

ABOUT THE TEST FoundationOne® Heme is a comprehensive genomic profiling test designed to identify genomic alterations within hundreds of cancer-related genes in hematologic malignancies and sarcomas.

PATIENT

DISEASE Soft tissue liposarcoma

DATE OF BIRTH 10 January 1941

SEX Male

MEDICAL RECORD # Not given

PHYSICIAN

MEDICAL FACILITY Arias Stella

ADDITIONAL RECIPIENT None

MEDICAL FACILITY ID 317319

PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Retroperitoneum

SPECIMEN ID 21-18594 9 (Q21-14642.9)

SPECIMEN TYPE Block

DATE OF COLLECTION 15 October 2021

SPECIMEN RECEIVED 21 January 2022

Biomarker Findings

Microsatellite status - MS-Stable
Tumor Mutational Burden - 2 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

CDK4 amplification
MDM2 amplification
CBL C384Y - subclonal[†]
FRS2 amplification
HMGA2 HMGA2-TSFM fusion

[†] See About the Test in appendix for details.

Report Highlights

- Variants with **diagnostic implications** that may indicate a specific cancer type: **CDK4 amplification** (p. 4), **MDM2 amplification** (p. 5)
- Targeted therapies with potential clinical benefit **approved in another tumor type**: Abemaciclib (p. 8)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 9)
- Variants that may represent **clonal hematopoiesis** and may originate from non-tumor sources: **CBL C384Y** (p. 6)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 2 Muts/Mb

GENOMIC FINDINGS

CDK4 - amplification

10 Trials see p. 9

MDM2 - amplification

5 Trials see p. 11

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE)

none

none

THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE)

Abemaciclib

none

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

CBL - C384Y - subclonal p. 6

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

CBL - C384Y - subclonal **p. 6** **HMGA2 - HMGA2-TSFM fusion** **p. 7**
FRS2 - amplification **p. 6**

NOTE Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type.
Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

ORDERED TEST # ORD-1289410-02

BIOMARKER FINDINGS
BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

In a computational analysis of paired tumor and normal sarcomas in the TCGA dataset, 25% of which were liposarcomas, only 0.8% (2/255) of samples were MSI-high (MSI-H)⁶. Smaller studies have reported MSI at any level in a subset of liposarcoma patients⁷⁻⁸ or reported as absent in 21 cases analyzed⁹. The prognostic significance of MSI in liposarcoma is unknown (PubMed, Jun 2021).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of

genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁰. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁰⁻¹². This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹³⁻¹⁵. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{10,12,14-15}.

BIOMARKER

Tumor Mutational Burden

RESULT

2 Muts/Mb

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁶⁻¹⁸, anti-PD-1 therapies¹⁶⁻¹⁹, and combination nivolumab and ipilimumab²⁰⁻²⁵. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors^{16-19,26}. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment for patients with 9 types of advanced tumors¹⁶. Analyses across several solid tumor types reported that patients with higher TMB (defined as ≥ 16 -20 Muts/Mb) achieved greater clinical benefit from

PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy²⁷ or those with lower TMB treated with PD-1 or PD-L1-targeting agents¹⁷. However, the KEYNOTE 158 trial of pembrolizumab monotherapy for patients with solid tumors found significant improvement in ORR for patients with TMB ≥ 10 Muts/Mb (based on this assay or others) compared to those with TMB < 10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials^{19,26}. Together, these studies suggest that patients with TMB ≥ 10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

FREQUENCY & PROGNOSIS

Liposarcoma harbors a median TMB of 1.7 mutations per megabase (mut/Mb), and 0.2% of cases have high TMB (> 20 muts/Mb)²⁸. Sarcomas in general harbor a median TMB of 2.5 muts/Mb, with angiosarcoma (13.4%) and malignant peripheral nerve sheath tumor (MPNST) (8.2%) having the highest percentage of cases with high TMB (> 20 muts/Mb)²⁸. Published data investigating the prognostic implications of TMB in sarcoma are limited (PubMed, Feb 2022). High

TMB was associated with improved PFS and metastasis-free survival in a study of undifferentiated sarcomas²⁹ and with reduced survival in a study of patients with rhabdomyosarcoma³⁰.

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma³¹⁻³² and cigarette smoke in lung cancer³³⁻³⁴, treatment with temozolomide-based chemotherapy in glioma³⁵⁻³⁶, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes³⁷⁻⁴¹, and microsatellite instability (MSI)^{37,40-41}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{17-18,26}.

ORDERED TEST # ORD-1289410-02

GENOMIC FINDINGS
GENE
CDK4
ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

CDK4 amplification or activation may predict sensitivity to CDK4/6 inhibitors such as abemaciclib, palbociclib, and ribociclib⁴²⁻⁴⁵. Clinical benefit has been reported for limited tumor types including patients with CDK4-amplified liposarcoma and sarcoma in response to treatment with abemaciclib⁴⁶, palbociclib^{42,47}, and ribociclib⁴⁸.

FREQUENCY & PROGNOSIS

In a genomic analysis of sarcomas, amplification of CDK4 was found in 24% of cases, including 88%

of dedifferentiated liposarcoma (DDLPS) cases, but was not observed in any of the 21 myxoid/round-cell liposarcoma cases or the 24 pleomorphic liposarcoma cases⁴⁹. Amplification of the chromosome subregion 12q13-15, with concomitant copy number gains of the CDK4 and MDM2 genes, is a hallmark genetic alteration in well-differentiated liposarcoma (WDLPS) and DDLPS, reported in up to 100% of cases^{42,50-51}. Amplification of CDK4 has been correlated with increased CDK4 protein expression in liposarcoma, undifferentiated high-grade pleomorphic sarcoma, leiomyosarcoma, and rhabdomyosarcoma⁵⁰⁻⁵³. CDK4 amplification predicts poor progression-free and disease-specific survival in patients with WDLPS or DDLPS as well as recurrence of WDLPS after surgical resection⁵⁴⁻⁵⁶.

FINDING SUMMARY

CDK4 encodes the cyclin-dependent kinase 4, which regulates the cell cycle, senescence, and

apoptosis⁵⁷. CDK4 and its functional homolog CDK6 are activated by D-type cyclins and promote cell cycle progression by inactivating the tumor suppressor Rb⁵⁸⁻⁵⁹. Amplification of the chromosomal region that includes CDK4 has been reported in multiple cancer types, including lung cancer, glioblastoma, and liposarcoma, and has been associated with overexpression of CDK4 protein^{42,50-53,60-62}.

