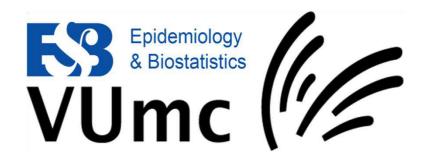


Integrative modeling of timecourse multi-level omics data from HPV-induced cervical carcinogenesis

Viktorian Miok





Contributors

Biostatistics department

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- Wessel van Wieringen
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Pathology department

- Saskia Wilting
- Annelieke Jaspers
- Renske Steenbergen
- Peter Snijders

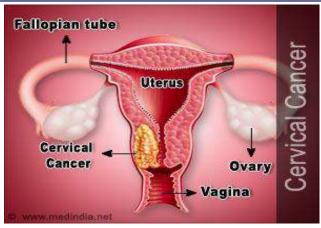


Introduction

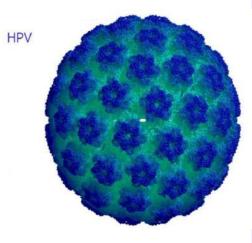


Cervical cancer study

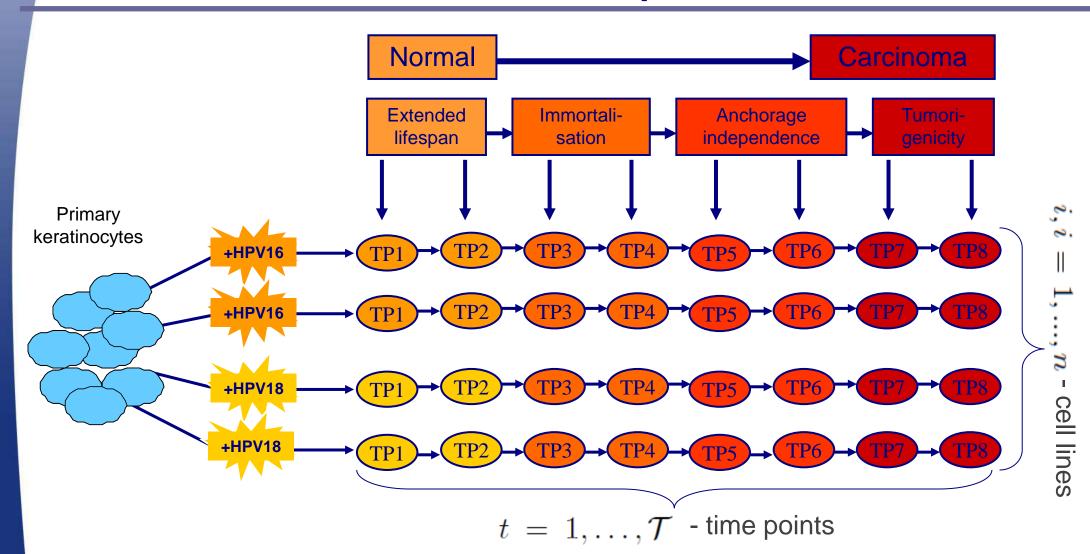
Second most common cancer in women worldwide.



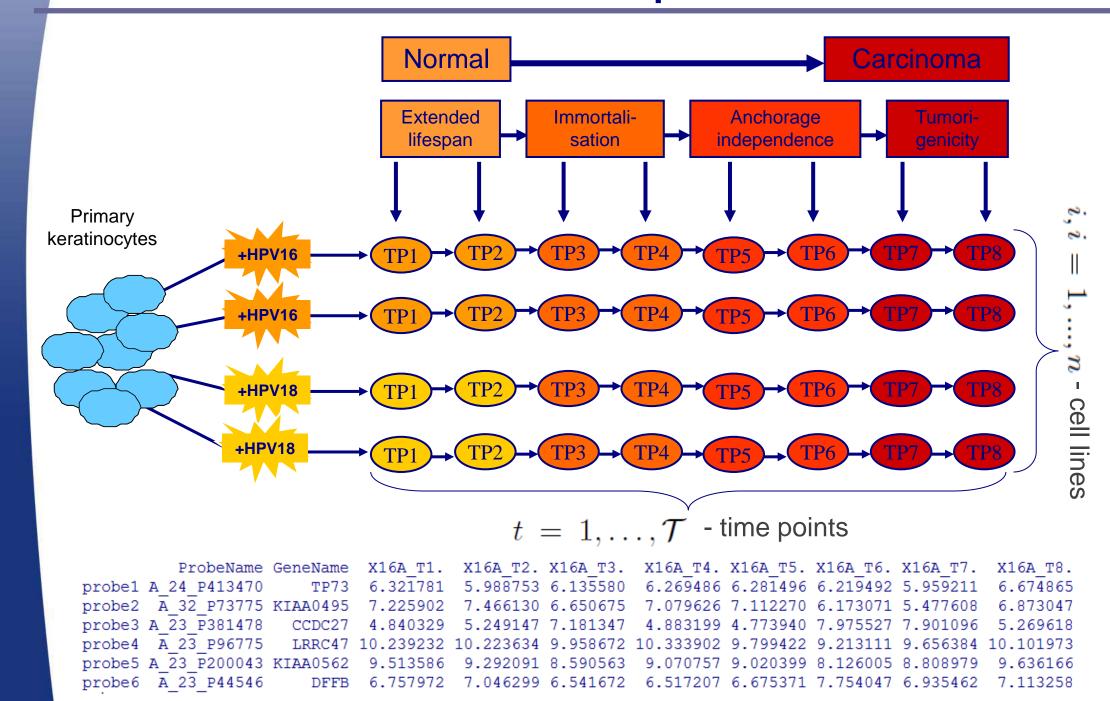
- Caused by HPV virus (70% cases HPV16 and HPV18) and followed by additional (epi)genetic abnormalities.
- Cell line model in vitro model system of HPV-induced transformation.
- Integration high-throughput multi level molecular data sets.
- Understand molecular mechanism driving cervical carcinogenesis



