

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Breast carcinoma (NOS)

DATE OF BIRTH 17 December 1957

SEX Female

MEDICAL RECORD # Not given

PHYSICIAN

MEDICAL FACILITY Arias Stella

ADDITIONAL RECIPIENT None

MEDICAL FACILITY ID 317319

PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Breast

SPECIMEN ID BP21 00246 -1 A

SPECIMEN TYPE Block

DATE OF COLLECTION 25 March 2021

SPECIMEN RECEIVED 03 June 2021

Biomarker Findings

Microsatellite status - MS-Stable

Tumor Mutational Burden - 6 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

PIK3CA H1047R

MYC amplification - equivocal[†]

PTEN Y174fs*15

NSD3 (WHSC1L1) amplification - equivocal[†]

SETD2 R1694fs*17

TET2 H266fs*24

TP53 R209fs*6

ZNF703 amplification - equivocal[†]

3 Disease relevant genes with no reportable alterations: *BRCA1*, *BRCA2*, *ERBB2*

[†] See About the Test in appendix for details.

3 Therapies with Clinical Benefit

22 Clinical Trials

0 Therapies with Lack of Response

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 6 Muts/Mb

GENOMIC FINDINGS

PIK3CA - H1047R

10 Trials *see p. 15*

MYC - amplification - equivocal

7 Trials *see p. 13*

PTEN - Y174fs*15

10 Trials *see p. 17*

ACTIONABILITY

No therapies or clinical trials. *see Biomarker Findings section*

No therapies or clinical trials. *see Biomarker Findings section*

THERAPIES WITH CLINICAL BENEFIT (IN PATIENT'S TUMOR TYPE)	THERAPIES WITH CLINICAL BENEFIT (IN OTHER TUMOR TYPE)
Alpelisib + Fulvestrant 1	Everolimus 2A
	Temsirolimus
none	none
none	none

NCCN category

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS (CH)

Genomic findings below may include nontumor somatic alterations, such as CH. The efficacy of targeting such nontumor somatic alterations is unknown. This content should be interpreted based on clinical context. Refer to appendix for additional information on CH.

TET2 - H266fs*24 p. 7

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIALS OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

NSD3 (WHSC1L1) - amplification - equivocal	p. 7	TP53 - R209fs*6	p. 8
SETD2 - R1694fs*17	p. 7	ZNF703 - amplification - equivocal	p. 9
TET2 - H266fs*24	p. 7		

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

ORDERED TEST # ORD-1110066-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and experienced a significantly higher ORR compared

with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI is extremely rare in breast cancer, reported in 0-1% of cases across studies⁶⁻¹¹. The incidence of MSI is increased in triple-negative breast cancer⁹⁻¹¹ and in tumors with homologous recombination defects, such as mutations in BRCA1/2^{9,11}. Notably, in Lynch syndrome-related breast cancer, MSI has been reported in 51-85% of cases¹²⁻¹⁷. A prospective study of 123 patients with breast cancer treated with chemotherapy reported an increase in the incidence of MSI-H following chemotherapy treatment (from 0% pre-treatment to 19% post-treatment) and a significant association between MSI and tumor recurrence¹⁸.

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁹. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁹⁻²¹. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers²²⁻²⁴. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{19,21,23-24}.

BIOMARKER

Tumor Mutational Burden

RESULT

6 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²⁵⁻²⁷, anti-PD-1 therapies²⁵⁻²⁸, and combination nivolumab and ipilimumab²⁹⁻³⁴. In multiple pan-tumor studies, higher TMB has been reported to be associated with increased ORR and OS from treatment with immune checkpoint inhibitors^{25-28,35}. Higher TMB was found to be significantly associated with improved OS upon immune checkpoint inhibitor treatment of patients with 9 types of advanced tumors²⁵. Analyses across several solid tumor types reported that patients with higher TMB (defined as $\geq 16-20$ Muts/Mb) achieved greater clinical benefit from PD-1 or PD-L1-targeting monotherapy, compared with patients with higher TMB treated with chemotherapy³⁶ or those with lower TMB treated with PD-1 or PD-L1-targeting agents²⁶. However, the KEYNOTE 158 trial of pembrolizumab monotherapy in patients with solid tumors found significant improvement in ORR in patients with TMB ≥ 10 Muts/Mb (based on this assay or others)

compared to those with TMB < 10 Muts/Mb, in a large cohort that included multiple tumor types; similar findings were observed in the KEYNOTE 028 and 012 trials^{28,35}. Together, these studies suggest that patients with TMB ≥ 10 Muts/Mb may derive clinical benefit from PD-1 or PD-L1 inhibitors.

FREQUENCY & PROGNOSIS

A study of 3,969 patients with breast cancer reported a median TMB of 2.63 mutations per megabase (Muts/Mb), with 5% of cases harboring TMB ≥ 10 Muts/Mb; median TMB was significantly higher in hormone receptor (HR)-negative and HER2-negative tumors than HR-positive or HER2-positive tumors³⁷. The Breast Invasive Carcinoma TCGA analysis reported an average (non-silent) mutation load of 0.84 Muts/Mb for luminal A tumors, 1.38 Muts/Mb for luminal B tumors, 2.05 Muts/Mb for HER2-enriched tumors, and 1.68 Muts/Mb for basal-like tumors³⁸. In breast cancer, TMB is significantly higher in recurrent versus primary tumors, metastatic versus localized cancers, triple-negative versus HR-positive tumors, and CDH1-mutated versus CDH1-wildtype tumors^{37,39-40}. Among metastatic tumors, TMB-high samples have been reported more frequently in invasive lobular carcinoma (9-17% of cases, depending on the TMB cutoff to designate TMB-high) than in invasive ductal carcinoma (2-8% of cases, depending on the cutoff), and TMB-high (at either cutoff) has not been observed in papillary carcinoma^{37,39-40}. Breast carcinoma harbors a

median TMB of 3.8 muts/Mb, and 3.1% of cases have high TMB (> 20 muts/Mb)⁴¹. In a large study of patients with breast cancer, hypermutation was more frequently observed in metastatic tumors than in primary tumors³⁷. In a study of 14,867 patients with breast cancer, high TMB was associated with older age and metastatic disease but was not significantly associated with PD-L1 positivity using the TMB cutoff of ≥ 10 Muts/Mb⁴⁰. In estrogen receptor-positive breast cancer, increased TMB in tissue samples ($> \text{mean of } 1.25$ Muts/Mb) associated with shorter OS (HR=2.02) in an analysis of the TCGA data⁴².

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁴³⁻⁴⁴ and cigarette smoke in lung cancer⁴⁵⁻⁴⁶, treatment with temozolomide-based chemotherapy in glioma⁴⁷⁻⁴⁸, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁴⁹⁻⁵³, and microsatellite instability (MSI)^{49,52-53}. This sample harbors a TMB level associated with lower rates of clinical benefit from treatment with PD-1- or PD-L1-targeting immune checkpoint inhibitors compared with patients with tumors harboring higher TMB levels, based on several studies in multiple solid tumor types^{26-27,35}.

ORDERED TEST # ORD-1110066-01

GENOMIC FINDINGS

GENE

PIK3CA

ALTERATION

H1047R

TRANSCRIPT ID

NM_006218

CODING SEQUENCE EFFECT

3140A>G

VARIANT ALLELE FREQUENCY (% VAF)

30.3%

POTENTIAL TREATMENT STRATEGIES

Clinical and preclinical data in various tumor types indicate that PIK3CA activating alterations may predict sensitivity to therapies targeting PI3K⁵⁴⁻⁵⁶, AKT⁵⁷, or mTOR⁵⁸⁻⁶⁵. In a Phase 1 trial of the dual PI3K/mTOR kinase inhibitor apitolisib, 79% (11/14) of patients with PIK3CA-mutated advanced solid tumors experienced disease control at the recommended Phase 2 dose (3/14 PRs, 8/14 SDs)⁶⁶. The addition of everolimus to exemestane to treat hormone-receptor-positive (HR+)/HER2-negative advanced breast cancer has shown clinical benefit regardless of PIK3CA status⁶⁷⁻⁶⁸. In the BELLE-2 trial for patients with endocrine-resistant HR+ breast cancer, the combination of the pan-PI3K inhibitor buparlisib with fulvestrant resulted in increased ORR (18% vs. 4%), median PFS (7.0 vs. 3.2 months; HR=0.58, p=0.001), and median OS (26.0 vs. 24.8 months; HR=0.81, p=0.127) compared with placebo with fulvestrant for patients with PIK3CA mutation; no significant improvement in ORR, PFS, or OS was observed for patients without PIK3CA mutation⁶⁹⁻⁷⁰. In a Phase 1 study, the p110 α -selective inhibitor inavolisib (GDC-0077) alone or in combination with endocrine therapy (letrozole or fulvestrant) with or without palbociclib yielded an ORR of 32% (23/73) for patients with PIK3CA-mutated HR+/HER2-negative breast cancer, with an ORR of 40% (6/15) observed for patients who received inavolisib plus palbociclib and fulvestrant⁷¹⁻⁷². A patient with previously treated HER2-negative metastatic breast cancer harboring a PIK3CA H1047R alteration achieved an

exceptional response with the pan-class I PI3K inhibitor copanlisib⁷³. However, studies of copanlisib and the pan-class I PI3K inhibitor buparlisib have demonstrated limited efficacy against PIK3CA-mutated tumors^{70,74-79}. PI3K-alpha-selective inhibitors such as alpelisib or PI3K-beta-sparing inhibitors such as taselisib may have bigger therapeutic windows than pan-PI3K inhibitors⁵⁴. In PIK3CA-mutated advanced solid tumors, alpelisib and taselisib have achieved low ORRs (0% [0/55] to 6% [7/111]) but high DCRs (55% [36/55] to 58% [64/111])⁵⁵. In the Phase 3 SOLAR-1 study, the addition of alpelisib to fulvestrant improved PFS (11.0 vs. 5.7 months, HR=0.65) and ORR (27% vs. 13%) in PIK3CA-mutated HR+/HER2- breast cancer compared with placebo with fulvestrant⁵⁷, but not in PIK3CA-wildtype HR+/HER2- breast cancer. In the Phase 3 SANDPIPER study, the addition of taselisib to fulvestrant improved PFS (7.4 vs. 5.4 months, HR=0.70) and ORR (27% vs. 12%) in PIK3CA-mutated HR+/HER2- breast cancer compared with placebo with fulvestrant⁸⁰; additionally, higher ORR was achieved for patients with multiple PIK3CA mutations following treatment with taselisib (30%, n=43) as compared with those treated with placebo (8.7%, n=23) and for patients with single PIK3CA-mutated tumors treated with either taselisib (18%, n=193) or placebo (10%, n=80)⁸¹. Data from the Phase 2 LOTUS trial reported an improved PFS for patients with PIK3CA/AKT1/PTEN-altered triple negative breast cancer treated with ipatasertib plus paclitaxel compared to paclitaxel alone (9.0 vs 4.9 months)⁸²; however, the Phase 3 IPATunity130 trial did not report a significant PFS improvement for first-line ipatasertib in combination with paclitaxel relative to paclitaxel alone (7.4 vs 6.1 months)⁸³. AKT inhibitors ipatasertib and capivasertib have also been tested in breast cancer. Two Phase 2 studies have reported improved PFS from the addition of either ipatasertib (9.0 vs. 4.9 months, HR = 0.44) or capivasertib (9.3 vs. 3.7 months, HR = 0.30) to paclitaxel in metastatic triple-negative breast cancer harboring PIK3CA/AKT1/PTEN alterations, compared with paclitaxel and placebo⁸⁴. Responses to capivasertib were also reported in 20% (3/15) of patients with PIK3CA-

