

ABOUT THE TEST FoundationOne®Liquid CDx is a next generation sequencing (NGS) assay that identifies clinically relevant genomic alterations in circulating cell-free DNA.

PATIENT

DISEASE Pancreas cancer (NOS)

DATE OF BIRTH 17 October 1949

SEX Male

MEDICAL RECORD # Not given

PHYSICIAN

MEDICAL FACILITY Arias Stella

ADDITIONAL RECIPIENT None

MEDICAL FACILITY ID 317319

PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN ID CJC8 10/17/1949

SPECIMEN TYPE Blood

DATE OF COLLECTION 23 February 2021

SPECIMEN RECEIVED 03 March 2021

Biomarker Findings

Blood Tumor Mutational Burden - 1 Muts/Mb

Microsatellite status - MSI-High Not Detected

Tumor Fraction - Cannot Be Determined

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

ATM R337C

TP53 P82L

4 Therapies with Clinical Benefit

10 Clinical Trials

0 Therapies with Lack of Response

PATHOLOGIST COMMENTS

Erik Williams, M.D. 11-Mar-2021

Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis (CH).

(1) This assay is not designed to distinguish germline (inherited) from somatic variants. However, the TP53 P82L variant found in this case has some characteristics suspicious for germline origin.

(2) The ATM R337C variant identified in this case may be secondary to CH rather than due to the patient's reported underlying malignancy.

(3) No somatic mutations definitively attributable to the patient's underlying reported pancreatic cancer were identified in this case, suggesting very low circulating tumor DNA.

Clinical correlation is advised.

BIOMARKER FINDINGS

Blood Tumor Mutational Burden - 1 Muts/Mb

Microsatellite status - MSI-High Not Detected

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. See Biomarker Findings section

MSI-High not detected. No evidence of microsatellite instability in this sample (see Appendix section).

BIOMARKER FINDINGS
Tumor Fraction - Cannot Be Determined

THERAPY AND CLINICAL TRIAL IMPLICATIONS
Tumor fraction is an estimate of the percentage of circulating-tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample based on observed aneuploid instability.
GENOMIC FINDINGS
VAF %
ATM - R337C 0.24%

10 Trials see p. 10

**THERAPIES WITH CLINICAL BENEFIT
(IN PATIENT'S TUMOR TYPE)**

Olaparib

**THERAPIES WITH CLINICAL BENEFIT
(IN OTHER TUMOR TYPE)**

Niraparib

Rucaparib

Talazoparib

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS

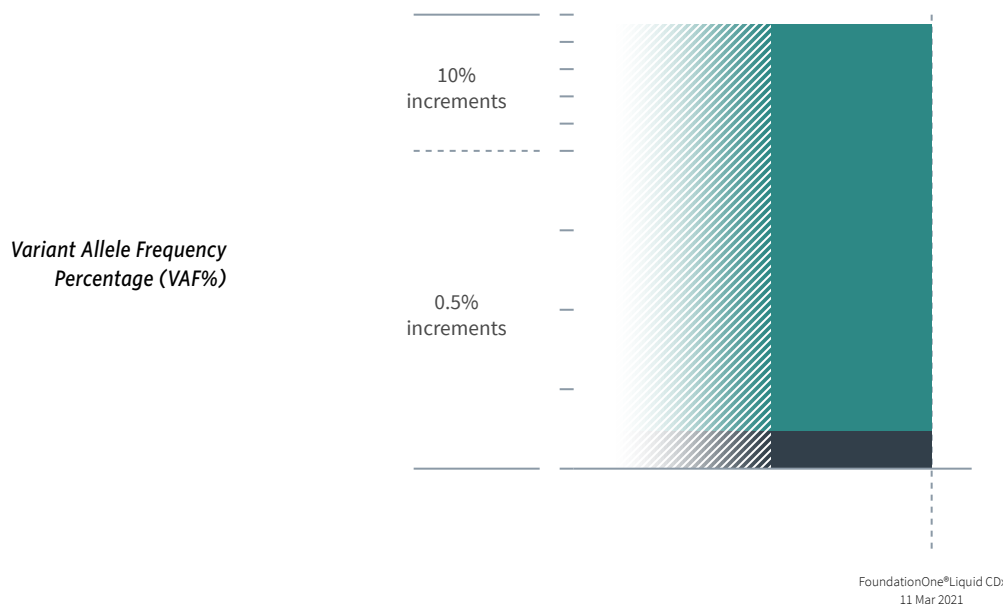
For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

TP53 - P82L **p. 6**

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the therapies listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and/or exhaustive. Neither the therapies nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies. Therapies contained in this report may have been approved by the US FDA or other national authorities; however, they might not have been approved in your respective country. In the appropriate clinical context, germline testing of *APC, BRCA1, BRCA2, BRIP1, MEN1, MLH1, MSH2, MSH6, MUTYH, NF2, PALB2, PMS2, PTEN, RAD51C, RAD51D, RB1, RET, SDHA, SDHB, SDHC, SDHD, SMAD4, STK11, TGFBR2, TP53, TSC1, TSC2, VHL*, and *WT1* is recommended.

Variant Allele Frequency is not applicable for copy number alterations.

ORDERED TEST # ORD-1033694-01



HISTORIC PATIENT FINDINGS		ORD-1033694-01 VAF%
Blood Tumor Mutational Burden		1 Muts/Mb
Microsatellite status		MSI-High Not Detected
Tumor Fraction		Cannot Be Determined
ATM	● R337C	0.24%
TP53	● P82L	48.4%

NOTE This comparison table refers only to genes and biomarkers assayed by prior FoundationOne®Liquid CDx, FoundationOne®Liquid, FoundationOne®, or FoundationOne®CDx tests. Up to five previous tests may be shown.

For some genes in FoundationOne Liquid CDx, only select exons are assayed. Therefore, an alteration found by a previous test may not have been confirmed despite overlapping gene lists. Please refer to the Appendix for the complete list of genes and exons assayed. The gene and biomarker list will be updated periodically to reflect new knowledge about cancer biology.

As new scientific information becomes available, alterations that had previously been listed as Variants of Unknown Significance (VUS) may become reportable.

Tissue Tumor Mutational Burden (TMB) and blood TMB (bTMB) are estimated from the number of synonymous and non-synonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.