Time-course experiment

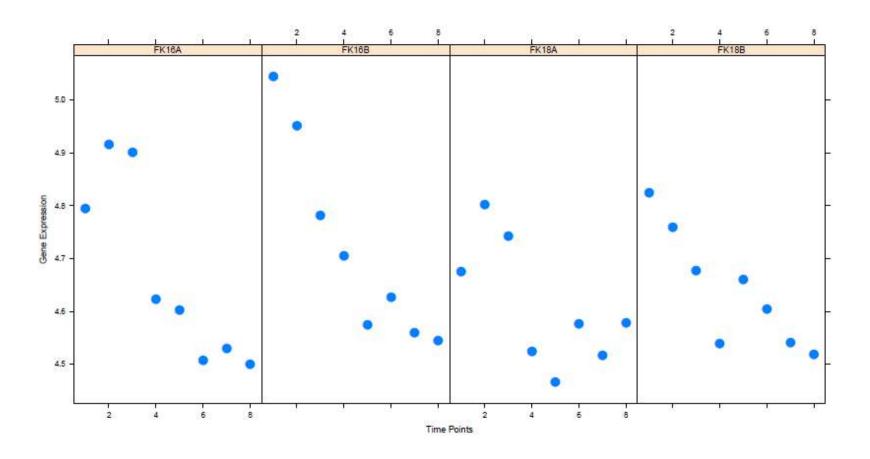


Time-course experiment



Why time-course experiments?

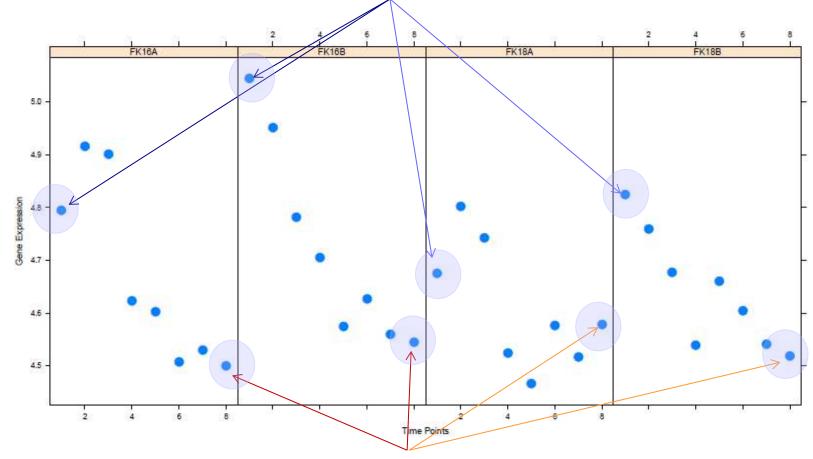
miR-218:



Pick one moment in time

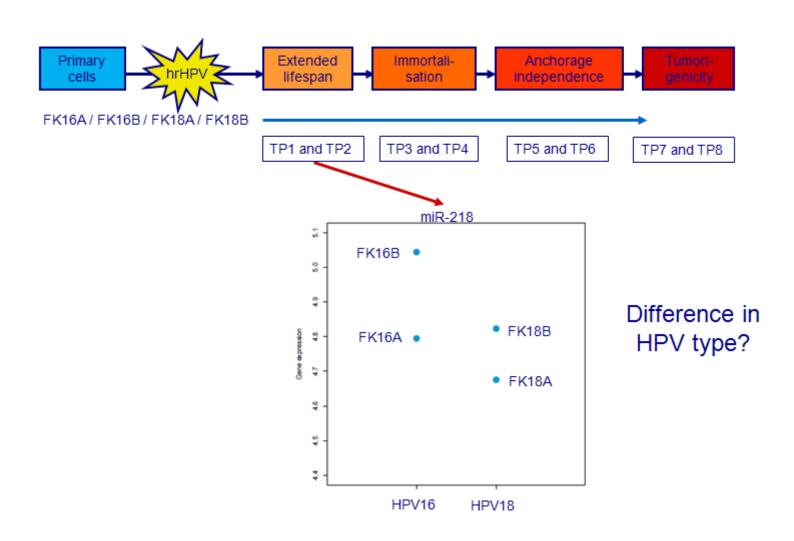
miR-218:

one moment in time: TP1

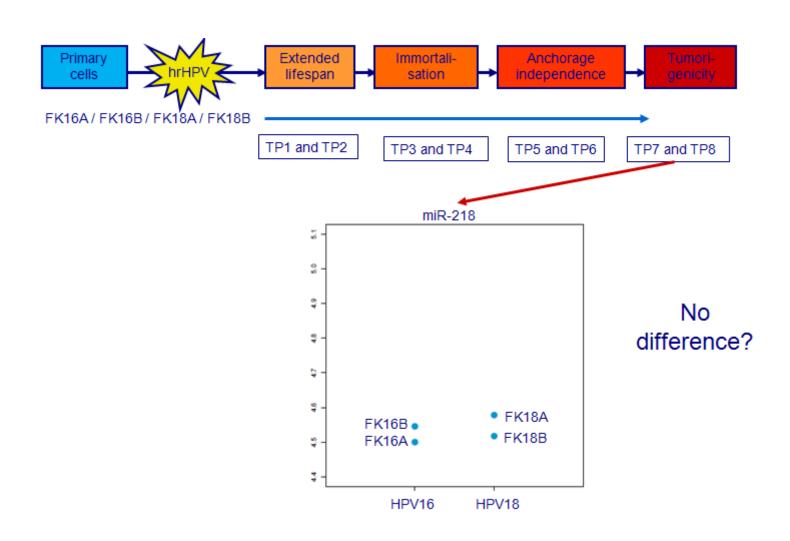


one moment in time: TP8

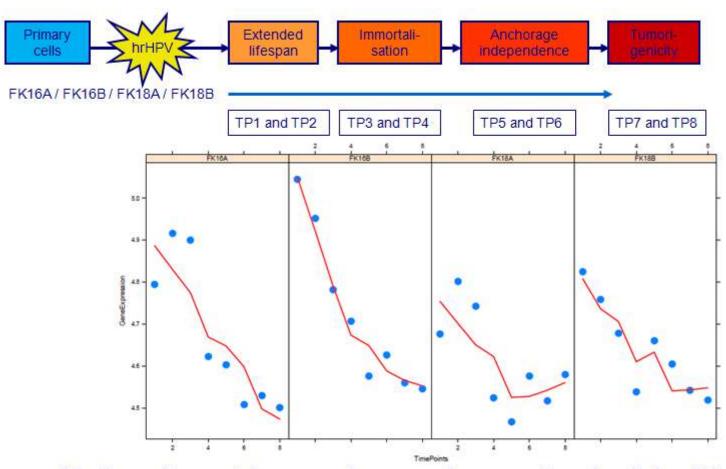
Inference based on TP1



Inference based on TP8



Strength of time-course



Similar pattern of decreased expression over time in all 4 cell lines

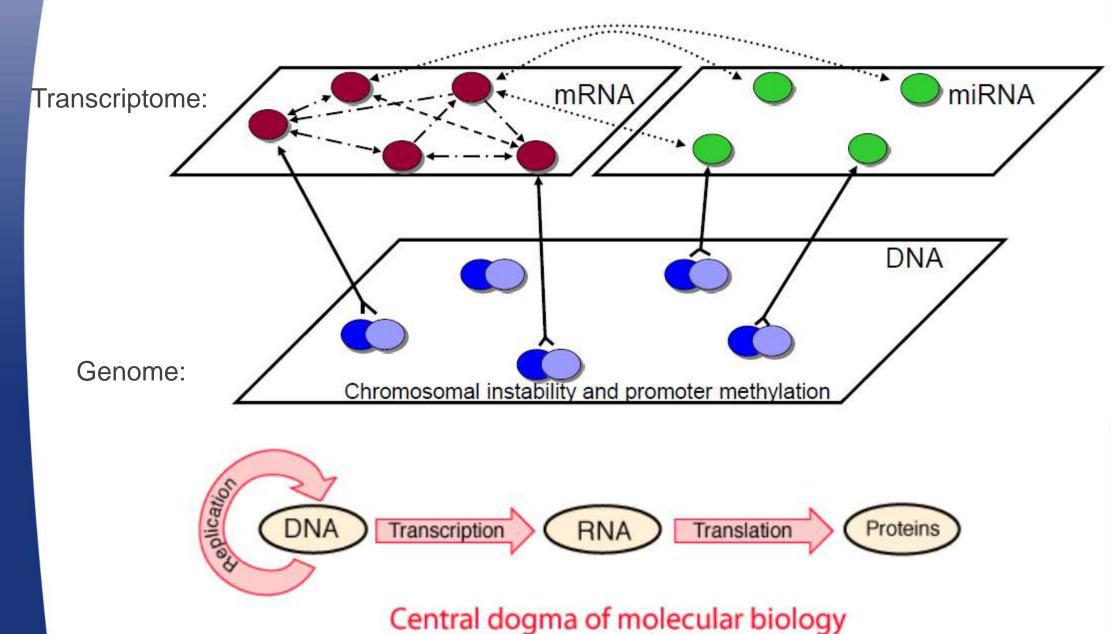


Why integration?



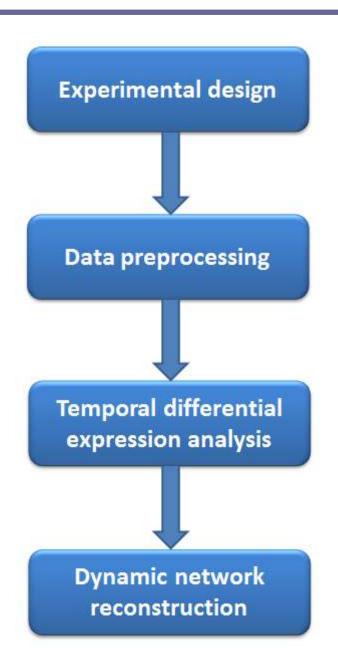
"Blind monks examining an elephant" by Itcho Hanabusa 1888

Multi-omics data integration





What we did?



