

Computational modeling of the human atrial anatomy and electrophysiology

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Abstract This review article gives a comprehensive survey of the progress made in computational modeling of the human atria during the last 10 years. Modeling the anatomy has emerged from simple “peanut”-like structures to very detailed models including atrial wall and fiber direction. Electrophysiological models started with just two cellular models in 1998. Today, five models exist considering e.g. details of intracellular compartments and atrial heterogeneity. On the pathological side, modeling atrial remodeling and fibrotic tissue are the other important aspects. The bridge to data that are measured in the catheter laboratory and on the body surface (ECG) is under construction. Every measurement can be used either for model personalization or for validation. Potential clinical applications are briefly outlined and future research perspectives are suggested.

Keywords Cardiac modeling · Atrial arrhythmia · Radio frequency ablation · Remodeling · Intraatrial electrograms

Abbreviations

AF	Atrial fibrillation
AP	Action potential
APD	Action potential duration
BB	Bachmann’s bundle

BCL	Basic cycle length
BSP(M)	Body surface potential (map)
CS	Coronary sinus
CT	Crista terminalis
CT	Computed tomography
CV	Conduction velocity
ECG	Electrocardiogram
EP	Electrophysiology
ERP	Effective refractory period
IVC	Inferior vena cava
LA	Left atrium
LAA	Left atrial appendage
LIPV	Left inferior pulmonary vein
LE-MRI	Late enhancement MRI
LSPV	Left superior pulmonary vein
MRI	Magnetic resonance imaging
PM	Pectinate muscle
PV	Pulmonary vein
PWd	P-wave duration
RA	Right atrium
RAA	Right atrial appendage
RF(A)	Radio frequency (ablation)
RIPV	Right inferior pulmonary vein
RSPV	Right superior pulmonary vein
SN	Sinus node
SVC	Superior vena cava
TEE	Transesophageal echocardiogram
WL	Wave length

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1 Introduction

Intensive research is carried out in the field of atrial physiology and pathophysiology, and both efforts and comprehension are increasing rapidly [215]. One

motivation for this endeavour lies in the fact that around 1 % of the population suffer from atrial fibrillation (AF). The incidence of AF increases significantly with age, that is why this topic is of utmost relevance in countries with aging population [215]. Since the year 2000, about 25 articles on comprehensive mathematical 3D-models of the atria and their electrophysiology and another 200 articles of various aspects on atrial modeling have been published. This strong research is motivated by the fact that computational modeling of the atria can contribute significantly to better comprehension of atrial physiology and pathophysiology. By that, it can lead to a more detailed diagnosis and improved therapy for millions of patients. Computational modeling enables a much deeper interpretation of experimental data and motivates new clinical research. We also find several review articles and book chapters on *in silico* modeling of the heart [29, 118, 166, 225, 239] and in particular the atria [97, 246]. The focus of this review article is 100 % on the atria and it will investigate the complete chain from the ion channels, the anatomy and the electrophysiology to the atrial intracardiac electrogram and the electrocardiogram (ECG). The review will demonstrate how simple models have evolved to sophisticated models that take into account heterogeneity of the atrial cells, details of the geometry and muscle strands, fiber direction and anisotropy, restitution curves, remodeling, fibrotic tissue and ablation scars. We will also try to bridge the gap between computational models and real measurements that are carried out every day in the electrophysiological laboratory (EP-lab) in the hospital.

2 Modeling the geometry of the atria

There is a strong link between the anatomy of the atria and the physiological excitation sequence. Also the onset and perpetuation of AF is connected to the anatomy [13, 85, 87, 90, 132]. Therefore, the following section briefly introduces gross atrial anatomy and focuses on the anatomical features relevant to atrial modeling. Comprehensive reviews about atrial anatomy can be found elsewhere [84, 85, 87].

2.1 Anatomy of the atria

The right atrium (RA) collects venous blood from the two caval veins and the coronary sinus (CS). The RA myocardium is separated into a smooth venous part and a rough free wall [87] by the crista terminalis (CT) [205]. The RA hosts the sinus node (SN), which is located at the junction of the superior caval vein (SVC) and the right atrial appendage (RAA) [206]. The triangular RAA clasps the aortic arch [205].

Usually, four pulmonary veins (PVs) connect to the left atrium (LA), which has a smooth endocardial surface. The posterior wall of the left atrium is close to the oesophagus [207]. The left atrial appendage (LAA) is clearly delineated and has a tubular shape with multiple finger-like branches [228]. The LA has a diameter of 3.7 cm [183, 240] and a volume of 42 ml in normal hearts [183]. An increase in left atrial volume is commonly linked to AF in clinical practice [214]. Atrial dilatation is therefore also subject to modeling studies (e.g. [198]).

The atria are connected via multiple interatrial bridges with Bachmann's bundle (BB) as the most prominent one, running from the area of the SN towards the LAA [88], bridging the interatrial groove [130, 248]. There is a high variability of the interatrial bundles between individuals [3, 88, 203].

2.1.1 Wall thickness

The wall of the human atria has a reported thickness between 1 and 3 mm [30, 40, 77, 165, 174, 180, 207, 229, 253]. Wall thickness in the RA is non-uniform due to the CT and pectinate muscles (PMs) [253]. The LA wall is thicker on the posterior and inferior side compared to the LA roof [77, 180]. Wang et al. [253] measured a thicker wall in the LA compared to the RA, whereas Neher et al. [165] reported the opposite (3.3 vs. 2.1 mm). Males have a slightly thicker atrial wall compared to females [77] and patients with AF often have a thinner left atrial wall (2.1–2.5 mm vs. 2.3–2.9 mm) [180]. The transmural of a lesion made with radio frequency (RF) ablation depends on the regional varying atrial wall thickness [40, 174] or vice-versa the wall thickness has an impact on the energy needed to create a transmural lesion [18, 88].

2.1.2 Pulmonary veins

Paroxysmal AF is most often triggered by ectopic activity in the PVs [75]. The ostia of the PVs are therefore of increasing interest for the electrophysiologist and are also subject of simulation studies (e.g. [260, 237]). The diameter of the PV ostia is larger in the superior veins than in the inferior veins (11 vs. 8 mm) and is often increased in patients with AF triggered by ectopic foci in the PVs [137]. The wall of the PVs is around 0.5 mm thick [81].

Typically, four PVs from the five lung lobes connect to the LA (two left, two right) [21]. The vein from the right middle lung lobe thereby usually joins the upper lobe vein before reaching the LA [241]. Various other patterns have been reported in literature as well [21, 78, 102, 106, 107, 149, 152, 159, 241]. The findings vary depending on the focus of the study as well as on the definition of the drainage pattern. Overall, common variants are a third middle right PV [241]

and a common trunk of the two left PVs [107, 159]. Kaseno et al. [106] observed a short common left trunk in more than half of 428 computed tomography (CT) scans of individual patients. In the detailed study of Marom et al. [152] four different patterns (1–2 ostia) on the left and ten different patterns (2–5 ostia) on the right side of the LA are distinguished.

The venous tissue of the PVs is covered by myocardial cells [85, 108]. These so-called sleeves have a variable length from the junction of the ostia and the left atrial body and are longer in the superior veins compared to the inferior veins [24, 88, 86, 202], which concurs with a higher rate of ectopic foci in the superior PVs [74]. The left superior PV (LSPV) was shown to have the longest sleeve, followed by the right superior PV (RSPV), left inferior PV (LIPV) and right inferior PV (RIPV) (41 vs. 30 vs. 21 vs. 13 mm) [24] and no significant difference between controls and AF patients was found. Measurements from Hassink et al. and Saito et al. confirmed the order of different length in PV sleeves, but experienced shorter extension of myocardium into the PVs (7.2–10.9 mm [81] and 6.0–14.8 mm [202]). Hassink et al. [81] also found the sleeves in the AF group to be longer than in controls (10.4 vs. 8.7 mm), whereas Saito et al. [202] did not find a significant difference.

2.1.3 Atrial isthmuses and interatrial septum

In the RA, the area between the IVC and the tricuspid valve forms an isthmus [19, 85]. This region is linked to macro-reentries in atrial flutter and is thus often target of ablation therapy [19], as it has a slower conduction velocity in these patients [55]. The LA hosts the mitral isthmus in the region between the mitral valve and the LIPV, which may also be target of ablation in AF patients [263].

The interatrial septum includes an electrically isolating layer [3, 84, 131]. Both sides of the septum are covered with conductive myocardium, except in the area of the fossa ovalis, where only the ring around the fossa ovalis is conductive.

2.2 Geometric models of the atria

Geometrical atrial models can be divided into different groups. One group consists of very detailed models of specific parts of mammal atria, most often of regions of the RAA, CT and PMs as well as the PV ostia (e.g. [22, 53, 271, 273]). Another group includes 3D models of human atria. This group can be subdivided into surface and volumetric models. Both model types represent a geometry in three dimensions, but surface models cannot reflect the finite wall thickness of the atria. The models represent either generalized atrial shapes (e.g. a two-sphere “peanut”), the anatomy of a single human derived from image data (magnetic

resonance imaging (MRI) or CT) or very recently reflect patient-specific atrial anatomies. Table 1 provides an overview of published 3D models of the atria and their properties.

Besides atrial models created for electrophysiological simulations, further atrial models, partly comprising fast conducting myocardial structures, were published without performing electrophysiological simulation [43, 165, 173, 200, 232, 258].

Geometric atrial models usually reflect the normal atrial anatomy, with four PVs and an average literature-based wall thickness, if this information could not be derived from the image data. An inconsistency exists in the modeling of the atrial septum, which is modeled either as fully conductive, or including an isolating layer (see Table 1). On the other hand, modeling of interatrial bundles is more consistent, as models commonly at least include the connection via BB and the CS (Table 1).

2.2.1 Emerging imaging and segmentation technology

Recent advances in cardiac imaging allow the creation of patient-specific atrial models from various data entities, such as CT [49, 50, 94], 4D CT [156], cone-beam CT [150], MRI [258], and TEE [252]. These advances will enable the use of such models for electrophysiological simulations accompanying the clinical workflow [124, 48].