Not Tested = not baited, not reported on test, or test preceded addition of biomarker or gene

Not Detected = baited but not detected on test

Detected = present (VAF% is not applicable)

VAF% = variant allele frequency percentage

Cannot Be Determined = Sample is not of sufficient data quality to confidently determine biomarker status

ORDERED TEST # ORD-1033694-01

BIOMARKER FINDINGS

BIOMARKER

Blood Tumor Mutational Burden

RESULT

1 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence in NSCLC and HSNCC, increased bTMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1¹⁻² and anti-PD-1³ therapies. In NSCLC, multiple clinical trials have shown patients with higher bTMB derive clinical benefit from immune checkpoint inhibitors following single agent or combination treatments with either CTLA4 inhibitors or chemotherapy, with reported high bTMB cutpoints ranging from 6 to 16 Muts/Mb¹. In HNSCC, a Phase 3 trial showed

that bTMB ≥ 16 Muts/Mb (approximate equivalency ≥ 8 Muts/Mb as measured by this assay) was associated with improved survival from treatment with a PD-L1 inhibitor alone or in combination with a CTLA-4 inhibitor⁴.

FREQUENCY & PROGNOSIS

Average bTMB levels in solid tumors other than NSCLC have not been evaluated (cBioPortal, COSMIC, PubMed, Mar 2021)⁵⁻⁷. Published data investigating the prognostic implications of bTMB levels in pancreatic carcinoma are limited (PubMed, Jul 2020). A study of patients with pancreatic ductal adenocarcinoma harboring mismatch repair gene mutations reported improved prognosis for patients with high TMB measured in tissue samples (defined as >50 mutations; survival 69-314 months) compared to those with lower TMB (average of 5.7 mutations; 10-42 months)⁸.

FINDING SUMMARY

Blood tumor mutational burden (bTMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations from circulating tumor DNA in blood. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁹⁻¹⁰ and cigarette smoke in lung cancer¹¹⁻¹², treatment with temozolomide-based chemotherapy in glioma¹³⁻¹⁴, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes¹⁵⁻¹⁹, and microsatellite instability (MSI)^{15,18-19}. High bTMB levels were not detected in this sample. It is unclear whether the bTMB levels in this sample would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents¹⁻³. Depending on the clinical context, TMB testing of an alternate sample or by another methodology could be considered.

BIOMARKER

Tumor Fraction

RESULT

Cannot Be Determined

POTENTIAL TREATMENT STRATEGIES

Specimens with high tumor fraction values have high circulating-tumor DNA (ctDNA) content, and thus high sensitivity for identifying genomic alterations. Such specimens are at low risk of false negative results. However, if tumor fraction is not detected as high, it does not exclude the presence of disease burden or compromise the confidence of reported alterations. Tumor fraction levels currently have limited implications for diagnosis, surveillance, or therapy and should not be overinterpreted or compared from one blood draw

to another. There are currently no targeted approaches to address specific tumor fraction levels. In the research setting, changes in tumor fraction estimates have been associated with treatment duration and clinical response and may be a useful indicator for future cancer management²⁰⁻²⁵.

FREQUENCY & PROGNOSIS

Detectable ctDNA levels have been reported in a variety of tumor types, with higher tumor fraction levels reported for patients with metastatic (Stage 4) tumors compared with patients with localized disease (Stages 1 to 3)²⁶. Elevated tumor fraction levels have been reported to be associated with worse prognosis in a variety of cancer types, including pancreatic cancer²⁷, Ewing sarcoma and osteosarcoma²⁸, prostate cancer²³, breast cancer²⁹, leiomyosarcoma³⁰, esophageal cancer³¹, colorectal cancer³², and gastrointestinal cancer³³.

FINDING SUMMARY

Tumor fraction provides an estimate of the percentage of ctDNA present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate for this sample is based on the observed level of aneuploid instability. The tumor fraction algorithm utilized for FoundationOne Liquid CDx uses the allele frequencies of approximately 1,000 single-nucleotide polymorphism (SNP) sites across the genome. Unlike the maximum somatic allele frequency (MSAF) method of estimating ctDNA content³⁴, the tumor fraction metric does not take into account the allele frequency of individual variants but rather produces a more holistic estimate of ctDNA content using data from across the genome. The amount of ctDNA detected may correlate with disease burden and response to therapy³⁵⁻³⁶. However, the tumor fraction estimate in this sample could not be determined with confidence.

ORDERED TEST # ORD-1033694-01

GENOMIC FINDINGS

GENE

ATM

ALTERATION

R337C

TRANSCRIPT ID

NM_000051

CODING SEQUENCE EFFECT

1009C>T

inhibitors. Preclinical experiments also indicate that loss of ATM causes dependency on DNA-PKcs in cancer cells; DNA-PKcs inhibitors promoted apoptosis in ATM-deficient cells and were active in a lymphoma mouse model lacking ATM activity⁵⁹. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

FINDING SUMMARY

ATM encodes the protein ataxia telangiectasia mutated, which is a serine/threonine protein kinase that plays a key role in the DNA damage response⁷². Loss of functional ATM promotes tumorigenesis⁷³. Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL GERMLINE IMPLICATIONS

ATM mutation carriers have increased cancer risk, with female carriers displaying a 38% lifetime risk of breast cancer⁷⁴. Biallelic mutations in ATM underlie the rare autosomal-recessive inherited disorder ataxia-telangiectasia (A-T), also referred to as genome instability or DNA damage response syndrome⁷⁵. This disease is characterized by genomic instability, sensitivity to DNA-damaging agents, and increased risk of developing cancer^{72,75}. The prevalence of A-T is estimated at 1:40,000 to 1:100,000 worldwide⁷⁵. In the appropriate clinical context, germline testing of ATM is recommended.

