

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT	DISEASE Lung adenocarcinoma	PHYSICIAN	MEDICAL FACILITY Arias Stella	SPECIMEN	SPECIMEN SITE Cervix
	DATE OF BIRTH 19 October 1947		ADDITIONAL RECIPIENT None		SPECIMEN ID 21-6827-2
	SEX Male		MEDICAL FACILITY ID 317319		SPECIMEN TYPE Block
	MEDICAL RECORD # Not given		PATHOLOGIST Not Provided		DATE OF COLLECTION 10 August 2021
					SPECIMEN RECEIVED 30 December 2021

Biomarker Findings

Microsatellite status - MS-Stable

Tumor Mutational Burden - 3 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

NF1 H1460fs*9

MTAP loss

PIK3R1 Y452_R461>I

BCL6 N73S

CDKN2A/B CDKN2A loss, CDKN2B loss

MAP2K4 loss exons 2-11

TP53 R196Q - subclonal[†]

8 Disease relevant genes with no reportable alterations: **ALK, BRAF, EGFR, ERBB2, KRAS, MET, RET, ROS1**

[†] See About the Test in appendix for details.

Report Highlights

- Targeted therapies with potential clinical benefit **approved in another tumor type: Selumetinib (p. 8), Trametinib (p. 8)**
- Evidence-matched clinical trial options based on this patient's genomic findings: (p. 9)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 3 Muts/Mb

GENOMIC FINDINGS

NF1 - H1460fs*9

10 Trials see p. 10

MTAP - loss

1 Trial see p. 9

PIK3R1 - Y452_R461>I

4 Trials see p. 12

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)
none	Selumetinib
	Trametinib
none	none
none	none

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

BCL6 - N73S	p. 5	MAP2K4 - loss exons 2-11	p. 6
CDKN2A/B - CDKN2A loss, CDKN2B loss	p. 6	TP53 - R196Q - subclonal	p. 7

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

ORDERED TEST # ORD-1275918-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI-H is generally infrequent in NSCLC, reported in fewer than 1% of samples across several large studies⁶⁻¹¹, whereas data on the reported incidence of MSI-H in SCLC has been limited and conflicting¹²⁻¹⁵. One study reported MSI-H in lung adenocarcinoma patients with smoking history, and 3 of 4 MSI-H patients examined also had metachronous carcinomas in other organs, although this has not been investigated in large scale studies⁶. Published data investigating the prognostic implications of MSI in NSCLC are limited (PubMed, Oct 2021).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁶. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁶⁻¹⁸. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹⁹⁻²¹. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{16,18,20-21}.

BIOMARKER

Tumor Mutational Burden

RESULT

3 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²²⁻²⁴, anti-PD-1 therapies²²⁻²⁵, and combination nivolumab and ipilimumab²⁶⁻³¹. Multiple clinical trials of PD-1- or PD-L1-targeting immune checkpoint inhibitors or combination of PD-1 and CTLA-4 inhibitors in NSCLC have reported that patients with tumors harboring TMB ≥ 10 Muts/Mb derive greater clinical benefit from these therapies than those with TMB < 10 Muts/Mb (based on this assay or others); similarly, higher efficacy of anti-PD-1 or anti-PD-L1 immunotherapy for treatment of patients with NSCLC, compared with the use of chemotherapy, has been observed more significantly in cases of TMB ≥ 10 Muts/Mb (based on this assay or others)^{22-23,26-28,32-39}. Improved OS of patients with

NSCLC treated with pembrolizumab plus chemotherapy relative to chemotherapy only⁴⁰, or those treated with nivolumab plus ipilimumab also relative to chemotherapy⁴¹, has been observed across all TMB levels.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that unspecified lung non-small cell lung carcinoma (NSCLC), lung adenocarcinoma, and lung squamous cell carcinoma (SCC) samples harbored median TMBs between 6.3 and 9 Muts/Mb, and 12% to 17% of cases had an elevated TMB of greater than 20 Muts/Mb⁴². Lower TMB is observed more commonly in NSCLCs harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are commonly observed in elevated TMB cases⁴³. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC⁴⁴⁻⁴⁵, several other large studies did find a strong association with increased TMB⁴⁶⁻⁴⁹. TMB > 10 muts/Mb was found to be more frequent in NSCLC metastases compared with primary tumors for both adenocarcinoma (38% vs. 25%) and SCC (41% vs. 35%) subtypes⁵⁰. A large study of Chinese patients with lung adenocarcinoma reported a shorter median OS for tumors with a higher number of mutations in a limited gene set compared with a

lower mutation number (48.4 vs. 61.0 months)⁴⁴. Another study of patients with NSCLC correlated elevated TMB with poorer prognosis and significantly associated lower TMB in combination with PD-L1 negative status with longer median survival in patients with lung adenocarcinoma⁵¹. However, no significant prognostic association of TMB and/or PD-L1 status with survival has been reported in patients with lung SCC⁵¹⁻⁵².

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁵³⁻⁵⁴ and cigarette smoke in lung cancer^{32,55}, treatment with temozolomide-based chemotherapy in glioma⁵⁶⁻⁵⁷, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁵⁸⁻⁶², and microsatellite instability (MSI)^{58,61-62}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{22-23,26-28,32-39,63}.

ORDERED TEST # ORD-1275918-01

GENOMIC FINDINGS

GENE

NF1

ALTERATION

H146Ofs*9

TRANSCRIPT ID

NM_001042492

CODING SEQUENCE EFFECT

4378delC

VARIANT ALLELE FREQUENCY (% VAF)

37.2%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in neurofibromatosis Type 1-associated neurofibroma⁶⁴⁻⁶⁷, glioma or glioblastoma⁶⁷⁻⁷¹, and non-small cell lung cancer⁷², NF1 inactivation may predict sensitivity to MEK inhibitors such as cobimetinib, trametinib, binimetinib, and selumetinib. Loss or inactivation of NF1 may also predict sensitivity to mTOR inhibitors, including

everolimus and temsirolimus, based on limited clinical data⁷³⁻⁷⁵ and strong preclinical data in models of malignant peripheral nerve sheath tumor (MPNST)⁷⁶⁻⁷⁷. A preclinical study suggests that combined mTOR and MEK inhibition is effective in a model of NF1-deficient MPNST⁷⁸. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors⁷⁹, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months⁸⁰.

FREQUENCY & PROGNOSIS

NF1 mutation has been observed in 6.9-11% of lung adenocarcinoma cases⁸¹ and 7.7-11% of lung squamous cell carcinoma cases⁸²⁻⁸⁴. Published data investigating the prognostic implications of NF1 alteration in lung cancer are limited (PubMed, Feb 2021). However, decreased NF1 expression was reported in 2 lung adenocarcinoma samples after

disease progression on first generation EGFR inhibitor and afatinib; neither sample harbored EGFR T790M mutation⁸⁵.

