Transmission-blocking activity of artesunate, chloroquine and methylene blue on Plasmodium vivax gametocytes

Victor Chaumeau & James A. Watson

2024-02-14

Contents

| Foreword | 1 |
|---|----------------------------|
| Session information | 2 |
| Summary of assay runs | 3 |
| Baseline characteristics of the samples Asexual parasitaemia Gametocytaemia Oocyst index Oocyst count Sporozoite index Sporozoite count | 3 3 3 4 4 4 |
| Evolution of parasite count during incubation ex vivo | 5 6 |
| Drug effects on parasite count and parasite index | 6 |
| Look at model coefficient etimates | 7 |
| Assay variability Inter-experiment variability | 9 9 10 |
| Correlation between oocyst and sporozoite count | 11 |
| Model assesment Traceplots | 13 13 15 |

Foreword

This analysis was carried out by Dr. Victor Chaumeau* under the supervision of Dr. James A. Waston, who wrote the Stan code for the Bayesian multi-level model used in this study.

^{*} For correspondence: victor@shoklo-unit.com

Session information

```
## R version 4.2.2 Patched (2022-11-10 r83330)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Debian GNU/Linux 12 (bookworm)
## Matrix products: default
           /usr/lib/x86_64-linux-gnu/atlas/libblas.so.3.10.3
## LAPACK: /usr/lib/x86 64-linux-gnu/atlas/liblapack.so.3.10.3
##
## locale:
  [1] LC CTYPE=en US.UTF-8
                                   LC NUMERIC=C
  [3] LC_TIME=en_US.UTF-8
                                   LC_COLLATE=en_US.UTF-8
   [5] LC_MONETARY=en_US.UTF-8
                                   LC_MESSAGES=en_US.UTF-8
## [7] LC_PAPER=en_US.UTF-8
                                   LC NAME=C
## [9] LC ADDRESS=C
                                   LC TELEPHONE=C
## [11] LC_MEASUREMENT=en_US.UTF-8 LC_IDENTIFICATION=C
##
## attached base packages:
## [1] stats
                 graphics grDevices utils
                                                datasets methods
                                                                    base
## other attached packages:
                                               forcats_1.0.0
## [1] lme4_1.1-35.1
                           Matrix_1.6-5
                                                                  stringr_1.5.1
  [5] dplyr_1.1.4
                           purrr_1.0.2
                                               readr_2.1.5
                                                                  tibble_3.2.1
                                               EnvStats_2.8.1
## [9] ggplot2_3.4.4
                           tidyverse_1.3.2
                                                                  scales_1.3.0
                           rstan 2.32.5
                                               StanHeaders_2.32.5 tidyr_1.3.1
## [13] plotrix_3.8-2
## [17] extrafont_0.19
                           RColorBrewer 1.1-3
## loaded via a namespace (and not attached):
## [1] httr_1.4.5
                            splines_4.2.2
                                                 jsonlite_1.8.8
## [4] QuickJSR_1.1.3
                            modelr_0.1.11
                                                 RcppParallel_5.1.7
## [7] stats4_4.2.2
                            googlesheets4_1.0.1 cellranger_1.1.0
## [10] yaml_2.3.8
                            lattice_0.22-5
                                                 Rttf2pt1_1.3.12
## [13] pillar_1.9.0
                            backports_1.4.1
                                                 glue_1.7.0
## [16] extrafontdb_1.0
                            digest_0.6.34
                                                 minqa_1.2.6
                                                 htmltools_0.5.7
## [19] rvest_1.0.3
                            colorspace_2.1-0
## [22] pkgconfig_2.0.3
                            broom_1.0.5
                                                 haven_2.5.4
## [25] tzdb_0.4.0
                            timechange_0.3.0
                                                 googledrive_2.0.0
## [28] generics_0.1.3
                            withr_3.0.0
                                                 cli_3.6.2
## [31] magrittr_2.0.3
                            crayon_1.5.2
                                                 readxl_1.4.3
## [34] evaluate_0.23
                            fs_1.6.3
                                                 fansi_1.0.6
## [37] nlme_3.1-162
                            MASS_7.3-60.0.1
                                                 xm12_1.3.3
                            tools_4.2.2
## [40] pkgbuild_1.4.3
                                                 100_2.6.0
## [43] hms 1.1.3
                            gargle 1.3.0
                                                 lifecycle 1.0.4
## [46] matrixStats_1.2.0
                            V8_4.2.2
                                                 munsell_0.5.0
## [49] reprex 2.1.0
                            compiler_4.2.2
                                                 rlang 1.1.3
## [52] nloptr_2.0.3
                            grid_4.2.2
                                                 rstudioapi_0.15.0
## [55] rmarkdown_2.25
                            boot_1.3-28.1
                                                 gtable_0.3.4
## [58] codetools_0.2-19
                                                 DBI_1.2.1
                            inline_0.3.19
## [61] curl_5.0.0
                            R6_2.5.1
                                                 gridExtra_2.3
## [64] lubridate_1.9.3
                            knitr_1.45
                                                 fastmap_1.1.1
## [67] utf8_1.2.4
                            stringi_1.8.3
                                                 parallel_4.2.2
## [70] Rcpp_1.0.12
                            vctrs_0.6.5
                                                 dbplyr_2.4.0
## [73] tidyselect_1.2.0
                            xfun_0.41
```

