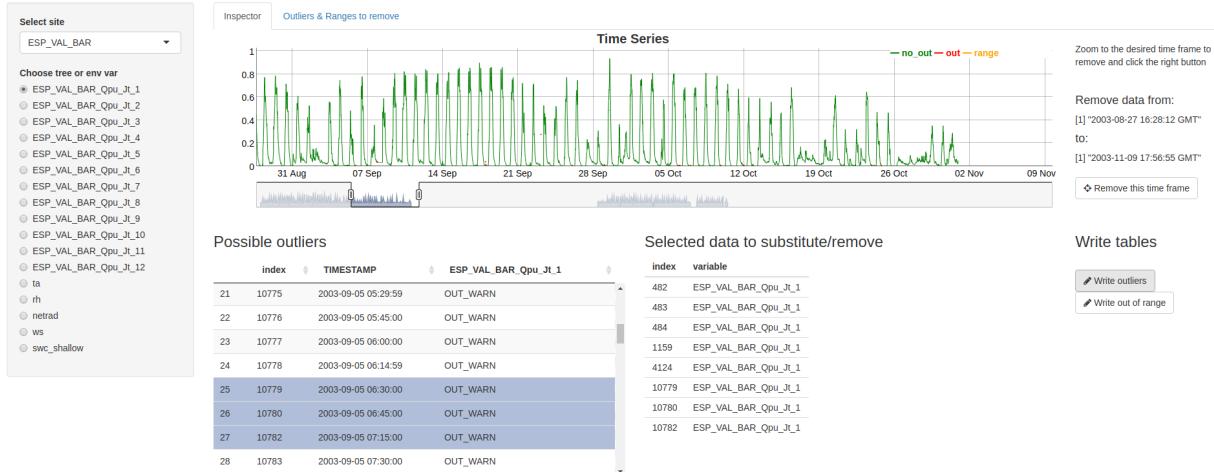


Figure B.3: Example screenshot of the app used for handling outliers and out of range values in time series. The left column shows dataset and variable selection. The central part shows the time series, with out of range values in red and possible outliers in yellow. Rows to replace or remove are selected in a table and written to a text file when done.

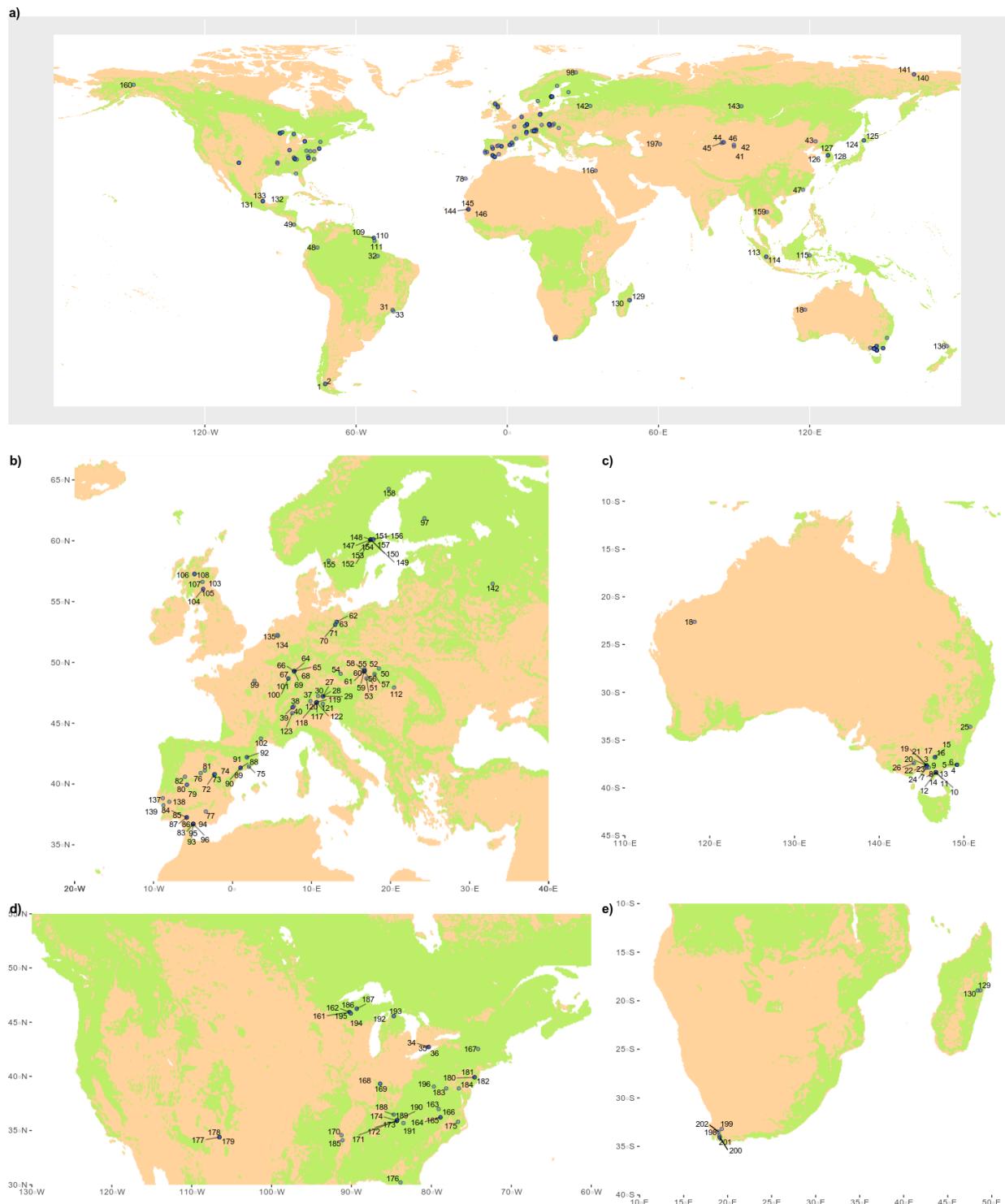


Figure B.4: Detailed geographic distribution of SAPFLUXNET datasets. Datasets are labelled by dataset number in Table S4. Woodland area from Crowther et al. (2015) shown in green.



B.2 Tables B

Table B.1: Data checks implemented in the first level of data quality control (QC1).

| Check | Description |
|----------------------------------|--|
| Metadata variables | All metadata variables are checked for presence and expected class (numeric, character, logical...). |
| Character variables values | All metadata character variables are checked against the possible values (factor levels) for that variable, raising a warning if some value is out of the expected. |
| E-mail check | E-mail provided by contributors is checked for validity |
| Coordinates and biome | Site coordinates are checked for correctness (are they inside the specified country?) and fixed if needed and possible. MAT and MAP values are obtained for that coordinates and the biome is calculated from that values. |
| Soil texture | Percentages of soil textures are used to calculate the USDA classification category if possible. |
| Species names | Species names in plant and species metadata are checked for spelling errors and the concordance between both metadata is also checked |
| Plant treatments | Check for uniformity in the treatment declared by plant. |
| Environmental variables presence | Check for concordance between the declared variables in the environmental metadata and the environmental data. |
| Timestamp | Format, NA presence (there is data, but there is no timestamp), concordance and continuity are checked. |
| Gap presence: | Data gaps (There is TIMESTAMP but there is no data) are summarised and visualized. |
| Soil water content | Check for percentage swc values and transform them to cm ³ /cm ³ |

Table B.2: Description of site metadata variables.

| Variable | Description | Type | Units |
|-------------------------|---|-----------|-----------------------------------|
| si_name | Site name given by contributors | Character | None |
| si_country | Country code (ISO) | Character | Fixed values |
| si_contact_firstname | Contributor first name | Character | None |
| si_contact_lastname | Contributor last name | Character | None |
| si_contact_email | Contributor email | Character | None |
| si_contact_institution | Contributor affiliation | Character | None |
| si_addcontr_firstname | Additional contributor first name | Character | None |
| si_addcontr_lastname | Additional contributor last name | Character | None |
| si_addcontr_email | Additional contributor email | Character | None |
| si_addcontr_institution | Additional contributor affiliation | Character | None |
| si_lat | Site latitude (i.e. 42.36) | Numeric | Latitude, decimal format (WGS84) |
| si_long | Site longitude (i.e. -8.23) | Numeric | Longitude, decimal format (WGS84) |
| si_elev | Elevation above sea level | Numeric | meters |
| si_paper | Paper with relevant information to understand the site as DOI links or DOI codes | Character | DOI link |
| si_dist_mgmt | Recent and historic disturbance and management events that affected the measurement years | Character | Fixed values |
| si_igbp | Vegetation type based on IGBP classification | Character | Fixed values |
| si_flux_network | Logical indicating if site is participating in the FLUXNET network | Logical | Fixed values |
| si_dendro_network | Logical indicating if site is participating in the DENDROGLOBAL network | Logical | Fixed values |
| si_remarks | Remarks and commentaries useful to grasp some site-specific peculiarities | Character | None |
| si_code | sapfluxnet site code, unique for each site | Character | Fixed value |
| si_mat | Site annual mean temperature, as obtained from WorldClim | Numeric | Celsius degrees |
| si_map | Site annual mean precipitation, as obtained from WorldClim | Numeric | mm |
| si_biome | Biome classification as per Whittaker diagram, based on mat and map obtained from WorldClim | Character | sapfluxnet calculated |

Table B.3: Description of stand metadata variables.

| Variable | Description | Type | Units |
|----------------------|---|-----------|---------------------------------|
| st_name | Stand name given by contributors | Character | None |
| st_growth_condition | Growth condition with respect to stand origin and management | Character | Fixed values |
| st_treatment | Treatment applied at stand level | Character | None |
| st_age | Mean stand age at the moment of sap flow measurements | Numeric | years |
| st_height | Canopy height | Numeric | meters |
| st_density | Total stem density for stand | Numeric | stems ha ⁻¹ |
| st_basal_area | Total stand basal area | Numeric | m ² ha ⁻¹ |
| st_lai | Total maximum stand leaf area (one-sided, projected) | Numeric | m ² m ⁻² |
| st_aspect | Aspect the stand is facing (exposure) | Character | Fixed values |
| st_terrain | Slope and/or relief of the stand | Character | Fixed values |
| st_soil_depth | Soil total depth | Numeric | cm |
| st_soil_texture | Soil texture class, based on simplified USDA classification | Character | Fixed values |
| st_sand_perc | Soil sand content, % mass | Numeric | % percentage |
| st_silt_perc | Soil silt content, % mass | Numeric | % percentage |
| st_clay_perc | Soil clay content, % mass | Numeric | % percentage |
| st_remarks | Remarks and commentaries useful to grasp some stand-specific peculiarities | Character | None |
| st_USDA_soil_texture | USDA soil classification based on the percentages provided by the contributor | Character | sapfluxnet calculated |

Table B.4: Description of species metadata variables.

| Variable | Description | Type | Units |
|--------------------|---|-----------|--|
| sp_name | Identity of each measured species | Character | Scientific name without author abbreviation, as accepted by The Plant List |
| sp_ntrees | Number of trees measured of each species | Numeric | number of trees |
| sp_leaf_habit | Leaf habit of the measured species | Character | Fixed values |
| sp_basal_area_perc | Basal area occupied by each measured species, in percentage over total stand basal area | Numeric | % percentage |

Table B.5: Description of plant metadata variables.

| Variable | Description | Type | Units |
|-------------------|--|-----------|--|
| pl_name | Plant code assigned by contributors | Character | None |
| pl_species | Species identity of the measured plant | Character | Scientific name without author abbreviation, as accepted by The Plant List |
| pl_treatment | Experimental treatment (if any) | Character | None |
| pl_dbh | Diameter at breast height of measured plants | Numeric | cm |
| pl_height | Height of measured plants | Numeric | m |
| pl_age | Plant age at the moment of measure | Numeric | years |
| pl_social | Plant social status | Character | Fixed values |
| pl_sapw_area | Cross-sectional sapwood area | Numeric | cm ² |
| pl_sapw_depth | Sapwood depth, measured at breast height | Numeric | cm |
| pl_bark_thick | Plant bark thickness | Numeric | mm |
| pl_leaf_area | Leaf area of each measured plant | Numeric | m ² |
| pl_sens_meth | Sap flow measures method | Character | Fixed values |
| pl_sens_man | Sap flow measures sensor manufacturer | Character | Fixed values |
| pl_sens_cor_grad | Correction for natural temperature gradients method | Character | Fixed values |
| pl_sens_cor_zero | Zero flow determination method | Character | Fixed values |
| pl_sens_calib | Was species-specific calibration used? | Logical | Fixed values |
| pl_sap_units | Uniformized sapfluxnet units for sapwood, leaf and plant level | Character | Fixed values |
| pl_sap_units_orig | Original contribution units (at sapwood or plant level) | Character | Fixed values |
| pl_sens_length | Length of the needles or electrodes forming the sensor | Numeric | mm |
| pl_sens_hgt | Sensor installation height, measured from the ground | Numeric | m |
| pl_sens_timestep | Subdaily time step of sensor measures | Numeric | minutes |
| pl_radial_int | | Character | Fixed values |
| pl_azimut_int | | Character | Fixed values |
| pl_remarks | Remarks and commentaries useful to grasp some plant-specific peculiarities | Character | None |
| pl_code | sapfluxnet plant code, unique for each plant | Character | Fixed value |

Table B.6: Description of environmental metadata variables.

| Variable | Description | Type | Units |
|-----------------------|---|-----------|--------------|
| env_time_zone | Time zone of site used in the TIMESTAMPS | Character | Fixed values |
| env_time_daylight | Is daylight saving time applied to the original timestamp? | Logical | Fixed values |
| env_timestep | Subdaily timestep of environmental measures | Numeric | minutes |
| env_ta | Location of air temperature sensor | Character | Fixed values |
| env_rh | Location of relative humidity sensor | Character | Fixed values |
| env_vpd | Location of relative vapour pressure deficit sensor | Character | Fixed values |
| env_sw_in | Location of shortwave incoming radiation sensor | Character | Fixed values |
| env_ppfd_in | Location of incoming photosynthetic photon flux density sensor | Character | Fixed values |
| env_netrad | Location of net radiation sensor | Character | Fixed values |
| env_ws | Location of wind speed sensor | Character | Fixed values |
| env_precip | Location of precipitation sensor | Character | Fixed values |
| env_swc_shallow_depth | Average depth for shallow soil water content measures | Numeric | cm |
| env_swc_deep_depth | Average depth for deep soil water content measures | Numeric | cm |
| env_plant_watpot | Availability of water potential values for the same measured plants during the sap flow measurements period | Character | Fixed values |
| env_leafarea_seasonal | Availability of seasonal course leaf area data and level | Character | Fixed values |
| env_remarks | Remarks and commentaries useful to grasp some environmental-specific peculiarities | Character | None |

Table B.7: Data checks implemented in the second level of data quality control (QC2).

| Data check | Description |
|--|--|
| Sap flow units harmonisation | Sap flow expressed in $\text{cm}^3 \text{ h}^{-1}$, sap flow per unit leafsapwood area in $\text{cm}^3 \text{ cm}^{-2} \text{ h}^{-1}$ |
| Out of range detection | Out of range values are flagged automatically, examined in a visual app and removed if confirmed |
| Outlier detection | Outliers are flagged automatically, examined in a visual app and removed if confirmed |
| Radiation transformations | Interconversion between global radiation (sw_in) and photosynthetically active radiation (ppfd_in) |
| VPD and relative humidity | Interconversion between VPD and relative humidity |
| Extraterrestrial radiation and solar timestamp | Calculation of extraterrestrial radiation and solar timestamp from timestamp and geographical data |
| Sap flow interconversions | When sapwood or leaf areas were available, interconversions were applied between the different expression levels for sap flow (per plant, per sapwood area or per leaf area) |

Table B.8: Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. Number of species per dataset, geographic coordinates and elevation are also shown. Negative coordinate values are shown for Southern and Western Hemispheres.

| si_code | si_name | si_lat | si_long | si_elev | # species |
|-----------------|-------------------------------------|--------|---------|---------|-----------|
| ARG_MAZ | Mazaruca_Patagonia | -51.58 | -72.29 | 550 | 1 |
| ARG_TRE | Tres Marias | -51.32 | -72.19 | 460 | 1 |
| AUS_BRI_BRI | Britannia Creek | -37.87 | 145.85 | 707 | 1 |
| AUS_CAN_ST1_EUC | Cann River | -37.58 | 149.17 | 180 | 1 |
| AUS_CAN_ST2_MIX | Cann River | -37.58 | 149.17 | 180 | 2 |
| AUS_CAN_ST3 ACA | Cann River | -37.58 | 149.17 | 180 | 1 |
| AUS_CAR THI_00F | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_0P0 | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_0PF | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_CON | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_T00 | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_T0F | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_TPO | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_CAR THI_TPF | Carrajung | -38.38 | 146.68 | 610 | 1 |
| AUS_ELL_HB_HIG | Ella | -36.78 | 146.58 | 705 | 2 |
| AUS_ELL_MB_MOD | Ella | -36.78 | 146.58 | 693 | 1 |
| AUS_ELL_UNB | Ella | -36.78 | 146.58 | 737 | 1 |
| AUS_KAR | Karijini NP | -22.62 | 118.22 | 710 | 1 |
| AUS_MAR_HSD_HIG | Maroondah | -37.64 | 145.58 | 468 | 2 |
| AUS_MAR_HSW_HIG | Maroondah | -37.65 | 145.57 | 297 | 2 |
| AUS_MAR_MSD_MOD | Maroondah | -37.64 | 145.58 | 467 | 2 |
| AUS_MAR_MSW_MOD | Maroondah | -37.65 | 145.57 | 261 | 2 |
| AUS_MAR_UBD | Maroondah | -37.69 | 145.56 | 303 | 3 |
| AUS_MAR_UBW | Maroondah | -37.89 | 145.57 | 336 | 3 |
| AUS_RIC_EUC_ELE | Richmond NSW EucFACE | -33.62 | 150.74 | 23 | 1 |
| AUS_WOM | WombatStateForest | -37.42 | 144.09 | 705 | 2 |
| AUT_PAT_FOR | Patscherkofel | 47.21 | 11.45 | 1950 | 1 |
| AUT_PAT_KRU | Patscherkofel | 47.21 | 11.45 | 2180 | 1 |
| AUT_PAT_TRE | Patscherkofel | 47.21 | 11.45 | 2110 | 1 |
| AUT_TSC | Tschirgant south | 47.23 | 10.84 | 750 | 1 |
| BRA_CAM | Campos do Jordão | -22.69 | -45.52 | 2000 | 1 |
| BRA_CAX_CON | Caxiuana | -1.79 | -51.43 | 15 | 8 |
| BRA_SAN | Santa Virgínia (PESM) | -23.28 | -45.18 | 1000 | 4 |
| CAN_TUR_P39_POS | TUR | 42.71 | -80.36 | 184 | 1 |
| CAN_TUR_P39_PRE | TUR | 42.71 | -80.36 | 184 | 1 |
| CAN_TUR_P74 | TUR | 42.71 | -80.35 | 184 | 1 |
| CHE_DAV_SEE | Davos | 46.82 | 9.86 | 1650 | 1 |
| CHE_LOT_NOR | Lotschental | 46.39 | 7.76 | 1300 | 2 |
| CHE_PFY_CON | Pfynwald | 46.30 | 7.60 | 615 | 1 |
| CHE_PFYIRR | Pfynwald | 46.30 | 7.60 | 615 | 1 |
| CHN_ARG_GWD | Arghan | 40.75 | 89.99 | 830 | 1 |
| CHN_ARG_GWS | Arghan | 41.38 | 89.94 | 830 | 1 |
| CHN_HOR_AFF | Horqin | 42.72 | 122.37 | 226 | 1 |
| CHN_YIN_ST1 | Yingbazar | 42.45 | 85.72 | 900 | 1 |
| CHN_YIN_ST2_DRO | Yingbazar | 42.11 | 85.13 | 930 | 1 |
| CHN_YIN_ST3_DRO | Yingbazar | 42.29 | 85.99 | 930 | 1 |
| CHN_YUN_YUN | Yunxiao | 23.92 | 117.42 | 0 | 2 |
| COL_MAC_SAF_RAD | Macagual Universidad de la Amazonia | 1.50 | -75.36 | 360 | 1 |
| CRI_TAM_TOW | TAMU Soltis Center | 10.39 | -84.63 | 600 | 17 |
| CZE_BIK | Bik | 49.49 | 18.53 | 875 | 1 |

Table B.8: Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. Number of species per dataset, geographic coordinates and elevation are also shown. Negative coordinate values are shown for Southern and Western Hemispheres. (*continued*)

| si_code | si_name | si_lat | si_long | si_elev | # species |
|-----------------|---|--------|---------|---------|-----------|
| CZE_BIL_BIL | Bilovice | 49.25 | 16.69 | 320 | 1 |
| CZE_KRT_KRT | Krtiny | 49.32 | 16.75 | 480 | 1 |
| CZE_LAN | Lanžhot | 48.68 | 16.95 | 150 | 3 |
| CZE_LIZ LES | Liz | 49.07 | 13.68 | 858 | 1 |
| CZE_RAJ_RAJ | Rajec | 49.44 | 16.70 | 600 | 1 |
| CZE_SOB_SOB | Šobesice | 49.25 | 16.69 | 320 | 1 |
| CZE_STI | Stitna nad Vlari | 49.04 | 17.97 | 550 | 1 |
| CZE_UTE_BEE | Utechov | 49.28 | 16.65 | 420 | 1 |
| CZE_UTE_BNA | Utechov | 49.28 | 16.65 | 390 | 1 |
| CZE_UTE_BPO | Utechov | 49.28 | 16.65 | 370 | 1 |
| CZE_UTE_SPR | Utechov | 49.28 | 16.65 | 360 | 1 |
| DEU_HIN_OAK | Hinnensee | 53.33 | 13.19 | 90 | 1 |
| DEU_HIN_TER | Hinnensee | 53.33 | 13.19 | 95 | 2 |
| DEU_MER_BEE_NON | Merzalben | 49.27 | 7.81 | 550 | 1 |
| DEU_MER_BEE THI | Merzalben | 49.27 | 7.81 | 550 | 1 |
| DEU_MER_DOU_NON | Merzalben | 49.27 | 7.81 | 550 | 1 |
| DEU_MER_DOU THI | Merzalben | 49.27 | 7.81 | 550 | 1 |
| DEU_MER_MIX_NON | Merzalben | 49.27 | 7.81 | 550 | 2 |
| DEU_MER_MIX THI | Merzalben | 49.27 | 7.81 | 550 | 2 |
| DEU_STE_2P3 | Stechlin | 53.10 | 13.00 | 78 | 1 |
| DEU_STE_4P5 | Stechlin | 53.10 | 13.00 | 78 | 1 |
| ESP_ALT_ARM | Alto Tajo | 40.78 | -2.33 | 1079 | 3 |
| ESP_ALT_HUE | Alto Tajo | 40.79 | -2.29 | 907 | 2 |
| ESP_ALT_TRI | Alto Tajo | 40.80 | -2.23 | 981 | 2 |
| ESP_CAN | Can Balasc | 41.43 | 2.07 | 270 | 4 |
| ESP_GUA_VAL | Guadarrama | 40.90 | -4.03 | 1140 | 1 |
| ESP_LAHC COM | LaHarina | 37.74 | -3.38 | 180 | 1 |
| ESP_LAS | Las Canadas, Teide national park tenerife | 28.31 | -16.57 | 2070 | 1 |
| ESP_MAJ_MAI | Majadas del Tietar | 39.94 | -5.77 | 260 | 1 |
| ESP_MAJ_NOR_LM1 | Majadas del Tietar | 39.94 | -5.77 | 260 | 1 |
| ESP_MON_SIE_NAT | Montejo | 41.12 | -3.50 | 1400 | 3 |
| ESP_RIN | Rinconada experimental catchment | 40.60 | -6.02 | 1200 | 1 |
| ESP_RON_PIL | Ronda | 36.69 | -5.02 | 1734 | 2 |
| ESP_SAN_A2_45I | Sanabria orchard | 37.25 | -5.80 | 49 | 1 |
| ESP_SAN_A_45I | Sanabria orchard | 37.25 | -5.80 | 49 | 1 |
| ESP_SAN_B_100 | Sanabria orchard | 37.25 | -5.80 | 49 | 1 |
| ESP_SAN_B2_100 | Sanabria orchard | 37.25 | -5.80 | 49 | 1 |
| ESP_TIL_MIX | Tillar | 41.33 | 1.01 | 1018 | 2 |
| ESP_TIL_OAK | Tillar | 41.33 | 1.01 | 1011 | 1 |
| ESP_TIL_PIN | Tillar | 41.33 | 1.01 | 1065 | 1 |
| ESP_VAL_BAR | Vallcebre | 42.20 | 1.82 | 1102 | 1 |
| ESP_VAL_SOR | Vallcebre | 42.20 | 1.81 | 1257 | 1 |
| ESP_YUN_C1 | Yunquera | 36.72 | -4.97 | 1220 | 1 |
| ESP_YUN_C2 | Yunquera | 36.72 | -4.97 | 1180 | 1 |
| ESP_YUN_T1 THI | Yunquera | 36.72 | -4.97 | 1190 | 1 |
| ESP_YUN_T3 THI | Yunquera | 36.72 | -4.97 | 1185 | 1 |
| FIN_HYY_SME | Hyttiala Forest Field Station | 61.85 | 24.29 | 185 | 2 |
| FIN_PET | Petsikko | 69.49 | 27.23 | 251 | 1 |
| FRA_FON | Fontainebleau-Barbeau | 48.48 | 2.78 | 105 | 2 |

Table B.8: Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. Number of species per dataset, geographic coordinates and elevation are also shown. Negative coordinate values are shown for Southern and Western Hemispheres. (*continued*)

| si_code | si_name | si_lat | si_long | si_elev | # species |
|-----------------|------------------------|--------|---------|---------|-----------|
| FRA_HES_HE1_NON | Hesse | 48.67 | 7.06 | 300 | 1 |
| FRA_HES_HE2_NON | Hesse | 48.67 | 7.06 | 300 | 1 |
| FRA_PUE | Puechabon | 43.74 | 3.60 | 270 | 1 |
| GBR_ABE_PLO | Aberfeldy | 56.62 | -3.80 | 340 | 1 |
| GBR_DEV_CON | Devilla | 56.03 | -3.72 | 75 | 1 |
| GBR_DEV_DRO | Devilla | 56.03 | -3.72 | 75 | 1 |
| GBR_GUI_ST1 | Guisachan | 57.27 | -4.82 | 300 | 1 |
| GBR_GUI_ST2 | Guisachan | 57.27 | -4.82 | 300 | 1 |
| GBR_GUI_ST3 | Guisachan | 57.27 | -4.82 | 300 | 1 |
| GUF_GUY_GUY | Guyaflux | 5.28 | -52.92 | 40 | 6 |
| GUF_GUY_ST2 | Guyaflux | 5.28 | -52.91 | 45 | 7 |
| GUF_NOU_PET | Nouragues station | 4.08 | -52.68 | 120 | 10 |
| HUN_SIK | Sikfokut | 47.93 | 20.44 | 330 | 2 |
| IDN_JAM_OIL | Jambi | -2.07 | 102.79 | 71 | 1 |
| IDN_JAM_RUB | Jambi | -2.10 | 102.78 | 90 | 1 |
| IDN_PON_STE | Pono | -1.49 | 120.06 | 1050 | 8 |
| ISR_YAT_YAT | Yatir | 31.34 | 35.05 | 650 | 1 |
| ITA_FEI_S17 | Feichtwald-Matsch | 46.69 | 10.61 | 1715 | 1 |
| ITA_KAE_S20 | Kaelbergangl-Matsch | 46.70 | 10.61 | 1990 | 1 |
| ITA_MAT_S21 | Matscher Alm-Matsch | 46.74 | 10.69 | 2100 | 2 |
| ITA_MUN | Muntatschinig-Matsch | 46.68 | 10.58 | 1160 | 1 |
| ITA_REN | Renon | 46.59 | 11.43 | 1794 | 3 |
| ITA_RUN_N20 | Runer Koepfl-Matsch | 46.70 | 10.64 | 2030 | 2 |
| ITA_TOR | Torgnon | 45.82 | 7.56 | 2100 | 1 |
| JPN_EBE_HYB | Ebetsu | 43.08 | 141.52 | 40 | 1 |
| JPN_EBE_SUG | Ebetsu | 43.08 | 141.52 | 40 | 1 |
| KOR_TAE_TC1_LOW | Taehwa | 37.30 | 127.32 | 160 | 1 |
| KOR_TAE_TC2_MED | Taehwa | 37.30 | 127.32 | 160 | 1 |
| KOR_TAE_TC3_EXT | Taehwa | 37.30 | 127.32 | 160 | 1 |
| MDG_SEM_TAL | Semi-mature forest | -18.93 | 48.71 | 950 | 6 |
| MDG_YOU_SHO | Young secondary forest | -18.95 | 48.40 | 990 | 1 |
| MEX_COR_YP | Cortadura | 19.49 | -97.04 | 2180 | 1 |
| MEX_VER_BSJ | VERACRUZ_BSJ | 19.51 | -96.98 | 1440 | 5 |
| MEX_VER_BSM | VERACRUZ_BSM | 19.53 | -96.99 | 1524 | 2 |
| NLD_LOO | Loobos | 52.17 | 5.74 | 25 | 1 |
| NLD_SPE_DOU | Speulderbos | 52.25 | 5.69 | 50 | 1 |
| NZL_HUA_HUA | Huapai | -36.80 | 174.49 | 90 | 1 |
| PRT_LEZ_ARN | LEZIRIAS | 38.83 | -8.82 | 15 | 1 |
| PRTMIT | MITRA II | 38.54 | -8.00 | 235 | 1 |
| PRT_PIN | Pinheiro da Cruz | 38.25 | -8.76 | 5 | 2 |
| RUS_CHE_LOW | Cherskii | 68.74 | 161.50 | 90 | 1 |
| RUS_CHE_Y4 | CHE | 68.74 | 161.41 | 6 | 1 |
| RUS_FYO | Fyodorovskoye | 56.46 | 32.92 | 260 | 3 |
| RUS_POG_VAR | Pogorelsky Bor | 56.36 | 92.95 | 243 | 3 |
| SEN_SOUIRR | Souilène | 16.34 | -15.43 | 10 | 1 |
| SEN_SOUPOS | Souilène | 16.34 | -15.43 | 10 | 1 |
| SEN_SOUPRE | Souilène | 16.34 | -15.43 | 10 | 1 |
| SWE_NOR_ST1_AF1 | Norunda | 60.09 | 17.48 | 45 | 2 |
| SWE_NOR_ST1_AF2 | Norunda | 60.09 | 17.48 | 45 | 2 |
| SWE_NOR_ST1_BEF | Norunda | 60.09 | 17.48 | 45 | 2 |
| SWE_NOR_ST2 | Norunda | 60.09 | 17.48 | 45 | 2 |
| SWE_NOR_ST3 | Norunda | 60.09 | 17.48 | 45 | 2 |

Table B.8: Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. Number of species per dataset, geographic coordinates and elevation are also shown. Negative coordinate values are shown for Southern and Western Hemispheres. (*continued*)

| si_code | si_name | si_lat | si_long | si_elev | # species |
|-----------------|---|--------|---------|---------|-----------|
| SWE_NOR_ST4_AFT | Norunda | 60.08 | 17.48 | 45 | 3 |
| SWE_NOR_ST4_BEF | Norunda | 60.08 | 17.48 | 45 | 2 |
| SWE_NOR_ST5_REF | Norunda | 60.08 | 17.48 | 45 | 3 |
| SWE_SKO_MIN | Skogaryd | 58.36 | 12.15 | 76 | 1 |
| SWE_SKY_38Y | Skyttorp | 60.13 | 17.84 | 50 | 1 |
| SWE_SKY_68Y | Skyttorp | 60.10 | 17.83 | 50 | 2 |
| SWE_SVA_MIX_NON | Svartberget | 64.26 | 19.77 | 267 | 2 |
| THA_KHU | Khu-Muang | 15.27 | 103.08 | 150 | 1 |
| USA_BNZ_BLA | BNZSPRC1 | 64.70 | -148.32 | 50 | 1 |
| USA_CHE_ASP | ChEAS | 45.94 | -90.27 | 477 | 6 |
| USA_CHE_MAP | ChEAS | 45.95 | -90.26 | 1565 | 2 |
| USA_DUK_HAR | Duke Blackwood | 36.98 | -79.09 | 163 | 6 |
| | Hardwood | | | | |
| USA_HIL_HF1_POS | Hill Demonstration Forest | 36.22 | -78.86 | 174 | 5 |
| USA_HIL_HF1_PRE | Hill Demonstration Forest | 36.22 | -78.86 | 174 | 5 |
| USA_HIL_HF2 | Hill Demonstration Forest | 36.22 | -78.86 | 174 | 7 |
| USA_HUY_LIN_NON | Huyck Preserve Lincoln | 42.53 | -74.16 | | 1 |
| | Pond | | | | |
| USA_INM | INMMSF | 39.32 | -86.41 | 286 | 6 |
| USA_MOR_SF | Morgan-Monroe State | 39.32 | -86.41 | 275 | 4 |
| | Forest | | | | |
| USA_NWH | NWhiteRiver | 34.58 | -91.26 | 48 | 2 |
| USA_ORN_ST1_AMB | ORNL-FACE | 35.90 | -84.33 | 227 | 1 |
| USA_ORN_ST2_AMB | ORNL-FACE | 35.90 | -84.33 | 227 | 1 |
| USA_ORN_ST3_ELE | ORNL-FACE | 35.90 | -84.33 | 227 | 1 |
| USA_ORN_ST4_ELE | ORNL-FACE | 35.90 | -84.33 | 227 | 1 |
| USA_PAR_FER | Parker Tract | 35.80 | -76.67 | 5 | 1 |
| USA_PER_PER | Perry | 30.21 | -83.87 | 14 | 1 |
| USA_PJS_P04_AMB | PJSEV -Rainfall | 34.39 | -106.53 | 1911 | 2 |
| | Manipulation Experiment - | | | | |
| | Sevilleta NWR, USA | | | | |
| USA_PJS_P08_AMB | PJSEV -Rainfall | 34.39 | -106.53 | 1911 | 2 |
| | Manipulation Experiment - | | | | |
| | Sevilleta NWR, USA | | | | |
| USA_PJS_P12_AMB | PJSEV -Rainfall | 34.39 | -106.53 | 1911 | 2 |
| | Manipulation Experiment - | | | | |
| | Sevilleta NWR, USA | | | | |
| USA_SIL_OAK_1PR | Silas Little Experimental Forest premortality | 39.92 | -74.60 | 33 | 4 |
| USA_SIL_OAK_2PR | Silas Little Experimental Forest premortality | 39.92 | -74.60 | 33 | 4 |
| USA_SIL_OAK_POS | Silas Little Experimental Forest premortality | 39.92 | -74.60 | 33 | 5 |
| USA_SMI_SCB | Smithsonian Conservation Biology Institute | 38.89 | -78.15 | 273 | 3 |
| USA_SMI_SER | Smithsonian Environmental Research Center | 38.89 | -76.56 | 19 | 5 |
| USA_SWH | SWhiteRiver | 34.11 | -91.13 | 44 | 2 |
| USA_SYL_HL1 | Sylvania Wilderness | 46.24 | -89.35 | 500 | 3 |
| USA_SYL_HL2 | Sylvania Wilderness | 46.24 | -89.35 | 500 | 4 |
| USA_TNB | TNBSF | 36.47 | -84.70 | 454 | 4 |

Table B.8: Datasets in the SAPFLUXNET database identified by numeric code, dataset code and site name. Number of species per dataset, geographic coordinates and elevation are also shown. Negative coordinate values are shown for Southern and Western Hemispheres. (*continued*)

| si_code | si_name | si_lat | si_long | si_elev | # species |
|----------------|-------------------------|--------|---------|---------|-----------|
| USA_TNO | TNOAK | 35.97 | -84.28 | 340 | 5 |
| USA_TNP | TNPINE | 35.96 | -84.29 | 342 | 5 |
| USA_TNY | TNYPOP | 35.69 | -83.50 | 850 | 3 |
| USA_UMB_CON | UMBS | 45.56 | -84.71 | 236 | 5 |
| USA_UMB_GIR | UMB | 45.56 | -84.70 | 239 | 4 |
| USA_WIL_WC1 | Willow Creek | 45.81 | -90.09 | 520 | 5 |
| USA_WIL_WC2 | Willow Creek | 45.81 | -90.09 | 520 | 4 |
| USA_WVF | WVFEF | 39.06 | -79.69 | 844 | 5 |
| UZB_YAN_DIS | Yangibazar | 41.65 | 60.62 | 101 | 2 |
| ZAF_FRA_FRA | Franshoek South Africa | -33.88 | 19.06 | 190 | 1 |
| ZAF_NOO_E3_IRR | Nooitgedacht farm | -33.20 | 19.34 | 1089 | 1 |
| ZAF_RAD | Radyn EGVV | -34.08 | 19.11 | 409 | 1 |
| ZAF_SOU_SOU | Southfield EGVV | -34.09 | 19.09 | 389 | 1 |
| ZAF_WEL_SOR | Wellington Western Cape | -33.48 | 18.96 | 81 | 1 |

Table B.9: Number of plants and number of datasets for each species present in the SAPFLUXNET database.

