

ABOUT THE TEST FoundationOne®CDx is a next-generation sequencing (NGS) based assay that identifies genomic findings within hundreds of cancer-related genes.

PATIENT

DISEASE Lung non-small cell lung carcinoma (NOS)
NAME Bravo Villanueva, Adolfo
DATE OF BIRTH 01 April 1946
SEX Male
MEDICAL RECORD # Not given

PHYSICIAN

MEDICAL FACILITY Arias Stella
ADDITIONAL RECIPIENT None
MEDICAL FACILITY ID 317319
PATHOLOGIST Not Provided

SPECIMEN

SPECIMEN SITE Thorax
SPECIMEN ID 22QX-523
SPECIMEN TYPE Block
DATE OF COLLECTION 03 February 2022
SPECIMEN RECEIVED 18 February 2022

Biomarker Findings

Microsatellite status - MS-Stable

Tumor Mutational Burden - 4 Muts/Mb

Genomic Findings

For a complete list of the genes assayed, please refer to the Appendix.

EGFR exon 19 deletion (E746_A750del)

MET amplification - equivocal[†]

CCNE1 amplification

MYC amplification

RNF43 R145*

RAD21 amplification

TP53 P151T

ZNF217 amplification

6 Disease relevant genes with no reportable alterations: **ALK, BRAF, ERBB2, KRAS, RET, ROS1**

[†] See About the Test in appendix for details.

Report Highlights

- Targeted therapies with **NCCN categories of evidence** in this tumor type: **Afatinib** (p. 10), **Dacomitinib** (p. 12), **Capmatinib** (p. 11), **Crizotinib** (p. 11), **Tepotinib** (p. 12)
- Targeted therapies with **potential resistance** based on this patient's genomic findings: **✖ Erlotinib** (p. 13), **Gefitinib** (p. 14), **Osimertinib** (p. 15)
- Evidence-matched **clinical trial options** based on this patient's genomic findings: (p. 17)

BIOMARKER FINDINGS

Microsatellite status - MS-Stable

Tumor Mutational Burden - 4 Muts/Mb

THERAPY AND CLINICAL TRIAL IMPLICATIONS

No therapies or clinical trials. see Biomarker Findings section

No therapies or clinical trials. see Biomarker Findings section

| GENOMIC FINDINGS | THERAPIES WITH CLINICAL RELEVANCE (IN PATIENT'S TUMOR TYPE) | THERAPIES WITH CLINICAL RELEVANCE (IN OTHER TUMOR TYPE) |
|---|---|---|
| EGFR - exon 19 deletion (E746_A750del) | Afatinib <input type="text" value="1"/> | none |
| | Dacomitinib <input type="text" value="1"/> | |
| | Erlotinib <input checked="" type="checkbox"/> | |
| | Gefitinib <input checked="" type="checkbox"/> | |
| | Osimertinib <input checked="" type="checkbox"/> | |
| 10 Trials see p. 18 | | |
| MET - amplification - equivocal | Capmatinib <input type="text" value="2A"/> | Cabozantinib |
| | Crizotinib <input type="text" value="2A"/> | |
| | Tepotinib <input type="text" value="2A"/> | |
| | Erlotinib <input checked="" type="checkbox"/> | |
| | Gefitinib <input checked="" type="checkbox"/> | |
| | Osimertinib <input checked="" type="checkbox"/> | |
| 10 Trials see p. 20 | | |
| CCNE1 - amplification | none | none |
| 3 Trials see p. 17 | | |
| MYC - amplification | none | none |
| 4 Trials see p. 22 | | |
| RNF43 - R145* | none | none |
| 3 Trials see p. 23 | | |

☒ Extensive evidence showing variant(s) in this sample may confer resistance to this therapy
 ☐ NCCN category

GENOMIC FINDINGS WITH NO REPORTABLE THERAPEUTIC OR CLINICAL TRIAL OPTIONS

For more information regarding biological and clinical significance, including prognostic, diagnostic, germline, and potential chemosensitivity implications, see the Genomic Findings section.