POTENTIAL DIAGNOSTIC IMPLICATIONS

Amplification of 12q13-q15, including CDK4 and MDM2, is characteristic of well-differentiated liposarcoma (WDLPS) and dedifferentiated (DDLPS) liposarcoma (NCCN Soft Tissue Sarcoma Guidelines, v3.2021)^{49-50,63-64}, and low-grade osteosarcomas, including parosteal osteosarcomas and low-grade central osteosarcomas as opposed to conventional high-grade osteosarcoma (NCCN Soft Tissue Sarcoma Guidelines, v3.2021)^{62,65-68}.

ORDERED TEST # ORD-1289410-02

GENOMIC FINDINGS

GENE

MDM2

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

MDM2 antagonists disrupt the MDM2-p53 interaction, thereby stabilizing p53⁶⁹. Preclinical studies have suggested that the amplification of MDM2, in the absence of concurrent TP53 mutations, may increase sensitivity to these agents⁷⁰⁻⁷¹. Preliminary Phase 1 studies of the MDM2-p53 antagonist alrizomadlin (APG-115) reported a PR in a patient with liposarcoma harboring an MDM2 amplification and wildtype for TP53 and SD in 21%-38% (6/28 and 5/13, respectively) of patients in genomically unselected solid tumors⁷²⁻⁷³. A Phase 2 trial of alrizomadlin in combination with pembrolizumab reported a PR in 1 of 3 patients with malignant peripheral nerve sheath tumor that had failed standard therapy, as well as PRs in patients with multiple types of solid tumors that had failed immunotherapy, including 1 out of 14 patients with non-small cell lung cancer; 1 out of 5 patients with urothelial carcinoma; and 2 out of 5, 1 out of 5, and 1 out of 11 patients with mucosal, uveal, and cutaneous melanoma, respectively⁷⁴. Phase 1b

studies of the MDM2 inhibitor idasanutlin for refractory AML in combination with cytarabine or venetoclax reported anti-leukemic response rates of 33% (25/75) and 37% (11/30), respectively⁷⁵⁻⁷⁶; clinical benefit (58% ORR, 7/12) with idasanutlin monotherapy has been reported for patients with polycythemia vera⁷⁷. The dual MDM2/MDM4 inhibitor ALRN-6924 led to an ORR of 27% (4/15) for patients with TP53 wildtype peripheral T-cell lymphoma in a Phase 2 study⁷⁸; responses have also been observed in TP53 wildtype AML, MDS, Merkel cell carcinoma, colorectal cancer, and liposarcoma⁷⁹⁻⁸⁰.

FREQUENCY & PROGNOSIS

Amplification of MDM2 has reported in 27-33% of sarcomas overall and in up to 90% of liposarcomas^{49,63-64}. Amplification of the chromosome subregion 12q13-q15, with concomitant copy number gains of the CDK4 and MDM2 genes, is a hallmark genetic alteration in well-differentiated liposarcomas (WDLPS) and dedifferentiated (DDLPS) liposarcomas⁵⁰. Patients with a diagnosis of WDLPS or DDLPS and the accompanying 12q13-15 amplification have a high chance of recurrence and a poor chance of survival if surgical resection is incomplete⁵⁵.

FINDING SUMMARY

MDM2 encodes an E3 ubiquitin protein ligase, which mediates the ubiquitination and subsequent

degradation of p53, Rb1, and other proteins⁸¹⁻⁸³. MDM2 acts to prevent the activity of the tumor suppressor p53; therefore, overexpression or amplification of MDM2 may be oncogenic⁸⁴⁻⁸⁵. Overexpression or amplification of MDM2 is frequent in cancer⁸⁶. Although two retrospective clinical studies suggest that MDM2 amplification may predict a short time-to-treatment failure on anti-PD-1/PD-L1 immune checkpoint inhibitors, with 4/5 patients with MDM2 amplification⁸⁷ and 2/3 patients with MDM2 or MDM4 amplification⁸⁸ experiencing tumor hyperprogression, amplification of MDM2 or MDM4 was not associated with shorter progression-free survival (PFS) in a retrospective analysis of non-small cell lung cancer (NSCLC) outcomes with immune checkpoint inhibitors (hazard ratio of 1.4, p=0.44)⁸⁹. The latter study reported PFS of >2 months for 5/8 patients with MDM2/MDM4 amplification⁸⁹.

POTENTIAL DIAGNOSTIC IMPLICATIONS

Amplification of 12q13-q15, including CDK4 and MDM2, is characteristic of well-differentiated liposarcoma (WDLPS) and dedifferentiated (DDLPS) liposarcoma (NCCN Soft Tissue Sarcoma Guidelines, v3.2021)^{49-50,63-64}, and low-grade osteosarcomas, including parosteal osteosarcomas and low-grade central osteosarcomas as opposed to conventional high-grade osteosarcoma (NCCN Soft Tissue Sarcoma Guidelines, v3.2021)^{62,65-68}.

ORDERED TEST # ORD-1289410-02

GENOMIC FINDINGS

GENE

CBL

ALTERATION

C384Y - subclonal

TRANSCRIPT ID

NM_005188

CODING SEQUENCE EFFECT

1151G>A

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

CBL inactivation may lead to the hyperactivation of various receptor tyrosine kinases (RTKs), including MET⁹⁰, PDGFRA⁹¹, KIT⁹², VEGFR2⁹³, and the TAM (TYRO3, AXL, MER) RTKs⁹⁴. These RTKs are targets of the multikinase inhibitor sitravatinib⁹⁵, which has shown activity in CBL-mutated advanced solid tumors⁹⁶. Among 8 patients with CBL inactivating alterations in a Phase 1b trial, sitravatinib produced 2 PRs (25% ORR), with 1 NSCLC and 1 melanoma responding for over 4 months, and 4 SD outcomes, with 3 prolonged SDs seen in a patient with NSCLC, a

patient with esophageal cancer, and a patient with a pancreatic neuroendocrine tumor⁹⁶. CBL has been shown to downregulate EGFR⁹⁷⁻¹⁰¹ and FLT3¹⁰²⁻¹⁰⁴. Preclinical models of myeloid malignancies have demonstrated that CBL inactivation confers sensitivity to the FLT3-targeting therapies sunitinib¹⁰², midostaurin¹⁰⁴, and quizartinib¹⁰⁵, as well as to dasatinib¹⁰⁶, although clinical evidence for this approach in solid tumors is lacking.