POTENTIAL TREATMENT STRATEGIES

Loss of functional ATM results in a defective DNA damage response and homologous recombination-mediated DNA repair and may predict sensitivity to PARP inhibitors³⁷. Clinical data in prostate cancer³⁸⁻⁴⁰, gastric cancer⁴¹, colorectal cancer⁴², breast cancer⁴², papillary renal cell carcinoma⁴³, and cholangiocarcinoma⁴⁴ indicate that loss or inactivation of ATM may confer sensitivity to PARP inhibitors⁴⁵⁻⁵². In Phase 1 trials of ATR inhibitors, a heavily pretreated patient with CRC who achieved a CR to berzosertib⁵³ and 4 out of 4 patients with diverse solid tumors who achieved PRs to BAY1895344⁵⁴ harbored ATM inactivation or protein loss; preclinical studies showing reduced cell viability and increased DNA damage in preclinical models of solid tumors⁵⁵⁻⁵⁷ and hematologic malignancies^{55,58} also support the increased sensitivity of ATM-deficient cells to ATR

FREQUENCY & PROGNOSIS

ATM mutations have been reported in 5% of pancreatic cancer cases (COSMIC, Oct 2020)⁷. Expression of the ATM protein has been observed in 67.7% of pancreatic tumors, where it was correlated with p53 expression⁶⁰. Published data investigating the prognostic implications of ATM alterations in pancreatic cancer are limited (PubMed, Oct 2020). Some familial mutations in ATM have been suggested to increase the risk of developing pancreatic cancer⁶¹⁻⁶². One study found that presence of a germline ATM pathogenic variant was significantly associated with frequency of pancreatic ductal adenocarcinoma (HR=7.73)⁶³. Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion⁶⁴⁻⁶⁹. Comprehensive genomic profiling of solid tumors may detect nontumor alterations that are due to CH^{68,70-71}. Patient-

ORDERED TEST # ORD-1033694-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

P82L

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

245C>T

POTENTIAL TREATMENT STRATEGIES

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib⁷⁶⁻⁷⁹, or p53 gene therapy and immunotherapeutics such as SGT-53⁸⁰⁻⁸⁴ and ALT-801⁸⁵. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% (17/176) and SDs in 53.4% (94/176) of patients with solid tumors; the response rate was 21.1% (4/19) in patients with TP53 mutations versus 12.1% (4/33) in patients who were TP53 wild-type⁸⁶. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 31.9% (30/94, 3 CR) ORR and a 73.4% (69/94) DCR in patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer⁸⁷. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 42.9% (9/21, 1 CR) ORR and a 76.2% (16/21) DCR in patients with platinum-refractory TP53-mutated ovarian cancer⁸⁸. The combination of adavosertib with paclitaxel and carboplatin in patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone⁸⁹. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24.0% (6/25) ORR with adavosertib combined with paclitaxel⁹⁰. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell

carcinoma (HNSCC) elicited a 71.4% (5/7) response rate for patients with TP53 alterations⁹¹. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75.0% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage⁸⁴. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53-mutated, but not TP53-wild-type, breast cancer xenotransplant mouse model⁹². Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutant p53 such as APR-246⁹³⁻⁹⁵. In a Phase 1b trial in patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR⁹⁶. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies^{58,97}; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies⁹⁸⁻⁹⁹. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition. It is not known whether these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

FREQUENCY & PROGNOSIS

TP53 mutations have been reported in 33-75% of pancreatic carcinomas, with the majority occurring as missense mutations, while deletion of TP53 has been found in 66% of pancreatic ductal adenocarcinoma cases¹⁰⁰⁻¹⁰³. TP53 mutations are common in pancreatic ductal adenocarcinomas and are known to occur in the process of pancreatic carcinogenesis¹⁰⁴⁻¹⁰⁵. Additionally, aberrant expression of p53 has been found in 54-81% of pancreatic ductal adenocarcinoma cases^{102,106-108}. Studies have found inconsistent results regarding the prognostic significance of

p53 expression in pancreatic ductal adenocarcinoma, although one study correlated low levels of TP53 mRNA with poor patient prognosis^{106,109-110}. Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion⁶⁴⁻⁶⁹. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy⁶⁴⁻⁶⁵. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease¹¹¹. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{68,70-71}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers¹¹². Although alterations such as seen here have not been fully characterized and are of unknown functional significance, similar alterations have been previously reported in the context of cancer, which may indicate biological relevance.

POTENTIAL GERMLINE IMPLICATIONS

Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers¹¹³⁻¹¹⁵, including sarcomas¹¹⁶⁻¹¹⁷. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000¹¹⁸ to 1:20,000¹¹⁷. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30¹¹⁹. In the appropriate clinical context, germline testing of TP53 is recommended.

ORDERED TEST # ORD-1033694-01

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Olaparib

Assay findings association
ATM
R337C

AREAS OF THERAPEUTIC USE

The PARP inhibitor olaparib is FDA approved to treat patients with epithelial ovarian, Fallopian tube, or primary peritoneal cancer, patients with deleterious or suspected deleterious gBRCA-mutated pancreatic adenocarcinoma or HER2-negative breast cancer, and patients with prostate cancer and mutations in homologous recombination repair genes. Olaparib is also approved in combination with bevacizumab to treat patients with ovarian, Fallopian tube, or primary peritoneal cancer with deleterious or suspected deleterious somatic or gBRCA mutation and/or genomic instability. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{38-40,120}, colorectal cancer⁴², breast cancer⁴², gastric cancer⁴¹, cholangiocarcinoma⁴⁴, and papillary renal cell carcinoma⁴³. It is not known whether this therapeutic approach would be relevant in

the context of alterations that have not been fully characterized, as seen here.

SUPPORTING DATA

The Phase 3 randomized, placebo-controlled POLO trial investigating maintenance olaparib for patients with platinum-sensitive, germline BRCA1/2-mutated metastatic pancreatic adenocarcinoma reported a significantly longer median PFS compared with placebo (7.4 vs. 3.8 months, HR=0.53); at 2-year follow-up, 22.1% of olaparib-treated patients were progression free, with 2 ongoing CRs reported, compared with 9.6% of patients receiving placebo; however, olaparib maintenance therapy did not impact health-related quality of life or OS (18.9 vs. 18.1 months, HR=0.91) relative to placebo¹²¹⁻¹²². A Phase 2 trial of olaparib monotherapy for patients with germline BRCA1/2-mutated recurrent pancreatic cancer reported a response rate of 21.7%¹²³. Parallel Phase 2 trials reported 2 PRs for patients with platinum-sensitive, DNA damage repair (DDR) deficient, germline BRCA mutation-negative pancreatic ductal adenocarcinoma; no responses were observed in platinum-refractory cases¹²⁴.