New image segmentation techniques also allow the patient individual determination of the PV variant [47, 78, 107], which will support the generation of detailed personalized geometrical atrial models. Advances in MRI enable in-vivo imaging of atrial wall thickness, which in future could augment personalized atrial models [165]. Imaging atrial fiber orientation in-vivo using diffusion tensor MRI, is not yet possible, although becoming feasible for the ventricles [238], because MRI resolution is in the range of atrial wall thickness and partial volume effects would therefore outweigh real data.

Another rising aspect is the generation and utilization of cardiac atlases and statistical models [264]. Atlases are generated from a multitude of image data and can be used to find a representation for the mean anatomy [173] and to show the variability of certain anatomical features [140]. Data for the generation of cardiac atlases as well as individual atrial models can be found in web-based databases (Cardiac Atlas Project (CAP) [7], Anatomical Model Database (AMDB) [65]). Statistical mean models are already used for cardiac image segmentation, but may in future also play a role in therapy planning and clinical training.

2.3 Fibers and bundles

Atrial fiber orientation with high anisotropy or other disturbances was shown to be linked to the onset and sustaining of AF [96, 177]. Preferred orientation of atrial

Table 1 3D geometrical atrial models

References	Basis	Species	Comp.	Geo.	Mesh	EPM	Prop.	Fibers	Sep.	Heterogeneities	Ext.
Lorange et al. [141]		Human (CT)	Four chamber	Surf-2D	Voxel	n/p	eikonal	–	c	–	ECG
Ellis et al. [54]		Generic	Biatrial	Surf-2D	Icosahedral	n/p	n/p	–	n/p	–	Abl
Gray et al. [70]		Generic	Two spheres	Surf-3D	Cubic	[61]	[70]	–	c	–	
Freudenberg et al. [62]		Visible female	Biatrial	Vol	Voxel	n/p	CA	–	c	CT	(G/-/-)
Harild et al. [80]		Human (MRI)	Biatrial	Vol	Hexahedral	Nyg	MD	B	i	BB, CT, PM, IS, FO	(G/CV/-)
Werner et al. [259]		Visible man	Four chamber	Vol	Voxel	n/p	CA	V	c	–	ECG
Vigmond et al. [249]		Generic	Two spheres	Surf-3D	Cables	[186]	[249]	ML	i	BB, CT, PM, CS, FO	(G/-/-)
Zemlin et al. [266]		Visible female	Biatrial	Surf-3D	Triangular	CRN	[266]	ML	c	CT, PM	(G/CV/-)
Blanc et al. [14]		Generic	Two spheres	Surf	Voxel	[11, 143]	MD	–	c	–	
Virag et al. [251]		Human (1, MRI)	Biatrial	Surf-3D	Triangular	CRN	MD	–	i	BB, CT, FO	(G/CV/-)
van Dam et al. [35]		Human (1, MRI)	Biatrial	Surf-3D	Triangular	n/a	FM	–	c	–	
Jacquemet et al. [37, 99, 100, 195–197, 243]		Human (1, MRI)	Biatrial	Surf-3D	Cubic	CRN	MD	ML	i	BB, CT, FO	(G/-/-)
Seemann et al. [219]		Visible female	Biatrial	Vol	Voxel	CRN	MD	B	c	BB, CT, PM, AVR, APG, SN	(G/CV/EP)
Ridler et al. [190]		Human (1, CT)	Biatrial	3D	Cables	[186]	MD	Yes	i	CS, FO, CS	(G/-/-)
Dokos et al. [46]		Generic	Two spheres	Surf-2D	Triangular	[193]	n/p	–	c	–	
Kuijpers et al. [128]		Human (1, MRI)	Biatrial	Surf-3D	Triangular	CRN	BD	–	c	–	
Gong et al. [66]		Human (averaged)	Biatrial	Vol	Hexahedral	CRN	n/p	B	i	BB, CT, PM, CS, FO, IS	(G/CV/-)
Plank et al. [178]		Visible female	Left atrium	Vol	Voxel	CRN	MD	V	n/a	–	
Tobón et al. [233–236]		Human (1, MRI)	Biatrial	Surf-3D	Triangular	Nyg	MD	ML	i	BB, CT, PM, CS, IS, FO	(G/CV/-)
Wieser et al. [261, 262]		Human (1, MRI)	Biatrial	Surf-3D	Triangular	[186]	MD	–	i	BB, CT, CS, FO	(G/CV/-)
Reumann et al. [188]		Visible female	Biatrial	Vol	Voxel	CRN	CA	B	c	BB, CT, PM	(G/-/-)
Kharche et al. [110]		Visible female	Biatrial	Vol	Voxel	CRN	CA	B	c	BB, CT, PM, AVR, APG, SNs	(G/CV/EP)
Krueger et al. [126]		Visible man	Biatrial	Vol	Voxel	CRN	MD	B	c	BB, CT, PM	(G/CV/EP)
Krueger et al. [123, 226]		Human (2, MRI)	Biatrial	Vol	Voxel	CRN	MD	B	c	BB, CT, PM, AVR, APG	(G/CV/EP)

Table 1 continued

References	Basis	Species	Comp.	Geo.	Mesh	EPM	Prop.	Fibers	Sep.	Heterogeneities	Ext.
Abed et al. [1]	Visible man		Biatial	Vol	Voxel	[1]	MD	–	n/p	BB, CT, CS, APG	(G/-)
Krueger et al. [124]	Human (15, CT, MRI)		Biatial	Vol	Voxel	CRN	MD	VL	c	BB, CT, PM, IS, AVR, APG	(G/CV/EP)
Lu et al. [142]	Human (1, MRI)		Four chambers	Vol	Spheroidal	Nyg	CA	–	n/p	BB, CT, PM, IS, FO	(G/-) ECG
Aslanidi et al. [6]	[219] Visible female		Biatial	Vol	Voxel	CRN	[6]	BS	c	BB, CT, PM, SN	(G/CV/EP) ECG
Cristoforetti et al. [33]	Human (1, CT)		Left atrium	Surf	Triangular	CRN	n/p	–	n/a	–	–
Ridder et al. [191]	[190] Human (1, CT) (dog size)		Biatial	3D	Cables	[114]	MD	–	i	CT, CS, FO	(G/-)
Burdumy et al. [16]	Human (4, CT)		Biatial	Vol	Voxel	CRN	CA	–	c	–	–
Zhao et al. [272]	Sheep (1, microscopy)		Biatial	Vol	Voxel	[57]	MD	V	c	BB, CT, PM, PV	(G/-)

EPM electrophysiological model, *CRN* model of Courtenanche et al. [31], *Nyg* model of Nygren et al. [168], *Comp.* compartments, *Prop.* propagation model, *Sep.* septum, *B* bundles, *V* volumetric, *ML* mono-layer, *VL* various layers, *BS* bundles and SN, *Ext.* model extension, *Abl* ablation lesion model, *ECG* ECG is computed, *Surf* surface model, *Surf-3D* 3D surface model, *vol* 3D volumetric model *CA* cellular automaton, *MD* monodomain, *BD* bidomain, *i* isolating, *c* connective, *n/a* not applicable, *n/p* not specified, *IS* right atrial isthmus, *AVR* atrio-ventricular ring, *APG* appendages, *G* structures in geometry, *CV* structures have different conductivity, *EP* structures have different electrophysiology

myocytes (myocardial fiber orientation) in the human atria is characterized by multiple overlapping, joining and separating fiber sheets and bundles. Some of the bundles have strong anisotropy and significantly influence the atrial depolarization sequence [13, 38, 151]. In the RA, the CT is the most prominent bundle and runs from the inferior vena cava (IVC) to the SVC while diminishing in width (9.1 vs. 6.1 mm) [205]. 15–20 PMs arise perpendicularly from the CT towards the muscular ring of the tricuspid valve [253]. The superior PM, so-called septum spurium, marks the beginning of the circular oriented myocardial fibers of the tip of the RAA and is thicker compared to the other PMs (4.2 mm vs. 1–3 mm) and shorter [205]. The inferior PMs encircle the orifice of the CS. From the circular fibers surrounding the SVC an epicardial fiber layer is extending over the smooth portion of the RA between the caval veins, crosses the CT epicardially and joins the fibers of the PMs [175].

Fiber orientation in the LA is made up of two myocardial layers overlapping each other transmurally. The septo-atrial bundle lies in the subendocardium and the septo-pulmonary bundle runs subepicardially [87]. The PV ostia and the atrio-ventricular valves are encircled by myocardial fibers [161]. An abrupt change of fiber direction can be present at the PV ostia [89]. The most comprehensive reviews of atrial fiber orientation can be found in Refs. [84] and [87].

2.3.1 Modeling atrial fibers

Fiber orientation can be included based on published anatomical observations (described above) into atrial models manually [80, 100, 219, 235, 249] or semi-automatically/rule-based [83, 123, 124, 126, 177, 200, 259, 266]. All models have in common, that surface-3D models cannot fully reproduce the complex multi-layer fiber arrangement described above. An example for rule-based, volumetric atrial fiber orientation is pictured in Fig. 1. In addition, more detailed animal models of specific atrial regions include fiber orientation derived from high-resolution imaging [22, 272, 273]. It has been shown, that some of the prominent bundles may be visible in high-resolution CT scans [208, 209]. This yields the possibility to include patient individual sizes of these structures in atrial models in the future. Recently, semi-automatic methods were presented to incorporate fiber orientation in patient-specific atrial models [83, 123, 124, 126].