Summary of assay runs

Table 1: Table 1. Summary of assay runs

| | | | | | | | | No. dis- | No. dis- | No. dis- |
|----|--------|--------|-----------------------|----------|--------------------|----------|----------|--------------|--------------------------|--------------|
| | | | | | No. dis- | No. dis- | No. dis- | sected pairs | sected pairs | sected pairs |
| N | lo. of | | | | sected | sected | sected | of salivary | of salivary | of salivary |
| b | lood | No. ba | sNo. co | on- | guts in | guts in | guts in | glands in | glands in | glands in |
| S | am- | line | trol | No. tre | a bad eline | control | treated | baseline | $\operatorname{control}$ | treated |
|] | ples | batche | esbatch | esbatche | s bacthes | bacthes | bacthes | batches | batches | batches |
| AS | 9 | 9 | 36 | 36 | 450 | 1797 | 1798 | 90 | 360 | 360 |
| CQ | 21 | 21 | 84 | 84 | 1008 | 4026 | 4036 | 263 | 1228 | 1177 |
| MB | 8 | 8 | 32 | 32 | 404 | 1592 | 1599 | 80 | 320 | 320 |
| | | | | | | | | | | |

38 vivax malaria patients took part in the study and provided a blood sample before receiving antimalarial drug regimen, 342 mosquito batches were fed on these samples and 20908 specimens were dissected for assessment of either oocyst or sporozoite count.

Baseline characteristics of the samples

Asexual parasitaemia

```
quantile(dta$AsexualParasitaemia, probs = c(0.5, 0.25, 0.75))
##
         50%
                   25%
                              75%
## 13160.700 6980.831 27798.019
aggregate(AsexualParasitaemia~Drug, FUN = quantile, probs = c(0.5,0.25,0.75), data = dta)
     Drug AsexualParasitaemia.50% AsexualParasitaemia.25% AsexualParasitaemia.75%
## 1
       AS
                         12108.60
                                                   8257.90
                                                                           16608.38
## 2
       CQ
                         12740.00
                                                   3610.45
                                                                           21652.31
## 3
       MB
                         17717.60
                                                  10116.38
                                                                           28675.35
```

Gametocytaemia

```
quantile(dta$Gametocytaemia, probs = c(0.5,0.25,0.75))
                   25%
                             75%
         50%
## 1091.5500
             473.2875 2009.2125
aggregate(Gametocytaemia~Drug, FUN = quantile, probs = c(0.5,0.25,0.75), data = dta)
     Drug Gametocytaemia.50% Gametocytaemia.25% Gametocytaemia.75%
##
## 1
       AS
                     766.700
                                         524.475
                                                            1350.250
## 2
       CQ
                     897.750
                                         195.000
                                                            1849.175
## 3
       MB
                    1821.050
                                        1308.666
                                                            4062.525
```