| Species | # trees | # sites | Species | # trees | # sites | Species | # trees | # sites |
|--------------------------------|---------|---------|---|---------|---------|---|---------|---------|
| <i>Pinus sylvestris</i> | 290 | 28 | <i>Acacia tortilis</i> | 9 | 3 | <i>Prunus serotina</i> | 3 | 2 |
| <i>Picea abies</i> | 178 | 19 | <i>Quercus spp.</i> | 9 | 2 | <i>Populus canescens</i> | 3 | 1 |
| <i>Acer saccharum</i> | 162 | 9 | <i>Kandelia obovata</i> | 8 | 1 | <i>Eucalyptus camaldulensis</i> | 3 | 1 |
| <i>Fagus sylvatica</i> | 116 | 16 | <i>Carpinus betulus</i> | 8 | 2 | <i>Qualea rosea</i> | 3 | 1 |
| <i>Pinus taeda</i> | 107 | 6 | <i>Castanopsis</i> <i>acuminatissima</i> | 8 | 1 | <i>Licania alba</i> | 3 | 1 |
| <i>Populus tremuloides</i> | 104 | 1 | <i>Pinus patula</i> | 8 | 1 | <i>Eucalyptus dives</i> | 2 | 1 |
| <i>Pinus koraiensis</i> | 96 | 3 | <i>Eucalyptus radiata</i> | 7 | 5 | <i>Licania octandra</i> | 2 | 1 |
| <i>Eucalyptus nitens</i> | 89 | 8 | <i>Betula pubescens subsp.</i> <i>czerapanovii</i> | 7 | 1 | <i>Swartzia racemosa</i> | 2 | 1 |
| <i>Pinus strobus</i> | 75 | 5 | <i>Avicennia marina</i> | 6 | 1 | <i>Manilkara bidentata</i> | 2 | 1 |
| <i>Liquidambar styraciflua</i> | 69 | 10 | <i>Quercus robur</i> | 6 | 1 | <i>Licania membranacea</i> | 2 | 2 |
| <i>Quercus ilex</i> | 62 | 6 | <i>Fraxinus excelsior</i> | 6 | 1 | <i>Eschweilera grandiflora</i> | 2 | 1 |
| <i>Acer rubrum</i> | 62 | 12 | <i>Cryptocarya laevigata</i> | 6 | 1 | <i>Pouteria viridis</i> | 2 | 1 |
| <i>Liriodendron tulipifera</i> | 51 | 11 | <i>Myrtaceae fam.</i> | 6 | 1 | <i>Ampelocera macrocarpa</i> | 2 | 1 |
| <i>Fagus grandifolia</i> | 48 | 4 | <i>Palaquium luzoniense</i> | 6 | 1 | <i>Otoba novogranatensis</i> | 2 | 1 |
| <i>Pinus resinosa</i> | 43 | 1 | <i>Platea excelsa</i> | 6 | 1 | <i>Mortoniodendron</i> <i>anisophyllum</i> | 2 | 1 |
| <i>Eucalyptus globulus</i> | 35 | 2 | <i>Pouteria firma</i> | 6 | 1 | <i>Meliosma idiopoda</i> | 2 | 1 |
| <i>Larix decidua</i> | 34 | 8 | <i>Agathis australis</i> | 6 | 1 | <i>Taxus baccata</i> | 2 | 1 |
| <i>Abies pinsapo</i> | 34 | 5 | <i>Ostrya virginiana</i> | 6 | 3 | <i>Sloanea sp</i> | 2 | 2 |
| <i>Acacia mearnsii</i> | 33 | 2 | <i>Picea mariana</i> | 6 | 1 | <i>Betula sp.</i> | 2 | 1 |
| <i>Quercus pyrenaica</i> | 32 | 2 | <i>Nothofagus pumilio</i> | 5 | 1 | <i>Picea glauca</i> | 2 | 1 |
| <i>Quercus rubra</i> | 32 | 6 | <i>Nothofagus</i> <i>cunninghamii</i> | 5 | 1 | <i>Fraxinus americana</i> | 2 | 1 |
| <i>Quercus petraea</i> | 31 | 5 | <i>Eucalyptus cypellocarpa</i> | 5 | 4 | <i>Carya cordiformis</i> | 2 | 1 |
| <i>Pseudotsuga menziesii</i> | 29 | 5 | <i>Eucalyptus rubida</i> | 5 | 1 | <i>Quercus prinus</i> | 2 | 1 |
| <i>Pinus halepensis</i> | 27 | 2 | <i>Drimys brasiliensis</i> | 5 | 1 | <i>Elaeagnus angustifolia</i> | 2 | 1 |
| <i>Quercus velutina</i> | 24 | 4 | <i>Alchornea triplinervia</i> | 5 | 1 | <i>Qualea tricolor</i> | 2 | 1 |
| <i>Tsuga canadensis</i> | 24 | 2 | <i>Santiria apiculata</i> | 5 | 1 | <i>Lecythis poiteauii</i> | 2 | 1 |
| <i>Larix cajanderi</i> | 23 | 2 | <i>Quercus michauxii</i> | 5 | 1 | <i>Quercus cerris</i> | 2 | 1 |

Table B.9: Number of plants and number of datasets for each species present in the SAPFLUXNET database. (continued)

| Species | # trees | # sites | Species | # trees | # sites | Species | # trees | # sites |
|--------------------------------|---------|---------|--------------------------------|---------|---------|---|---------|---------|
| <i>Betula papyrifera</i> | 21 | 2 | <i>Quercus phellos</i> | 5 | 1 | <i>Pleuranthodendron lindenii</i> | 1 | 1 |
| <i>Quercus montana</i> | 21 | 3 | <i>Pinus rigida</i> | 5 | 3 | <i>Inga sp.</i> | 1 | 1 |
| <i>Quercus pubescens</i> | 19 | 2 | <i>Tilia americana</i> | 5 | 2 | <i>Cupania macrophylla</i> | 1 | 1 |
| <i>Abies balsamea</i> | 19 | 1 | <i>Nothofagus antarctica</i> | 4 | 1 | <i>Genipa americana</i> | 1 | 1 |
| <i>Quercus alba</i> | 19 | 7 | <i>Eucalyptus baxteri</i> | 4 | 2 | <i>Brosimum alicastrum</i> | 1 | 1 |
| <i>Pinus cembra</i> | 18 | 6 | <i>Coprosma quadrifida</i> | 4 | 2 | <i>Pouteria sp.</i> | 1 | 1 |
| <i>Populus euphratica</i> | 16 | 6 | <i>Eschweilera coriacea</i> | 4 | 2 | <i>Macrolobium costaricense</i> | 1 | 1 |
| <i>Olea europaea</i> | 16 | 5 | <i>Arbutus unedo</i> | 4 | 1 | <i>Eschweilera sp.</i> | 1 | 1 |
| <i>Quercus rotundifolia</i> | 16 | 3 | <i>Psiadia altissima</i> | 4 | 1 | <i>Aspidosperma desmanthum</i> | 1 | 1 |
| <i>Hevea brasiliensis</i> | 16 | 2 | <i>Quercus suber</i> | 4 | 1 | <i>Trophis mexicana</i> | 1 | 1 |
| <i>Betula alleghaniensis</i> | 16 | 2 | <i>Betula pubescens</i> | 4 | 2 | <i>Betula pendula</i> | 1 | 1 |
| <i>Pinus nigra</i> | 15 | 3 | <i>Fraxinus pennsylvanica</i> | 4 | 2 | <i>Iryanthera sagotiana</i> | 1 | 1 |
| <i>Picea sitchensis</i> | 15 | 1 | <i>Pouteria anomala</i> | 3 | 1 | <i>Vantanea sp</i> | 1 | 1 |
| <i>Pinus edulis</i> | 15 | 3 | <i>Protium tenuifolium</i> | 3 | 1 | <i>Recordoxylon speciosum</i> | 1 | 1 |
| <i>Juniperus monosperma</i> | 15 | 3 | <i>Hieronyma alchorneoides</i> | 3 | 1 | <i>Larix kaempferi x Larix gmelinii</i> | 1 | 1 |
| <i>Eucalyptus victrix</i> | 14 | 1 | <i>Mollinedia schottiana</i> | 3 | 1 | <i>Cryptomeria japonica</i> | 1 | 1 |
| <i>Eucalyptus obliqua</i> | 14 | 5 | <i>Rustia formosa</i> | 3 | 1 | <i>Eugenia spp.</i> | 1 | 1 |
| <i>Quercus lyrata</i> | 13 | 2 | <i>Theobroma cacao</i> | 3 | 1 | <i>Ocotea samosa</i> | 1 | 1 |
| <i>Celtis laevigata</i> | 13 | 2 | <i>Carapa guianensis</i> | 3 | 1 | <i>Leptolaena sp.</i> | 1 | 1 |
| <i>Quercus coccinea</i> | 13 | 4 | <i>Gymnanthes riparia</i> | 3 | 1 | <i>Abrahama ditimena</i> | 1 | 1 |
| <i>Populus grandidentata</i> | 12 | 1 | <i>Ilex aquifolium</i> | 3 | 1 | <i>Brachylaena ramiflora</i> | 1 | 1 |
| <i>Malus domestica</i> | 11 | 3 | <i>Vouacapoua americana</i> | 3 | 3 | <i>Cryptocarya sp.</i> | 1 | 1 |
| <i>Eucalyptus tereticornis</i> | 10 | 1 | <i>Oxandra asbeckii</i> | 3 | 2 | <i>Sauraia pedunculata</i> | 1 | 1 |
| <i>Quercus faginea</i> | 10 | 2 | <i>Gouania glabra</i> | 3 | 3 | <i>Turpinia insignis</i> | 1 | 1 |
| <i>Pinus canariensis</i> | 10 | 1 | <i>Vernonia arborea</i> | 3 | 1 | <i>Sassafras albidum</i> | 1 | 1 |
| <i>Elaeis guineensis</i> | 10 | 1 | <i>Platanus mexicana</i> | 3 | 1 | <i>Ulmus americana</i> | 1 | 1 |
| <i>Acacia longifolia</i> | 10 | 1 | <i>Clethra macrophylla</i> | 3 | 2 | <i>Carya glabra</i> | 1 | 1 |
| <i>Pinus pinaster</i> | 10 | 1 | <i>Larix sibirica</i> | 3 | 1 | <i>Quercus falcata</i> | 1 | 1 |
| <i>Thuja occidentalis</i> | 10 | 1 | <i>Larix gmelinii</i> | 3 | 1 | <i>Cornus florida</i> | 1 | 1 |
| <i>Carya tomentosa</i> | 10 | 2 | <i>Pinus sibirica</i> | 3 | 1 | <i>Licania Rodriguesii</i> | 1 | 1 |

Table B.9: Number of plants and number of datasets for each species present in the SAPFLUXNET database. (*continued*)

| Species | # trees | # sites | Species | # trees | # sites | Species | # trees | # sites |
|-----------------------------|---------|---------|-------------------------|---------|---------|-----------------------|---------|---------|
| <i>Dicorynia guianensis</i> | 9 | 2 | <i>Pinus virginiana</i> | 3 | 1 | <i>Sextonia rubra</i> | 1 | 1 |

Table B.10: Number of plants per genus present in the SAPFLUXNET database.

| Genus | # trees | Genus | # trees | Genus | # trees |
|---------------------|---------|-----------------------|---------|--------------------------|---------|
| <i>Pinus</i> | 725 | <i>Cryptocarya</i> | 7 | <i>Ampelocera</i> | 2 |
| <i>Quercus</i> | 326 | <i>Avicennia</i> | 6 | <i>Otoba</i> | 2 |
| <i>Acer</i> | 224 | <i>Myrtaceae fam.</i> | 6 | <i>Mortoniodendron</i> | 2 |
| <i>Picea</i> | 201 | <i>Palaquium</i> | 6 | <i>Meliosma</i> | 2 |
| <i>Eucalyptus</i> | 188 | <i>Platea</i> | 6 | <i>Taxus</i> | 2 |
| <i>Fagus</i> | 164 | <i>Agathis</i> | 6 | <i>Sloanea</i> | 2 |
| <i>Populus</i> | 135 | <i>Ostrya</i> | 6 | <i>Elaeagnus</i> | 2 |
| <i>Liquidambar</i> | 69 | <i>Drimys</i> | 5 | <i>Lecythis</i> | 2 |
| <i>Larix</i> | 64 | <i>Alchornea</i> | 5 | <i>Pleuranthodendron</i> | 1 |
| <i>Abies</i> | 53 | <i>Santiria</i> | 5 | <i>Inga</i> | 1 |
| <i>Acacia</i> | 52 | <i>Tilia</i> | 5 | <i>Cupania</i> | 1 |
| <i>Betula</i> | 51 | <i>Qualea</i> | 5 | <i>Genipa</i> | 1 |
| <i>Liriodendron</i> | 51 | <i>Coprosma</i> | 4 | <i>Brosimum</i> | 1 |
| <i>Pseudotsuga</i> | 29 | <i>Arbutus</i> | 4 | <i>Macrolobium</i> | 1 |
| <i>Tsuga</i> | 24 | <i>Psiadia</i> | 4 | <i>Aspidosperma</i> | 1 |
| <i>Olea</i> | 16 | <i>Protium</i> | 3 | <i>Trophis</i> | 1 |
| <i>Hevea</i> | 16 | <i>Hieronyma</i> | 3 | <i>Iryanthera</i> | 1 |
| <i>Juniperus</i> | 15 | <i>Mollinedia</i> | 3 | <i>Vantanea</i> | 1 |
| <i>Nothofagus</i> | 14 | <i>Rustia</i> | 3 | <i>Recordoxylon</i> | 1 |
| <i>Carya</i> | 13 | <i>Theobroma</i> | 3 | <i>Cryptomeria</i> | 1 |
| <i>Celtis</i> | 13 | <i>Carapa</i> | 3 | <i>Eugenia</i> | 1 |
| <i>Pouteria</i> | 12 | <i>Gymnanthes</i> | 3 | <i>Ocotea</i> | 1 |
| <i>Fraxinus</i> | 12 | <i>Ilex</i> | 3 | <i>Leptolaena</i> | 1 |
| <i>Malus</i> | 11 | <i>Vouacapoua</i> | 3 | <i>Abrahamaia</i> | 1 |
| <i>Elaeis</i> | 10 | <i>Oxandra</i> | 3 | <i>Brachylaena</i> | 1 |
| <i>Thuja</i> | 10 | <i>Gouphia</i> | 3 | <i>Sauraia</i> | 1 |
| <i>Dicorynia</i> | 9 | <i>Vernonia</i> | 3 | <i>Turpinia</i> | 1 |
| <i>Licania</i> | 8 | <i>Platanus</i> | 3 | <i>Sassafras</i> | 1 |
| <i>Kandelia</i> | 8 | <i>Clethra</i> | 3 | <i>Ulmus</i> | 1 |
| <i>Carpinus</i> | 8 | <i>Prunus</i> | 3 | <i>Cornus</i> | 1 |
| <i>Castanopsis</i> | 8 | <i>Swartzia</i> | 2 | <i>Sextonia</i> | 1 |
| <i>Eschweilera</i> | 7 | <i>Manilkara</i> | 2 | | |

C

Appendix Chapter 4

C.1 Figures C

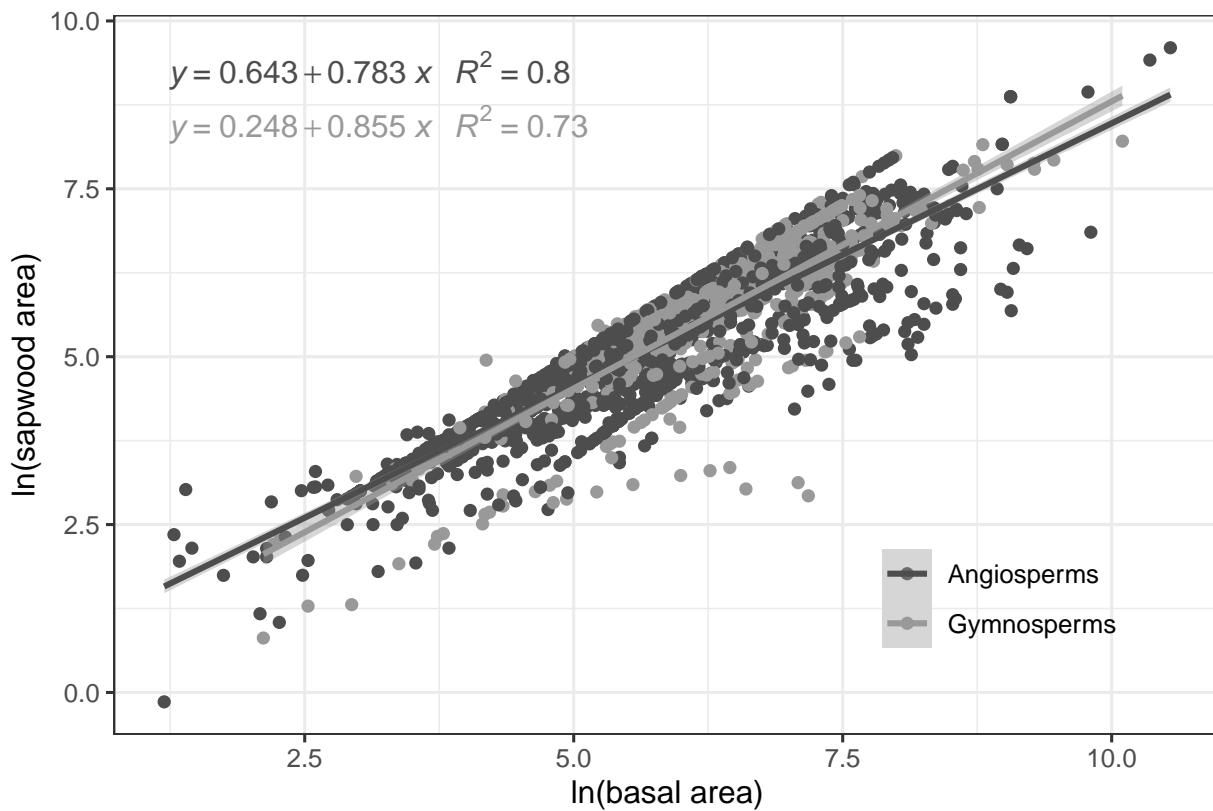


Figure C.1: SAPFLUXNET global scaling relationship between basal area and sapwood area. Shaded areas are 95% model confidence interval.

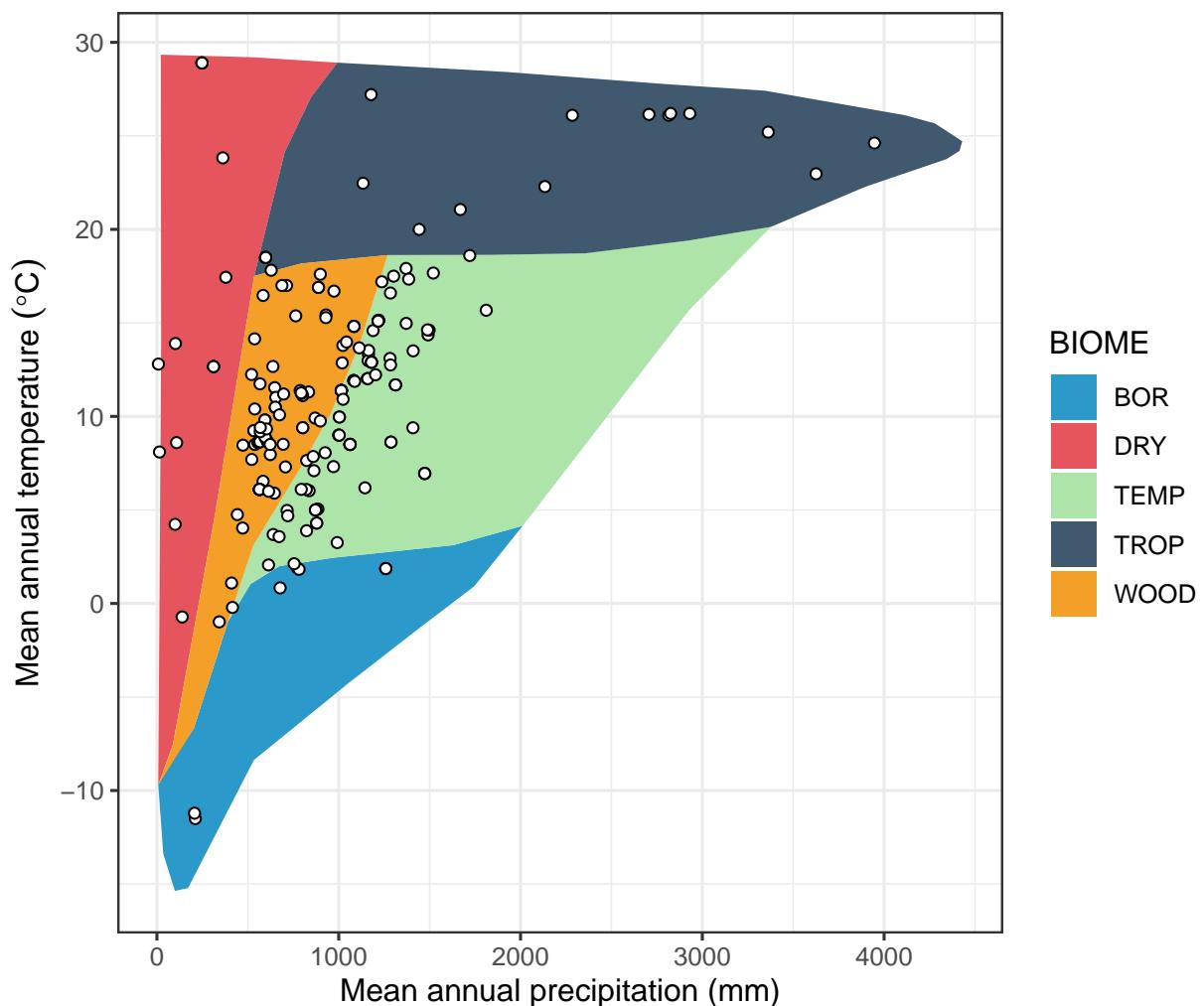


Figure C.2: Bioclimatic distribution of the SAPFLUXNET datasets used in the study. Points show the different datasets in a Whittaker diagram showing the classification of the aggregated biomes used in the study.

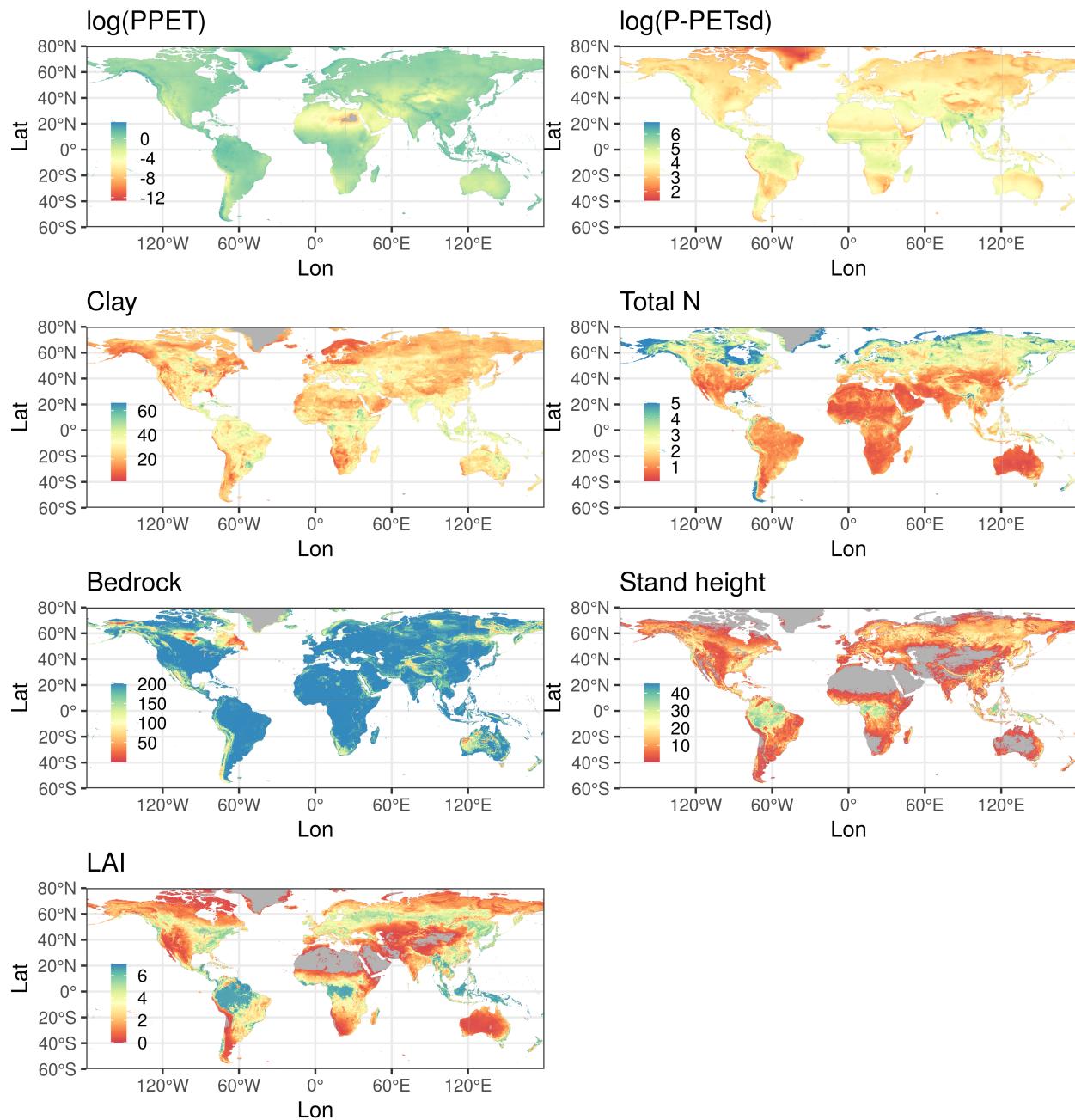


Figure C.3: Global projection of climatic, soil and stand structure variables. log(PPET): logarithm of precipitation over potential evapotranspiration [log(mm mm⁻¹)]; log(P-PET_{sd}): logarithm of the standard deviation of the difference between precipitation and potential evapotranspiration [log(mm)]; Clay: percentage of clay in the soil; Total N: total nitrogen in the soil [g kg⁻¹]; Bedrock [cm]; Stand height [m]; LAI: leaf area index [m² m⁻²]. Total N values above 5 g kg⁻¹ were truncated.

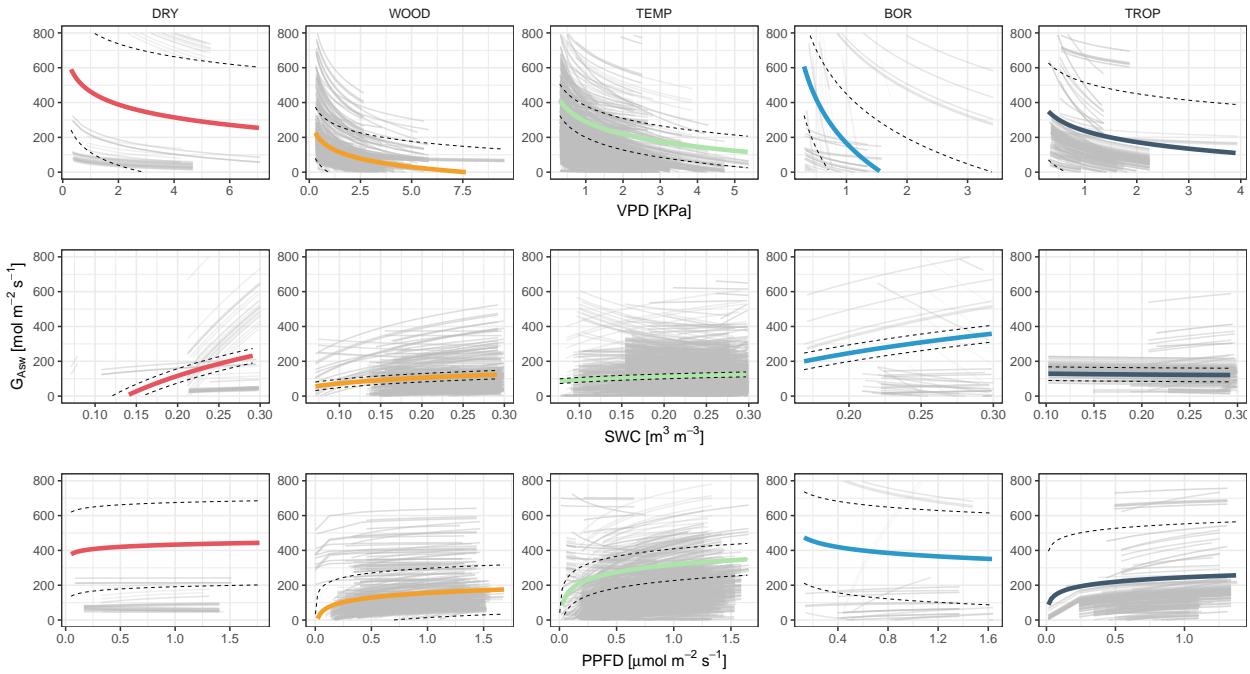


Figure C.4: Log relationships of the three environmental variables estimated with the TOTAL model (VPD + SWC + PPFD) and grouped by biome. Coloured lines are biome average models calculated from the models predictions using LMM with G_{Asw} as response variable and the neperian logarithm of the environmental constrains as explanatory variables. Dashed line shows standard error of the average models calculated with bootstrap prediction using 100 simulations.

C.2 Tables C

Table C.1: SAPFLUXNET sites included in the study. Biome was calculated using Whittaker diagram. *Indicates that the biome was manually adjusted and confirmed by SAPFLUXNET contributors.

| Site code | Latitude | Longitude | Biome | # Tree-days | # Species | # Trees |
|-----------------|----------|-----------|-------|-------------|-----------|---------|
| AUS_CAN_ST1_EUC | -37.58 | 149.17 | WOOD | 337 | 1 | 12 |
| AUS_CAN_ST2_MIX | -37.58 | 149.17 | WOOD | 712 | 2 | 22 |
| AUS_CAN_ST3 ACA | -37.58 | 149.17 | WOOD | 409 | 1 | 12 |
| AUS_CAR THI CON | -38.38 | 146.68 | TEMP | 54 | 1 | 7 |
| AUS_ELL_UNB | -36.78 | 146.58 | TEMP | 105 | 1 | 2 |
| AUS_MAR_UBD | -37.69 | 145.56 | TEMP | 50 | 3 | 5 |
| AUS_MAR_UBW | -37.89 | 145.57 | TEMP | 105 | 3 | 5 |
| AUS_WOM | -37.42 | 144.09 | TEMP | 2130 | 2 | 11 |
| AUT_PAT_FOR | 47.21 | 11.45 | BOR | 149 | 1 | 3 |
| AUT_PAT_KRU | 47.21 | 11.45 | BOR | 70 | 1 | 3 |
| AUT_PAT_TRE | 47.21 | 11.45 | BOR | 81 | 1 | 3 |
| BRA_CAM | -22.69 | -45.52 | TROP* | 79 | 1 | 5 |
| BRA_CAX_CON | -1.79 | -51.43 | TROP | 525 | 8 | 15 |
| CAN_TUR_P39_PRE | 42.71 | -80.36 | TEMP | 1021 | 1 | 18 |
| CAN_TUR_P74 | 42.71 | -80.35 | TEMP | 1997 | 1 | 16 |
| CHN_ARG_GWS | 41.38 | 89.94 | DRY | 174 | 1 | 2 |
| CHN_HOR_AFF | 42.72 | 122.37 | WOOD | 1366 | 1 | 16 |
| CHN_YIN_ST1 | 42.45 | 85.72 | DRY | 105 | 1 | 5 |
| CRI_TAM_TOW | 10.39 | -84.63 | TROP | 666 | 17 | 26 |
| CZE_BIL_BIL | 49.25 | 16.69 | TEMP* | 238 | 1 | 6 |
| CZE_KRT_KRT | 49.32 | 16.75 | TEMP* | 238 | 1 | 6 |
| CZE_LAN | 48.68 | 16.95 | TEMP* | 1093 | 3 | 17 |
| CZE_RAJ_RAJ | 49.44 | 16.70 | TEMP* | 274 | 1 | 6 |
| CZE_SOBO_SOBO | 49.25 | 16.69 | TEMP* | 655 | 1 | 6 |
| CZE_STI | 49.04 | 17.97 | TEMP | 263 | 1 | 8 |
| CZE_UTE_BPO | 49.28 | 16.65 | TEMP* | 234 | 1 | 6 |
| DEU_HIN_OAK | 53.33 | 13.19 | TEMP* | 482 | 1 | 8 |
| DEU_HIN_TER | 53.33 | 13.19 | TEMP* | 1052 | 2 | 16 |
| DEU_MER_BEE_NON | 49.27 | 7.81 | TEMP | 495 | 1 | 8 |
| DEU_MER_DOU_NON | 49.27 | 7.81 | TEMP | 491 | 1 | 7 |
| DEU_MER_MIX_NON | 49.27 | 7.81 | TEMP | 1108 | 2 | 17 |
| DEU_STE_2P3 | 53.10 | 13.00 | TEMP* | 722 | 1 | 10 |
| DEU_STE_4P5 | 53.10 | 13.00 | TEMP* | 327 | 1 | 10 |
| ESP_ALT_ARM | 40.78 | -2.33 | WOOD | 1990 | 3 | 15 |
| ESP_ALT_HUE | 40.79 | -2.29 | WOOD | 967 | 2 | 8 |
| ESP_ALT_TRI | 40.80 | -2.23 | WOOD | 1522 | 2 | 12 |
| ESP_CAN | 41.43 | 2.07 | WOOD | 2317 | 4 | 21 |
| ESP_GUA_VAL | 40.90 | -4.03 | WOOD | 2100 | 1 | 24 |
| ESP_LAS | 28.31 | -16.57 | WOOD | 1778 | 1 | 10 |
| ESP_MAJ_MAI | 39.94 | -5.77 | WOOD | 978 | 1 | 6 |
| ESP_MON_SIE_NAT | 41.12 | -3.50 | WOOD | 1250 | 3 | 20 |
| ESP_RIN | 40.60 | -6.02 | WOOD | 502 | 1 | 8 |
| ESP_RON_PIL | 36.69 | -5.02 | TEMP | 911 | 2 | 12 |
| ESP_TIL_MIX | 41.33 | 1.01 | WOOD | 3434 | 2 | 32 |
| ESP_TIL_OAK | 41.33 | 1.01 | WOOD | 717 | 1 | 10 |
| ESP_TIL_PIN | 41.33 | 1.01 | WOOD | 589 | 1 | 9 |
| ESP_VAL_BAR | 42.20 | 1.82 | WOOD | 837 | 1 | 12 |
| ESP_VAL_SOR | 42.20 | 1.81 | WOOD | 1109 | 1 | 13 |
| ESP_YUN_C1 | 36.72 | -4.97 | WOOD | 619 | 1 | 6 |

Table C.1: SAPFLUXNET sites included in the study. Biome was calculated using Whittaker diagram. *Indicates that the biome was manually adjusted and confirmed by SAPFLUXNET contributors. (*continued*)

| Site code | Latitude | Longitude | Biome | # Tree-days | # Species | # Trees |
|-----------------|----------|-----------|-------|-------------|-----------|---------|
| ESP_YUN_C2 | 36.72 | -4.97 | WOOD | 288 | 1 | 6 |
| FIN_HYY_SME | 61.85 | 24.29 | TEMP | 34 | 2 | 4 |
| FIN_PET | 69.49 | 27.23 | BOR* | 118 | 1 | 7 |
| FRA_FON | 48.48 | 2.78 | TEMP* | 276 | 1 | 3 |
| FRA_HES_HE1_NON | 48.67 | 7.06 | TEMP | 620 | 1 | 10 |
| FRA_HES_HE2_NON | 48.67 | 7.06 | TEMP | 1347 | 1 | 10 |
| FRA_PUE | 43.74 | 3.60 | WOOD | 5229 | 1 | 25 |
| GBR_ABE_PLO | 56.62 | -3.80 | TEMP | 486 | 1 | 15 |
| GBR_DEV_CON | 56.03 | -3.72 | TEMP* | 133 | 1 | 4 |
| GBR_GUI_ST1 | 57.27 | -4.82 | TEMP | 398 | 1 | 15 |
| GBR_GUI_ST2 | 57.27 | -4.82 | TEMP | 298 | 1 | 9 |
| GBR_GUI_ST3 | 57.27 | -4.82 | TEMP | 249 | 1 | 8 |
| GUF_GUY_GUY | 5.28 | -52.92 | TROP | 246 | 6 | 6 |
| GUF_GUY_ST2 | 5.28 | -52.91 | TROP | 369 | 7 | 11 |
| GUF_NOU_PET | 4.08 | -52.68 | TROP | 562 | 10 | 22 |
| HUN_SI | 47.93 | 20.44 | WOOD | 365 | 2 | 4 |
| ISR_YAT_YAT | 31.34 | 35.05 | DRY | 3704 | 1 | 24 |
| ITA_FEI_S17 | 46.69 | 10.61 | TEMP | 244 | 1 | 6 |
| ITA_KAE_S20 | 46.70 | 10.61 | BOR | 325 | 1 | 6 |
| ITA_MUN | 46.68 | 10.58 | TEMP* | 384 | 1 | 6 |
| ITA_REN | 46.59 | 11.43 | TEMP | 247 | 3 | 8 |
| ITA_RUN_N20 | 46.70 | 10.64 | BOR | 331 | 2 | 8 |
| MEX_COR_YP | 19.49 | -97.04 | TEMP | 119 | 1 | 8 |
| NLD_LOO | 52.17 | 5.74 | TEMP* | 621 | 1 | 6 |
| NLD_SPE_DOU | 52.25 | 5.69 | TEMP* | 107 | 1 | 3 |
| NZL_HUA_HUA | -36.80 | 174.49 | TEMP | 243 | 1 | 6 |
| PRT_LEZ_ARN | 38.83 | -8.82 | WOOD | 403 | 1 | 4 |
| PRT_MIT | 38.54 | -8.00 | WOOD | 494 | 1 | 4 |
| PRT_PIN | 38.25 | -8.76 | WOOD | 1233 | 2 | 20 |
| RUS_CHE_Y4 | 68.74 | 161.41 | BOR | 447 | 1 | 11 |
| RUS_FYO | 56.46 | 32.92 | TEMP | 1132 | 3 | 17 |
| RUS_POG_VAR | 56.36 | 92.95 | TEMP* | 603 | 3 | 9 |
| SEN_SOUPRE | 16.34 | -15.43 | DRY | 466 | 1 | 3 |
| SWE_NOR_ST1_BEF | 60.09 | 17.48 | TEMP* | 653 | 2 | 22 |
| SWE_NOR_ST2 | 60.09 | 17.48 | TEMP* | 175 | 2 | 12 |
| SWE_NOR_ST3 | 60.09 | 17.48 | TEMP* | 810 | 2 | 37 |
| SWE_NOR_ST5_REF | 60.08 | 17.48 | TEMP* | 712 | 3 | 35 |
| SWE_SKO_MIN | 58.36 | 12.15 | TEMP | 533 | 1 | 11 |
| SWE_SKY_38Y | 60.13 | 17.84 | TEMP* | 326 | 1 | 12 |
| SWE_SKY_68Y | 60.10 | 17.83 | TEMP* | 664 | 2 | 12 |
| SWE_SVA_MIX_NON | 64.26 | 19.77 | TEMP | 861 | 2 | 20 |
| THA_KHU | 15.27 | 103.08 | TROP | 411 | 1 | 6 |
| USA_BNZ_BLA | 64.70 | -148.32 | BOR* | 797 | 1 | 6 |
| USA_CHE_ASP | 45.94 | -90.27 | TEMP | 3548 | 6 | 149 |
| USA_CHE_MAP | 45.95 | -90.26 | TEMP | 2651 | 2 | 153 |
| USA_DUK_HAR | 36.98 | -79.09 | TEMP | 495 | 6 | 34 |
| USA_HIL_HF2 | 36.22 | -78.86 | TEMP | 228 | 5 | 23 |
| USA_INM | 39.32 | -86.41 | TEMP | 766 | 6 | 9 |
| USA_MOR_SF | 39.32 | -86.41 | TEMP | 285 | 4 | 6 |
| USA_NWH | 34.58 | -91.26 | TEMP | 248 | 2 | 10 |
| USA_ORN_ST1_AMB | 35.90 | -84.33 | TEMP | 247 | 1 | 8 |
| USA_PAR_FER | 35.80 | -76.67 | TEMP | 467 | 1 | 8 |
| USA_PER_PER | 30.21 | -83.87 | TROP | 6269 | 1 | 80 |