ORDERED TEST # ORD-1033694-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Niraparib

Assay findings association

ATM
R337C

AREAS OF THERAPEUTIC USE

The PARP inhibitor niraparib is FDA approved to treat patients with epithelial ovarian, fallopian tube, or primary peritoneal cancer, with or without homologous recombination deficiency (HRD)-positive status. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{38-40,120}, colorectal cancer⁴², breast cancer⁴², gastric cancer⁴¹, cholangiocarcinoma⁴⁴, and papillary renal cell carcinoma⁴³. It is not known whether this therapeutic approach would be relevant in the context of alterations that have not been fully characterized, as seen here.

SUPPORTING DATA

Clinical data on the efficacy of niraparib for the treatment of pancreatic cancer are limited (PubMed, Jan 2021).

Niraparib has been primarily evaluated in the context of ovarian cancer. In a Phase 3 study of patients with platinum-sensitive, recurrent ovarian cancer, niraparib significantly increased median PFS, as compared to placebo, in patients with germline BRCA mutations (21 vs. 5.5 months) and in patients without germline BRCA mutations (9.3 vs. 3.9 months) as well as in a subgroup of the patients without germline BRCA mutations with homologous recombination-deficient tumors (12.9 vs. 3.8 months)¹²⁵. In a Phase 1 study of niraparib treatment for patients with solid tumors, 40% (8/20) of patients with ovarian cancer and BRCA mutations and 50% (2/4) of patients with breast cancer and BRCA mutations experienced a PR, and 43% (9/21) of patients with castration-resistant prostate cancer and 100% (2/2) of patients with non-small cell lung cancer achieved SD¹²⁶. A Phase 1 study of the combination of niraparib and bevacizumab in patients with platinum-sensitive, high-grade ovarian cancer reported a DCR of 91% (10/11), with a response rate of 45% (5/11)¹²⁷.

Rucaparib

Assay findings association

ATM
R337C

AREAS OF THERAPEUTIC USE

The PARP inhibitor rucaparib is FDA approved to treat patients with metastatic castration-resistant prostate cancer (mCRPC) or epithelial ovarian, Fallopian tube, or primary peritoneal cancer and deleterious somatic or germline BRCA mutations. Rucaparib is also approved as a maintenance treatment of patients with recurrent epithelial ovarian, Fallopian tube, or primary peritoneal cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{38-40,120}, colorectal cancer⁴²,

breast cancer⁴², gastric cancer⁴¹, cholangiocarcinoma⁴⁴, and papillary renal cell carcinoma⁴³. It is not known whether this therapeutic approach would be relevant in the context of alterations that have not been fully characterized, as seen here.

SUPPORTING DATA

In the RUCAPANC Phase 2 study, rucaparib elicited an ORR of 15.8% and a DCR of 31.6% in BRCA1/2-mutated advanced or metastatic pancreatic cancer, with 1 CR and 2 PRs confirmed and 1 additional unconfirmed CR out of 19 treated patients¹²⁸. In a Phase 2 study, rucaparib monotherapy elicited 1 CR, 6 PRs, an ORR of 36.8%, and a DCR (CR+PR+SD) of 89.5% in a cohort of 19 patients with advanced pancreatic carcinoma harboring germline or somatic mutations in BRCA or PALB2¹²⁹.

ORDERED TEST # ORD-1033694-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Talazoparib

Assay findings association

ATM
R337C

AREAS OF THERAPEUTIC USE

The PARP inhibitor talazoparib is FDA approved to treat HER2-negative locally advanced or metastatic breast cancer with deleterious or suspected deleterious germline BRCA mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

ATM inactivation may predict sensitivity to PARP inhibitors. ATM alterations have been associated with clinical benefit from PARP inhibitors in solid tumors, including prostate cancer^{38-40,120}, colorectal cancer⁴², breast cancer⁴², gastric cancer⁴¹, cholangiocarcinoma⁴⁴, and papillary renal cell carcinoma⁴³. It is not known whether this therapeutic approach would be relevant in the context of alterations that have not been fully characterized, as seen here.

SUPPORTING DATA

A Phase 1 study of talazoparib reported 2 PRs for patients

with pancreatic cancer and a BRCA2 or PALB2 mutation¹³⁰. Talazoparib has been studied primarily in the context of BRCA-mutated, HER2-negative breast cancer, where patients achieved significantly longer median PFS (8.6 vs. 5.6 months, HR=0.54), a higher ORR (62.6% vs. 27.2%), and improved quality of life on talazoparib compared with standard chemotherapy in a Phase 3 study¹³¹⁻¹³². In a Phase 2 study of talazoparib for BRCA1/2-wildtype patients with homologous recombination pathway alterations, the best outcome in non-breast tumors was SD ≥ 6 months for 2/7 patients who had colon cancer with germline ATM alteration or testicular cancer with germline CHEK2 and somatic ATM alteration⁴². Clinical activity of single-agent talazoparib has been observed in numerous other solid tumors, including responses in BRCA-mutated ovarian, pancreatic, prostate, and ampulla of Vater cancers; PALB2-mutated pancreatic and bladder cancers; ATM-mutated cholangiocarcinoma; and small cell lung cancer^{130,133-135}.

NOTE Genomic alterations detected may be associated with activity of certain US FDA or other specific country approved therapies; however, the therapies listed in this report may have varied evidence in the patient's tumor type. The listed therapies are not ranked in order of potential or predicted efficacy for this patient or in order of level of evidence for this patient's tumor type. The therapies listed in this report may not be complete and/or exhaustive. Furthermore, the listed therapies are limited to US FDA approved pharmaceutical drug products that are linked to a specific genomic alteration. There may also be US FDA approved pharmaceutical drug products that are not linked to a genomic alteration. Further there may also exist pharmaceutical drug products that are not approved by the US FDA or other national authorities. There may also be other treatment modalities available than pharmaceutical drug products.

ORDERED TEST # ORD-1033694-01

CLINICAL TRIALS

IMPORTANT Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or

research staff. This is not a comprehensive list of all available clinical trials. There may also be compassionate use or early access programs available, which are not listed in this report. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials are not ranked in order of potential or predicted efficacy for this patient or

in order of level of evidence for this patient's tumor type. Clinical trials listed here may have additional enrollment criteria that may require medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see clinicaltrials.gov. However, clinicaltrials.gov does not list all clinical trials that might be available.