FINDING SUMMARY

NF1 encodes neurofibromin, a GTPase-activating protein (GAP) that is a key negative regulator of the RAS signaling pathway⁸⁶. Neurofibromin acts as a tumor suppressor by repressing RAS signaling⁸⁷. Alterations such as seen here may disrupt NF1 function or expression⁸⁷⁻⁹⁶.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in NF1 cause the autosomal dominant disorder neurofibromatosis type 1, which is characterized in part by increased risk of developing various tumors, including sarcoma, glioma, breast carcinoma, and neuroendocrine and hematological neoplasms⁹⁷⁻⁹⁹. Estimates for the prevalence of the disorder in the general population range from 1:2,500 to 1:3,000¹⁰⁰⁻¹⁰¹, and in the appropriate clinical context, germline testing of NF1 is recommended.

GENE

MTAP

ALTERATION

loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical and limited clinical evidence indicate that MTAP inactivation produces specific metabolic vulnerabilities. MTAP inactivation may confer sensitivity to MAT2A inhibitors¹⁰². A Phase 1 trial of MAT2A inhibitor AG-270 reported 1 PR and 2 SDs lasting longer than 6 months for patients with advanced solid tumors displaying MTAP loss¹⁰³. Although preclinical data have suggested that MTAP loss sensitizes cells to PRMT5 inhibition^{102,104-105}, MTAP loss may not be a biomarker of response to previously developed small-molecule SAM-uncompetitive PRMT5 inhibitors¹⁰⁶; dual PRMT1 and PRMT5 inhibition may be more effective¹⁰⁷⁻¹⁰⁹. In preclinical cancer models, MTAP inactivation showed increased

sensitivity to inhibitors of purine synthesis or purine analogs, especially upon addition of exogenous MTA, which is converted to adenine in normal cells, thereby providing competition to purine poisons lacking in MTAP-deficient cells¹¹⁰⁻¹²⁰. A Phase 2 study of L-alanosine, an inhibitor of adenine synthesis, as a monotherapy for 65 patients with MTAP-deficient cancers reported no responses and stable disease in 23.6% (13/55) of patients¹²¹.

FREQUENCY & PROGNOSIS

MTAP loss/homozygous deletion as well as loss of expression has been reported in a wide variety of solid tumors and hematologic cancers¹²²⁻¹²³; such events have been correlated with poor prognosis in a variety of cancer types, including hepatocellular carcinoma¹²⁴, gastrointestinal stromal tumors¹²⁵, mantle cell lymphoma (MCL)¹²⁶, melanoma¹²⁷⁻¹²⁸, gastric cancer¹²⁹, myxofibrosarcoma¹³⁰, nasopharyngeal carcinoma¹³¹, ovarian carcinoma¹²² and non-small cell lung cancer¹³². MTAP loss was not prognostic in pediatric B-cell acute lymphocytic leukemia¹³³ or in astrocytoma¹³⁴. However, MTAP has also

been reported to be overexpressed in colorectal cancer (CRC) samples¹³⁵, and MTAP retention is thought to be important for prostate cancer growth due to continuous supply of SAM¹³⁶. Germline SNPs in MTAP have been correlated with the development of cutaneous melanoma¹³⁷⁻¹³⁸, esophageal cancer¹³⁹⁻¹⁴⁰, osteosarcoma¹⁴¹, and CRC¹⁴².

FINDING SUMMARY

MTAP encodes S-methyl-5'-thioadenosine (MTA) phosphorylase, a tumor suppressor involved in polyamine metabolism and methionine synthesis, although its enzymatic function is dispensable for its tumor suppressor activity¹⁴³⁻¹⁴⁴. Decreased expression of MTAP leads to MTA accumulation within tumor cells and their microenvironment^{124,145-146}, thereby reducing intracellular arginine methylation^{102,104,147} and altering cell signaling^{146,148}. MTAP is located at 9p21, adjacent to CDKN2A and CDKN2B, with which it is frequently co-deleted in various cancers. Other alterations in MTAP are rare and have not been extensively characterized.

ORDERED TEST # ORD-1275918-01

GENOMIC FINDINGS

GENE

PIK3R1

ALTERATION

Y452_R461>I

TRANSCRIPT ID

NM_181523

CODING SEQUENCE EFFECT

1354_1382>AT

VARIANT ALLELE FREQUENCY (% VAF)

8.6%

CRs have been achieved by patients with endometrial cancer treated with the pan-PI3K inhibitor pilaralisib¹⁴⁹, and 1 PR has been achieved by a patient with breast cancer treated with the PI3K-alpha inhibitor alpelisib in combination with ribociclib and letrozole¹⁵³. Limited clinical and preclinical data suggest that PIK3R1 alterations may also be sensitive to inhibitors of mTOR^{152,154-157} or AKT¹⁵⁸⁻¹⁵⁹. One preclinical study reported that PIK3R1 truncation mutations in the 299-370 range confer sensitivity to MEK inhibitors¹⁶⁰.

(80%) was observed in endometrial carcinoma¹⁶³⁻¹⁶⁵, although PIK3R1 indels have been reported in other cancer types such as GBM, cervical squamous cell carcinoma, and urothelial bladder carcinoma¹⁶³. On the basis of limited clinical data, reduced PIK3R1 expression has been associated with reduced disease-free survival in prostate cancer¹⁶⁶ and metastasis-free survival in breast cancer¹⁶⁷. PIK3R1 expression is not associated with overall survival in neuroendocrine tumors¹⁶⁸.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical¹⁴⁹⁻¹⁵⁰ and preclinical¹⁵¹⁻¹⁵² data, PIK3R1 alteration may predict sensitivity to pan-PI3K or PI3K-alpha-selective inhibitors. In patients with PIK3R1 mutation and no other alterations in the PI3K-AKT-mTOR pathway, 2

FREQUENCY & PROGNOSIS

In the TCGA datasets, PIK3R1 mutation is most frequently observed in endometrial carcinoma (33%)⁵⁸, glioblastoma (GBM; 11%)¹⁶¹, uterine carcinosarcoma (11%)(cBioPortal, Jan 2022)⁸³⁻⁸⁴, and lower grade glioma (5%)¹⁶². PIK3R1 is often inactivated by in-frame insertions or deletions (indels), and the majority of this class of mutation

FINDING SUMMARY

PIK3R1 encodes the p85-alpha regulatory subunit of phosphatidylinositol 3-kinase (PI3K)¹⁶⁹. Loss of PIK3R1 has been shown to result in increased PI3K signaling¹⁷⁰⁻¹⁷³, promote tumorigenesis^{151,158,170}, and promote hyperplasia in the context of PTEN-deficiency¹⁷⁴. Alterations such as seen here may disrupt PIK3R1 function or expression^{152,159-160,164-165,175-183}.

GENE

BCL6

ALTERATION

N73S

TRANSCRIPT ID

NM_001706

CODING SEQUENCE EFFECT

218A>G

VARIANT ALLELE FREQUENCY (% VAF)

50.3%

alterations in BCL6.