mutated breast cancer in an earlier study⁸⁵. However, a Phase 1 trial reported no PFS benefit for patients with PIK3CA-mutated, ER+/HER2- metastatic breast cancer from the addition of capivasertib to paclitaxel compared with paclitaxel plus placebo (10.9 vs. 10.8 months)⁸⁶. Activating mutations in PIK3CA may confer resistance to HER2-targeted therapies; combined inhibition of HER2 and the PI3K pathway may be required in HER2-positive tumors with PIK3CA mutation⁸⁷⁻⁹¹. For patients with PIK3CA-mutated breast cancer, PTEN loss or mutation may be associated with resistance to alpelisib in combination with fulvestrant or as a single agent^{54,92-93}.

FREQUENCY & PROGNOSIS

Mutations in PIK3CA have been reported in 25-40% of breast cancer cases^{38,94-98}. In the randomized Phase 2 SAFIRO2 trial, PIK3CA mutations were associated with reduced OS in patients with hormone-receptor-positive (HR+)/HER2 negative (HER-) metastatic breast cancer but with improved OS in patients with mTNBC compared to patients with PIK3CA wildtype status⁹⁸. Although double PIK3CA mutations were frequently observed in HR+/HER2- breast cancers, as compared with other receptor subtypes (15.4% vs. 5.4%, p=0.004), this did not impact invasive disease-free survival or OS for patients when compared with single PIK3CA mutations by univariate and multivariate analysis in 1 retrospective study⁸¹. Mutations in coding exon 20 (H1047R) of PIK3CA have been associated with a better prognosis in breast carcinoma than mutations occurring in coding exon 9 (E542K)⁹⁹.

FINDING SUMMARY

PIK3CA encodes p110-alpha, which is the catalytic subunit of phosphatidylinositol 3-kinase (PI3K). The PI3K pathway is involved in cell signaling that regulates a number of critical cellular functions, including cell growth, proliferation, differentiation, motility, and survival¹⁰⁰⁻¹⁰¹. PIK3CA alterations that have been characterized as activating, such as observed here, are predicted to be oncogenic¹⁰²⁻¹²².

ORDERED TEST # ORD-1110066-01

GENOMIC FINDINGS

GENE

MYC

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

There are no available therapies that directly target MYC. However, preclinical data indicate that MYC overexpression may predict sensitivity to investigational agents targeting CDK1¹²³⁻¹²⁴, CDK2¹²⁵, Aurora kinase A¹²⁶⁻¹³³, Aurora kinase B¹³⁴⁻¹³⁷, glutaminase¹³⁸⁻¹⁴¹, or BET bromodomain-containing proteins¹⁴²⁻¹⁴⁵, as well as agents targeting both HDAC and PI3K¹⁴⁶⁻¹⁴⁸. A Phase 2 study reported a PFS benefit associated with a combination of the Aurora A kinase inhibitor

alisertib and paclitaxel as second-line therapy for patients with MYC-overexpressed small cell lung cancer but not for patients without MYC overexpression¹⁴⁹. A patient with MYC-amplified invasive ductal breast carcinoma experienced a PR to an Aurora kinase inhibitor¹⁵⁰. The glutaminase inhibitor CB-839, in combination with either everolimus or cabozantinib, has demonstrated encouraging efficacy in Phase 1 and 2 studies enrolling patients with pretreated advanced renal cell carcinoma¹⁵¹⁻¹⁵². MYC amplification has also been suggested to predict response to chemotherapy in patients with breast cancer in some studies¹⁵³⁻¹⁵⁴. Preclinical evidence suggests that colon cancer cells with MYC amplification may be more sensitive to 5-fluorouracil and paclitaxel¹⁵⁵⁻¹⁵⁶.

FREQUENCY & PROGNOSIS

In the TCGA dataset, MYC amplification was observed in 15% of breast invasive carcinoma cases³⁸. MYC amplification has been associated with an aggressive phenotype, early onset, and poor prognosis in patients with breast cancer, although the data have been conflicting^{153,157-159}.

FINDING SUMMARY

MYC (c-MYC) encodes a transcription factor that regulates many genes related to cell cycle regulation and cell growth. It is an oncogene and may be activated in as many as 20% of cancers¹⁶⁰. MYC dysregulation (amplification, overexpression, translocation) has been identified in a number of different cancer types¹⁶¹. MYC amplification has been significantly linked with increased mRNA and protein levels and results in the dysregulation of a large number of target genes^{160,162-163}.

ORDERED TEST # ORD-1110066-01

GENOMIC FINDINGS

GENE

PTEN

ALTERATION

Y174fs*15

TRANSCRIPT ID

NM_000314

CODING SEQUENCE EFFECT

518_519insAGGGAGTAACATTCCAGTCAGAGGCG

VARIANT ALLELE FREQUENCY (% VAF)

38.5%

POTENTIAL TREATMENT STRATEGIES

PTEN loss or mutation leads to activation of the PI3K-AKT-mTOR pathway and may predict sensitivity to inhibitors of this pathway^{75,164-168} such as the mTOR inhibitors temsirolimus and everolimus or the PI3K inhibitor copanlisib. While PTEN loss correlated with longer PFS and response to single-agent everolimus for patients with prostate cancer¹⁶⁴, clinical studies in renal cell carcinoma¹⁶⁹⁻¹⁷³, glioblastoma¹⁷⁴⁻¹⁷⁵, cervical carcinoma¹⁷⁶, breast cancer¹⁷⁷⁻¹⁷⁸, endometrial cancer¹⁷⁹⁻¹⁸⁴, urothelial carcinoma¹⁸⁵⁻¹⁸⁷, leiomyosarcoma¹⁸⁸, neuroendocrine tumors¹⁸⁹, gastric cancer¹⁹⁰, and peripheral T-cell lymphomas¹⁹¹ did not observe a correlation of PTEN deficiency with response to everolimus or temsirolimus. Preclinical studies suggest that PTEN-deficient cancers, in the absence of other oncogenic mutations, depend primarily on the beta isoform of PI3K (PI3K-beta)¹⁹²⁻¹⁹⁴, and PI3K-beta-selective inhibitors are in clinical trials for PTEN-deficient tumors. However, the NCI-MATCH Phase 2 study observed limited activity of the PI3K-beta-selective inhibitor GSK2636771 as monotherapy in PTEN-deficient cancers, with a median PFS of 1.8 months. The best outcomes were 1 PR (1/22, prostate cancer), SD (7/22) for patients with PTEN deletion/mutation, and SD (9/34) for patients with PTEN protein loss¹⁹⁵. Clinical data in breast¹⁹⁶⁻¹⁹⁷ and prostate cancer¹⁹⁸⁻¹⁹⁹ suggest that PTEN alterations may predict sensitivity to pan-AKT inhibitors such as ipatasertib or capivasertib. Phase 2 studies have reported improved PFS from the addition of either

ipatasertib (9.0 vs. 4.9 months, HR=0.44) or capivasertib (9.3 vs. 3.7 months, HR=0.30) to paclitaxel, compared with paclitaxel and placebo, for patients with metastatic triple-negative breast cancer harboring PIK3CA/AKT1/PTEN alterations⁸⁴. However, data from the Phase 2 LOTUS trial reported an improved PFS for patients with PIK3CA/AKT1/PTEN-altered triple negative breast cancer treated with ipatasertib plus paclitaxel compared to paclitaxel alone (9.0 vs 4.9 months)⁸²; however, the Phase 3 IPATunity130 trial did not report a significant PFS improvement for first-line ipatasertib in combination with paclitaxel relative to paclitaxel alone (7.4 vs 6.1 months)⁸³. Preclinical data indicate that PTEN loss or inactivation may predict sensitivity to PARP inhibitors²⁰⁰⁻²⁰⁴, and clinical benefit has been observed for patients with PTEN-altered breast cancer²⁰⁵, ovarian cancer²⁰⁶, endometrial cancer²⁰⁴, and other tumor types²⁰⁷ treated with PARP inhibitors. However, several studies have reported a lack of association between PTEN mutation and PARP inhibitor sensitivity^{203,208-209}. Limited clinical evidence in glioblastoma²¹⁰, leiomyosarcoma²¹¹, NSCLC²¹², and melanoma²¹³ suggests that PTEN alterations may predict a lack of response to anti-PD-1 therapy. In an analysis of 39 patients with metastatic melanoma treated with pembrolizumab or nivolumab, patients with PTEN-expressing tumors achieved significantly greater reduction of tumor size than those with reduction or loss of PTEN expression²¹³. In a retrospective analysis of 66 patients with glioblastoma, tumors from nivolumab or pembrolizumab non-responders were significantly enriched for PTEN mutations²¹⁰. In a patient with uterine leiomyosarcoma treated with pembrolizumab monotherapy, a treatment-resistant tumor arose that harbored PTEN loss²¹¹. A patient with NSCLC whose tumor harbored a PTEN alteration exhibited a lack of response to nivolumab and pembrolizumab²¹². Clinical and preclinical evidence suggests that PTEN loss or mutation may predict resistance to PI3K inhibitors^{92,214-215}, and to CDK inhibitors such as palbociclib, ribociclib, and abemaciclib^{214,216}. For patients with PIK3CA-mutated breast cancer, PTEN loss or mutation may be associated with

resistance to alpelisib in combination with fulvestrant or as a single agent^{54,92-93}.

