

MPXV ONT Galaxy Workflow


—
Presenter name

Objectives

- Import and run the MPXV (Mpox) ONT Galaxy workflow
- Understand the outputs of the pipeline
- Download and save the consensus genomes
- Perform quality control (QC) on the consensus genomes

The workflow

edited about 1 month ago

MPXV (Mpox) Analysis Pipeline for ONT 

2025-01-29T23:42:10.546Z

MPXV (Mpox) sequence analysis pipeline for ONT data using the ARTIC minion pipeline (v.1.6.0).

ONT ✕

ARTIC ✕

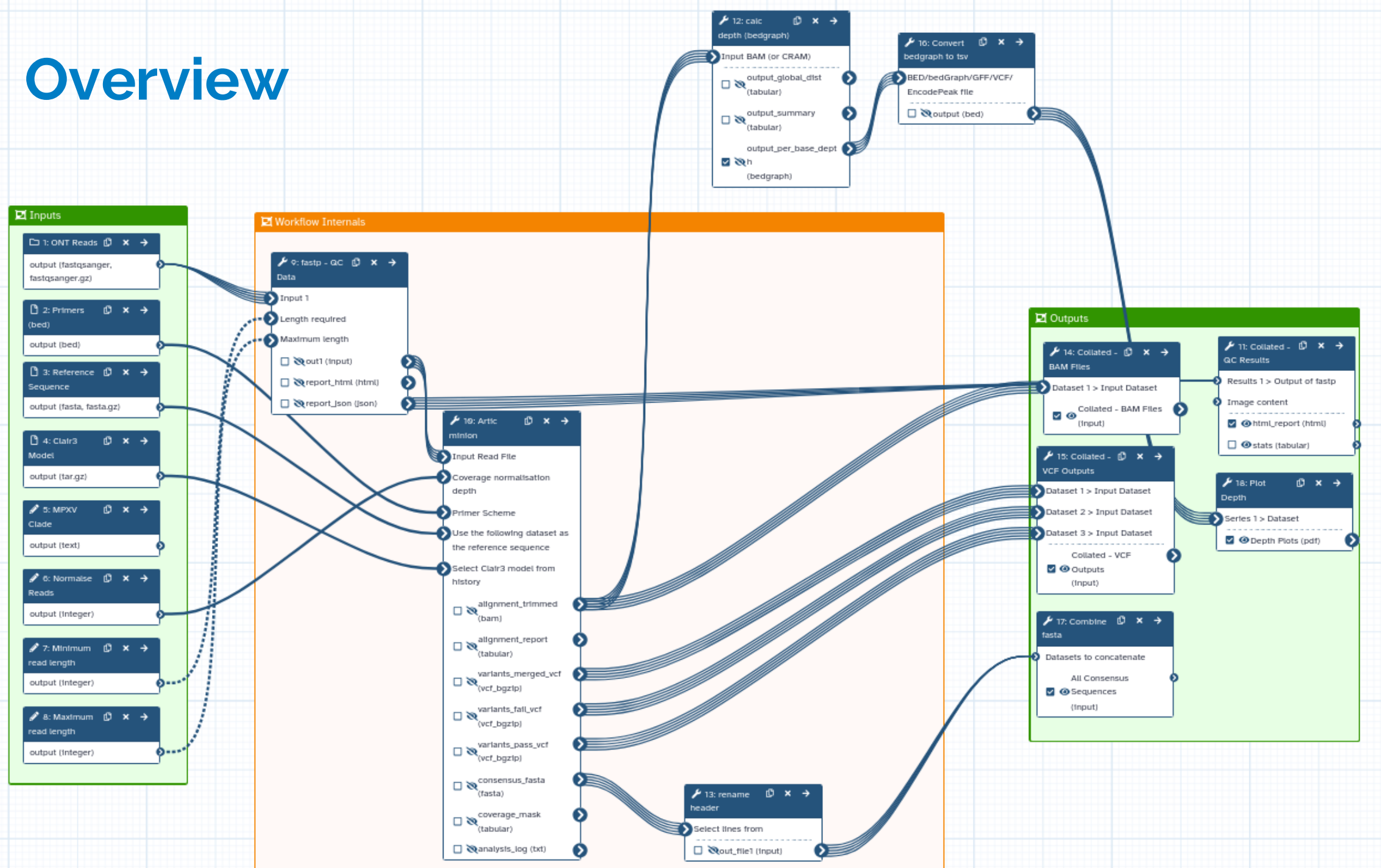
MPXV ✕

Mpox ✕

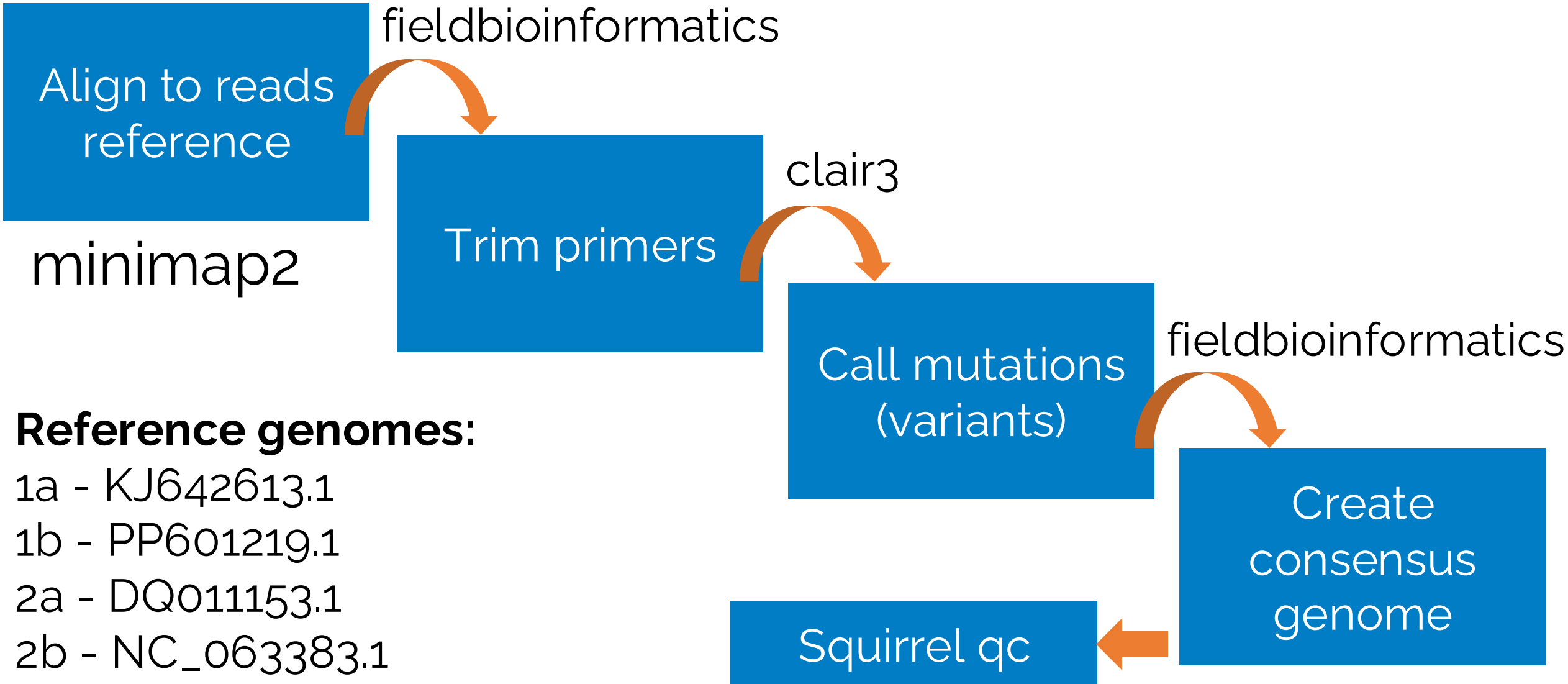
Add Tags 

- Design for ONT platform
- Performed genome mapping, variant calling and consensus assembly

Overview



Pipeline Steps - simplified



Let's run the pipeline!



Let's run the pipeline! ... not yet

Pipeline Inputs



Let's run the pipeline! ... not yet

Workflow inputs for MPXV pipeline:

- Reads (fastq.gz)
- Primer file (tab)
- Reference (fasta)
- Clair3 model (zip)
- Specify Clade (Clade1/2)

Pipeline inputs – fastq files

Workflow inputs for MPXV pipeline:

- ☒ Reads (fastq.gz)
- ☐ Primer file (tab)
- ☐ Reference (fasta)
- ☐ Clair3 model (zip)
- ☐ Specify Clade (Clade1/2)

Pipeline inputs – primer/reference files

Workflow inputs for MPXV pipeline:



Reads (fastq.gz)



Primer file (tab)

Get Primers from



Reference (fasta)

<https://labs.primalscheme.com>



Clair3 model (zip)



Specify Clade (Clade1/2)

Pipeline inputs – primer/reference files

Workflow inputs for MPXV pipeline:

- ✓ Reads (fastq.gz)
- ✓ Primer file (tab) **Get Primers from**
- ✓ Reference (fasta) <https://labs.primalscheme.com>

IMPORTANT! Ensure you download the fasta/primers of the same scheme.