RAD21 - amplification p. 7 **ZNF217** - amplification p. 9
TP53 - P151T p. 8

NOTE Genomic alterations detected may be associated with activity of certain approved therapies; however, the agents listed in this report may have varied clinical evidence in the patient's tumor type. Therapies and the clinical trials listed in this report may not be complete and exhaustive. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type. This report should be regarded and used as a supplementary source of information and not as the single basis for the making of a therapy decision. All treatment decisions remain the full and final responsibility of the treating physician and physicians should refer to approved prescribing information for all therapies.

Therapies contained in this report may have been approved by the US FDA.

ORDERED TEST # ORD-1306820-01

BIOMARKER FINDINGS

BIOMARKER

Microsatellite status

RESULT

MS-Stable

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence, MSS tumors are significantly less likely than MSI-H tumors to respond to anti-PD-1 immune checkpoint inhibitors¹⁻³, including approved therapies nivolumab and pembrolizumab⁴. In a retrospective analysis of 361 patients with solid tumors treated with pembrolizumab, 3% were MSI-H and

experienced a significantly higher ORR compared with non-MSI-H cases (70% vs. 12%, $p=0.001$)⁵.

FREQUENCY & PROGNOSIS

MSI-H is generally infrequent in NSCLC, reported in fewer than 1% of samples across several large studies⁶⁻¹¹, whereas data on the reported incidence of MSI-H in SCLC has been limited and conflicting¹²⁻¹⁵. One study reported MSI-H in lung adenocarcinoma patients with smoking history, and 3 of 4 MSI-H patients examined also had metachronous carcinomas in other organs, although this has not been investigated in large scale studies⁶. Published data investigating the prognostic implications of MSI in NSCLC are limited (PubMed, Oct 2021).

FINDING SUMMARY

Microsatellite instability (MSI) is a condition of genetic hypermutability that generates excessive amounts of short insertion/deletion mutations in the genome; it generally occurs at microsatellite DNA sequences and is caused by a deficiency in DNA mismatch repair (MMR) in the tumor¹⁶. Defective MMR and consequent MSI occur as a result of genetic or epigenetic inactivation of one of the MMR pathway proteins, primarily MLH1, MSH2, MSH6, or PMS2¹⁶⁻¹⁸. This sample is microsatellite-stable (MSS), equivalent to the clinical definition of an MSS tumor: one with mutations in none of the tested microsatellite markers¹⁹⁻²¹. MSS status indicates MMR proficiency and typically correlates with intact expression of all MMR family proteins^{16,18,20-21}.

BIOMARKER

Tumor Mutational Burden

RESULT

4 Muts/Mb

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of clinical evidence in solid tumors, increased TMB may be associated with greater sensitivity to immunotherapeutic agents, including anti-PD-L1²²⁻²⁴, anti-PD-1 therapies²²⁻²⁵, and combination nivolumab and ipilimumab²⁶⁻³¹. Multiple clinical trials of PD-1- or PD-L1-targeting immune checkpoint inhibitors or combination of PD-1 and CTLA-4 inhibitors in NSCLC have reported that patients with tumors harboring TMB ≥ 10 Muts/Mb derive greater clinical benefit from these therapies than those with TMB < 10 Muts/Mb (based on this assay or others); similarly, higher efficacy of anti-PD-1 or anti-PD-L1 immunotherapy for treatment of patients with NSCLC, compared with the use of chemotherapy, has been observed more significantly in cases of TMB ≥ 10 Muts/Mb (based on this assay or others)^{22-23,26-28,32-39}. Improved OS of patients with

NSCLC treated with pembrolizumab plus chemotherapy relative to chemotherapy only⁴⁰, or those treated with nivolumab plus ipilimumab also relative to chemotherapy⁴¹, has been observed across all TMB levels.