FREQUENCY & PROGNOSIS

In the TCGA dataset, PTEN mutation has been reported in 4% of breast invasive carcinomas, while putative homozygous deletion of PTEN has been reported in 2% of cases³⁸. PTEN mutation has also been observed in 5.3% (1/19) of metaplastic breast cancers²¹⁷ and 2% of invasive lobular carcinoma tumors analyzed²¹⁸. PTEN mutations are associated more frequently with triple-negative breast cancer than with HER2- or hormone-positive breast cancer²¹⁹⁻²²⁰. Loss or reduction of PTEN expression has been observed in 28% of invasive ductal breast carcinomas and has been correlated with metastasis and poor patient prognosis, including decreased 2-year disease-free survival²²¹⁻²²³.

FINDING SUMMARY

PTEN encodes an inositol phosphatase that functions as a tumor suppressor by negatively regulating the PI3K-AKT-mTOR pathway; loss of PTEN can lead to uncontrolled cell growth and suppression of apoptosis¹⁶⁷. Alterations such as seen here may disrupt PTEN function or expression²²⁴⁻²⁶⁴.

POTENTIAL GERMLINE IMPLICATIONS

PTEN mutations underlie several inherited disorders, collectively termed PTEN hamartoma tumor syndrome (PHTS), which include Cowden syndrome (CS) and its variant Lhermitte-Duclos disease (LD), Bannayan-Riley-Ruvalcaba syndrome (BRRS), PTEN-related Proteus syndrome (PS), and Proteus-like syndrome²⁶⁵⁻²⁶⁶. The mutation rate for PTEN in these disorders ranges from 20 to 85% of patients^{265,267}. The estimated incidence of Cowden syndrome is 1/200,000, which may be an underestimate due to the high variability of this disorder²⁶⁵. Given the association between PTEN and these inherited syndromes, in the appropriate clinical context, germline testing for mutations affecting PTEN is recommended.

ORDERED TEST # ORD-1110066-01

GENOMIC FINDINGS

GENE

NSD3 (WHSC1L1)

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

There are no targeted therapies available to address genomic alterations in NSD3.

FREQUENCY & PROGNOSIS

In TCGA datasets, NSD3 amplification has been most frequently observed in lung squamous cell carcinoma (17%)²⁶⁸, breast invasive carcinoma (13%)²⁶⁹, bladder urothelial carcinoma (9%)²⁷⁰, and head and neck squamous cell carcinoma (9%)²⁷¹ samples²⁷²⁻²⁷³. Amplification of at least one member of the NSD3-CHD8-BRD4 pathway has been associated with worse overall survival in ovarian high-grade serous carcinoma and endometrial cancer²⁷⁴. In endometrial cancers,

amplification of this pathway was more frequent in endometrial serous and endometrioid serous-like carcinomas compared to low-grade endometrioid endometrial adenocarcinomas²⁷⁴.

FINDING SUMMARY

NSD3, also known as WHSC1L1, encodes an enzyme that mediates histone methylation²⁷⁵. NSD3 has been shown to be amplified in various cancers²⁷⁶⁻²⁷⁸.

GENE

SETD2

ALTERATION

R1694fs*17

TRANSCRIPT ID

NM_014159

CODING SEQUENCE EFFECT

5082_5083delAG

VARIANT ALLELE FREQUENCY (% VAF)

8.3%

POTENTIAL TREATMENT STRATEGIES

There are no targeted therapies available to address genomic alterations in SETD2.

FREQUENCY & PROGNOSIS

Somatic inactivating alterations of SETD2 are documented to occur at low frequency in a number of solid tumors, most commonly in renal carcinoma²⁷⁹. SETD2 mutations have been detected in 6-12% of acute lymphoblastic leukemias (ALL) and reportedly increase chromosomal abnormalities and contribute to

leukemia development²⁸⁰⁻²⁸².

FINDING SUMMARY

SETD2 encodes a histone lysine-36 methyltransferase²⁸³ that preferentially interacts with the expanded N-terminal polyglutamine tracts present in mutant huntingtin, implicating it in the pathogenesis of Huntington disease²⁸⁴. SETD2 mRNA expression has been observed to be consistently reduced in breast tumors relative to adjacent non-tumor tissue, suggesting a potential tumor suppressor role²⁸⁵.

GENE

TET2

ALTERATION

H266fs*24

TRANSCRIPT ID

NM_001127208

CODING SEQUENCE EFFECT

797_806delACCCATCGCA

VARIANT ALLELE FREQUENCY (% VAF)

11.7%

FREQUENCY & PROGNOSIS

TET2 alterations have been reported at relatively low frequencies in solid tumors and are more prevalent in hematological malignancies (cBioPortal, Jan 2021)²⁷²⁻²⁷³. Published data investigating the prognostic implications of TET2 alterations in solid tumors are limited (PubMed, Jan 2021).

FINDING SUMMARY

TET2 encodes a tumor suppressor involved in reversing DNA methylation marks, a process critical for proper gene regulation²⁸⁶⁻²⁸⁷. Alterations such as seen here may disrupt TET2 function or expression²⁸⁸⁻²⁹².

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to

occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion²⁹³⁻²⁹⁸. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy²⁹³⁻²⁹⁴. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease²⁹⁹. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{297,300-301}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

POTENTIAL TREATMENT STRATEGIES

There are no targeted therapies available to address genomic alterations in TET2 in solid tumors.

ORDERED TEST # ORD-1110066-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

R209fs*6

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

626_627delGA

VARIANT ALLELE FREQUENCY (% VAF)

52.0%

POTENTIAL TREATMENT STRATEGIES

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib³⁰²⁻³⁰⁵, or p53 gene therapy and immunotherapeutics such as SGT-53³⁰⁶⁻³¹⁰ and ALT-801³¹¹. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% (17/176) and SDs in 53.4% (94/176) of patients with solid tumors; the response rate was 21.1% (4/19) in patients with TP53 mutations versus 12.1% (4/33) in patients who were TP53 wild-type³¹². A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 31.9% (30/94, 3 CR) ORR and a 73.4% (69/94) DCR in patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer³¹³. A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 42.9% (9/21, 1 CR) ORR and a 76.2% (16/21) DCR in patients with platinum-refractory TP53-mutated ovarian cancer³¹⁴. The combination of adavosertib with paclitaxel and carboplatin in patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone³¹⁵. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24.0% (6/25) ORR with adavosertib combined with

paclitaxel³¹⁶. A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71.4% (5/7) response rate for patients with TP53 alterations³¹⁷. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel in patients with solid tumors, 75.0% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage³¹⁰. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53-mutated, but not TP53-wild-type, breast cancer xenotransplant mouse model³¹⁸. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies³¹⁹⁻³²⁰; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies³²¹⁻³²². Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in breast cancer; mutations in this gene have been identified in 27-37% of breast carcinoma samples^{38,323-327}. TP53 mutations that are located within the region encoding the DNA binding domain are associated with poor prognosis in patients with breast cancer^{325,328-329}. TP53 mutation is also implicated in breast cancer susceptibility, as TP53 mutation carriers have an 18-60 fold increased risk for early onset breast cancer³³⁰⁻³³².

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers³³³. Alterations such as seen here may disrupt TP53 function or expression³³⁴⁻³³⁸.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Mar 2021)³³⁹. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers³⁴⁰⁻³⁴², including sarcomas³⁴³⁻³⁴⁴. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000³⁴⁵ to 1:20,000³⁴⁴. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30³⁴⁶. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion²⁹³⁻²⁹⁸. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy²⁹³⁻²⁹⁴. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease²⁹⁹. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{297,300-301}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # **ORD-1110066-01**
GENOMIC FINDINGS
GENE
ZNF703
ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

There are no available targeted therapies to directly address ZNF703 alterations in cancer. One preclinical study suggested that ZNF703 expression in breast cancer cell lines is associated with reduced sensitivity to tamoxifen through AKT-mTOR activation³⁴⁷, although these findings

have not been verified in the clinical setting.

FREQUENCY & PROGNOSIS

Amplification and high expression of ZNF703 has been observed in luminal B breast tumors, a subtype associated with aggressive disease progression and poor patient outcomes³⁴⁸⁻³⁵⁰. ZNF703 expression has also been linked with aggressive tumor characteristics in patients with gastric and colorectal cancers³⁵¹⁻³⁵². Putative high-level amplification of ZNF703 has been reported with the highest frequency in breast carcinoma, bladder urothelial carcinoma, uterine carcinosarcoma, lung squamous cell carcinoma (SCC), esophageal carcinoma and head and neck

SCC (5-13% of samples)(cBioPortal, 2021)²⁷²⁻²⁷³.

FINDING SUMMARY

ZNF703 encodes a transcriptional repressor that plays roles in stem cell proliferation, cell cycle progression, and other key cellular functions^{349,353}. Amplification of ZNF703 has been correlated with protein expression³⁴⁸⁻³⁴⁹. ZNF703 was established as a breast cancer oncoprotein by studies showing that ZNF703 expression resulted in transformation and increased proliferation of cultured cells^{348-349,354}, as well as increased lung metastases in a breast cancer xenograft model³⁵⁴.

ORDERED TEST # ORD-1110066-01

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Alpelisib + Fulvestrant

Assay findings association

PIK3CA
H1047R

AREAS OF THERAPEUTIC USE

Alpelisib is a phosphatidylinositol 3-kinase (PI3K) inhibitor with selective activity against the alpha isoform (PI3K-alpha), and fulvestrant is an estrogen receptor (ER) antagonist and selective estrogen receptor degrader (SERD). The combination is FDA approved to treat men and postmenopausal women with hormone receptor (HR)-positive, human epidermal growth factor receptor 2 (HER2)-negative, PIK3CA-mutated advanced breast cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of prospective clinical data, PIK3CA mutations including C420R, E542K, E545A, E545G, E545K, E545D, Q546E, Q546R, H1047L, H1047Y, and H1047R are associated with sensitivity to alpelisib in combination with fulvestrant. In ER+/HER2- breast cancer, PFS benefit from the addition of alpelisib to fulvestrant was specifically observed for patients with PIK3CA mutations (11.0 vs. 5.7 months, HR=0.65), including patients with PIK3CA exon 9 or exon 20 mutations⁵⁷. Preclinical and limited clinical evidence suggest that PTEN inactivation may be associated with resistance to PI3K alpha-selective inhibitors^{54,92-93}. PTEN mutation or loss has been reported for a small number of patients with PIK3CA-mutated HR+/HER2- breast cancer who progressed on combination treatment with alpelisib and fulvestrant, including as an acquired alteration for 1 patient^{93,355}.