FREQUENCY & PROGNOSIS

BCL6 mutations have been reported in <1% of lung adenocarcinomas and in 3% of lung squamous cell carcinomas⁸¹⁻⁸². BCL6 has been shown to be methylated in non-small cell lung cancer (NSCLC), and BCL6 mRNA levels were downregulated in analyzed NSCLC samples¹⁸⁴. The prognostic impact of BCL6 alterations in the context of non-small cell lung carcinoma has not been extensively investigated (PubMed, Apr 2021).

frequently rearranged or mutated in lymphomas¹⁸⁶, having been observed in follicular lymphoma (FL)¹⁸⁷, DLBCL¹⁸⁸⁻¹⁹⁰, Hodgkin's lymphoma¹⁹¹ and in MALT lymphoma¹⁹², where they are thought to promote transformation to large cell marginal zone B-cell lymphoma (MZBCL)¹⁹³. These events typically occur in the 5' regulatory region and lead to increased expression of BCL6¹⁹⁴⁻¹⁹⁵. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no therapies available to directly target

FINDING SUMMARY

BCL6 encodes B-cell lymphoma protein 6, a transcriptional repressor involved in the normal development of B-lymphocytes¹⁸⁵. This gene is

ORDERED TEST # ORD-1275918-01

GENOMIC FINDINGS

GENE

CDKN2A/B

ALTERATION

CDKN2A loss, CDKN2B loss

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical data suggest that tumors with loss of p16INK4a function may be sensitive to CDK4/6 inhibitors, such as abemaciclib, ribociclib, and palbociclib¹⁹⁶⁻¹⁹⁹. Although case studies have reported that patients with breast cancer or uterine leiomyosarcoma harboring CDKN2A loss responded to palbociclib treatment²⁰⁰⁻²⁰¹, multiple other clinical studies have shown no significant correlation between p16INK4a loss or inactivation and therapeutic benefit of these agents²⁰²⁻²⁰⁸; it is not known whether CDK4/6 inhibitors would be beneficial in this case. Although preclinical studies have suggested that loss of p14ARF function may be associated with reduced sensitivity to MDM2 inhibitors²⁰⁹⁻²¹⁰, the clinical relevance of p14ARF as a predictive biomarker is not clear. There are no drugs that directly target the mutation or loss of CDKN2B in cancer. Because the p15INK4b protein encoded by CDKN2B is known to inhibit CDK4, tumors with CDKN2B mutation or loss may predict sensitivity to CDK4/6 inhibitors, such as ribociclib, abemaciclib, and

palbociclib^{203,205-206,211-213}.

FREQUENCY & PROGNOSIS

CDKN2A/B loss and CDKN2A mutation have been reported in approximately 19% and 4% of lung adenocarcinomas, respectively⁸¹. CDKN2A/B loss and CDKN2A mutation have been reported in 26% and 17% of lung squamous cell carcinoma (SCC) samples analyzed in the TCGA dataset, respectively⁸². Loss of p16INK4a protein expression, through CDKN2A mutation, homozygous deletion, or promoter methylation, has been described in 49-68% of non-small cell lung cancer (NSCLC) samples, whereas low p14ARF protein expression has been detected in 21-72% of NSCLC samples^{82,214-219}. In patients with lung SCC, loss of CDKN2B associated with poor survival in one study²²⁰. Loss of p16INK4a protein as well as CDKN2A promoter hypermethylation correlate with poor survival in patients with NSCLC^{216,221-223}.

FINDING SUMMARY

CDKN2A encodes two different, unrelated tumor suppressor proteins, p16INK4a and p14ARF, whereas CDKN2B encodes the tumor suppressor p15INK4b²²⁴⁻²²⁵. Both p15INK4b and p16INK4a bind to and inhibit CDK4 and CDK6, thereby maintaining the growth-suppressive activity of the Rb tumor suppressor; loss or inactivation of either p15INK4b or p16INK4a contributes to

dysregulation of the CDK4/6-cyclin-Rb pathway and loss of cell cycle control^{215,226}. The tumor suppressive functions of p14ARF involve stabilization and activation of p53, via a mechanism of MDM2 inhibition²²⁷⁻²²⁸. One or more alterations observed here are predicted to result in p16INK4a loss of function²²⁹⁻²⁵⁰. One or more alterations seen here are predicted to result in p14ARF loss of function^{233,250-253}. CDKN2B alterations such as seen here are predicted to inactivate p15INK4b²⁵⁴.

POTENTIAL GERMLINE IMPLICATIONS

Germline CDKN2A mutation is associated with melanoma-pancreatic cancer syndrome, a condition marked by increased risk of developing malignant melanoma and/or pancreatic cancer²⁵⁵. Mutation carriers within families may develop either or both types of cancer, and melanoma cases may be referred to as familial or hereditary melanoma²⁵⁶⁻²⁵⁷. CDKN2A is the most implicated gene in familial melanoma, with germline mutations present in 16% to 20% of familial melanoma cases²⁵⁸⁻²⁶⁰. CDKN2A alteration has also been implicated in familial melanoma-astrocytoma syndrome, an extremely rare tumor association characterized by dual predisposition to melanoma and nervous system tumors²⁶¹⁻²⁶³. In the appropriate clinical context, germline testing of CDKN2A is recommended.

GENE

MAP2K4

ALTERATION

loss exons 2-11

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no targeted therapies to address MAP2K4 loss or inactivation.

FREQUENCY & PROGNOSIS

MAP2K4 mutation has been reported in 1% of lung adenocarcinoma samples and <1% of lung squamous cell carcinoma samples analyzed in the TCGA datasets⁸¹⁻⁸². In these same datasets, deletion of MAP2K4 was reported in 2% of lung adenocarcinoma samples, but not in any lung squamous cell carcinoma samples⁸¹⁻⁸². Whether MAP2K4 acts as an oncogene²⁶⁴⁻²⁶⁵ or tumor suppressor²⁶⁶⁻²⁶⁸ in the context of lung cancer is unclear. Published data investigating the prognostic implications of MAP2K4 in lung cancer are limited (PubMed, Feb 2021).

FINDING SUMMARY

MAP2K4 encodes the protein kinase MKK4, a member of a MAPK signaling cascade that leads to apoptosis in response to cellular stress²⁶⁶. MAP2K4 has been proposed to act as a tumor suppressor in several cancer types, including pancreatic cancer, but has also been suggested to act as an oncogene in certain situations²⁶⁴⁻²⁷⁰. Alterations such as seen here may disrupt MAP2K4 function or expression^{267,269,271-272}.