FREQUENCY & PROGNOSIS

CBL alterations have been reported in 1% of liposarcoma cases (COSMIC, Oct 2021)¹⁰⁷. Published data investigating the prognostic implications of CBL alterations in liposarcoma are limited (PubMed, Oct 2021).

FINDING SUMMARY

CBL encodes an E3 ubiquitin protein ligase that is involved in cell signaling and ubiquitination, targeting proteins such as EGFR, FGFR1, FGFR2, PDGFR-alpha, PDGFR-beta, FLT3, and SRC for degradation by the proteasome¹⁰⁸⁻¹¹². CBL alterations that result in loss or disruption of the tyrosine kinase binding domain, RING finger

domain, and/or tail domain, as observed here, are predicted to be inactivating and to promote tumorigenesis¹¹³⁻¹³⁰.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion¹³¹⁻¹³⁶. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy¹³¹⁻¹³². Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹³⁷. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{135,138-139}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

GENE

FRS2

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies that target alterations in FRS2. Amplification of FRS2 can lead to activation of the FGFR and MAPK-ERK pathways, and preliminary studies in liposarcoma cell lines have shown that cells with this alteration are sensitive to FGFR inhibitors^{51,140}.

FREQUENCY & PROGNOSIS

FRS2 amplification is particularly prevalent in liposarcoma, where amplification of the 12q13-15 region of chromosome 12 is considered to be a hallmark genetic alteration, although the effects of amplification of CDK4 and MDM2, also located in this region, have been studied in more detail than FRS2 in this context¹⁴¹⁻¹⁴². Amplification of FRS2 has been observed in 93%-100% of dedifferentiated liposarcoma, 32% of undifferentiated high-grade pleomorphic sarcoma, and 100% of well-differentiated liposarcoma^{51,143}. Amplification of the 12p15 chromosomal region containing FRS2, but not CDK4 or MDM2, was found in 12.5% of high-grade serous ovarian carcinomas, and knockdown of FRS2 in these cells resulted in apoptosis, indicating that cells with

12p15 amplification require FRS2¹⁴⁰.

FINDING SUMMARY

FRS2 encodes the fibroblast growth factor receptor (FGFR) substrate 2, an adaptor protein involved in FGFR signaling, which may also mediate signaling through EGFR, NTRK, and VEGF receptors¹⁴⁴⁻¹⁴⁷. FRS2 amplification was found to correlate with overexpression in high-grade serous ovarian tumors, and FRS2 overexpression promoted tumorigenesis in a preclinical study¹⁴⁰.

ORDERED TEST # ORD-1289410-02

GENOMIC FINDINGS
GENE

HMGA2

ALTERATION

HMGA2-TSFM fusion

POTENTIAL TREATMENT STRATEGIES
— Targeted Therapies —

There are no targeted therapies available to address fusions involving the HMGA2 gene. However, HMGA2 expression has been reported to be the driver of embryonic rhabdomyosarcoma (ERMS) via up-regulation of NRAS expression¹⁴⁸, suggesting that MEK inhibitors that are predicted to inhibit NRAS-dependent signaling may be

beneficial for patients with HMGA2 activation or overexpression. However, it is unknown whether this approach would be beneficial here.

FREQUENCY & PROGNOSIS

HMGA2 rearrangements have been most frequently identified in benign neoplasms such as lipomas, uterine leiomyomas, angiomyxomas, as well as in malignant tumors such as well-differentiated liposarcomas and inflammatory myofibroblastic tumors¹⁴⁹⁻¹⁵⁴. Amplifications of HMGA2 have also been described in a variety of tumors^{148,151,155-156}, including in up to 25% of sarcomas, including uterine leiomyomas, leiomyosarcomas, embryonic rhabdomyosarcomas (ERMS), and lipomas^{152,157-159}.

FINDING SUMMARY

HMGA2 encodes a protein that may act as a transcriptional regulator¹⁵⁰. Rearrangements involving HMGA2 typically result in a separation of the three DNA-binding domains, encoded by exons 1-3 (or sometimes two DNA-binding domains encoded by exons 1-2), from the modulatory acidic tail encoded by exons 4-5 as well as the 3' UTR that has been reported to inhibit HMGA2 expression^{151-154,160}. Both truncations of HMGA2 at exon 3 and HMGA2 (exon 1-3)-involving fusions have been reported to transform cultured cells¹⁶¹. HMGA2-RAD51B fusions have been reported to be produced in t(12;14) uterine leiomyomas and reported to lead to HMGA2 overexpression¹⁶²⁻¹⁶³.

ORDERED TEST # ORD-1289410-02

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Abemaciclib

Assay findings association
CDK4
amplification

AREAS OF THERAPEUTIC USE

Abemaciclib inhibits the cyclin-dependent kinases 4 and 6 (CDK4/6) and is FDA approved to treat adults with hormone-receptor-positive (HR+), HER2-negative (HER2-) breast cancer as monotherapy as well as in combination with tamoxifen or an aromatase inhibitor, including anastrozole, letrozole, and exemestane. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Targeting CDK4 amplification with abemaciclib may be an effective strategy to treat advanced liposarcoma based on a positive Phase 2 study⁴⁶.

SUPPORTING DATA

A Phase 2 trial of abemaciclib for patients with metastatic or recurrent dedifferentiated liposarcoma met the primary endpoint with a 12-week PFS rate of 76% (22/29) and reported a median PFS of 30 weeks and an ORR of 3% (1/29), with 3 additional patients experiencing tumor size reduction by >10%; all evaluable patients had CDK4 and MDM2 amplification and intact RB1⁴⁶. Studies with other CDK4/6 inhibitors also observed clinical benefit for a subset of patients with CDK4-amplified liposarcoma^{42,45,47,164}.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

ORDERED TEST # ORD-1289410-02

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
CDK4
RATIONALE
CDK4 amplification may predict sensitivity to

CDK4/6 inhibitors.