mRNA: 45K probes arrays

miRNA: 60K probes arrays

CN: 180K probes arrays

mRNA: 27637 genes

miRNA: 1187 genes

CN: 27637 genes

mRNA: 3642 genes

miRNA: 106 genes

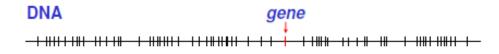
mRNA: 64 genes linked to p53 signaling pathway

miRNA: 106 genes which target mRNA

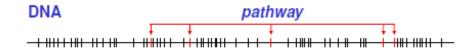


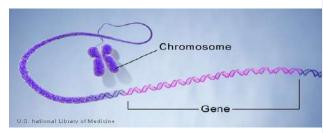
Statistical unit

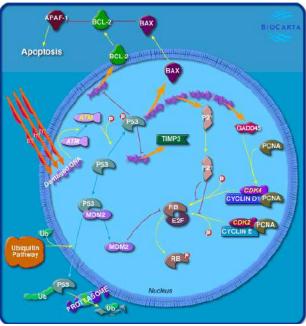
• Gene – measured part of the genome



Pathway – group of genes which work together









Statistical unit

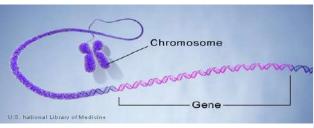
• Gene – measured part of the genome

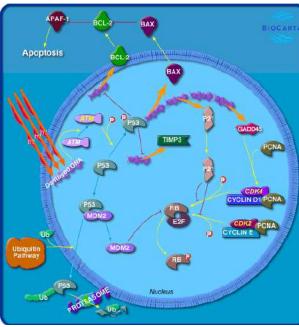


Pathway – group of genes which work together



Approach	Statistical Unit
- Restrict dimension model - Test model across genome - Employ familywise error control	Individual features
 Employ regularization Enabling estimation and inference when p > n 	Pathways







Temporal differential expression analysis



Model

$$j, j = 1, ..., p$$
 -genes

$$\mathbf{Y}_{*,*,t} = (\mathbf{Y}_{1,*,t},...,\mathbf{Y}_{n,*,t})$$
 - mRNA gene expression

Bayesian GLMM: $Y_{i,j,t} \sim \mathcal{N}(\mu_{i,j,t}, \sigma_{\varepsilon,j}^2)$

$$Y_{i,j,t} \sim \mathcal{N}(\mu_{i,j,t}, \sigma_{\varepsilon,j}^2)$$

GE = CL + Time

Model

$$j, j = 1, ..., p$$
 - genes

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Bayesian GLMM:
$$Y_{i,j,t} \sim \mathcal{N}(\mu_{i,j,t}, \sigma_{\varepsilon,j}^2)$$

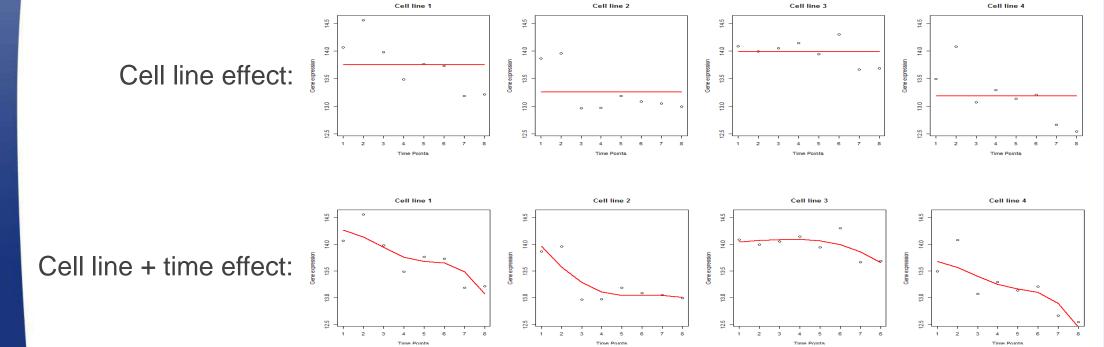
GE = CL + Time

fixed (cell line) effect random (splines) effect

$$\mu_{i,j,t} = f(i; \alpha_j) + h(t; \gamma_j)$$

 α, γ - Gaussian distribution assumption

Fixed and random effects

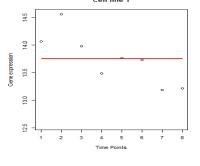


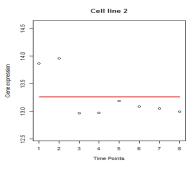
Fixed and random effects

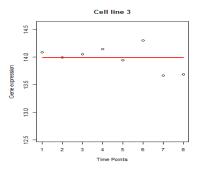
Fixed effect: $f(i; \alpha_j) = \alpha_{i,j}$ cell line

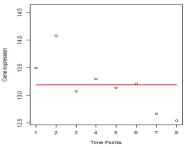
Random effect:
$$h(t; \gamma_j) = \sum_{k=1}^{K} \gamma_{j,k} |t - \kappa_k|^3$$
 time

Cell line effect:

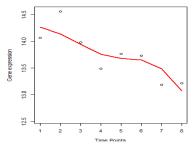


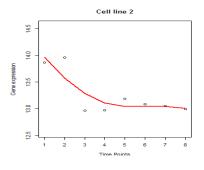


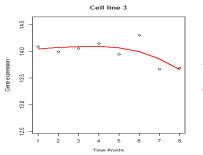


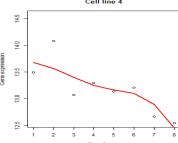


Cell line + time effect:









Matrix notation:

$$Y_{i,j,t} =$$

$$Y_{i,j,t} = \alpha_{i,j} + \tilde{\mathbf{Z}}_t \tilde{\boldsymbol{\gamma}}_j + \varepsilon_{i,j,t}$$

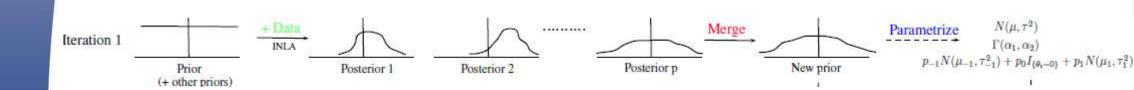
Spline basis:

$$\mathbf{Z}_t = (|t - \kappa_1|^3, \dots, |t - \kappa_K|^3)$$

Spline coefficients:

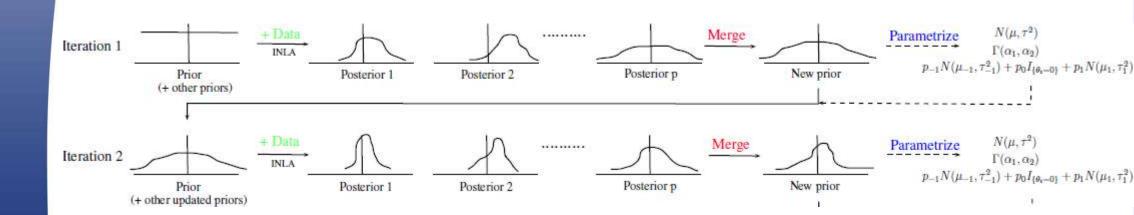
$$\boldsymbol{\gamma}_j = (\gamma_j, \dots, \gamma_{j,K})^{\mathrm{T}}$$

Model parameters estimation

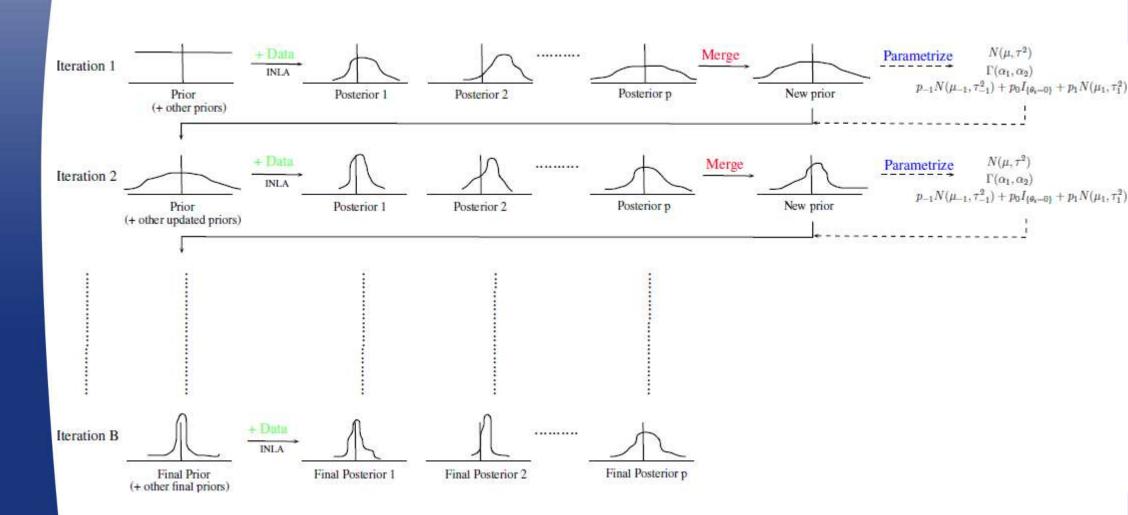




Model parameters estimation



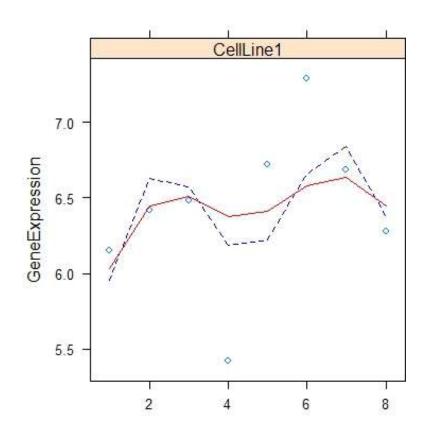
Model parameters estimation



INLA: Marginal posterior are estimated using integrated nested Laplace approximation

Shrinkage

- borrowing information across the genes
- better control of false positives
- improvement of reproducibility
- leads to more stable estimates