Do not mix different schemes together!

Pipeline inputs – primer/reference files

<https://labs.primalscheme.com/detail/artic-inrb-mpox/2500/v1.0.0/>

artic-inrb-mpox / 2500 / v1.0.0

primer.bed



download

Toggle to show

reference.fasta



download

Toggle to show

Let's run the pipeline – clair3 models

Workflow inputs for MPXV pipeline:

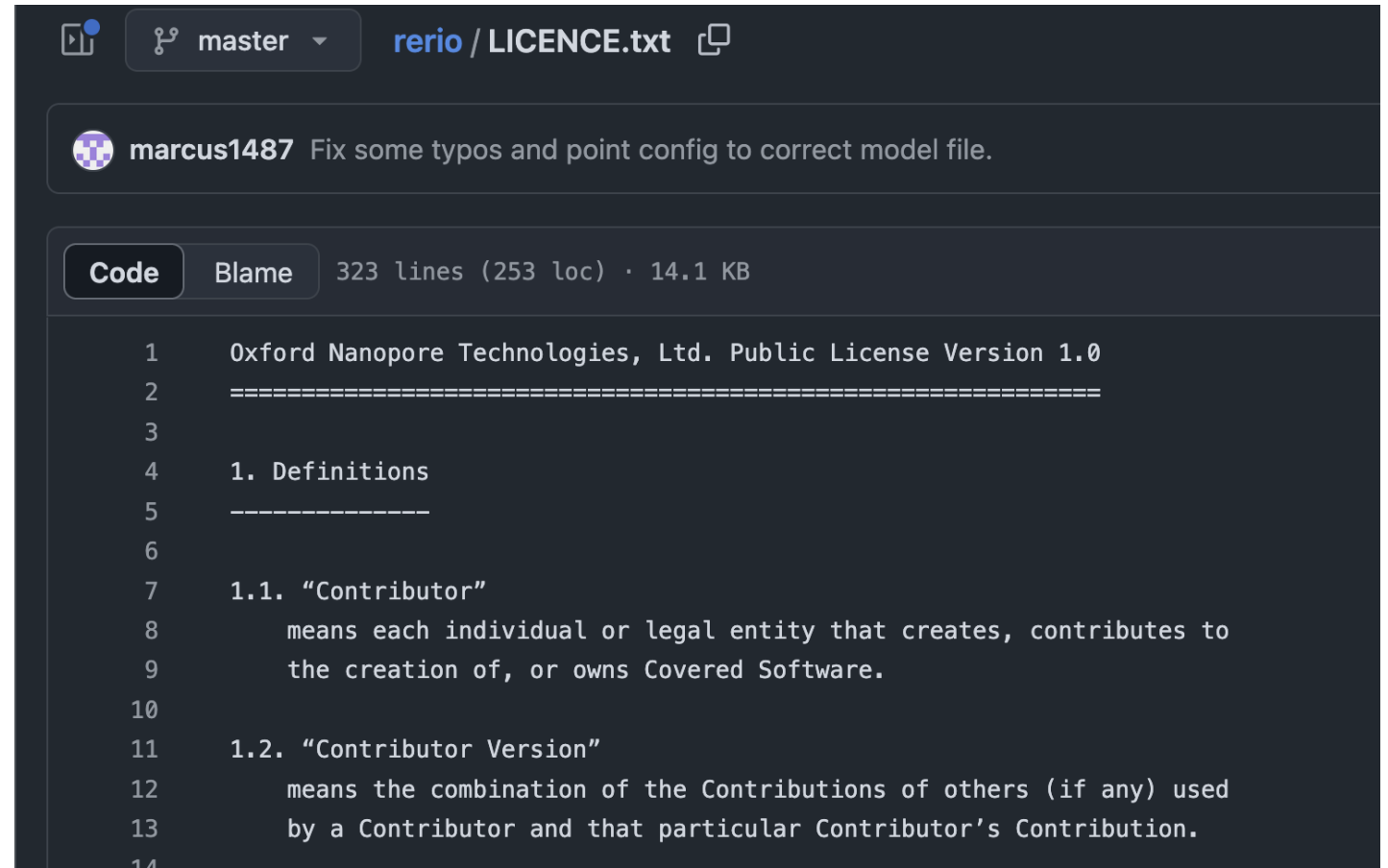
- ☒ Reads (fastq.gz)
- ☒ Primer file (tab)
- ☒ Reference (fasta)
- ☐ Clair3 model (zip)
- ☐ Specify Clade (Clade1/2)