FREQUENCY & PROGNOSIS

A large-scale genomic analysis found that unspecified lung non-small cell lung carcinoma (NSCLC), lung adenocarcinoma, and lung squamous cell carcinoma (SCC) samples harbored median TMBs between 6.3 and 9 Muts/Mb, and 12% to 17% of cases had an elevated TMB of greater than 20 Muts/Mb⁴². Lower TMB is observed more commonly in NSCLCs harboring known driver mutations (EGFR, ALK, ROS1, or MET) with the exception of BRAF or KRAS mutations, which are commonly observed in elevated TMB cases⁴³. Although some studies have reported a lack of association between smoking and mutational burden in NSCLC⁴⁴⁻⁴⁵, several other large studies did find a strong association with increased TMB⁴⁶⁻⁴⁹. TMB > 10 muts/Mb was found to be more frequent in NSCLC metastases compared with primary tumors for both adenocarcinoma (38% vs. 25%) and SCC (41% vs. 35%) subtypes⁵⁰. A large study of Chinese patients with lung adenocarcinoma reported a shorter median OS for tumors with a higher number of mutations in a limited gene set compared with a

lower mutation number (48.4 vs. 61.0 months)⁴⁴. Another study of patients with NSCLC correlated elevated TMB with poorer prognosis and significantly associated lower TMB in combination with PD-L1 negative status with longer median survival in patients with lung adenocarcinoma⁵¹. However, no significant prognostic association of TMB and/or PD-L1 status with survival has been reported in patients with lung SCC⁵¹⁻⁵².

FINDING SUMMARY

Tumor mutation burden (TMB, also known as mutation load) is a measure of the number of somatic protein-coding base substitution and insertion/deletion mutations occurring in a tumor specimen. TMB is affected by a variety of causes, including exposure to mutagens such as ultraviolet light in melanoma⁵³⁻⁵⁴ and cigarette smoke in lung cancer^{32,55}, treatment with temozolomide-based chemotherapy in glioma⁵⁶⁻⁵⁷, mutations in the proofreading domains of DNA polymerases encoded by the POLE and POLD1 genes⁵⁸⁻⁶², and microsatellite instability (MSI)^{58,61-62}. This sample harbors a TMB below levels that would be predicted to be associated with sensitivity to PD-1- or PD-L1-targeting immune checkpoint inhibitors, alone or in combination with other agents^{22-23,26-28,32-39,63}.

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

EGFR

ALTERATION

exon 19 deletion (E746_A750del)

TRANSCRIPT ID

NM_005228

CODING SEQUENCE EFFECT

2235_2249delGGAATTAAGAGAAGC

VARIANT ALLELE FREQUENCY (% VAF)

33.5%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

For patients with non-small cell lung cancer, EGFR activating mutations may predict sensitivity to EGFR TKIs, including erlotinib⁶⁴, gefitinib⁶⁵, afatinib⁶⁶, dacomitinib⁶⁷, and osimertinib⁶⁸; however, the data for patients with other tumor types are limited⁶⁹⁻⁷⁴. The Phase 1 CHRYSALIS study of amivantamab monotherapy or in combination with lazertinib for the treatment of EGFR-mutated non-small cell lung cancer (NSCLC) has produced encouraging preliminary results for treatment-naïve patients and patients who relapsed after treatment with osimertinib with and without chemotherapy, including osimertinib-relapsed patients with biomarkers indicating EGFR/MET-based osimertinib resistance⁷⁵⁻⁷⁸. In a Phase 1 trial, the HER3-targeted antibody patritumab deruxtecan elicited an ORR of 39% (22/57, 1 CR) and a median PFS of 8.2 months for patients with non-small cell lung cancer previously treated with an EGFR TKI, many of whom displayed TKI resistance alterations⁷⁹. A Phase 1 trial evaluating the EGFR inhibitor AZD3759 reported a reduction in the volume of brain metastases in 40% (8/20) of

patients with previously treated non-small cell lung cancer (NSCLC) harboring either the EGFR L858R alteration or EGFR exon 19 deletion, including 3 confirmed PRs and 3 unconfirmed PRs⁸⁰⁻⁸¹. In a Phase 1/2 trial for advanced NSCLC, the brain-penetrant third-generation EGFR TKI lazertinib enabled ORRs of 54% (69/127) for all evaluable patients and 44% (8/18, intracranial) for patients with brain metastases⁸².