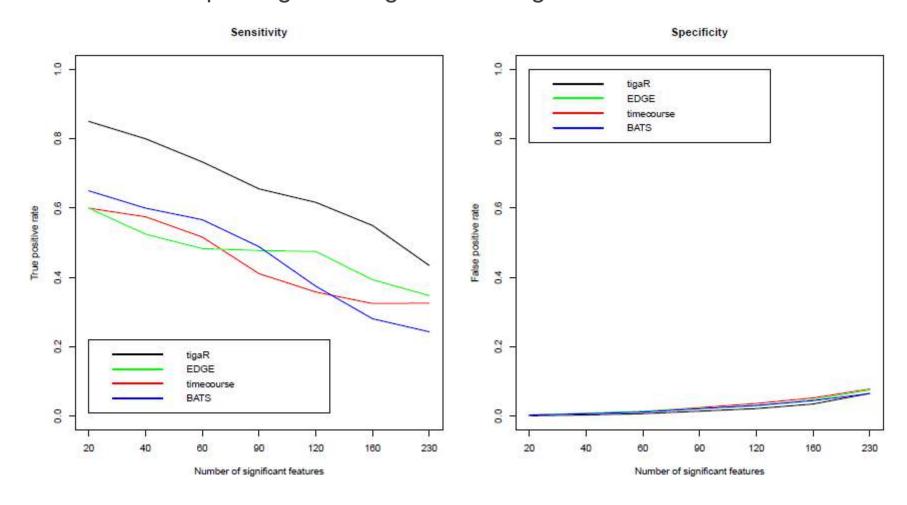
Comparison

- Comparison of following methods:
 - ➤ timecourse Tai and Speed, Annals of Statistics, 2006.
 - ➤ **EDGE** Storey et al., PNAS, 2005.
 - ➤ BATS Angelini et al., BMC Bioinformatics, 2008.
 - ➤ tigaR Miok et al., BMC Bioinformatics, 2014.

- Method is applied on two data sets
 - > Data from our experiment (only mRNA data)
 - > Data from Storey et al., PNAS, 2005.

Sensitivity and specificity

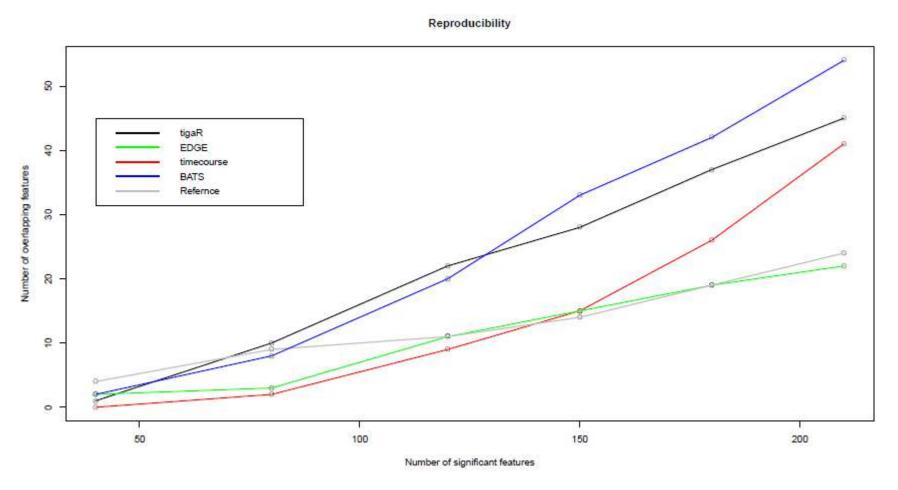
• Truth – overlap of significant genes among methods.





Reproducibility

Equally divided data set in two groups.



DNA copy number (CN)

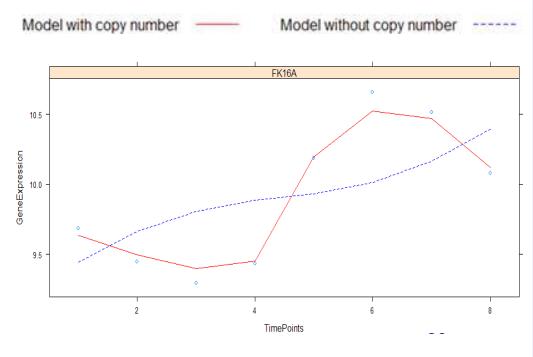
GE = CL + CN + Time

DNA copy number (CN)

$$GE = CL + CN + Time$$

Gene GSTM3:

$$\mathbf{X}_{*,*,t} = (\mathbf{X}_{1,*,t},...,\mathbf{X}_{n,*,t})$$
 - CN Cell line CN Time Error $Y_{i,j,t} = \alpha_{i,j} + \beta_j \, x_{i,j,t} + \mathbf{\tilde{Z}}_t \, \mathbf{\tilde{\gamma}}_j + \varepsilon_{i,j,t}$



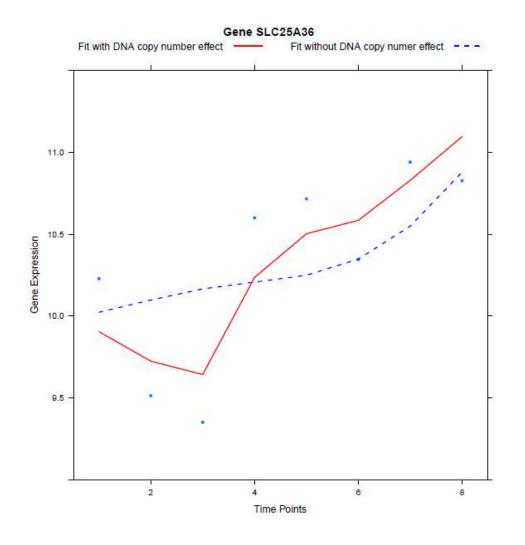
Hypothesis testing

- Questions?
 - ➤ Is there differential expression over time?
 - Does DNA copy number drive gene expression?
 - > Is there a difference between the cell lines?
- Hypothesis are evaluated by means of the likelihood ratio statistics

$$D_{j} = \log \ \left[L \begin{pmatrix} (H_{A}) \\ \widehat{\boldsymbol{\alpha}} \\ j \end{pmatrix}, \widehat{\boldsymbol{\beta}}_{j}^{(H_{A})}, \widehat{\boldsymbol{\sigma}}_{\gamma,j}^{2,(H_{A})}, \widehat{\boldsymbol{\sigma}}_{\varepsilon,j}^{2,(H_{A})} \right] - \log \ \left[L \begin{pmatrix} (H_{0}) \\ \widehat{\boldsymbol{\alpha}} \\ j \end{pmatrix}, 0, \widehat{\boldsymbol{\sigma}}_{\gamma,j}^{2,(H_{0})}, \widehat{\boldsymbol{\sigma}}_{\varepsilon,j}^{2,(H_{0})} \right]$$