Table C.1: SAPFLUXNET sites included in the study. Biome was calculated using Whittaker diagram. *Indicates that the biome was manually adjusted and confirmed by SAPFLUXNET contributors. (*continued*)

| Site code | Latitude | Longitude | Biome | # Tree-days | # Species | # Trees |
|-----------------|----------|-----------|-------|-------------|-----------|---------|
| USA_PJS_P04_AMB | 34.39 | -106.53 | DRY | 2313 | 2 | 10 |
| USA_PJS_P08_AMB | 34.39 | -106.53 | DRY | 2262 | 2 | 10 |
| USA_PJS_P12_AMB | 34.39 | -106.53 | DRY | 2350 | 2 | 10 |
| USA_SIL_OAK_1PR | 39.92 | -74.60 | TEMP | 1210 | 4 | 18 |
| USA_SIL_OAK_2PR | 39.92 | -74.60 | TEMP | 2275 | 4 | 22 |
| USA_SMI_SER | 38.89 | -76.56 | TEMP* | 1045 | 5 | 31 |
| USA_SWH | 34.11 | -91.13 | TEMP | 511 | 2 | 16 |
| USA_SYL_HL1 | 46.24 | -89.35 | TEMP | 3130 | 3 | 48 |
| USA_SYL_HL2 | 46.24 | -89.35 | TEMP | 1631 | 4 | 20 |
| USA_TNB | 36.47 | -84.70 | TEMP | 583 | 4 | 8 |
| USA_TNO | 35.97 | -84.28 | TEMP | 680 | 5 | 9 |
| USA_TNP | 35.96 | -84.29 | TEMP | 806 | 5 | 9 |
| USA_UMB_CON | 45.56 | -84.71 | TEMP | 5840 | 5 | 57 |
| USA_UMB_GIR | 45.56 | -84.70 | TEMP | 5867 | 4 | 57 |
| USA_WIL_WC1 | 45.81 | -90.09 | TEMP | 639 | 5 | 16 |
| USA_WVF | 39.06 | -79.69 | TEMP | 488 | 5 | 8 |
| ZAF_FRA_FRA | -33.88 | 19.06 | WOOD | 220 | 1 | 3 |
| ZAF_RAD | -34.08 | 19.11 | WOOD | 303 | 1 | 3 |
| ZAF_SOU_SOU | -34.09 | 19.09 | WOOD | 198 | 1 | 2 |
| ZAF_WEL_SOR | -33.48 | 18.96 | WOOD* | 356 | 1 | 3 |

Table C.2: SAPFLUXNET stand treatments included in the this study (Chapter 3).

| Plot treatment |
|---|
| None |
| Control |
| control |
| Ambient Control |
| Control - Unthinned |
| natural conditions |
| Reference |
| 1Premortality |
| 2premortality |
| destructive sampling |
| Girdling early successional |
| Pre-thinning |
| Before thinning |
| Before Thinning |
| non thinned |
| none (periodict thinning every 5-6 years 20 to 25% of basal area) |
| Radiation Level |
| AMBIENT CO2 FACE rings |
| fertilization at plantation |
| AcaciaMonoculture |
| MixtureEucalyptusAndAcacia |
| EucalyptusMonoculture |
| Pre Irrigation |

Table C.3: Summary table of site level R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} , climate, soil properties and vegetation structure data. PPET is in [mm mm⁻¹], P-PET_{sd} is in [mm], Clay and Sand are in [%], Total N is in [g kg⁻¹], Stand height is in [m], LAI is in [m²_{leaves} m²_{soil}]. Letters show data source: a = SAPFLUXNET, b = Global rasters, c = SAPFLUXNET plant height.

| Site code | R^2_{VPD} | R^2_{SWC} | R^2_{PPFD} | Relimp VPD | Relimp SWC | Relimp PPFD | PPET | P - PET _{sd} | Clay | Sand | Total N | Bedrock | Stand height | LAI |
|-----------------|-------------|-------------|--------------|------------|------------|-------------|------|-----------------------|---------|---------|---------|---------|--------------|--------|
| AUS_CAN_ST1_EUC | 0.74 | 0.44 | 0.54 | 0.65 | 0.35 | 0.00 | 1.23 | 47.52 | 26.30 b | 45.10 b | 1.02 | 184 | 22.00 a | 1.39 a |
| AUS_CAN_ST2_MIX | 0.91 | 0.65 | 0.76 | 0.87 | 0.13 | 0.00 | 1.23 | 47.52 | 26.30 b | 45.10 b | 1.02 | 184 | 21.80 a | 2.07 a |
| AUS_CAN_ST3 ACA | 0.85 | 0.67 | 0.74 | 0.87 | 0.12 | 0.01 | 1.23 | 47.52 | 26.30 b | 45.10 b | 1.02 | 184 | 11.80 a | 1.35 a |
| AUS_CAR THI CON | 0.67 | 0.00 | 0.00 | 0.99 | 0.00 | 0.01 | 1.36 | 49.01 | 27.20 b | 44.30 b | 2.34 | 111 | 17.21 a | 4.80 a |
| AUS_ELL_UNB | 0.87 | 0.44 | 0.76 | 0.97 | 0.00 | 0.03 | 1.08 | 67.16 | 26.70 b | 48.50 b | 1.95 | 63 | 25.00 a | 6.20 b |
| AUS_MAR_UBD | 0.79 | 0.53 | 0.69 | 0.71 | 0.29 | 0.00 | 1.35 | 70.37 | 26.60 b | 44.60 b | 1.90 | 89 | 25.00 a | 2.10 a |
| AUS_MAR_UBW | 0.90 | 0.81 | 0.82 | 0.90 | 0.00 | 0.09 | 1.21 | 65.38 | 27.90 b | 43.90 b | 2.00 | 173 | 40.00 a | 2.30 a |
| AUS_WOM | 0.82 | 0.52 | 0.49 | 0.79 | 0.00 | 0.20 | 1.09 | 69.35 | 25.90 b | 52.90 b | 1.97 | 172 | 22.00 a | 2.20 a |
| AUT_PAT_FOR | 0.78 | 0.73 | 0.68 | 0.82 | 0.09 | 0.09 | 2.17 | 16.78 | 5.00 a | 60.00 a | 3.94 | 180 | 12.00 a | 4.30 b |
| AUT_PAT_KRU | 0.61 | 0.45 | 0.50 | 0.77 | 0.06 | 0.17 | 2.17 | 16.78 | 5.00 a | 60.00 a | 3.94 | 180 | 0.75 a | 4.30 b |
| AUT_PAT_TRE | 0.58 | 0.27 | 0.19 | 0.70 | 0.30 | 0.00 | 2.17 | 16.78 | 5.00 a | 60.00 a | 3.94 | 180 | 4.00 a | 4.30 b |
| BRA_CAM | 0.83 | 0.68 | 0.69 | 0.65 | 0.28 | 0.07 | 1.66 | 88.82 | 27.60 b | 52.00 b | 2.26 | 200 | 12.00 a | 5.30 a |
| BRA_CAX_CON | 0.79 | 0.74 | 0.73 | 0.70 | 0.00 | 0.30 | 1.90 | 122.90 | 8.00 a | 79.00 a | 1.45 | 197 | 38.00 b | 5.30 a |
| CAN_TUR_P39_PRE | 0.59 | 0.37 | 0.34 | 0.71 | 0.08 | 0.21 | 1.39 | 42.08 | 1.00 a | 98.00 a | 1.58 | 200 | 23.40 a | 5.30 a |
| CAN_TUR_P74 | 0.26 | 0.47 | 0.14 | 0.32 | 0.38 | 0.30 | 1.39 | 41.87 | 1.00 a | 98.00 a | 1.60 | 200 | 16.20 a | 6.70 a |
| CHN_ARG_GWS | 0.42 | 0.32 | 0.27 | 0.51 | 0.47 | 0.01 | 0.01 | 63.51 | 17.70 b | 46.00 b | 0.70 | 172 | 7.90 a | 0.36 a |
| CHN_HOR_AFF | 0.39 | 0.38 | 0.34 | 0.25 | 0.75 | 0.00 | 0.59 | 31.24 | 8.00 a | 83.00 a | 1.00 | 200 | 9.05 a | 1.61 a |
| CHN_YIN_ST1 | 0.47 | 0.45 | 0.44 | 0.63 | 0.18 | 0.20 | 0.19 | 35.09 | 20.80 b | 32.90 b | 2.41 | 148 | 10.60 a | 0.50 b |
| CRI_TAM_TOW | 0.73 | 0.73 | 0.73 | 0.46 | 0.11 | 0.44 | 3.57 | 159.99 | 36.10 b | 34.70 b | 2.75 | 200 | 30.60 a | 3.30 a |
| CZE_BIL_BIL | 0.54 | 0.50 | 0.41 | 0.57 | 0.20 | 0.23 | 0.71 | 28.98 | 29.60 b | 27.40 b | 1.91 | 200 | 14.00 a | 6.00 b |
| CZE_KRT_KRT | 0.57 | 0.45 | 0.30 | 0.58 | 0.03 | 0.39 | 0.85 | 27.00 | 26.00 b | 27.40 b | 2.10 | 200 | 17.00 a | 5.70 b |
| CZE_LAN | 0.79 | 0.76 | 0.72 | 0.67 | 0.08 | 0.25 | 0.66 | 37.49 | 17.80 a | 71.80 a | 2.46 | 200 | 36.00 a | 6.04 a |
| CZE_RAJ_RAJ | 0.38 | 0.40 | 0.43 | 0.33 | 0.07 | 0.60 | 0.99 | 26.14 | 21.80 b | 33.90 b | 1.96 | 200 | 18.00 a | 4.60 b |
| CZE_SOZ_SOZ | 0.48 | 0.45 | 0.14 | 0.52 | 0.25 | 0.23 | 0.71 | 28.98 | 29.60 b | 27.40 b | 1.91 | 200 | 21.00 a | 6.00 b |
| CZE_STI | 0.54 | 0.32 | 0.39 | 0.70 | 0.18 | 0.12 | 1.13 | 27.10 | 34.20 a | 47.60 a | 1.65 | 200 | 31.00 a | 5.50 a |
| CZE_UTE_BPO | 0.62 | 0.65 | 0.52 | 0.46 | 0.16 | 0.38 | 0.75 | 29.86 | 26.70 b | 23.80 b | 2.71 | 200 | 18.00 a | 6.10 b |
| DEU_HIN_OAK | 0.43 | 0.19 | 0.33 | 0.92 | 0.07 | 0.01 | 0.95 | 35.10 | 17.90 b | 49.90 b | 2.42 | 200 | 31.45 c | 5.70 b |
| DEU_HIN_TER | 0.31 | 0.22 | 0.24 | 0.76 | 0.01 | 0.24 | 0.95 | 35.10 | 18.00 b | 50.50 b | 2.05 | 200 | 24.43 c | 5.60 b |
| DEU_MER_BEE_NON | 0.47 | 0.27 | 0.29 | 0.76 | 0.06 | 0.18 | 1.48 | 47.33 | 4.00 a | 71.00 a | 2.56 | 200 | 23.00 a | 5.90 a |
| DEU_MER_DOU_NON | 0.45 | 0.25 | 0.22 | 0.60 | 0.31 | 0.10 | 1.48 | 47.33 | 4.00 a | 71.00 a | 2.56 | 200 | 29.00 a | 5.30 a |
| DEU_MER_MIX_NON | 0.41 | 0.23 | 0.26 | 0.82 | 0.02 | 0.16 | 1.48 | 47.33 | 4.00 a | 71.00 a | 2.56 | 200 | 30.00 a | 6.10 a |
| DEU_STE_2P3 | 0.56 | 0.15 | 0.30 | 0.86 | 0.09 | 0.05 | 0.90 | 37.45 | 2.50 a | 92.50 a | 3.28 | 200 | 27.20 a | 4.30 b |
| DEU_STE_4P5 | 0.48 | 0.27 | 0.31 | 0.64 | 0.28 | 0.09 | 0.90 | 37.45 | 2.50 a | 92.50 a | 3.28 | 200 | 27.20 a | 4.30 b |
| ESP_ALT_ARM | 0.51 | 0.30 | 0.29 | 0.91 | 0.01 | 0.09 | 0.66 | 65.80 | 21.90 b | 41.50 b | 1.27 | 187 | 19.00 b | 1.09 a |

Table C.3: Summary table of site level R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} , climate, soil properties and vegetation structure data. PPET is in [mm mm⁻¹], P-PET_{sd} is in [mm], Clay and Sand are in [%], Total N is in [g kg⁻¹], Stand height is in [m], LAI is in [$m^2_{leaves} m^2_{soil}$]. Letters show data source: a = SAPFLUXNET, b = Global rasters, c = SAPFLUXNET plant height. (*continued*)

| Site code | R^2_{VPD} | R^2_{SWC} | R^2_{PPFD} | Relimp VPD | Relimp SWC | Relimp PPFD | PPET | P - PET _{sd} | Clay | Sand | Total N | Bedrock | Stand height | LAI |
|---------------------|-------------|-------------|--------------|------------|------------|-------------|-------|-----------------------|---------|---------|---------|---------|--------------|--------|
| ESP_ALT_HUE | 0.47 | 0.22 | 0.18 | 0.78 | 0.02 | 0.21 | 0.51 | 63.08 | 21.60 b | 35.90 b | 1.46 | 200 | 8.64 c | 1.50 b |
| ESP_ALT_TRI | 0.58 | 0.44 | 0.31 | 0.80 | 0.07 | 0.12 | 0.57 | 63.48 | 21.00 b | 40.00 b | 1.31 | 196 | 4.89 a | 1.60 b |
| ESP_CAN | 0.55 | 0.40 | 0.34 | 0.74 | 0.02 | 0.24 | 0.94 | 46.91 | 32.90 b | 28.30 b | 1.76 | 179 | 10.80 a | 3.30 a |
| ESP_GUA_VAL | 0.51 | 0.28 | 0.23 | 0.69 | 0.00 | 0.31 | 0.68 | 69.09 | 24.80 b | 40.90 b | 1.27 | 200 | 12.00 a | 3.80 a |
| ESP_LAS | 0.37 | 0.25 | 0.07 | 0.57 | 0.37 | 0.06 | 1.63 | 37.89 | 1.00 a | 70.00 a | 1.65 | 197 | 10.30 a | 3.60 a |
| ESP_MAJ_MAI | 0.56 | 0.43 | 0.28 | 0.73 | 0.14 | 0.12 | 0.76 | 97.45 | 9.00 a | 80.00 a | 1.18 | 200 | 7.00 a | 0.30 a |
| ESP_MON_SIE_NAT | 0.25 | 0.20 | 0.22 | 0.49 | 0.09 | 0.42 | 0.62 | 63.05 | 20.80 b | 41.90 b | 1.45 | 200 | 22.00 a | 3.30 b |
| ESP_RIN | 0.83 | 0.61 | 0.59 | 0.97 | 0.00 | 0.03 | 0.85 | 76.30 | 15.00 a | 9.00 a | 2.17 | 200 | 7.40 a | 3.40 a |
| ESP_RON_PIL | 0.37 | 0.25 | 0.17 | 0.60 | 0.07 | 0.33 | 1.05 | 93.66 | 18.00 a | 30.00 a | 1.86 | 200 | 2.60 a | 0.90 b |
| ESP_TIL_MIX | 0.45 | 0.36 | 0.22 | 0.55 | 0.09 | 0.36 | 0.77 | 48.10 | 20.00 a | 60.00 a | 1.44 | 162 | 14.20 a | 3.27 a |
| ESP_TIL_OAK | 0.31 | 0.38 | 0.18 | 0.45 | 0.15 | 0.40 | 0.77 | 48.10 | 20.00 a | 60.00 a | 1.44 | 162 | 5.00 a | 4.59 a |
| ESP_TIL_PIN | 0.37 | 0.45 | 0.08 | 0.43 | 0.37 | 0.20 | 0.79 | 48.10 | 20.00 a | 60.00 a | 1.78 | 188 | 18.30 a | 1.02 a |
| ESP_VAL_BAR | 0.61 | 0.22 | 0.29 | 0.90 | 0.00 | 0.10 | 0.70 | 34.07 | 32.63 a | 9.81 a | 1.94 | 200 | 10.60 a | 2.10 a |
| ESP_VAL_SOR | 0.58 | 0.36 | 0.26 | 0.73 | 0.14 | 0.13 | 0.78 | 32.15 | 20.00 a | 60.00 a | 2.04 | 200 | 11.00 a | 2.40 a |
| ESP_YUN_C1 | 0.37 | 0.50 | 0.24 | 0.28 | 0.56 | 0.16 | 0.83 | 93.65 | 29.00 a | 22.00 a | 1.37 | 197 | 10.60 a | 2.20 b |
| ESP_YUN_C2 | 0.28 | 0.62 | 0.27 | 0.21 | 0.43 | 0.37 | 0.78 | 91.33 | 29.00 a | 22.00 a | 1.37 | 188 | 11.60 a | 2.50 b |
| FIN_HYY_SME | 0.57 | 0.49 | 0.62 | 0.12 | 0.86 | 0.02 | 1.20 | 38.32 | 6.50 a | 37.00 a | 1.67 | 200 | 18.00 a | 1.30 a |
| FIN_PET | 0.59 | 0.50 | 0.58 | 0.47 | 0.37 | 0.16 | 1.13 | 26.34 | 7.30 b | 60.80 b | 5.08 | 200 | 3.76 a | 0.61 a |
| FRA_FON | 0.78 | 0.68 | 0.68 | 0.70 | 0.17 | 0.13 | 0.89 | 45.10 | 19.00 a | 37.00 a | 1.26 | 200 | 28.00 a | 6.00 a |
| FRA_HES_HE1_NON0.46 | 0.57 | 0.41 | 0.45 | 0.25 | 0.30 | 1.31 | 47.72 | 25.00 a | 8.00 a | 1.41 | 200 | 12.80 a | 6.00 a | |
| FRA_HES_HE2_NON0.33 | 0.50 | 0.16 | 0.42 | 0.27 | 0.31 | 1.31 | 47.72 | 25.00 a | 8.00 a | 1.41 | 200 | 13.00 a | 6.00 a | |
| FRA_PUE | 0.49 | 0.52 | 0.33 | 0.51 | 0.18 | 0.30 | 1.27 | 70.16 | 39.00 a | 26.00 a | 1.69 | 195 | 5.00 a | 2.40 a |
| GBR_ABE_PLO | 0.40 | 0.37 | 0.35 | 0.51 | 0.22 | 0.27 | 1.92 | 47.48 | 10.00 a | 60.00 a | 3.70 | 179 | 10.00 a | 6.00 a |
| GBR_DEV_CON | 0.89 | 0.45 | 0.58 | 0.95 | 0.01 | 0.04 | 1.43 | 44.38 | 14.80 b | 56.90 b | 3.44 | 200 | 15.00 a | 1.92 a |
| GBR_GUI_ST1 | 0.84 | 0.79 | 0.78 | 0.68 | 0.00 | 0.32 | 3.19 | 68.11 | 3.70 b | 80.40 b | 14.26 | 197 | 11.00 a | 0.92 a |
| GBR_GUI_ST2 | 0.64 | 0.55 | 0.49 | 0.59 | 0.03 | 0.38 | 3.19 | 68.11 | 3.70 b | 80.40 b | 14.26 | 197 | 13.30 a | 0.94 a |
| GBR_GUI_ST3 | 0.86 | 0.82 | 0.79 | 0.73 | 0.03 | 0.24 | 3.19 | 68.11 | 3.70 b | 80.40 b | 14.26 | 197 | 14.30 a | 1.57 a |
| GUF_GUY_GUY | 0.97 | 0.89 | 0.94 | 0.82 | 0.05 | 0.12 | 2.88 | 135.18 | 43.00 a | 48.00 a | 1.53 | 200 | 35.00 a | 7.00 a |
| GUF_GUY_ST2 | 0.80 | 0.77 | 0.76 | 0.61 | 0.30 | 0.09 | 3.02 | 141.34 | 43.20 a | 47.80 a | 1.66 | 200 | 35.00 a | 6.70 a |
| GUF_NOU_PET | 0.76 | 0.53 | 0.66 | 1.00 | 0.00 | 0.00 | 2.69 | 158.16 | 59.20 a | 33.20 a | 2.22 | 200 | 35.00 a | 5.50 a |
| HUN_SIK | 0.86 | 0.42 | 0.56 | 0.90 | 0.00 | 0.09 | 0.70 | 39.64 | 30.40 b | 44.00 b | 1.64 | 200 | 20.00 a | 7.00 a |
| ISR_YAT_YAT | 0.40 | 0.32 | 0.18 | 0.62 | 0.37 | 0.01 | 0.28 | 83.43 | 28.00 a | 31.00 a | 0.71 | 178 | 11.00 a | 1.70 a |
| ITA_FEI_S17 | 0.53 | 0.37 | 0.30 | 0.66 | 0.14 | 0.19 | 1.08 | 22.97 | 8.00 a | 76.00 a | 3.11 | 117 | 20.00 a | 3.10 b |
| ITA_KAE_S20 | 0.68 | 0.45 | 0.46 | 0.75 | 0.07 | 0.18 | 1.24 | 22.97 | 17.00 a | 50.00 a | 3.64 | 121 | 14.00 a | 2.60 b |

Table C.3: Summary table of site level R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} , climate, soil properties and vegetation structure data. PPET is in [mm mm⁻¹], P-PET_{sd} is in [mm], Clay and Sand are in [%], Total N is in [g kg⁻¹], Stand height is in [m], LAI is in [m²_{leaves} m²_{soil}]. Letters show data source: a = SAPFLUXNET, b = Global rasters, c = SAPFLUXNET plant height. (*continued*)

| Site code | R^2_{VPD} | R^2_{SWC} | R^2_{PPFD} | Relimp VPD | Relimp SWC | Relimp PPFD | PPET | P - PET _{sd} | Clay | Sand | Total N | Bedrock | Stand height | LAI |
|---------------------|-------------|-------------|--------------|------------|------------|-------------|-------|-----------------------|---------|---------|---------|---------|--------------|--------|
| ITA_MUN | 0.63 | 0.48 | 0.42 | 0.62 | 0.31 | 0.06 | 0.80 | 29.87 | 7.00 a | 55.00 a | 1.93 | 188 | 18.00 a | 2.20 b |
| ITA_REN | 0.85 | 0.77 | 0.79 | 0.95 | 0.02 | 0.04 | 1.61 | 12.59 | 17.70 b | 47.90 b | 2.73 | 143 | 27.00 b | 4.60 b |
| ITA_RUN_N20 | 0.84 | 0.73 | 0.74 | 0.94 | 0.02 | 0.04 | 1.39 | 15.28 | 14.00 a | 54.00 a | 3.33 | 123 | 18.70 a | 5.70 b |
| MEX_COR_YP | 0.68 | 0.24 | 0.31 | 0.78 | 0.05 | 0.18 | 1.42 | 81.77 | 22.20 b | 46.40 b | 2.94 | 200 | 7.00 a | 5.20 a |
| NLD_LOO | 0.55 | 0.31 | 0.27 | 0.74 | 0.02 | 0.24 | 1.33 | 41.67 | 1.00 a | 99.00 a | 2.61 | 200 | 18.00 a | 2.20 a |
| NLD_SPE_DOU | 0.76 | 0.60 | 0.63 | 0.81 | 0.09 | 0.11 | 1.42 | 39.80 | 4.80 b | 80.70 b | 1.62 | 200 | 30.00 a | 4.50 a |
| NZL_HUA_HUA | 0.71 | 0.62 | 0.61 | 0.75 | 0.17 | 0.08 | 2.62 | 42.52 | 71.20 a | 13.20 a | 1.73 | 200 | 27.00 a | 6.60 b |
| PRT_LEZ_ARN | 0.70 | 0.23 | 0.25 | 0.77 | 0.02 | 0.21 | 0.72 | 77.42 | 5.04 a | 90.38 a | 1.52 | 200 | 12.00 a | 1.50 a |
| PRTMIT | 0.76 | 0.47 | 0.36 | 0.78 | 0.18 | 0.04 | 0.51 | 80.80 | 16.10 b | 64.50 b | 1.33 | 200 | 7.50 a | 0.55 a |
| PRT_PIN | 0.65 | 0.50 | 0.36 | 0.66 | 0.33 | 0.01 | 0.76 | 74.76 | 16.60 b | 61.20 b | 1.26 | 200 | 12.60 a | 1.10 b |
| RUS_CHE_Y4 | 0.31 | 0.21 | 0.23 | 0.95 | 0.01 | 0.04 | 0.62 | 34.23 | 21.10 b | 23.20 b | 4.96 | 200 | 7.00 a | 1.30 b |
| RUS_FYO | 0.72 | 0.62 | 0.62 | 0.88 | 0.00 | 0.12 | 1.24 | 30.87 | 18.20 b | 48.80 b | 3.77 | 198 | 23.50 a | 3.50 a |
| RUS_POG_VAR | 0.78 | 0.49 | 0.61 | 0.89 | 0.00 | 0.11 | 0.70 | 33.02 | 28.60 b | 37.50 b | 2.64 | 200 | 22.00 a | 2.80 b |
| SEN_SOU_PRE | 0.75 | 0.35 | 0.16 | 0.79 | 0.21 | 0.00 | 0.13 | 43.94 | 6.00 a | 90.00 a | 0.23 | 200 | 7.00 a | 0.22 a |
| SWE_NOR_ST1_BEFO.75 | 0.62 | 0.62 | 0.68 | 0.19 | 0.13 | 1.07 | 36.70 | 5.80 a | 58.60 a | 2.63 | 185 | 28.70 a | 4.18 a | |
| SWE_NOR_ST2 | 0.26 | 0.21 | 0.17 | 0.56 | 0.00 | 0.44 | 1.07 | 36.70 | 5.80 a | 58.60 a | 2.63 | 185 | 27.70 a | 6.15 a |
| SWE_NOR_ST3 | 0.62 | 0.61 | 0.60 | 0.67 | 0.08 | 0.25 | 1.07 | 36.70 | 5.80 a | 58.60 a | 2.63 | 185 | 27.20 a | 4.55 a |
| SWE_NOR_ST5_REF0.59 | 0.62 | 0.57 | 0.50 | 0.20 | 0.30 | 1.07 | 36.55 | 19.20 b | 43.50 b | 2.83 | 190 | 20.00 a | 5.00 a | |
| SWE_SKO_MIN | 0.76 | 0.74 | 0.70 | 0.55 | 0.04 | 0.41 | 1.60 | 45.86 | 17.30 b | 52.00 b | 2.48 | 133 | 28.00 a | 6.50 a |
| SWE_SKY_38Y | 0.38 | 0.46 | 0.40 | 0.12 | 0.88 | 0.00 | 1.39 | 33.61 | 21.70 b | 43.80 b | 3.93 | 184 | 13.60 a | 3.98 a |
| SWE_SKY_68Y | 0.38 | 0.57 | 0.37 | 0.10 | 0.76 | 0.14 | 1.30 | 33.80 | 18.90 b | 46.50 b | 4.15 | 184 | 20.30 a | 3.83 a |
| SWE_SVA_MIX_NON0.64 | 0.47 | 0.54 | 0.88 | 0.11 | 0.00 | 1.33 | 34.34 | 0.50 a | 92.50 a | 1.67 | 200 | 15.00 a | 3.80 b | |
| THA_KHU | 0.51 | 0.38 | 0.39 | 0.71 | 0.25 | 0.05 | 0.83 | 84.24 | 10.00 a | 65.00 a | 0.75 | 200 | 15.00 a | 3.90 a |
| USA_BNZ_BLA | 0.66 | 0.43 | 0.56 | 0.78 | 0.17 | 0.06 | 0.69 | 33.86 | 10.30 b | 36.80 b | 2.57 | 200 | 3.00 a | 3.60 b |
| USA_CHE_ASP | 0.69 | 0.34 | 0.30 | 0.92 | 0.02 | 0.06 | 1.23 | 20.06 | 12.00 a | 74.00 a | 1.52 | 200 | 10.00 a | 4.50 a |
| USA_CHE_MAP | 0.71 | 0.62 | 0.65 | 0.81 | 0.02 | 0.17 | 1.22 | 19.85 | 6.63 a | 59.31 a | 2.54 | 200 | 18.00 a | 3.90 a |
| USA_DUK_HAR | 0.72 | 0.62 | 0.68 | 0.86 | 0.03 | 0.10 | 1.12 | 41.33 | 33.90 b | 31.00 b | 0.76 | 200 | 25.00 a | 7.03 a |
| USA_HIL_HF2 | 0.72 | 0.67 | 0.71 | 0.59 | 0.00 | 0.41 | 1.14 | 37.46 | 26.00 a | 43.00 a | 0.71 | 200 | 15.00 a | 5.50 a |
| USA_INM | 0.52 | 0.42 | 0.47 | 0.53 | 0.00 | 0.47 | 1.18 | 39.20 | 26.70 b | 8.00 b | 1.05 | 200 | 30.00 a | 4.90 a |
| USA_MOR_SF | 0.73 | 0.52 | 0.50 | 0.89 | 0.10 | 0.01 | 1.18 | 39.20 | 30.00 a | 10.00 a | 1.05 | 200 | 27.00 a | 5.00 a |
| USA_NWH | 0.87 | 0.82 | 0.70 | 0.84 | 0.03 | 0.14 | 1.05 | 59.56 | 36.70 b | 4.90 b | 0.80 | 200 | 22.70 a | 5.60 b |
| USA_ORN_ST1_AMB0.64 | 0.62 | 0.56 | 0.52 | 0.08 | 0.41 | 1.14 | 61.36 | 24.00 a | 21.00 a | 0.85 | 200 | 17.90 a | 5.50 a | |
| USA_PAR_FER | 0.49 | 0.16 | 0.23 | 0.68 | 0.03 | 0.29 | 1.32 | 25.96 | 10.00 a | 60.00 a | 1.75 | 200 | 18.00 a | 4.20 a |
| USA_PER_PER | 0.61 | 0.30 | 0.33 | 0.80 | 0.01 | 0.19 | 1.32 | 34.41 | 3.40 b | 89.20 b | 6.13 | 200 | 12.00 a | 4.10 a |

Table C.3: Summary table of site level R^2_{VPD} , R^2_{SWC} , R^2_{PPFD} , climate, soil properties and vegetation structure data. PPET is in [mm mm⁻¹], P-PET_{sd} is in [mm], Clay and Sand are in [%], Total N is in [g kg⁻¹], Stand height is in [m], LAI is in [$m^2_{leaves} m^2_{soil}$]. Letters show data source: a = SAPFLUXNET, b = Global rasters, c = SAPFLUXNET plant height. (*continued*)

| Site code | R^2_{VPD} | R^2_{SWC} | R^2_{PPFD} | Relimp VPD | Relimp SWC | Relimp PPFD | PPET | P - PET _{sd} | Clay | Sand | Total N | Bedrock | Stand height | LAI |
|-----------------|-------------|-------------|--------------|------------|------------|-------------|------|-----------------------|---------|---------|---------|---------|--------------|--------|
| USA_PJS_P04_AMB | 0.46 | 0.15 | 0.31 | 0.78 | 0.12 | 0.10 | 0.25 | 49.32 | 6.00 a | 52.00 a | 0.82 | 186 | 4.20 a | 0.71 a |
| USA_PJS_P08_AMB | 0.50 | 0.16 | 0.26 | 0.93 | 0.07 | 0.00 | 0.25 | 49.32 | 3.00 a | 49.00 a | 0.82 | 186 | 4.10 a | 0.90 a |
| USA_PJS_P12_AMB | 0.36 | 0.17 | 0.11 | 0.65 | 0.30 | 0.05 | 0.25 | 49.32 | 6.00 a | 54.00 a | 0.82 | 186 | 4.00 a | 0.72 a |
| USA_SIL_OAK_1PR | 0.44 | 0.49 | 0.41 | 0.33 | 0.49 | 0.18 | 1.36 | 38.70 | 1.00 a | 98.00 a | 0.74 | 200 | 9.50 a | 3.60 a |
| USA_SIL_OAK_2PR | 0.43 | 0.33 | 0.39 | 0.94 | 0.06 | 0.00 | 1.36 | 38.70 | 1.00 a | 98.00 a | 0.74 | 200 | 9.50 a | 3.60 a |
| USA_SMI_SER | 0.58 | 0.46 | 0.39 | 0.64 | 0.29 | 0.06 | 1.03 | 40.05 | 28.70 b | 30.90 b | 0.82 | 200 | 40.00 a | 5.80 b |
| USA_SWH | 0.82 | 0.56 | 0.52 | 0.92 | 0.02 | 0.06 | 1.09 | 62.29 | 43.10 b | 6.30 b | 0.69 | 200 | 24.20 a | 4.00 b |
| USA_SYL_HL1 | 0.57 | 0.42 | 0.44 | 0.92 | 0.08 | 0.00 | 1.27 | 25.01 | 8.90 b | 51.00 b | 1.41 | 200 | 27.00 a | 5.40 b |
| USA_SYL_HL2 | 0.53 | 0.50 | 0.50 | 0.66 | 0.02 | 0.31 | 1.27 | 25.01 | 8.90 b | 51.00 b | 1.41 | 200 | 27.00 a | 5.40 b |
| USA_TNB | 0.31 | 0.33 | 0.30 | 0.37 | 0.13 | 0.50 | 1.39 | 48.33 | 21.60 b | 34.90 b | 0.84 | 200 | 25.00 a | 4.70 a |
| USA_TNO | 0.38 | 0.39 | 0.37 | 0.41 | 0.22 | 0.38 | 1.41 | 60.02 | 29.60 b | 30.20 b | 0.83 | 200 | 30.00 a | 6.60 a |
| USA_TNP | 0.34 | 0.38 | 0.31 | 0.50 | 0.18 | 0.32 | 1.41 | 61.60 | 31.60 b | 26.60 b | 0.81 | 200 | 25.00 a | 4.50 a |
| USA_UMB_CON | 0.61 | 0.46 | 0.44 | 0.76 | 0.02 | 0.22 | 1.30 | 30.60 | 1.00 a | 92.00 a | 2.02 | 200 | 29.00 a | 3.50 a |
| USA_UMB_GIR | 0.57 | 0.43 | 0.39 | 0.76 | 0.05 | 0.19 | 1.25 | 30.69 | 1.00 a | 92.00 a | 2.49 | 200 | 29.00 a | 3.50 a |
| USA_WIL_WC1 | 0.54 | 0.30 | 0.23 | 0.81 | 0.18 | 0.01 | 1.19 | 20.23 | 6.90 b | 53.20 b | 1.01 | 200 | 24.30 a | 6.20 b |
| USA_WVF | 0.43 | 0.35 | 0.35 | 0.65 | 0.02 | 0.33 | 1.63 | 30.35 | 24.90 b | 29.90 b | 1.37 | 200 | 30.00 a | 6.90 a |
| ZAF_FRA_FRA | 0.61 | 0.08 | 0.24 | 0.96 | 0.01 | 0.03 | 0.90 | 99.17 | 20.00 b | 69.90 b | 0.95 | 200 | 20.00 a | 1.80 a |
| ZAF_RAD | 0.45 | 0.33 | 0.32 | 0.58 | 0.06 | 0.37 | 0.95 | 82.73 | 21.30 b | 61.40 b | 1.18 | 200 | 3.50 a | 2.70 a |
| ZAF_SOUL_SOUL | 0.43 | 0.18 | 0.17 | 0.58 | 0.05 | 0.38 | 0.97 | 86.39 | 23.00 b | 61.90 b | 1.13 | 200 | 4.00 a | 3.00 a |
| ZAF_WEL_SOR | 0.63 | 0.29 | 0.32 | 0.66 | 0.05 | 0.29 | 0.50 | 79.71 | 20.00 a | 60.00 a | 0.81 | 179 | 25.00 a | 1.80 a |

Table C.4: Table of equivalence between Whittaker biomes and the groups of biomes used in the study.

| Original biome name | Study biome group |
|----------------------------------|-------------------|
| Desert | DRY |
| Temperate grassland desert | DRY |
| Subtropical desert | DRY |
| Woodland/shrubland | WOOD |
| Temperate forest | TEMP |
| Boreal forest | BOR |
| Tundra | BOR |
| Tropical rainforest | TROP |
| Tropical seasonal forest/savanna | TROP |

D

Appendix Chapter 5

D.1 Notes D

Notes S1: Whole-tree stomatal conductance (G'_{Asw}) calculation procedure.

To account for aerodynamic effect on whole-tree canopy conductance (G_{Asw}) in trees where wind speed was available, we subtracted from G_{Asw} (calculated with eq. 2 of the main text) the aerodynamic conductance calculated following Tan *et al.* 2019. We first calculate the aerodynamic conductance for momentum (eq. 14 in Tan *et al.* 2019):

$$g_{aM} = \frac{\kappa^2 u}{\ln[(z - d)/z_{0M}] \ln[(z - d)/z_{0H}]} \quad (\text{D.1})$$

Where κ is the Von Karman constant and equivalent to 0.41 and u is wind speed [m s^{-1}] and:

$$z = \text{tree height} + 2, \quad d = 0.6(\text{tree height}), \quad z_{0M} = 0.1(\text{tree height}), \quad z_{0H} = 0.135(z_{0M}) \quad (\text{D.2})$$

Lately, we calculated the boundary layer conductance following eq. 10 in Tan *et al.* (2019):

$$g_{bN} = \frac{\kappa u*}{\ln\left(\frac{z_{0M}}{z_{0H}}\right)} \quad (\text{D.3})$$

Where $u*$ [m s^{-1}] is the friction velocity calculated inverting eq. 2 in Chu *et al.* 2018:

$$u* = \frac{\kappa u}{\ln[(z - d)/z_{0M}] + \ln(1.25)} \quad (\text{D.4})$$

Ultimately, G'_{Asw} was calculated as:

$$G'_{\text{Asw}} = \left(\frac{1}{1/G_{\text{Asw}} - (1/g_{aM} + 1/g_{bN})} \right) \quad (\text{D.5})$$

Being G_{Asw} in [m s^{-1}]. Finally, G'_{Asw} was transformed to [$\text{mol m}_{\text{Asw}}^{-2} \text{s}^{-1}$].

D.2 Figures D

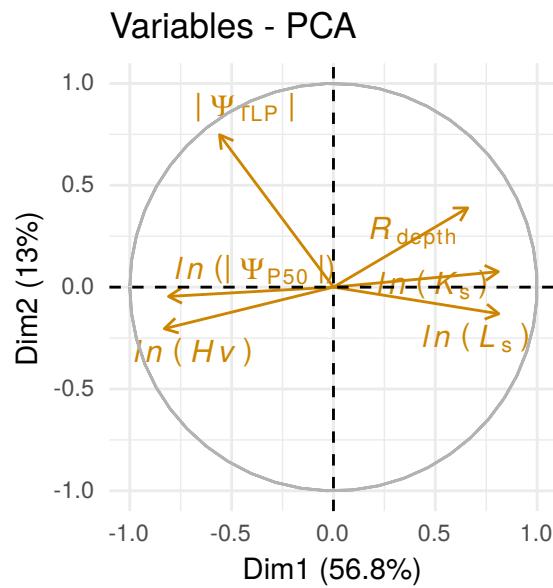


Figure D.1: PCA plot of water relations traits using imputations. $\ln(|\Psi_{P50}|)$: logarithm of absolute water potential at 50% water conductivity loss; $\ln(K_s)$: logarithm of maximum sapwood water conductivity; $\ln(Hv)$: logarithm of Huber value; $|\Psi_{TLP}|$: absolute water potential at turgor-loss point; R_{depth} : rooting depth; $\ln(L_s)$: logarithm of individual leaf area.