— Potential Resistance —

Patients with NSCLC harboring EGFR mutation and MET amplification are unlikely to respond to first- and third-generation EGFR inhibitors (NCCN NSCLC Guidelines, v2.2021). A retrospective study reported significantly shorter PFS and OS on gefitinib treatment for patients with EGFR-mutated non-small cell lung cancer (NSCLC) additionally harboring MET amplification than for those with EGFR-mutated tumors without MET amplification⁸³. Additionally, numerous clinical studies have reported MET amplification as an emergent alteration associated with acquired resistance of EGFR-mutated NSCLC to first-generation EGFR inhibitors such as erlotinib and gefitinib⁸⁴⁻¹⁰⁰ and third-generation EGFR inhibitors such as osimertinib^{91,101-109}. For a small number of patients with EGFR-mutated NSCLC, studies have also reported MET amplification in association with resistance to second-generation EGFR inhibitors such as afatinib and dacomitinib^{97,110-113}. Patients with NSCLC harboring EGFR mutation and MET amplification may benefit from combination treatment with MET- and EGFR-targeting agents^{87,90,104,106,112,114-116}.

— Nontargeted Approaches —

Patients with EGFR-mutated non-squamous metastatic non-small cell lung cancer previously

treated with EGFR TKI have benefited from immune checkpoint inhibitors combined with anti-angiogenic and chemotherapy, particularly atezolizumab plus bevacizumab plus carboplatin and paclitaxel (OS HR 0.61 compared with bevacizumab/chemotherapy)¹¹⁷⁻¹¹⁹ or sintilimab plus bevacizumab biosimilar plus cisplatin and pemetrexed (PFS HR 0.46 compared with chemotherapy alone)¹²⁰.

FREQUENCY & PROGNOSIS

EGFR mutation has been reported in 12-36% of lung adenocarcinomas^{48,121-122} and in 4% of lung squamous cell carcinomas¹²³. EGFR protein expression/overexpression has been reported in up to 70% of NSCLC cases¹²⁴⁻¹²⁹. In addition, expression of EGFR protein has been shown to be higher in lung squamous cell carcinoma samples as compared to lung adenocarcinoma¹³⁰⁻¹³¹. In patients with lung adenocarcinoma, EGFR mutation was a predictor of poor overall survival¹³²⁻¹³³. However, EGFR mutations have been reported to predict improved survival in patients with resected Stage 1-3 lung adenocarcinoma¹³⁴ or resected Stage 1 NSCLC¹³⁵.

FINDING SUMMARY

EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide¹³⁶. EGFR exon 19 deletion mutations, such as seen here, have been shown to activate the tyrosine kinase activity of EGFR and to confer sensitivity to EGFR tyrosine kinase inhibitors such as erlotinib, gefitinib¹³⁷⁻¹³⁹, afatinib¹⁴⁰, osimertinib¹⁴¹, and dacomitinib^{67,142}, although limited preclinical data suggest reduced sensitivity to lapatinib¹⁴³⁻¹⁴⁴.