GENE ATM

ALTERATION R337C

RATIONALE
Loss or inactivation of ATM may increase sensitivity to PARP inhibitors, ATR inhibitors, or DNA-PKcs inhibitors. It is not known whether

these therapeutic approaches would be relevant in the context of alterations that have not been fully characterized, as seen here.

NCT03742895

PHASE 2

Efficacy and Safety of Olaparib (MK-7339) in Participants With Previously Treated, Homologous Recombination Repair Mutation (HRRm) or Homologous Recombination Deficiency (HRD) Positive Advanced Cancer (MK-7339-002 / LYNK-002)

TARGETS
PARP

LOCATIONS: Lima (Peru), Trujillo (Peru), Cali (Colombia), Bogota (Colombia), Medellin (Colombia), Monteria (Colombia), Valledupar (Colombia), Buenos Aires (Argentina), Ciudad de Buenos Aires (Argentina), Berazategui (Argentina)

NCT04123366

PHASE 2

Study of Olaparib (MK-7339) in Combination With Pembrolizumab (MK-3475) in the Treatment of Homologous Recombination Repair Mutation (HRRm) and/or Homologous Recombination Deficiency (HRD)-Positive Advanced Cancer (MK-7339-007/KEYLYNK-007)

TARGETS
PARP, PD-1

LOCATIONS: Lima (Peru), Bellavista (Peru), Cuzco (Peru), Cali (Colombia), Medellin (Colombia), Bucaramanga (Colombia), Barranquilla (Colombia), Buenos Aires (Argentina), Ciudad de Buenos Aires (Argentina), Guatemala (Guatemala)

NCT02693535

PHASE 2

TAPUR: Testing the Use of Food and Drug Administration (FDA) Approved Drugs That Target a Specific Abnormality in a Tumor Gene in People With Advanced Stage Cancer

TARGETS
VEGFRs, ABL, SRC, ALK, AXL, MET, ROS1, TRKA, TRKC, CDK4, CDK6, CSF1R, FLT3, KIT, RET, mTOR, EGFR, ERBB2, ERBB3, MEK, BRAF, SMO, DDR2, PARP, PD-1, CTLA-4, ERBB4

LOCATIONS: Florida, Texas, Georgia, Alabama, North Carolina, Virginia, Oklahoma, Pennsylvania, Indiana

NCT02498613

PHASE 2

A Phase 2 Study of Cediranib in Combination With Olaparib in Advanced Solid Tumors

TARGETS
PARP, VEGFRs

LOCATIONS: Florida, Texas, Tennessee, Virginia, Connecticut, Massachusetts, Toronto (Canada), California

ORDERED TEST # ORD-1033694-01

CLINICAL TRIALS
NCT03188965
PHASE 1

First-in-human Study of ATR Inhibitor BAY1895344 in Patients With Advanced Solid Tumors and Lymphomas

TARGETS
ATR

LOCATIONS: Florida, Texas, Georgia, Virginia, Pennsylvania, New York, Ohio, Massachusetts

NCT02595931
PHASE 1

ATR Kinase Inhibitor VX-970 and Irinotecan Hydrochloride in Treating Patients With Solid Tumors That Are Metastatic or Cannot Be Removed by Surgery

TARGETS
ATR

LOCATIONS: Florida, North Carolina, Tennessee, Missouri, Pennsylvania, Connecticut, Massachusetts, California

NCT03907969
PHASE 1/2

A Clinical Trial to Evaluate AZD7648 Alone and in Combination With Other Anti-cancer Agents in Patients With Advanced Cancers

TARGETS
PARP, DNA-PK

LOCATIONS: Texas, Connecticut, Newcastle upon Tyne (United Kingdom)

NCT03337087
PHASE 1/2

Liposomal Irinotecan, Fluorouracil, Leucovorin Calcium, and Rucaparib in Treating Patients With Metastatic Pancreatic, Colorectal, Gastroesophageal, or Biliary Cancer

TARGETS
PARP, TOP1

LOCATIONS: Georgia, Arizona, Minnesota

NCT02769962
PHASE 1/2

Trial of CRLX101, a Nanoparticle Camptothecin With Olaparib in People With Relapsed/Refractory Small Cell Lung Cancer

TARGETS
PARP, TOP1

LOCATIONS: Maryland

NCT02487095
PHASE 1/2

Trial of Topotecan With VX-970, an ATR Kinase Inhibitor, in Small Cell Cancers

TARGETS
ATR

LOCATIONS: Maryland

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APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

CDH1
A298T

CHEK2
G229D

KDR
E207G

LTK
R647W

MET
V313L

MLH1
R9W

PDCD1LG2 (PD-L2)
E11fs*7

SMO
G767R

SPEN
S2306del and S2426F

TSC1
K587R

APPENDIX
Genes assayed in FoundationOne®Liquid CDx

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FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

ABL1 Exons 4-9	ACVR1B	AKT1 Exon 3	AKT2	AKT3	ALK Exons 20-29, Introns 18, 19	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF Exons 4, 5, 7, 11, 13, 15, 16	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BCR* Introns 8, 13, 14	BRAF Exons 11-18, Introns 7-10	BRCA1 Introns 2, 7, 8, 12, 16, 19, 20	BRCA2 Intron 2	BRD4	BRIP1	BTG1
BTG2	BTK Exons 2, 15	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL
CCND1	CCND2	CCND3	CCNE1	CD22	CD70	CD74* Introns 6-8	CD79A	CD79B
CD274 (PD-L1)	CDC73	CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B
CDKN2A	CDKN2B	CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL
CSF1R	CSF3R	CTCF	CTNNA1	CTNNB1 Exon 3	CUL3	CUL4A	CXCR4	CYP17A1
DAXX	DDR1	DDR2 Exons 5, 17, 18	DIS3	DNMT3A	DOT1L	EED	EGFR Introns 7, 15, 24-27	EP300
EPHA3	EPHB1	EPHB4	ERBB2	ERBB3 Exons 3, 6, 7, 8, 10, 12, 20, 21, 23, 24, 25	ERBB4	ERCC4	ERG	ERRF1
ESR1 Exons 4-8	ETV4* Intron 8	ETV5* Introns 6, 7	ETV6* Introns 5, 6	EWSR1* Introns 7-13	EZH2 Exons 4, 16, 17, 18	EZR* Introns 9-11	FAM46C	FANCA
FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12	FGF14	FGF19
FGF23	FGF3	FGF4	FGF6	FGFR1 Introns 1, 5, Intron 17	FGFR2 Intron 1, Intron 17	FGFR3 Exons 7, 9 (alternative designation exon 10), 14, 18, Intron 17	FGFR4	FH
FLCN	FLT1	FLT3 Exons 14, 15, 20	FOXL2	FUBP1	GABRA6	GATA3	GATA4	GATA6
GNA11 Exons 4, 5	GNA13	GNAQ Exons 4, 5	GNAS Exons 1, 8	GRM3	GSK3B	H3F3A	HDAC1	HGF
HNFI1A	HRAS Exons 2, 3	HSD3B1	ID3	IDH1 Exon 4	IDH2 Exon 4	IGF1R	IKBKE	IKZF1
INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2 Exon 14	JAK3 Exons 5, 11, 12, 13, 15, 16	JUN	KDMSA
KDMSC	KDM6A	KDR	KEAP1	KEL	KIT Exons 8, 9, 11, 12, 13, 17, Intron 16	KLHL6	KMT2A (MLL) Introns 6, 8-11, Intron 7	KMT2D (MLL2)