Get clair3 model from

https://cdn.oxfordnanoportal.com/software/analysis/models/_e82_400bps_hac_v500.tar.gz (right click save as)

Rerio Clair3 models are not Open Source

Licensed – for research, public health purposes only.



The screenshot shows a GitHub repository interface for the 'rerio' project. The file 'LICENCE.txt' is selected, showing a commit by 'marcus1487' with the message 'Fix some typos and point config to correct model file.' The file content is displayed in a dark-themed editor, showing the beginning of a license agreement for Oxford Nanopore Technologies, Ltd. The visible text includes the title 'Oxford Nanopore Technologies, Ltd. Public License Version 1.0', a section header '1. Definitions', and two sub-sections: '1.1. "Contributor"' and '1.2. "Contributor Version"', each followed by a definition. The file statistics show 323 lines (253 loc) and 14.1 KB.

```
1 Oxford Nanopore Technologies, Ltd. Public License Version 1.0
2 =====
3
4 1. Definitions
5 -----
6
7 1.1. "Contributor"
8     means each individual or legal entity that creates, contributes to
9     the creation of, or owns Covered Software.
10
11 1.2. "Contributor Version"
12     means the combination of the Contributions of others (if any) used
13     by a Contributor and that particular Contributor's Contribution.
14
```

<https://github.com/nanoporetech/rerio/blob/master/LICENCE.txt>

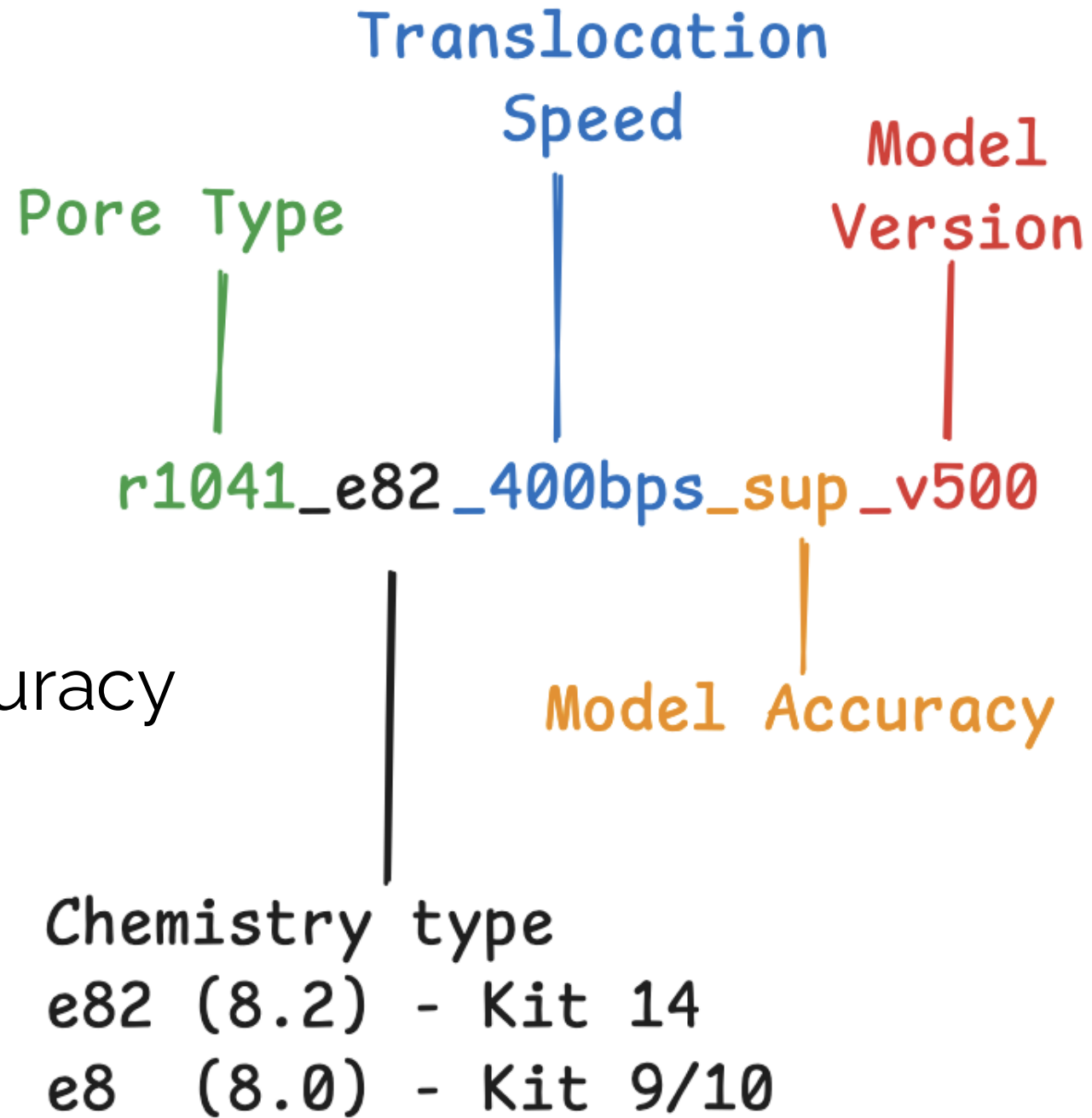
Which model to download?