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

MET

ALTERATION

amplification - equivocal

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

On the basis of extensive clinical evidence, MET amplification or activating mutations may predict sensitivity to MET-targeting therapies such as kinase inhibitors crizotinib, capmatinib, tepotinib, and cabozantinib. Crizotinib has benefited patients with MET-amplified non-small cell lung cancer (NSCLC) of varied histologies¹⁴⁵⁻¹⁴⁸, gastroesophageal cancer¹⁴⁹, glioblastoma¹⁵⁰, and carcinoma of unknown primary¹⁵¹. Capmatinib has demonstrated clinical efficacy for patients with MET-amplified NSCLC both as a monotherapy¹⁵²⁻¹⁵³ and in combination with an EGFR-TKI for patients with concurrent activating EGFR mutations^{114,154-155}. Tepotinib has demonstrated efficacy for patients with MET-amplified hepatocellular carcinoma¹⁵⁶ and NSCLC¹⁵⁷ as a monotherapy, as well as in combination with gefitinib for patients with MET-amplified and EGFR-mutated NSCLC¹⁵⁸⁻¹⁶⁰. Savolitinib elicited responses in patients with MET-amplified papillary renal cell carcinoma¹⁶¹ and gastric cancer either alone or in combination with docetaxel¹⁶²⁻¹⁶³. AMG 337 elicited an ORR of 50% (5/10), including 1 CR, for patients with MET-amplified gastric, esophageal, or gastroesophageal junction cancer¹⁶⁴. Patients with MET-amplified NSCLC¹⁶⁵ or MET-amplified gastric cancer¹⁶⁶ treated with the MET-targeting antibody onartuzumab (MetMab) achieved clinical responses. In addition, high MET expression has been suggested to predict patient response to therapies such as the monoclonal HGF-targeting antibody rilotumumab, as well as the combination of ramucirumab and the monoclonal MET-targeting antibody emibetuzumab¹⁶⁷. A first-in-human Phase 1 trial of telisotuzumab vedotin (teliso-V), a MET

antibody-drug conjugate, reported activity in a subset of patients with MET-positive NSCLC, with an ORR of 19% (3/16) and a DCR of 56% (9/16); no responses were observed in any other patients¹⁶⁸. A subsequent Phase 2 trial of teliso-V in patients with MET-positive NSCLC reported a 35% (13/37) ORR in patients with non-squamous, EGFR-wildtype tumors, which met the prespecified criteria for transition to the next stage; lower ORRs were observed in patients with squamous (14%; 3/21) or non-squamous EGFR-mutated (13%; 4/30) tumors¹⁶⁹. A Phase 1 study for patients with MET-altered NSCLC treated with MET inhibitor bozitinib monotherapy reported an overall ORR of 30.6% (11/36) and DCR of 97.2% (35/36) with MET overexpression, amplification, and exon 14 skipping demonstrating ORRs of 35.7% (5/14), 41.2% (7/17), and 66.7% (10/15), respectively; increased ORRs were observed in patients with both exon 14 skipping and amplification (100%, 4/4) and with both amplification and overexpression (50%, 3/6)¹⁷⁰.

— Potential Resistance —

Patients with NSCLC harboring EGFR mutation and MET amplification are unlikely to respond to first- and third-generation EGFR inhibitors (NCCN NSCLC Guidelines, v2.2021). A retrospective study reported significantly shorter PFS and OS on gefitinib treatment for patients with EGFR-mutated non-small cell lung cancer (NSCLC) additionally harboring MET amplification than for those with EGFR-mutated tumors without MET amplification⁸³. Additionally, numerous clinical studies have reported MET amplification as an emergent alteration associated with acquired resistance of EGFR-mutated NSCLC to first-generation EGFR inhibitors such as erlotinib and gefitinib⁸⁴⁻¹⁰⁰ and third-generation EGFR inhibitors such as osimertinib^{91,101-109}. For a small number of patients with EGFR-mutated NSCLC, studies have also reported MET amplification in association with resistance to second-generation EGFR inhibitors such as afatinib and dacomitinib^{97,110-113}. Patients with NSCLC harboring EGFR mutation and MET amplification may benefit from combination

treatment with MET- and EGFR-targeting agents^{87,90,104,106,112,114-116}.

FREQUENCY & PROGNOSIS

MET amplification has been reported at incidences of 14-48% in non-small cell lung cancer (NSCLC), is correlated with increased MET protein expression, and occurs more frequently following treatment with EGFR inhibitors^{127,165,171-177}. In the Phase 2 VISION study of patients with NSCLC, MET amplification was reported in 4.9% of samples¹⁷⁸. Studies on the effect of MET amplification on prognosis in NSCLC have yielded conflicting results^{127,171,175,179-183}, although concurrent MET amplification and EGFR mutation have been correlated with reduced disease-free survival¹⁸⁴. MET exon 14 splice alteration, which has predominantly been observed in lung cancer, was found to be an independent poor prognostic factor in a study of 687 patients with NSCLC¹⁸⁵. However, other studies did not find MET exon 14 splice alteration as a major risk factor for overall survival for NSCLC patients, although recurrence rate was significantly higher in patients with exon 14 splice alteration compared to those with ALK fusion¹⁸⁶⁻¹⁸⁷. Among NSCLC patients with exon 14 alterations that had not been previously treated with a MET inhibitor, a non-significant trend for reduced survival was noted in the context of concurrent MET amplification (5.2 vs 10.5 months, $p = 0.06$)¹⁸⁸.