To account for multiplicity the False Discovery Rate is controlled

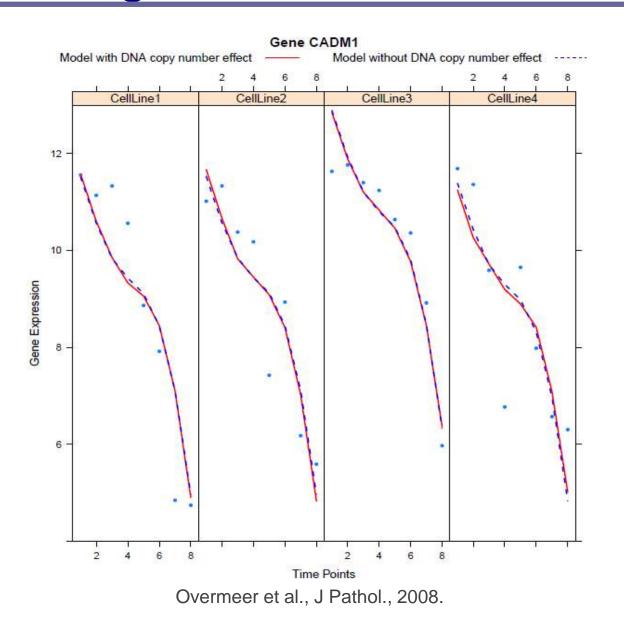
SLC25A36 – gene with CN effect



Wilting et al., Genes, Chromosomes and Cancer, 2008.



CADM1- gene without CN effect

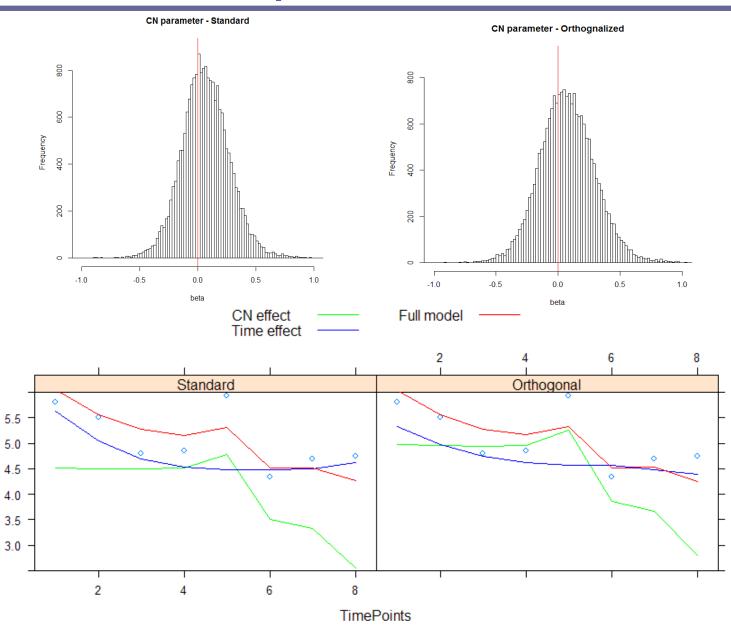


Orthogonalization splines onto CN

CN parameter standard vs. orthogonal:

Fit of the model standard vs. orthogonal:

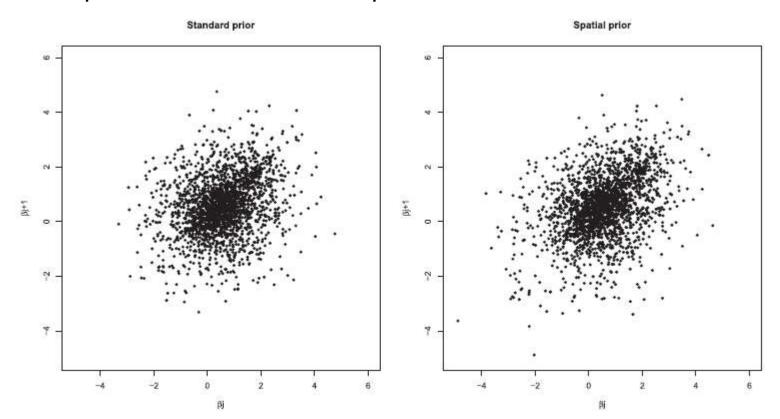
GeneExpression



Spatial multivariate prior for CN

$$\text{Multivariate prior:} \quad \begin{pmatrix} \beta_{j-1} \\ \beta_{j} \\ \beta_{j+1} \end{pmatrix} \sim \mathcal{N} \begin{pmatrix} \begin{pmatrix} 0 \\ 0 \\ 0 \end{pmatrix}, \begin{pmatrix} \sigma_{j-1}^2 & \sigma_{j-1}\sigma_{j}\rho & \sigma_{j-1}\sigma_{j+1}\rho^2 \\ \sigma_{j-1}\sigma_{j}\rho & \sigma_{j}^2 & \sigma_{j}\sigma_{j+1}\rho \\ \sigma_{j-1}\sigma_{j+1}\rho^2 & \sigma_{j}\sigma_{j+1}\rho & \sigma_{j+1}^2 \end{pmatrix} \right)$$