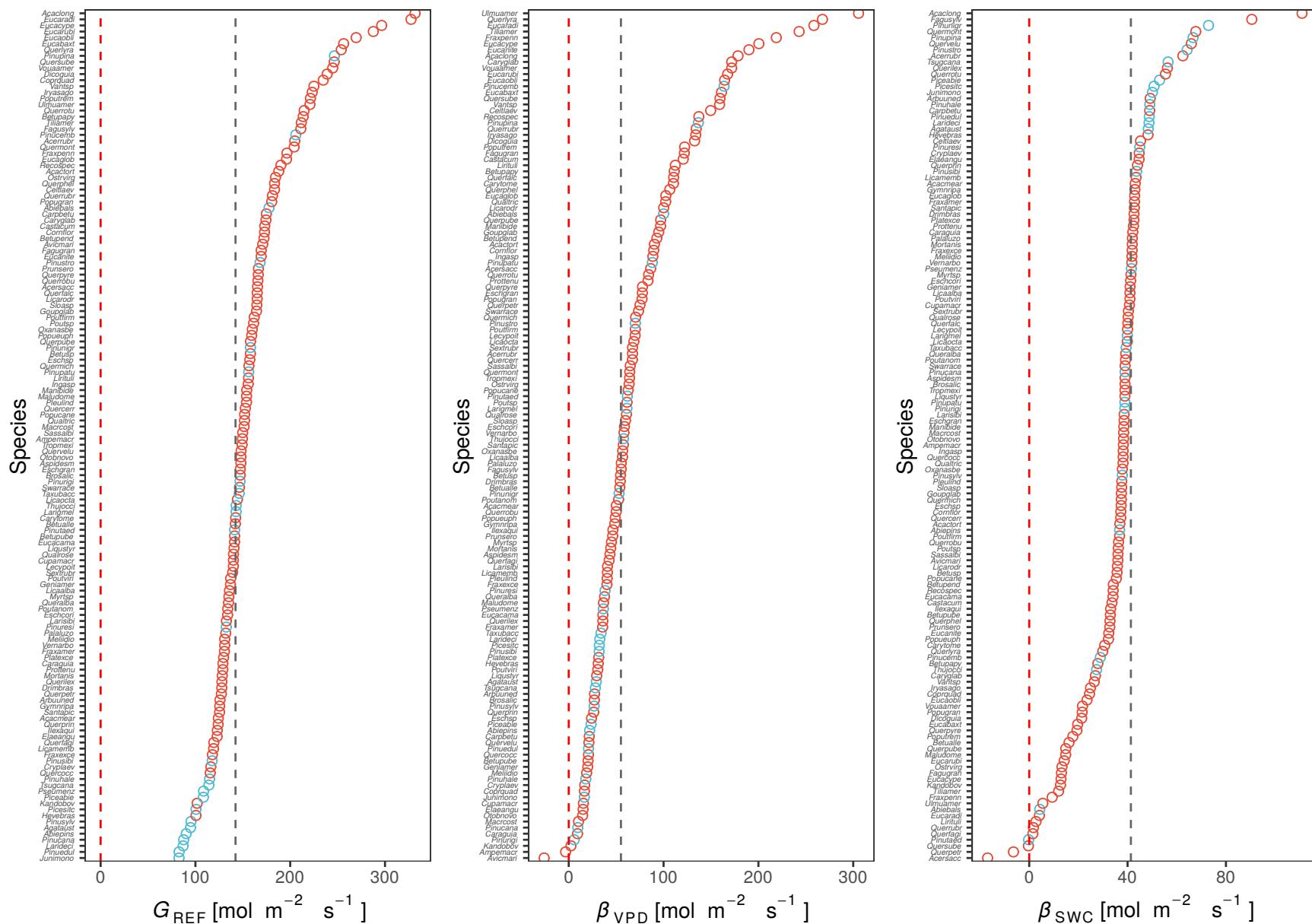
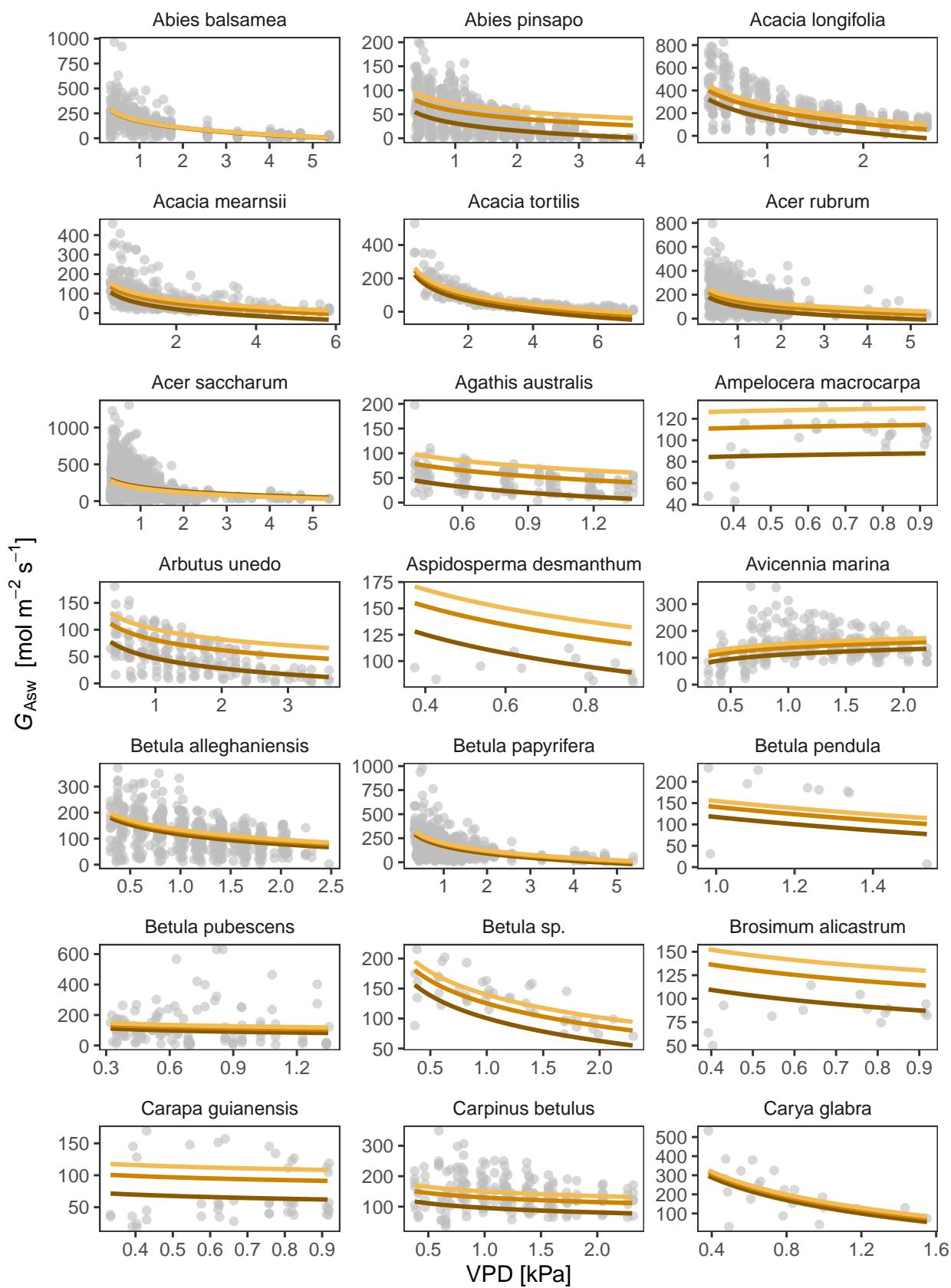
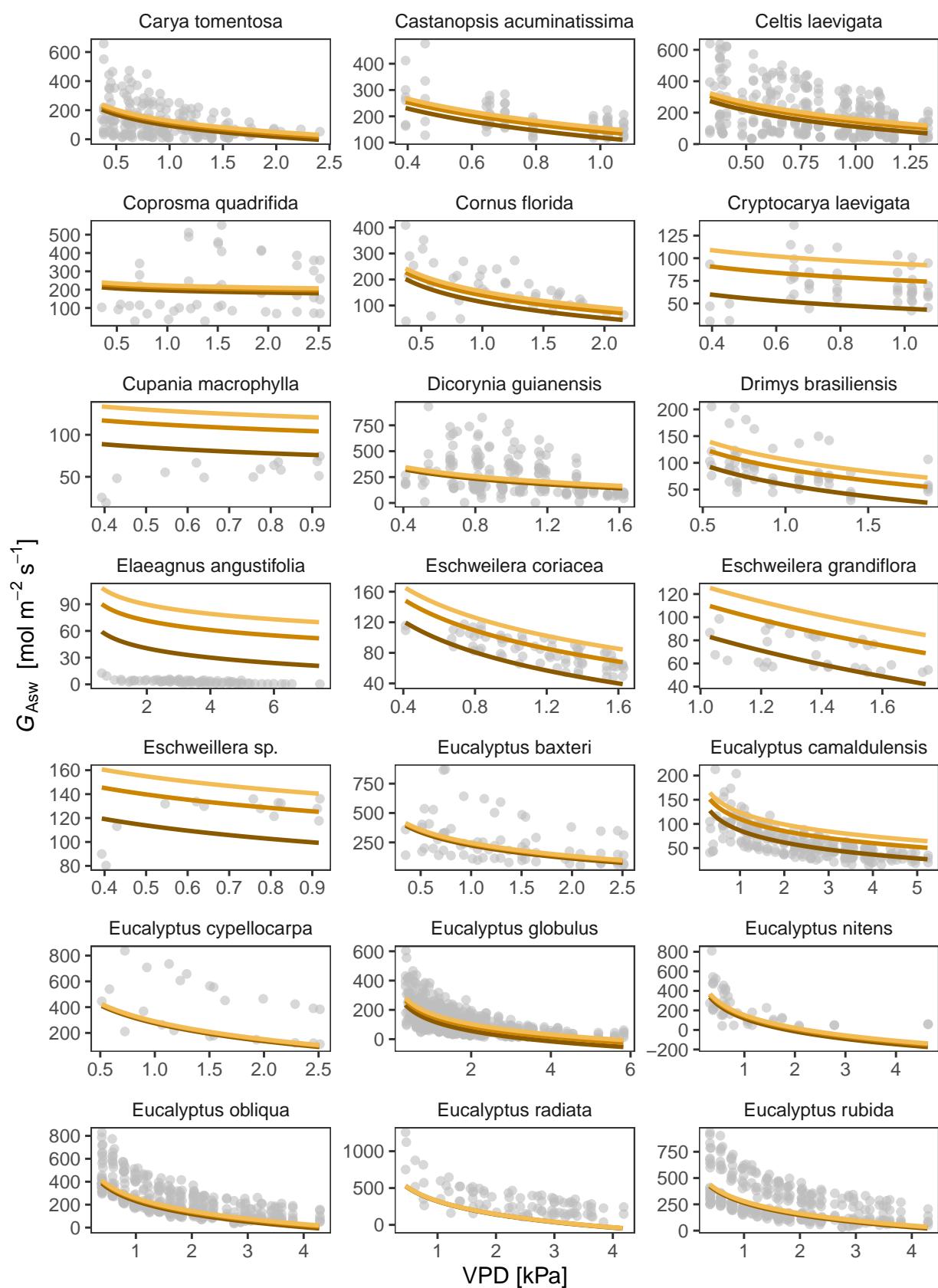
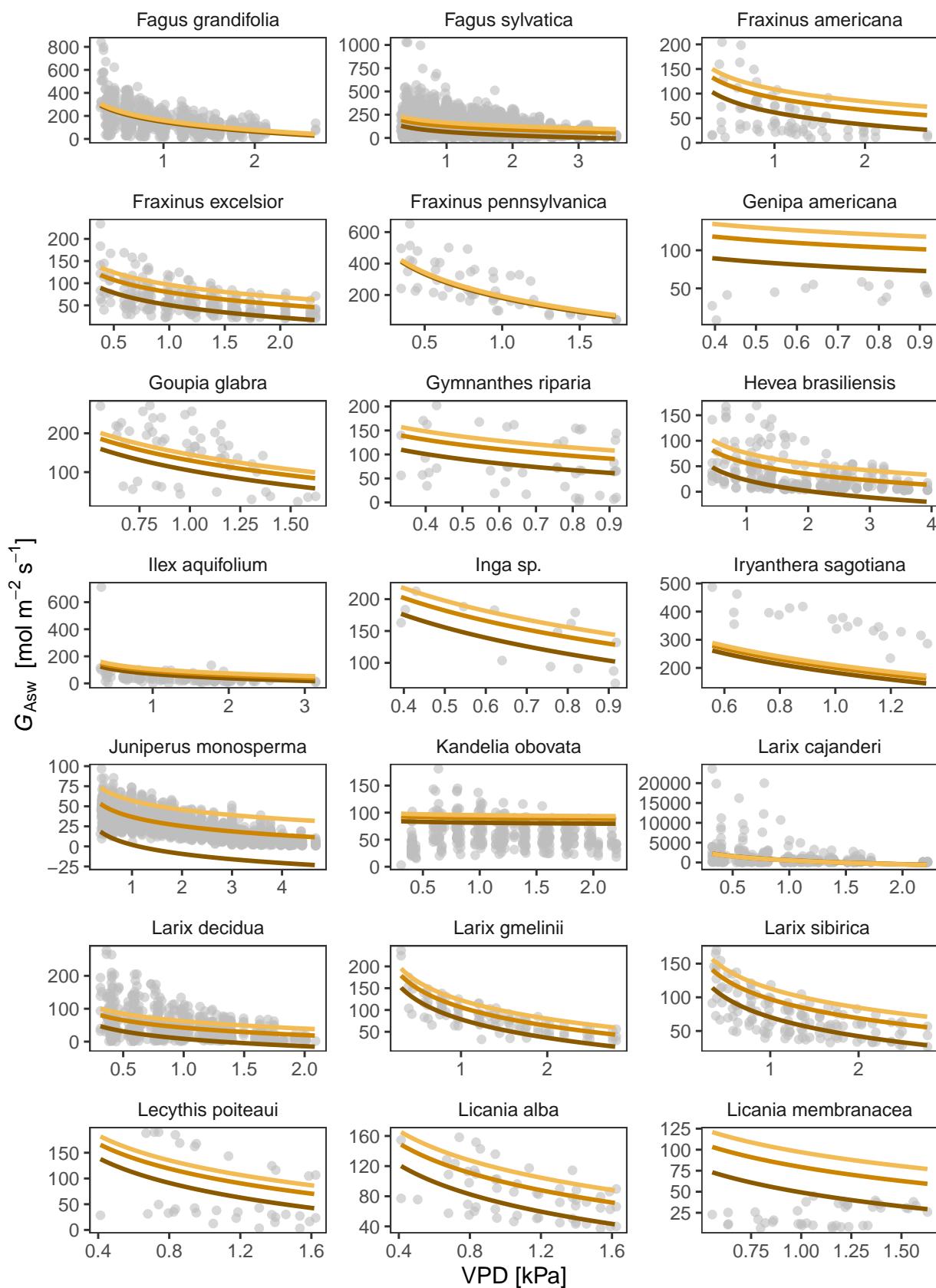
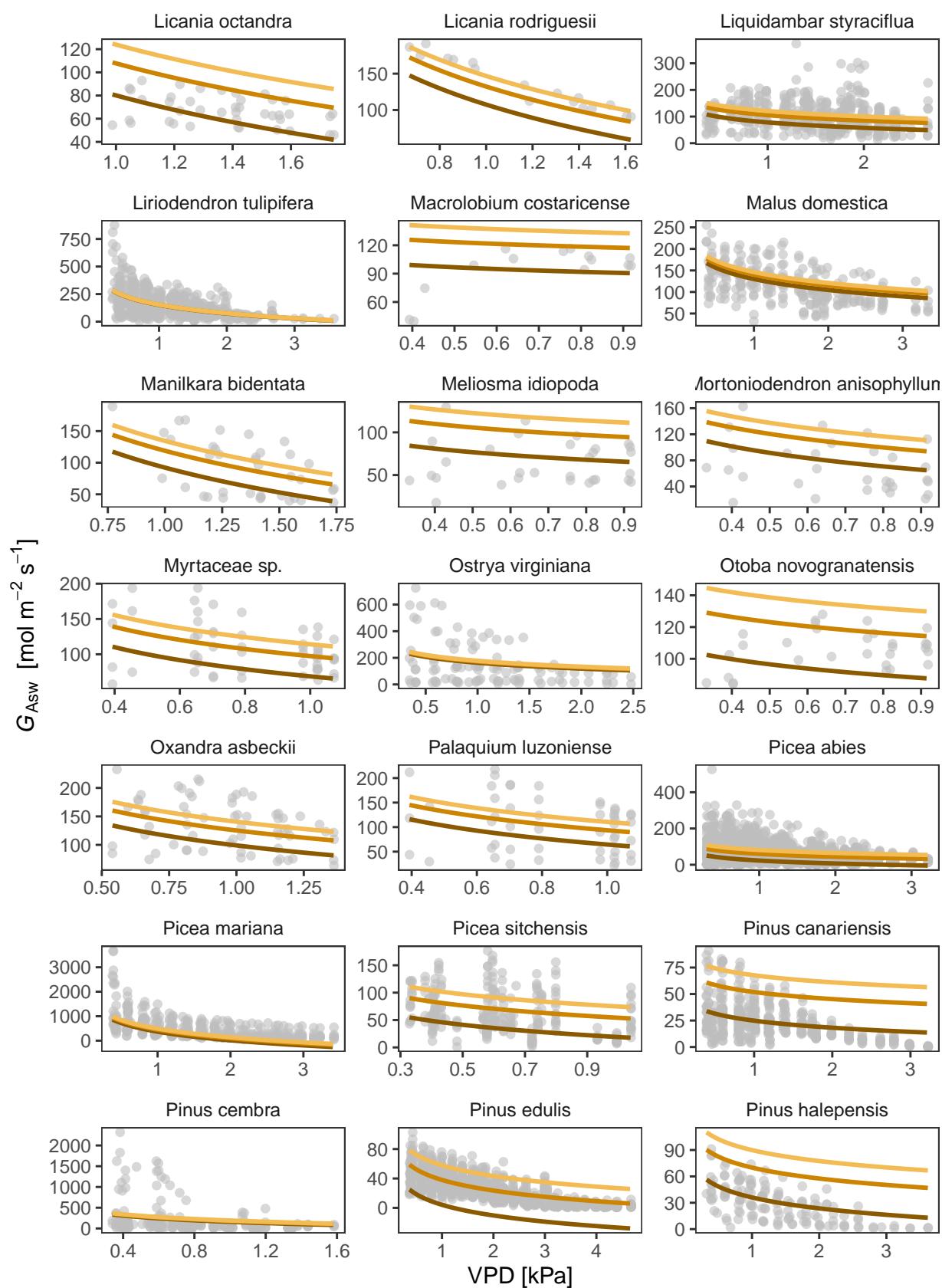


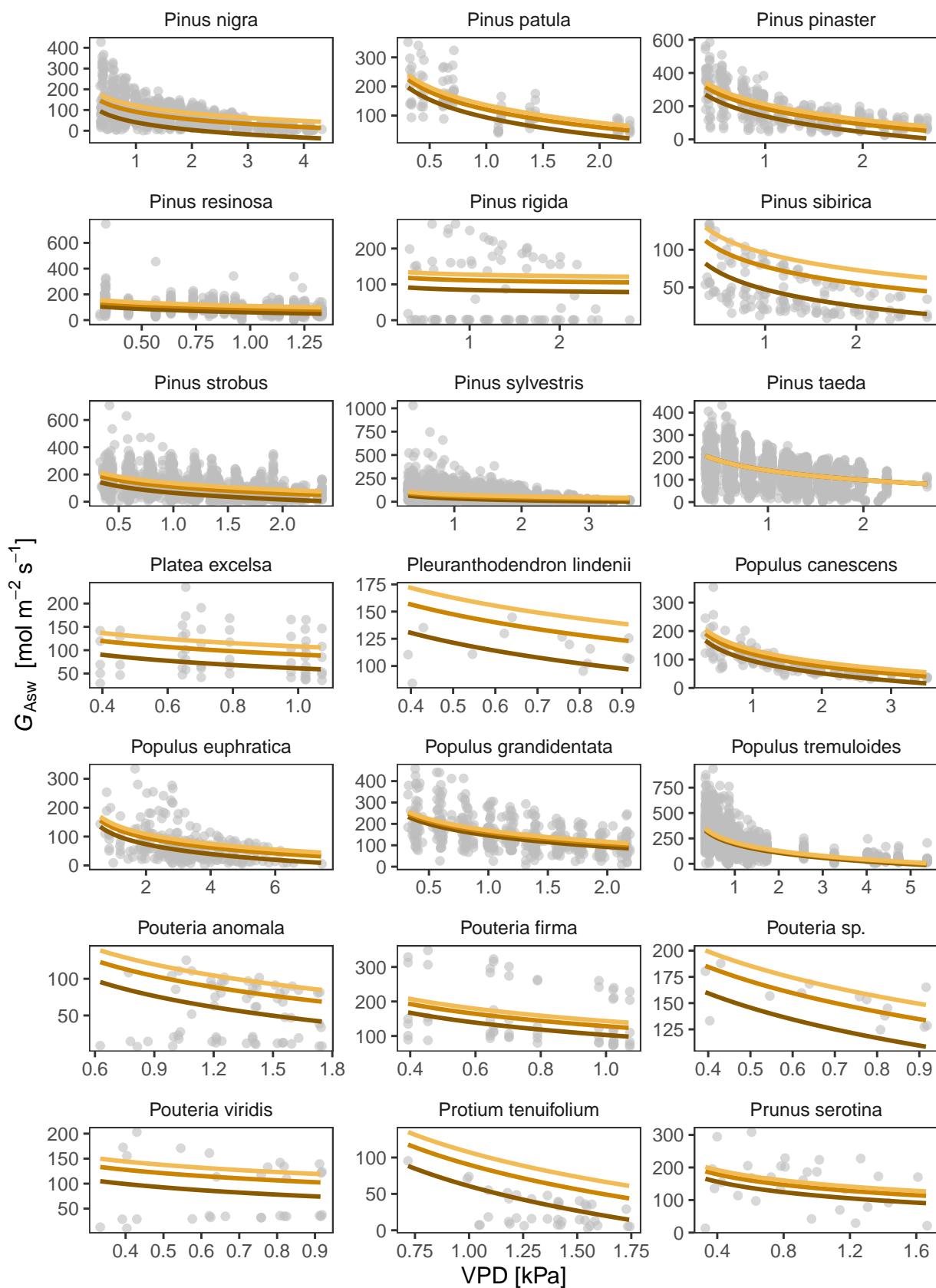
Figure D.2: Water use parameters at the species level. Grey dashed lines are the weighted mean of the parameters

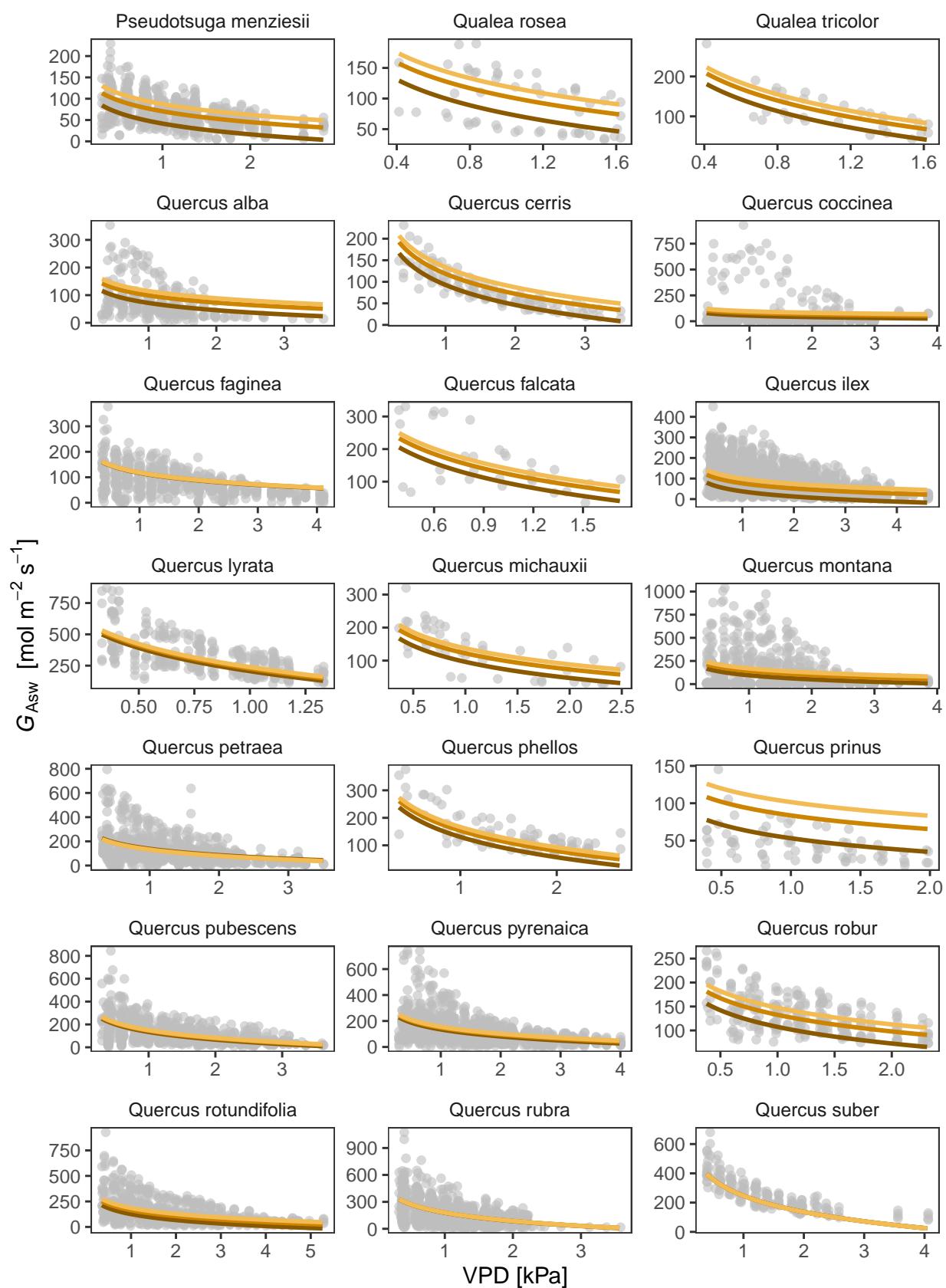












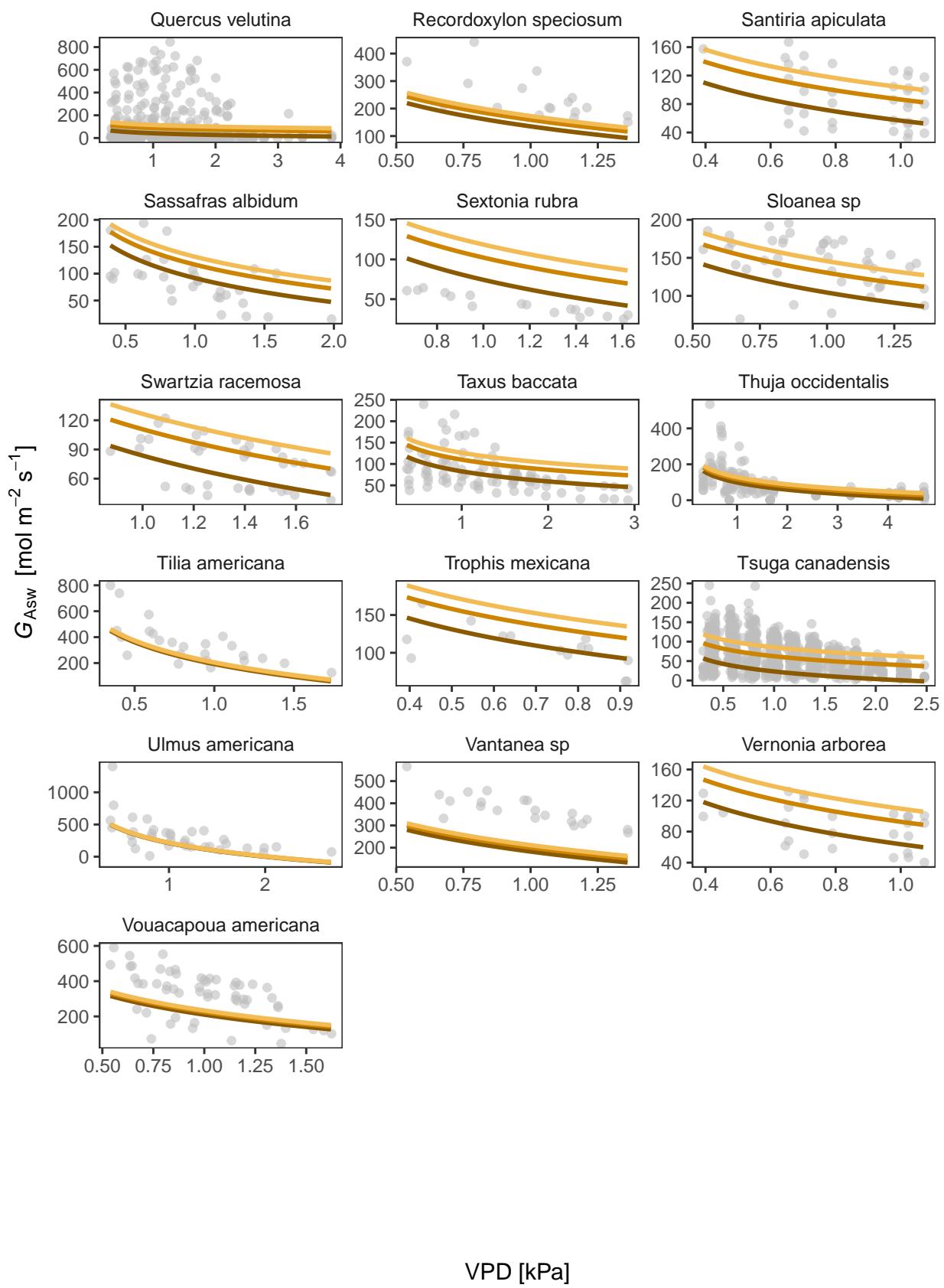
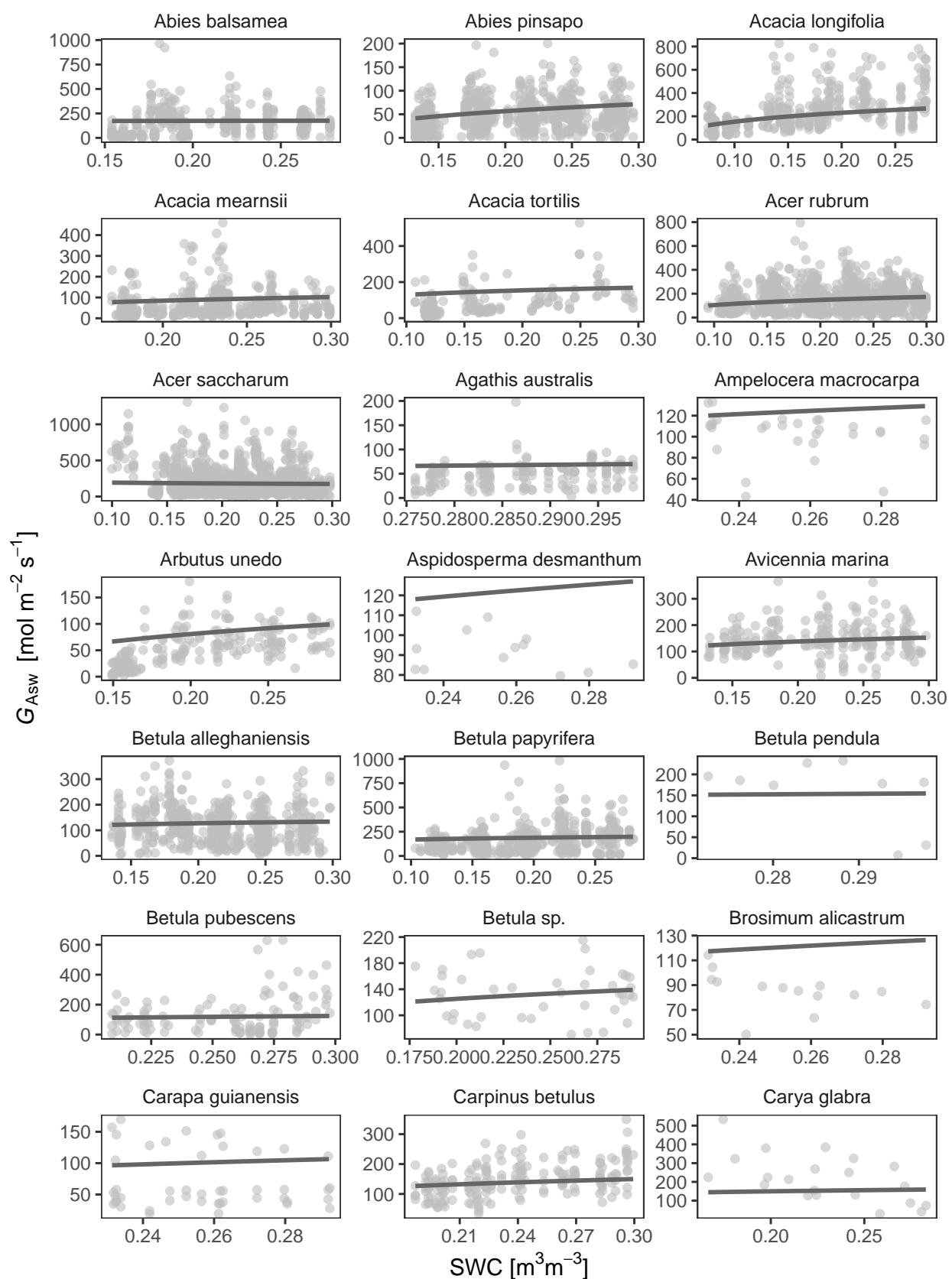
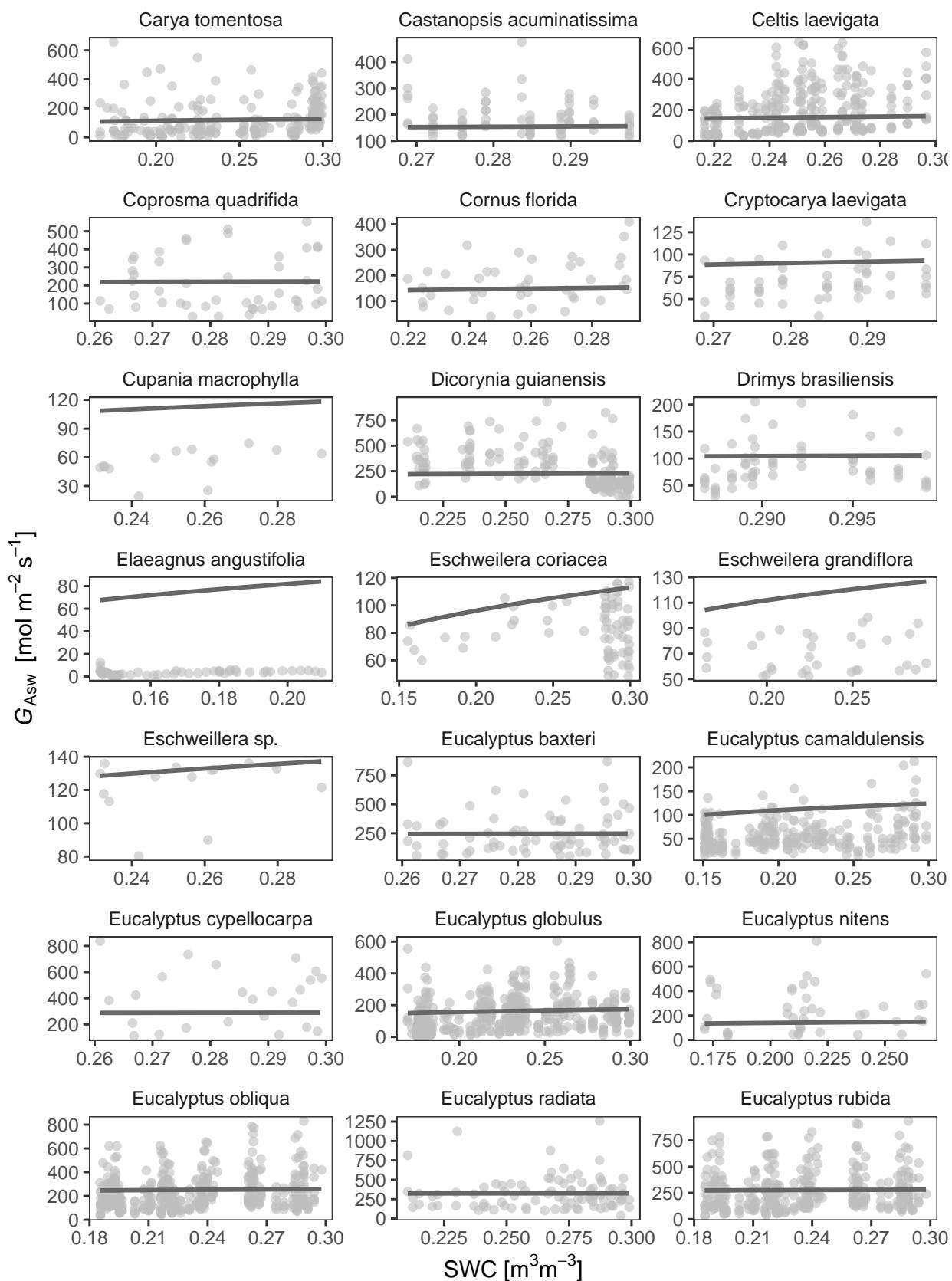
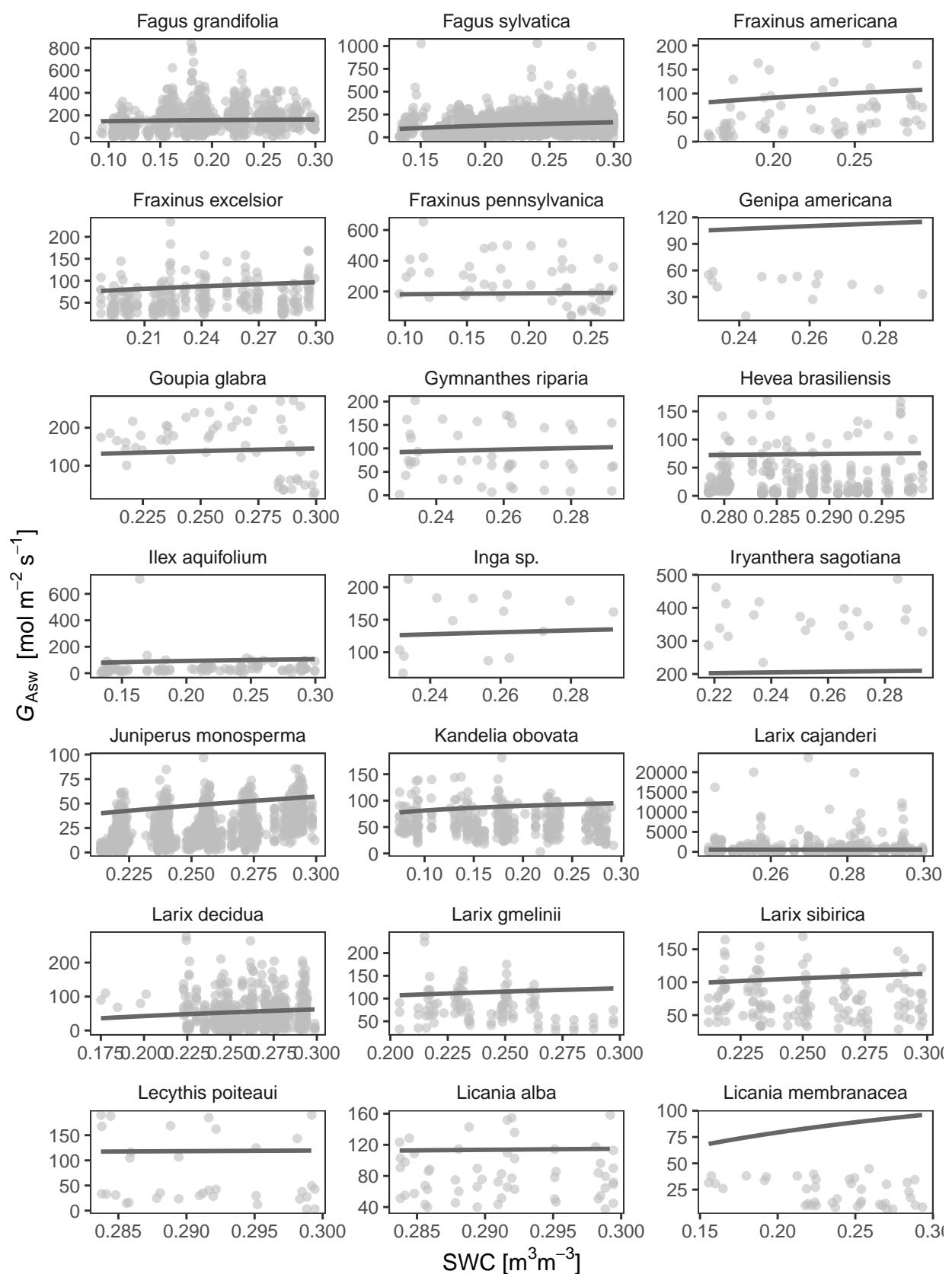
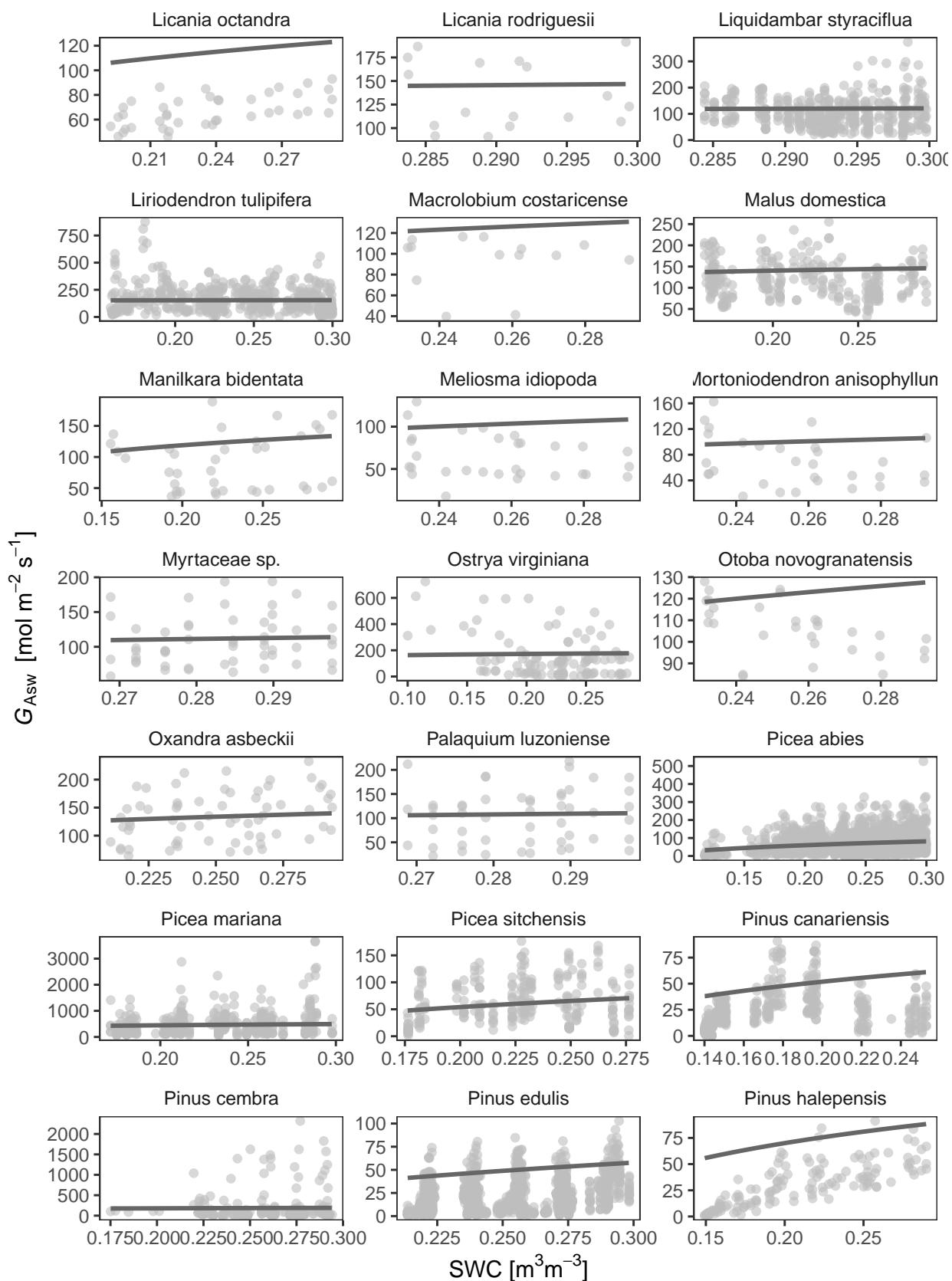