Oocyst index

0.96 0.84 0.98

```
quantile(dta_agg$`NbOocysts > 0`, probs = c(0.5,0.25,0.75))
## 50% 25% 75%
```

```
aggregate(`NbOocysts > 0`~Drug, FUN = quantile, probs = c(0.5,0.25,0.75), data = dta_agg)
     Drug NbOocysts > 0.50% NbOocysts > 0.25% NbOocysts > 0.75%
##
## 1
       AS
                  0.9800000
                                     0.9400000
                                                       0.9800000
## 2
       CQ
                  0.9591837
                                     0.6800000
                                                       0.9800000
## 3
       MB
                  0.9600000
                                     0.9158824
                                                       0.9850000
Oocyst count
##
       50%
               25%
                       75%
##
    63.750
             4.125 124.875
     Drug NbOocysts.50% NbOocysts.25% NbOocysts.75%
##
                 95.000
                                48.500
                                             127.000
## 1
       AS
       CQ
                 18.500
                                 2.000
                                              92.000
## 2
##
       MB
                114.500
                                54.625
                                             183.875
Sporozoite index
quantile(dta_agg$`NbSporozoites2 > 0`, probs = c(0.5,0.25,0.75))
    50% 25% 75%
## 0.70 0.35 0.90
aggregate(`NbSporozoites2 > 0`~Drug, FUN = quantile, probs = c(0.5,0.25,0.75), data = dta_agg)
##
     Drug NbSporozoites2 > 0.50% NbSporozoites2 > 0.25% NbSporozoites2 > 0.75%
## 1
       AS
                         0.60000
                                                 0.10000
                                                                         0.70000
## 2
       CQ
                         0.65000
                                                 0.27500
                                                                         0.85625
## 3
       MB
                         0.95000
                                                                         1.00000
                                                 0.87500
Sporozoite count
quantile(dta agg$NbSporozoites2, probs = c(0.5,0.25,0.75))
    50%
         25%
##
             75%
##
    211
           5 4321
aggregate(NbSporozoites2~Drug, FUN = quantile, probs = c(0.5,0.25,0.75), data = dta_agg)
##
     Drug NbSporozoites2.50% NbSporozoites2.25% NbSporozoites2.75%
## 1
       AS
                        33.0
                                             0.0
                                                              211.0
## 2
       CQ
                       122.0
                                             2.5
                                                              2377.5
```

Overall, the median asexual parasitaemia was 13161 parasites /uL (inter-quartile range, IQR: 6981 to 27798) and the median gametocytaemia was 1092 gametocytes /uL (IQR: 473 to 2009). All blood samples but one were infectious to mosquitoes (97.4%, the sample that was not infectious during baseline became infectious after 24 hours of incubation). The median oocyst index (i.e., the proportion of mosquitoes harboring malaria oocysts) and oocyst count in mosquito samples were 0.96 (IQR: 0.84 to 0.98) and 63.8 oocysts per mosquito (IQR: 4.1 to 124.9). The corresponding figures for the sporozoite stage were 0.7 (IQR: 0.35 to 0.9) and 211 sporozoites per mosquito (IQR: 5 to 4321).