SUPPORTING DATA

In the Phase 3 SOLAR-1 study for patients with HR+/HER2- endocrine therapy-resistant advanced breast cancer, addition of alpelisib to fulvestrant significantly improved median PFS (mPFS; 11.0 vs. 5.7 months, HR=0.65), ORR (27% vs. 13%), and clinical benefit rate (CBR; 62% vs. 45%), and numerically improved median OS (39.3 vs. 31.4 months, HR=0.86) for patients with PIK3CA mutation^{57,356}. Benefit was observed for patients with PIK3CA exon 9 or exon 20 mutations⁵⁷; for PIK3CA-wildtype patients, the addition of alpelisib to fulvestrant did not significantly improve mPFS (7.4 vs. 5.6 months, HR=0.85)⁵⁷. This trial excluded patients with active brain metastases; however, control of progressive brain metastases (1/4 PR and 2/4 SDs by RANO brain metastases criteria) was reported in a case series of 4 patients with PIK3CA-mutated HR+/HER2- breast cancer treated with alpelisib in combination with either fulvestrant or exemestane³⁵⁷. The Phase 2 BYLieve study for previously treated patients with PIK3CA-mutated HR+/HER2- advanced breast cancer reported an ORR of 17%, a CBR of 46%, and mPFS of 7.3 months for patients treated with alpelisib plus fulvestrant following progression on a CDK4/6 inhibitor in combination with an aromatase inhibitor³⁵⁸ and, similarly in another cohort, an ORR of 16%, CBR of 32%, and mPFS of 5.7 months for patients treated with alpelisib plus letrozole following progression on a CDK4/6 inhibitor in combination with fulvestrant³⁵⁹.

ORDERED TEST # ORD-1110066-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Everolimus

Assay findings association
PIK3CA
H1047R

AREAS OF THERAPEUTIC USE

Everolimus is an orally available mTOR inhibitor that is FDA approved to treat renal cell carcinoma (RCC) following antiangiogenic therapy; pancreatic neuroendocrine tumors; and well-differentiated non-functional neuroendocrine tumors of the lung or gastrointestinal tract. Everolimus is also approved to treat either renal angiomyolipoma or subependymal giant cell astrocytoma in association with tuberous sclerosis complex (TSC). Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence⁵⁸⁻⁶⁵, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors^{62-65,360-364}.

SUPPORTING DATA

Clinical benefit has been reported for patients with PIK3CA-mutated breast cancer treated with everolimus as a single agent³⁶⁵ or in combination with gemcitabine and cisplatin³⁶⁶. For patients with HR+, HER2- advanced breast cancer, the BOLERO-2, SAFIRTOR, and EVE biomarker studies demonstrated clinical benefit from addition of everolimus to exemestane regardless of baseline PIK3CA mutation status^{67-68,367-368}. In the Phase 3 BOLERO-2 study for hormone receptor-positive (HR+), HER2-negative (HER2-) breast cancer, the addition of everolimus to exemestane improved median PFS (mPFS)

in both the first-line exploratory cohort (11.5 vs. 4.1 months, HR=0.39)³⁶⁹ and second-line cohort (7.8 vs. 3.2 months, HR=0.45)³⁷⁰⁻³⁷² compared with exemestane alone. Combination everolimus and exemestane modestly improved mPFS compared with everolimus alone in BOLERO-6 (8.4 vs. 6.8 months, HR=0.74)³⁷³. Patients with HR+, HER2- breast cancer also benefited from everolimus combined with other anti-estrogen therapies, including letrozole, tamoxifen, and anastrozole³⁷⁴⁻³⁷⁶. For patients with HR+, HER2- breast cancer who progressed on anti-estrogen therapies, addition of everolimus to the most recent endocrine therapy elicited mPFS of 6.6 months³⁷⁷. For patients with HER2+ breast cancer, everolimus combined with trastuzumab plus paclitaxel did not significantly improve mPFS in the full study population (15.0 months with everolimus vs. 14.5 months with placebo), but increased PFS in the HR-negative subpopulation (20.3 vs. 13.1 months)³⁷⁸. For patients with trastuzumab-resistant HER2+ breast cancer, the addition of everolimus to trastuzumab plus vinorelbine prolonged mPFS (7.0 vs. 5.8 months)³⁷⁹, whereas for HER2- breast cancer, addition of everolimus to vinorelbine in the second-line did not improve mPFS (4.0 vs. 4.1 months)³⁸⁰. Patients with metastatic triple-negative breast cancer treated with everolimus plus carboplatin achieved a clinical benefit rate of 36% (9/25)³⁸¹. Whereas frequent adverse events precluded a recommended Phase 2 dose and schedule for the combination of trametinib and everolimus in a Phase 1b trial for solid tumors³⁸², a retrospective study for heavily pretreated patients with solid tumors reported tolerable regimens of the combination for 23/31 patients, with 16 patients treated >3 months and evaluable patients achieving a median PFS of 6.5 months³⁸³.

ORDERED TEST # ORD-1110066-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Temsirolimus

Assay findings association

PIK3CA
H1047R

AREAS OF THERAPEUTIC USE

Temsirolimus is an intravenous mTOR inhibitor that is FDA approved for the treatment of advanced renal cell carcinoma. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical evidence⁵⁸⁻⁶⁵, PIK3CA activating mutations may predict sensitivity to mTOR inhibitors such as everolimus and temsirolimus. Studies have reported modest activity of these therapies as single agents (ORRs of 0-4%), but improved activity has been observed when they are combined with other agents such as bevacizumab and doxorubicin (ORRs of 25-44%), for patients with PIK3CA-mutated solid tumors^{62-65,360-364}.

SUPPORTING DATA

Clinical benefit has been reported for patients with PIK3CA-mutated breast cancer treated with temsirolimus as a single agent¹⁷⁸ or in combination with doxorubicin and bevacizumab^{64-65,363,384-386}. A Phase 1 trial examining the combination of temsirolimus, liposomal doxorubicin,

and bevacizumab in 74 patients with breast and gynecological malignancies reported that 37.9% of patients experienced either a CR (1.4%), PR (18.9%), or SD (17.6%); among 25 patients with PIK3CA mutation or PTEN loss, 52% experienced a CR, PR (36%), or SD (16%)⁶⁵. Another Phase 1 trial including patients with several types of cancer reported a 42% incidence of complete or partial responses in patients with metastatic breast cancer⁶³. However, a Phase 2 study of temsirolimus in pretreated patients with metastatic breast cancer reported minimal clinical activity and no association with PTEN protein or PIK3CA mutation status¹⁷⁸. A Phase 3 placebo-controlled trial of letrozole plus oral temsirolimus as first-line endocrine therapy in postmenopausal women with locally advanced or metastatic breast cancer was terminated at the second interim since the addition of temsirolimus to letrozole did not improve PFS as a first-line therapy³⁸⁷. A study examining the efficacy of temsirolimus-involving regimens in 24 patients with mesenchymal/metaplastic breast cancer (MpBCs) reported 2 CRs, 4 PRs, 2 instances of SD longer than 6 months, and 4 instances of SD shorter than 6 months³⁶³.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

ORDERED TEST # **ORD-1110066-01**
CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
MYC
ALTERATION
amplification - equivocal
RATIONALE
MYC overexpression may predict sensitivity to inhibition of CDKs, especially CDK1 and CDK2, of Aurora kinases, including Aurora kinase A and B,

and of BET domain proteins, which are reported to downregulate MYC expression and MYC-dependent transcriptional programs.

NCT03297424
PHASE 1/2

A Study of PLX2853 in Advanced Malignancies.

TARGETS
BRD4
LOCATIONS: Florida, Texas, Virginia, New York, Arizona

NCT03901469
PHASE 2

A Study of ZEN003694 and Talazoparib in Patients With Triple Negative Breast Cancer

TARGETS
BRD2, BRD3, BRD4, BRDT, PARP
LOCATIONS: Texas, Tennessee, Pennsylvania, New York, Kansas, Arizona, Madrid (Spain), Barcelona (Spain), Brussels (Belgium), Leuven (Belgium)

NCT04555837
PHASE 1/2

Alistertib and Pembrolizumab for the Treatment of Patients With Rb-deficient Head and Neck Squamous Cell Cancer

TARGETS
Aurora kinase A, PD-1
LOCATIONS: Texas

NCT02516553
PHASE 1

BI 894999 First in Human Dose Finding Study in Advanced Malignancies

TARGETS
BRD2, BRD3, BRD4, BRDT
LOCATIONS: Texas, New York, Ohio, Massachusetts, California, Madrid (Spain), Nantes (France), Barcelona (Spain), Villejuif (France), Paris (France)

NCT03654547
PHASE 1

Safety of TT-00420 Monotherapy in Patients With Advanced Solid Tumors and Triple Negative Breast Cancer

TARGETS
Aurora kinase A, Aurora kinase B
LOCATIONS: Texas

NCT01434316
PHASE 1

Veliparib and Dinaciclib in Treating Patients With Advanced Solid Tumors

TARGETS
PARP, CDK1, CDK2, CDK5, CDK9
LOCATIONS: Massachusetts

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Electronically signed by Douglas A. Mata, MD, MPH | 15 June 2021
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Sample Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

ORDERED TEST # **ORD-1110066-01**
CLINICAL TRIALS
NCT03220347
PHASE 1

A Study to Assess the Safety, Tolerability, Pharmacokinetics and Preliminary Efficacy of CC-90010 in Subjects With Advanced Solid Tumors and Relapsed/Refractory Non-Hodgkin's Lymphomas

TARGETS
BRD2, BRD3, BRD4, BRDT
LOCATIONS: Madrid (Spain), Bordeaux (France), Barcelona (Spain), Villejuif (France), Rozzano (MI) (Italy), Meldola (Italy), Napoli, Campania (Italy), Kashiwa (Japan)

ORDERED TEST # ORD-1110066-01

CLINICAL TRIALS

GENE
PIK3CA
ALTERATION
H1047R

RATIONALE
PIK3CA activating mutations may lead to activation of the PI3K-AKT-mTOR pathway and may therefore indicate sensitivity to inhibitors of

this pathway. Strong clinical data support sensitivity of PIK3CA-mutated solid tumors to the PI3K-alpha inhibitor alpelisib.