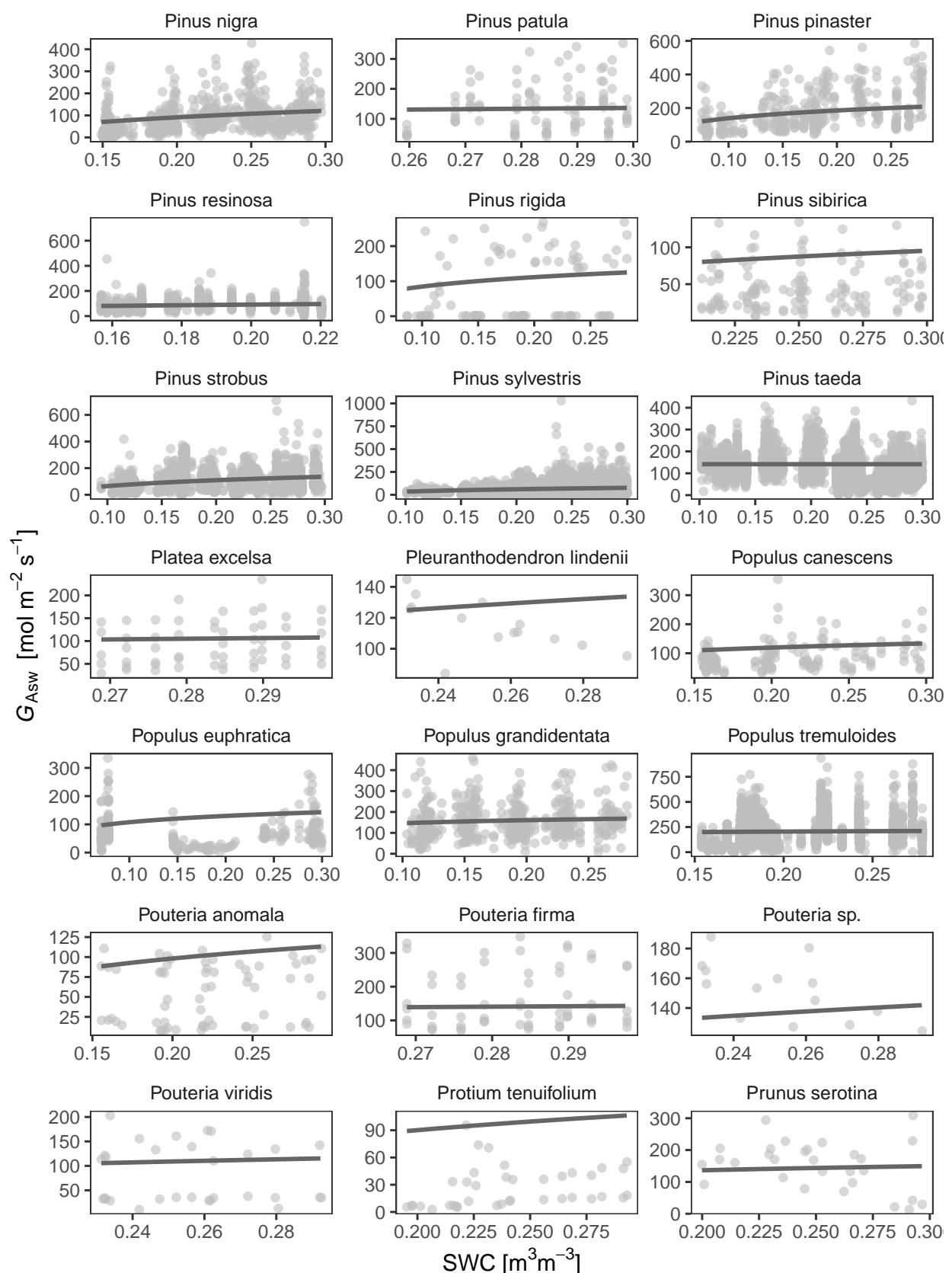
Figure D.3: Species G_{Asw} responses to VPD. Each species have different curves representing distinct levels of SWC (from dark to light, 0.1, 0.2, 0.3 $[\text{m}^3 \text{m}^{-3}]$).

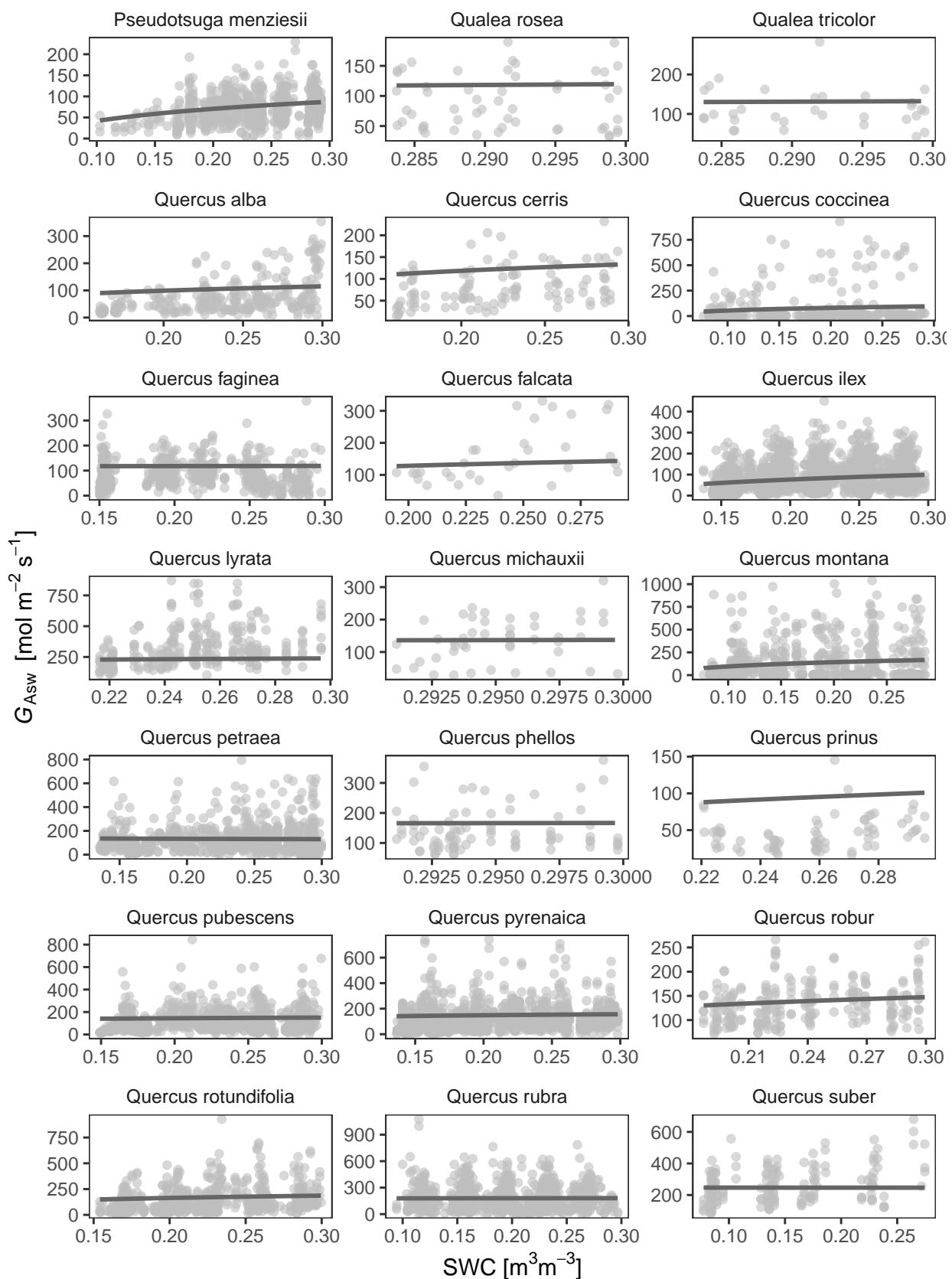












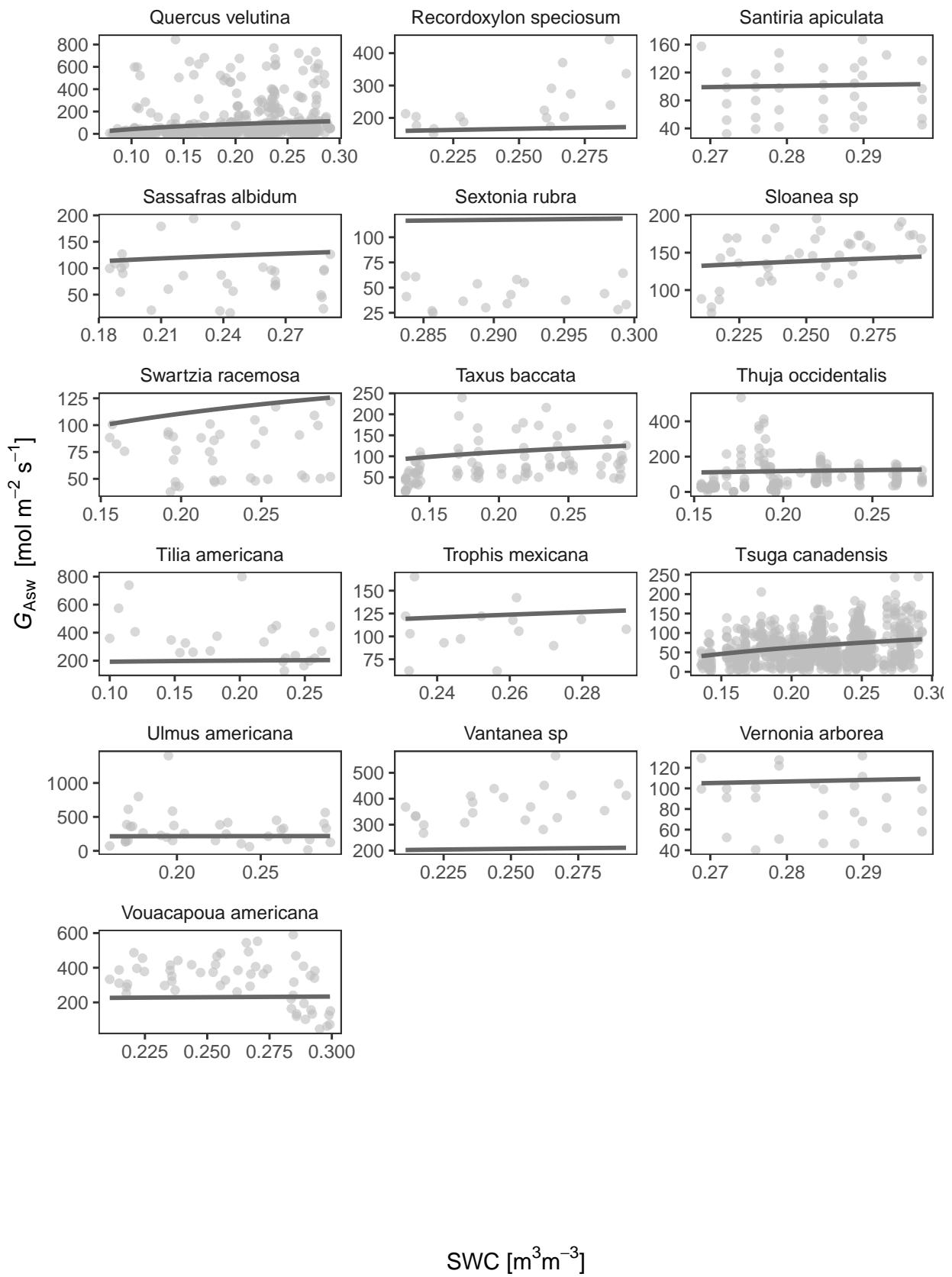


Figure D.4: Species G_{Asw} responses to SWC. Curves are fitted using a VPD reference level of 1 kPa.

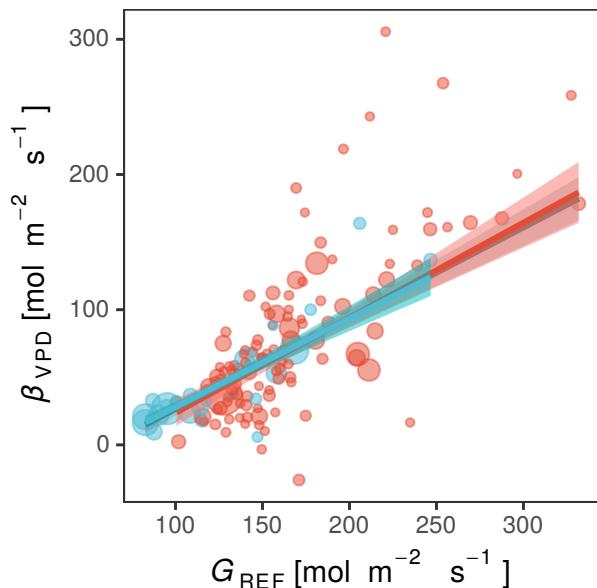


Figure D.5: Scatterplot between G_{REF} and β_{VPD} parameters. Grey line is global relationship. Blue and red lines are angiosperms and gymnosperms relationships, respectively. Shadow areas are 95% confidence interval.

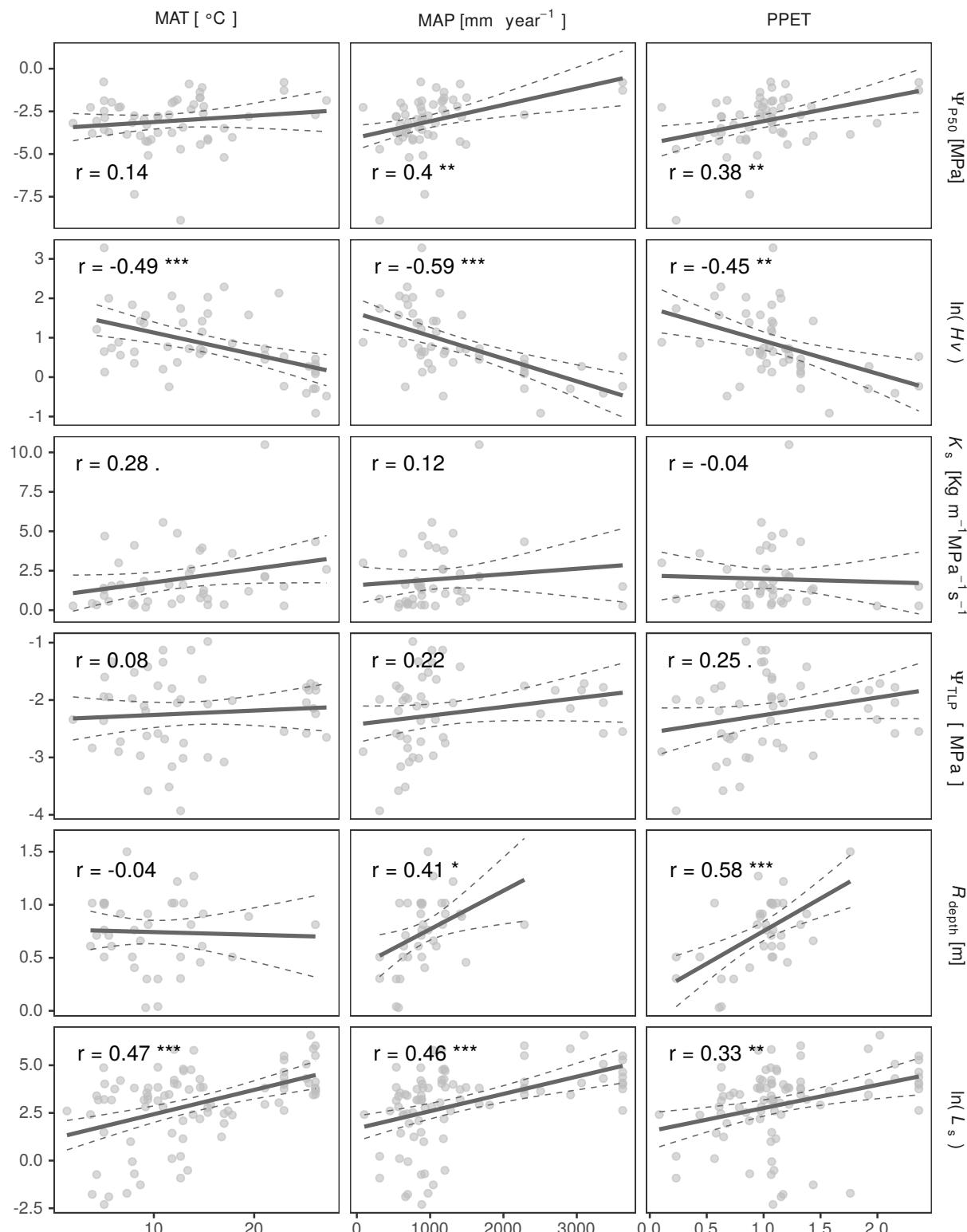


Figure D.6: Species' climatic variables and water relations traits relationships.
MAP: mean annual precipitation, MAT: mean annual temperature, PPET: mean annual precipitation over potential evapotranspiration. All climatic variables were calculated as the weighted average of the characteristic plots of the trees of each species, using tree-days as weighting factor.

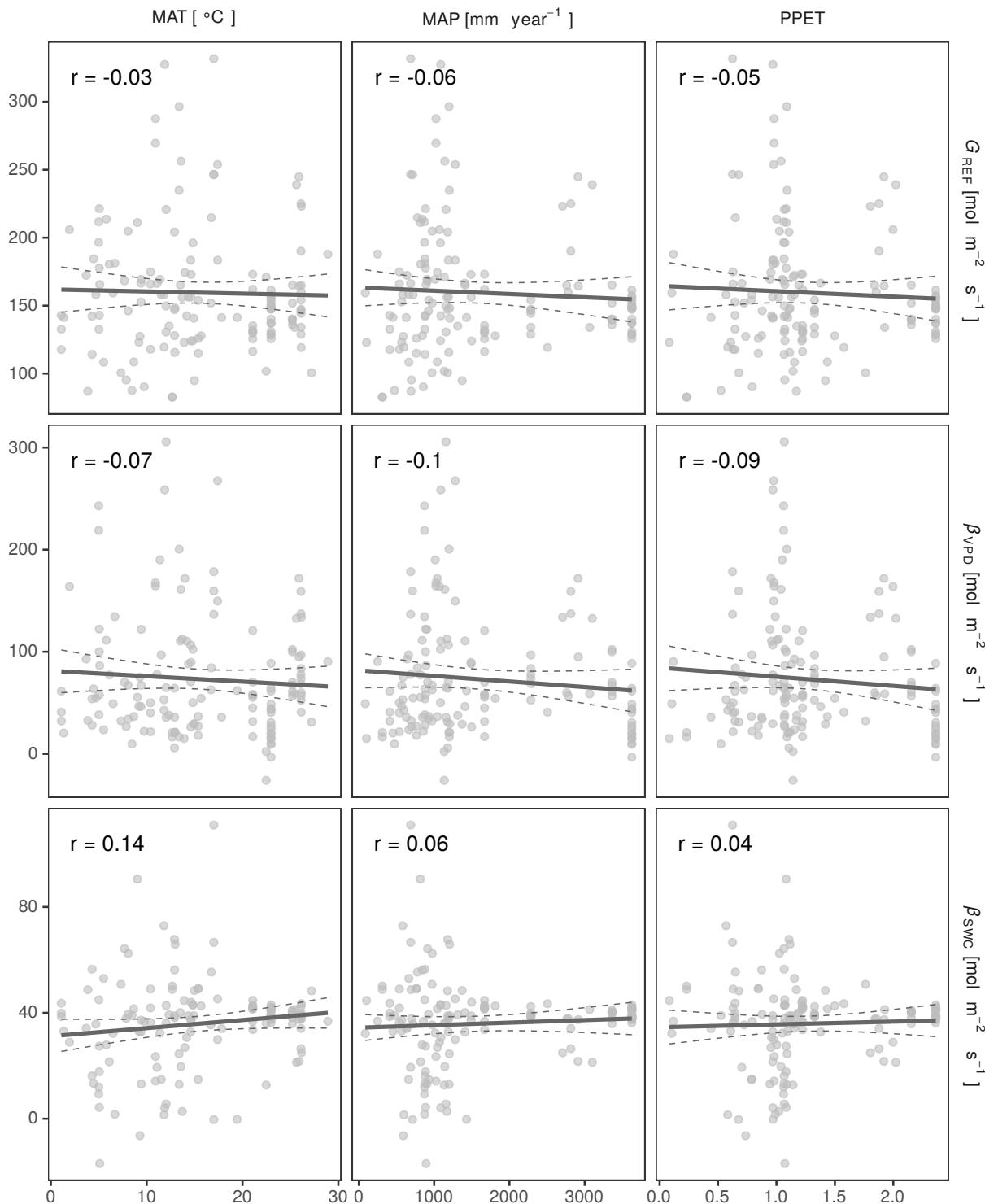


Figure D.7: Species' climatic variables and water use parameters relationships. MAP: mean annual precipitation, MAT: mean annual temperature, PPET: mean annual precipitation over potential evapotranspiration. All climatic variables were calculated as the weighted average of the characteristic plots of the trees of each species, using tree-days as weighting factor.

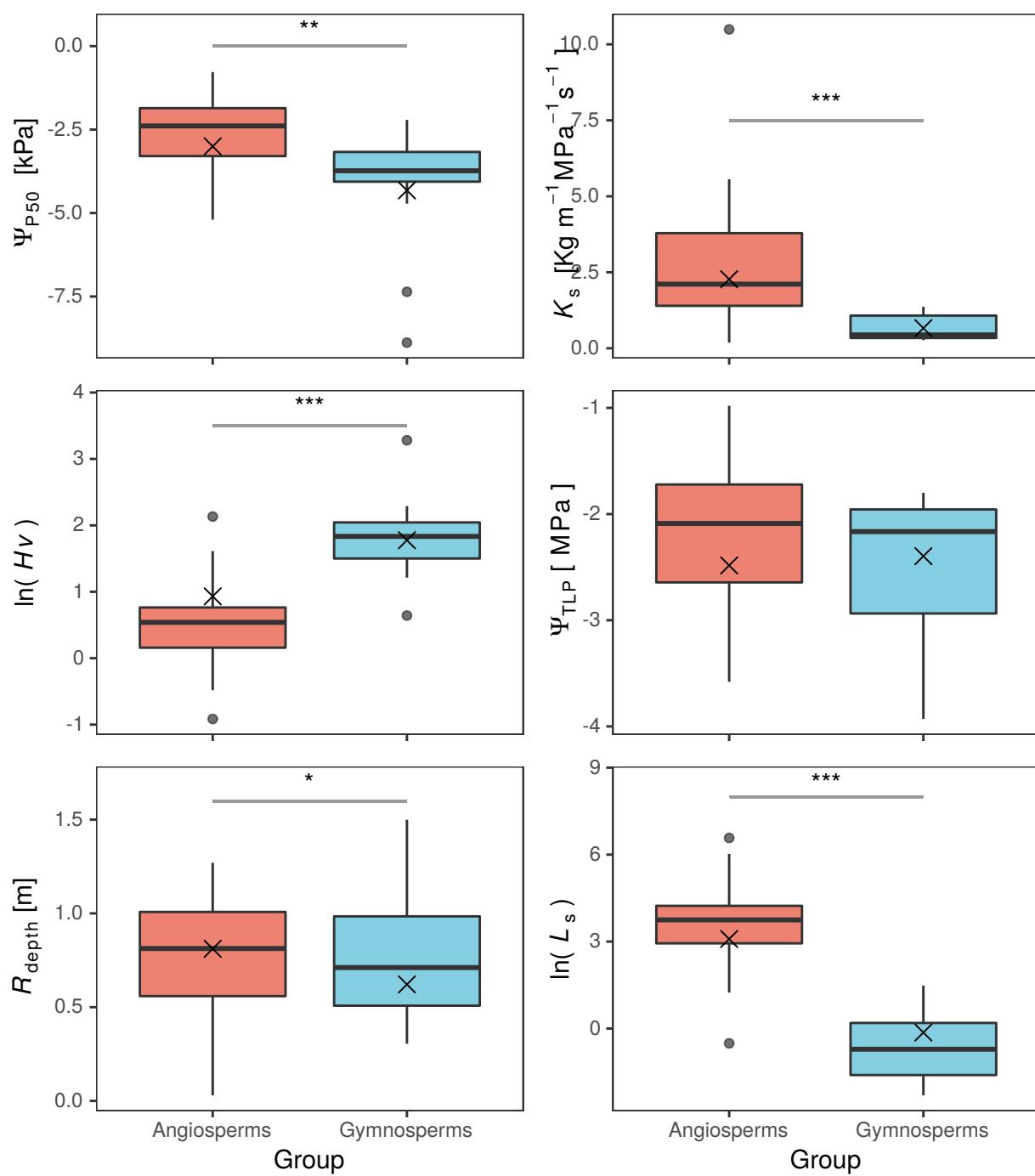


Figure D.8: Boxplots of water relations traits for angiosperms and gymnosperms. Statistical significance level is showed as symbols: ., $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$. Crosses are weighted means of each trait by groups

D.3 Tables D

Table D.1: SAPFLUXNET plot treatments included in this study (Chapter 3).

| Plot treatment |
|---|
| None |
| Control |
| control |
| Ambient Control |
| Control - Unthinned |
| natural conditions |
| Reference |
| 1Premortality |
| 2premortality |
| distructive sampling |
| Girdling early successional |
| Pre-thinning |
| Before thinning |
| Before Thinning |
| non thinned |
| none (periodict thinning every 5-6 years 20 to 25% of basal area) |
| Radiation Level |
| AMBIENT CO2 FACE rings |
| fertilization at plantation |
| AcaciaMonoculture |
| MixtureEucalyptusAndAcacia |
| EucalyptusMonoculture |
| Pre Irrigation |

Table D.2: Species resume table. CEP names are Cornell Ecology Programs species names. Ψ_{P50} : water potential at 50% water conductivity loss [MPa], K_s : maximum sapwood water conductivity [$\text{Kg m}^{-1} \text{ MPa}^{-1} \text{ s}^{-1}$], H_v : Huber value [$\text{cm}_\text{Asw}^2 \text{ m}_\text{leaf area}^{-2}$], Ψ_{TLP} : water potential at turgor-loss point [MPa], R_depth : rooting depth [m], L_s : individual leaf area [cm^2].

| Species | CEP names | Group | # tree-days | # trees | # plots | Ψ_{P50} | K_s | Ψ_{TLP} | H_v | R_depth | L_s |
|-------------------------------------|-----------|-------------|-------------|---------|---------|--------------|--------|--------------|--------|------------------|---------|
| <i>Abies balsamea</i> | Abiebals | Gymnosperms | 855 | 19 | 1 | -2.479 | 1.292 | | 26.600 | 0.508 | |
| <i>Abies pinsapo</i> | Abiepins | Gymnosperms | 11822 | 22 | 3 | -4.150 | | | | | |
| <i>Acacia longifolia</i> | Acaclong | Angiosperms | 2130 | 10 | 1 | | | | | | 11.715 |
| <i>Acacia mearnsii</i> | Acacmear | Angiosperms | 4555 | 23 | 2 | | | | | | 4.134 |
| <i>Acacia tortilis</i> | Acactort | Angiosperms | 1740 | 3 | 1 | | | | | | |
| <i>Acer rubrum</i> | Acerrubr | Angiosperms | 17230 | 49 | 9 | -2.755 | 4.108 | -1.520 | 1.422 | 0.762 | 44.983 |
| <i>Acer saccharum</i> | Acersacc | Angiosperms | 10928 | 156 | 8 | -2.873 | 4.695 | -1.600 | 1.134 | 1.016 | 55.675 |
| <i>Agathis australis</i> | Agataust | Gymnosperms | 1272 | 6 | 1 | -2.210 | 1.250 | | | | |
| <i>Ampelocera macrocarpa</i> | Ampemacr | Angiosperms | 100 | 2 | 1 | | | | | | 103.560 |
| <i>Arbutus unedo</i> | Arbuuned | Angiosperms | 1900 | 4 | 1 | -4.198 | 0.714 | -0.980 | 5.017 | | 12.470 |
| <i>Aspidosperma desmanthum</i> | Aspidesdm | Angiosperms | 35 | 1 | 1 | | | | | | 42.925 |
| <i>Avicennia marina</i> | Avicmari | Angiosperms | 828 | 6 | 1 | | | | 8.435 | | 20.800 |
| <i>Betula alleghaniensis</i> | Betulle | Angiosperms | 3750 | 16 | 2 | | | | | | 29.938 |
| <i>Betula papyrifera</i> | Betupapy | Angiosperms | 3869 | 21 | 2 | -1.966 | 1.538 | -1.330 | 2.096 | 0.610 | 23.942 |
| <i>Betula pendula</i> | Betupend | Angiosperms | 9 | 1 | 1 | -2.265 | | | | 0.610 | 11.177 |
| <i>Betula pubescens</i> | Betupube | Angiosperms | 397 | 9 | 2 | | | | | | 13.510 |
| <i>Betula sp.</i> | Betuspb | Angiosperms | 204 | 2 | 1 | | | | | | |
| <i>Brosimum alicastrum</i> | Brosalic | Angiosperms | 40 | 1 | 1 | | | | 1.685 | | 58.953 |
| <i>Carapa guianensis</i> | Caraguia | Angiosperms | 150 | 3 | 1 | -0.800 | 0.270 | | 0.793 | | |
| <i>Carpinus betulus</i> | Carpbetu | Angiosperms | 530 | 5 | 1 | -3.750 | | | | | 23.160 |
| <i>Carya glabra</i> | Caryglab | Angiosperms | 47 | 1 | 1 | -2.100 | | | | 1.270 | 324.425 |
| <i>Carya tomentosa</i> | Carytome | Angiosperms | 645 | 10 | 2 | | | | | | |
| <i>Castanopsis acuminatissima</i> | Castacum | Angiosperms | 96 | 8 | 1 | | 10.490 | | 1.570 | | 45.000 |
| <i>Celtis laevigata</i> | Celtlaev | Angiosperms | 745 | 13 | 2 | | | | | | 13.580 |
| <i>Coprosma quadrifida</i> | Coprquad | Angiosperms | 57 | 4 | 2 | | | | | | 0.600 |
| <i>Cornus florida</i> | Cornflor | Angiosperms | 126 | 1 | 1 | -4.442 | 0.768 | | 2.034 | 0.457 | 46.343 |
| <i>Cryptocarya laevigata</i> | Cryplaev | Angiosperms | 72 | 6 | 1 | | 2.140 | | 1.750 | | 19.075 |
| <i>Cupania macrophylla</i> | Cupamacr | Angiosperms | 40 | 1 | 1 | | | | | | |
| <i>Dicorynia guianensis</i> | Dicoguia | Angiosperms | 596 | 9 | 2 | | | -1.712 | | | 717.500 |
| <i>Drimys brasiliensis</i> | Drimbras | Angiosperms | 90 | 5 | 1 | | | | | | 18.420 |
| <i>Elaeagnus angustifolia</i> | Elaeangu | Angiosperms | 402 | 2 | 1 | | | | | | 11.200 |
| <i>Eschweilera coriacea</i> | Eschcori | Angiosperms | 249 | 4 | 2 | | | -1.828 | 1.312 | | 58.654 |
| <i>Eschweilera grandiflora</i> | Eschgranc | Angiosperms | 378 | 2 | 1 | | | -1.754 | 0.746 | | 62.085 |
| <i>Eschweilera sp.</i> | Eschsp | Angiosperms | 40 | 1 | 1 | | | | | | |
| <i>Eucalyptus baxteri</i> | Eucabaxt | Angiosperms | 112 | 4 | 2 | | | | | | |
| <i>Eucalyptus camaldulensis</i> | Eucacama | Angiosperms | 564 | 3 | 1 | -4.025 | 3.600 | -2.010 | 2.364 | 0.508 | 10.910 |
| <i>Eucalyptus cypellocarpa</i> | Eucacype | Angiosperms | 29 | 2 | 2 | | | | | | |
| <i>Eucalyptus globulus</i> | Eucaglob | Angiosperms | 4555 | 23 | 2 | -1.100 | 3.950 | -1.640 | 3.155 | 0.610 | |
| <i>Eucalyptus nitens</i> | Eucanite | Angiosperms | 378 | 7 | 1 | | | | | 2.163 | |
| <i>Eucalyptus obliqua</i> | Eucaboli | Angiosperms | 2466 | 6 | 1 | | | -1.340 | | | 16.790 |
| <i>Eucalyptus radiata</i> | Eucaradi | Angiosperms | 170 | 2 | 1 | | | | | | |
| <i>Eucalyptus rubida</i> | Eucarubi | Angiosperms | 2055 | 5 | 1 | | 5.560 | -1.130 | 1.222 | | |
| <i>Fagus grandifolia</i> | Fagugran | Angiosperms | 6815 | 32 | 3 | -5.080 | | | | 0.813 | 45.167 |
| <i>Fagus sylvatica</i> | Fagusylv | Angiosperms | 15075 | 93 | 12 | -2.972 | 1.830 | -2.107 | 3.934 | | 24.030 |
| <i>Fraxinus americana</i> | Fraxamer | Angiosperms | 378 | 2 | 1 | -1.920 | | | | 1.016 | 339.552 |
| <i>Fraxinus excelsior</i> | Fraxexce | Angiosperms | 636 | 6 | 1 | -2.805 | | -1.750 | | 0.040 | 133.800 |
| <i>Fraxinus pennsylvanica</i> | Fraxpenn | Angiosperms | 178 | 2 | 1 | -0.777 | 1.397 | -2.360 | | 1.016 | |
| <i>Genipa americana</i> | Geniamer | Angiosperms | 40 | 1 | 1 | -1.270 | 1.500 | -2.550 | | | |
| <i>Gouania glabra</i> | Gouplab | Angiosperms | 172 | 3 | 3 | | | | | | 31.288 |
| <i>Gymnanthes riparia</i> | Gymnripa | Angiosperms | 150 | 3 | 1 | | | | | | |
| <i>Hevea brasiliensis</i> | Hevebras | Angiosperms | 1302 | 6 | 1 | -1.859 | 2.582 | -2.650 | 0.618 | | |
| <i>Ilex aquifolium</i> | Ilexaqui | Angiosperms | 567 | 3 | 1 | -4.241 | 0.186 | -2.190 | 4.825 | 0.030 | 16.770 |
| <i>Inga sp.</i> | Ingasp | Angiosperms | 40 | 1 | 1 | | | | | | |
| <i>Iryanthera sagotiana</i> | Iryasago | Angiosperms | 72 | 1 | 1 | | | -1.830 | | | |
| <i>Juniperus monosperma</i> | Junimono | Gymnosperms | 23650 | 15 | 3 | -8.882 | 0.710 | -3.930 | 5.700 | 0.305 | 0.400 |
| <i>Kandelia obovata</i> | Kandobov | Angiosperms | 2584 | 8 | 1 | | | | | | |
| <i>Larix decidua</i> | Larideci | Gymnosperms | 3127 | 23 | 5 | -3.791 | 0.434 | -2.835 | | 1.016 | 0.170 |
| <i>Larix gmelinii</i> | Larigmel | Gymnosperms | 303 | 3 | 1 | | | | | | |
| <i>Larix sibirica</i> | Larisibi | Angiosperms | 516 | 3 | 1 | | | | | | |
| <i>Lecythis poiteau</i> | Lecypoit | Angiosperms | 90 | 2 | 1 | | | -2.581 | | | |
| <i>Licania alba</i> | Licaalba | Angiosperms | 141 | 3 | 1 | | | -2.044 | 0.664 | | 157.435 |
| <i>Licania membranacea</i> | Licamemb | Angiosperms | 217 | 2 | 2 | | | -2.239 | 0.401 | | 38.313 |
| <i>Licania octandra</i> | Licaocta | Angiosperms | 256 | 2 | 1 | | | | 1.088 | | 32.005 |
| <i>Licania rodriquesii</i> | Licarodr | Angiosperms | 34 | 1 | 1 | | | | | 0.914 | 47.104 |
| <i>Liquidambar styraciflua</i> | Liquistyr | Angiosperms | 742 | 23 | 3 | -2.622 | 1.042 | | 1.949 | | |
| <i>Liriodendron tulipifera</i> | Lirituli | Angiosperms | 2246 | 27 | 7 | -2.390 | 2.610 | -1.130 | | 0.813 | 116.933 |
| <i>Macrolobium costaricense</i> | Macrcost | Angiosperms | 40 | 1 | 1 | | | | | | 79.840 |
| <i>Malus domestica</i> | Maludome | Angiosperms | 1175 | 5 | 2 | | | -2.070 | | | 16.200 |
| <i>Manilkara bidentata</i> | Manibide | Angiosperms | 500 | 2 | 1 | -2.700 | 4.333 | | | 1.309 | 0.813 |
| <i>Meliocoma idiopoda</i> | Meliidio | Angiosperms | 100 | 2 | 1 | | | | | | 45.229 |
| <i>Mortoniodendron anisophyllum</i> | Mortanis | Angiosperms | 100 | 2 | 1 | | | | | | |
| <i>Myrtaceae sp.</i> | Myrtsp | Angiosperms | 72 | 6 | 1 | | | | | | |
| <i>Ostrya virginiana</i> | Ostrvirg | Angiosperms | 549 | 3 | 2 | | | | | | 23.994 |

