6th June, 2018:

→ Passaging of P6 cells -

↳ Cells are at 100% confluency

↳ One 60mm plate trypsinized first - trypsin wash, followed by treatment - both with 500µL 0.25% trypsin - EDTA [aliquot from Rutika].

↳ After 1 min incubation, trypsin was inhibited with an equal amount of DMEM - FBS - we have used about 5mL of the mix aliquot as we only have 15mL Falcon tubes, but 1.5mL Eppendorf tubes would be ideal for these volumes (<1mL).

↳ Cells are ~~fa~~ ~~pelted~~ ~~pelleted~~ spun down in about 6mL of DMEM : FBS (pooled from both 60mm dishes).

↳ This pooled set has split between 2 100mm dishes and one frozen stock.

↳ Proportion of components are roughly 1:2 from 60mm to 100mm plates - about 12-15mL of media and ~1mL of cell suspension.

↳ In this batch, pen-strep [aliquot from Rutika] was added directly to the dishes as we forgot to add to the DMEM: FBS aliquot - 200µL in each dish, and in the remaining media aliquot.

↳ Proportions for 1mL stock - 700µL media + 200µL cells + 100µL DMSO.

↳ Finally: 2 100mm dishes of ~~100mm~~ HEK293 - P7
& 1 cryovial of 1mL stock of HEK293-P6.

→ No passaging is expected tomorrow.