2402.5

10876.0

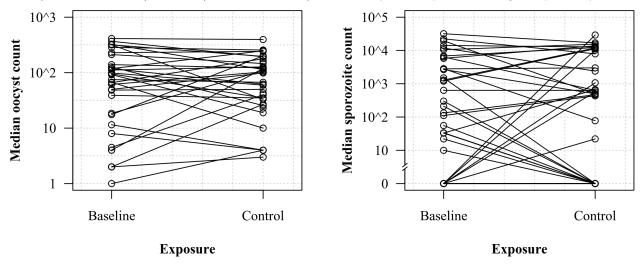
4604.5

3

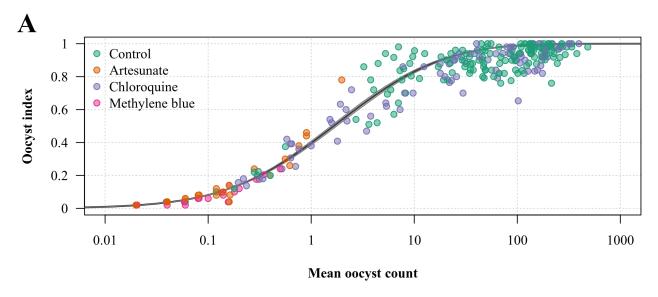
MB

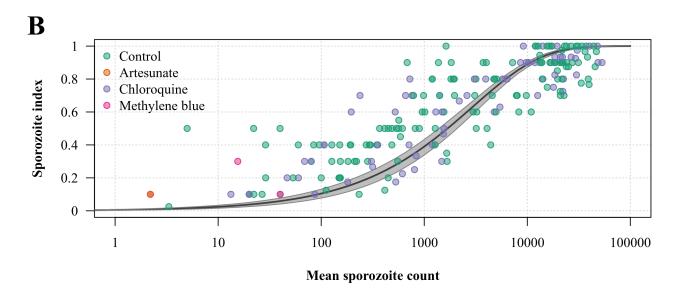
Evolution of parasite count during incubation ex vivo

The median ratio of the median parasite count in the controls to the median parasite count at baseline was 0.93 (IQR: 0.51 to 3.68) and 0.9 (IQR: 0.01 to 6.56) for the oocyst and sporozoite stage, respectively.



Prevalence estimates

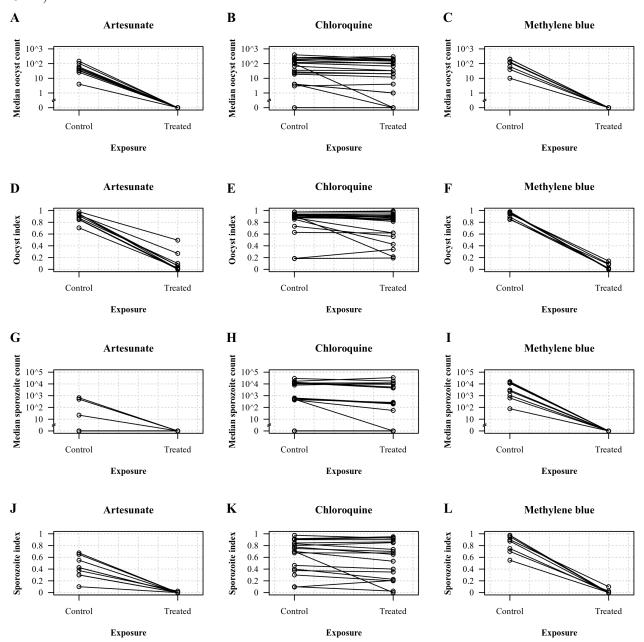




Drug effects on parasite count and parasite index

Chloroquine exhibited little transmission-blocking activity on P. vivax gametocyte transmission, despite the high concentrations used (Figure 1). Of all dissected mosquitoes in the chloroquine spiked samples, 2974/4036 (73.69%) carried oocysts in the treated replicates compared with 3299/4026 (81.94%) in the controls (relative risk, RR: 0.85 [95%CI 0.7 to 0.98], $\log 10$ [p-value] = -24.73), and 701/1177 (59.56%) carried sporozoites in the treated replicates compared with 785/1228 (63.93%) in the controls (RR: 0.89 [95%CI 0.86 to 0.92], $\log 10$ [p-value] = -2.3). In contrast, artesunate and methylene blue almost completely interrupted gametocyte transmission. For artesunate, only 207/1798 (11.51%) of the mosquitoes carried oocysts in the treated replicates compared with 1591/1797 (88.54%) in the controls (RR: 0.04 [95%CI 4e-04 to 0.94], $\log 10$ [p-value] = -258.52) and 1/360 (0.28%) dissected mosquitoes carried sporozoites in the treated replicates versus 152/360 (42.22%) in the controls (RR: 0.0078 [95%CI 2e-04 to 0.31], $\log 10$ [p-value] = -8.14); for methylene blue, only 76/1599 (4.75%) carried oocysts in the treated replicates versus 1470/1592 (92.34%) in the controls (RR:

0.017 [95%CI 1e-04 to 0.93], $\log 10$ [p-value] = -255.89) and only 5/320 (1.56%) carried sporozoites in the treated replicates versus 267/320 (83.44%) in the controls (RR: 0.0045 [95%CI 2e-05 to 0.6], $\log 10$ [p-value] = -32.12).



Look at model coefficient etimates

As observed previously, there was considerable heterogeneity in the count data across mosquitos and a considerable variability in the count data across blood samples and experimental batches. To account for this heterogeneity and variability, we estimated the drug effects under a Bayesian hierarchical model (mixed effects) whereby the count data are modelled as negative binomial with the dispersion parameter as a parametric function of the mean count (Table 2; see Methods) (20). In contrast to the previous data describing proportions of mosquitos with parasites, the model parameterizes the drug effect as a reduction in the mean number of parasites per mosquito, accounting for variability across blood samples and mosquito

batches.

Table 2: Parameter estimates given by the output of transmissionblocking activity model fitted to oocyst and sporozoite data

| | $Oocyst_Median$ | Oocyst_CI | ${\bf Sporozoite_Median}$ | Sporozoite_CI |
|------------------------|------------------|--------------------|----------------------------|--------------------|
| mu_population | 44.6000 | 31.3 to 63.7 | 4906.6000 | 2875 to 8158.8 |
| sigma_patient | 0.9200 | 0.74 to 1.15 | 1.2200 | 0.98 to 1.54 |
| sigma_batch | 0.4600 | 0.41 to 0.53 | 0.3100 | 0.23 to 0.41 |
| a0 | 0.4091 | 0.3719 to 0.4469 | 0.0007 | 4e-04 to 0.0012 |
| a1 | 0.1865 | 0.1637 to 0.2099 | 0.6103 | 0.5556 to 0.6826 |
| artesunate_effect | 0.0021 | 0.0015 to 0.0029 | 0.0068 | 0.0021 to 0.0165 |
| chloroquine_effect | 0.7142 | 0.606 to 0.8359 | 0.7444 | 0.6032 to 0.8951 |
| methylene_blue_effect | 0.0007 | 5e-04 to 0.001 | 0.0019 | 8e-04 to 0.0041 |
| baseline_oocyst_count | 1.8000 | 1.2 to 2.5 | 1.2000 | 0.7 to 2 |
| baseline_parasitemia | 1.1000 | 0.6 to 2.1 | 1.2000 | 0.5 to 3 |
| baseline_gametocytemia | 6.5000 | 3 to 13.4 | 3.6000 | 1.2 to 9.2 |
| wash | 0.8000 | 0.4 to 1.7 | 0.3000 | 0.1 to 0.6 |