ALTERATION
amplification

NCT02693535
PHASE 2

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

TARGETS
VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, CDK4, CDK6, FLT3, CSF1R, KIT, RET, mTOR, EGFR, ERBB2, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4

LOCATIONS: Florida, Georgia, South Carolina, Texas, Alabama, North Carolina

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
TRKB, ALK, TRKC, ROS1, TRKA, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Florida, Louisiana, Texas, Mississippi, Georgia

NCT04801966
PHASE NULL

Safety and Oversight of the Individually Tailored Treatment Approach: A Novel Pilot Study

TARGETS
CDK4, CDK6, PI3K-alpha, PD-L1, MEK, PARP, PD-1, BRAF

LOCATIONS: Melbourne (Australia)

NCT03310879
PHASE 2

Study of the CDK4/6 Inhibitor Abemaciclib in Solid Tumors Harboring Genetic Alterations in Genes Encoding D-type Cyclins or Amplification of CDK4 or CDK6

TARGETS
CDK4, CDK6

LOCATIONS: Massachusetts

ORDERED TEST # ORD-1289410-02

CLINICAL TRIALS
NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS

VEGFRs, ABL, SRC, ALK, ROS1, AXL, TRKA, MET, TRKC, DDR2, KIT, EGFR, PD-1, CTLA-4, PARP, CDK4, CDK6, FLT3, CSF1R, RET, mTOR, ERBB2, MEK, BRAF, SMO

LOCATIONS: London (Canada), Toronto (Canada), Kingston (Canada), Ottawa (Canada), Montreal (Canada), Regina (Canada), Saskatoon (Canada), Edmonton (Canada), Vancouver (Canada)

NCT03784014
PHASE 3

MOLECULAR PROFILING OF ADVANCED SOFT-TISSUE SARCOMAS

TARGETS

ABL, KIT, ROS1, ALK, MET, ERBB2, EGFR, BRAF, MEK, PARP, PD-L1, CDK4, CDK6

LOCATIONS: Bordeaux (France), Saint-Herblain (France), Clermont-Ferrand (France), Villejuif (France), Paris (France), Marseille (France), Lyon (France), Dijon (France)

NCT04557449
PHASE 1

Study to Test the Safety and Tolerability of PF-07220060 in Participants With Advance Solid Tumors

TARGETS

CDK4, Aromatase, ER

LOCATIONS: Texas, Tennessee, Connecticut, Massachusetts, Michigan

NCT03114527
PHASE 2

Phase II Trial of Ribociclib and Everolimus in Advanced Dedifferentiated Liposarcoma (DDL) and Leiomyosarcoma (LMS)

TARGETS

mTOR, CDK6, CDK4

LOCATIONS: Pennsylvania

NCT04438824
PHASE 2

Palbociclib and INCMGA00012 in People With Advanced Liposarcoma

TARGETS

CDK4, CDK6, PD-1

LOCATIONS: New Jersey, New York

NCT03065062
PHASE 1

Study of the CDK4/6 Inhibitor Palbociclib (PD-0332991) in Combination With the PI3K/mTOR Inhibitor Gedatolisib (PF-05212384) for Patients With Advanced Squamous Cell Lung, Pancreatic, Head & Neck and Other Solid Tumors

TARGETS

PI3K-alpha, PI3K-gamma, mTORC1, mTORC2, CDK4, CDK6

LOCATIONS: Massachusetts

ORDERED TEST # ORD-1289410-02

CLINICAL TRIALS

GENE
MDM2

RATIONALE
Inhibitors of the MDM2-p53 interaction are being tested in clinical trials. Overexpression or amplification of MDM2 may increase sensitivity to these agents, but more data are required.

ALTERATION
amplification

NCT04979442
PHASE 3

Treatment of Milademetan Versus Trabectedin in Patient With Dedifferentiated Liposarcoma

TARGETS
FUS-DDIT3

LOCATIONS: Florida, Texas, Pennsylvania, Missouri, New York, Ohio, Massachusetts, Michigan, Toronto (Canada)

NCT04589845
PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

TARGETS
TRKB, ALK, TRKC, ROS1, TRKA, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

LOCATIONS: Sao Paulo (Brazil), San Juan (Puerto Rico), Florida, Alabama, Texas, Georgia, South Carolina

NCT03611868
PHASE 1/2

A Study of APG-115 in Combination With Pembrolizumab in Patients With Metastatic Melanomas or Advanced Solid Tumors

TARGETS
MDM2, PD-1

LOCATIONS: Florida, Texas, Tennessee, Virginia, Arkansas, District of Columbia, Pennsylvania, Missouri

NCT03449381
PHASE 1

This Study Aims to Find the Best Dose of BI 907828 in Patients With Different Types of Advanced Cancer (Solid Tumors)

TARGETS
MDM2

LOCATIONS: Florida, Tennessee, New York, Connecticut, Ottawa (Canada), Barcelona (Spain), Leuven (Belgium), Tübingen (Germany), Berlin (Germany), Tokyo, Chuo-ku (Japan)

NCT03725436
PHASE 1

ALRN-6924 and Paclitaxel in Treating Patients With Advanced, Metastatic, or Unresectable Solid Tumors

TARGETS
MDM2, MDM4

LOCATIONS: Texas

ORDERED TEST # ORD-1289410-02

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future. Please note that some VUS rearrangements between targeted genes and unknown fusion partners or intergenic regions detected by RNA sequencing may not be reported.

APC

S881F

CARD11

L706F

CUX1

Q1245R

ECT2L

amplification

ESR1

amplification

FOXO3

amplification

GATA3

amplification

GSK3B

amplification

INPP5D

L1061P

KMT2A (MLL)

A53V

MAP3K7

amplification

NOTCH1

D740H

PRDM1

amplification

SGK1

amplification

SPEN

S2306del

TNFAIP3

amplification

TSC1

K587R

WISP3

amplification

ORDERED TEST # ORD-1289410-02

APPENDIX
Genes Assayed in FoundationOne®Heme

FoundationOne Heme is designed to include genes known to be somatically altered in human hematologic malignancies and sarcomas that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay utilizes DNA sequencing to interrogate 406 genes as well as selected introns of 31 genes involved in rearrangements, in addition to RNA sequencing of 265 genes. The assay will be updated periodically to reflect new knowledge about cancer biology.