NCT04305496
PHASE 3

Capiwasertib+Fulvestrant vs Placebo+Fulvestrant as Treatment for Locally Advanced (Inoperable) or Metastatic HR+/HER2- Breast Cancer

TARGETS
ER, AKTs

LOCATIONS: Lima (Peru), Arequipa (Peru), La Rioja (Argentina), Rosario (Argentina), Ciudad Autonoma De Buenos Aire (Argentina), Berazategui (Argentina), Viedma (Argentina), Florida, Georgia

NCT03997123
PHASE 3

Capiwasertib+Paclitaxel as First Line Treatment for Patients With Locally Advanced or Metastatic TNBC

TARGETS
AKTs

LOCATIONS: Lima (Peru), Callao (Peru), Rosario (Argentina), Londrina (Brazil), Goiania (Brazil), São José do Rio Preto (Brazil), Mar del Plata (Argentina), Caba (Argentina), Ciudad Autonoma De Buenos Aire (Argentina), Ciudad Autónoma de Bs. As. (Argentina)

NCT04251533
PHASE 3

Study Assessing the Efficacy and Safety of Alpelisib + Nab-paclitaxel in Subjects With Advanced TNBC Who Carry Either a PIK3CA Mutation or Have PTEN Loss Without PIK3CA Mutation

TARGETS
PI3K-alpha

LOCATIONS: Bogota (Colombia), Barretos (Brazil), Caxias do Sul (Brazil), Sao Paulo (Brazil), Florida, Louisiana, Texas

NCT04191499
PHASE 2/3

A Study Evaluating the Efficacy and Safety of GDC-0077 + Palbociclib + Fulvestrant vs Placebo + Palbociclib + Fulvestrant in Patients With PIK3CA-Mutant, Hormone Receptor-Positive, Her2-Negative, Locally Advanced or Metastatic Breast Cancer

TARGETS
PI3K-alpha, CDK4, CDK6, ER

LOCATIONS: Florida, Texas, Georgia

NCT04589845
PHASE 2

Tumor-Agnostic Precision Immuno-Oncology and Somatic Targeting Rational for You (TAPISTRY) Platform Study

TARGETS
ALK, ROS1, TRKA, TRKB, TRKC, RET, PD-L1, AKTs, ERBB2, MDM2, PI3K-alpha

LOCATIONS: Florida, Alabama, Texas, Georgia, Tennessee, Pennsylvania

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Florida, Louisiana, Texas

ORDERED TEST # ORD-1110066-01

CLINICAL TRIALS

NCT04632992
PHASE 2

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

TARGETS

ALK, ROS1, TRKA, TRKB, TRKC, PD-L1, ERBB2, ERBB3, PI3K-alpha, RET, AKTs

LOCATIONS: Florida, Louisiana, Tennessee, Texas, New Jersey

NCT04188548
PHASE 1

A Study of LY3484356 in Participants With Advanced or Metastatic Breast Cancer or Endometrial Cancer

TARGETS

mTOR, Aromatase, CDK4, CDK6, ER, PI3K-alpha, ERBB2

LOCATIONS: Florida, Texas, Georgia, North Carolina, Tennessee, Virginia

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS

mTORC1, mTORC2, PD-1

LOCATIONS: Chengdu (China), Chongqing (China)

NCT03280563
PHASE 1/2

A Study of Multiple Immunotherapy-Based Treatment Combinations in Hormone Receptor (HR)-Positive Human Epidermal Growth Factor Receptor 2 (HER2)-Negative Breast Cancer

TARGETS

PD-L1, ER, HDAC, AKTs, CDK4, CDK6

LOCATIONS: North Carolina, Tennessee, Maryland, Pennsylvania, New York, Illinois, California

ORDERED TEST # ORD-1110066-01

CLINICAL TRIALS

GENE
PTEN
ALTERATION

Y174fs*15

RATIONALE

PTEN loss or inactivating mutations may lead to increased activation of the PI3K-AKT-mTOR pathway and may indicate sensitivity to inhibitors

of this pathway. PTEN loss or inactivation may also predict sensitivity to PARP inhibitors.

NCT04305496
PHASE 3

Capiivasertib+Fulvestrant vs Placebo+Fulvestrant as Treatment for Locally Advanced (Inoperable) or Metastatic HR+/HER2- Breast Cancer

TARGETS
ER, AKTs

LOCATIONS: Lima (Peru), Arequipa (Peru), La Rioja (Argentina), Rosario (Argentina), Ciudad Autonoma De Buenos Aire (Argentina), Berazategui (Argentina), Viedma (Argentina), Florida, Georgia

NCT03997123
PHASE 3

Capiivasertib+Paclitaxel as First Line Treatment for Patients With Locally Advanced or Metastatic TNBC

TARGETS
AKTs

LOCATIONS: Lima (Peru), Callao (Peru), Rosario (Argentina), Londrina (Brazil), Goiania (Brazil), São José do Rio Preto (Brazil), Mar del Plata (Argentina), Caba (Argentina), Ciudad Autonoma De Buenos Aire (Argentina), Ciudad Autónoma de Bs. As. (Argentina)

NCT04191135
PHASE 2/3

Study of Olaparib Plus Pembrolizumab Versus Chemotherapy Plus Pembrolizumab After Induction With First-Line Chemotherapy Plus Pembrolizumab in Triple Negative Breast Cancer (TNBC) (MK-7339-009/KEYLYNK-009)

TARGETS
PD-1, PARP

LOCATIONS: Cali (Colombia), Medellin (Colombia), La Serena (Chile), Monteria (Colombia), Vina del Mar (Chile), Santiago (Chile), Barranquilla (Colombia), Temuco (Chile), Florida, Texas

NCT03598257
PHASE 2

Radiation Therapy With or Without Olaparib in Treating Patients With Inflammatory Breast Cancer

TARGETS
PARP

LOCATIONS: San Juan (Puerto Rico), Florida, Louisiana, Georgia, South Carolina, Texas

NCT03994796
PHASE 2

Genetic Testing in Guiding Treatment for Patients With Brain Metastases

TARGETS
ALK, ROS1, TRKA, TRKB, TRKC, CDK4, CDK6, PI3K, mTOR

LOCATIONS: Florida, Louisiana, Texas

NCT04632992
PHASE 2

A Study Evaluating Targeted Therapies in Participants Who Have Advanced Solid Tumors With Genomic Alterations or Protein Expression Patterns Predictive of Response

TARGETS
ALK, ROS1, TRKA, TRKB, TRKC, PD-L1, ERBB2, ERBB3, PI3K-alpha, RET, AKTs

LOCATIONS: Florida, Louisiana, Tennessee, Texas, New Jersey

ORDERED TEST # ORD-1110066-01

CLINICAL TRIALS
NCT02498613
PHASE 2

A Phase 2 Study of Cediranib in Combination With Olaparib in Advanced Solid Tumors

TARGETS
PARP, VEGFRs

LOCATIONS: Florida, Texas, Tennessee, Virginia, Connecticut, Massachusetts, Toronto (Canada), California

NCT01042379
PHASE 2

I-SPY 2 TRIAL: Neoadjuvant and Personalized Adaptive Novel Agents to Treat Breast Cancer

TARGETS
PARP, PD-L1, ERBB2, ERBB3, PD-1,
TLR9, LAG-3

LOCATIONS: Florida, Georgia, Alabama, North Carolina, Tennessee, District of Columbia, Pennsylvania, New York, Connecticut

NCT03329001
PHASE 1

Crossover Study to Assess the Relative Bioavailability and Bioequivalence of Niraparib Tablet Compared to Niraparib Capsule

TARGETS
PARP

LOCATIONS: Florida, Georgia, Texas, Tennessee, Oklahoma, Connecticut, Michigan, Colorado, California

NCT04337463
PHASE NULL

ATG-008 Combined With Toripalimab in Advanced Solid Tumors

TARGETS
mTORC1, mTORC2, PD-1

LOCATIONS: Chengdu (China), Chongqing (China)

ORDERED TEST # ORD-1110066-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BCL6

Q397H

GATA3

amplification

MERTK

P81fs*27

MTOR

G305A

NBN

amplification

RAD21

amplification

TSC1

K587R

XPO1

I87L

ORDERED TEST # ORD-1110066-01

APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

ABL1	ACVR1B	AKT1	AKT2	AKT3	ALK	ALOX12B	AMER1 (FAM123B)	APC
AR	ARAF	ARFRP1	ARID1A	ASXL1	ATM	ATR	ATRX	AURKA
AURKB	AXIN1	AXL	BAP1	BARD1	BCL2	BCL2L1	BCL2L2	BCL6
BCOR	BCORL1	BRAF	BRCA1	BRCA2	BRD4	BRIP1	BTG1	BTG2
BTB	C11orf30 (EMSY)	C17orf39 (GID4)	CALR	CARD11	CASP8	CBFB	CBL	CCND1
CCND2	CCND3	CCNE1	CD22	CD274 (PD-L1)	CD70	CD79A	CD79B	CDC73
CDH1	CDK12	CDK4	CDK6	CDK8	CDKN1A	CDKN1B	CDKN2A	CDKN2B
CDKN2C	CEBPA	CHEK1	CHEK2	CIC	CREBBP	CRKL	CSF1R	CSF3R
CTCF	CTNNA1	CTNNB1	CUL3	CUL4A	CXCR4	CYP17A1	DAXX	DDR1
DDR2	DIS3	DNMT3A	DOT1L	EED	EGFR	EP300	EPHA3	EPHB1
EPHB4	ERBB2	ERBB3	ERBB4	ERCC4	ERG	ERRF1	ESR1	EZH2
FAM46C	FANCA	FANCC	FANCG	FANCL	FAS	FBXW7	FGF10	FGF12
FGF14	FGF19	FGF23	FGF3	FGF4	FGF6	FGFR1	FGFR2	FGFR3
FGFR4	FH	FLCN	FLT1	FLT3	FOXO2	FUBP1	GABRA6	GATA3
GATA4	GATA6	GNA11	GNA13	GNAQ	GNAS	GRM3	GSK3B	H3F3A
HDAC1	HGF	HNF1A	HRAS	HSD3B1	ID3	IDH1	IDH2	IGF1R
IKBKE	IKZF1	INPP4B	IRF2	IRF4	IRS2	JAK1	JAK2	JAK3
JUN	KDMSA	KDM5C	KDM6A	KDR	KEAP1	KEL	KIT	KLHL6
KMT2A (MLL)	KMT2D (MLL2)	KRAS	LTK	LYN	MAF	MAP2K1 (MEK1)	MAP2K2 (MEK2)	MAP2K4
MAP3K1	MAP3K13	MAPK1	MCL1	MDM2	MDM4	MED12	MEF2B	MEN1
MERTK	MET	MITF	MKNK1	MLH1	MPL	MRE11A	MSH2	MSH3
MSH6	MST1R	MTAP	MTOR	MUTYH	MYC	MYCL (MYCL1)	MYCN	MYD88
NBN	NF1	NF2	NFE2L2	NFKB1A	NKX2-1	NOTCH1	NOTCH2	NOTCH3
NPM1	NRAS	NSD3 (WHSC1L1)	NT5C2	NTRK1	NTRK2	NTRK3	P2RY8	PALB2
PARK2	PARP1	PARP2	PARP3	PAX5	PBRM1	PDCD1 (PD-1)	PDCD1LG2 (PD-L2)	PDGFRA
PDGFRB	PDK1	PIK3C2B	PIK3C2G	PIK3CA	PIK3CB	PIK3R1	PIM1	PMS2
POLD1	POLE	PPARG	PPP2R1A	PPP2R2A	PRDM1	PRKAR1A	PRKCI	PTCH1
PTEN	PTPN11	PTPRO	QKI	RAC1	RAD21	RAD51	RAD51B	RAD51C
RAD51D	RAD52	RAD54L	RAF1	RARA	RB1	RBM10	REL	RET
RICTOR	RNF43	ROS1	RPTOR	SDHA	SDHB	SDHC	SDHD	SETD2
SF3B1	SGK1	SMAD2	SMAD4	SMARCA4	SMARCB1	SMO	SNCAIP	SOC3
SOX2	SOX9	SPEN	SPOP	SRC	STAG2	STAT3	STK11	SUFU
SYK	TBX3	TEK	TET2	TGFBR2	TIPARP	TNFAIP3	TNFRSF14	TP53
TSC1	TSC2	TYRO3	U2AF1	VEGFA	VHL	WHSC1	WT1	XPO1
XRCC2	ZNF217	ZNF703						