Decoding model names

Which is our model?

We used:

- Chemistry version K14
- Flow cell R10.4
- Model accuracy was high accuracy



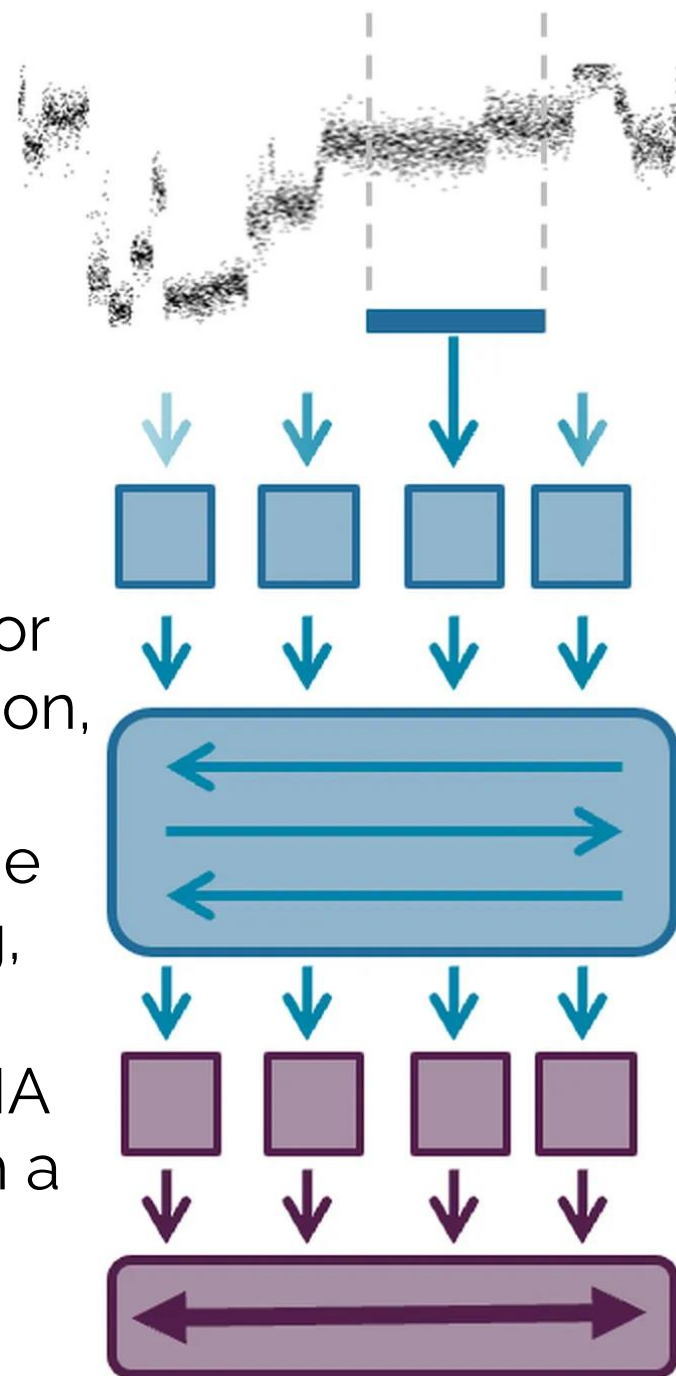
Where to download?