HEMATOLOGICAL MALIGNANCY DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACTB	AKT1	AKT2	AKT3	ALK	AMER1 (FAM123B or WTX)	APC
APH1A	AR	ARAF	ARFRP1	ARHGAP26 (GRAF)		ARID1A	ARID2
ASXL1	ATM	ATR	ATRX	AURKA	AURKB	AXIN1	AXL
BAP1	BARD1	BCL10	BCL11B	BCL2	BCL2L2	BCL6	BCL7A
BCORL1	BIRC3	BLM	BRAF	BRCA1	BRCA2	BRD4	BRIP1
BTG2	BTK	BTLA	C11orf30 (EMSY)	CAD	CALR*	CARD11	CBFB
CCND1	CCND2	CCND3	CCNE1	CCT6B	CD22	CD274 (PD-L1)	CD36
CD70	CD79A	CD79B	CDC73	CDH1	CDK12	CDK4	CDK6
CDKN1B	CDKN2A	CDKN2B	CDKN2C	CEBPA	CHD2	CHEK1	CHEK2
CIITA	CKS1B	CPS1	CREBBP	CRKL	CRLF2	CSF1R	CSF3R
CTNNA1	CTNNB1	CUX1	CXCR4	DAXX	DDR2	DDX3X	DNM2
DOT1L	DTX1	DUSP2	DUSP9	EBF1	ECT2L	EED	EGFR
EP300	EPHA3	EPHA5	EPHA7	EPHB1	ERBB2	ERBB3	ERBB4
ESR1	ETS1	ETV6	EXOSC6	EZH2	FAF1	FAM46C	FANCA
FANCD2	FANCE	FANCF	FANCG	FANCL	FAS (TNFRSF6)	FBXO11	FBXO31
FGF10	FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1
FGFR3	FGFR4	FHIT	FLCN	FLT1	FLT3	FLT4	FLYWCH1
FOXO1	FOXO3	FOXP1	FRS2	GADD45B	GATA1	GATA2	GATA3
GNA11	GNA12	GNA13	GNAQ	GNAS	GPR124	GRIN2A	GSK3B
HDAC1	HDAC4	HDAC7	HGF	HIST1H1C	HIST1H1D	HIST1H1E	HIST1H2AC
HIST1H2AL	HIST1H2AM	HIST1H2BC	HIST1H2BJ	HIST1H2BK	HIST1H2BO	HIST1H3B	HNF1A
HSP90AA1	ICK	ID3	IDH1	IDH2	IGF1R	IKBKE	IKZF1
IKZF3	IL7R	INHBA	INPP4B	INPP5D (SHIP)	IRF1	IRF4	IRF8
JAK1	JAK2	JAK3	JARID2	JUN	KAT6A (MYST3)	KDM2B	KDM4C
KDM5C	KDM6A	KDR	KEAP1	KIT	KLHL6	KMT2A (MLL)	KMT2C (MLL3)
KRAS	LEF1	LRP1B	LRRK2	MAF	MAFB	MAGED1	MALT1
MAP2K2	MAP2K4	MAP3K1	MAP3K14	MAP3K6	MAP3K7	MAPK1	MCL1
MDM4	MED12	MEF2B	MEF2C	MEN1	MET	MIB1	MITF
MLH1	MPL	MRE11A	MSH2	MSH3	MSH6	MTOR	MUTYH
MYCL (MYCL1)	MYCN	MYD88	MYO18A	NCOR2	NCSTN	NF1	NF2
NFKBIA	NKX2-1	NOD1	NOTCH1	NOTCH2	NPM1	NRAS	NT5C2
NTRK2	NTRK3	NUP93	NUP98	P2RY8	PAG1	PAK3	PALB2
PAX5	PBRM1	PC	PCBP1	PCLO	PDCD1	PDCD11	PDCD1LG2 (PD-L2)
PDGFRB	PDK1	PHF6	PIK3CA	PIK3CG	PIK3R1	PIK3R2	PIM1
POT1	PPP2R1A	PRDM1	PRKAR1A	PRKDC	PRSS8	PTCH1	PTEN
PTPN2	PTPN6 (SHP-1)	PTPRO	RAD21	RAD50	RAD51	RAF1	RARA
RB1	RELN	RET	RHOA	RICTOR	RNF43	ROS1	RPTOR
S1PR2	SDHA	SDHB	SDHC	SDHD	SERP2	SETBP1	SETD2
SGK1	SMAD2	SMAD4	SMARCA1	SMARCA4	SMARCB1	SMC1A	SMC3
SOC1	SOC2	SOC3	SOX10	SOX2	SPEN	SPOP	SRC
STAG2	STAT3	STAT4	STAT5A	STAT5B	STAT6	STK11	SUFU
TAF1	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TET2	TGFBR2	TLL2	TMEM30A
TMSB4XP8 (TMSL3)		TNFAIP3	TNFRSF11A	TNFRSF14	TNFRSF17	TOP1	TP53
TRAF2	TRAF3	TRAF5	TSC1	TSC2	TSHR	TUSC3	TYK2
U2AF2	VHL	WDR90	WHSC1 (MMSET or NSD2)		WISP3	WT1	XBP1
YY1AP1	ZMYM3	ZNF217	ZNF24 (ZSCAN3)	ZNF703	ZRSR2		

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Chelsea Marcus, M.D. | 28 February 2022
Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1289410-02

APPENDIX
Genes Assayed in FoundationOne®Heme

*Note: the assay was updated on 11/8/2016 to include the detection of alterations in CALR

HEMATOLOGICAL MALIGNANCY DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCL6	BCR	BRAF	CCND1	CRLF2	EGFR	EPOR
ETV1	ETV4	ETV5	ETV6	EWSR1	FGFR2	IGH	IGK	IGL
JAK1	JAK2	KMT2A (MLL)	MYC	NTRK1	PDGFRA	PDGFRB	RAF1	RARA
RET	ROS1	TMPRSS2	TRG					

HEMATOLOGICAL MALIGNANCY RNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS*