3 Electrophysiology

3.1 General atrial electrophysiology

The resting membrane voltage of a human atrial myocyte is about –80 to –90 mV. An external stimulus leads to the

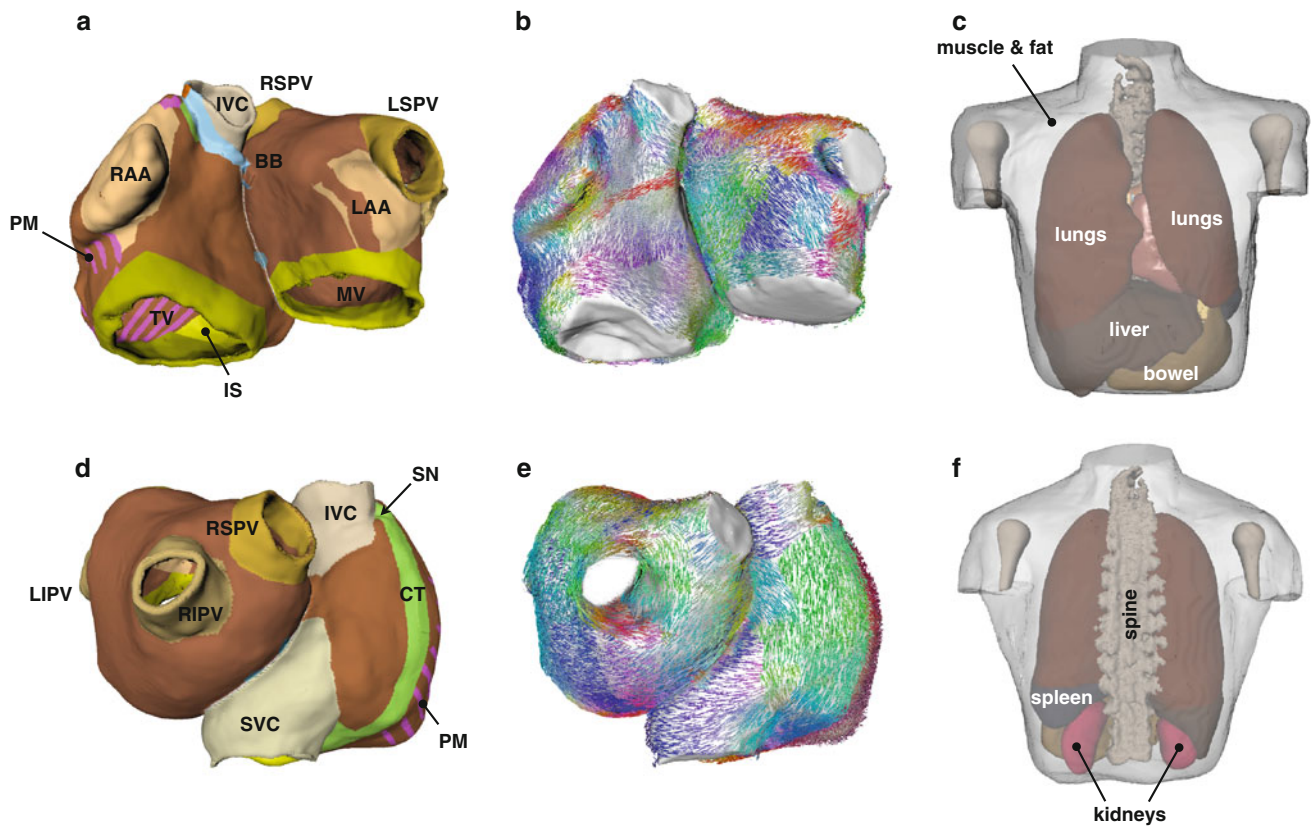


Fig. 1 **a, d** Anatomical model of an AF patient with structures relevant for EP simulations (colored). **b, e** Rule-based volumetric fiber orientation in the atrial model [124]. Orientation of fibers is

color-coded. **c, f** Anterior and posterior view of a thorax model of a volunteer with segmented organs [123]

initiation of a characteristic action potential (AP). The AP starts with the opening of fast Na^+ channels, which are rapidly inactivated causing the refractory period. After the overshoot, fast repolarizing K^+ outward currents lead to a decrease of the transmembrane voltage. This process is prolonged by a slow Ca^{2+} inward current. Afterwards, repolarizing delayed rectifier K^+ channels restore the resting membrane voltage. The duration of an atrial AP ranges between 200 and 300 ms. The differences in ion concentrations of Na^+ and K^+ between intra- and extra-cellular space are maintained by energy-consuming ion pumps and exchangers.

The pacemaker of the whole heart is the SN, where specialized cells spontaneously activate the surrounding tissue. There, a hyperpolarization-activated funny current I_f in combination with other currents as e.g. the sodium calcium exchanger slowly depolarize the membrane, which results in the initiation of an AP [129]. In the different regions of the right and left atrium, AP shape and duration (APD) are heterogeneously distributed. Differences between the APs of the RAA, CT, PM and atrio-ventricular ring as well as between RA and LA were investigated in canine atria [56, 134]. Furthermore, AP parameters of

human atrial myocytes such as, e.g., APD_{90} or effective refractory period (ERP) decrease with shorter diastolic intervals [111, 265].

3.2 Electrophysiological models

On the cellular level, the mathematical description of cardiac electrophysiology is mostly based on the work of Hodgkin and Huxley [91], who introduced a mathematical model of neural cells in 1952. Depending on the model's level of detail, several non-linearly coupled ODEs describe the cell membrane and the different ion concentrations. The change of transmembrane voltage V_m is calculated as follows:

$$dV_m/dt = -(I_{\text{ion}} + I_{\text{stim}})/C_m, \quad (1)$$

where I_{ion} is the sum of ion currents flowing through channels, pumps and exchangers across the membrane, I_{stim} an externally applied stimulus current and C_m the membrane capacity. The different ion currents I_x are computed using conductivities g_x scaling the amplitude and the product of gating variables γ_i describing the open probability of the channels depending on V_m :

$$I_x = g_x \prod_i \gamma_i (V_m - E_{\text{Nernst},x}), \quad (2)$$

$$\frac{d\gamma_i}{dt} = \alpha_{\gamma_i}(1 - \gamma_i) - \beta_{\gamma_i}\gamma_i, \quad (3)$$

where $E_{\text{Nernst},x}$ is the reversal potential of the ion type x and α_{γ_i} and β_{γ_i} are rate constants describing the transition from closed to open state and vice versa. Furthermore, intracellular calcium handling including formulations for the Ca^{2+} uptake and release of the sarcoplasmic reticulum are considered in current models. The setup and solution of the equations used in the models is shown more detailed in Ref. [59].

3.2.1 Modeling human atrial electrophysiology

The first models of human atrial electrophysiology are the ones published by Courtemanche et al. [31] and Nygren et al. [168] in 1998. Both models are mostly based on the same human experimental data. However, some of the formulations depend on experimental data of other mammals, where human data are missing. Although they have a similar base, the APs of the models have a different morphology (compare Fig. 2a). The Courtemanche model shows a spike-and-dome morphology, whereas that of Nygren et al. presents a more triangular shape. Furthermore, the APD_{90} restitution curves of both models differ (see Fig. 2b). At high basic cycle lengths (BCLs), the APD of the Courtemanche model is almost 70 ms longer and it decreases towards shorter BCLs, whereas that of the Nygren model decreases towards a BCL of 470 ms and then increases again. The differences and similarities of both models were compared in detail in e.g. Ref. [27, 169]. The Nygren model was refined by Maleckar et al. [148] in 2008, with a focus on improving the rate dependence of the AP and the formulations of repolarizing currents. The AP of the Maleckar model is similar in shape and duration to that of Nygren. The APD_{90} restitution curve is also similar to that of the previous model, although the APD is shorter

and stagnates towards shorter BCLs. Also based on the Nygren model, Koivumäki et al. [119] added a more detailed atria-specific description of the sarcoplasmic reticulum considering spatial effects of Ca^{2+} uptake and release in 2011. As a consequence, the AP mainly differs from the Nygren model in the first half of the plateau phase. The APD restitution curve has a similar shape as the Courtemanche model although the APD is shorter in the Koivumäki model. Also in 2011, Grandi et al. [68] published a human atrial model that also focuses on intracellular Ca^{2+} dynamics. This model furthermore considers the impact of β -adrenergic and cholinergic stimulation on human atrial APs. Since it is based on a ventricular model of the same group [69], also the ionic bases for the differences between atrial and ventricular APs were investigated. The AP shows a similar shape as the Nygren-based models, but has an APD comparable to that of the Courtemanche model. The APD_{90} increases towards a BCL of 450 ms and then decreases again. The shape of the APs of the models and also the restitution properties can also be adapted to other models or experimental data, as e.g. shown in Ref. [230]. An overview of the model properties is given in Table 2.

3.2.2 Restitution properties

The models of human atrial electrophysiology show different restitution properties (see Fig. 3) of the conduction velocity (CV), ERP and WL ($\text{WL} = \text{CV} \cdot \text{ERP}$) when analyzed in a 1D tissue patch. These restitution curves are good indicators of the stability of reentrant circuits [112]. The intracellular conductivity is adjusted so that a CV of approximately 75 cm/s is obtained at a BCL of 1 s. In general, the CV decreases towards shorter BCLs. The Grandi model presents a rapid decline of the CV down to a BCL of 550 ms. Only the Courtemanche model presents an increase towards 500 ms and then decreases again. In contrast to the other models, the ERP of this model decreases towards short BCLs. The ERPs of the Nygren,

Fig. 2 **a** Action potentials and **b** APD_{90} restitution of the different models of human atrial electrophysiology