ORDERED TEST # ORD-1275918-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

R196Q - subclonal

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

587G>A

VARIANT ALLELE FREQUENCY (% VAF)

0.81%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib²⁷³⁻²⁷⁶, or p53 gene therapy and immunotherapeutics such as SGT-53²⁷⁷⁻²⁸¹ and ALT-801²⁸². In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% (17/176) and SDs in 53.4% (94/176) of patients with solid tumors; the response rate was 21.1% (4/19) for patients with TP53 mutations versus 12.1% (4/33) for patients who were TP53 wild-type²⁸³. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 31.9% (30/94, 3 CR) ORR and a 73.4% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer²⁸⁴. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 42.9% (9/21, 1 CR) ORR and a 76.2% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer²⁸⁵. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone²⁸⁶. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24.0% (6/25) ORR with adavosertib combined with paclitaxel²⁸⁷. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and

docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71.4% (5/7) response rate for patients with TP53 alterations²⁸⁸. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75.0% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage²⁸¹. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53-mutated, but not TP53-wild-type, breast cancer xenotransplant mouse model²⁸⁹. Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246²⁹⁰⁻²⁹². In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR²⁹³. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies²⁹⁴⁻²⁹⁵; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies²⁹⁶⁻²⁹⁷. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer; mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)^{81-82,218,298-302}, including 42-52% of lung adenocarcinomas and 58-83% of lung squamous cell carcinomas (cBioPortal, COSMIC, Feb 2021)^{48-49,81-82}. TP53 homozygous deletion has been observed in 1.4% of lung adenocarcinoma and <1% of lung squamous cell carcinoma cases (cBioPortal, Feb 2021)⁸³⁻⁸⁴. In one study of 55 patients with lung adenocarcinoma, TP53 alterations correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to

PD-1 inhibitors pembrolizumab and nivolumab in this study³⁰³. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma³⁰⁴.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers³⁰⁵. Alterations such as seen here may disrupt TP53 function or expression³⁰⁶⁻³¹⁰.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers³¹¹⁻³¹³, including sarcomas³¹⁴⁻³¹⁵. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000³¹⁶ to 1:20,000³¹⁵. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30³¹⁷. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion³¹⁸⁻³²³. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy³¹⁸⁻³¹⁹. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease³²⁴. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{322,325-326}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # ORD-1275918-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Selumetinib

Assay findings association
NF1
H1460fs*9

AREAS OF THERAPEUTIC USE

Selumetinib is a MEK inhibitor that is FDA approved to treat pediatric patients 2 years of age and older with neurofibromatosis type 1 (NF1) who have symptomatic, inoperable plexiform neurofibromas (PNs). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence in neurofibromatosis type 1 (NF1)-associated neurofibroma^{64-67,327-331}, glioma^{67-71,332}, and non-small cell lung cancer⁷², NF1 inactivation may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

In the Phase 2 umbrella trial National Lung Matrix Trial, selumetinib plus docetaxel yielded an ORR of 29% (4/14) for patients with lung adenocarcinoma harboring NF1 loss⁷². In a Phase 2 study of selumetinib monotherapy to treat patients with lung cancer who were selected for mutation in KRAS, HRAS, NRAS, or BRAF, a mPFS of 2.3 months and mOS of 6.5 months was observed³³³. In a Phase 2 study of patients with NSCLC who had failed on at least 2 prior chemotherapeutic regimens, selumetinib as a monotherapy did not improve survival as compared to

pemetrexed (67 vs 90 days, HR= 1.08); however, 2 PRs were reported³³⁴. A Phase 2 study of selumetinib combined with docetaxel for patients with advanced or metastatic KRAS wild-type NSCLC who were previously treated did not report improved survival benefit compared to docetaxel alone³³⁵. A Phase 2 study of selumetinib combined with pemetrexed and platinum based chemotherapy for treatment of patients with advanced non-squamous NSCLC showed improved ORR (35% with intermittent dosing and 62% for continuous dosing) compared to chemotherapy alone (24%) but did not report a statistically significant improvement in mPFS³³⁶. The combination of selumetinib with platinum doublet chemotherapy has been studied in a Phase 1 trial for patients with advanced NSCLC in the first line setting and has reported 4/21 PRs in the selumetinib + pemetrexed/carboplatin cohort and 2/15 PRs in the pemetrexed/cisplatin cohort; selumetinib in combination with gemcitabine regimens was not tolerated³³⁷. A Phase 1b study of selumetinib in combination with osimertinib for patients with EGFR-mutated lung cancer who had progressed on previous TKI treatment reported an ORR of 41.7% (15/36)³³⁸.

Trametinib

Assay findings association
NF1
H1460fs*9

AREAS OF THERAPEUTIC USE

Trametinib is a MEK inhibitor that is FDA approved as a monotherapy to treat patients with melanoma with BRAF V600E or V600K mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence in neurofibromatosis type 1 (NF1)-associated neurofibroma^{64-67,327-331}, glioma^{67-71,332}, and non-small cell lung cancer⁷², NF1 inactivation may predict sensitivity to MEK inhibitors.

SUPPORTING DATA

Phase 1 and 2 monotherapy trials of MEK inhibitors such as trametinib and RO4987655 have shown low response rates in patients with non-small cell lung cancer (NSCLC), irrespective of KRAS mutation status, and no improvement in PFS compared to docetaxel³³⁹⁻³⁴¹. However, Phase 1 and 2 trials of MEK inhibitors in combination with docetaxel or pemetrexed in NSCLC have shown improved clinical activity and patient survival compared to chemotherapeutics alone, although no association was observed between response and KRAS

mutation status³⁴²⁻³⁴⁴. In contrast, although 3 objective responses were observed in patients with NSCLC treated with the MEK inhibitor selumetinib in combination with erlotinib in a Phase 2 trial, there was no significant increase in either PFS or OS relative to patients treated with selumetinib alone; further, the combination increased toxicity relative to monotherapy³⁴⁵. Preclinical and early clinical studies have shown synergistic antitumorigenic effects when the combination of MEK and PI3K inhibitors was used to treat KRAS-driven NSCLC³⁴⁶⁻³⁴⁸. A Phase 1b combination trial of trametinib and the pan-PI3K inhibitor BKM120 reported a DCR of 59% in patients with NSCLC, including 1 confirmed PR in 17 patients; although the reported adverse effects were prevalent and often severe, the study recommended a Phase 2 dose³⁴⁹. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors⁷⁹, a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months⁸⁰.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 13 January 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # **ORD-1275918-01**
CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
MTAP
RATIONALE
MTAP loss may predict sensitivity to MAT2A inhibitors.

ALTERATION
loss

NCT03435250
PHASE 1

Study of AG-270 in Participants With Advanced Solid Tumors or Lymphoma With MTAP Loss

TARGETS
MAT2A
LOCATIONS: Tennessee, New York, Connecticut, Massachusetts, Barcelona (Spain), Villejuif Cedex (France)

ORDERED TEST # ORD-1275918-01

CLINICAL TRIALS

GENE

NF1

ALTERATION

H1460fs*9

RATIONALE

On the basis of clinical evidence and strong preclinical evidence, NF1 inactivation may predict sensitivity to MEK inhibitors. Limited clinical

data and strong preclinical data indicate that loss or inactivation of NF1 may also predict sensitivity to mTOR inhibitors.

NCT03600701
PHASE 2

Atezolizumab and Cobimetinib in Treating Patients With Metastatic, Recurrent, or Refractory Non-small Cell Lung Cancer

TARGETS
PD-L1, MEK

LOCATIONS: Florida, Alabama, North Carolina, Virginia, District of Columbia, Oklahoma, Ohio, Michigan, New Hampshire

NCT03989115
PHASE 1/2

Dose-Escalation and Dose-Expansion of RMC-4630 and Cobimetinib in Relapsed/Refractory Solid Tumors

TARGETS
SHP2, MEK

LOCATIONS: Florida, Georgia, Texas, North Carolina, Tennessee, Virginia, Oklahoma, Maryland, Pennsylvania, Ohio

NCT01737502
PHASE 1/2

Sirolimus and Auranofin in Treating Patients With Advanced or Recurrent Non-Small Cell Lung Cancer or Small Cell Lung Cancer

TARGETS
mTOR

LOCATIONS: Florida

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03905148
PHASE 1/2

Study of the Safety and Pharmacokinetics of BGB-283 and PD-0325901 in Patients With Advanced or Refractory Solid Tumors

TARGETS
RAFs, EGFR, MEK

LOCATIONS: Texas, Randwick (Australia), Blacktown (Australia), Melbourne (Australia), Nedlands (Australia)

NCT03334617
PHASE 2

Phase II Umbrella Study of Novel Anti-cancer Agents in Patients With NSCLC Who Progressed on an Anti-PD-1/PD-L1 Containing Therapy.