Table D.2: Species resume table. CEP names are Cornell Ecology Programs species names. Ψ_{P50} : water potential at 50% water conductivity loss [MPa], K_s : maximum sapwood water conductivity [$\text{Kg m}^{-1} \text{ MPa}^{-1} \text{ s}^{-1}$], H_v : Huber value [$\text{cm}^2 \text{ Asw m}_{\text{leaf area}}^{-2}$], Ψ_{TLP} : water potential at turgor-loss point [MPa], R_{depth} : rooting depth [m], L_s : individual leaf area [cm^2]. (*continued*)

| Species | CEP names | Group | # tree-days | # trees | # plots | Ψ_{P50} | K_s | Ψ_{TLP} | H_v | R_{depth} | V^s |
|--------------------------------|-----------|-------------|-------------|---------|---------|--------------|-------|--------------|-------|--------------------|---------|
| <i>Otoba novogranatensis</i> | Otobnovo | Angiosperms | 100 | 2 | 1 | | | | | | 196.140 |
| <i>Ozandria asbeckii</i> | Oxanasbe | Angiosperms | 263 | 3 | 2 | | | | | | |
| <i>Palaquium luzonicense</i> | Palaluzo | Angiosperms | 72 | 6 | 1 | | | | | | |
| <i>Picea abies</i> | Piceabie | Gymnosperms | 11873 | 109 | 11 | -3.714 | 0.558 | -1.950 | 7.375 | 0.711 | 0.150 |
| <i>Picea stichensis</i> | Picesitc | Gymnosperms | 990 | 15 | 1 | -3.850 | | | | 1.500 | 0.180 |
| <i>Pinus canariensis</i> | Pinucana | Gymnosperms | 4890 | 10 | 1 | | | | | | |
| <i>Pinus cembra</i> | Pinucemb | Gymnosperms | 1151 | 14 | 5 | -3.192 | 0.267 | -2.340 | | | |
| <i>Pinus edulis</i> | Pinuedul | Gymnosperms | 23715 | 15 | 3 | -4.718 | | -1.990 | | 0.508 | 1.250 |
| <i>Pinus halepensis</i> | Pinuhale | Gymnosperms | 1329 | 3 | 1 | -4.105 | 0.335 | -3.000 | 7.590 | | |
| <i>Pinus nigra</i> | Pinunigr | Gymnosperms | 11246 | 15 | 3 | -3.753 | 0.407 | -1.800 | 7.860 | 1.016 | 1.110 |
| <i>Pinus patula</i> | Pinupatu | Gymnosperms | 152 | 8 | 1 | | | | | | |
| <i>Pinus pinaster</i> | Pinupina | Gymnosperms | 2130 | 10 | 1 | -3.489 | 0.352 | | 9.870 | | |
| <i>Pinus resinosa</i> | Pinuresi | Gymnosperms | 1176 | 42 | 1 | | | -1.940 | | 1.016 | 4.400 |
| <i>Pinus rigida</i> | Pinurigi | Gymnosperms | 489 | 2 | 2 | | | | | | 3.150 |
| <i>Pinus sibirica</i> | Pinusbici | Gymnosperms | 516 | 3 | 1 | | | | | | |
| <i>Pinus strobus</i> | Pinustro | Gymnosperms | 32538 | 60 | 4 | | | | | | 2.700 |
| <i>Pinus sylvestris</i> | Pinusylv | Gymnosperms | 42463 | 215 | 21 | -3.163 | 0.448 | -1.975 | 6.257 | 0.508 | 0.947 |
| <i>Pinus taeda</i> | Pinutaed | Gymnosperms | 24380 | 97 | 4 | -2.810 | 1.197 | | 4.848 | 0.889 | |
| <i>Plates excelsa</i> | Platexce | Angiosperms | 72 | 6 | 1 | | | | | | 53.633 |
| <i>Pleuranthodendron</i> | Pleulind | Angiosperms | 40 | 1 | 1 | | | | | | |
| <i>lindenii</i> | | | | | | | | | | | |
| <i>Populus canescens</i> | Popucane | Angiosperms | 597 | 3 | 1 | | | | | | |
| <i>Populus euphratica</i> | Popueuph | Angiosperms | 841 | 9 | 3 | -2.264 | 2.998 | -2.900 | 2.420 | | |
| <i>Populus grandidentata</i> | Popugran | Angiosperms | 6888 | 12 | 1 | | | | | | 42.608 |
| <i>Populus tremuloides</i> | Poputrem | Angiosperms | 4455 | 99 | 1 | -1.876 | 0.934 | | 1.910 | 1.000 | 24.690 |
| <i>Pouteria anomala</i> | Poutanom | Angiosperms | 912 | 3 | 1 | | | | 1.392 | | 42.295 |
| <i>Pouteria firma</i> | Poutfirm | Angiosperms | 72 | 6 | 1 | | | | | | |
| <i>Pouteria sp.</i> | Poutsp | Angiosperms | 40 | 1 | 1 | | | | | | |
| <i>Pouteria viridis</i> | Poutviri | Angiosperms | 100 | 2 | 1 | | | | | | |
| <i>Protium tenuifolium</i> | Prottenu | Angiosperms | 242 | 2 | 1 | | | | 1.587 | | 413.810 |
| <i>Prunus serotina</i> | Prunsero | Angiosperms | 100 | 1 | 1 | -4.270 | 0.550 | -1.420 | | 0.914 | 24.759 |
| <i>Pseudotsuga menziesii</i> | Pseumenz | Gymnosperms | 2985 | 18 | 3 | -3.926 | 1.366 | -2.970 | 4.153 | 0.660 | 0.280 |
| <i>Qualea rosea</i> | Qualrose | Angiosperms | 141 | 3 | 1 | | | -1.779 | | | |
| <i>Qualea tricolor</i> | Qualtric | Angiosperms | 78 | 2 | 1 | | | | | | |
| <i>Quercus alba</i> | Queralba | Angiosperms | 1094 | 9 | 5 | -1.818 | 4.877 | -2.050 | 1.454 | 1.219 | 54.302 |
| <i>Quercus cerris</i> | Quercerr | Angiosperms | 284 | 2 | 1 | | | -3.580 | | | 29.190 |
| <i>Quercus coccinea</i> | Quercocc | Angiosperms | 5343 | 11 | 3 | | | | | | 121.678 |
| <i>Quercus faginea</i> | Querfagi | Angiosperms | 6784 | 10 | 2 | -2.002 | | -3.160 | | | 4.210 |
| <i>Quercus falcata</i> | Querfalc | Angiosperms | 134 | 1 | 1 | -0.893 | 2.323 | | | 2.057 | 41.635 |
| <i>Quercus ilex</i> | Querilex | Angiosperms | 65295 | 62 | 6 | -3.438 | 1.595 | -3.015 | 3.980 | | 8.078 |
| <i>Quercus lyrata</i> | Querlyra | Angiosperms | 745 | 13 | 2 | | | | | | |
| <i>Quercus michauxii</i> | Quermich | Angiosperms | 132 | 4 | 1 | -1.700 | | | | | 71.498 |
| <i>Quercus montana</i> | Quermont | Angiosperms | 5488 | 14 | 2 | | | | | | |
| <i>Quercus petraea</i> | Querpetr | Angiosperms | 4617 | 29 | 5 | -3.294 | | -2.624 | | 0.300 | 35.000 |
| <i>Quercus phellos</i> | Querphel | Angiosperms | 185 | 5 | 1 | -1.375 | 3.787 | | 1.806 | | 8.935 |
| <i>Quercus prinus</i> | Querprin | Angiosperms | 316 | 2 | 1 | -1.700 | | | | | |
| <i>Quercus pubescens</i> | Querpube | Angiosperms | 6505 | 19 | 2 | -2.696 | 1.612 | -3.515 | 0.782 | | 32.200 |
| <i>Quercus pyrenaica</i> | Querpyre | Angiosperms | 6680 | 32 | 2 | | | -2.680 | | | 30.260 |
| <i>Quercus robur</i> | Querrobt | Angiosperms | 636 | 6 | 1 | -2.802 | | -2.583 | | 0.300 | 46.537 |
| <i>Quercus rotundifolia</i> | Querrotu | Angiosperms | 4796 | 10 | 2 | | | | | | 3.460 |
| <i>Quercus rubra</i> | Querrubr | Angiosperms | 15185 | 32 | 6 | -2.237 | 1.602 | -2.725 | 1.746 | 0.914 | 66.641 |
| <i>Quercus suber</i> | Quersube | Angiosperms | 1816 | 4 | 1 | -5.200 | | -3.080 | | | 7.030 |
| <i>Quercus velutina</i> | Quervelu | Angiosperms | 5132 | 16 | 3 | | | -2.750 | | 1.016 | 110.450 |
| <i>Recordroxylon speciosum</i> | Recospec | Angiosperms | 41 | 1 | 1 | | | | | | |
| <i>Santiria apiculata</i> | Santapic | Angiosperms | 55 | 5 | 1 | | | | | | |
| <i>Sassafras albidum</i> | Sassalbi | Angiosperms | 117 | 1 | 1 | | | | | | 68.080 |
| <i>Sextonia rubra</i> | Sextrubr | Angiosperms | 34 | 1 | 1 | | | | | | 74.000 |
| <i>Sloanea sp</i> | Sloasp | Angiosperms | 156 | 2 | 2 | | | | | | |
| <i>Swartzia racemosa</i> | Swarrace | Angiosperms | 476 | 2 | 1 | | | 1.156 | | | 246.450 |
| <i>Taxus baccata</i> | Taxubacc | Gymnosperms | 922 | 2 | 1 | -7.360 | 0.320 | | 1.900 | 0.406 | 0.500 |
| <i>Thuja occidentalis</i> | Thujocci | Gymnosperms | 340 | 10 | 1 | -3.570 | | | | 0.762 | 0.100 |
| <i>Tilia americana</i> | Tiliamer | Angiosperms | 72 | 1 | 1 | | | | | | 130.715 |
| <i>Trophis mexicana</i> | Tropmexi | Angiosperms | 40 | 1 | 1 | | | | | | 13.880 |
| <i>Tsuga canadensis</i> | Tsugcana | Gymnosperms | 5739 | 24 | 2 | -3.070 | 0.329 | | 3.362 | 0.711 | 0.480 |
| <i>Ulmus americana</i> | Ulmuamer | Angiosperms | 133 | 1 | 1 | | | | | | 56.993 |
| <i>Vantanea sp</i> | Vantsp | Angiosperms | 69 | 1 | 1 | | | | | | |
| <i>Vernonia arborea</i> | Vernarbo | Angiosperms | 36 | 3 | 1 | | | 2.110 | | 2.060 | |
| <i>Vouacapoua americana</i> | Vouaamer | Angiosperms | 178 | 3 | 3 | | | -2.147 | 0.744 | | 353.583 |

Table D.3: Results of the linear models relating water use parameters calculated using G'_{Asw} to water relations traits. Parameters are explained by individual traits using simple linear models with number of species-days as weighting factor.

| Parameter | Trait | N Species | Intercept | Slope | R^2 |
|-----------------------|-----------------------|-----------|-------------|-------------|-------|
| G'_{REF} | $\ln(\Psi_{P50})$ | 25 | 333.019 *** | -111.725 ** | 0.331 |
| | $\ln(K_s)$ | 19 | 196.081 *** | 81.935 ** | 0.359 |
| | $\ln(Hv)$ | 20 | 297.029 *** | -71.225 * | 0.188 |
| | $ \Psi_{\text{TLP}} $ | 25 | 196.754 ** | -4.398 | 0.000 |
| | R_{depth} | 13 | -13.266 | 303.191 ** | 0.508 |
| | $\ln(L_s)$ | 26 | 151.531 *** | 31.494 *** | 0.497 |
| | $\ln(\Psi_{P50})$ | 25 | 235.774 *** | -95.494 * | 0.215 |
| | $\ln(K_s)$ | 19 | 125.553 *** | 95.013 ** | 0.448 |
| | $\ln(Hv)$ | 20 | 221.274 *** | -69.362 * | 0.153 |
| β'_{VPD} | $ \Psi_{\text{TLP}} $ | 25 | 62.062 | 17.932 | 0.000 |
| | R_{depth} | 13 | -74.639 * | 245.287 *** | 0.681 |
| | $\ln(L_s)$ | 26 | 69.367 *** | 31.574 *** | 0.409 |
| | $\ln(\Psi_{P50})$ | 25 | 34.052 . | 21.703 | 0.066 |
| β'_{SWC} | $\ln(K_s)$ | 19 | 57.145 *** | -19.246 . | 0.111 |
| | $\ln(Hv)$ | 20 | 39.018 . | 13.555 | 0.008 |
| | $ \Psi_{\text{TLP}} $ | 25 | 86.217 *** | -9.086 | 0.011 |
| | R_{depth} | 13 | 94.44 *** | -29.023 | 0.038 |
| | $\ln(L_s)$ | 26 | 73.395 *** | -8.696 ** | 0.234 |

Statistical significant levels: ., $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

Table D.4: Results of the linear models relating water use parameters to water relations traits using REW instead of SWC. Parameters are explained by individual traits using simple linear models with number of species-days as weighting factor.

| Parameter | Trait | N Species | Intercept | Slope | R^2 |
|----------------------|-----------------------|-----------|-------------|-------------|-------|
| G_{REF} | $\ln(\Psi_{P50})$ | 55 | 198.741 *** | -70.836 *** | 0.328 |
| | $\ln(K_s)$ | 43 | 112.403 *** | 37.208 *** | 0.375 |
| | $\ln(Hv)$ | 49 | 177.173 *** | -45.734 *** | 0.389 |
| | $ \Psi_{\text{TLP}} $ | 48 | 171.577 *** | -24.124 ** | 0.118 |
| | R_{depth} | 37 | 28.834 | 122.446 *** | 0.373 |
| | $\ln(L_s)$ | 86 | 86.92 *** | 17.171 *** | 0.488 |
| β_{VPD} | $\ln(\Psi_{P50})$ | 55 | 109.036 *** | -48.046 *** | 0.240 |
| | $\ln(K_s)$ | 43 | 49.473 *** | 20.603 ** | 0.181 |
| | $\ln(Hv)$ | 49 | 96.583 *** | -34.355 *** | 0.369 |
| | $ \Psi_{\text{TLP}} $ | 48 | 79.314 *** | -12.005 | 0.037 |
| | R_{depth} | 37 | -1.771 | 79.374 *** | 0.252 |
| | $\ln(L_s)$ | 86 | 33.569 *** | 11.392 *** | 0.336 |
| β_{REW} | $\ln(\Psi_{P50})$ | 55 | 13.956 ** | -3 | 0.000 |
| | $\ln(K_s)$ | 43 | 11.426 *** | 1.256 | 0.000 |
| | $\ln(Hv)$ | 49 | 11.166 *** | 0.339 | 0.000 |
| | $ \Psi_{\text{TLP}} $ | 48 | 18.869 *** | -3.274 . | 0.054 |
| | R_{depth} | 37 | 5.99 | 5.114 | 0.000 |
| | $\ln(L_s)$ | 86 | 15.63 *** | -0.373 | 0.000 |

Statistical significant levels: ., $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

Table D.5: Results of the linear models relating water use parameters calculated using G'_{Asw} , water relations traits, climate and tree height. Water use parameters are explained by individual traits, MAP (mean annual precipitation) and H (tree height) using simple linear models using number of species-days as weighting factor. β values are the slopes for each explanatory variable. NI = not included variable after model selection.

| Parameter | Trait | N Species | Intercept | β_{trait} | β_{MAP} | β_H | R^2 |
|-----------------------|-----------------------|-----------|-------------|------------------------|----------------------|-----------|-------|
| G'_{REF} | $\ln(\Psi_{P50})$ | 25 | 328.177 *** | -106.498 ** | NI | NI | 0.262 |
| | $\ln(K_s)$ | 19 | 199.257 *** | 74.161 ** | NI | NI | 0.317 |
| | $\ln(Hv)$ | 20 | 296.21 *** | -68.395 * | NI | NI | 0.177 |
| | $ \Psi_{\text{TLP}} $ | 25 | 72.695 | NI | 0.175 ** | NI | 0.240 |
| | R_{depth} | 13 | 0.75 | 280.314 ** | NI | NI | 0.468 |
| | $\ln(L_s)$ | 26 | 158.682 *** | 28.552 *** | NI | NI | 0.435 |
| β'_{VPD} | $\ln(\Psi_{P50})$ | 25 | -10.852 | NI | 0.274 *** | -5.198 ** | 0.476 |
| | $\ln(K_s)$ | 19 | 71.809 | 60.751 * | 0.153 . | -4.342 . | 0.539 |
| | $\ln(Hv)$ | 20 | 149.585 . | -54.146 . | 0.177 * | -6.093 * | 0.432 |
| | $ \Psi_{\text{TLP}} $ | 25 | -17.511 | NI | 0.29 *** | -5.639 ** | 0.522 |
| | R_{depth} | 13 | -63.756 . | 226.165 *** | NI | NI | 0.630 |
| | $\ln(L_s)$ | 26 | 23.708 | 23.07 ** | 0.182 ** | -5.54 *** | 0.615 |
| β'_{SWC} | $\ln(\Psi_{P50})$ | 25 | 95.16 *** | NI | -0.084 ** | 1.953 ** | 0.366 |
| | $\ln(K_s)$ | 19 | 84.858 *** | NI | -0.072 * | 1.91 * | 0.261 |
| | $\ln(Hv)$ | 20 | 78.013 ** | NI | -0.06 * | 1.809 . | 0.191 |
| | $ \Psi_{\text{TLP}} $ | 25 | 98.488 *** | NI | -0.089 *** | 1.91 * | 0.388 |
| | R_{depth} | 13 | 92.124 *** | NI | NI | -1.025 . | 0.173 |
| | $\ln(L_s)$ | 26 | 90.501 *** | -5.107 | -0.066 * | 2.011 ** | 0.402 |

Statistical significant levels: ., $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

References

- Ackerly D. **2004**. Functional strategies of chaparral shrubs in relation to seasonal water deficit and disturbance. *Ecological Monographs* **74**: 25–44.
- Aemisegger F, Pfahl S, Sodemann H, Lehner I, Seneviratne SI, Wernli H. **2014**. Deuterium excess as a proxy for continental moisture recycling and plant transpiration. *Atmospheric Chemistry and Physics* **14**: 4029–4054.
- Alarcón JJ, Ortuño MF, Nicolás E, Torres R, Torrecillas A. **2005**. Compensation heat-pulse measurements of sap flow for estimating transpiration in young lemon trees. *Biologia Plantarum* **49**: 527–532.
- Allen CD, Breshears DD, McDowell NG. **2015**. On underestimation of global vulnerability to tree mortality and forest die-off from hotter drought in the anthropocene. *Ecosphere* **6**: art129.
- Allen ST, Kirchner JW, Braun S, Siegwolf RTW, Goldsmith GR. **2019**. Seasonal origins of soil water used by trees. *Hydrology and Earth System Sciences* **23**: 1199–1210.
- Ambrose AR, Sillett SC, Koch GW, Van Pelt R, Antoine ME, Dawson TE. **2010**. Effects of height on treetop transpiration and stomatal conductance in coast redwood (*Sequoia sempervirens*). *Tree Physiology* **30**: 1260–1272.
- Anderegg WRL. **2015**. Spatial and temporal variation in plant hydraulic traits and their relevance for climate change impacts on vegetation. *New Phytologist* **205**: 1008–1014.
- Anderegg WRL, Venturas MD. **2020**. Plant hydraulics play a critical role in earth system fluxes. *New Phytologist* **226**: 1535–1538.
- Anderegg WRL, Klein T, Bartlett M, Sack L, Pellegrini AFA, Choat B, Jansen S. **2016**. Meta-analysis reveals that hydraulic traits explain cross-species patterns of drought-induced tree mortality across the globe. *Proceedings of the National Academy of Sciences* **113**: 5024–5029.
- Anderegg WRL, Konings AG, Trugman AT, Yu K, Bowling DR, Gabbitas R, Karp DS, Pacala S, Sperry JS, Sulman BN *et al.* **2018**. Hydraulic diversity of forests regulates ecosystem resilience during drought. *Nature* **561**: 538–541.
- Anderegg WRL, Trugman AT, Badgley G, Anderson CM, Bartuska A, Ciais P, Cullenward D, Field CB, Freeman J, Goetz SJ *et al.* **2020**. Climate-driven risks to the climate mitigation potential of forests. *Science* **368**.
- Anderegg WRL, Trugman AT, Bowling DR, Salvucci G, Tuttle SE. **2019**. Plant functional traits and climate influence drought intensification and land–atmosphere feedbacks. *Proceedings of*

- the National Academy of Sciences **116**: 14071–14076.
- Anfodillo T, Sigalotti GB, Tomasi M, Semenzato P, Valentini R. **1993**. Applications of a thermal imaging technique in the study of the ascent of sap in woody species. *Plant, Cell & Environment* **16**: 997–1001.
- Aparecido LMT, Woo S, Suazo C, Hultine KR, Blonder B. **2020**. High water use in desert plants exposed to extreme heat (J Penuelas, Ed.). *Ecology Letters* **23**: 1189–1200.
- Asbjornsen H, Goldsmith GR, Alvarado-Barrientos MS, Rebel K, Van Osch FP, Rietkerk M, Chen J, Gotsch S, Tobón C, Geissert DR *et al.* **2011**. Ecohydrological advances and applications in plant–water relations research: a review. *Journal of Plant Ecology* **4**: 3–22.
- Aspinwall MJ, King JS, Domec J-C, McKeand SE, Isik F. **2011**. Genetic effects on transpiration, canopy conductance, stomatal sensitivity to vapour pressure deficit, and cavitation resistance in loblolly pine. *Ecohydrology* **4**: 168–182.
- Babst F, Bouriaud O, Poulter B, Trouet V, Girardin MP, Frank DC. **2019**. Twentieth century redistribution in climatic drivers of global tree growth. *Science Advances* **5**: eaat4313.
- Bacelar ELVA, Moutinho-Pereira JM, Gonçalves BMC, Brito CVQ, Gomes-Laranjo J, Ferreira HMF, Correia CM. **2012**. Water Use Strategies of Plants Under Drought Conditions. In: Aroca R, ed. *Plant Responses to Drought Stress*. Berlin, Heidelberg: Springer Berlin Heidelberg, 145–170.
- Baker JM, Van Bavel CHM. **1987**. Measurement of mass flow of water in the stems of herbaceous plants. *Plant, Cell & Environment* **10**: 777–782.
- Ball JT, Woodrow IE, Berry JA. **1987**. A model predicting stomatal conductance and its contribution to the control of photosynthesis under different environmental conditions. In: Biggins J, ed. *Progress in photosynthesis research: Volume 4 proceedings of the viith international congress on photosynthesis providence, rhode island, usa, august 10–15, 1986*. Dordrecht: Springer Netherlands, 221–224.
- Ballester C, Castel JR, Sanz F, Yeves A, Intrigliolo DS, Castel JR. **2011**. Can sap flow probes be used for determining transpiration of citrus trees under different irrigation regimes? *Acta Horticulturae* **922**: 221–228.
- Barbeta A, Peñuelas J. **2017**. Relative contribution of groundwater to plant transpiration estimated with stable isotopes. *Scientific Reports* **7**: 10580.
- Barnard HR, Ryan MG. **2003**. A test of the hydraulic limitation hypothesis in fast-growing *Eucalyptus saligna*. *Plant, Cell and Environment* **26**: 1235–1245.
- Barrett DJ, Hatton TJ, Ash JE, Ball MC. **1995**. Evaluation of the heat pulse velocity technique for measurement of sap flow In rainforest and eucalypt forest species of south-eastern Australia.: 463–469.
- Bartlett MK, Klein T, Jansen S, Choat B, Sack L. **2016**. The correlations and sequence of plant stomatal, hydraulic, and wilting responses to drought. *Proceedings of the National Academy of*

- Sciences* **113**: 13098–13103.
- Bartlett MK, Scoffoni C, Sack L. **2012**. The determinants of leaf turgor loss point and prediction of drought tolerance of species and biomes: A global meta-analysis: Drivers of plant drought tolerance. *Ecology Letters* **15**: 393–405.
- Bartoń K. **2017**. *MuMIn: Multi-Model Inference*.
- Bartoń K. **2020**. MuMIn: Multi-Model Inference. *R package version 1.43.17*.
- Bates D, Mächler M, Bolker B, Walker S. **2015**. Fitting Linear Mixed-Effects Models Using **lme4**. *Journal of Statistical Software* **67**.
- Becker P. **1998**. Limitations of a compensation heat pulse velocity system at low sap flow: implications for measurements at night and in shaded trees. *Tree Physiology* **18**: 177–184.
- Beer C, Reichstein M, Tomelleri E, Ciais P, Jung M, Carvalhais N, Rödenbeck C, Arain MA, Baldocchi D, Bonan GB *et al.* **2010**. Terrestrial gross carbon dioxide uptake: Global distribution and covariation with climate. *Science* **329**: 834–838.
- Beerling DJ. **2015**. Gas valves, forests and global change: A commentary on Jarvis (1976) ‘The interpretation of the variations in leaf water potential and stomatal conductance found in canopies in the field’. *Philosophical Transactions of the Royal Society B: Biological Sciences* **370**: 20140311.
- Benyon RG, Lane PNJ, Jaskierniak D, Kuczera G, Haydon SR. **2015**. Use of a forest sapwood area index to explain long-term variability in mean annual evapotranspiration and streamflow in moist eucalypt forests. *Water Resources Research* **51**: 5318–5331.
- Berdanier AB, Miniat CF, Clark JS. **2016**. Predictive models for radial sap flux variation in coniferous, diffuse-porous and ring-porous temperate trees. *Tree Physiology* **36**: 932–941.
- Berry ZC, Looker N, Holwerda F, Gómez Aguilar LR, Ortiz Colin P, González Martínez T, Asbjornsen H. **2018**. Why size matters: The interactive influences of tree diameter distribution and sap flow parameters on upscaled transpiration. *Tree Physiology* **38**: 263–275.
- Bleby TM, Burgess SSO, Adams MA. **2004**. A validation, comparison and error analysis of two heat-pulse methods for measuring sap flow in *Eucalyptus marginata* saplings. *Functional Plant Biology* **31**: 645–658.
- Bleby T, McElrone A, Burgess S. **2008**. Limitations of the HRM: great at low flow rates, but not yet up to speed? In: 7th sap flow workshop. Seville.
- Boese S, Jung M, Carvalhais N, Teuling AJ, Reichstein M. **2018**. *Carbon–Water Flux Coupling Under Progressive Drought*. Biogeochemistry: Air - Land Exchange.
- Bohrer G, Mourad H, Laursen TA, Drewry D, Avissar R, Poggi D, Oren R, Katul GG. **2005**. Finite element tree crown hydrodynamics model (FETCH) using porous media flow within branching elements: A new representation of tree hydrodynamics. *Water Resources Research*

- 41: W11404.
- Bonan GB, Levis S, Sitch S, Vertenstein M, Oleson KW. **2003**. A dynamic global vegetation model for use with climate models: Concepts and description of simulated vegetation dynamics. *Global Change Biology* **9**: 1543–1566.
- Bond-Lamberty B. **2018**. Data Sharing and Scientific Impact in Eddy Covariance Research. *Journal of Geophysical Research (Biogeosciences)* **123**: 1440–1443.
- Bond-Lamberty B, Thomson A. **2010**. A global database of soil respiration data. *Biogeosciences* **7**: 1915–1926.
- Bourne AE, Creek D, Peters JMR, Ellsworth DS, Choat B. **2017**. Species climate range influences hydraulic and stomatal traits in Eucalyptus species. *Annals of Botany* **120**: 123–133.
- Bowen IS. **1926**. The ratio of heat losses by conduction and by evaporation from any water surface. *Phys. Rev.* **27**: 779–787.
- Braun P, Schmid J. **1999**. Sap flow measurements in grapevines (*Vitis vinifera L.*) 2. Granier measurements. *Plant and Soil* **215**: 47–55.
- Bretfeld M, Ewers BE, Hall JS. **2018**. Plant water use responses along secondary forest succession during the 2015-2016 El Niño drought in Panama. *New Phytologist* **219**: 885–899.
- Bréda N, Huc R, Granier A, Dreyer E. **2006**. Temperate forest trees and stands under severe drought: A review of ecophysiological responses, adaptation processes and long-term consequences. *Ann. For. Sci.* **63**: 625–644.
- Brinkmann N, Eugster W, Zweifel R, Buchmann N, Kahmen A. **2016**. Temperate tree species show identical response in tree water deficit but different sensitivities in sap flow to summer soil drying. *Tree Physiology* **36**: 1508–1519.
- Brodersen CR, Roddy AB, Wason JW, McElrone AJ. **2019**. Functional status of xylem through time. *Annual Review of Plant Biology* **70**: 407–433.
- Brodribb TJ, Holbrook NM. **2003**. Stomatal Closure during Leaf Dehydration, Correlation with Other Leaf Physiological Traits. *Plant Physiology* **132**: 2166–2173.
- Brodribb TJ, Powers J, Cochard H, Choat B. **2020**. Hanging by a thread? Forests and drought. *Science* **368**: 261–266.
- Brutsaert W. **1982**. *Evaporation into the Atmosphere*. Dordrecht: Springer Netherlands.
- Buckley TN. **2005**. The control of stomata by water balance. *New Phytologist* **168**: 275–292.
- Buckley TN. **2019**. How do stomata respond to water status? *New Phytologist* **224**: 21–36.
- Buckley TN, Turnbull TL, Adams MA. **2012**. Simple models for stomatal conductance derived from a process model: Cross-validation against sap flux data. *Plant, Cell & Environment* **35**: 1647–1662.
- Burgess SS, Adams M a, Turner NC, Beverly CR, Ong CK, Khan a a, Bleby TM. **2001**. An

- improved heat pulse method to measure low and reverse rates of sap flow in woody plants. *Tree physiology* **21**: 589–598.
- Bush SE, Hultine KR, Sperry JS, Ehleringer JR, Phillips N. **2010**. Calibration of thermal dissipation sap flow probes for ring- and diffuse-porous trees. *Tree Physiology* **30**: 1545–1554.
- (C3S) CCCS. **2017**. ERA5: Fifth generation of ECMWF atmospheric reanalyses of the global climate .
- (C3S) CCCS. **2019**. C3S ERA5-Land reanalysis . Copernicus Climate Change Service, May 2020.
- Cabibel B, Do F, Horoyan J. **1991**. Mesures thermiques des flux de sève dans les troncs et les racines et fonctionnement hydrique des arbres. i. analyse théorique des erreurs sur la mesure des flux et validation des mesures en présence de gradients thermiques extérieurs. *Agronomie* **11**: 669–678.
- Cain R. **2009**. The climatic significance of tropical forest edges and their representation in global climate models.
- Caspari HW, Green SR, Edwards WR. **1993**. Transpiration of well-watered and water-stressed Asian pear trees as determined by lysimetry, heat-pulse, and estimated by a Penman-Monteith model. *Agricultural and Forest Meteorology* **67**: 13–27.
- Castelan-Estrada M, Vivin P, Gaudillière JP. **2002**. Allometric relationships to estimate seasonal above-ground vegetative and reproductive biomass of *Vitis vinifera* L. *Annals of Botany* **89**: 401–408.
- Caterina GL, Will RE, Turton DJ, Wilson DS, Zou CB. **2013**. Water use of *Juniperus virginiana* trees encroached into mesic prairies in Oklahoma, USA. *Ecohydrology* **7**: 1124–1134.
- Center EROAS (EROS). **2017**. Shuttle Radar Topography Mission (SRTM) 1 Arc-Second Global.
- Cerný M, Mazal P, Cermák J, Nohál L. **2011**. Potentials of sap flow evaluation by means of acoustic emission measurements. *Acta Universitatis Agriculturae et Silviculturae Mendelianae Brunensis* **59**: 105–110.
- Cerveny RS, Lawrimore J, Edwards R, Landsea C. **2007**. Extreme weather records: Compilation, adjudication, and publication. *Bulletin of the American Meteorological Society* **88**: 853–860.
- Chan AM. **2015**. *Tree Transpiration from Two Forests in the Wasatch Mountains, Utah*. University of Utah; Department of Biology, University of Utah.
- Chen Y-J, Cao K-F, Schnitzer SA, Fan Z-X, Zhang J-L, Bongers F. **2015**. Water-use advantage for lianas over trees in tropical seasonal forests. *New Phytologist* **205**: 128–136.
- Chen X, Miller GR, Rubin Y, Baldocchi DD. **2012**. A statistical method for estimating wood thermal diffusivity and probe geometry using in situ heat response curves from sap flow measurements. *Tree Physiology* **32**: 1458–1470.
- Choat B, Brodribb TJ, Brodersen CR, Duursma RA, López R, Medlyn BE. **2018**. Triggers of

- tree mortality under drought. *Nature* **558**: 531–539.
- Choat B, Jansen S, Brodribb TJ, Cochard H, Delzon S, Bhaskar R, Bucci SJ, Feild TS, Gleason SM, Hacke UG *et al.* **2012**. Global convergence in the vulnerability of forests to drought. *Nature* **491**: 752–755.
- Christoffersen BO, Gloor M, Fauset S, Fyllas NM, Galbraith DR, Baker TR, Kruijt B, Rowland L, Fisher RA, Binks OJ *et al.* **2016**. Linking hydraulic traits to tropical forest function in a size-structured and trait-driven model (tfs v.1-hydro). *Geoscientific Model Development* **9**: 4227–4255.
- Chu H, Baldocchi DD, Poindexter C, Abraha M, Desai AR, Bohrer G, Arain MA, Griffis T, Blanken PD, O'Halloran TL *et al.* **2018**. Temporal Dynamics of Aerodynamic Canopy Height Derived From Eddy Covariance Momentum Flux Data Across North American Flux Networks. *Geophysical Research Letters* **45**: 9275–9287.
- Chuang Y-L, Oren R, Bertozzi AL, Phillips N, Katul GG. **2006**. The porous media model for the hydraulic system of a conifer tree: Linking sap flux data to transpiration rate. *Ecological Modelling* **191**: 447–468.
- Clearwater MJ, Luo Z, Mazzeo M, Dichio B. **2009**. An external heat pulse method for measurement of sap flow through fruit pedicels, leaf petioles and other small-diameter stems. *Plant, Cell and Environment* **32**: 1652–1663.
- Clearwater MJ, Meinzer FC, Andrade JL, Goldstein G, Holbrook NM. **1999**. Potential errors in measurement of nonuniform sap flow using heat dissipation probes. *Tree Physiology* **19**: 681–687.
- Cohen Y, Cohen S, Cantuarias-Avilés T, Schiller G. **2008**. Variations in the radial gradient of sap velocity in trunks of forest and fruit trees. *Plant and Soil* **305**: 49–59.
- Cohen M, Doltra J, Oncins JA, Bonany J, Camps F, Salvia J. **1998**. Heat pulse method for measuring sap flow in fruit trees. In: 4th international workshop on measuring sap flow in intact plants. Zidlochovice, Czech Republic: IUFRO Publications, 12–21.
- Cohen Y, Fuchs M, Green GC. **1981**. Improvement of the heat pulse method for determining sap flow in trees. *Plant, Cell and Environment* **4**: 391–397.
- Corelli-Grappadelli L, Magnanini E. **1993**. A whole-tree system for gas-exchange studies. *HortScience HortSci* **28**: 41–45.
- Costa ACL da, Rowland L, Oliveira RS, Oliveira AAR, Binks OJ, Salmon Y, Vasconcelos SS, Junior JAS, Ferreira LV, Poyatos R *et al.* **2018**. Stand dynamics modulate water cycling and mortality risk in droughted tropical forest. *Global Change Biology* **24**: 249–258.
- Crowther TW, Glick HB, Covey KR, Bettigole C, Maynard DS, Thomas SM, Smith JR, Hintler G, Duguid MC, Amatulli G *et al.* **2015**. Mapping tree density at a global scale. *Nature* **525**: 201–205.
- Cunningham S. **2004**. Stomatal sensitivity to vapour pressure deficit of temperate and tropical

- evergreen rainforest trees of Australia. *Trees* **18**.
- Čermák J, Deml M, Penka M. **1973**. A new method of sap flow rate determination in trees. *Biologia Plantarum* **15**: 171–178.
- Čermák J, Kučera J, Nadezhina N. **2004**. Sap flow measurements with some thermodynamic methods, flow integration within trees and scaling up from sample trees to entire forest stands. *Trees - Structure and Function* **18**: 529–546.
- Dai A. **2013**. Increasing drought under global warming in observations and models. *Nature Climate Change* **3**: 52–58.
- Dai S-Q, Li H, Xiong J, Ma J, Guo H-Q, Xiao X, Zhao B. **2018**. Assessing the extent and impact of online data sharing in eddy covariance flux research. *Journal of Geophysical Research: Biogeosciences* **123**: 129–137.
- Damour G, Simonneau T, Cochard H, Urban L. **2010**. An overview of models of stomatal conductance at the leaf level. *Plant, Cell & Environment* **33**: 1419–1438.
- Daum CR. **1967**. A method for determining water transport in trees. *Ecology* **48**: 425–431.
- de Cáceres M, Mencuccini M, Martin-StPaul N, Limousin J-M, Coll L, Poyatos R, Cabon A, Granda V, Forner A, Valladares F *et al.* **2021**. Unravelling the effect of species mixing on water use and drought stress in mediterranean forests: A modelling approach. *Agricultural and Forest Meteorology* **296**: 108233.
- de Oliveira Reis F, Campostrini E, Sousa EF de, Silva MGe. **2006**. Sap flow in papaya plants: Laboratory calibrations and relationships with gas exchanges under field conditions. *Scientia Horticulturae* **110**: 254–259.
- Diawara a, Loustau D, Berbigier P. **1991**. Comparison of 2 methods for estimating the evaporation of a Pinus pinaster stand - Sap flow and energy-balance with sensible heat-flux measurements by an Eddy Covariance Method. *Agricultural and Forest Meteorology* **54**: 49–66.
- Dierick D, Hölscher D. **2009**. Species-specific tree water use characteristics in reforestation stands in the philippines. *Agricultural and Forest Meteorology* **149**: 1317–1326.
- Ding J, Johnson EA, Martin YE. **2020**. Optimization of leaf morphology in relation to leaf water status: A theory. *Ecology and Evolution* **10**: 1510–1525.
- Do F, Rocheteau A. **2002a**. Influence of natural temperature gradients on measurements of xylem sap flow with thermal dissipation probes. 2. Advantages and calibration of a noncontinuous heating system. *Tree Physiology* **22**: 649–654.
- Do F, Rocheteau A. **2002b**. Influence of natural temperature gradients on measurements of xylem sap flow with thermal dissipation probes. 1. Field observations and possible remedies. *Tree Physiology* **22**: 641–648.
- Domec J-C, Johnson DM. **2012**. Does homeostasis or disturbance of homeostasis in minimum leaf water potential explain the isohydric versus anisohydric behavior of *Vitis vinifera* L. cultivars?