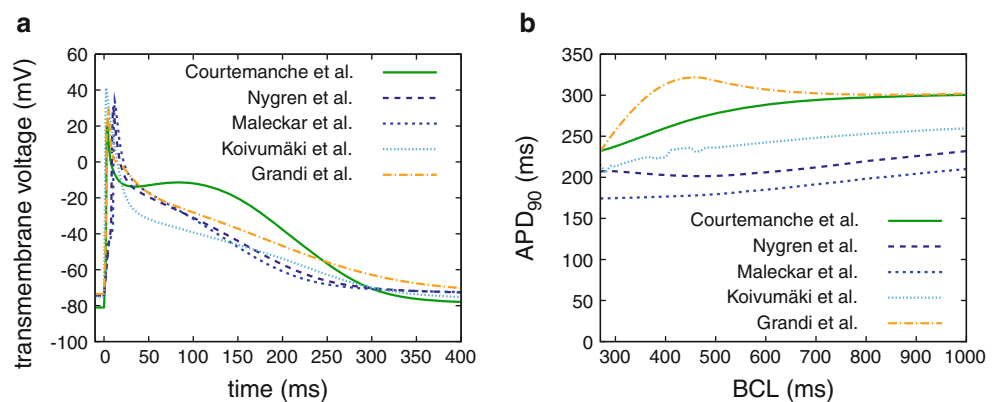
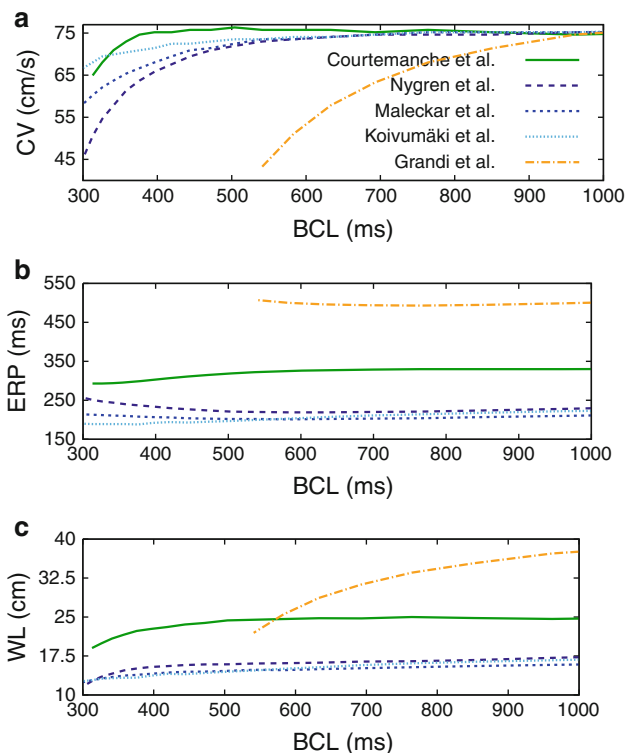


Table 2 Properties of models of atrial electrophysiology

Author	Reference	Transmembrane currents	Ca ²⁺ Compartments	Intended investigations	Extensions	Parent model
Courtemanche et al.	[31]	Common ^a	Intracellular, extracellular	Rate-dependent adaptation of human atrial AP, block of I_{CaL} and I_{NaCa}	AF: [27, 32, 242, 268], long-term stability: [172]	[144]
Nygren et al.	[168]	Common ^a	Intracellular, dyadic cleft, cleft space ^b , extracellular	Differences between rabbit and human atrial electrophysiology, especially repolarizing currents	AF: [27, 268]	[138]
Maleckar et al.	[148]	Common ^a + acetylcholine-activated K ⁺ current I_{KACH}	Intracellular, dyadic cleft, cleft space ^b , extracellular	Rate-dependent repolarization of human atrial AP, coupling between myocytes and fibroblasts	acetylcholine: [148]	[168]
Koivumäki et al.	[119]	Common ^a + hyperpolarization-activated inward K ⁺ current I_f	Four spatially varying intracellular spaces, subsarcolemmal, extracellular	Detailed description of calcium dynamics and impact on AP morphology		[168]
Grandi et al.	[68]	Common ^a + I_{KACH} + Cl ⁻ currents (background I_{bCl} , Ca ²⁺ -activated $I_{Cl(Ca)}$)	Intracellular, dyadic cleft, subsarcolemmal, extracellular	Differences between human atrial and ventricular electrophysiology with focus on calcium handling	AF: [68], β -adrenergic: [68], acetylcholine: [68]	[69]

^a Common to all models: fast Na⁺ current I_{Na} , L-type Ca²⁺ channel I_{CaL} , transient outward current I_{to} , ultrarapid delayed rectifier K⁺ current I_{Kur} , slow and rapid delayed rectifier K⁺ current I_{Ks} and I_{Kr} , inward rectifier K⁺ current I_{K1} , Na⁺ and Ca²⁺ background currents I_{bNa} and I_{bCa} , Na⁺/Ca²⁺ exchanger I_{NaCa} , Na⁺ – K⁺ pump current I_{NaK} , sarcolemmal Ca²⁺ pump current I_{pCa}

^b Cleft space: cell surrounded by restricted space, where ions accumulate or deplete

**Fig. 3** **a** CV, **b** ERP and **c** WL restitution of the different models of human atrial electrophysiology

Maleckar and Koivumäki models are much shorter. The ERP of the Nygren and Maleckar model even increases towards short BCLs. The highest ERP is obtained by the Grandi model, which therefore can only be computed to BCLs down to 550 ms. The resulting WL decreases in all models towards shorter BCLs. The WL of the Grandi model is longer than that of the Courtemanche model. The Nygren-based models have the shortest WL indicating a more probable initiation and persistence of arrhythmias.

3.2.3 Atrial heterogeneity

In several computational studies, heterogeneities of atrial electrophysiology were investigated. However, all of these studies were based on animal experimental data, e.g. [56, 134]. In Ref. [186], a canine atrial model considering heterogeneous APs of the RAA, CT, PM and atrio-ventricular ring was developed. The same experimental data were used for the integration of these heterogeneities into the human atrial model of Courtemanche et al. in Ref. [219] and into the model of Nygren et al. in Ref. [235]. In Ref. [273], the human Courtemanche model was adapted to APs of the canine CT and a modified version of the Luo-Rudy model [185] was adapted to APs of the canine PM. Zhao et al. altered the model of Courtemanche et al. to reproduce APs from the CT [273]. In a recent computational study [6], the Courtemanche model was

modified to simulate different APs in the RAA, CT, PM and atrio-ventricular ring, as well as in the LA.

3.2.4 Sinus node

Although very detailed models of the SN of rabbits are reported [269], only few models of human SN exist. In Ref. [219], the Courtemanche model was modified based on a heterogeneous model of the rabbit SN [269]. Furthermore, Aslanidi et al. [6] used modifications of the Courtemanche model published by Chandler et al. [23] to simulate human SN APs.

4 Excitation propagation

4.1 Propagation properties

The myocardium is built mainly of discrete cardiomyocytes. These myocytes are surrounded by the extracellular space and are coupled electrically by gap junctions. A transmembrane voltage difference in adjacent cells generates a current through gap junctions and through the extracellular space in such a way that the activation can spread. Both the intracellular and extracellular conductivities show anisotropic behavior allowing a faster excitation propagation along the main axis of the cell than across.

4.1.1 Activation pattern during sinus rhythm

The physiological excitation of the heart driven by the SN is called sinus rhythm. The frequency adaptation of the SN is controlled by the autonomous nerve system and by hormones [15]. After activation of the SN, the fast right atrial pathways, i.e. CT and PM, conduct excitation preferentially towards the atrioventricular region. During that phase the RA becomes depolarized [13]. At the same time, BB conducts the excitation towards the LA. Finally, the whole atrium gets depolarized after approximately 100 ms. All conduction pathways are characterized by a large conductivity along the fiber direction leading to a fast conduction velocity along their main axis.

4.1.2 Conduction velocity and anisotropy

The measured longitudinal conduction velocity (CV) in the atrium lies between 51 and 120 cm/s for non-AF patients (see Table 3). Under pathological conditions the variance increases to values between 37 and 133 cm/s. With age, the CV decreases [120]. In general, the CV along the conduction bundles is faster than in the working myocardium. Only little is known about anisotropic features of the CV. Spach et al. [227] measured an averaged ratio of 4.76:1 in the human atrium, but it is probably less pronounced since

this measurement included conduction pathways. In CT, an anisotropy ratio of around 8:1 was described [201].

4.2 Modeling excitation propagation

Commonly, excitation conduction is calculated with macroscopic approaches describing the atrial tissue as a continuum [6, 35, 80, 95, 113, 160, 188, 199, 219, 233, 249, 251]. One computational node is reproducing the properties of several real myocytes. The used models can be divided into three degrees of complexity: rule-based methods (i.e. cellular automaton), simplified propagation models (e.g. the shortest path algorithm and eikonal models) and models of the electrical current combined with detailed electrophysiological descriptions (e.g. mono- and bidomain models).

4.2.1 Rule-based method

Cellular automata (CA) have a finite number of states to describe electrophysiological properties. Very early studies used this approach [160] as the computational cost is very low. Newer CA can consider tissue anisotropy and appropriate wavefront curvature [199] as well as rate-dependent APD adaptations [188]. The disadvantage of the CA is the limited possibility to adjust electrophysiologically relevant parameters and the lack of electrotonic interactions.

4.2.2 Simplified conduction methods

The shortest path algorithm is the most simple propagation model as it does not include any electrophysiological state. In combination with a precise geometrical model, it can be used to illustrate routes of excitation [35]. Eikonal models are still simplified approaches but consider anisotropy and electrophysiological states [42, 122]. These models can be implemented efficiently using e.g. a multi-front fast-marching algorithm. Eikonal models have been used recently to investigate reentrant behavior of atrial tissue [95]. Also here the disadvantage is that cellular measurement data can poorly be integrated into the model.