TARGETS
PD-L1, PARP, mTORC1, mTORC2, ATR, CD73, STAT3

LOCATIONS: Texas, Tennessee, Virginia, District of Columbia, Maryland, Missouri, Pennsylvania, New York, Massachusetts

ORDERED TEST # ORD-1275918-01

CLINICAL TRIALS
NCT03337698
PHASE 1/2

A Study Of Multiple Immunotherapy-Based Treatment Combinations In Participants With Metastatic Non-Small Cell Lung Cancer (Morpheus- Non-Small Cell Lung Cancer)

TARGETS
PD-L1, MEK, CEA, CXCR4, EZH2,
MDM2, ADORA2A

LOCATIONS: Tennessee, Ohio, Nevada, Malaga (Spain), Madrid (Spain), Valencia (Spain), Pamplona (Spain), Saint Herblain (France), Barcelona (Spain), Toulouse (France)

NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS
VEGFRs, ABL, SRC, ALK, ROS1, AXL,
TRKA, MET, TRKC, DDR2, KIT, EGFR,
PD-1, CTLA-4, PARP, CDK4, CDK6,
FLT3, CSF1R, RET, mTOR, ERBB2,
ERBB3, MEK, BRAF, SMO

LOCATIONS: London (Canada), Toronto (Canada), Kingston (Canada), Ottawa (Canada), Montreal (Canada), Regina (Canada), Saskatoon (Canada), Edmonton (Canada), Vancouver (Canada)

NCT02664935
PHASE 2

National Lung Matrix Trial: Multi-drug Phase II Trial in Non-Small Cell Lung Cancer

TARGETS
FGFRs, mTORC1, mTORC2, CDK4,
CDK6, ALK, ROS1, AXL, TRKA, MET,
TRKC, MEK, AKTs, EGFR, PD-L1, KIT,
DDR2, VEGFRs, PDGFRA, FLT3, RET,
TRKB

LOCATIONS: Exeter (United Kingdom), Belfast (United Kingdom), Cardiff (United Kingdom), Bristol (United Kingdom), Wirral (United Kingdom), Southampton (United Kingdom), Glasgow (United Kingdom), Birmingham (United Kingdom), Manchester (United Kingdom), Oxford (United Kingdom)

NCT04185831
PHASE 2

A MolEcularly Guided Anti-Cancer Drug Off-Label Trial

TARGETS
PD-L1, MEK, mTOR

LOCATIONS: Gothenburg (Sweden), Uppsala (Sweden)

ORDERED TEST # ORD-1275918-01

CLINICAL TRIALS

GENE
PIK3R1

RATIONALE
On the basis of clinical and strong preclinical data, sensitivity to pan-PI3K or PI3K-alpha-selective inhibitors.
PIK3R1 loss or inactivation may indicate

ALTERATION
Y452_R461>I

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03711058
PHASE 1/2

Study of PI3Kinase Inhibition (Copanlisib) and Anti-PD-1 Antibody Nivolumab in Relapsed/Refractory Solid Tumors With Expansions in Mismatch-repair Proficient (MSS) Colorectal Cancer

TARGETS
PD-1, PI3K

LOCATIONS: Maryland

NCT03502733
PHASE 1

Copanlisib and Nivolumab in Treating Patients With Metastatic Solid Tumors or Lymphoma

TARGETS
PI3K, PD-1

LOCATIONS: Texas, Maryland

NCT04895579
PHASE 1

Lung Cancer With Copanlisib and Durvalumab

TARGETS
PD-L1, PI3K

LOCATIONS: Kentucky

ORDERED TEST # ORD-1275918-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

CDH1
G661D

CIC
K468R

HGF
loss

KDM5C
R828Q

PARP3
I452F

RAD51D
G74R

TP53
P34R

ORDERED TEST # ORD-1275918-01

APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXO2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDMS5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TPRSS2

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

ORDERED TEST # ORD-1275918-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.


ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials
Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 13 January 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1275918-01

APPENDIX

About FoundationOne®CDx

- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
- TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 - The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
 - Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 - Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

- Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	%CV*
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	%CV*
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