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

ALK	BCL2	BCR	BRAF	BRCA1	BRCA2	CD74	EGFR	ETV4
ETV5	ETV6	EWSR1	EZR	FGFR1	FGFR2	FGFR3	KIT	KMT2A (MLL)
MSH2	MYB	MYC	NOTCH2	NTRK1	NTRK2	NUTM1	PDGFRA	RAF1
RARA	RET	ROS1	RSP02	SDC4	SLC34A2	TERC*	TERT**	TMPS2

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score
Microsatellite (MS) status
Tumor Mutational Burden (TMB)

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About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a., Cipalstraat 3, 2440 Geel, Belgium.


ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Alterations and Therapies
Biomarker and Genomic Findings

Therapies are ranked based on the following criteria: Therapies with clinical benefit in patient's tumor type (ranked alphabetically within each NCCN category) followed by therapies with clinical benefit in other tumor type (ranked alphabetically within each NCCN category).

Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2021. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. The MSI-H/MSS designation by FMI F1CDx test is based on genome wide analysis of 95 microsatellite loci and not based on the 5 or 7 MSI loci described in current clinical practice guidelines. The threshold for MSI-H/MSS was determined by analytical concordance to comparator assays (IHC and PCR) using uterine, cecum and colorectal cancer FFPE tissue. The clinical validity of the qualitative MSI designation has not been established. For Microsatellite Instability (MSI) results, confirmatory testing using a validated orthogonal method should be considered.
2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit.

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Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.

3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine whether the patient is a candidate for biopsy.
6. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH

test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of HER2 overexpression as 20% in breast cancer. Based on the F1CDx HER2 CDx concordance study, approximately 10% of HER2 amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

BASE SUBSTITUTIONS	
Repeatability	5.11 - 10.40
Reproducibility	5.95 - 12.31
INDELS	
Repeatability	6.29 - 10.00
Reproducibility	7.33 - 11.71

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*,

MSH2, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides

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entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

ABBREVIATION	DEFINITION
CR	Complete response
DCR	Disease control rate
DNMT	DNA methyltransferase
HR	Hazard ratio
ITD	Internal tandem duplication
MMR	Mismatch repair
mut/Mb	Mutations per megabase
NOS	Not otherwise specified
ORR	Objective response rate
OS	Overall survival
PD	Progressive disease
PFS	Progression-free survival
PR	Partial response
SD	Stable disease
TKI	Tyrosine kinase inhibitor

MR Suite Version 4.1.0

The median exon coverage for this sample is 685x

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Electronically signed by Douglas A. Mata, MD, MPH | 15 June 2021
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Sample Preparation: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531
Post-Sequencing Analysis: 150 Second St., 1st Floor, Cambridge, MA 02141 · CLIA: 22D2027531

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References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Anbazhagan R, et al. Clin. Cancer Res. (1999) PMID: 10213220
7. Adem C, et al. Int. J. Cancer (2003) PMID: 14520695
8. Horimoto Y, et al. Cancer Sci (2020) PMID: 32449246
9. Heeke AL, et al. Breast Cancer Res Treat (2020) PMID: 32776290
10. Kurata K, et al. Breast Cancer (2020) PMID: 31907878
11. Sivapiragasam A, et al. Cancer Med (2020) PMID: 33314633
12. Walsh MD, et al. Clin. Cancer Res. (2010) PMID: 20215533
13. Risinger JI, et al. Cancer (1996) PMID: 8646682
14. de Leeuw WJ, et al. Cancer Res. (2003) PMID: 12615735
15. Shanley S, et al. Fam. Cancer (2009) PMID: 19123071
16. Buerki N, et al. Genes Chromosomes Cancer (2012) PMID: 22034109
17. Yee CJ, et al. Cancer Res. (1994) PMID: 8137273
18. Kamat N, et al. BMC Cancer (2012) PMID: 22928966
19. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
20. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
21. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
22. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
23. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
24. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
25. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
26. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
27. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
28. Cristescu R, et al. Science (2018) PMID: 30309915
29. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
30. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
31. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
32. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
33. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
34. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
35. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
36. Legrand et al., 2018; ASCO Abstract 12000
37. Barroso-Sousa R, et al. Ann. Oncol. (2020) PMID: 32067680
38. Nature (2012) PMID: 23000897
39. Sokol ES, et al. Ann. Oncol. (2019) PMID: 30423024
40. Chumsri S, et al. J Natl Compr Canc Netw (2020) PMID: 32380464
41. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
42. Haricharan S, et al. Breast Cancer Res. Treat. (2014) PMID: 24839032
43. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
44. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
45. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
46. Rizvi NA, et al. Science (2015) PMID: 25765070
47. Johnson BE, et al. Science (2014) PMID: 24336570
48. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
49. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
50. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
51. Heitzer E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
52. Nature (2012) PMID: 22810696
53. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
54. Fritsch C, et al. Mol. Cancer Ther. (2014) PMID: 24608574
55. Juric D, et al. J. Clin. Oncol. (2018) PMID: 29401002
56. Gallant JN, et al. NPJ Precis Oncol (2019) PMID: 30793038
57. André F, et al. N. Engl. J. Med. (2019) PMID: 31091374
58. Park HS, et al. PLoS ONE (2016) PMID: 27105424
59. Lim SM, et al. Oncotarget (2016) PMID: 26859683
60. Hou MM, et al. Oncotarget (2014) PMID: 25426553
61. Varnier R, et al. Eur J Cancer (2019) PMID: 31351267
62. Janku D, et al. Cell Rep (2014) PMID: 24440717
63. Moroney J, et al. Clin. Cancer Res. (2012) PMID: 22927482
64. Basho RK, et al. JAMA Oncol (2017) PMID: 27893038
65. Moroney JW, et al. Clin. Cancer Res. (2011) PMID: 21890452
66. Dolly SO, et al. Clin. Cancer Res. (2016) PMID: 26787751
67. Moynahan ME, et al. Br. J. Cancer (2017) PMID: 28183140
68. Hortobagyi GN, et al. J Clin Oncol (2016) PMID: 26503204
69. Baselga J, et al. Lancet Oncol (2017) PMID: 28576675
70. Campone M, et al. Eur. J. Cancer (2018) PMID: 30241001
71. Jhaveri et al., 2020; Abstract PS5-12
72. Bedard et al., 2020; SABCS Abstract PD1-02
73. Spathas et al., 2020; DOI: 10.1200/PO.19.00049
74. Santin AD, et al. Gynecol Oncol Rep (2020) PMID: 31934607
75. Patnaik A, et al. Ann. Oncol. (2016) PMID: 27672108
76. Rodon J, et al. Invest New Drugs (2014) PMID: 24652201
77. Bendell JC, et al. J. Clin. Oncol. (2012) PMID: 22162589
78. Heudel PE, et al. Br. J. Cancer (2017) PMID: 28072765
79. Vansteenkiste JF, et al. J Thorac Oncol (2015) PMID: 26098748
80. Baselga et al., 2018; ASCO Abstract LBA1006
81. Vasan N, et al. Science (2019) PMID: 31699932
82. Kim SB, et al. Lancet Oncol (2017) PMID: 28800861
83. Kim et al., 2020; SABCS Abstract GS3-04
84. Schmid P, et al. J. Clin. Oncol. (2019) PMID: 31841354
85. Banerji et al., 2015; ASCO Abstract 2500
86. Turner NC, et al. Ann. Oncol. (2019) PMID: 30860570
87. Esteva FJ, et al. Am. J. Pathol. (2010) PMID: 20813970
88. Baselga J, et al. J. Clin. Oncol. (2014) PMID: 25332247
89. Chakrabarty A, et al. Oncogene (2010) PMID: 20581867
90. Kataoka Y, et al. Ann. Oncol. (2010) PMID: 19633047
91. Wang L, et al. BMC Cancer (2011) PMID: 21676217
92. Juric D, et al. Nature (2015) PMID: 25409150
93. Hoste G, et al. Clin Drug Investig (2018) PMID: 30187361
94. Loi S, et al. PLoS ONE (2013) PMID: 23301057
95. Christgen M, et al. Genes Chromosomes Cancer (2013) PMID: 22997091
96. Ramirez-Ardila DE, et al. Breast Cancer Res. Treat. (2013) PMID: 23592373
97. Kalinsky K, et al. Clin. Cancer Res. (2009) PMID: 19671852
98. Mosele F, et al. Ann. Oncol. (2020) PMID: 32067679
99. Barbareschi M, et al. Clin. Cancer Res. (2007) PMID: 17947469
100. Samuels Y, et al. Cancer Cell (2005) PMID: 15950905
101. Nat. Rev. Cancer (2009) PMID: 19629070
102. Kang S, et al. Proc. Natl. Acad. Sci. U.S.A. (2005) PMID: 15647370
103. Ikenoue T, et al. Cancer Res. (2005) PMID: 15930273
104. Gymnopoulos M, et al. Proc. Natl. Acad. Sci. U.S.A. (2007) PMID: 17376864
105. Horn S, et al. Oncogene (2008) PMID: 18317450
106. Rudd ML, et al. Clin. Cancer Res. (2011) PMID: 21266528
107. Hon WC, et al. Oncogene (2012) PMID: 22120714
108. Burke JE, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22949682
109. Wu H, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19915146
110. Laurenti R, et al. Rev Saude Publica (1990) PMID: 2103068
111. Dan S, et al. Cancer Res. (2010) PMID: 20530683
112. Oda K, et al. Cancer Res. (2008) PMID: 18829572
113. Zhao L, et al. Oncogene (2008) PMID: 18794883
114. Lui VW, et al. Cancer Discov (2013) PMID: 23619167
115. Ross RL, et al. Oncogene (2013) PMID: 22430209
116. Rivière JB, et al. Nat. Genet. (2012) PMID: 22729224
117. Shibata T, et al. Cancer Lett. (2009) PMID: 19394761
118. Dogruluk T, et al. Cancer Res. (2015) PMID: 26627007
119. Croessmann S, et al. Clin. Cancer Res. (2018) PMID: 29284706
120. Ng PK, et al. Cancer Cell (2018) PMID: 29533785
121. Spangle JM, et al. (2020) PMID: 32929011
122. Chen L, et al. Nat Commun (2018) PMID: 29636477
123. Horiuchi D, et al. J. Exp. Med. (2012) PMID: 22430491
124. Goga A, et al. Nat. Med. (2007) PMID: 17589519
125. Molenaar JJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19525400
126. Dammert MA, et al. Nat Commun (2019) PMID: 31375684
127. Mollaoglu G, et al. Cancer Cell (2017) PMID: 28089889
128. Cardnell RJ, et al. Oncotarget (2017) PMID: 29088717
129. Wang L, et al. Mol Oncol (2017) PMID: 28417568
130. Takahashi Y, et al. Ann. Oncol. (2015) PMID: 25632068
131. Li Y, et al. Thyroid (2018) PMID: 30226440
132. Mahadevan D, et al. PLoS ONE (2014) PMID: 24893165
133. Park SI, et al. Target Oncol (2019) PMID: 31429028
134. Helfrich BA, et al. Mol. Cancer Ther. (2016) PMID: 27496133
135. Hook KE, et al. Mol. Cancer Ther. (2012) PMID: 22222631
136. Yang D, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20643922
137. He J, et al. Anticancer Drugs (2019) PMID: 30540594
138. Shroff EH, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25964345
139. Effenberger M, et al. Oncotarget (2017) PMID: 29156762
140. Qu X, et al. Biochem. Biophys. Res. Commun. (2018) PMID: 30103944
141. Xiang Y, et al. J. Clin. Invest. (2015) PMID: 25915584
142. Delmore JE, et al. Cell (2011) PMID: 21889194
143. Bandopadhyay P, et al. Clin. Cancer Res. (2014) PMID: 24297863
144. Lovén J, et al. Cell (2013) PMID: 23582323
145. Otto C, et al. Neoplasia (2019) PMID: 31734632
146. Dong LH, et al. J Hematol Oncol (2013) PMID: 23866964
147. Pei Y, et al. Cancer Cell (2016) PMID: 26977882
148. Fu XH, et al. Acta Pharmacol. Sin. (2019) PMID: 30224636
149. Owonikoko TK, et al. J Thorac Oncol (2020) PMID: 31655296
150. Ganesan P, et al. Mol. Cancer Ther. (2014) PMID:

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ORDERED TEST # **ORD-1110066-01**
APPENDIX
References

- 25253784
151. Tannir et al., 2018; ASCO GU Abstract 603
152. Motzer et al., 2019; ESMO Abstract LBA54
153. Pereira CB, et al. PLoS ONE (2013) pmid: 23555992
154. Yasojima H, et al. Eur. J. Cancer (2011) pmid: 21741827
155. Arango D, et al. Cancer Res. (2001) pmid: 11406570
156. Bottone MG, et al. Exp. Cell Res. (2003) pmid: 14516787
157. Chen Y, et al. Expert Rev Anticancer Ther (2008) pmid: 18925859
158. Xu J, et al. Genes Cancer (2010) pmid: 21779462
159. Colak D, et al. PLoS ONE (2013) pmid: 23704896
160. Dang CV, et al. Semin. Cancer Biol. (2006) pmid: 16904903
161. Nesbit CE, et al. Oncogene (1999) pmid: 10378696
162. Blancato J, et al. Br. J. Cancer (2004) pmid: 15083194
163. Fromont G, et al. Hum. Pathol. (2013) pmid: 23574779
164. Templeton AJ, et al. Eur. Urol. (2013) pmid: 23582881
165. Courtney KD, et al. J. Clin. Oncol. (2010) pmid: 20085938
166. Wu R, et al. Clin. Cancer Res. (2011) pmid: 21903772
167. Simpson L, et al. Exp. Cell Res. (2001) pmid: 11237521
168. Dreyling M, et al. Ann. Oncol. (2017) pmid: 28633365
169. Hsieh JJ, et al. Eur. Urol. (2017) pmid: 27751729
170. Roldan-Romero JM, et al. Int J Cancer (2020) pmid: 31335987
171. Kwiatkowski DJ, et al. Clin. Cancer Res. (2016) pmid: 26831717
172. Figlin RA, et al. Cancer (2009) pmid: 19526589
173. Cho D, et al. Clin Genitourin Cancer (2007) pmid: 17956710
174. Galanis E, et al. J. Clin. Oncol. (2005) pmid: 15998902
175. Cloughesy TF, et al. PLoS Med. (2008) pmid: 18215105
176. Tinker AV, et al. Gynecol. Oncol. (2013) pmid: 23672928
177. Ellard SL, et al. J Clin Oncol (2009) pmid: 19687332
178. Fleming GF, et al. Breast Cancer Res. Treat. (2012) pmid: 22245973
179. Meyer LA, et al. Int J Gynecol Cancer (2014) pmid: 24651628
180. Oza AM, et al. J. Clin. Oncol. (2011) pmid: 21788564
181. Mackay HJ, et al. Cancer (2014) pmid: 24166148
182. Fleming GF, et al. Gynecol. Oncol. (2014) pmid: 24456823
183. Tsoref D, et al. Gynecol. Oncol. (2014) pmid: 25173583
184. Myers AP, et al. Gynecol. Oncol. (2016) pmid: 27016228
185. Seront E, et al. Ann. Oncol. (2012) pmid: 22473592
186. Seront E, et al. Br. J. Cancer (2013) pmid: 23989949
187. Milowsky MI, et al. BJU Int. (2013) pmid: 23551593
188. Italiano A, et al. Anticancer Drugs (2011) pmid: 21301319
189. Khushman et al., 2015; ASCO GI Abstract 333
190. Voss MH, et al. Clin. Cancer Res. (2018) pmid: 30327302
191. Kim SJ, et al. Ann. Oncol. (2016) pmid: 26861608
192. Wee S, et al. Proc. Natl. Acad. Sci. U.S.A. (2008) pmid: 18755892
193. Jia S, et al. Nature (2008) pmid: 18594509
194. Schmit F, et al. Proc. Natl. Acad. Sci. U.S.A. (2014) pmid: 24737887
195. Janku et al., 2018; ESMO Abstract 418PD
196. Dent et al., 2018; ASCO Abstract 1008
197. Schmid et al., 2018; ASCO Abstract 1007
198. de Bono JS, et al. Clin. Cancer Res. (2019) pmid: 30037818
199. Saura C, et al. Cancer Discov (2017) pmid: 27872130
200. Mendes-Pereira AM, et al. EMBO Mol Med (2009) pmid: 20049735
201. Shen Y, et al. Clin. Cancer Res. (2013) pmid: 23881923
202. Chatterjee P, et al. PLoS ONE (2013) pmid: 23565244
203. McCormick A, et al. Int. J. Gynecol. Cancer (2016) pmid: 26905328
204. Forster MD, et al. Nat Rev Clin Oncol (2011) pmid: 21468130
205. Gruber et al., 2019; ASCO Abstract 3006
206. Dougherty et al., 2014; ASCO Abstract 5536
207. Piha-Paul et al., 2018; AACR Abstract A096
208. Ihnen M, et al. Mol. Cancer Ther. (2013) pmid: 23729402
209. Sandhu SK, et al. Lancet Oncol. (2013) pmid: 23810788
210. Zhao J, et al. Nat. Med. (2019) pmid: 30742119
211. George S, et al. Immunity (2017) pmid: 28228279
212. Parikh AR, et al. Lung Cancer (Auckl) (2018) pmid: 29844707
213. Peng W, et al. Cancer Discov (2016) pmid: 26645196
214. Costa C, et al. Cancer Discov (2019) pmid: 31594766
215. Le X, et al. Cancer Discov (2016) pmid: 27604488
216. Chen SH, et al. Oncogene (2018) pmid: 29059158
217. Hennessy BT, et al. Cancer Res. (2009) pmid: 19435916
218. Mercapide J, et al. Mol. Carcinog. (2002) pmid: 12203362
219. Hohensee I, et al. Am. J. Pathol. (2013) pmid: 23665199
220. Perez EA, et al. J. Clin. Oncol. (2013) pmid: 23650412
221. Tsutsui S, et al. Oncology (2005) pmid: 16020969
222. Zhang HY, et al. Oncol Lett (2013) pmid: 23946797
223. Capodanno A, et al. Hum. Pathol. (2009) pmid: 19428048
224. Campbell RB, et al. J. Biol. Chem. (2003) pmid: 12857747
225. Rodriguez-Escudero I, et al. Hum. Mol. Genet. (2011) pmid: 21828076
226. He X, et al. Cancer Res. (2013) pmid: 23475934
227. Han SY, et al. Cancer Res. (2000) pmid: 10866302
228. Myers MP, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) pmid: 9811831
229. Pradella LM, et al. BMC Cancer (2014) pmid: 24498881
230. Kim JS, et al. Mol. Cell. Biol. (2011) pmid: 21536651
231. Denning G, et al. Oncogene (2007) pmid: 17213812
232. Hlobilkova A, et al. Anticancer Res. (2016) pmid: 16619501
233. Redfern RE, et al. Protein Sci. (2010) pmid: 20718038
234. Shenoy S, et al. PLoS ONE (2012) pmid: 22505997
235. Wang Y, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19329485
236. Okumura K, et al. J. Biol. Chem. (2006) pmid: 16829519
237. Lee JO, et al. Cell (1999) pmid: 10555148
238. Maxwell GL, et al. Cancer Res. (1998) pmid: 9635567
239. Risinger JJ, et al. Clin. Cancer Res. (1998) pmid: 9865913
240. Kato H, et al. Clin. Cancer Res. (2000) pmid: 11051241
241. Fenton TR, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) pmid: 22891331
242. Ngeow J, et al. J. Clin. Endocrinol. Metab. (2012) pmid: 23066114
243. Lobo GP, et al. Hum. Mol. Genet. (2009) pmid: 19457929
244. Liu J, et al. Oncogene (2014) pmid: 23995781
245. Maehama T, et al. Annu. Rev. Biochem. (2001) pmid: 11395408
246. De Vivo I, et al. J. Med. Genet. (2000) pmid: 10807691
247. Ramaswamy S, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) pmid: 10051603
248. Liu JL, et al. Mol. Cell. Biol. (2005) pmid: 15988030
249. Karoui M, et al. Br. J. Cancer (2004) pmid: 15026806
250. Gil A, et al. PLoS ONE (2015) pmid: 25875300
251. Furnari FB, et al. Cancer Res. (1998) pmid: 9823298
252. Spinelli L, et al. J. Med. Genet. (2015) pmid: 25527629
253. Mingo J, et al. Eur. J. Hum. Genet. (2018) pmid: 29706633
254. Wang Q, et al. J. Mol. Graph. Model. (2010) pmid: 20538496
255. Andrés-Pons A, et al. Cancer Res. (2007) pmid: 17942903
256. Butler MG, et al. J. Med. Genet. (2005) pmid: 15805158
257. Georgescu MM, et al. Proc. Natl. Acad. Sci. U.S.A. (1999) pmid: 10468583
258. Staal FJ, et al. Br. J. Cancer (2002) pmid: 12085208
259. Nguyen HN, et al. Oncogene (2014) pmid: 24292679
260. Rahdar M, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) pmid: 19114656
261. Das S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12808147
262. Wang X, et al. Biochem. J. (2008) pmid: 18498243
263. Valiente M, et al. J. Biol. Chem. (2005) pmid: 15951562
264. Nguyen HN, et al. Oncogene (2015) pmid: 25263454
265. Blumenthal GM, et al. Eur. J. Hum. Genet. (2008) pmid: 18781191
266. Orloff MS, et al. Oncogene (2008) pmid: 18794875
267. Zbuk KM, et al. Nat. Rev. Cancer (2007) pmid: 17167516
268. Nature (2012) pmid: 22960745
269. Ciriello G, et al. Cell (2015) pmid: 26451490
270. Nature (2014) pmid: 24476821
271. Nature (2015) pmid: 25631445
272. Cerami E, et al. Cancer Discov (2012) pmid: 22588877
273. Gao J, et al. Sci Signal (2013) pmid: 23550210
274. Jones DH, et al. Mol Clin Oncol (2017) pmid: 28781807
275. Kim SM, et al. Biochem. Biophys. Res. Commun. (2006) pmid: 16682010
276. Kang D, et al. Genes Chromosomes Cancer (2013) pmid: 23011637
277. Chen Y, et al. PLoS ONE (2014) pmid: 24874471
278. Morishita M, et al. Biochim. Biophys. Acta (2011) pmid: 21664949
279. Varela I, et al. Nature (2011) pmid: 21248752
280. Mar BG, et al. Nat Commun (2014) pmid: 24662245
281. Wang Q, et al. Sci China Life Sci (2014) pmid: 25077743
282. Zhu X, et al. Nat. Genet. (2014) pmid: 24509477
283. Sun XJ, et al. J. Biol. Chem. (2005) pmid: 16118227
284. Faber PW, et al. Hum. Mol. Genet. (1998) pmid: 9700202
285. Al Sarakbi W, et al. BMC Cancer (2009) pmid: 19698110
286. Ito S, et al. Nature (2010) pmid: 20639862
287. Guo JU, et al. Cell (2011) pmid: 21496894
288. Iyer LM, et al. Cell Cycle (2009) pmid: 19411852
289. Ko M, et al. Nature (2010) pmid: 21057493
290. Yang H, et al. Oncogene (2013) pmid: 22391558
291. Hu L, et al. Cell (2013) pmid: 24315485
292. Wang Y, et al. Mol. Cell (2015) pmid: 25601757
293. Jaiswal S, et al. N. Engl. J. Med. (2014) pmid: 25426837
294. Genovesi G, et al. N. Engl. J. Med. (2014) pmid: 25426838
295. Xie M, et al. Nat. Med. (2014) pmid: 25326804
296. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) pmid: 28669404
297. Severson EA, et al. Blood (2018) pmid: 29678827
298. Fuster JJ, et al. Circ. Res. (2018) pmid: 29420212
299. Hematology Am Soc Hematol Educ Program (2018) pmid: 30504320
300. Chabon JJ, et al. Nature (2020) pmid: 32269342
301. Razavi P, et al. Nat. Med. (2019) pmid: 31768066
302. Hirai H, et al. Cancer Biol. Ther. (2010) pmid: 20107315
303. Bridges KA, et al. Clin. Cancer Res. (2011) pmid: 21799033