4.2.3 Detailed reaction-diffusion models

Mono- and bidomain models consider the electrical current of ions between adjacent cells [29, 82]. The bidomain model is taking into account the current in intra- and extracellular space as well as through gap junctions and the cell membrane. It is based on the two Poisson equations:

$$\nabla \cdot (\sigma_e \nabla \Phi_e) = -\beta I_m - I_{se} \quad (4)$$

$$\nabla \cdot (\sigma_i \nabla \Phi_i) = \beta I_m - I_{si} \quad (5)$$

Table 3 Conduction velocities reported in the literature from measurements in human tissue

Study	Subjects	Region	Longitudinal (cm/s)	Transversal (cm/s)	BCL (ms)	Condition
[55]	9	septum	58–61		300–600	H
		RA FW	58–64			
		isthmus	51–55			
[79]	12	RA FW	88		SR	H
[136]	21	RA septum	96–98		200–500	H
		RA FW	94–100			
[227]	49	PM	70	8.2	800	H
[45]	20	RA	120		SR	H
	20	LA	120		SR	H
[105]	16	RA FW	83		1006	CAB
			77		500	
[55]	9	septum	50–54		300–600	Aflut
		RA FW	53–57			
		isthmus	37–42			
[136]	25	RA septum	86–91		200–500	Aflut
		RA FW	91–98			
[212]	8	isthmus	77		240	Aflut
		RA smooth	108			
		PM	133			
[222]	9	isthmus	70		243	Aflut
		RA sup	120			Aflut
		RA lat	110			Aflut
		RA septum	110			
[45]	20	RA	80		SR	OSA
		LA	90			
[255]	5	RA, LA post & roof	108		SR	AF/FAT
		RA, LA post & roof	86		500	
		RA, LA post & roof	77		300	

FW free wall, Aflut atrial flutter, FAT focal atrial tachycardia, OSA obstructive sleep apnea, H no arrhythmia, CAB coronary artery bypass.

with the intra- and extracellular potentials Φ_i and Φ_e , respectively, the corresponding conductivity tensors σ_i and σ_e , the transmembrane current density I_m , the membrane surface to cell volume ratio β , and externally applied current sources I_{si} and I_{se} in the intra- and extracellular domain, respectively. I_m is the sum of capacitive and ionic membrane currents:

$$I_m = C_m \frac{dV_m}{dt} + I_{ion} \quad (6)$$

with I_{ion} being the sum of all transmembrane currents I_x from Eq. 2.

Under the assumption of equal anisotropy ratios, this model can be reduced to the monodomain model calculating the current through the intracellular space and through gap junctions:

$$\nabla \cdot (\sigma \nabla V_m) = \beta \left(C_m \frac{dV_m}{dt} + I_{ion} \right) \quad (7)$$

In both the cases, the model of current is connected in each computational node to detailed electrophysiological models of e.g. Courtemanche or Nygren. Most of the atrial modeling studies have used the monodomain model [6, 80, 113, 219, 233, 249, 251]. Figure 4 illustrates the propagation simulation on a schematic 2D patch and on a realistic 3D geometrical model using the monodomain method. The disadvantage of mono- and bidomain modeling is the large computational cost and the large number of parameters in the model that cannot be determined with sparse measurement data.

4.2.4 Conduction velocity and anisotropy

Many modeling studies lie within the range of measured conduction velocities (36 to 130 cm/s) and also consider the velocity differences in different tissue types (e.g. isthmus 36 cm/s, atrial myocardium 65 cm/s, bundles 210 cm/s in [66]). In addition, anisotropy ratios of the CV have been

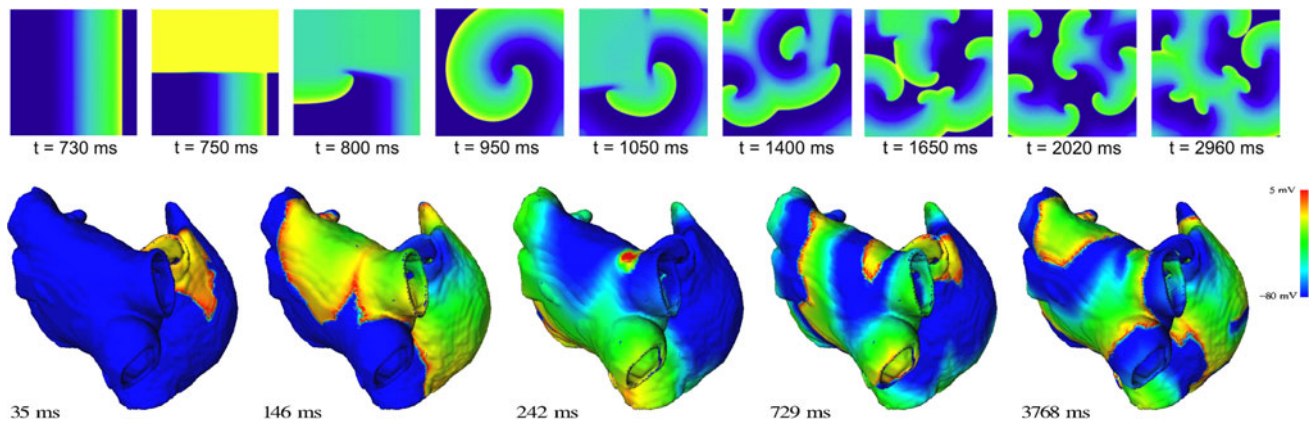


Fig. 4 *Top row* simulation of AF rotors in a 2D patch of atrial tissue [218]. Rotors are generated using a cross-field stimulus protocol. *Bottom row* example of simulation of AF in a 3D model. AF is triggered by ectopic activity in the RSPV (241 ms) after initial sinus rhythm (35 ms)

set e.g. to 1.9:1 [6], 2.4:1 [113, 133] or 3:1 [266] in the working myocardium and to e.g. 6:1 [262] or 9:1 [100, 219] for the conductivity in the conducting pathways. The working myocardium is often treated isotropically since fiber orientation information in the human atrium is rarely included in the models (Table 3).

4.2.5 Stimulation of tissue

In order to simulate sinus rhythm, the reported approaches are either to apply an intracellular stimulus current in the region where the SN is located [80, 251] or to set nodes close to the SN to be self-excitatory cells [6, 219]. A crossfield stimulation can be used to generate a single rotor in the tissue (see Fig. 4 upper row, left). Here, a first planar excitation is initiated and then in the half refractory/half excitable period a perpendicular second current is applied [218]. In order to generate multiple reentrant waves, pulse trains can be used reproducing ectopic focal activity. These pulse trains can be with constant position and rate e.g. in the pulmonary vein [188] or vary in position and rate to more easily produce multiple reentries (see Fig. 4 upper row, right).

5 ECG

5.1 Computation of ECG and electrograms

Body surface potentials are commonly computed in a two-step approach. In a first step, transmembrane voltages are simulated in the atrial models using an arbitrary excitation propagation model (see Sect. 4 excitation propagation). From the transmembrane voltages V_m the impressed current sources I_{imp} can be calculated

$$I_{\text{imp}} = -\nabla \cdot (\sigma_i \nabla V_m). \quad (8)$$

In a second step, Poisson's equation as the parabolic part of the bidomain formulation [82, 184] is solved

$$\nabla \cdot [(\sigma_i + \sigma_e) \nabla \Phi_e] = I_{\text{imp}}. \quad (9)$$

Thereby Dirichlet and Neumann boundary conditions are set for the reference potential and the thorax-air boundary. Considering these conditions, the bidomain formulation is transformed into a linear set of equations using either the finite element method (FEM) (e.g. [109, 125, 259]) or the boundary element method (BEM) (e.g. [142, 171]). The FEM has the advantage that it can consider anisotropy within the organs (e.g. skeletal muscle fiber orientation), whereas the BEM is computationally more efficient. Measurements of the organ conductivities show a large variability leading to some uncertainties in solving the forward problem [109]. ECG signals can then be extracted at surface nodes of the thorax model. In order to compare simulated and measured signals, the measurement electrodes need to be tracked with respect to the torso model during ECG recording [123].

Intracardiac electrograms can be simulated using the methods described above [256] or by computing pseudo-electrograms

$$\Phi_e(r, t) = \frac{1}{4\pi\sigma_e} \int \left(dr' \frac{I_m(r', t)}{|r - r'|} \right), \quad (10)$$

where r is the electrode location vector, r' the current source location vector and I_m the transmembrane current per unit area of tissue surface [236, 251].

5.2 Characteristics of the P- and Ta-wave

The atrial depolarization sequence can be seen in the body surface ECG as the P-wave. The signal of the P-wave is strongest in the Einthoven II lead and shows a Gaussian-like shape with a maximum amplitude of 0.25 mV. The P-wave has a smooth contour and is monophasic positive in

lead II, inverted in aVR and often biphasic in V1 [216]. Left atrial enlargement may cause a bifid P-wave (P mitrale) and right atrial enlargement may cause a peaked P-wave (P pulmonale) [216]. Inversion of the P-wave is caused by ectopic triggers and a variable morphology is caused by multifocal arrhythmias [216].

The duration of the P-wave (PWD) reflects the time of complete atrial depolarization. AF patients often have a longer P-wave duration (120 ms) compared to controls (100 ms) [44, 71], which can be used to detect AF based on the P-wave duration [39]. Nevertheless, automatic detection based on P-wave analysis shows only moderate specificity and sensitivity [28, 63]. The integrated body surface potential maps (BSPMs) of the P-wave show a dipole on the front chest from the right shoulder to the left hip with the zero potential line crossing from the left upper chest to the lower right chest [135, 158, 224]. The body surface potential distribution can vary depending on the site of the origin of atrial activity [224].

Atrial repolarization is usually not visible in the body surface ECG, as the repolarization signal is occluded by the much larger QRS complex due to the ventricular depolarization. In patients with third degree AV block, the repolarization signal can be measured. The polarity of the so-called Ta-wave is inverted compared to the P-wave and has a smaller amplitude [92].

Right-to-left atrial conduction varies between individuals [13, 90, 93, 132, 151, 179]. The most common sites of septal breakthrough are over Bachmann's bundle at the anterior side of the LA, the musculature of the CS and the region near the fossa ovalis [13, 38, 93, 131]. The individual interatrial conduction can be deduced from the polarity of the vectorcardiogram (VCG) [93].

5.3 Simulation of atrial ECG

Only few groups have simulated body surface ECG from bi-atrial electrophysiological simulations. The first efforts go back to the early 1990s, when Kafer examined interatrial conduction in the VCG with a computer model [103]. Later on, the first four chamber simulations with ECG computation became computationally feasible [141, 259].

Van Dam, van Oosterom, and Jacquemet computed the P-wave and concentrated on volume conductor effects [36] and showed that an equivalent double layer source model was sufficient to reproduce a measured P-wave [171]. Later on, Jacquemet et al. examined the potential of the VCG to characterize AF using the methods introduced earlier [98].