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Electronically signed by Erik Williams, M.D. | 11 March 2021
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Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

APPENDIX
Genes assayed in FoundationOne®Liquid CDx

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KRAS	LTK	LYN	MAF	MAP2K1 (MEK1) Exons 2, 3	MAP2K2 (MEK2) Exons 2-4, 6, 7	MAP2K4	MAP3K1	MAP3K13
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APPENDIX
Genes assayed in FoundationOne®Liquid CDx

ORDERED TEST # ORD-1033694-01

FoundationOne Liquid CDx interrogates 324 genes, including 309 genes with complete exonic (coding) coverage and 15 genes with only select non-coding coverage (indicated with an *); 75 genes (indicated in bold) are captured with increased sensitivity and have complete exonic (coding) coverage unless otherwise noted.

MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1	MERTK	MET
MITF	MKNK1	MLH1	MPL Exon 10	MRE11A	MSH2 Intron 5	MSH3	MSH6	MST1R
MTAP	MTOR Exons 19, 30, 39, 40, 43-45, 47, 48, 53, 56	MUTYH	MYB* Intron 14	MYC Intron 1	MYCL (MYCL1)	MYCN	MYD88 Exon 4	NBN
NF1	NF2	NFE2L2	NFKBIA	NKX2-1	NOTCH1	NOTCH2 Intron 26	NOTCH3	NPM1 Exons 4-6, 8, 10
NRAS Exons 2, 3	NSD3 (WHSC1L1)	NTSC2	NTRK1 Exons 14, 15, Introns 8-11	NTRK2 Intron 12	NTRK3 Exons 16, 17	NUTM1* Intron 1	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA Exons 12, 18, Introns 7, 9, 11
PDGFRB Exons 12-21, 23	PDK1	PIK3C2B	PIK3C2G	PIK3CA Exons 2, 3, 5-8, 10, 14, 19, 21 (Coding Exons 1, 2, 4-7, 9, 13, 18, 20)	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1 Exons 3, 4, 6, 7, 10, 14, 15, 17, Introns 4-8	RARA Intron 2	RB1	RBM10	REL	RET Introns 7, 8, Exons 11, 13-16, Introns 9-11
RICTOR	RNF43	ROS1 Exons 31, 36-38, 40, Introns 31-35	RPTOR	RSPO2* Intron 1	SDC4* Intron 2	SDHA	SDHB	SDHC
SDHD	SETD2	SF3B1	SGK1	SLC34A2* Intron 4	SMAD2	SMAD4	SMARCA4	SMARCB1
SMO	SNCAIP	SOC31	SOX2	SOX9	SPEN	SPOP	SRC	STAG2
STAT3	STK11	SUFU	SYK	TBX3	TEK	TERC* ncRNA	TERT* Promoter	TET2
TGFBR2	TIPARP	TMPPRSS2* Introns 1-3	TNFAIP3	TNFRSF14	TP53	TSC1	TSC2	TYRO3
U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1	XRCC2	ZNF217	ZNF703

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Microsatellite (MS) status

Blood Tumor Mutational Burden (bTMB)

Tumor Fraction

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APPENDIX

About FoundationOne®Liquid CDx

FoundationOne Liquid CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium. The CE-IVD regulatory status of FoundationOne Liquid CDx is applicable in countries that accept and/or recognize the CE mark.



ABOUT FOUNDATIONONE LIQUID CDx

FoundationOne Liquid CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne Liquid CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details.

INTENDED USE

FoundationOne Liquid CDx is a next generation sequencing based *in vitro* diagnostic device that analyzes 324 genes. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The test also detects the genomic signatures blood tumor mutational burden (bTMB), microsatellite instability (MSI), and tumor fraction. FoundationOne Liquid CDx utilizes circulating cell-free DNA (cfDNA) isolated from plasma derived from the anti-coagulated peripheral whole blood of cancer patients. The test is intended to be used as a companion diagnostic to identify patients who may benefit from treatment with targeted therapies in accordance with the approved therapeutic product labeling. Additionally, FoundationOne Liquid CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with malignant neoplasms.

TEST PRINCIPLES

The FoundationOne Liquid CDx assay is performed exclusively as a laboratory service using circulating cell-free DNA (cfDNA) isolated from plasma derived from anti-coagulated peripheral whole blood from patients with solid malignant neoplasms. The assay employs a single DNA extraction method to obtain cfDNA from plasma from whole blood. Extracted

cfDNA undergoes whole-genome shotgun library construction and hybridization-based capture of 324 cancer-related genes including coding exons and select introns of 309 genes, as well as only select intronic regions or non-coding regions of 15 genes. Hybrid-capture selected libraries are sequenced with deep coverage using the NovaSeq® 6000 platform. Sequence data are processed using a customized analysis pipeline designed to accurately detect genomic alterations, including base substitutions, indels, select copy number variants, and select genomic rearrangements. Substitutions and insertion and deletion alterations (indels) are reported in 311 genes, copy number alterations (CNAs) are reported in 310 genes, and gene rearrangements are reported in 324 genes. The assay also detects select genomic rearrangements, select copy number alterations, tumor fraction, and genomic signatures including MSI and bTMB. A subset of targeted regions in 75 genes is baited for increased sensitivity.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. *Note:* A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

QUALIFIED ALTERATION CALLS (EQUIVOCAL)

All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls.