ORDERED TEST # ORD-1275918-01

APPENDIX
About FoundationOne®CDx

cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 5.2.0

The median exon coverage for this sample is 1,017x

ORDERED TEST # ORD-1275918-01

APPENDIX

References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Warth A, et al. Virchows Arch. (2016) PMID: 26637197
7. Ninomiya H, et al. Br. J. Cancer (2006) PMID: 16641899
8. Vanderwalde A, et al. Cancer Med (2018) PMID: 29436178
9. Zang YS, et al. Cancer Med (2019) PMID: 31270941
10. Dudley JC, et al. Clin. Cancer Res. (2016) PMID: 26880610
11. Takamochi K, et al. Lung Cancer (2017) PMID: 28676214
12. Pyllkänen L, et al. Environ. Mol. Mutagen. (1997) PMID: 9329646
13. Gonzalez R, et al. Ann. Oncol. (2000) PMID: 11061602
14. Chen XQ, et al. Nat. Med. (1996) PMID: 8782463
15. Merlo A, et al. Cancer Res. (1994) PMID: 8174113
16. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
17. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
18. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
19. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
20. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
21. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
22. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
23. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
24. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
25. Cristescu R, et al. Science (2018) PMID: 30309915
26. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
27. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
28. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
29. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
30. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
31. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
32. Rizvi NA, et al. Science (2015) PMID: 25765070
33. Colli LM, et al. Cancer Res. (2016) PMID: 27197178
34. Wang VE, et al. J Immunother Cancer (2017) PMID: 28923100
35. Carbone DP, et al. N. Engl. J. Med. (2017) PMID: 28636851
36. Rizvi H, et al. J. Clin. Oncol. (2018) PMID: 29337640
37. Forde PM, et al. N. Engl. J. Med. (2018) PMID: 29658848
38. Miao D, et al. Nat. Genet. (2018) PMID: 30150660
39. Chae YK, et al. Clin Lung Cancer (2019) PMID: 30425022
40. Paz-Ares et al., 2019; ESMO Abstract LBA80
41. Hellmann MD, et al. N. Engl. J. Med. (2019) PMID: 31562796
42. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
43. Spigel et al., 2016; ASCO Abstract 9017
44. Xiao D, et al. Oncotarget (2016) PMID: 27009843
45. Shim HS, et al. J Thorac Oncol (2015) PMID: 26200269
46. Govindan R, et al. Cell (2012) PMID: 22980976
47. Ding L, et al. Nature (2008) PMID: 18948947
48. Imielinski M, et al. Cell (2012) PMID: 22980975
49. Kim Y, et al. J. Clin. Oncol. (2014) PMID: 24323028
50. Stein et al., 2019; DOI: 10.1200/PO.18.00376
51. Chen Y, et al. J. Exp. Clin. Cancer Res. (2019) PMID: 31088500
52. Yu H, et al. J Thorac Oncol (2019) PMID: 30253973
53. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
54. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
55. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
56. Johnson BE, et al. Science (2014) PMID: 24336570
57. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
58. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
59. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
60. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
61. Nature (2012) PMID: 22810696
62. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
63. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
64. Dombi E, et al. N. Engl. J. Med. (2016) PMID: 28029918
65. Schalkwijk S, et al. Cancer Chemother Pharmacol (2021) PMID: 33903938
66. Toledano H, et al. Childs Nerv Syst (2021) PMID: 33751171
67. Ronsley R, et al. Cancer Med (2021) PMID: 33939292
68. Fangusaro J, et al. Lancet Oncol. (2019) PMID: 31151904
69. Manoharan N, et al. J Neurooncol (2020) PMID: 32780261
70. Kondyli M, et al. J Neurooncol (2018) PMID: 30097824
71. Awada G, et al. Case Rep Oncol () PMID: 33082744
72. Middleton G, et al. Nature (2020) PMID: 32669708
73. Lim SM, et al. Oncotarget (2016) PMID: 26859683
74. Weiss B, et al. Neuro-oncology (2015) PMID: 25314964
75. Janku F, et al. Oncotarget (2014) PMID: 24931142
76. Johannessen CM, et al. Curr. Biol. (2008) PMID: 18164202
77. Johannessen CM, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15937108
78. Malone CF, et al. Cancer Discov (2014) PMID: 24913553
79. Tolcher AW, et al. Ann. Oncol. (2015) PMID: 25344362
80. Patterson et al., 2018; AACR Abstract 3891
81. Nature (2014) PMID: 25079552
82. Nature (2012) PMID: 22960745
83. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
84. Gao J, et al. Sci Signal (2013) PMID: 23550210
85. de Bruin EC, et al. Cancer Discov (2014) PMID: 24535670
86. Hattori S, et al. Biochem. Biophys. Res. Commun. (1991) PMID: 1904223
87. Morcos P, et al. Mol. Cell. Biol. (1996) PMID: 8628317
88. Ballester R, et al. Cell (1990) PMID: 2121371
89. Xu GF, et al. Cell (1990) PMID: 2116237
90. Martin GA, et al. Cell (1990) PMID: 2121370
91. Thomas L, et al. Hum. Mutat. (2012) PMID: 22807134
92. Skuse GR, et al. Hum. Mol. Genet. (1997) PMID: 9300663
93. Messiaen LM, et al. Genet. Med. () PMID: 11258625
94. Ars E, et al. Hum. Mol. Genet. (2000) PMID: 10607834
95. Messiaen LM, et al. J. Med. Genet. (2005) PMID: 15863657
96. Poullet P, et al. Mol. Cell. Biol. (1994) PMID: 8264648
97. Jett K, et al. Genet. Med. (2010) PMID: 20027112
98. Patil S, et al. Oncologist (2012) PMID: 22240541
99. Evans DG, et al. Clin Sarcoma Res (2012) PMID: 23036231
100. Upadhyaya M, et al. J. Med. Genet. (1995) PMID: 8544190
101. Williams VC, et al. Pediatrics (2009) PMID: 19117870
102. Marjon K, et al. Cell Rep (2016) PMID: 27068473
103. Heist et al., 2019; AACR-NCI-EORTC Abstract B116
104. Mavrikis KJ, et al. Science (2016) PMID: 26912361
105. Endoscopy (1989) PMID: 2691236
106. Guccione E, et al. Nat. Rev. Mol. Cell Biol. (2019) PMID: 31350521
107. Fedoriv A, et al. Cancer Cell (2019) PMID: 31257072
108. Srour N, et al. Cancer Cell (2019) PMID: 31287990
109. Gao G, et al. Nucleic Acids Res. (2019) PMID: 30916320
110. Hansen LJ, et al. Cancer Res. (2019) PMID: 31040154
111. Tang B, et al. Cancer Res. (2018) PMID: 29844120
112. Munshi PN, et al. Oncologist (2014) PMID: 24928612
113. de Oliveira SF, et al. PLoS ONE (2016) PMID: 26751376
114. Lubin M, et al. PLoS ONE (2009) PMID: 19478948
115. Tang B, et al. Cancer Biol. Ther. (2012) PMID: 22825330
116. Collins CC, et al. Mol. Cancer Ther. (2012) PMID: 22252602
117. Bertino JR, et al. Cancer Biol. Ther. (2011) PMID: 21301207
118. Coulthard SA, et al. Mol. Cancer Ther. (2011) PMID: 21282358
119. Miyazaki S, et al. Int. J. Oncol. (2007) PMID: 17912432
120. Efferth T, et al. Blood Cells Mol. Dis. () PMID: 11987241
121. Kindler HL, et al. Invest New Drugs (2009) PMID: 18618081
122. Wei R, et al. Sci Rep (2016) PMID: 27929028
123. Zhao M, et al. BMC Genomics (2016) PMID: 27556634
124. Kirovski G, et al. Am. J. Pathol. (2011) PMID: 21356366
125. Huang HY, et al. Clin. Cancer Res. (2009) PMID: 19887491
126. Marcé S, et al. Clin. Cancer Res. (2006) PMID: 16778103
127. Meyer S, et al. Exp. Dermatol. (2010) PMID: 20500769
128. Wild PJ, et al. Arch Dermatol (2006) PMID: 16618867
129. Kim J, et al. Genes Chromosomes Cancer (2011) PMID: 21412930
130. Li CF, et al. Oncotarget (2014) PMID: 25426549
131. He HL, et al. Medicine (Baltimore) (2015) PMID: 26656376
132. Su CY, et al. Eur J Surg Oncol (2014) PMID: 24969958
133. Mirebeau D, et al. Haematologica (2006) PMID: 16818274
134. Becker AP, et al. Pathobiology (2015) PMID: 26088413
135. Snezhkina AV, et al. Oxid Med Cell Longev (2016) PMID: 27433286
136. Bistulfi G, et al. Oncotarget (2016) PMID: 26910893
137. Antonopoulou K, et al. J. Invest. Dermatol. (2015) PMID: 25407435
138. Maccioni L, et al. BMC Cancer (2013) PMID: 23816148
139. Hyland PL, et al. Int J Epidemiol (2016) PMID: 26635288
140. Lin X, et al. Cancer Sci. (2017) PMID: 27960044
141. Zhi L, et al. J Cancer (2016) PMID: 27994653
142. Gu F, et al. Br. J. Cancer (2013) PMID: 23361049
143. Limm K, et al. PLoS ONE (2016) PMID: 27479139
144. Tang B, et al. G3 (Bethesda) (2014) PMID: 25387827
145. Limm K, et al. Eur. J. Cancer (2013) PMID: 23265702
146. Stevens AP, et al. J. Cell. Biochem. (2009) PMID: 19097084
147. Kryukov GV, et al. Science (2016) PMID: 26912360
148. Limm K, et al. Eur. J. Cancer (2014) PMID: 25087184
149. Matulonis U, et al. Gynecol. Oncol. (2015) PMID: 25528496
150. Pitz MW, et al. Neuro-oncology (2015) PMID: 25605819
151. Thorpe LM, et al. Proc. Natl. Acad. Sci. U.S.A. (2017) PMID: 28630349
152. Sun M, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20027531