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ORDERED TEST # **ORD-1110066-01**
APPENDIX
References

304. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) pmid: 21389100
305. Osman AA, et al. Mol. Cancer Ther. (2015) pmid: 25504633
306. Xu L, et al. Mol. Cancer Ther. (2002) pmid: 12489850
307. Xu L, et al. Mol. Med. (2001) pmid: 11713371
308. Camp ER, et al. Cancer Gene Ther. (2013) pmid: 23470564
309. Kim SS, et al. Nanomedicine (2015) pmid: 25240597
310. Pirolo KF, et al. Mol. Ther. (2016) pmid: 27357628
311. Hajdenberg et al., 2012; ASCO Abstract e15010
312. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27601554
313. Moore et al., 2019; ASCO Abstract 5513
314. Leijen S, et al. J. Clin. Oncol. (2016) pmid: 27998224
315. Oza et al., 2015; ASCO Abstract 5506
316. Lee J, et al. Cancer Discov (2019) pmid: 31315834
317. Méndez E, et al. Clin. Cancer Res. (2018) pmid: 29535125
318. Ma CX, et al. J. Clin. Invest. (2012) pmid: 22446188
319. Kwok M, et al. Blood (2016) pmid: 26563132
320. Boudny M, et al. Haematologica (2019) pmid: 30975914
321. Dillon MT, et al. Mol. Cancer Ther. (2017) pmid: 28062704
322. Middleton FK, et al. Cancers (Basel) (2018) pmid: 30127241
323. Banerji S, et al. Nature (2012) pmid: 22722202
324. Stephens PJ, et al. Nature (2012) pmid: 22722201
325. Alsner J, et al. Acta Oncol (2008) pmid: 18465328
326. Alkam Y, et al. Histopathology (2013) pmid: 24004112
327. Uji K, et al. Cancer Lett. (2014) pmid: 23973262
328. Olivier M, et al. Clin. Cancer Res. (2006) pmid: 16489069
329. Végran F, et al. PLoS ONE (2013) pmid: 23359294
330. Walsh T, et al. JAMA (2006) pmid: 16551709
331. Garber JE, et al. J. Clin. Oncol. (2005) pmid: 15637391
332. Apostolou P, et al. Biomed Res Int (2013) pmid: 23586058
333. Brown CJ, et al. Nat. Rev. Cancer (2009) pmid: 19935675
334. Joerger AC, et al. Annu. Rev. Biochem. (2008) pmid: 18410249
335. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) pmid: 12826609
336. Kamada R, et al. J. Biol. Chem. (2011) pmid: 20978130
337. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) pmid: 28472496
338. Yamada H, et al. Carcinogenesis (2007) pmid: 17690113
339. Landrum MJ, et al. Nucleic Acids Res. (2018) pmid: 29165669
340. Bougeard G, et al. J. Clin. Oncol. (2015) pmid: 26014290
341. Sorrell AD, et al. Mol Diagn Ther (2013) pmid: 23355100
342. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) pmid: 11219776
343. Kleihues P, et al. Am. J. Pathol. (1997) pmid: 9006316
344. Gonzalez KD, et al. J. Clin. Oncol. (2009) pmid: 19204208
345. Lalloo F, et al. Lancet (2003) pmid: 12672316
346. Mandelker D, et al. Ann. Oncol. (2019) pmid: 31050713
347. Zhang X, et al. PLoS ONE (2013) pmid: 23991038
348. Holland DG, et al. EMBO Mol Med (2011) pmid: 21337521
349. Sircoulomb F, et al. EMBO Mol Med (2011) pmid: 21328542
350. Reynisdottir I, et al. Cancer Med (2013) pmid: 24156016
351. Yang G, et al. Oncol. Rep. (2014) pmid: 24481460
352. Ma F, et al. Oncol. Rep. (2014) pmid: 25017610
353. Bazarov AV, et al. Breast Cancer Res. (2011) pmid: 21635707
354. Slorach EM, et al. Genes Dev. (2011) pmid: 21317240
355. Juric D, et al. JAMA Oncol (2018) pmid: 30543347
356. Andre et al., 2020; ESMO Abstract LBA18
357. Batalini F, et al. JCO Precis Oncol (2020) pmid: 32923889
358. Rugo et al., 2020; ASCO Abstract 1006
359. Rugo et al., 2020; SABCS Abstract PD2-07
360. Janku F, et al. Cancer Res. (2013) pmid: 23066039
361. Janku F, et al. J. Clin. Oncol. (2012) pmid: 22271473
362. Janku F, et al. Mol. Cancer Ther. (2011) pmid: 21216929
363. Moulder S, et al. Ann. Oncol. (2015) pmid: 25878190
364. Byeon et al., 2020; doi: 10.21037/tcr.2020.04.07
365. Yuan Y, et al. Oncotarget (2017) pmid: 28061482
366. Park IH, et al. J Cancer (2018) pmid: 29675095
367. Kruger DT, et al. Mol Oncol (2020) pmid: 31841262
368. Bachelot et al., 2019; ASCO Abstract 1024
369. Beck JT, et al. Breast Cancer Res. Treat. (2014) pmid: 24362951
370. Yardley DA, et al. Adv Ther (2013) pmid: 24158787
371. Baselga J, et al. N. Engl. J. Med. (2012) pmid: 22149876
372. Piccart M, et al. Ann. Oncol. (2014) pmid: 25231953
373. Jerusalem G, et al. JAMA Oncol (2018) pmid: 29862411
374. Baselga J, et al. J. Clin. Oncol. (2009) pmid: 19380449
375. Bachelot T, et al. J. Clin. Oncol. (2012) pmid: 22565002
376. Wheler JJ, et al. Oncotarget (2014) pmid: 24912489
377. Yardley DA, et al. Clin. Breast Cancer (2019) pmid: 31932237
378. Hurvitz SA, et al. Lancet Oncol. (2015) pmid: 26092818
379. André F, et al. Lancet Oncol. (2014) pmid: 24742739
380. Decker T, et al. Breast Cancer Res Treat (2019) pmid: 31115844
381. Singh J, et al. Breast Cancer Res. (2014) pmid: 24684785
382. Tolcher AW, et al. Ann. Oncol. (2015) pmid: 25344362
383. Patterson et al., 2018; AACR Abstract 3891
384. Basho RK, et al. Oncologist (2018) pmid: 30139837
385. Moulder S, et al. J Clin Oncol (2011) pmid: 21482991
386. Agarwal R, et al. J Breast Cancer (2014) pmid: 25320628
387. Wolff AC, et al. J. Clin. Oncol. (2013) pmid: 23233719