Recently, more groups started to simulate the P-wave and body surface potentials from atrial depolarization. In two patient models the atrial ECG was computed. The generated P-waves were compared to measured P-wave signals from the same patients [123]. Figure 5 provides an

example of the comparison between simulated and measured atrial electrical signals, which correspond on multiple scales. The torso and atria were segmented from MRI data of the patients and a number of organs in the torso were distinguished. Lu et al. [142] were able to compute a reasonable P-wave using a generic homogeneous thorax model. Similar work was done by Aslanidi et al. [6], although they incorporated the lungs into their model and utilized a different mathematical approach.

Furthermore, the computation of the P-wave was recently used to test influences of clinical therapies. Thereby the influence of hemodialysis therapy on atrial electrophysiology in a heterogeneous atria and thorax model was evaluated and a P-wave prolongation due to the therapy was revealed [125] (see also Sect. 8.2). In this study, also the atrial repolarization was less uniform during hypokalemia resulting in a larger Ta-wave amplitude.

The influence of the conductivity of different organs on the computed P-wave was evaluated, showing that muscle, blood, lungs and fat were the organs with the greatest sensitivity for the P-wave morphology [109]. Uncertainties in the organ conductivities can be predicted using a fast principal component analysis (PCA) based method [254].

6 Patient measurements for model validation

As described above, electrophysiological models are typically built on the ion channel level, based on voltage clamp or patch clamp techniques. Wherever available, human data are used, but the remaining channels are modeled from animal data. Although in many cases animal data may give a good approximation for humans, several uncertainties remain. Apart from that, a good parametrization on the single-cell level does not guarantee that the model resembles macroscopic excitation patterns in the atria, because coupling between the cells may influence excitation propagation. Therefore, it is of interest to obtain macroscopic measurements from patients or healthy subjects, and if possible also to compare them to properties of atrial models.

Patient data of interest are frequently acquired during EP studies, especially electrograms measured with multi-electrode catheters. The first step is to quantitatively analyze such data. In a second step, such measurements can be compared to model simulations. Furthermore, optical measurements allow for visualizing excitation in tissue patches, but are restricted to animal experiments so far.

6.1 Substrate characterization from electrophysiological studies

Electrophysiological studies are performed by inserting one or several catheters into the atria, most of which have

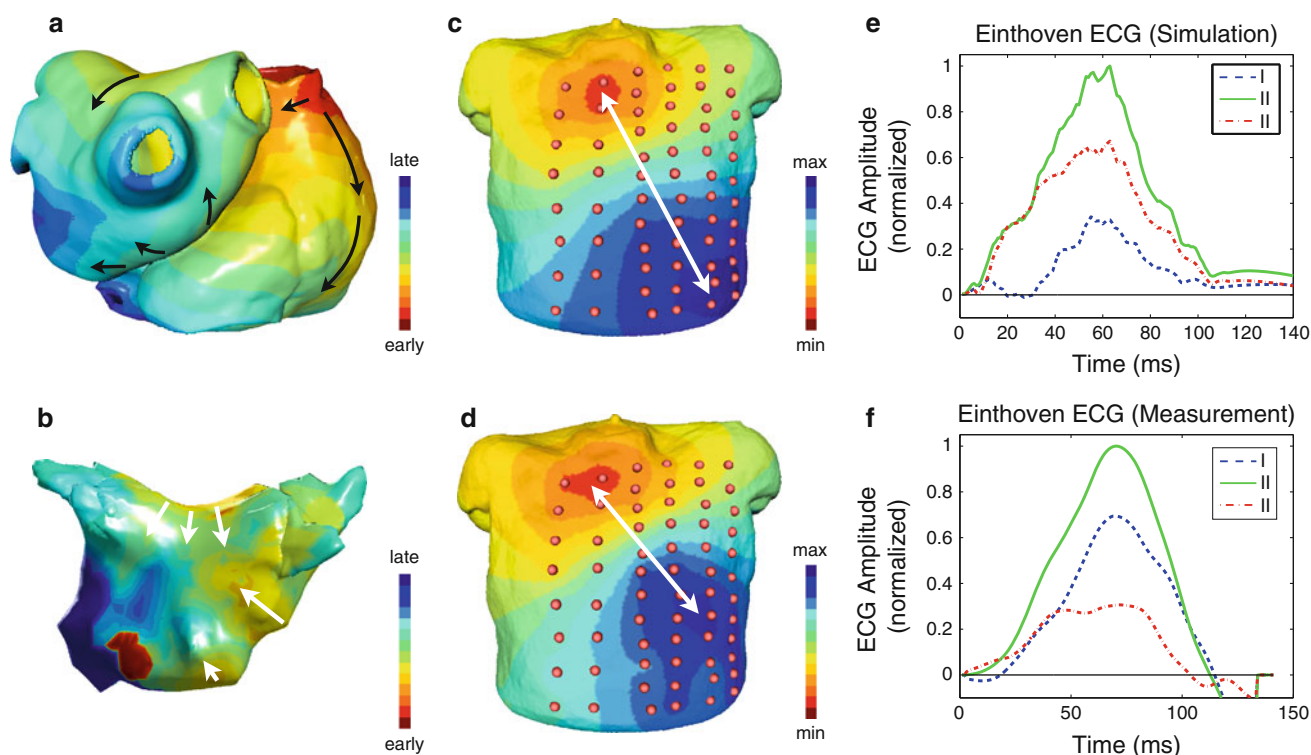


Fig. 5 **a** Simulated activation times in an atrial model. *Black arrows* indicate preferred conduction routes. **b** Measured local activation times in a left atrium. *White arrows* indicate preferred conduction routes. **c** Body surface potentials integrated over a simulated P-wave. *White arrow* marks the dipole from atrial excitation. **d** Body surface potentials integrated over a measured P-wave (same subject as in (c)).

White arrow marks the dipole from atrial excitation. **e** Simulated Einthoven ECG (*dotted blue* lead I, *solid green* lead II, *dashed red* lead III). **f** Measured Einthoven ECG (same subject as in (e)). The simulation approach is similar to the one used in Refs. [123, 125], but data were acquired from different individuals here

multiple electrodes for recording electrical activation of the surrounding tissue [2, 213, 223]. It is further possible to apply a stimulation current via certain electrodes to induce an excitation wave, if the underlying tissue is not refractory. From such measurements, relevant data for atrial modeling can be obtained.

Catheters do not measure transmembrane voltages, but the extracellular potential difference between two electrodes that shows up during depolarization at the wave front. There are techniques to estimate APD from such intracardiac electrograms using the activation recovery interval [157]. However, because repolarization in the atria is slower than in the ventricles, repolarization currents are weaker, which makes the method more challenging in the atria. Vigmond et al. demonstrated an accuracy of around 40 ms for atrial APDs [250].

For simulating arrhythmias, it is even more important to determine the ERP. This is done during electrophysiological studies by programmed S1–S2 stimulation. An initial train of driving stimuli at cycle length S1 is followed by a shorter coupling interval S2. At a distant location, it is determined if the S2 pulse generated an excitation wave. In this case, the stimulation protocol is repeated with a shortened S2 until the

S2 pulse falls into the refractory period. This way, atrial ERPs of 233 ms for the LA and 226 ms for the RA were measured at an S1 cycle length of 440 ms [194]. Such measurements can be combined with measurements of other excitation properties, such as electrogram voltage, degree of fractionation, conduction time, or CV [45].

CV measurements during EP studies are normally approximated from interpolated local activation maps. From the time delay and spatial distance between two or more points along the steepest gradient, the CV is estimated [45, 120, 121, 211]. More advanced techniques are based on vector field analysis of activation maps or data from epicardial electrode arrays [9, 10, 60]. Interpolation of atrial activation time fields based on sparse data was performed using radial basis functions [154]. Neither beat-to-beat variations in CV nor changes in the excitation pattern during map acquisition are considered because the activation map is recorded over many cardiac cycles. CV measurements on single wavefronts in the human RA were performed using epicardial electrode arrays during open-heart surgery [79]. A single-shot measurement of CV during EP studies was demonstrated for plane wavefronts using a circular mapping catheter [256]. With the same

technique, CV restitution was determined from pacing at different cycle lengths [255].

6.2 Comparison of models and clinical measurement

With patient data available, the outcome of atrial simulations can be compared to the measured data. Total activation times of the atria were simulated in a monolayer model generated from an MRI scan of a single patient. Simulated activation times for stimulation at different locations were comparable to mean P-wave durations from nine subjects [67, 262]. The average deviation over all stimulus sites was 8 ms. Stimulation protocols from electrophysiological studies were reproduced *in silico* on isotropic anatomical geometries derived from CT scans [16]. The propagation direction at catheter measurement sites in the LA deviated by 15° on average.

6.3 Optical mapping

Excitation in cardiac tissue can be visualized using voltage-sensitive fluorescent dyes [51, 52]. The fluorescence signal depends on the local transmembrane voltages and thus allows to image relative changes in tissue excitation. The dye is typically excited by a laser in the 532 nm range, and the resulting fluorescence signal is recorded by a CCD camera. With such data available, simulated excitation propagation can be compared to measurements.

Cherry et al. [26] demonstrated optical mapping of AF in tissue preparations from isolated perfused canine atria. Under the influence of acetylcholine, a sustained mother rotor was observed and compared to simulated data. Arora et al. [4] visualized arrhythmogenic substrate in pulmonary vein preparations with conduction slowing in the proximal part and reentrant loops. Umapathy et al. [247] mapped stable rotors in monolayers of murine atrial tissue. Such studies are helpful to validate the dynamic evolution of 2D and 3D atrial simulations.

Kalifa et al. [104] presented a steerable cardio-endoscope for fluorescence mapping in the intact LA. Different propagation patterns were recorded in sheep hearts that had undergone a stretch-induced AF model. Although such measurements in the intact atrium are a step towards clinical patient measurements with optical mapping, a critical limitation is the phototoxicity of currently available fluorescence dyes [104].