Improvement in partial correlation of CN parameters:



Model fit with same and different splines

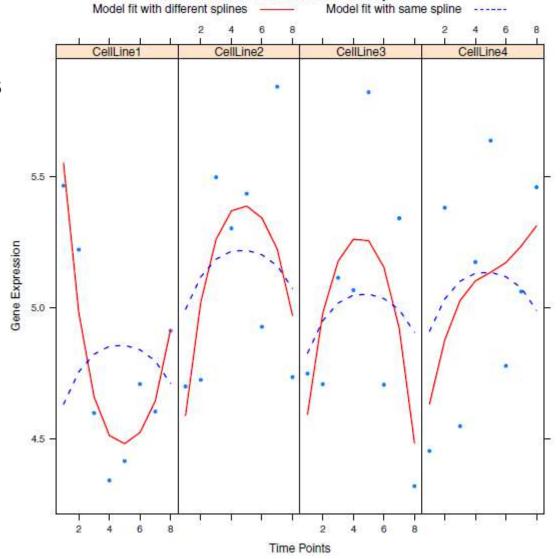
Splines flexibility

Same spline – up/down regulated genes

$$ilde{\mathbf{Z}} = ilde{\mathbf{Z}} \otimes \mathbf{1}_{n imes n}$$

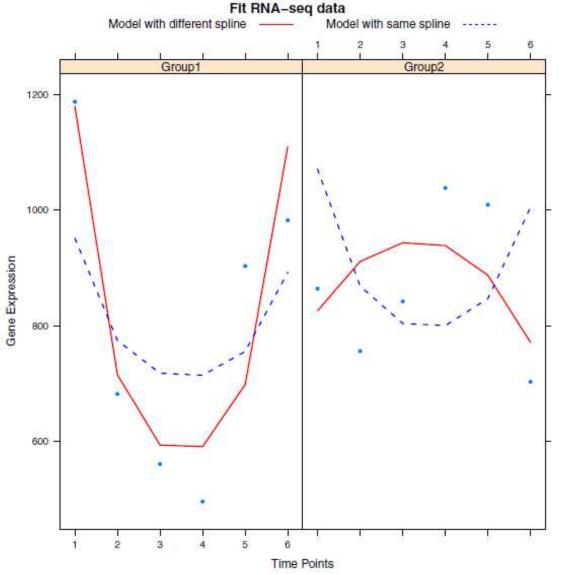
Different spline – allow more flexibility

$$\tilde{\mathbf{Z}} = \tilde{\mathbf{Z}} \otimes \mathbf{I}_{n \times n}$$



RNA-seq data

- Changing link function method can deal with count data.
- Two group time-course RNA-seq data.



Article + R-package

Miok et al. BMC Bioinformatics 2014, 15:327 http://www.biomedcentral.com/1471-2105/15/327



METHODOLOGY ARTICLE

Open Access

tigaR: integrative significance analysis of temporal differential gene expression induced by genomic abnormalities

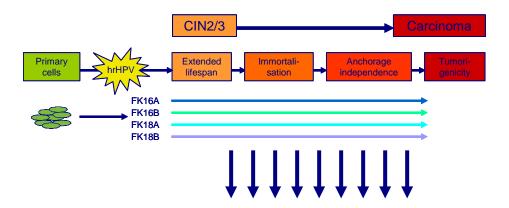
Viktorian Miok^{1,2}, Saskia M Wilting², Mark A van de Wiel^{1,3}, Annelieke Jaspers², Paula I van Noort⁴, Ruud H Brakenhoff⁵, Peter JF Snijders², Renske DM Steenbergen² and Wessel N van Wieringen^{1,3*}



tigaR: temporal integrative genomic analysis in R

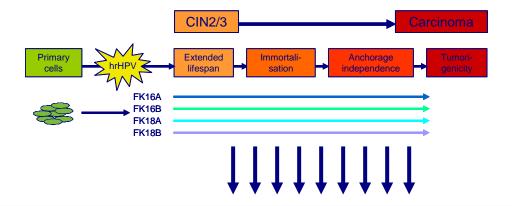
https://github.com/viktormiok/tigaR

tigaR analysis



Measured expression of 1187 miRNAs and 27637 mRNAs

tigaR analysis



Measured expression of 1187 miRNAs and 27637 mRNAs

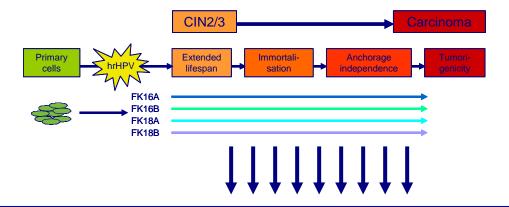




(concordant change in expression in at least 3 cell lines)



tigaR analysis



Measured expression of 1187 miRNAs and 27637 mRNAs





(concordant change in expression in at least 3 cell lines)



36 miRNAs and 1233 mRNAs linked with CN

(~34% of altered expression in both cases)





Thank you for your attention!