ABI1	ABL1	ABL2	ACSL6	AFF1	AFF4	ALK	ARHGAP26 (GRAF)	
ARHGEF12	ARID1A	ARNT	ASXL1	ATF1	ATG5	ATIC	BCL10	BCL11A
BCL11B	BCL2	BCL3	BCL6	BCL7A	BCL9	BCOR	BCR	BIRC3
BRAF	BTG1	CAMTA1	CARS	CBFA2T3	CBFB	CBL	CCND1	CCND2
CCND3	CD274 (PD-L1)	CDK6	CDX2	CHIC2	CHN1	CIC	CIITA	CLP1
CLTC	CLTCL1	CNTRL (CEP110)	COL1A1	CREB3L1	CREB3L2	CREBBP	CRLF2	CSF1
CTNNB1	DDIT3	DDX10	DDX6	DEK	DUSP22	EGFR	EIF4A2	ELF4
ELL	ELN	EML4	EP300	EPOR	EPS15	ERBB2	ERG	ETS1
ETV1	ETV4	ETV5	ETV6	EWSR1	FCGR2B	FCRL4	FEV	FGFR1
FGFR1OP	FGFR2	FGFR3	FLI1	FBNP1	FOXO1	FOXO3	FOXO4	FOXP1
FSTL3	FUS	GAS7	GLI1	GMPS	GPHN	HERPUD1	HEY1	HIP1
HIST1H4I	HLF	HMGA1	HMGA2	HOXA11	HOXA13	HOXA3	HOXA9	HOXC11
HOXC13	HOXD11	HOXD13	HSP90AA1	HSP90AB1	IGH	IGK	IGL	IKZF1
IL21R	IL3	IRF4	ITK	JAK1	JAK2	JAK3	JAZF1	KAT6A (MYST3)
KDSR	KIF5B	KMT2A (MLL)	LASP1	LCP1	LMO1	LMO2	LPP	LYL1
MAF	MAFB	MALT1	MDS2	MECOM	MKL1	MLF1	MLLT1 (ENL)	MLLT10 (AF10)
MLLT3	MLLT4 (AF6)	MLLT6	MN1	MNX1	MSI2	MSN	MUC1	MYB
MYC	MYH11	MYH9	NACA	NBEAP1 (BCL8)	NCOA2	NDRG1	NF1	NF2
NFKB2	NIN	NOTCH1	NPM1	NR4A3	NSD1	NTRK1	NTRK2	NTRK3
NUMA1	NUP214	NUP98	NUTM2A	OMD	P2RY8	PFAFH1B2	PAX3	PAX5
PAX7	PBX1	PCM1	PCSK7	PDCD1LG2 (PD-L2)	PDE4DIP	PDGFB	PDGFRA	PDGFRB
PER1	PHF1	PICALM	PIM1	PLAG1	PML	POU2AF1	PPP1CB	PRDM1
PRDM16	PRRX1	PSIP1	PTCH1	PTK7	RABEP1	RAF1	RALGDS	RAP1GDS1
RARA	RBM15	RET	RHOH	RNF213	ROS1	RPL22	RPN1	RUNX1
RUNX1T1 (ETO)	RUNX2	SEC31A	SEPT5	SEPT6	SEPT9	SET	SH3GL1	SLC1A2
SNX29 (RUNDC2A)	SRSF3	SS18	SSX1	SSX2	SSX4	STAT6	STL	SYK
TAF15	TAL1	TAL2	TBL1XR1	TCF3 (E2A)	TCL1A (TCL1)	TEC	TET1	TFE3
TFG	TFPT	TFRC	TLX1	TLX3	TMPRSS2	TNFRSF11A	TOP1	TP63
TPM3	TPM4	TRIM24	TRIP11	TTL	TYK2	USP6	WHSC1 (MMSET or NSD2)	
WHSC1L1	YPEL5	ZBTB16	ZMYM2	ZNF384	ZNF521			

*Note: some VUS rearrangements between targeted genes and unknown fusion partners or intergenic regions detected by RNA sequencing may not be reported.

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Microsatellite (MS) status
Tumor Mutational Burden (TMB)

ORDERED TEST # ORD-1289410-02

APPENDIX
Performance Specifications

The median exon coverage for this sample is 853x

ACCURACY

Sensitivity: Base Substitutions	At $\geq 5\%$ Minor Allele Frequency	>99.0%
Sensitivity: Insertions/Deletions (1-40bp)	At $\geq 10\%$ Minor Allele Frequency	98.0%
Sensitivity: Focal Copy Number Alterations (Homozygous Deletions or Amplifications)	At ≥ 8 copies	>95.0%
Sensitivity: Microsatellite Instability-High (MSI-H) status	Positive Predictive Agreement (PPA)	100.0% (87.54%-100.00%)*
Sensitivity: Microsatellite Stable (MSS) status	Positive Predictive Agreement (PPA)	89.66% (81.50%, 94.46%)*
Sensitivity: Known Gene Fusions	>95.0%	
Specificity: Base Substitutions, Insertions/Deletions, and Focal Copy Number Alterations	Positive Predictive Value (PPV)	>99.0%
Specificity: Known Gene Fusions	Positive Predictive Value (PPV)	>95.0%
Specificity: Microsatellite Instability-High (MSI-H) status	Negative Predictive Agreement (NPA)	97.44% (91.12%-99.29%)*
Specificity: Microsatellite Stable (MSS) status	Negative Predictive Agreement (NPA)	94.44% (86.57%, 97.82%)*
Accuracy: Tumor Mutation Burden	At $\geq 20\%$ tumor nuclei	>90.0%
Reproducibility (average concordance between replicates)	97.0% inter-batch precision 97.0% intra-batch precision 95.0% microsatellite status precision 96.0% tumor mutation burden precision	

*95% Confidence Interval

Assay specifications were determined for typical median exon coverage of approximately 500X. For additional information regarding the validation of FoundationOne®Heme, please refer to the article He, J. et al. Integrated genomic DNA/RNA profiling of hematologic malignancies in the clinical setting, Blood (2016 Jun. 16).

In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the FoundationOne Heme assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator

assay using a pan-tumor sample set. Patients with results categorized as "MS-Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.

Tumor Mutational Burden (TMB) is determined by measuring the number of somatic mutations in sequenced genes on the FoundationOne Heme test and extrapolating to the genome as a whole. TMB is assayed for all FoundationOne Heme samples and is reported as the number of mutations per megabase (Muts/Mb). Tumor Mutational Burden is reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine Tumor Mutational Burden.

© 2022 Foundation Medicine, Inc. All rights reserved.