7 Modeling atrial pathologies

The major application of human atrial models is to investigate atrial flutter and fibrillation. In 2008, Jacquemet et al. have published a comprehensive review article about the numerical modeling of atrial arrhythmias [97]. Since

then, further atrial arrhythmia modeling work considering realistic atrial geometry was published [6, 33, 142, 178, 188, 191, 233–236]. In this review article, we want to focus on modeling the changes in electrophysiology and structure favoring the initiation and continuation of atrial arrhythmias. Therefore, we present some examples how modeling of pathologies is done and what kind of influence it has on cellular electrophysiology and excitation propagation. The results of these modeling studies can be used to cross-validate measurement results or to test hypotheses.

7.1 Remodeling

The effects of sustained AF can be e.g. severe congestive heart failure, thromboembolism, cardiomyopathy, ventricular arrhythmia and remodeling of the atria. This remodeling can be electrical, structural and contractile [162]. Electrical remodeling leads to a reduced APD due to a calcium overload compensation mechanism. The structural remodeling mainly reduces the CV. Both of these remodeling effects lead to a reduced wavelength favoring the continuation of AF. Contractile remodeling reduces the active tension development in atrial cells. Schotten et al. presented recently a comprehensive review article about underlying mechanisms of AF [215]. The reduction of APD is based on an increase of I_{K1} by around 110 % and a decrease of both $I_{Ca,L}$ and I_{to} by around 65 %. The structural remodeling decreases gap junction density reducing the intracellular conductivity by up to 30 %.

These values for changed atrial electrophysiology were implemented by several modeling groups, e.g. [218, 236, 267], to investigate the effects of remodeling on atrial electrophysiology, excitation propagation and vulnerability. As an example, the Courtemanche model in conjunction with the monodomain model was used to reconstruct the remodeling effects and simulate re-entry mechanisms in a schematic 2D model [218]. Both electrophysiological and structural remodeling was implemented based on literature values. The APD in tissue was reduced from 322 ms in the control case to 144 ms under remodeling. The CV was decreased from 71.5 cm/s in the physiological case to 54 cm/s. The tissue simulation showed that re-entrant patterns can be generated only for the case of remodeling and cannot be sustained in the normal atrial tissue.

7.2 Fibrosis

Atrial fibrosis increases with age and develops in conjunction with cardiomyopathy and heart failure. Fibrosis has been linked to an increased probability of arrhythmias, including AF [117]. Fibrosis is characterized by both an expansion of the extracellular space and an increase in the number of cardiac fibroblasts. The increase in extracellular

matrix reduces the conduction but only little is known about fibroblasts. As fibroblasts have mostly passive electrophysiological properties but can express functional gap junctions with myocytes, conduction can also be altered by fibroblasts [117].

Several modeling studies illustrated the impact of fibrosis on atrial electrophysiology and conduction as well as on ECGs [96, 147, 178]. Plank et al. [178] showed in a realistic atrial anatomy that increased anisotropy in the atria due to fibrosis can be responsible for the breakup of PV ectopic waves into multiple re-entrant circuits. Maleckar et al. [147] coupled a human atrial myocyte to a variable number of fibroblasts and investigated the effect of altering the intercellular coupling conductance, electrophysiological fibroblast properties, and stimulation rate on the atrial AP. The results demonstrate that the myocyte resting potential and AP waveform are modulated strongly by the properties and number of coupled fibroblasts, the degree of coupling, and the pacing frequency. Jacquemet et al. developed a 2D model of atrial tissue including microfibrosis incorporated as a set of collagenous septa disconnecting transverse coupling [96]. The density and length of these septa were varied. The analysis of unipolar electrograms showed that the septa decreased CV by up to 75% and increased the amount of fractionation and asymmetry of the electrograms.

7.3 Effects of vagal activity

Vagal nerve stimulation releases acetylcholine (ACh) as a transmitter to the whole atrium. The main effect of ACh is to slow down heart rate by decreasing the SN activity. Furthermore, ACh-dependent potassium channels also exist in atrial working myocardium. If ACh is released, the APD shortens. As the ACh release was measured to be inhomogeneous [114], APD differences can exist throughout the atrium due to vagal stimulation.

Kneller et al. [114] combined their measurements with a computational study to investigate the arrhythmogenic effects of these ACh-induced inhomogeneities of the APD. A 2D patch including periodic changes in ACh concentration was used to investigate the effects of vagal stimulation on the reentrant properties. It was found that one initial stable rotor can breakup into three re-entries under influence of ACh because the heterogeneity of refractoriness generates inhomogeneous wavefront-waveback interactions. Thereby, the model was able to support the hypothesis that vagal stimulation can have an effect on arrhythmia generation.

7.4 Pathological dilation of the atrium

Stretch-activated channels might play a role in the initiation and continuation of AF [270]. Under pathological

condition, the atrium can be dilated and therefore be under a constant stretch.

Two modeling approaches have shown the potential impact of dilation to the initiation and perpetuation of AF [127, 198]. Ruiz-Villa et al. [198] showed that the vulnerability to ectopic beats is higher using a dilation model than a purely electrophysiological remodeling approach. Three different ectopic beat locations were investigated and the resulting arrhythmia properties corresponded well to measured data. Kuijpers et al. [127] simulated acute atrial dilation by increasing stretch throughout the atrial wall. This stretch was heterogeneously distributed because of geometrical effects and resulted in a higher dispersion of ERP, a slowing of conduction and local conduction block through stretch-activated currents. All these factors were shown to favor the existence of AF.

7.5 Genetic defects

Additional to other initiating factors, AF has been shown to be a heritable disorder. Recent studies have identified several genetic defects affecting atrial ion channels (e.g. [25, 155]). Chen et al. [25] studied a family with persistent AF. The family had a mutation (S140G) in the *KCNQ1* gene, which forms the α -subunit of the I_{Ks} channel. The mutation caused a gain-of-function effect, an instantaneous activation and deactivation as well as an almost linear current–voltage relationship. McPate et al. [155] measured the properties of the mutation N588K in the gene *KCNH2*, which encodes the hERG protein. This protein forms the α -subunit of the I_{Kr} channel. This mutation also causes a gain-of-function effect.

For both mutations, a computational equivalent was developed based on the Courtemanche model [217, 220]. To this end, the model parameters were adjusted to the measured voltage-clamp data defining the difference between measured and simulated ion current response to voltage steps and minimizing this difference. In case of the I_{Ks} mutation it was shown that APD_{90} was reduced to 35 % of the physiological value leading to a significant decrease in wavelength [220]. The I_{Kr} channel mutation also showed a reduction in APD (62 %) [217]. Interesting here was that the ERP in tissue showed a less pronounced reduction (27 %) as it depended on the maximum upstroke reached during depolarization leading to a less-pronounced increase of the mutant I_{Kr} in tissue compared to single cell. Nevertheless, the wavelength was also in this case reduced building a substrate for AF.

8 Towards clinical applications

The usage of atrial models has various faces. In the previous section, research applications of atrial models were presented, which help to gain a better understanding of

atrial pathologies. Another potential field of application is the usage of atrial models for a more therapy-oriented clinical research. In future, a goal could be to use computational models in clinical practice, as part of the therapy planning procedure. This section gives four examples for atrial modeling moving towards clinical application.

8.1 Improving RF ablation

Patients suffering from paroxysmal AF can be treated conventionally by drugs which suppress the symptoms of AF. Another option is the curative RF ablation therapy [20]. The minimally invasive ablation procedure was preceded by surgical procedures, such as the MAZE I–III. Although more frequently undertaken in the recent years, the success rate of ablation procedures is still variable and may differ between clinical centers and medical practitioners. Although for atrial flutter and paroxysmal AF high success rates are achieved, success rates for persistent AF are significantly smaller [20].

The treatment of AF with ablation is reviewed in Ref. [20]. Reviews about the biophysics of lesion formation during RF ablation can be found elsewhere [72, 73, 164] and modeling of the creation of a RF ablation lesion is subject to thermodynamical modeling [12].

8.1.1 Ablation lesion pattern models

Atrial models were used to find the best possible surgical pattern [196, 197, 234] and to evaluate different idealized ablation patterns on their ability to terminate AF [37, 54, 188, 195]. Some of these studies also introduced incomplete lesions to test the impact on AF recurrence [37, 188, 195, 196].

8.1.2 Simulation of gaps in RF lesions

Small gaps in ablation lesions show an isthmus-like conduction behavior. Cabo et al. [17] were able to show such slow conduction in lesion gaps both experimentally and in silico. They also discovered a distinct wave-front curvature caused by the isthmus in the lesion. Thomas et al. [231] found the behavior to depend on the size of the gap, which was confirmed recently in a combined measurement and simulation study [187]. Perez et al. [176] showed experimental evidence that conduction through discontinuities in ablation lesions is associated with gap geometry. Pop et al. [182] simulated the influence of conductivity and fiber orientation on the conduction through a gap between two scars in ventricular myocardium. Ranjan et al. [187] were also able to show a conductivity-dependent conduction behavior in silico.

8.1.3 Ablation lesions from LE-MRI

Late enhancement MRI (LE-MRI) using contrast agents provides a tool to image myocardium with reduced perfusion. Several groups have shown to image pathological regions prior to RFA [170] as well as acute and chronic scars after RFA [8, 116].

MacLeod et al. [146] highlighted the importance to include such information about structural changes of the atrial myocardium into geometrical models. Recently, first approaches have been undertaken to use patient-specific atrial models to evaluate and predict the outcome of RF ablation procedures in silico using information from LE-MRI [226]. An example for such an attempt can be seen in Fig. 6.