- Tree Physiology **32**: 245–248.
- Dragoni D, Lakso AN, Piccioni RM. **2005**. Transpiration of apple trees in a humid climate using heat pulse sap flow gauges calibrated with whole-canopy gas exchange chambers. *Agricultural and Forest Meteorology* **130**: 85–94.
- Drake JE, Tjoelker MG, Vårhammar A, Medlyn BE, Reich PB, Leigh A, Pfautsch S, Blackman CJ, López R, Aspinwall MJ *et al.* **2018**. Trees tolerate an extreme heatwave via sustained transpirational cooling and increased leaf thermal tolerance. *Global Change Biology* **24**: 2390–2402.
- Dye P, Soko S, Poulter A. **1996**. Evaluation of the heat pulse velocity method for measuring sap flow in *< i>Pinus patula</i>*. *Journal of Experimental Botany* **47**: 975–981.
- Edwards WR, Becker P, Čermák J. **1997**. A unified nomenclature for sap flow measurements. *Tree Physiology* **17**: 65–67.
- Ehlneringer JR, Phillips SL, Schuster WSF, Sandquist DR. **1991**. Differential utilization of summer rains by desert plants. *Oecologia* **88**: 430–434.
- Eller C, Bittencourt P, Oliveira R. **2018**. Using sap flow to measure whole-tree hydraulic conductance loss in response to drought. *Acta Horticulturae*: 75–84.
- Eller CB, Rowland L, Mencuccini M, Rosas T, Williams K, Harper A, Medlyn BE, Wagner Y, Klein T, Teodoro GS *et al.* **2020**. Stomatal optimization based on xylem hydraulics (sox) improves land surface model simulation of vegetation responses to climate. *New Phytologist* **226**: 1622–1637.
- Evans JR, Santiago LS. **2014**. PrometheusWiki Gold Leaf Protocol: Gas exchange using LI-COR 6400. *Functional Plant Biology* **41**: 223.
- Evaristo J, McDonnell JJ. **2017**. Prevalence and magnitude of groundwater use by vegetation: A global stable isotope meta-analysis. *Scientific Reports* **7**.
- Ewers BE, Oren R. **2000**. Analyses of assumptions and errors in the calculation of stomatal conductance from sap flux measurements. *Tree Physiology* **20**: 579–589.
- Ewers B, Oren I, Albaugh TJ, Dougherty P. **1999**. Carry-over effects of water and nutrient supply on water use of *pinus taeda*. *Ecological Applications* **9**: 513–525.
- Ewers BE, Oren R, Phillips N, Strömgren M, Linder S. **2001**. Mean canopy stomatal conductance responses to water and nutrient availabilities in *Picea abies* and *Pinus taeda*. *Tree Physiology* **21**: 841–850.
- Falster DS, Duursma RA, Ishihara MI, Barneche DR, FitzJohn RG, Vårhammar A, Aiba M, Ando M, Anten N, Aspinwall MJ *et al.* **2015**. BAAD: A biomass and allometry database for woody plants. *Ecology* **96**: 1445–1445.
- Fan J, Guyot A, Ostergaard KT, Lockington DA. **2018**. Effects of earlywood and latewood on sap flux density-based transpiration estimates in conifers. *Agricultural and Forest Meteorology*

- 249:** 264–274.
- Fan Y, Miguez-Macho G, Jobbágy EG, Jackson RB, Otero-Casal C. **2017.** Hydrologic regulation of plant rooting depth. *Proceedings of the National Academy of Sciences* **114:** 10572–10577.
- Fathi L. **2014.** Structural and mechanical properties of the wood from coconut palms, oil palms and date palms.
- Fatichi S, Pappas C, Ivanov VY. **2016.** Modeling plant-water interactions: an ecohydrological overview from the cell to the global scale. *Wiley Interdisciplinary Reviews: Water* **3:** 327–368.
- Fauset S, Freitas HC, Galbraith DR, Sullivan MJ, Aidar MP, Joly CA, Phillips OL, Vieira SA, Gloor MU. **2018.** Differences in leaf thermoregulation and water use strategies between three co-occurring Atlantic forest tree species: Leaf energy balance of Atlantic forest trees. *Plant, Cell & Environment* **41:** 1618–1631.
- Feng X, Ackerly DD, Dawson TE, Manzoni S, Skelton RP, Vico G, Thompson SE. **2018.** The ecohydrological context of drought and classification of plant responses (H Maherli, Ed.). *Ecology Letters* **21:** 1723–1736.
- Feng X, Dawson TE, Ackerly DD, Santiago LS, Thompson SE. **2017.** Reconciling seasonal hydraulic risk and plant water use through probabilistic soil–plant dynamics. *Global Change Biology* **23:** 3758–3769.
- Fernández JE, Duran PJ, Palomo MJ, Diaz-Espejo A, Chamorro V, Giron IF. **2006.** Calibration of sap flow estimated by the compensation heat pulse method in olive, plum and orange trees: relationships with xylem anatomy. *Tree Physiology* **26:** 719–728.
- Fernández JE, Palomo MJ, Díaz-Espejo A, Girón I. **1999.** Calibrating the Compensation Heat-Pulse Technique for measuring sap flow in olive. *Acta Horticulturae* **474:** 455–458.
- Ficklin DL, Novick KA. **2017.** Historic and projected changes in vapor pressure deficit suggest a continental-scale drying of the United States atmosphere. *Journal of Geophysical Research: Atmospheres* **122:** 2061–2079.
- Fisher RA, Koven CD. **2020.** Perspectives on the future of land surface models and the challenges of representing complex terrestrial systems. *Journal of Advances in Modeling Earth Systems* **12:** e2018MS001453.
- Fisher RA, Koven CD, Anderegg WRL, Christoffersen BO, Dietze MC, Farrior CE, Holm JA, Hurt GC, Knox RG, Lawrence PJ *et al.* **2018.** Vegetation demographics in earth system models: A review of progress and priorities. *Global Change Biology* **24:** 35–54.
- Flexas J, Díaz-Espejo A, Conesa MA, Coopman RE, Douthe C, Gago J, Gallé A, Galmés J, Medrano H, Ribas-Carbo M *et al.* **2016.** Mesophyll conductance to co₂ and rubisco as targets for improving intrinsic water use efficiency in c₃ plants. *Plant, Cell & Environment* **39:** 965–982.
- Flo V, Martinez-Vilalta J, Steppe K, Schuldt B, Poyatos R. **2019.** A synthesis of bias and

- uncertainty in sap flow methods. *Agricultural and Forest Meteorology* **271**: 362–374.
- Ford CR, Hubbard RM, Kloepel BD, Vose JM. **2007**. A comparison of sap flux-based evapotranspiration estimates with catchment-scale water balance. *Agricultural and Forest Meteorology* **145**: 176–185.
- Forster MA. **2014**. How significant is nocturnal sap flow? *Tree Physiology* **34**: 757–765.
- Forster MA. **2019**. The dual method approach (dma) resolves measurement range limitations of heat pulse velocity sap flow sensors. *Forests* **10**: 46.
- Frank DC, Poulter B, Saurer M, Esper J, Huntingford C, Helle G, Treydte K, Zimmermann NE, Schleser GH, Ahlström A *et al.* **2015**. Water-use efficiency and transpiration across European forests during the Anthropocene. *Nature Climate Change* **5**: 579–583.
- Franks PJ. **2004**. Stomatal control and hydraulic conductance, with special reference to tall trees. *Tree Physiology* **24**: 865–878.
- Fuchs S, Leuschner C, Link R, Coners H, Schuldt B. **2017**. Calibration and comparison of thermal dissipation, heat ratio and heat field deformation sap flow probes for diffuse-porous trees. *Agricultural and Forest Meteorology* **244–245**: 151–161.
- Gallagher R, Falster D, Maitner B, Salguero-Gómez R, Vandvik V, Pearse W, Schneider F, Kattge J, Poelen J, Madin J *et al.* **2020**. Open science principles for accelerating trait-based science across the tree of life. *Nature Ecology & Evolution* **4**: 294–303.
- Gentine P, Massmann A, Lintner BR, Hamed Alemohammad S, Fu R, Green JK, Kennedy D, Vilà-Guerau de Arellano J. **2019**. Land–atmosphere interactions in the tropics – a review. *Hydrology and Earth System Sciences* **23**: 4171–4197.
- Giardina F, Konings AG, Kennedy D, Alemohammad SH, Oliveira RS, Uriarte M, Gentine P. **2018**. Tall Amazonian forests are less sensitive to precipitation variability. *Nature Geoscience* **11**: 405–409.
- Gleason SM, Westoby M, Jansen S, Choat B, Hacke UG, Pratt RB, Bhaskar R, Brodribb TJ, Bucci SJ, Cao K-F *et al.* **2016**. Weak tradeoff between xylem safety and xylem-specific hydraulic efficiency across the world's woody plant species. *New Phytologist* **209**: 123–136.
- Gonzales-Altozano P, Ruiz-Sin B, Castel J. **1998**. Sap flow determination in citrus trees by heat pulse techniques. In: Cermák J, Nadezhina N, eds. 4th international workshop on measuring sap flow in intact plants. Zidlochovice, Czech Republic: IUFRO.
- Good SP, Moore GW, Miralles DG. **2017**. A mesic maximum in biological water use demarcates biome sensitivity to aridity shifts. *Nature Ecology & Evolution* **1**: 1883–1888.
- Gorelick N, Hancher M, Dixon M, Ilyushchenko S, Thau D, Moore R. **2017**. Google Earth Engine: Planetary-scale geospatial analysis for everyone. *Remote Sensing of Environment* **202**: 18–27.
- Granda V, Poyatos R, Flo V, Nelson JA, Team SC. **2020**. Sapfluxnetr: Working with 'Sapfluxnet'

- Project Data. *R package version 0.1.1.*
- Granda V, Poyatos R, Flo V, Nelson JA, Sirot M, Bagaria G. **2016**. sapfluxnetQC1: R package with functions related to sapfluxnet project. [online] Available from: <https://github.com/sapfluxnet/sapfluxn>
- Granier A. **1985**. Une nouvelle methode pour la mesure du flux de seve brute dans le tronc des arbres. *Annales des Sciences Forestieres* **42**(2): 193–200.
- Granier A. **1987**. Evaluation of transpiration in a Douglas-fir stand by means of sap flow measurements. *Tree Physiology* **3**: 309–320.
- Granier A, Biron P, Köstner B, Gay LW, Najjar G. **1996**. Comparisons of xylem sap flow and water vapour flux at the stand level and derivation of canopy conductance for Scots pine. *Theoretical and Applied Climatology* **53**: 115–122.
- Green SR, Clothier BE. **1988**. Water use of kiwifruit vines and apple trees by the heat pulse technique. *Journal of Experimental Botany* **39**: 115–123.
- Green S, Clothier B, Jardine B. **2003**. Theory and Practical Application of Heat Pulse to Measure Sap Flow. *Agriculture*.
- Green S, Clothier B, Perie E. **2009**. A re-analysis of heat pulse theory across a wide range of sap flows. *Acta Horticulturae* **846**: 95–104.
- Green JK, Seneviratne SI, Berg AM, Findell KL, Hagemann S, Lawrence DM, Gentine P. **2019**. Large influence of soil moisture on long-term terrestrial carbon uptake. *Nature* **565**: 476–479.
- Grissino-Mayer HD, Fritts HC. **1997**. The international tree-ring data bank: An enhanced global database serving the global scientific community. *The Holocene* **7**: 235–238.
- Grossiord C, Buckley TN, Cernusak LA, Novick KA, Poulter B, Siegwolf RTW, Sperry JS, McDowell NG. **2020**. Plant responses to rising vapor pressure deficit. *New Phytologist* **226**: 1550–1566.
- Grossiord C, Christoffersen B, Alonso-Rodríguez AM, Anderson-Teixeira K, Asbjørnsen H, Aparecido LMT, Carter Berry Z, Baraloto C, Bonal D, Borrego I *et al.* **2019**. Precipitation mediates sap flux sensitivity to evaporative demand in the neotropics. *Oecologia* **191**: 519–530.
- Grossiord C, Sevanto S, Adams HD, Collins AD, Dickman LT, McBranch N, Michaletz ST, Stockton EA, Vigil M, McDowell NG. **2017**. Precipitation, not air temperature, drives functional responses of trees in semi-arid ecosystems (L Huenneke, Ed.). *Journal of Ecology* **105**: 163–175.
- Grossiord C, Sevanto S, Limousin J-M, Meir P, Mencuccini M, Pangle RE, Pockman WT, Salmon Y, Zweifel R, McDowell NG. **2018**. Manipulative experiments demonstrate how long-term soil moisture changes alter controls of plant water use. *Environmental and Experimental Botany* **152**: 19–27.
- Gutierrez MV, Harrington RA, Meinzer FC, Fownes JH. **1994**. the Effect of Environmentally-Induced Stem Temperature-Gradients on Transpiration Estimates From the Heat-Balance

- Method in 2 Tropical Woody Species. *Tree Physiology* **14**: 179–190.
- Gutiérrez-Soto M, Mora E, Araya JM, Torres J, Castillo A. **2012**. HRM - Heat Ratio Method to Measure Sap Flow in Papaya & Oil Palm. Performance of the heat ratio method (HRM) to measure sap flow in Papaya (Carica papaya L.) ‘Pocosí’ hybrid, and Oil Palm (Elaeis guineensis Jack.) compact clones.: 2–7.
- Hampel FR. **1974**. The influence curve and its role in robust estimation. *Journal of the American Statistical Association* **69**: 383–393.
- Hanssens J, De Swaef T, Nadezhina N, Steppe K. **2013**. Measurement of sap flow dynamics through the tomato peduncle using a non-invasive sensor based on the heat field deformation method. *Acta Horticulturae* **991**: 409–416.
- Hassler SK, Weiler M, Blume T. **2018**. Tree-, stand- and site-specific controls on landscape-scale patterns of transpiration. *Hydrology and Earth System Sciences* **22**: 13–30.
- Hatton T. **1990**. Integration of sapflow velocity to estimate plant water use. *Tree Physiology* **6**: 201–209.
- Hatton TJ, Moore SJ, Reece PH. **1995**. Estimating stand transpiration in a Eucalyptus populnea woodland with the heat pulse method: measurement errors and sampling strategies. *Tree physiology* **15**: 219–227.
- Heilman J, Ham J. **1990**. Measurement of mass flow rate of sap in Ligustrum japonicum. *HortScience* **25**: 465–467.
- Hengl T, Mendes de Jesus J, Heuvelink GBM, Ruiperez Gonzalez M, Kilibarda M, Blagotić A, Shangguan W, Wright MN, Geng X, Bauer-Marschallinger B *et al.* **2017**. SoilGrids250m: Global gridded soil information based on machine learning (B Bond-Lamberty, Ed.). *PLOS ONE* **12**: e0169748.
- Henry C, John GP, Pan R, Bartlett MK, Fletcher LR, Scoffoni C, Sack L. **2019**. A stomatal safety-efficiency trade-off constrains responses to leaf dehydration. *Nature Communications* **10**: 3398.
- Herbst M, Roberts JM, Rosier PTW, Gowing DJ. **2007**. Seasonal and interannual variability of canopy transpiration of a hedgerow in southern England. *Tree physiology* **27**: 321–33.
- Hernandez-Santana V, Hernandez-Hernandez A, Vadéboncoeur MA, Asbjornsen H. **2015**. Scaling from single-point sap velocity measurements to stand transpiration in a multispecies deciduous forest: uncertainty sources, stand structure effect, and future scenarios. *Canadian Journal of Forest Research* **45**: 1489–1497.
- Hijmans RJ. **2020**. Raster: Geographic Data Analysis and Modeling. *R package version 3.3-13*.
- Hillel D. **1998**. *Environmental soil physics*. San Diego, CA: Academic Press.
- Hogg EH, Black TA, Hartog G den, Neumann HH, Zimmermann R, Hurdle PA, Blanken PD, Nesic Z, Yang PC, Staebler RM *et al.* **1997**. A comparison of sap flow and eddy fluxes of

- water vapor from a boreal deciduous forest. *Journal of Geophysical Research* **102**: 28929.
- Hoshika Y, Osada Y, Marco A de, Peñuelas J, Paoletti E. **2018**. Global diurnal and nocturnal parameters of stomatal conductance in woody plants and major crops. *Global Ecology and Biogeography* **27**: 257–275.
- Howell T. **2005**. Lysimetry. In: Hillel D, ed. Encyclopedia of soils in the environment. Oxford: Elsevier, 379–386.
- Hölttä T, Linkosalo T, Riikonen A, Sevanto S, Nikinmaa E. **2015**. An analysis of Granier sap flow method, its sensitivity to heat storage and a new approach to improve its time dynamics. *Agricultural and Forest Meteorology* **211-212**: 2–12.
- Hu J, Moore D, Riveros-Iregui D, Burns S, Monson R. **2010**. Modeling whole-tree carbon assimilation rate using observed transpiration rates and needle sugar carbon isotope ratios. *New Phytologist* **185**: 1000–1015.
- Huber B. **1932**. Beobachtung und messung pflanzlicher sartströme. *Berichte der Deutschen Botanischen Gesellschaft* **50**: 89–109.
- Hultine KR, Nagler PL, Morino K, Bush SE, Burtch KG, Dennison PE, Glenn EP, Ehleringer JR. **2010**. Sap flux-scaled transpiration by tamarisk (*Tamarix* spp.) before, during and after episodic defoliation by the saltcedar leaf beetle (*Diorhabda carinulata*). *Agricultural and Forest Meteorology* **150**: 1467–1475.
- Intrigliolo DS, Lakso AN, Piccioni RM. **2009**. Using the heat pulse ‘Tmax’ procedure to estimate grapevine water use in a humid climate. *Acta Horticulturae* **846**: 177–184.
- Isarangkool Na Ayutthaya S, Do FC, Pannengpetch K, Junjittakarn J, Maeght JL, Rocheteau A, Cochard H. **2010**. Transient thermal dissipation method of xylem sap flow measurement: Multi-species calibration and field evaluation. *Tree Physiology* **30**: 139–148.
- Jackson PC, Cavelier J, Goldstein G, Meinzer FC, Holbrook NM. **1995**. Partitioning of water resources among plants of a lowland tropical forest. *Oecologia* **101**: 197–203.
- Jacobsen AL, Pratt RB, Davis SD, Ewers FW. **2007**. Cavitation resistance and seasonal hydraulics differ among three arid californian plant communities. *Plant, Cell & Environment* **30**: 1599–1609.
- Jaeger B. **2017**. R2glmm: Computes R Squared for Mixed (Multilevel) Models. *CRAN version 0.1.2*.
- Jarvis PG. **1976**. The interpretation of the variations in leaf water potential and stomatal conductance found in canopies in the field. *Philosophical Transactions of the Royal Society of London. B, Biological Sciences* **273**: 593–610.
- Jarvis PG. **1995**. Scaling processes and problems. *Plant, Cell & Environment* **18**: 1079–1089.
- Jarvis PG, McNaughton KG. **1986**. Stomatal control of transpiration: scaling up from leaf to

- region. In: Advances in Ecological Research. Elsevier, 1–49.
- Jasechko S, Sharp ZD, Gibson JJ, Birks SJ, Yi Y, Fawcett PJ. **2013**. Terrestrial water fluxes dominated by transpiration. *Nature* **496**: 347–350.
- Johnson DM, McCulloh KA, Woodruff DR, Meinzer FC. **2012**. Hydraulic safety margins and embolism reversal in stems and leaves: Why are conifers and angiosperms so different? *Plant Science* **195**: 48–53.
- Jones HG. **1997**. Stomatal control of photosynthesis and transpiration. *Journal of Experimental Botany* **49**: 387–398.
- Jones TS, Winbourne JB, Hutyra LR. **2020**. Ribbonized sap flow: An emerging technology for the integration of sap flow sensor components onto a single platform. *Ecosphere* **11**.
- Josse J, Husson F. **2016**. missMDA : A Package for Handling Missing Values in Multivariate Data Analysis. *Journal of Statistical Software* **70**.
- Jung M, Koirala S, Weber U, Ichii K, Gans F, Camps-Valls G, Papale D, Schwalm C, Tramontana G, Reichstein M. **2019**. The fluxcom ensemble of global land-atmosphere energy fluxes. *Scientific data* **6**.
- Jung M, Reichstein M, Schwalm CR, Huntingford C, Sitch S, Ahlström A, Arneth A, Camps-Valls G, Ciais P, Friedlingstein P *et al.* **2017**. Compensatory water effects link yearly global land CO₂ sink changes to temperature. *Nature* **541**: 516–520.
- Kallarackal J, Otieno D, Reineking B, Jung E, Schmidt M, Granier A, Tenhunen J. **2013**. Functional convergence in water use of trees from different geographical regions: A meta-analysis. *Trees* **27**: 787–799.
- Kannenberg SA, Schwalm CR, Anderegg WRL. **2020**. Ghosts of the past: How drought legacy effects shape forest functioning and carbon cycling (M Rejmanek, Ed.). *Ecology Letters* **23**: 891–901.
- Karger DN, Conrad O, Böhner J, Kawohl T, Kreft H, Soria-Auza RW, Zimmermann NE, Linder HP, Kessler M. **2017**. Climatologies at high resolution for the earth's land surface areas. *Scientific Data* **4**: 170122.
- Kasurinen V, Alfredsen K, Kolari P, Mammarella I, Alekseychik P, Rinne J, Vesala T, Bernier P, Boike J, Langer M *et al.* **2014**. Latent heat exchange in the boreal and arctic biomes. *Global Change Biology* **20**: 3439–3456.
- Kattge J, Bönisch G, Díaz S, Lavorel S, Prentice IC, Leadley P, Tautenhahn S, Werner GDA, Aakala T, Abedi M *et al.* **2020**. TRY plant trait database – enhanced coverage and open access. *Global Change Biology* **26**: 119–188.
- Kattge J, Díaz S, Lavorel S, Prentice IC, Leadley P, Bönisch G, Garnier E, Westoby M, Reich PB, Wright IJ *et al.* **2011**. TRY - a global database of plant traits. *Global Change Biology*

- 17: 2905–2935.
- Katul GG, Oren R, Manzoni S, Higgins C, Parlange MB. **2012**. Evapotranspiration: A process driving mass transport and energy exchange in the soil-plant-atmosphere-climate system: Evapotranspiration and climate. *Reviews of Geophysics* **50**.
- Katul G, Porporato A, Oren R. **2007**. Stochastic dynamics of plant-water interactions. *Annual Review of Ecology, Evolution, and Systematics* **38**: 767–791.
- Kauwe MG de, Medlyn BE, Knauer J, Williams CA. **2017**. Ideas and perspectives: How coupled is the vegetation to the boundary layer? *Biogeosciences* **14**: 4435–4453.
- Keenan TF, Hollinger DY, Bohrer G, Dragoni D, Munger JW, Schmid HP, Richardson AD. **2013**. Increase in forest water-use efficiency as atmospheric carbon dioxide concentrations rise. *Nature* **499**: 324–327.
- Kempe A, Lautenschl??ger T, Lange A, Neinhuis C. **2014**. How to become a tree without wood - biomechanical analysis of the stem of Carica papaya L. *Plant Biology* **16**: 264–271.
- Kennedy D, Swenson S, Oleson KW, Lawrence DM, Fisher R, Lola da Costa AC, Gentine P. **2019**. Implementing plant hydraulics in the community land model, version 5. *Journal of Advances in Modeling Earth Systems* **11**: 485–513.
- Klein T. **2014**. The variability of stomatal sensitivity to leaf water potential across tree species indicates a continuum between isohydric and anisohydric behaviours. *Functional Ecology* **28**: 1313–1320.
- Klein T, Rotenberg E, Tatarinov F, Yakir D. **2016**. Association between sap flow-derived and eddy covariance-derived measurements of forest canopy co₂ uptake. *New Phytologist* **209**: 436–446.
- Knauer J, Werner C, Zaehle S. **2015**. Evaluating stomatal models and their atmospheric drought response in a land surface scheme: A multibiome analysis. *Journal of Geophysical Research: Biogeosciences* **120**: 1894–1911.
- Knowles JE, Frederick C. **2016**. merTools: Tools for analyzing mixed effect regression models. *R package version 0.3. 0*.
- Konings AG, Williams AP, Gentine P. **2017**. Sensitivity of grassland productivity to aridity controlled by stomatal and xylem regulation. *Nature Geoscience* **10**: 284–288.
- Kool D, Agam N, Lazarovitch N, Heitman JL, Sauer TJ, Ben-gal a. **2014**. A review of approaches for evapotranspiration partitioning Author ' s personal copy. *Agricultural and Forest Meteorology* **184**: 56–70.
- Köstner B, Granier A, Cermák J. **1998**. Original article Sapflow measurements in forest stands : methods and uncertainties b Granier Xylem sapflow techniques provide temperature and air. *Annals of Forest Science* **55**: 13–27.
- Lamigueiro OP. **2012**. SolaR: Solar radiation and photovoltaic systems with r. *Journal of Statistical Software* **49**: 1–37.

- tical Software, Articles **50**: 1–32.
- Lemeur R, Fernández J, Steppe K. **2009**. Symbols, SI units and physical quantities within the scope of sap flow studies. *Acta Horticulturae*: 21–32.
- Lenth RV. **2016**. Least-Squares Means: The {R} Package *{lsmeans}*. *Journal of Statistical Software* **69**: 1–33.
- Li X, Gentine P, Lin C, Zhou S, Sun Z, Zheng Y, Liu J, Zheng C. **2019**. A simple and objective method to partition evapotranspiration into transpiration and evaporation at eddy-covariance sites. *Agricultural and Forest Meteorology* **265**: 171–182.
- Lin C, Gentine P, Huang Y, Guan K, Kimm H, Zhou S. **2018**. Diel ecosystem conductance response to vapor pressure deficit is suboptimal and independent of soil moisture. *Agricultural and Forest Meteorology* **250–251**: 24–34.
- Lin Y-S, Medlyn BE, Duursma RA, Prentice IC, Wang H, Baig S, Eamus D, Dios VR de, Mitchell P, Ellsworth DS *et al.* **2015**. Optimal stomatal behaviour around the world. *Nature Climate Change* **5**: 459–464.
- Litvak E, McCarthy HR, Pataki DE. **2012**. Transpiration sensitivity of urban trees in a semi-arid climate is constrained by xylem vulnerability to cavitation. *Tree Physiology* **32**: 373–388.
- Liu H, Gleason SM, Hao G, Hua L, He P, Goldstein G, Ye Q. **2019**. Hydraulic traits are coordinated with maximum plant height at the global scale. *Science Advances* **5**: eaav1332.
- Liu L, Gudmundsson L, Hauser M, Qin D, Li S, Seneviratne SI. **2020**. Soil moisture dominates dryness stress on ecosystem production globally. *Nature Communications* **11**: 4892.
- Liu B, Zhao W, Jin B. **2011**. The response of sap flow in desert shrubs to environmental variables in an arid region of China. *Ecohydrology* **4**: 448–457.
- Looker N, Martin J, Jencso K, Hu J. **2016**. Contribution of sapwood traits to uncertainty in conifer sap flow as estimated with the heat-ratio method. *Agricultural and Forest Meteorology* **223**: 60–71.
- López-Bernal Á, Testi L, Villalobos FJ. **2017**. A single-probe heat pulse method for estimating sap velocity in trees. *New Phytologist*: 321–329.
- Lu P. **2002**. Whole-plant water use of some tropical and subtropical tree crops and its application in irrigation management.: 781–789.
- Lu P, Chacko E. **1998**. Evaluation of Granier's sap flux in young mango trees sensor. *Agronomie* **18**: 461–471.
- Lu Y, Duursma RA, Farrior CE, Medlyn BE, Feng X. **2020**. Optimal stomatal drought response shaped by competition for water and hydraulic risk can explain plant trait covariation. *New Phytologist* **225**: 1206–1217.
- Lu P, Müller WJ, Chacko EK. **2000**. Spatial variations in xylem sap flux density in the trunk of orchard-grown, mature mango trees under changing soil water conditions. *Tree Physiology* **20**:

- 683–692.
- Lu P, Urban L, Zhao P. **2004**. Granier's thermal dissipation probe (TDP) method for measuring sap flow in trees: theory and practice. *Acta Botanica Sinica* **46**: 631–646.
- Lusk CH, Wright I, Reich PB. **2003**. Photosynthetic differences contribute to competitive advantage of evergreen angiosperm trees over evergreen conifers in productive habitats. *New Phytologist* **160**: 329–336.
- Lüdecke, Makowski, Waggoner, Patil. **2020**. Assessment of Regression Models Performance. *CRAN*.
- Mackay DS, Ewers BE, Loranty MM, Kruger EL. **2010**. On the representativeness of plot size and location for scaling transpiration from trees to a stand. *Journal of Geophysical Research: Biogeosciences* **115**.
- MacLean J. **1941**. Thermal Conductivity of wood. *Heating, piping & air conditioning* **13**: 380–391.
- Madurapperuma WS, Bleby TM, Burgess SSO. **2009**. Evaluation of sap flow methods to determine water use by cultivated palms. *Environmental and Experimental Botany* **66**: 372–380.
- Magnani F, Leonardi S, Tognetti R, Grace J, Borghetti M. **1998**. Modelling the surface conductance of a broad-leaf canopy: Effects of partial decoupling from the atmosphere. *Plant, Cell & Environment* **21**: 867–879.
- Maherali H, Moura CF, Caldeira MC, Willson CJ, Jackson RB. **2006**. Functional coordination between leaf gas exchange and vulnerability to xylem cavitation in temperate forest trees. *Plant, Cell and Environment* **29**: 571–583.
- Manzoni S, Vico G, Katul G, Fay PA, Polley W, Palmroth S, Porporato A. **2011**. Optimizing stomatal conductance for maximum carbon gain under water stress: A meta-analysis across plant functional types and climates. *Functional Ecology* **25**: 456–467.
- Manzoni S, Vico G, Katul G, Palmroth S, Jackson RB, Porporato A. **2013**. Hydraulic limits on maximum plant transpiration and the emergence of the safety-efficiency trade-off. *The New phytologist* **198**: 169–78.
- Marañón-Jiménez S, Bulcke J den, Piayda A, Van Acker J, Cuntz M, Rebmann C, Steppe K. **2018**. X-ray computed microtomography characterizes the wound effect that causes sap flow underestimation by thermal dissipation sensors. *Tree Physiology* **38**: 287–301.
- Maréchaux I, Bonal D, Bartlett MK, Burban B, Coste S, Courtois EA, Dulormne M, Goret J, Mira E, Mirabel A *et al.* **2018**. Dry-season decline in tree sapflux is correlated with leaf turgor loss point in a tropical rainforest (A Sala, Ed.). *Functional Ecology* **32**: 2285–2297.
- Marshall DC. **1958**. Measurement of sap flow in conifers by heat transport. *Plant Physiology* **33**: 385–396.
- Martens B, Miralles DG, Lievens H, Schalie R van der, Jeu RAM de, Fernández-Prieto D, Beck

- HE, Dorigo WA, Verhoest NEC. **2017**. GLEAM v3: Satellite-based land evaporation and root-zone soil moisture. *Geoscientific Model Development* **10**: 1903–1925.
- Martin-StPaul N, Delzon S, Cochard H. **2017**. Plant resistance to drought depends on timely stomatal closure (H Maherali, Ed.). *Ecology Letters* **20**: 1437–1447.
- Martínez-Vilalta J, Garcia-Forner N. **2017**. Water potential regulation, stomatal behaviour and hydraulic transport under drought: Deconstructing the iso/anisohydric concept: Deconstructing the iso/anisohydric concept. *Plant, Cell & Environment* **40**: 962–976.
- Martínez-Vilalta J, Korakaki E, Vanderklein D, Mencuccini M. **2007**. Below-ground hydraulic conductance is a function of environmental conditions and tree size in Scots pine. *Functional Ecology* **21**: 1072–1083.
- Martínez-Vilalta J, Poyatos R, Aguadé D, Retana J, Mencuccini M. **2014**. A new look at water transport regulation in plants. *New Phytologist* **204**: 105–115.
- Martínez-Vilalta J, Sala A, Piñol J. **2004**. The hydraulic architecture of Pinaceae – a review. *Plant Ecology (formerly Vegetation)* **171**: 3–13.
- Massmann A, Gentile P, Lin C. **2019**. When does vapor pressure deficit drive or reduce evapotranspiration? *Journal of Advances in Modeling Earth Systems* **11**: 3305–3320.
- Matheny AM, Bohrer G, Garrity SR, Morin TH, Howard CJ, Vogel CS. **2015**. Observations of stem water storage in trees of opposing hydraulic strategies. *Ecosphere* **6**: art165.
- Matheny AM, Bohrer G, Stoy PC, Baker IT, Black AT, Desai AR, Dietze MC, Gough CM, Ivanov VY, Jassal RS *et al.* **2014**. Characterizing the diurnal patterns of errors in the prediction of evapotranspiration by several land-surface models: An nacp analysis. *Journal of Geophysical Research: Biogeosciences* **119**: 1458–1473.
- Matheny AM, Mirfenderesgi G, Bohrer G. **2017**. Trait-based representation of hydrological functional properties of plants in weather and ecosystem models. *Plant Diversity* **39**: 1–12.
- Mazal P, Cerný M, Vlasic F, Nohál L. **2012**. Correlation of sap flow changes in trees with signal of acoustic emission during field measurements.
- McCulloh KA, Domec J-C, Johnson DM, Smith DD, Meinzer FC. **2019**. A dynamic yet vulnerable pipeline: Integration and coordination of hydraulic traits across whole plants. *Plant, Cell & Environment* **42**: 2789–2807.
- McCulloh KA, Winter K, Meinzer FC, Garcia M, Aranda J, Lachenbruch B. **2007**. A comparison of daily water use estimates derived from constant-heat sap-flow probe values and gravimetric measurements in pot-grown saplings. *Tree physiology* **27**: 1355–1360.
- McDowell NG, Allen CD, Anderson-Teixeira K, Aukema BH, Bond-Lamberty B, Chini L, Clark JS, Dietze M, Grossiord C, Hanbury-Brown A *et al.* **2020**. Pervasive shifts in forest dynamics in a changing world. *Science* **368**.
- McDowell N, Pockman WT, Allen CD, Breshears DD, Cobb N, Kolb T, Plaut J, Sperry J, West

- A, Williams DG *et al.* 2008. Mechanisms of plant survival and mortality during drought: Why do some plants survive while others succumb to drought? *New Phytologist* **178**: 719–739.
- Meinzer FC. 2002. Co-ordination of vapour and liquid phase water transport properties in plants. *Plant, Cell and Environment* **25**: 265–274.
- Meinzer FC, Bond BJ, Warren JM, Woodruff DR. 2005. Does water transport scale universally with tree size? *Functional Ecology* **19**: 558–565.
- Meinzer FC, Woodruff DR, Marias DE, Mcculloh KA, Sevanto S. 2014. Dynamics of leaf water relations components in co-occurring iso- and anisohydric conifer species: Dynamics of leaf water relations components. *Plant, Cell & Environment* **37**: 2577–2586.
- Mencuccini M. 2003. The ecological significance of long-distance water transport: Short-term regulation, long-term acclimation and the hydraulic costs of stature across plant life forms. *Plant, Cell and Environment* **26**: 163–182.
- Mencuccini M, Manzoni S, Christoffersen B. 2019a. Modelling water fluxes in plants: From tissues to biosphere. *New Phytologist* **222**: 1207–1222.
- Mencuccini M, Rosas T, Rowland L, Choat B, Cornelissen H, Jansen S, Kramer K, Lapenis A, Manzoni S, Niinemets Ü *et al.* 2019b. Leaf economics and plant hydraulics drive leaf : Wood area ratios. *New Phytologist* **224**: 1544–1556.
- Merlin M, Solarik KA, Landhäusser SM. 2020. Quantification of uncertainties introduced by data-processing procedures of sap flow measurements using the cut-tree method on a large mature tree. *Agricultural and Forest Meteorology* **287**: 107926.
- Miralles DG, De Jeu RA, Gash JH, Holmes TR, Dolman AJ. 2011. Magnitude and variability of land evaporation and its components at the global scale. *Hydrology and Earth System Sciences* **15**: 967–981.
- Miralles DG, Gentile P, Seneviratne SI, Teuling AJ. 2019. Land-atmospheric feedbacks during droughts and heatwaves: State of the science and current challenges. *Annals of the New York Academy of Sciences* **1436**: 19–35.
- Miralles DG, Jiménez C, Jung M, Michel D, Ershadi A, McCabe MF, Hirschi M, Martens B, Dolman AJ, Fisher JB *et al.* 2016. The wacmos-et project – part 2: Evaluation of global terrestrial evaporation data sets. *Hydrology and Earth System Sciences* **20**: 823–842.
- Mirfenderesgi G, Bohrer G, Matheny AM, Fatichi S, Moraes Frasson RP de, Schäfer KVR. 2016. Tree level hydrodynamic approach for resolving aboveground water storage and stomatal conductance and modeling the effects of tree hydraulic strategy. *Journal of Geophysical Research: Biogeosciences* **121**: 1792–1813.
- Mitchell RJ, Irwin RE, Flanagan RJ, Karron JD. 2009. Ecology and evolution of plant-pollinator interactions. *Annals of botany* **103**: 1355–1363.
- Mitchell PJ, O'Grady AP, Tissue DT, White DA, Ottenschlaeger ML, Pinkard EA. 2013. Drought response strategies define the relative contributions of hydraulic dysfunction and car-