Electronically signed by Chelsea Marcus, M.D. | 28 February 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1289410-02

APPENDIX

About FoundationOne®Heme

ABOUT FOUNDATIONONE HEME

FoundationOne®Heme is a comprehensive genomic profiling test for hematologic malignancies and sarcomas. The test is designed to provide physicians with clinically actionable information to help with diagnostic sub-classification, prognosis assessment, and targeted therapeutic selection. Test results provide information about clinically significant alterations, potential targeted therapies, available clinical trials, and quantitative markers that may support immunotherapy clinical trial enrollment. FoundationOne Heme is analytically validated to detect all classes of genomic alterations in more than 400 cancer-related genes. In addition to DNA sequencing, FoundationOne Heme employs RNA sequencing across 265 genes to capture a broad range of gene fusions, common drivers of hematologic malignancies and sarcomas.

FoundationOne Heme was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Heme has not been cleared or approved by the United States Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. FoundationOne Heme may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high-complexity clinical testing.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne Heme identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls
(Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne Heme for identifying a copy number amplification is five (5) for *ERBB2* and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that FoundationOne Heme data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that FoundationOne Heme analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials
Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source

or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Heme.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. These include: subclonal alterations in heterogeneous samples, low sample quality or with homozygous losses of <3 exons; and deletions and insertions >40bp, or in repetitive/high homology sequences. FoundationOne Heme is performed using DNA and RNA derived from tumor, and as such germline events may not be reported.

The following targets typically have low coverage resulting in a reduction in sensitivity: SDHD exon 4, TNFRSF11A exon1, and TP53 exon 1.

FoundationOne Heme fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a., Cipalstraat 3, 2440 Geel, Belgium.