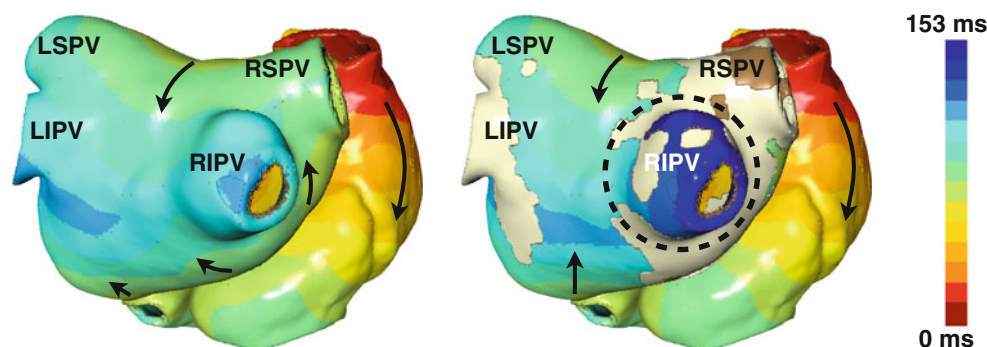


Fig. 6 Local activation times in a patient-specific model. *Left* Regular model. *Right* Model with ablation lesions derived from the same patient using LE-MRI. Data presented in Ref. [226]. *Right* inferior pulmonary vein is not completely isolated (*encircled*), whereas the right superior pulmonary vein is completely isolated

from left atrial tissue. Activation pattern of the left atrium changes after ablation, as activation over the upper and middle posterior interatrial bridges cannot propagate any further due to the ablation lesion

8.2 Hemodialysis

End-stage renal disease patients show a higher prevalence for suffering from AF [64]. Hemodialysis therapy compensates for loss of renal function, but also alters the concentrations of blood electrolytes. A hemodialysis session lasts for several hours, so it is reasonable to assume that with the blood electrolyte also the extracellular ion concentration in the cardiac tissue will change. It was shown that alterations in electrolyte concentrations before and after a hemodialysis session impact atrial electrophysiology, slow down excitation propagation, decrease WL and prolong the P-wave both in simulations and measurements [125, 221]. The modeling approach was thereby able to close the gap in the chain of evidence between single cell measurements and body surface ECG measurements and supported the explanation of the potential risk of pro-arrhythmic effects during hemodialysis therapy. In the future, the model could be used to find the appropriate dialysis solution to minimize the risk of developing AF.

8.3 Development of signal analysis techniques

Besides the presented applications to clinical research, atrial models bear the potential as development tool for improving signal analysis and measurement techniques. To validate such techniques, they can be tested *in silico* in a controlled environment with no unknown variables. Especially, the quantity or property to determine is well-defined in the model. Such a validation may be used to estimate possible errors, or improve the system before any clinical studies or animal experiments are carried out.

On a single-electrogram level, it was studied how intracardiac electrogram morphology depended on the angle of the catheter tip with respect to the myocardium. To this end, different orientations were simulated using the bidomain equations and compared to clinical measurements [181]. In an arrhythmia simulation with stable rotors and multiple wavebreaks, simulated electrograms, e.g. at the rotor tip, were compared to clinical complex fractionated atrial electrogram (CFAE) signals [163].

In the context of catheter activation patterns, atrial simulations were used to develop methods to quantitatively analyze intracardiac electrogram recordings from multi-spline catheters [153] and circular mapping catheters [255–257]. Richter et al. [189] validated a propagation pattern analysis for AF signals in two simulation scenarios before applying it to clinical data. A method for interpolating local activation times on the complete atria from sparse measurement data with the help of a simulation model was presented for a simulated case of typical atrial flutter [139].

With respect to AF cycle length, Haissaguerre et al. demonstrated in 3D atrial simulations that the AF cycle length was inversely related to the number of active trigger sources, and then applied the cycle length analysis to clinical data [76]. Sassi et al. [210] used simulated atrial ECGs to validate methods for determining the dominant AF frequency from the T-Q interval, i.e. without removing the ventricular signal.

8.4 Atrial overdrive pacing

Besides pharmaceutical management, AF can also be terminated electrically using cardioversion or antitachycardia pacing [115], so-called overdrive pacing. External pacing with a frequency higher than the tachycardia frequency will override the tachycardia, as the stimulation introduces refractoriness in areas that previously conducted the tachycardia. Success rates are variable, and computer models bear the potential to provide a better understanding of the mechanisms of overdrive pacing and may allow an optimization of the therapy. For example, simulation results of a whole heart revealed that atrial tissue properties such as CV and APD determine the response to the therapy [244, 245]. Atrial models could therefore in future be used to determine the proper pacing site and pacing cycle length [245, 246]. Low-energy pacing can establish intramural wave sources at heterogeneities in the fibrillatory substrate and thus directly target cores of the electrical turbulences [145]. Fenton et al. [58] were able to provide a better understanding of the formation of virtual electrodes due to pulsed far-field stimulation in a combined simulation and measurement study.

9 Perspectives

Although this review article illustrates the advances that have been made in the last years in the field of atrial modeling, it is obvious that we have not yet progressed far enough to enable the simple use of the models in all areas of research and clinical application. In the following, some emergent aspects are highlighted which need further focus in future.

9.1 Computer modeling for basic research

A better understanding of initiation and perpetuation of AF is a major issue of current research. We do not yet clearly know what type of changes appear in the atrial tissue before atrial flutter and AF emerge. It becomes evident that many different processes are responsible for AF and in different patients different aspects are dominant. Computer modeling of pathological atria will contribute to the answer

of many open questions. Altered heterogeneities and anisotropies of atrial tissue promote AF and they have to be included into modeling as realistically as possible. In particular, modeling atrial tissue on cellular level including extracellular matrix (collagen) and fibroblasts will play an important role, e.g. in highlighting the origin and characteristics of CFAEs and their correlation with fibrotic tissue. Furthermore, computational modeling can help to understand atrial remodeling and the slow process of reverse remodeling after a therapeutic intervention.

9.2 Integrating elastomechanics and flow

Modeling the ventricles is in the leading edge in terms of elastomechanics and flow, probably because the fiber orientation is known better and the walls are much thicker and thus easier to model. First attempts to model the elastomechanics of the atria have been published without any link to electrophysiology [41, 101]. By integrating elastomechanics and flow into the atrial model, the filling of the ventricles with blood can be investigated and the risk for clots leading to stroke for patients suffering from AF can be estimated. Also structural remodeling, e.g. as a consequence of hypertension or persistent AF, can be investigated with the support of elastomechanical modeling.

9.3 Personalization

A fast way to build computational models of the atria of an individual patient has to be found. A versatile clinical workflow has to be implemented that includes all measurements that are essential and only adds more diagnostic procedures if unavoidable. This chain will start with improved imaging techniques [150], move on with new segmentation tools, include BSPM and end with catheter-based measurements like electrograms with specific stimulation protocols. Modeling would have an important impact, if it could lead to significantly higher therapeutic success rates in particular in case of ablation of persistent AF. However, modeling will only be accepted in clinical practice, if the price is not a longer therapeutic procedure. Presumably, patients will be sorted into groups that fit best to their specific problem and treated along a new set of group-specific clinical pathways. Algorithms to adapt model parameters to patient data have to be improved [122, 139].

9.4 Electrocardiographic imaging (ECGI)

Solving the inverse problem of electrocardiology can produce images of electrophysiology like e.g. localization of ectopic foci, activation time maps, spatially resolved CV, and visualization of fibrotic regions. The method is

completely non-invasive, only a BSPM and an MRI thorax scan are needed. Unfortunately, the inverse problem is severely ill-posed in particular in the atria as the signal is lower than in the ventricles and the atria are more distant from the body surface. Promising preliminary results [34] have to be validated and new methods of regularization have to be discovered before ECGI can be used for parametrization of individual atrial models.

9.5 Drug research

The idea to support pharmacological research with computational modeling is appealing [167, 192]. Today, we encounter several problems e.g. that antiarrhythmic drugs most often change the properties of several ion channels and that the data needed for computational modeling may not be available. Nevertheless, first efforts have been undertaken to model drug effects in the atria [5, 204]. Specific information about the genetic profile of the patient can or must be included in the long run if the best drug is to be selected for an individual patient.

9.6 Ablation therapy planning

Today, generalized approaches to validate the success chance of various ablation lines using computational models have been presented. The future will be an individualized planning of ablation lines. In silico modeling of the atria must be reduced to the most essential parts since it must deliver a reliable result in real-time. Important information delivered by the models can only have impact in the EP-lab when the cardiologist gets the answer to some crucial question within a few minutes. Measurements of intraatrial electrograms with multielectrode catheters must find a straightforward way into the personalization of the model. Also ablation lines that have already been realized must be considered in the model immediately. If patients can be grouped into typical cases, e.g. using ECGI, these calculations can be done beforehand and only the case that fits best to the patient has to be selected in the EP-lab.

9.7 Prevention from atrial arrhythmias

A completely new aspect of computational modeling can be the prevention from AF. With modeling, we might be able to identify new biomarkers on different levels e.g. proteins, cell and tissue properties and in the BSPM of persons that show an increased risk for AF. In addition, specific drugs can be designed to hinder the onset of AF for those persons. Also, the models in combination with measurement studies can give hints to high risk people how to change their way of life, so that AF will not establish.

9.8 Merging imaging, electrophysiology and genomics

The authors of this review are convinced that on the long run computational modeling will be the best and may be even the only way to merge the information from imaging, electrophysiology and genomics so that all this information can be used in an optimal way for diagnosis and therapy planning.

10 Conclusion

This review has demonstrated that tremendous progress has been made in the last 10 years in modeling human atria. The contributions to research and comprehension of physiology and pathophysiology are obvious. The impact on clinical procedures is not yet as strong as envisaged. Pathologies of the atria are much more complex than expected by modeling experts. More and precise measurements on human atria are urgently needed. These data can help to select which of the currently available five cellular models fits best to reality and which one reproduces most accurately e.g. heterogeneity, restitution curves and remodeling. We need to learn how we have to model excitation propagation, so that we can replicate the real physiological and pathological case, e.g. fibrotic tissue. The community of computational modeling of the atria is encouraged to build a much stronger link to real measured data. Virtual atria with no reference to real measurements are of limited clinical use. However, continuously merging computational models with experimental and clinical data will significantly impact on a better comprehension, detailed diagnosis and a better therapy of atrial diseases.

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