- bohydrate depletion during tree mortality. *New Phytologist* **197**: 862–872.
- Moffat AM, Papale D, Reichstein M, Hollinger DY, Richardson AD, Barr AG, Beckstein C, Braswell BH, Churkina G, Desai AR *et al.* **2007**. Comprehensive comparison of gap-filling techniques for eddy covariance net carbon fluxes. *Agricultural and Forest Meteorology* **147**: 209–232.
- Monasterio M, Sarmiento G. **1976**. Phenological Strategies of Plant Species in the Tropical Savanna and the Semi-Deciduous Forest of the Venezuelan Llanos. *Journal of Biogeography* **3**: 325.
- Montague T, Kjelgren R. **2006**. Use of Thermal Dissipation Probes to Estimate Water Loss of Containerized Landscape Trees. *Journal of Environmental Horticulture* **24**: 95–104.
- Morán-López T, Poyatos R, Llorens P, Sabaté S. **2014**. Effects of past growth trends and current water use strategies on scots pine and pubescent oak drought sensitivity. *European Journal of Forest Research* **133**: 369–382.
- Moreno-Martínez Á, Camps-Valls G, Kattge J, Robinson N, Reichstein M, Bodegom P van, Kramer K, Cornelissen JHC, Reich P, Bahn M *et al.* **2018**. A methodology to derive global maps of leaf traits using remote sensing and climate data. *Remote Sensing of Environment* **218**: 69–88.
- Mursinna A, McCormick E, Van Horn K, Sartin L, Matheny A. **2018**. Plant Hydraulic Trait Covariation: A Global Meta-Analysis to Reduce Degrees of Freedom in Trait-Based Hydrologic Models. *Forests* **9**: 446.
- Myneni YK. **2015**. MCD15A3H MODIS/Terra+Aqua Leaf Area Index/FPAR 4-day L4 Global 500m SIN Grid V006.
- Nadezhina N. **2018**. Revisiting the heat field deformation (HFD) method for measuring sap flow. *IForest* **11**: 118–130.
- Nadezhina N, Cermak J, Ceulemans R. **2002**. Radial patterns of sap flow in woody stems of dominant and understory species: Scaling errors associated with positioning of sensors. *Tree physiology* **22** **13**: 907–18.
- Nadezhina N, Cermák J, Nadezhdin V. **1998**. Heat field deformation method for sap flow measurements.: 72–92.
- Nadezhina N, Steppe K, De Pauw DJ, Bequet R, Čermak J, Ceulemans R. **2009**. Stem-mediated hydraulic redistribution in large roots on opposing sides of a douglas-fir tree following localized irrigation. *New Phytologist* **184**: 932–943.
- Nakagawa S, Schielzeth H. **2013**. A general and simple method for obtaining R2 from generalized linear mixed-effects models. *Methods in Ecology and Evolution* **4**: 133–142.
- Nelson JA, Pérez-Priego O, Zhou S, Poyatos R, Zhang Y, Blanken PD, Gimeno TE, Wohlfahrt G, Desai AR, Gioli B *et al.* **2020**. Ecosystem transpiration and evaporation: Insights from three

- water flux partitioning methods across FLUXNET sites. *Global Change Biology*: gcb.15314.
- Nortes PA, Baille A, Egea G, González-Real MM, Domingo R. **2009**. Comparison of almond tree transpiration determined by sap flow measurements and lysimetry. *Acta Horticulturae* **846**: 359–366.
- Novick KA, Ficklin DL, Stoy PC, Williams CA, Bohrer G, Oishi ACC, Papuga SA, Blanken PD, Noormets A, Sulman BN *et al.* **2016**. The increasing importance of atmospheric demand for ecosystem water and carbon fluxes. *Nature Climate Change* **1**: 1–5.
- Novick K, Oren R, Stoy P, Juang J-Y, Siqueira M, Katul G. **2009**. The relationship between reference canopy conductance and simplified hydraulic architecture. *Advances in Water Resources* **32**: 809–819.
- Ogle K, Barber JJ, Barron-Gafford GA, Bentley LP, Young JM, Huxman TE, Loik ME, Tissue DT. **2015**. Quantifying ecological memory in plant and ecosystem processes (E Cleland, Ed.). *Ecology Letters* **18**: 221–235.
- Oishi AC, Hawthorne DA, Oren R. **2016**. Baseline: An open-source, interactive tool for processing sap flux data from thermal dissipation probes. *SoftwareX* **5**: 139–143.
- Oishi AC, Oren R, Stoy PC. **2008**. Estimating components of forest evapotranspiration: A footprint approach for scaling sap flux measurements. *Agricultural and Forest Meteorology* **148**: 1719–1732.
- Oishi AC, Oren R, Novick KA, Palmroth S, Katul GG. **2010**. Interannual invariability of forest evapotranspiration and its consequence to water flow downstream. *Ecosystems*. **13**: 421–436.
- Oki T, Kanae S. **2006**. Global hydrological cycles and world water resources. *Science* **313**: 1068–1072.
- Oliveras I, Llorens P. **2001**. Medium-term sap flux monitoring in a Scots pine stand: analysis of the operability of the heat dissipation method for hydrological purposes. *Tree Physiology* **21**: 473–480.
- Olson ME, Soriano D, Rosell JA, Anfodillo T, Donoghue MJ, Edwards EJ, León-Gómez C, Dawson T, Camarero Martínez JJ, Castorena M *et al.* **2018**. Plant height and hydraulic vulnerability to drought and cold. *Proceedings of the National Academy of Sciences* **115**: 7551–7556.
- Oren R, Phillips N, Ewers B, Pataki D, Megonigal J. **1999a**. Sap-flux-scaled transpiration responses to light, vapor pressure deficit, and leaf area reduction in a flooded taxodium distichum forest. *Tree physiology* **19** **6**: 337–347.
- Oren R, Sperry J, Katul G, Pataki D, Ewers B, Phillips N, Schäfer K. **1999b**. Survey and synthesis of intra-and interspecific variation in stomatal sensitivity to vapour pressure deficit. *Plant, Cell & Environment* **22**: 1515–1526.
- O'Brien JJ, Oberbauer SF, Clark DB. **2004**. Whole tree xylem sap flow responses to multiple

- environmental variables in a wet tropical forest. *Plant, Cell & Environment* **27**: 551–567.
- Passioura JB. **1988**. Water transport in and to roots. *Annual Review of Plant Physiology and Plant Molecular Biology* **39**: 245–265.
- Pastorello T G. **2020**. The FLUXNET2015 dataset and the ONEFlux processing pipeline for eddy covariance data. *Sci Data*.
- Paudel I, Kanety T, Cohen S. **2013**. Inactive xylem can explain differences in calibration factors for thermal dissipation probe sap flow measurements. *Tree Physiology* **33**: 986–1001.
- Pearsall KR, Williams LE, Castorani S, Bleby TM, McElrone AJ. **2014**. Evaluating the potential of a novel dual heat-pulse sensor to measure volumetric water use in grapevines under a range of flow conditions. *Functional Plant Biology* **41**: 874–883.
- Pebesma E. **2020**. Stars: Spatiotemporal Arrays, Raster and Vector Data Cubes. *R package version 0.4-3*.
- Peel MC, McMahon TA, Finlayson BL. **2010**. Vegetation impact on mean annual evapotranspiration at a global catchment scale: VEGETATION IMPACT ON EVAPOTRANSPIRATION. *Water Resources Research* **46**.
- Peters RL, Fonti P, Frank DC, Poyatos R, Pappas C, Kahmen A, Carraro V, Prendin AL, Schneider L, Baltzer JL *et al.* **2018**. Quantification of uncertainties in tree sap flow measured with the thermal dissipation method. *Under Review*.
- Peters RL, Pappas C, Hurley AG, Poyatos R, Flo V, Zweifel R, Goossens W, Steppe K. **2020**. Assimilate, process and analyse thermal dissipation sap flow data using the trex r package. *Methods in Ecology and Evolution* **n/a**: 1–9.
- Pérez-Priego O, Testi L, Orgaz F, Villalobos FJ. **2010**. A large closed canopy chamber for measuring CO₂ and water vapour exchange of whole trees. *Environmental and Experimental Botany* **68**: 131–138.
- Phillips N, Oren R. **1998**. A comparison of daily representations of canopy conductance based on two conditional time-averaging methods and the dependence of daily conductance on environmental factors. *Annales des Sciences Forestières* **55**: 217–235.
- Phillips N, Nagchaudhuri A, Oren R, Katul G. **1997**. Time constant for water transport in loblolly pine trees estimated from time series of evaporative demand and stem sapflow. *Trees - Structure and Function* **11**: 412–419.
- Phillips N, Oren R, Zimmermann R. **1996**. Radial patterns of xylem sap flow in non-, diffuse- and ring-porous tree species. *Plant, Cell & Environment* **19**: 983–990.
- Phillips NG, Oren R, Licata J, Linder S. **2004**. Time series diagnosis of tree hydraulic characteristics. *Tree Physiology* **24**: 879–890.
- Phillips NG, Scholz FG, Bucci SJ, Goldstein G, Meinzer FC. **2009**. Using branch and basal trunk sap flow measurements to estimate whole-plant water capacitance: Comment on Burgess and

- dawson (2008). *Plant Soil* **315**: 315–324.
- Poulter B, Ciais P, Hodson E, Lischke H, Maignan F, Plummer S, Zimmermann NE. **2011**. Plant functional type mapping for earth system models. *Geoscientific Model Development* **4**: 993–1010.
- Poyatos R, Flo V, Granda V, Steppe K, Mencuccini M, Martínez-Vilalta J. **2020a**. Using the SAPFLUXNET database to understand transpiration regulation of trees and forests. In: Acta Horticulturae. International Society for Horticultural Science (ISHS), Leuven, Belgium, 179–186.
- Poyatos R, Granda V, Flo V, Adams MA, Adorján B, Aguadé D, Aidar MP, Allen S, Alvarado-Barrientos MS, Anderson-Teixeira KJ *et al.* **2020b**. *Global transpiration data from sap flow measurements: The SAPFLUXNET database*. Earth System Science Data.
- Poyatos R, Granda V, Flo V, Molowny-Horas R, Steppe K, Mencuccini M, Martínez-Vilalta J. **2020c**. SAPFLUXNET: A global database of sap flow measurements.
- Poyatos R, Granda V, Molowny-Horas R, Mencuccini M, Steppe K, Martínez-Vilalta J. **2016**. SAPFLUXNET: towards a global database of sap flow measurements. *Tree Physiology* **36**: 1449–1455.
- Poyatos R, Martínez-Vilalta J, Čermák J, Ceulemans R, Granier A, Irvine J, Köstner B, Lagergren F, Meiresonne L, Nadezhina N *et al.* **2007**. Plasticity in hydraulic architecture of Scots pine across Eurasia. *Oecologia* **153**: 245–259.
- Prasad PVV, Staggenborg SA, Ristic Z. **2008**. Impacts of drought and/or heat stress on physiological, developmental, growth, and yield processes of crop plants. In: Response of crops to limited water. John Wiley & Sons, Ltd, 301–355.
- Prendergast PT, Astill MS, Green SR, Mills TM, Clothier BE. **2007**. Water use by a kiwifruit vine: Calibration, measurements and a model. *Acta Horticulturae* **753**: 535–538.
- R Core Team. **2017**. *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.
- Rascher KG, Mágus C, Werner C. **2010**. On the use of phloem sap ^{13}C as an indicator of canopy carbon discrimination. *Tree Physiology* **30**: 1499–1514.
- Reich PB. **2014**. The world-wide ‘fast-slow’ plant economics spectrum: A traits manifesto (H Cornelissen, Ed.). *Journal of Ecology* **102**: 275–301.
- Reichstein M, Bahn M, Mahecha MD, Kattge J, Baldocchi DD. **2014**. Linking plant and ecosystem functional biogeography. *Proceedings of the National Academy of Sciences*.
- Ren R, Liu G, Wen M, Horton R, Li B, Si B. **2017**. The effects of probe misalignment on sap flux density measurements and in situ probe spacing correction methods. *Agricultural and Forest Meteorology* **232**: 176–185.
- Renninger HJ, Phillips N, Salvucci GD. **2010**. Wet- vs. Dry-Season Transpiration in an Amazonian Rainforest. *Tree Physiology* **30**: 1499–1514.

- nian Rain Forest Palm *Iriartea deltoidea*: Transpiration Seasonality in Palms. *Biotropica* **42**: 470–478.
- Resco de Dios V, Chowdhury FI, Granda E, Yao Y, Tissue DT. **2019**. Assessing the potential functions of nocturnal stomatal conductance in c3 and c4 plants. *New Phytologist* **223**: 1696–1706.
- Resco de Dios V, Roy J, Ferrio JP, Alday JG, Landais D, Milcu A, Gessler A. **2015**. Processes driving nocturnal transpiration and implications for estimating land evapotranspiration. *Scientific reports* **5**: 10975.
- Reyes-Acosta JL, Vandegehuchte MW, Steppe K, Lubczynski MW. **2012**. Novel, cyclic heat dissipation method for the correction of natural temperature gradients in sap flow measurements. Part 2. Laboratory validation. *Tree Physiology* **32**: 913–929.
- Rodell M, Beaudoin HK, L'Ecuyer TS, Olson WS, Famiglietti JS, Houser PR, Adler R, Bosilovich MG, Clayson CA, Chambers D *et al.* **2015**. The observed state of the water cycle in the early twenty-first century. *Journal of Climate* **28**: 8289–8318.
- Rosas Torrent T, Martínez Vilalta J, Mencuccini M, Saura Mas S, Universitat Autònoma de Barcelona, Departament de Biologia Animal de BV i d. **2019**. Integrating plant hydraulics into functional traits framework to understand plant adjustments along a water availability gradient.
- Rosas T, Mencuccini M, Barba J, Cochard H, Saura-Mas S, Martínez-Vilalta J. **2019**. Adjustments and coordination of hydraulic, leaf and stem traits along a water availability gradient. *New Phytologist* **223**: 632–646.
- Rosseel Y. **2012**. Lavaan : An R Package for Structural Equation Modeling. *Journal of Statistical Software* **48**.
- Rubilar RA, Hubbard RM, Yañez MA, Medina AM, Valenzuela HE. **2017**. Quantifying differences in thermal dissipation probe calibrations for *Eucalyptus globulus* species and *E. nitens* × *globulus* hybrid. *Trees - Structure and Function* **31**: 1263–1270.
- Running SW, Nemani RR, Heinsch FA, Zhao M, Reeves M, Hashimoto H. **2004**. A Continuous Satellite-Derived Measure of Global Terrestrial Primary Production. *BioScience* **54**: 547–560.
- Sage RF. **2002**. Variation in the kcat of Rubisco in C3 and C4 plants and some implications for photosynthetic performance at high and low temperature. *Journal of Experimental Botany* **53**: 609–620.
- Sakuratani T. **1981**. A Heat Balance Method for Measuring Water Flux in the Stem of Intact Plants. *J. Agr. Met.* **37**: 9–17.
- Salomón RL, Limousin J-M, Ourcival J-M, Rodríguez-Calcerrada J, Steppe K. **2017**. Stem hydraulic capacitance decreases with drought stress: Implications for modelling tree hydraulics in the mediterranean oak *quercus ilex*. *Plant, Cell & Environment* **40**: 1379–1391.
- Sanchez-Martinez P, Martínez-Vilalta J, Dexter KG, Segovia RA, Mencuccini M. **2020**. Adapta-

- tion and coordinated evolution of plant hydraulic traits. *Ecology Letters* **23**: 1599–1610.
- Satorra A, Bentler PM. **2001**. A scaled difference chi-square test statistic for moment structure analysis. *Psychometrika* **66**: 507–514.
- Sánchez-Costa E, Poyatos R, Sabaté S. **2015**. Contrasting growth and water use strategies in four co-occurring mediterranean tree species revealed by concurrent measurements of sap flow and stem diameter variations. *Agricultural and Forest Meteorology* **207**: 24–37.
- Schäfer KVR, Oren R, Tenhunen JD. **2000**. The effect of tree height on crown level stomatal conductance. *Plant, Cell & Environment* **23**: 365–375.
- Scheidegger AE. **1974**. *The physics of flow through porous media (3rd edition)*. University of Toronto Press.
- Schielzeth H, Nakagawa S. **2013**. Nested by design: Model fitting and interpretation in a mixed model era. *Methods in Ecology and Evolution* **4**: 14–24.
- Schlesinger WH, Jasechko S. **2014**. Transpiration in the global water cycle. *Agricultural and Forest Meteorology* **189–190**: 115–117.
- Schöley J, Kashnitsky I. **2020**. Tricolore: A Flexible Color Scale for Ternary Compositions.
- Schreel JDM, Steppe K. **2018**. Analysis of sap flow dynamics in saplings with mini-HFD (heat field deformation) sensors. In: *Acta horticulturae*. International Society for Horticultural Science (ISHS), Leuven, Belgium, 161–166.
- Schreiber SG, Hacke UG, Chamberland S, Lowe CW, Kamelchuk D, Bräutigam K, Campbell MM, Thomas BR. **2016**. Leaf size serves as a proxy for xylem vulnerability to cavitation in plantation trees: Hydraulic trait variation in plantation trees. *Plant, Cell & Environment* **39**: 272–281.
- Schulze ED, Čermák J, Matyssek M, Penka M, Zimmermann R, Vasícek F, Gries W, Kučera J. **1985**. Canopy transpiration and water fluxes in the xylem of the trunk of Larix and Picea trees — a comparison of xylem flow, porometer and cuvette measurements. *Oecologia* **66**: 475–483.
- Schwalm CR, Anderegg WRL, Michalak AM, Fisher JB, Biondi F, Koch G, Litvak M, Ogle K, Shaw JD, Wolf A *et al.* **2017**. Global patterns of drought recovery. *Nature* **548**: 202–205.
- Sellers PJ. **1985**. Canopy reflectance, photosynthesis and transpiration. *International Journal of Remote Sensing* **6**: 1335–1372.
- Serra-Diaz JM, Enquist BJ, Maitner B, Merow C, Svenning J-C. **2017**. Big data of tree species distributions: How big and how good? *Forest Ecosystems* **4**: 30.
- Shackel KA, Johnson RS, Medawar CK, Phene CJ. **1992**. Substantial errors in estimates of sap flow using the heat balance technique on woody stems under field conditions. *Journal of the American Society for Horticultural Science* **117**: 351–356.
- Shimizu T, Kumagai T, Kobayashi M, Tamai K, Iida S, Kabeya N, Ikawa R, Tateishi M, Miyazawa Y, Shimizu A. **2015**. Estimation of annual forest evapotranspiration from a coniferous planta-

- tion watershed in Japan (2): Comparison of eddy covariance, water budget and sap-flow plus interception loss. *Journal of Hydrology* **522**: 250–264.
- Shimshi D. **1970**. The effect of nitrogen supply on transpiration and stomatal behaviour of beans (*Phaseolus vulgaris L.*). *New Phytologist* **69**: 405–412.
- Shuttleworth WJ. **2007**. Putting the ‘vap’ into evaporation. *Hydrology and Earth System Sciences* **11**: 210–244.
- Siefert A, Violle C, Chalmandrier L, Albert CH, Taudiere A, Fajardo A, Aarssen LW, Baraloto C, Carlucci MB, Cianciaruso MV *et al.* **2015**. A global meta-analysis of the relative extent of intraspecific trait variation in plant communities. *Ecology Letters* **18**: 1406–1419.
- Silvertown J, Araya Y, Gowing D. **2015**. Hydrological niches in terrestrial plant communities: A review. *Journal of Ecology* **103**: 93–108.
- Simard M, Pinto N, Fisher JB, Baccini A. **2011**. Mapping forest canopy height globally with spaceborne lidar. *Journal of Geophysical Research* **116**: G04021.
- Simonin K, Kolb T, Montes-Helu M, Koch G. **2007**. The influence of thinning on components of stand water balance in a ponderosa pine forest stand during and after extreme drought. *Agricultural and Forest Meteorology* **143**: 266–276.
- Simpson WT. **1993**. Specific Gravity , Moisture Content , and Density Relationship for Wood. *Gen Tech Rep FPLGTR76 Madison WI US Department of Agriculture Forest Service Forest Products Laboratory 13 p Gen. Tech.*: 13.
- Smith DM. **1995**. Water Use by Windbreak Trees in the Sahel.
- Smith DM, Allen SJ. **1996**. Measurement of sap flow in plant stems. *Journal of Experimental Botany* **47**: 1833–1844.
- Smith DD, Sperry JS. **2014**. Coordination between water transport capacity, biomass growth, metabolic scaling and species stature in co-occurring shrub and tree species. *Plant, Cell & Environment* **37**: 2679–2690.
- Smith TM, Shugart HH, Bonan GB, Smith JB. **1992**. Modeling the potential response of vegetation to global climate change. In: Begon M, Fitter A, Macfadyen A, eds. *Advances in ecological research. The ecological consequences of global climate change*. Academic Press, 93–116.
- Speckman H, Ewers BE, Beverly DP. **2020**. AquaFlux: Rapid, transparent and replicable analyses of plant transpiration. *Methods in Ecology and Evolution* **11**: 44–50.
- Sperling O, Shapira O, Cohen S, Tripler E, Schwartz A, Lazarovitch N. **2012**. Estimating sap flux densities in date palm trees using the heat dissipation method and weighing lysimeters. *Tree Physiology* **32**: 1171–1178.
- Sperry JS, Hacke UG, Oren R, Comstock JP. **2002**. Water deficits and hydraulic limits to leaf water supply. *Plant, Cell and Environment* **25**: 251–263.
- Sperry JS, Venturas MD, Todd HN, Trugman AT, Anderegg WRL, Wang Y, Tai X. **2019**. The

- impact of rising co₂ and acclimation on the response of us forests to global warming. *Proceedings of the National Academy of Sciences* **116**: 25734–25744.
- Sperry JS, Wang Y, Wolfe BT, Mackay DS, Anderegg WRL, McDowell NG, Pockman WT. **2016**. Pragmatic hydraulic theory predicts stomatal responses to climatic water deficits. *New Phytologist* **212**: 577–589.
- Spracklen DV, Arnold SR, Taylor CM. **2012**. Observations of increased tropical rainfall preceded by air passage over forests. *Nature* **489**: 282–285.
- Staal A, Tuinenburg O, Bosmans J, Holmgren M, van Nes E, Scheffer M, Zemp D, Dekker S. **2018**. Forest-rainfall cascades buffer against drought across the amazon. *Nature Climate Change* **8**: 539–543.
- Steppe K, De Pauw DJW, Doody TM, Teskey RO. **2010**. A comparison of sap flux density using thermal dissipation, heat pulse velocity and heat field deformation methods. *Agricultural and Forest Meteorology* **150**: 1046–1056.
- Steppe K, De Pauw DJW, Lemeur R, Vanrolleghem PA. **2006**. A mathematical model linking tree sap flow dynamics to daily stem diameter fluctuations and radial stem growth. *Tree Physiology* **26**: 257–273.
- Steppe K, Vandegehuchte MW, Tognetti R, Mencuccini M. **2015**. Sap flow as a key trait in the understanding of plant hydraulic functioning. *Tree Physiology* **35**: 341–345.
- Stoy PC, El-Madany TS, Fisher JB, Gentine P, Gerken T, Good SP, Klosterhalfen A, Liu S, Miralles DG, Perez-Priego O *et al.* **2019**. Reviews and syntheses: Turning the challenges of partitioning ecosystem evaporation and transpiration into opportunities. *Biogeosciences* **16**: 3747–3775.
- Suleiman BM, Larfeldt J, Leckner B, Gustavsson M. **1999**. Thermal conductivity and diffusivity of wood. *Wood Science and Technology* **33**: 465–473.
- Sun H, Aubrey DP, Teskey RO. **2012**. A simple calibration improved the accuracy of the thermal dissipation technique for sap flow measurements in juvenile trees of six species. *Trees - Structure and Function* **26**: 631–640.
- Swanson R. **1983**. Numerical and experimental analyses of implanted probe heat pulse velocity theory.
- Swanson RH. **1994**. Significant historical developments in thermal methods for measuring sap flow in trees. *Agricultural and Forest Meteorology* **72**: 113–132.
- Swanson R, Whitfield W. **1981**. A numerical analysis of heat pulse velocity theory. *Journal of Experimental Botany* **32**: 221–239.
- Talsma CJ, Good SP, Jimenez C, Martens B, Fisher JB, Miralles DG, McCabe MF, Purdy AJ. **2018**. Partitioning of evapotranspiration in remote sensing-based models. *Agricultural and*

- Forest Meteorology* **260-261**: 131–143.
- Tan Z-H, Zhao J-F, Wang G-Z, Chen M-P, Yang L-Y, He C-S, Restrepo-Coupe N, Peng S-S, Liu X-Y, Rocha HR da *et al.* **2019**. Surface conductance for evapotranspiration of tropical forests: Calculations, variations, and controls. *Agricultural and Forest Meteorology* **275**: 317–328.
- Taneda H, Sperry JS. **2008**. A case-study of water transport in co-occurring ring- versus diffuse-porous trees: Contrasts in water-status, conducting capacity, cavitation and vessel refilling. *Tree Physiology* **28**: 1641–1651.
- Tardieu F, Zhang J, Gowing DJG. **1993**. Stomatal control by both [aba] in the xylem sap and leaf water status: A test of a model for draughted or aba-fed field-grown maize. *Plant, Cell & Environment* **16**: 413–420.
- Testi L, Villalobos FJ. **2009**. New approach for measuring low sap velocities in trees. *Agricultural and Forest Meteorology* **149**: 730–734.
- Torngern P, Oren R, Oishi AC, Uebelherr JM, Palmroth S, Tarvainen L, Ottosson-Löfvenius M, Linder S, Domec J-C, Näsholm T. **2017**. Ecophysiological variation of transpiration of pine forests: Synthesis of new and published results. *Ecological Applications* **27**: 118–133.
- Torngern P, Oren R, Palmroth S, Novick K, Oishi A, Linder S, Ottosson-Löfvenius M, Näsholm T. **2018**. Water balance of pine forests: Synthesis of new and published results. *Agricultural and Forest Meteorology* **259**: 107–117.
- Trabucco A, Zomer R. **2019**. Global Aridity Index and Potential Evapotranspiration (ET0) Climate Database v2.
- Trugman AT, Anderegg LDL, Shaw JD, Anderegg WRL. **2020**. Trait velocities reveal that mortality has driven widespread coordinated shifts in forest hydraulic trait composition. *Proceedings of the National Academy of Sciences* **117**: 8532–8538.
- Trumbore S, Brando P, Hartmann H. **2015**. Forest health and global change. *Science* **349**: 814–818.
- Tyree MT. **1997**. The cohesion-tension theory of sap ascent: Current controversies. *Journal of Experimental Botany* **48**: 1753–1765.
- Tyree MT, Ewers FW. **1991**. The hydraulic architecture of trees and other woody plants. *New Phytologist* **119**: 345–360.
- Tyree MT, Sperry JS. **1988**. Do woody plants operate near the point of catastrophic xylem dysfunction caused by dynamic water stress?: answers from a model. *Plant Physiology* **88**: 574–580.
- Tyree MT, Zimmermann MH. **2002**. *Xylem Structure and the Ascent of Sap* (TE Timell, Ed.). Berlin, Heidelberg: Springer Berlin Heidelberg.
- Uddling J, Teclaw RM, Pregitzer KS, Ellsworth DS. **2009**. Leaf and canopy conductance in aspen and aspen-birch forests under free-air enrichment of carbon dioxide and ozone. *Tree Physiology*

- 29:** 1367–1380.
- Urban J, Krofta K, Kučera J. **2012**. Calibration of Stem Heat Balance Sensors Upon a Study of Water Balance of the Hop Plantation. *Acta Horticulturae*: 79–86.
- van der Molen M, Dolman A, Ciais P, Eglin T, Gobron N, Law B, Meir P, Peters W, Phillips O, Reichstein M *et al.* **2011**. Drought and ecosystem carbon cycling. *Agricultural and Forest Meteorology* **151**: 765–773.
- Vandegehuchte MW, Steppe K. **2012a**. Interpreting the Heat Field Deformation method: Erroneous use of thermal diffusivity and improved correlation between temperature ratio and sap flux density. *Agricultural and Forest Meteorology* **162–163**: 91–97.
- Vandegehuchte MW, Steppe K. **2012b**. A triple-probe heat-pulse method for measurement of thermal diffusivity in trees. *Agricultural and Forest Meteorology* **160**: 90–99.
- Vandegehuchte MW, Steppe K. **2012c**. Sapflow+: A four-needle heat-pulse sap flow sensor enabling nonempirical sap flux density and water content measurements. *New Phytologist* **196**: 306–317.
- Vandegehuchte MW, Steppe K. **2013**. Sap-flux density measurement methods : working principles and applicability. *Functional Plant Biology*: 213–223.
- Vandegehuchte MW, Burgess SS, Downey A, Steppe K. **2015**. Influence of stem temperature changes on heat pulse sap flux density measurements. *Tree Physiology* **35**: 346–353.
- Vandegehuchte MW, Steppe K, Phillips N. **2012**. Improving sap flux density measurements by correctly determining thermal diffusivity, differentiating between bound and unbound water. *Tree Physiology* **32**: 930–942.
- Vellame LM, Coelho Filho Ma, Paz VPS, Coelho EF. **2010**. Stem heat balance method to estimate transpiration of young orange and mango plants. *Revista Brasileira de Engenharia Agrícola e Ambiental* **14**: 594–599.
- Venturas MD, Sperry JS, Hacke UG. **2017**. Plant xylem hydraulics: What we understand, current research, and future challenges: Plant xylem hydraulics. *Journal of Integrative Plant Biology* **59**: 356–389.
- Vergeynst LL, Vandegehuchte MW, McGuire MA, Teskey RO, Steppe K. **2014**. Changes in stem water content influence sap flux density measurements with thermal dissipation probes. *Trees - Structure and Function* **28**: 949–955.
- Vernay A, Tian X, Chi J, Linder S, Mäkelä A, Oren R, Peichl M, Stangl ZR, Tor-Ngern P, Marshall JD. **2020**. Estimating canopy gross primary production by combining phloem stable isotopes with canopy and mesophyll conductances. *Plant, Cell & Environment* **43**: 2124–2142.
- Vieweg GH, Ziegler H. **1960**. Thermoelektrische registrierung der geschwindigkeit des transpirationsstromes. *Ber Dtsch Bot Ges* **73**: 221–226.
- Vilà-Cabrera A, Martínez-Vilalta J, Retana J. **2015**. Functional trait variation along environ-

- mental gradients in temperate and Mediterranean trees. *Global Ecology and Biogeography* **24**: 1377–1389.
- Vitale D, Fratini G, Bilancia M, Nicolini G, Sabbatini S, Papale D. **2020**. A robust data cleaning procedure for eddy covariance flux measurements. *Biogeosciences* **17**: 1367–1391.
- Vose JM, Miniat CF, Luce CH, Asbjornsen H, Caldwell PV, Campbell JL, Grant GE, Isaak DJ, Loheide SP, Sun G. **2016**. Ecohydrological implications of drought for forests in the united states. *Forest Ecology and Management* **380**: 335–345.
- Vuichard N, Papale D. **2015**. Filling the gaps in meteorological continuous data measured at fluxnet sites with era-interim reanalysis. *Earth System Science Data* **7**: 157–171.
- Walker AP, De Kauwe MG, Bastos A, Belmecheri S, Georgiou K, Keeling RF, McMahon SM, Medlyn BE, Moore DJP, Norby RJ *et al.* **2020**. Integrating the evidence for a terrestrial carbon sink caused by increasing atmospheric co₂. *New Phytologist*.
- Wang K, Dickinson RE. **2012**. A review of global terrestrial evapotranspiration: Observation, modeling, climatology, and climatic variability. *Reviews of Geophysics* **50**.
- Wang Y, Sperry JS, Anderegg WRL, Venturas MD, Trugman AT. **2020**. A theoretical and empirical assessment of stomatal optimization modeling. *New Phytologist* **227**: 311–325.
- Wang-Erlandsson L, Ent RJ van der, Gordon LJ, Savenije HHG. **2014**. Contrasting roles of interception and transpiration in the hydrological cycle – part 1: Temporal characteristics over land. *Earth System Dynamics* **5**: 441–469.
- Ward EJ, Bell DM, Clark JS, Oren R. **2013**. Hydraulic time constants for transpiration of loblolly pine at a free-air carbon dioxide enrichment site. *Tree Physiology* **33**: 123–134.
- Ward EJ, Domec J-C, King J, Sun G, McNulty S, Noormets A. **2017**. TRACC: An open source software for processing sap flux data from thermal dissipation probes. *Trees* **31**.
- Wei Z, Yoshimura K, Wang L, Miralles DG, Jasechko S, Lee X. **2017**. Revisiting the contribution of transpiration to global terrestrial evapotranspiration. *Geophysical Research Letters* **44**: 2792–2801.
- Wheeler EA. **2011**. Inside Wood – A Web resource for hardwood anatomy. *IAWA Journal* **32**: 199–211.
- Whitehead D. **1998**. Regulation of stomatal conductance and transpiration in forest canopies. *Tree Physiology* **18**: 633–644.
- Whitley R, Taylor D, Macinnis-Ng C, Zeppel M, Yunusa I, O’Grady A, Froend R, Medlyn B, Eamus D. **2013**. Developing an empirical model of canopy water flux describing the common response of transpiration to solar radiation and vpd across five contrasting woodlands and forests. *Hydrological Processes* **27**: 1133–1146.
- Wiedemann A, Jiménez SM, Rebman C, Cuntz M, Herbst M. **2013**. Empirical study of wound

- response dynamics on sap flow measured with thermal dissipation probes.
- Wiedemann A, Marañón-Jiménez S, Rebmann C, Herbst M, Cuntz M, Walther BA, Moore JL, Rahbek C. **2016**. An empirical study of the wound effect on sap flux density measured with thermal dissipation probes. *Tree Physiology* **36**: 1471–1484.
- Wild M, Folini D, Hakuba MZ, Schär C, Seneviratne SI, Kato S, Rutan D, Amman C, Wood EF, König-Langlo G. **2015**. The energy balance over land and oceans: An assessment based on direct observations and cmip5 climate models. *Climate Dynamics* **44**: 3393–3429.
- Williams M, Bond BJ, Ryan MG. **2001**. Evaluating different soil and plant hydraulic constraints on tree function using a model and sap flow data from ponderosa pine. *Plant, Cell & Environment* **24**: 679–690.
- Williams CA, Reichstein M, Buchmann N, Baldocchi D, Beer C, Schwalm C, Wohlfahrt G, Hasler N, Bernhofer C, Foken T *et al.* **2012**. Climate and vegetation controls on the surface water balance: Synthesis of evapotranspiration measured across a global network of flux towers: Climate and vegetation controls on surface water balance. *Water Resources Research* **48**.
- Wilson KB, Hanson PJ, Mulholland PJ, Baldocchi DD, Wullschleger SD. **2001**. A comparison of methods for determining forest evapotranspiration and its components: Sap-flow, soil water budget, eddy covariance and catchment water balance. *Agricultural and Forest Meteorology* **106**: 153–168.
- Wood SN. **2011**. Fast stable restricted maximum likelihood and marginal likelihood estimation of semiparametric generalized linear models: Estimation of Semiparametric Generalized Linear Models. *Journal of the Royal Statistical Society: Series B (Statistical Methodology)* **73**: 3–36.
- Wright IJ, Dong N, Maire V, Prentice IC, Westoby M, Díaz S, Gallagher RV, Jacobs BF, Kooyman R, Law EA *et al.* **2017**. Global climatic drivers of leaf size. *Science* **357**: 917–921.
- Wullschleger SD, Childs KW, King AW, Hanson PJ. **2011**. A model of heat transfer in sapwood and implications for sap flux density measurements using thermal dissipation probes. *Tree Physiology* **31**: 669–679.
- Wullschleger SD, Epstein HE, Box EO, Euskirchen ES, Goswami S, Iversen CM, Kattge J, Norby RJ, Bodegom PM van, Xu X. **2014**. Plant functional types in Earth system models: past experiences and future directions for application of dynamic vegetation models in high-latitude ecosystems. *Annals of Botany* **114**: 1–16.
- Wullschleger SD, Meinzer FC, Vertessy RA. **1998**. A review of whole-plant water use studies in tree. *Tree Physiology* **18**: 499–512.
- Wutzler T, Lucas-Moffat A, Migliavacca M, Knauer J, Sickel K, Šigut L, Menzer O, Reichstein M. **2018**. Basic and extensible post-processing of eddy covariance flux data with redyproc. *Biogeosciences* **15**: 5015–5030.
- Xie J, Wan X. **2018**. The accuracy of the thermal dissipation technique for estimating sap flow is affected by the radial distribution of conduit diameter and density. *Acta Physiologiae Plantarum*

tarum.

- Yang Y, Zhu Q, Peng C, Wang H, Chen H. **2015**. From plant functional types to plant functional traits: A new paradigm in modelling global vegetation dynamics. *Progress in Physical Geography: Earth and Environment* **39**: 514–535.
- Yin J, Bauerle TL. **2017**. A global analysis of plant recovery performance from water stress. *Oikos* **126**: 1377–1388.
- Yuan W, Zheng Y, Piao S, Ciais P, Lombardozzi D, Wang Y, Ryu Y, Chen G, Dong W, Hu Z *et al.* **2019**. Increased atmospheric vapor pressure deficit reduces global vegetation growth. *Science Advances* **5**: eaax1396.
- Zanne AE, Westoby M, Falster DS, Ackerly DD, Loarie SR, Arnold SE, Coomes DA. **2010**. Angiosperm wood structure: Global patterns in vessel anatomy and their relation to wood density and potential conductivity. *American Journal of Botany* **97**: 207–215.
- Zeppel M, Lewis J, Phillips N, Tissue D. **2014**. Consequences of nocturnal water loss: a synthesis of regulating factors and implications for capacitance, embolism and use in models. *Tree Physiology* **34**: 1047–1055.
- Zhang Q, Manzoni S, Katul G, Porporato A, Yang D. **2014**. The hysteretic evapotranspiration—Vapor pressure deficit relation. *Journal of Geophysical Research: Biogeosciences* **119**: 125–140.
- Zhang Y, Oren R, Kang S. **2012**. Spatiotemporal variation of crown-scale stomatal conductance in an arid *Vitis vinifera* L. cv. Merlot vineyard: Direct effects of hydraulic properties and indirect effects of canopy leaf area. *Tree Physiology* **32**: 262–279.
- Zhang ZZ, Zhao P, McCarthy HR, Zhao XH, Niu JF, Zhu LW, Ni GY, Ouyang L, Huang YQ. **2016**. Influence of the decoupling degree on the estimation of canopy stomatal conductance for two broadleaf tree species. *Agricultural and Forest Meteorology* **221**: 230–241.
- Zhao WL, Gentine P, Reichstein M, Zhang Y, Zhou S, Wen Y, Lin C, Li X, Qiu GY. **2019a**. Physics-Constrained Machine Learning of Evapotranspiration. *Geophysical Research Letters* **46**: 14496–14507.
- Zhao S, Pederson N, D'Orangeville L, HilleRisLambers J, Boose E, Penone C, Bauer B, Jiang Y, Manzanedo RD. **2019b**. The international tree-ring data bank (itrdb) revisited: Data availability and global ecological representativity. *Journal of Biogeography* **46**: 355–368.
- Zhou S, Williams AP, Berg AM, Cook BI, Zhang Y, Hagemann S, Lorenz R, Seneviratne SI, Gentine P. **2019a**. Land–atmosphere feedbacks exacerbate concurrent soil drought and atmospheric aridity. *Proceedings of the National Academy of Sciences* **116**: 18848–18853.
- Zhou S, Zhang Y, Park Williams A, Gentine P. **2019b**. Projected increases in intensity, frequency, and terrestrial carbon costs of compound drought and aridity events. *Science Advances* **5**: eaau5740.
- Zweifel R, Steppe K, Sterck FJ. **2007**. Stomatal regulation by microclimate and tree water relations: interpreting ecophysiological field data with a hydraulic plant model. *Journal of*

Experimental Botany **58**: 2113–2131.