RANKING OF ALTERATIONS AND THERAPIES

Biomarker and Genomic Findings

Therapies are ranked based on the following criteria: Therapies with clinical benefit in patient's tumor type (ranked alphabetically within each NCCN category) followed by therapies with clinical benefit in other tumor type (ranked alphabetically within each NCCN category).

Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

LIMITATIONS

1. For *in vitro* diagnostic use.
2. For prescription use only. This test must be ordered by a qualified medical professional in accordance with clinical laboratory regulations.
3. A negative result does not rule out the presence of a mutation below the limits of detection of the assay. Patients for whom no companion diagnostic alterations are detected should be considered for confirmation with an appropriately validated tumor tissue test, if available.
4. The FoundationOne Liquid CDx assay does not detect heterozygous deletions.
5. The test is not intended to provide information on cancer predisposition.
6. Performance has not been validated for cfDNA input below the specified minimum input.
7. Tissue TMB and blood TMB (bTMB) are estimated from the number of synonymous and nonsynonymous single-nucleotide variants (SNVs) and insertions and deletions (indels) per area of coding genome sampled, after the removal of known and likely oncogenic driver events and germline SNPs. Tissue TMB is calculated based on variants with an allele frequency of $\geq 5\%$, and bTMB is calculated based on variants with an allele frequency of $\geq 0.5\%$.
8. Tumor fraction is the percentage of circulating-tumor DNA (ctDNA) present in a cell-free DNA (cfDNA) sample. The tumor fraction estimate is computationally derived from observed aneuploid instability in the sample.
9. Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the tumor genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor. The MSI algorithm is based on genome wide analysis of 1765 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines for solid tissue testing.
10. Genomic findings from circulating cell-free DNA (cfDNA) may originate from circulating tumor DNA fragments, germline alterations, or non-tumor somatic alterations, such as clonal hematopoiesis of indeterminate potential (CHIP). Genes with alterations that may be derived from CHIP include, but are not limited

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APPENDIX

About FoundationOne®Liquid CDx

to: *ASXL1*, *ATM*, *CBL*, *CHEK2*, *DNMT3A*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, *TP53*, and *U2AF1*.

11. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. If a reported alteration is suspected to be germline, confirmatory testing should be considered in the appropriate clinical context.
12. The test is not intended to replace germline testing or to provide information about cancer predisposition.

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters with no conflicts), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >30%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories please refer to the NCCN Compendium®.

Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2020. All rights reserved. To view the most recent and complete version of the guideline, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne Liquid CDx.

TREATMENT DECISIONS ARE THE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this test or the information contained in this report.

Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or

repetitive/high homology sequences. FoundationOne Liquid CDx is performed using cell-free DNA, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
Muts/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 3.0.0