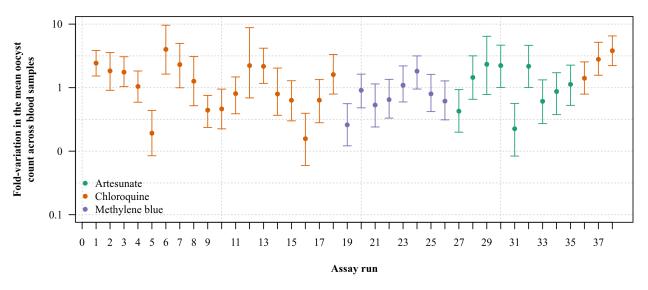
Under the model, gametocyte exposure to chloroquine decreased the mean oocyst count 1.4-fold [95% credible interval [CrI]: 1.65 to 1.2-fold] (from 100.33 to 69.44 oocysts per mosquito in the controls and treated replicates, respectively) and the mean sporozoite count 1.34-fold [95%CrI 1.66 to 1.12-fold] (from 14414.5 to 11131.9 sporozoites per mosquito in the controls and treated replicates, respectively). The corresponding figures for artesunate and methylene blue were a 469.42-fold reduction [95%CrI 650.45 to 344.73-fold] (from 60.56 to 0.22 oocysts per mosquito in the controls and treated replicates, respectively) and a 1437.88-fold reduction [95%CI 2063.86 to 969.55-fold] (from 107.25 to 0.08 oocysts per mosquito in the controls and treated replicates, respectively) in oocyst count respectively and a 147.77-fold [95%CrI 469.56 to 60.64-fold] (from 1303.3 to 0.1 sporozoites per mosquito in the controls and treated replicates, respectively) and a 536.1-fold [95%CrI 1310.51 to 245.99-fold] (from 13914.5 to 1.8 sporozoites per mosquito in the controls and treated replicates, respectively) reduction in sporozoite count, respectively.

To consider baseline variations in blood meal infectiousness to mosquitoes across blood samples, the log10oocyst count, log10asexual parasitaemia and log10gametocytaemia assessed on admission (i.e., on the collection day before the 24-hour incubation time with or without drug) were introduced as linear predictors of the mean parasite count in mosquito samples of the experimental replicates (i.e. after 24 hours of incubation with or without drug). A 10-fold increase in the mean oocyst count and gametocytaemia at baseline were respectively associated with a 1.76 [95%CI: 1.2 to 2.48] and a 6.47-fold increase [95%CI: 3.04 to 13.37] in the mean oocyst count in the experimental replicates; there was no significant association between the mean oocyst count in experimental replicate and baseline asexual parasitaemia (model coefficient: 1.08 [95%CI 0.56 to 2.11]) or artesunate wash off (model coefficient: 0.84 [95%CI 0.41 to 1.71]). A 10-fold increase in baseline gametocytaemia and artesunate wash off were respectively associated with a 3.56-fold increase [95%CI 1.21 to 9.2] and a 0.26-fold decrease [95%CI 0.1 to 0.61] in the mean sporozoite count in the experimental replicates; there was no significant association between the mean sporozoite count in the experimental replicate and mean oocyst count (model coefficient: 1.21 [95%CI 0.72 to 1.96]) or asexual parasitaemia (model coefficient: 1.22 [95%CI 0.48 to 2.96]) at baseline.