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 13 January 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1275918-01

APPENDIX

References

- 20713702
153. Juric et al., 2016; SABCS Abstract P3-14-01
154. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
155. Myers AP, et al. Gynecol. Oncol. (2016) PMID: 27016228
156. Day TA, et al. Clin. Cancer Res. (2019) PMID: 30420444
157. Ou O, et al. Cancer Lett. (2014) PMID: 25193464
158. Li X, et al. Nat Commun (2019) PMID: 30755611
159. Quayle SN, et al. PLoS ONE (2012) PMID: 23166678
160. Cheung LW, et al. Cancer Cell (2014) PMID: 25284480
161. Brennan CW, et al. Cell (2012) PMID: 24120142
162. Cancer Genome Atlas Research Network, et al. N. Engl. J. Med. (2015) PMID: 26061751
163. Ye K, et al. Nat. Med. (2016) PMID: 26657142
164. Cheung LW, et al. Cancer Discov (2011) PMID: 21984976
165. Urick ME, et al. Cancer Res. (2011) PMID: 21478295
166. Munkley J, et al. Oncoscience (2015) PMID: 26501081
167. Cizkova M, et al. BMC Cancer (2013) PMID: 24229379
168. Qian ZR, et al. J. Clin. Oncol. (2013) PMID: 23980085
169. Huang CH, et al. Cell Cycle (2008) PMID: 18418043
170. Taniguchi CM, et al. Cancer Res. (2010) PMID: 20530665
171. Luo J, et al. Cell Metab. (2006) PMID: 16679293
172. Ueki K, et al. J. Biol. Chem. (2003) PMID: 14504291
173. Mauvais-Jarvis F, et al. J. Clin. Invest. (2002) PMID: 11781359
174. Luo J, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 16006513
175. Jaiswal BS, et al. Cancer Cell (2009) PMID: 19962665
176. Ko HR, et al. Cell Death Dis (2014) PMID: 24651434
177. Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19915146
178. Huang CH, et al. Science (2007) PMID: 18079394
179. Bousquet C, et al. EMBO J. (2006) PMID: 16917505
180. Oliver MD, et al. Biosci. Rep. (2017) PMID: 28143957
181. Philp AJ, et al. Cancer Res. (2001) PMID: 11606375
182. Lucas CL, et al. J. Exp. Med. (2014) PMID: 25488983
183. Chen L, et al. Nat Commun (2018) PMID: 29636477
184. Dmitriev AA, et al. Epigenetics (2012) PMID: 22491060
185. Eur. J. Immunol. (2011) PMID: 21792874
186. Wagner SD, et al. Br. J. Haematol. (2011) PMID: 21083654
187. Otsuki T, et al. Blood (1995) PMID: 7742550
188. Tibiletti MG, et al. Hum. Pathol. (2009) PMID: 19144384
189. Tzankov A, et al. Hum. Pathol. (2009) PMID: 19524106
190. Offit K, et al. N. Engl. J. Med. (1994) PMID: 8208268
191. Seitz V, et al. Blood (2001) PMID: 11290603
192. Ye H, et al. Haematologica (2008) PMID: 18166802
193. Flossbach L, et al. Int. J. Cancer (2011) PMID: 20830719
194. Ye BH, et al. EMBO J. (1995) PMID: 8557040
195. J Clin Exp Hematop (2006) PMID: 17142954
196. Konecny GE, et al. Clin. Cancer Res. (2011) PMID: 21278246
197. Katsumi Y, et al. Biochem. Biophys. Res. Commun. (2011) PMID: 21871868
198. Cen L, et al. Neuro-oncology (2012) PMID: 22711607
199. Logan JE, et al. Anticancer Res. (2013) PMID: 23898052
200. Elvin JA, et al. Oncologist (2017) PMID: 28283584
201. Gao J, et al. Curr Oncol (2015) PMID: 26715889
202. Gopalan et al., 2014; ASCO Abstract 8077
203. Peguero et al., 2016; ASCO Abstract 2528
204. Konecny et al., 2016; ASCO Abstract 5557
205. DeMichele A, et al. Clin. Cancer Res. (2015) PMID: 25501126
206. Finn RS, et al. Lancet Oncol. (2015) PMID: 25524798
207. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
208. Johnson DB, et al. Oncologist (2014) PMID: 24797823
209. Van Maerken T, et al. Mol. Cancer Ther. (2011) PMID: 21460101
210. Gamble LD, et al. Oncogene (2012) PMID: 21725357
211. Shapiro et al., 2013; ASCO Abstract 2500
212. Flaherty KT, et al. Clin. Cancer Res. (2012) PMID: 22090362
213. Dickson MA, et al. J. Clin. Oncol. (2013) PMID: 23569312
214. Dostader EE, et al. Hum. Pathol. (2012) PMID: 21840041
215. Gazzeri S, et al. Oncogene (1998) PMID: 9484839
216. Kratzke RA, et al. Cancer Res. (1996) PMID: 8758904
217. Lee JU, et al. Tuberc Respir Dis (Seoul) (2012) PMID: 23101020
218. Cortot AB, et al. Clin Lung Cancer (2014) PMID: 24169260
219. Mounawar M, et al. Cancer Res. (2007) PMID: 17575133
220. Zhao Y, et al. Clin Lung Cancer (2011) PMID: 21889114
221. Kawabuchi B, et al. Int. J. Cancer (1999) PMID: 9988232
222. Xing XB, et al. PLoS ONE (2013) PMID: 23805242
223. Lou-Qian Z, et al. PLoS ONE (2013) PMID: 23372805
224. Quelle DE, et al. Cell (1995) PMID: 8521522
225. Mutat. Res. (2005) PMID: 15878778
226. Oncogene (1999) PMID: 10498883
227. Sherr CJ, et al. Cold Spring Harb. Symp. Quant. Biol. (2005) PMID: 16869746
228. Ozenne P, et al. Int. J. Cancer (2010) PMID: 20549699
229. Ruas M, et al. Oncogene (1999) PMID: 10498896
230. Jones R, et al. Cancer Res. (2007) PMID: 17909018
231. Haferkamp S, et al. Aging Cell (2008) PMID: 18843795
232. Mutat. Res. (2002) PMID: 12417717
233. Rizo H, et al. J. Biol. Chem. (2001) PMID: 11518711
234. Gombart AF, et al. Leukemia (1997) PMID: 9324288
235. Yang R, et al. Cancer Res. (1995) PMID: 7780957
236. Parry D, et al. Mol. Cell. Biol. (1996) PMID: 8668202
237. Greenblatt MS, et al. Oncogene (2003) PMID: 12606942
238. Yarbrough WG, et al. J. Natl. Cancer Inst. (1999) PMID: 10491434
239. Poi MJ, et al. Mol. Carcinog. (2001) PMID: 11255261
240. Byeon IJ, et al. Mol. Cell (1998) PMID: 9660926
241. Kannengiesser C, et al. Hum. Mutat. (2009) PMID: 19260062
242. Lal G, et al. Genes Chromosomes Cancer (2000) PMID: 10719365
243. Koh J, et al. Nature (1995) PMID: 7777061
244. McKenzie HA, et al. Hum. Mutat. (2010) PMID: 20340136
245. Miller PJ, et al. Hum. Mutat. (2011) PMID: 21462282
246. Kutscher CL, et al. Physiol. Behav. (1977) PMID: 905385
247. Scaini MC, et al. Hum. Mutat. (2014) PMID: 24659262
248. Jenkins NC, et al. J. Invest. Dermatol. (2013) PMID: 23190892
249. Walker GJ, et al. Int. J. Cancer (1999) PMID: 10389768
250. Rutter JL, et al. Oncogene (2003) PMID: 12853981
251. Itahana K, et al. Cancer Cell (2008) PMID: 18538737
252. Zhang Y, et al. Mol. Cell (1999) PMID: 10360174
253. Zhang Y, et al. Cell (1998) PMID: 9529249
254. Jafri M, et al. Cancer Discov (2015) PMID: 25873077
255. Whelan AJ, et al. N Engl J Med (1995) PMID: 7666917
256. Adv Exp Med Biol (2010) PMID: 20687502
257. Hogg D, et al. J Cutan Med Surg (1998) PMID: 9479083
258. De Unamuno B, et al. Melanoma Res (2018) PMID: 29543703
259. Soura E, et al. J Am Acad Dermatol (2016) PMID: 26892650
260. Huerta C, et al. Acta Derm Venereol (2018) PMID: 29405243
261. Kaufman DK, et al. Neurology (1993) PMID: 8414022
262. Bahau M, et al. Cancer Res (1998) PMID: 9622062
263. Chan AK, et al. Clin Neuropathol () PMID: 28699883
264. Lee HY, et al. Clin. Cancer Res. (2005) PMID: 16115952
265. Lee HY, et al. J. Biol. Chem. (2003) PMID: 12714585
266. Curtis C, et al. Nature (2012) PMID: 22522925
267. Ahn YH, et al. Mol. Cell. Biol. (2011) PMID: 21896780
268. Davis SJ, et al. BMC Cancer (2011) PMID: 21575258
269. Teng DH, et al. Cancer Res. (1997) PMID: 9331070
270. Cunningham SC, et al. Cancer Res. (2006) PMID: 16740690
271. Kan Z, et al. Nature (2010) PMID: 20668451
272. Takekawa M, et al. Mol. Cell (2005) PMID: 15866172
273. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
274. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
275. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
276. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633
277. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
278. Xu L, et al. Mol. Med. (2001) PMID: 11713371
279. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
280. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
281. Pirolo KF, et al. Mol. Ther. (2016) PMID: 27357628
282. Hajdenberg et al., 2012; ASCO Abstract e15010
283. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
284. Moore et al., 2019; ASCO Abstract 5513
285. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27998224
286. Oza et al., 2015; ASCO Abstract 5506
287. Lee J, et al. Cancer Discov (2019) PMID: 31315834
288. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
289. Ma CX, et al. J. Clin. Invest. (2012) PMID: 22446188
290. Lehmann S, et al. J. Clin. Oncol. (2012) PMID: 22965953
291. Mohell N, et al. Cell Death Dis (2015) PMID: 26086967
292. Fransson Å, et al. J Ovarian Res (2016) PMID: 27179933
293. Gourley et al., 2016; ASCO Abstract 5571
294. Kwok M, et al. Blood (2016) PMID: 26563132
295. Boudny M, et al. Haematologica (2019) PMID: 30975914
296. Dillon MT, et al. Mol. Cancer Ther. (2017) PMID: 28062704
297. Middleton FK, et al. Cancers (Basel) (2018) PMID: 30127241
298. Mogi A, et al. J. Biomed. Biotechnol. (2011) PMID: 21331359
299. Tekpli X, et al. Int. J. Cancer (2013) PMID: 23011884
300. Vignot S, et al. J. Clin. Oncol. (2013) PMID: 23630207
301. Maeng CH, et al. Anticancer Res. (2013) PMID: 24222160
302. Itakura M, et al. Br. J. Cancer (2013) PMID: 23922113
303. Dong ZY, et al. Clin. Cancer Res. (2017) PMID: 28039262
304. Seo JS, et al. Genome Res. (2012) PMID: 22975805
305. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
306. Joerger AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
307. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
308. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
309. Zerdoury Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
310. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Erik Williams, M.D. | 13 January 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # **ORD-1275918-01**
APPENDIX
References

311. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
312. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
313. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
314. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
315. Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
316. Lalloo F, et al. Lancet (2003) pmid: 12672316
317. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713
318. Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
319. Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838
320. Xie M, et al. Nat. Med. (2014) pmid: 25326804
321. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
322. Severson EA, et al. Blood (2018) pmid: 29678827
323. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
324. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
325. Chabon JJ, et al. Nature (2020) pmid: 32269342
326. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
327. Glassberg et al., 2020; ASPHO Abstract 2015
328. Coyne et al., 2020; ASCO Abstract 3612
329. McCowage et al., 2018; ASCO Abstract 10504
330. Mueller et al., 2020; SNO Abstract NFB-17
331. Waldner et al., 2020; DOI: 10.1055/s-0040-1715638
332. Romo et al., 2019; SNO Abstract RARE-54
333. Lopez-Chavez A, et al. J. Clin. Oncol. (2015) pmid: 25667274
334. Hainsworth JD, et al. J Thorac Oncol (2010) pmid: 20802351
335. Soria JC, et al. Ann. Oncol. (2017) pmid: 29045535
336. Melosky B, et al. Lung Cancer (2019) pmid: 31200828
337. Greystoke A, et al. Br. J. Cancer (2017) pmid: 28950288
338. Oxnard GR, et al. Ann. Oncol. (2020) pmid: 32139298
339. Blumenschein GR, et al. Ann. Oncol. (2015) pmid: 25722381
340. Leijen S, et al. Clin. Cancer Res. (2012) pmid: 22767668
341. Zimmer L, et al. Clin. Cancer Res. (2014) pmid: 24947927
342. Kelly et al., 2013; ASCO Abstract 8027
343. Gandara et al., 2013; ASCO Abstract 8028
344. Jänne PA, et al. Lancet Oncol. (2013) pmid: 23200175
345. Carter CA, et al. Ann. Oncol. (2016) pmid: 26802155
346. Banerji et al., 2014; ASCO Abstract e13559
347. Castellano E, et al. Cancer Cell (2013) pmid: 24229709
348. Ku BM, et al. Invest New Drugs (2015) pmid: 25342139
349. Bedard PL, et al. Clin. Cancer Res. (2015) pmid: 25500057