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1. Gandara DR, et al. Nat. Med. (2018) pmid: 30082870
2. Wang Z, et al. JAMA Oncol (2019) pmid: 30816954
3. Aggarwal C, et al. Clin. Cancer Res. (2020) pmid: 32102950
4. Li et al., 2020; ASCO Abstract 6511
5. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
6. Gao J, et al. Sci Signal (2013) pmid: 23550210
7. Tate JG, et al. Nucleic Acids Res. (2019) pmid: 30371878
8. Hu et al., 2017; ASCO Abstract e15791
9. Pfeifer GP, et al. Mutat. Res. (2005) pmid: 15748635
10. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) pmid: 23875803
11. Pfeifer GP, et al. Oncogene (2002) pmid: 12379884
12. Rizvi NA, et al. Science (2015) pmid: 25765070
13. Johnson BE, et al. Science (2014) pmid: 24336570
14. Choi S, et al. Neuro-oncology (2018) pmid: 29452419
15. Cancer Genome Atlas Research Network, et al. Nature (2013) pmid: 23636398
16. Briggs S, et al. J. Pathol. (2013) pmid: 23447401
17. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) pmid: 24583393
18. Nature (2012) pmid: 22810696
19. Roberts SA, et al. Nat. Rev. Cancer (2014) pmid: 25568919
20. Bronkhorst AJ, et al. Biomol Detect Quantif (2019) pmid: 30923679
21. Raja R, et al. Clin. Cancer Res. (2018) pmid: 30093454
22. Hrebien S, et al. Ann. Oncol. (2019) pmid: 30860573
23. Choudhury AD, et al. JCI Insight (2018) pmid: 30385733
24. Goodall J, et al. Cancer Discov (2017) pmid: 28450425
25. Goldberg SB, et al. Clin. Cancer Res. (2018) pmid: 29330207
26. Bettgowda C, et al. Sci Transl Med (2014) pmid: 24553385
27. Lapin M, et al. J Transl Med (2018) pmid: 30400802
28. Shulman DS, et al. Br. J. Cancer (2018) pmid: 30131550
29. Stover DG, et al. J. Clin. Oncol. (2018) pmid: 29298117
30. Hemming ML, et al. JCO Precis Oncol (2019) pmid: 30793095
31. Egyud M, et al. Ann. Thorac. Surg. (2019) pmid: 31059681
32. Fan G, et al. PLoS ONE (2017) pmid: 28187169
33. Vu et al., 2020; DOI: 10.1200/PO.19.00204
34. Li G, et al. J Gastrointest Oncol (2019) pmid: 31602320
35. Zhang EW, et al. Cancer (2020) pmid: 32757294
36. Butler TM, et al. Cold Spring Harb Mol Case Stud (2019) pmid: 30833418
37. Michels J, et al. Oncogene (2014) pmid: 24037533
38. Mateo J, et al. N. Engl. J. Med. (2015) pmid: 26510020
39. Mateo J, et al. Lancet Oncol. (2019) pmid: 31806540
40. Abida W, et al. Clin. Cancer Res. (2020) pmid: 32086346
41. Bang YJ, et al. J. Clin. Oncol. (2015) pmid: 26282658
42. Gruber et al., 2019; ASCO Abstract 3006
43. Olson D, et al. Clin Genitourin Cancer (2016) pmid: 27079472
44. Piha-Paul et al., 2018; AACR-NCI-EORTC Abstract A096
45. Weston VJ, et al. Blood (2010) pmid: 20739657
46. Williamson CT, et al. Mol. Cancer Ther. (2010) pmid: 20124459
47. Gilardini Montani MS, et al. J. Exp. Clin. Cancer Res. (2013) pmid: 24252502
48. Bryant HE, et al. Nucleic Acids Res. (2006) pmid: 16556909
49. Ihnen M, et al. Mol. Cancer Ther. (2013) pmid: 23729402
50. Williamson CT, et al. EMBO Mol Med (2012) pmid: 22416035
51. Kubota E, et al. Cell Cycle (2014) pmid: 24841718
52. Huehls AM, et al. Mol. Pharmacol. (2012) pmid: 22833573
53. O'Carrigan et al., 2016; ASCO Abstract 2504
54. De Bono et al., 2019; ASCO Abstract 3007
55. Menezes DL, et al. Mol. Cancer Res. (2015) pmid: 25232030
56. Vendetti FP, et al. Oncotarget (2015) pmid: 26517239
57. Min A, et al. Mol. Cancer Ther. (2017) pmid: 28138034
58. Kwok M, et al. Blood (2016) pmid: 26563132
59. Riabinska A, et al. Sci Transl Med (2013) pmid: 23761041
60. Yu G, et al. Pancreas (2004) pmid: 15097860
61. Roberts NJ, et al. Cancer Discov (2012) pmid: 22585167
62. Geoffroy-Perez B, et al. Int. J. Cancer (2001) pmid: 11410879
63. Cremin C, et al. Cancer Med (2020) pmid: 32255556
64. Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
65. Genovese G, et al. N. Engl. J. Med. (2014) pmid: 25426838
66. Xie M, et al. Nat. Med. (2014) pmid: 25326804
67. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
68. Severson EA, et al. Blood (2018) pmid: 29678827
69. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
70. Chabon JJ, et al. Nature (2020) pmid: 32269342
71. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
72. Shiloh Y, et al. Nat. Rev. Mol. Cell Biol. (2013) pmid: 23847781
73. Cremona CA, et al. Oncogene (2014) pmid: 23851492
74. van Os NJ, et al. Clin Genet (2016) pmid: 26662178
75. Rothblum-Oviatt C, et al. Orphanet J Rare Dis (2016) pmid: 27884168
76. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315
77. Bridges KA, et al. Clin. Cancer Res. (2011) pmid: 21799033
78. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid: 21389100
79. Osman AA, et al. Mol. Cancer Ther. (2015) pmid: 25504633
80. Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
81. Xu L, et al. Mol. Med. (2001) pmid: 11713371
82. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
83. Kim SS, et al. Nanomedicine (2015) pmid: 25240597
84. Pirolo KF, et al. Mol. Ther. (2016) pmid: 27357628
85. Hajdenberg et al., 2012; ASCO Abstract e15010
86. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
87. Moore et al., 2019; ASCO Abstract 5513
88. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27998224
89. Oza et al., 2015; ASCO Abstract 5506
90. Lee J, et al. Cancer Discov (2019) pmid: 31315834
91. Méndez E, et al. Clin. Cancer Res. (2018) pmid: 29535125
92. Ma CX, et al. J. Clin. Invest. (2012) pmid: 22446188
93. Lehmann S, et al. J. Clin. Oncol. (2012) pmid: 22965953
94. Mohell N, et al. Cell Death Dis (2015) pmid: 26086967
95. Franssón A, et al. J Ovarian Res (2016) pmid: 27179933
96. Gourley et al., 2016; ASCO Abstract 5571
97. Boudny M, et al. Haematologica (2019) pmid: 30975914
98. Dillon MT, et al. Mol. Cancer Ther. (2017) pmid: 28062704
99. Middleton FK, et al. Cancers (Basel) (2018) pmid: 30127241
100. Biankin AV, et al. Nature (2012) pmid: 23103869
101. Morton JP, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) pmid: 20018721
102. Scarpa A, et al. Am. J. Pathol. (1993) pmid: 8494051
103. Luo Y, et al. Pathol. Oncol. Res. (2013) pmid: 22782330
104. Iacobuzio-Donahue CA, et al. Clin. Cancer Res. (2012) pmid: 22896692
105. Macgregor-Das AM, et al. J Surg Oncol (2013) pmid: 22806689
106. Oshima M, et al. Ann. Surg. (2013) pmid: 23470568
107. Ottenhof NA, et al. Cell Oncol (Dordr) (2012) pmid: 22351431
108. Tsiambas E, et al. J BUON () pmid: 20414934
109. Ansari D, et al. Br J Surg (2011) pmid: 21644238
110. Grochola LF, et al. Pancreas (2011) pmid: 21404460
111. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
112. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
113. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
114. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
115. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
116. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
117. Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
118. Laloo F, et al. Lancet (2003) pmid: 12672316
119. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713
120. de Bono et al., 2020; ASCO GU Abstract 119
121. Golan T, et al. N. Engl. J. Med. (2019) pmid: 31157963
122. Oh et al., 2019; ASCO Abstract 1260
123. Kaufman B, et al. J. Clin. Oncol. (2015) pmid: 25366685
124. Golan et al., 2018; ASCO Abstract 297
125. Mirza MR, et al. N. Engl. J. Med. (2016) pmid: 27717299
126. Sandhu SK, et al. Lancet Oncol. (2013) pmid: 23810788
127. Mirza et al., 2016; ASCO Abstract 5555
128. Shroff RT, et al. JCO Precis Oncol (2018) pmid: 30051098
129. Binder et al., 2019; AACR Abstract CT234
130. de Bono J, et al. Cancer Discov (2017) pmid: 28242752
131. Litton JK, et al. N. Engl. J. Med. (2018) pmid: 30110579
132. Ettl J, et al. Ann. Oncol. (2018) pmid: 30124753
133. Lu E, et al. J Natl Compr Canc Netw (2018) pmid: 30099369
134. Piha-Paul et al., 2017; EORTC-NCI-AACR Abstract A096
135. Meehan et al., 2017; AACR Abstract 4687