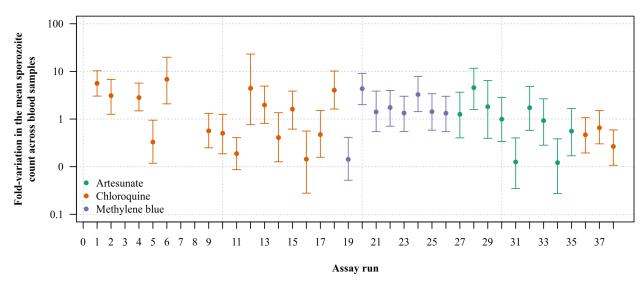
Assay variability

Inter-experiment variability

\mathbf{A}



B



These are the figures for inter-experiment variability (in fold-variation in the mean parasite count across blood samples):

```
quantile(my_table$median.oocyst)

## 0% 25% 50% 75% 100%

## 0.1569302 0.6182925 1.0669434 2.0812507 4.0001822

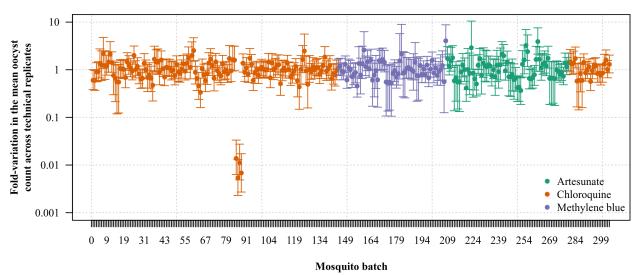
quantile(my_table$median.sporozoite, na.rm = T)

## 0% 25% 50% 75% 100%

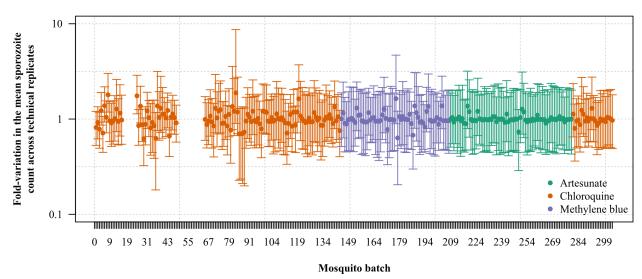
## 0.1215025 0.4695308 1.3234454 2.3978970 6.8286956
```

Intra-experiment variability





B



These are the figures for intra-experiment variability (in fold-variation in the mean parasite count accross technical replicates):

```
quantile(my_table$median.oocyst)

## 0% 25% 50% 75% 100%

## 0.005421552 0.768113629 1.001994206 1.298951808 4.045166536

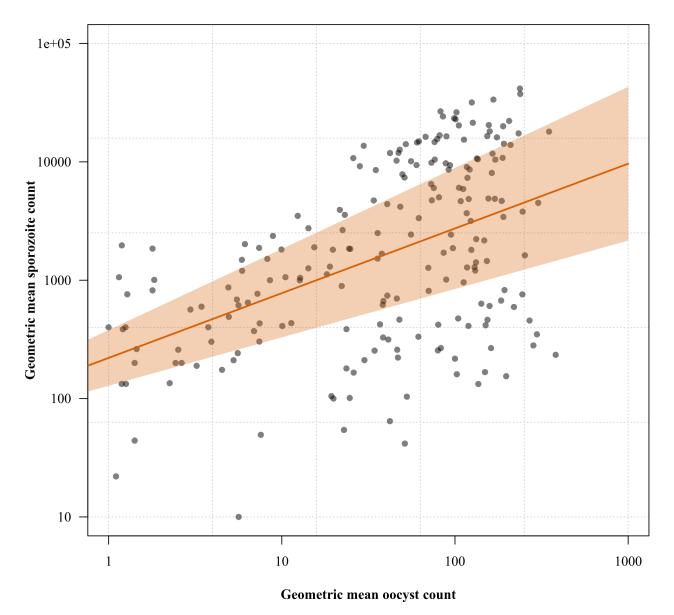
quantile(my_table$median.sporozoite, na.rm = T)

## 0% 25% 50% 75% 100%

## 0.6208217 0.9386475 0.9875390 1.0514436 1.8824427
```

Correlation between oocyst and sporozoite count

```
##
## Call:
## lm(formula = yval ~ xval)
## Residuals:
      Min 1Q Median 3Q
##
## -1.75436 -0.41638 0.08999 0.52068 1.03513
## Coefficients:
            Estimate Std. Error t value Pr(>|t|)
## (Intercept) 2.34347 0.11931 19.642 < 2e-16 ***
             0.54698
                       0.07005 7.809 2.98e-13 ***
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.05 '.' 0.1 ' ' 1
\mbox{\tt \#\#} Residual standard error: 0.6681 on 204 degrees of freedom
## Multiple R-squared: 0.2301, Adjusted R-squared: 0.2263
## F-statistic: 60.98 on 1 and 204 DF, p-value: 2.979e-13
```