ORDERED TEST # ORD-1289410-02

APPENDIX

About FoundationOne®Heme

CE

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*),

MPL, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 6.0.0

ORDERED TEST # ORD-1289410-02

APPENDIX

References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Bonneville R, et al. JCO Precis Oncol (2017) PMID: 29850653
7. Davis JL, et al. Arch. Pathol. Lab. Med. (2014) PMID: 24878023
8. Suwa K, et al. J Orthop Sci (1999) PMID: 10370164
9. Schneider-Stock R, et al. Int. J. Oncol. (1999) PMID: 10087320
10. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
11. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
12. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
13. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
14. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
15. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
16. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
17. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
18. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
19. Cristescu R, et al. Science (2018) PMID: 30309915
20. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
21. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
22. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
23. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
24. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
25. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
26. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
27. Legrand et al., 2018; ASCO Abstract 12000
28. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
29. Steele CD, et al. Cancer Cell (2019) PMID: 30889380
30. Casey DL, et al. Clin Cancer Res (2020) PMID: 31699828
31. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
32. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
33. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
34. Rizvi NA, et al. Science (2015) PMID: 25765070
35. Johnson BE, et al. Science (2014) PMID: 24336570
36. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
37. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
38. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
39. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
40. Nature (2012) PMID: 22810696
41. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
42. Dickson MA, et al. J. Clin. Oncol. (2013) PMID: 23569312
43. Flaherty KT, et al. Clin. Cancer Res. (2012) PMID: 22090362
44. Patnaik A, et al. Cancer Discov (2016) PMID: 27217383
45. Infante JR, et al. Clin. Cancer Res. (2016) PMID: 27542767
46. Dickson et al., 2019; ASCO Abstract 11004
47. Dickson MA, et al. JAMA Oncol (2016) PMID: 27124835
48. Peguero et al., 2016; ASCO Abstract 2528
49. Barretina J, et al. Nat. Genet. (2010) PMID: 20601955
50. Horvai AE, et al. Mod. Pathol. (2009) PMID: 19734852
51. Zhang K, et al. Cancer Res. (2013) PMID: 23393200
52. Chung L, et al. Am. J. Surg. Pathol. (2009) PMID: 19574885
53. Ragazzini P, et al. Histol. Histopathol. (2004) PMID: 15024701
54. Lee SE, et al. Histol. Histopathol. (2014) PMID: 23852861
55. Crago AM, et al. Curr Opin Oncol (2011) PMID: 21552124
56. Lee S, et al. PLoS ONE (2014) PMID: 25121597
57. Choi YJ, et al. Oncogene (2014) PMID: 23644662
58. Cell (1995) PMID: 7736585
59. Musgrave EA, et al. Nat. Rev. Cancer (2011) PMID: 21734724
60. Wikman H, et al. Genes Chromosomes Cancer (2005) PMID: 15543620
61. Rao SK, et al. J. Neurooncol. (2010) PMID: 19609742
62. Dujardin F, et al. Mod. Pathol. (2011) PMID: 21336260
63. Oliner JD, et al. Nature (1992) PMID: 1614537
64. Kashima T, et al. Mod. Pathol. (2012) PMID: 22699518
65. Mejia-Guerrero S, et al. Genes Chromosomes Cancer (2010) PMID: 20196171
66. Yoshida A, et al. Am. J. Surg. Pathol. (2012) PMID: 22301501
67. Park HR, et al. Pathol. Res. Pract. (2004) PMID: 15310147
68. Wunder JS, et al. Oncogene (1999) PMID: 9989829
69. Cheok CF, et al. Nat Rev Clin Oncol (2011) PMID: 20975744
70. Ohnstad HO, et al. Cancer (2013) PMID: 23165797
71. Gamble LD, et al. Oncogene (2012) PMID: 21725357
72. Zhang et al., 2019; ASCO Abstract 3124
73. Rasco et al., 2019; ASCO Abstract 3126
74. Tolcher et al., 2021; ASCO Abstract 2506
75. Martinelli et al., 2016; EHA21 Abstract S504
76. Daver et al., 2018; ASH Abstract 767
77. Mascarenhas et al., 2019; ASH Abstract 134
78. Shustov et al., 2018; ASH Abstract 1623
79. Sallman et al., 2018; ASH Abstract 4066
80. Meric-Bernstam et al., 2017; ASCO Abstract 2505
81. Sdek P, et al. Mol. Cell (2005) PMID: 16337594
82. Brady M, et al. Mol. Cell. Biol. (2005) PMID: 15632057
83. Li M, et al. Mol. Cell (2004) PMID: 15053880
84. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
85. Cordon-Cardo C, et al. Cancer Res. (1994) PMID: 8306343
86. Beroukhim R, et al. Nature (2010) PMID: 20164920
87. Kato S, et al. Clin. Cancer Res. (2017) PMID: 28351930
88. Singavi et al., 2017; ESMO Abstract 1140PD
89. Rizvi H, et al. J. Clin. Oncol. (2018) PMID: 29337640
90. Mancini A, et al. J. Biol. Chem. (2002) PMID: 11847211
91. Miyake S, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) PMID: 9653117
92. Masson K, et al. Biochem. J. (2006) PMID: 16780420
93. Singh AJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17372230
94. Paolino M, et al. Nature (2014) PMID: 24553136
95. Patwardhan PP, et al. Oncotarget (2016) PMID: 26675259
96. Bazhenova et al., 2018; ESMO Abstract 4080
97. Shtiegman K, et al. Oncogene (2007) PMID: 17486068
98. Padrón D, et al. Cancer Res. (2007) PMID: 17699773
99. Hosaka T, et al. Anticancer Res. () PMID: 17695511
100. Han W, et al. Cancer Biol. Ther. (2006) PMID: 16969069
101. Yang S, et al. Cancer Res. (2006) PMID: 16849543
102. Sargin B, et al. Blood (2007) PMID: 17446348
103. Oshikawa G, et al. J. Biol. Chem. (2011) PMID: 21768087
104. Reindl C, et al. Clin. Cancer Res. (2009) PMID: 19276253
105. Taylor SJ, et al. Blood (2012) PMID: 22990016
106. Makishima H, et al. Leukemia (2012) PMID: 22246246
107. Tate JG, et al. Nucleic Acids Res. (2019) PMID: 30371878
108. Bacher U, et al. Ann. Hematol. (2010) PMID: 20195608
109. Miyake S, et al. J. Biol. Chem. (1999) PMID: 10347229
110. Polzer H, et al. Exp. Hematol. (2013) PMID: 23217761
111. Levkowitz G, et al. Genes Dev. (1998) PMID: 9851973
112. Bunda S, et al. Cancer Res. (2013) PMID: 23400592
113. Andoniou CE, et al. EMBO J. (1994) PMID: 7925293
114. Aranaz P, et al. Haematologica (2012) PMID: 22315494
115. Fernandes MS, et al. J. Biol. Chem. (2010) PMID: 20622007
116. Grand FH, et al. Blood (2009) PMID: 19387008
117. Javadi M, et al. J. Biol. Chem. (2013) PMID: 23696637
118. Kassenbrock CK, et al. J. Biol. Chem. (2004) PMID: 15117950
119. Levkowitz G, et al. Mol. Cell (1999) PMID: 10635327
120. Loh ML, et al. Blood (2009) PMID: 19571318
121. Martinelli S, et al. Am. J. Hum. Genet. (2010) PMID: 20619386
122. Saito Y, et al. Leuk. Res. (2012) PMID: 22591685
123. Sanada M, et al. Nature (2009) PMID: 19620960
124. Score J, et al. Blood (2012) PMID: 22053108
125. Shiba N, et al. Leukemia (2011) PMID: 21494262
126. Standaert ML, et al. Biochemistry (2004) PMID: 15581361
127. Tan YH, et al. PLoS ONE (2010) PMID: 20126411
128. Thien CB, et al. Mol. Cell (2001) PMID: 11239464
129. Visser GD, et al. Exp. Cell Res. (2005) PMID: 16246327
130. Li M, et al. Cancer Res. (2016) PMID: 26676746
131. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
132. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
133. Xie M, et al. Nat. Med. (2014) PMID: 25326804
134. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
135. Severson EA, et al. Blood (2018) PMID: 29678827
136. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
137. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
138. Chabon JJ, et al. Nature (2020) PMID: 32269342
139. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
140. Luo LY, et al. Mol. Cancer Res. (2015) PMID: 25368431
141. Pilotti S, et al. J. Pathol. (1998) PMID: 9713346
142. Segura-Sánchez J, et al. Anticancer Res. () PMID: 17214366
143. Wang X, et al. Genes Chromosomes Cancer (2011) PMID: 21793095
144. Kanodia J, et al. Cell Commun. Signal (2014) PMID: 24885272
145. Wu Y, et al. Biol. Chem. (2003) PMID: 12974390
146. Chen PY, et al. Proc. Natl. Acad. Sci. U.S.A. (2014) PMID: 24706887
147. Yan KS, et al. J. Biol. Chem. (2002) PMID: 11877385
148. Li Z, et al. Cancer Res. (2013) PMID: 23536553
149. Panagopoulos I, et al. Int. J. Oncol. (2015) PMID: 26202160
150. Cleyne I, et al. Int. J. Oncol. (2008) PMID: 18202751
151. Henriksen J, et al. BMC Cancer (2010) PMID: 20576167
152. Berner JM, et al. Oncogene (1997) PMID: 9205100
153. Dahlén A, et al. Mod. Pathol. (2003) PMID: 14614053
154. Meza-Zepeda LA, et al. Genes Chromosomes Cancer (2001) PMID: 11391797
155. Romero-Pérez L, et al. Hum. Pathol. (2013) PMID: 22974476
156. Zhang K, et al. Clin. Cancer Res. (2014) PMID: 24423609
157. Bertsch E, et al. Mod. Pathol. (2014) PMID: 24390224

© 2022 Foundation Medicine, Inc. All rights reserved.

 Electronically signed by Chelsea Marcus, M.D. | 28 February 2022
 Julia Elvin, M.D., Ph.D., Laboratory Director CLIA: 22D2027531
 Nimesh R. Patel, M.D., Laboratory Director CLIA: 34D2044309
 Foundation Medicine, Inc. | 1.888.988.3639

 Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
 Sample Preparation: 7010 Kit Creek Road, Morrisville, NC 27560 · CLIA: 34D2044309
 Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # ORD-1289410-02

APPENDIX
References

- | | | |
|--|---|--|
| <p>158. Italiano A, et al. Int. J. Cancer (2008) pmid: 18214854</p> <p>159. Biochem. Pharmacol. (2012) pmid: 22387046</p> <p>160. Shi G, et al. J. Cell. Mol. Med. (2009) pmid: 19602040</p> | <p>161. Fedele M, et al. Oncogene (1998) pmid: 9696033</p> <p>162. Schoenmakers EF, et al. Cancer Res. (1999) pmid: 9892177</p> | <p>163. Schoenmakers EF, et al. Genes Chromosomes Cancer (2013) pmid: 22965931</p> <p>164. Razak et al., 2018; AACR Abstract CT009</p> |
|--|---|--|