See table:

<https://github.com/nanoporetech/rerio?tab=readme-ov-file#clair3-models>

Why do we need a model?

Basecalling is the process of translating the raw, electrical signals captured by our sequencing devices into a sequence of nucleotides (DNA or RNA). This is like voice recognition, where spoken language is translated into written text. In the context of genomic sequencing, our devices detect changes in electrical current caused by DNA or RNA strands passing through a nanopore.



Parameters learned from training data

Extraction of blocks of features

Bidirectional information flow

Multi-base prediction

Decode to sequence

Let's run the pipeline!

Workflow inputs for MPXV pipeline:

- ☒ Reads (fastq.gz)
- ☒ Primer file (tab)
- ☒ Reference (fasta)
- ☒ Clair3 model (zip)
- ☐ Specify Clade (Clade1/2)

Choosing Clade

- Choosing the correct clade is important
 - MPXV clades have major genomic differences.
- If you are unsure, run the pipeline twice:
 - once using clade 1 as reference
 - once using clade 2 as reference
- Compare results
 - Check greatest coverage % and best depth

Let's run the pipeline!

Workflow inputs for MPXV pipeline:

- ✓ Reads (fastq.gz)
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- ✓ Specify Clade (Clade1/2)

Finally!

Let's run the pipeline!

Your inputs
Need to be
this



History

search datasets

⌵

×

MPXV Training

261 MB

4

137

45

☒

143: r1041_e82_400bps_hac_v5

00.tar.gz

142: reference.fasta

141: primer.bed

92: raw_data

a list with 2 fastqsanger.gz datasets

Let's run the pipeline!

edited about 9 hours ago

workflow runs: 2 ☆ ▼

MPXV (Mpox) Analysis pipeline for ONT

MPXV (Mpox) pipeline for ONT platform. Using the latest Fieldbioinformatics Artic Minion pipeline that uses clair3.

ONT × MPXV × Mpox × #amplicon × Virus ×

Add Tags ➞

Copy Download Share Edit ▶

Click the play button
To enter workflow





 edited about 9 hours ago

 workflow runs: 2

MPXV (Mpox) pipeline for ONT platform. Using the latest Fieldbioinformatics Artic Minion pipeline that uses clair3.

ONT

MPXV

Mpox

#amplicon

Virus

ONT Reads *

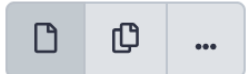


192: fastp on collection 191: Read 1 output

accepted formats ▼

ONT reads from ARTIC assay with fastqsanger encoding

Primers (bed) *

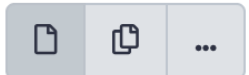


141: primer.bed (as bed)

accepted formats ▼

Primer (bed) file for use as reference. Must be the same reference as the input Reference sequence input. Bed files can be found: https://github.com/artic-network/artic-mpxv-nf/tree/master/data/primer_schemes/MPXV

Reference Sequence *

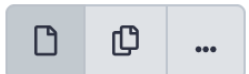


230: all.consensus.fasta

accepted formats ▼

Fasta file for use as reference. Must be the same reference as the input bed file. Reference sequences files can be found: https://github.com/artic-network/artic-mpxv-nf/tree/master/data/primer_schemes/MPXV

Clair3 Model *



144: r1041_e82_400bps_hac_v500.tar.gz

accepted formats ▼

Model must be tar.gz

MPXV Clade *

Clade I

Choose a Clade for downstream phylogenetic processing.

Normalse Reads *

400

Sample at most this number of reads per amplicon and strand. default is 400

Minimum read length - optional

200

Primer scheme

Maximum read length - optional

2600

Primer Scheme

Expand to full workflow form.



 Run Workflow

Click to run the pipeline
Watch the pipeline deploy!

That was a lot! Well done! 10 minute break.

MPXV pipeline outputs

Workflow inputs for MPXV pipeline:

- ✓ Reads (fastq.gz)
- ✓ Primer file (tab)
- ✓ Reference (fasta)

IMPORTANT! Ensure you download the fasta/primers of the same scheme.

Do not mix different schemes together!

Questions? + Resources

- ONT - How basecalling works
 - <https://nanoporetech.com/platform/technology/basecalling>
- Clair3 models:
 - https://github.com/nanoporetech/nerio/tree/master/clair3_models
- To understand model names read this:
 - <https://github.com/nanoporetech/dorado?tab=readme-ov-file#decoding-dorado-model-names>