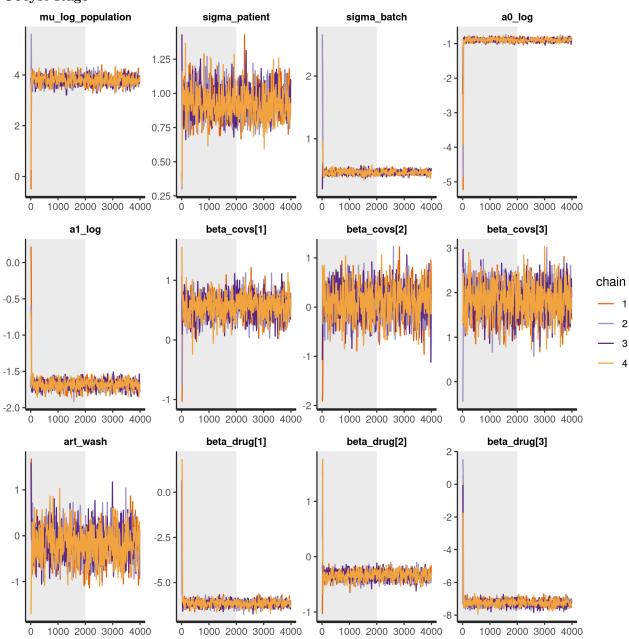


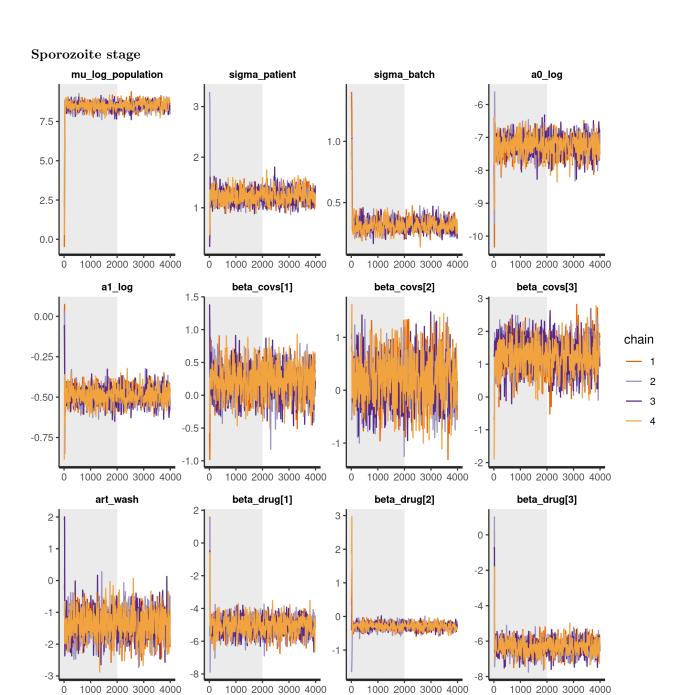
The fit line equation is y = 0.5469815*x + 2.3434715.

Model assesment

Traceplots

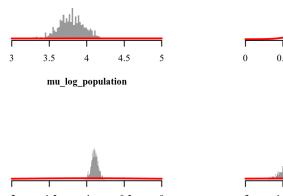


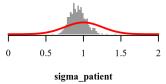


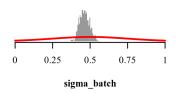


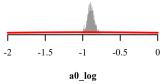
Comparison of priors and posteriors

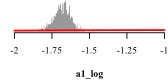
Oocyst stage

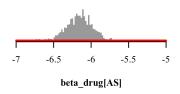


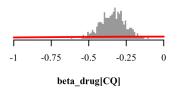


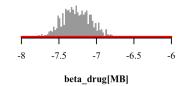


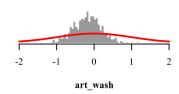


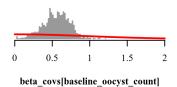


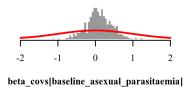


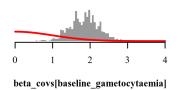












Sporozoite stage