FINDING SUMMARY

MET encodes a receptor tyrosine kinase, also known as c-MET or hepatocyte growth factor receptor (HGFR), that is activated by the ligand HGF; MET activation results in signaling mediated partly by the RAS-RAF-MAPK and PI3K pathways to promote proliferation¹⁸⁹⁻¹⁹⁰. MET has been reported to be amplified in cancer¹⁹¹, with amplification positively correlating with protein expression in some cancer types^{171,192-195} and associating with therapeutic response to MET inhibitors in a variety of cancer types^{145-147,149-151,196-197}.

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

CCNE1

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies that directly target CCNE1 alterations. Because amplification or overexpression of CCNE1 leads to increased genomic instability through the ATR-CHK1-WEE1 pathway¹⁹⁸⁻¹⁹⁹ and cyclin E1 promotes cell cycle progression in a complex with CDK2²⁰⁰, clinical and preclinical studies have investigated inhibitors of CHK1, ATR, CDK2, and WEE1 as potential therapeutic approaches for tumors with CCNE1 activation. Clinical benefit has been reported for patients with recurrent high-grade serous ovarian carcinoma (HGSOC) with CCNE1 amplification or expression in response to treatment with the CHK1 inhibitor prexasertib²⁰¹. Studies of the WEE1 inhibitor adavosertib observed PRs in

patients with CCNE1-amplified HGSOC and ovarian cancer²⁰²⁻²⁰³. Similarly, in a Phase 2 study of patients with CCNE1-amplified solid tumors, adavosertib elicited an ORR of 26% with PRs reported for patients with ovarian cancer, urothelial carcinoma, or melanoma²⁰⁴. Preclinical studies have demonstrated that cell lines with CCNE1 amplification or overexpression were sensitive to inhibitors of ATR²⁰⁵⁻²⁰⁶, CDK2²⁰⁷, or WEE1^{199,208}. However, other studies have shown that sensitivity of various cell lines to CDK2 inhibitors, including SNS-032, dinaciclib, and seliciclib, at clinically achievable doses, is largely independent of CCNE1 copy number or expression²⁰⁹⁻²¹². One study has reported a reduction in tumor CCNE1 levels in 4/6 lung and esophageal cancer cases following treatment with the HDAC inhibitor vorinostat²¹³.

FREQUENCY & PROGNOSIS

In the Lung Adenocarcinoma and Lung Squamous Cell Carcinoma TCGA datasets, putative high-level CCNE1 amplification has been reported in 2.6%¹²² and 5.6%¹²³ of cases, respectively. CCNE1 amplification was identified in 6% (6/98) of

patients with non-small cell lung cancer (NSCLC) and was associated with TP53 mutation²¹⁴. A study of 68 NSCLC samples observed cyclin E1 overexpression to significantly correlate with centrosome abnormalities²¹⁵. Published data investigating the prognostic implications of CCNE1 in NSCLC are limited (PubMed, Jul 2021).

FINDING SUMMARY

CCNE1 encodes the protein cyclin E1, which plays a role in the regulated transition from the G1 to S phase by binding to and activating cyclin-dependent protein kinase 2 (CDK2). It also has a direct role in initiation of replication and the maintenance of genomic stability²⁰⁰. Amplification of chromosomal region 19q12-q13 has been demonstrated in many types of cancer, and CCNE1 is a well-studied gene within this amplicon²¹⁶⁻²¹⁷. Increased copy number of CCNE1 is highly associated with overexpression of the cyclin E1 protein²¹⁸⁻²¹⁹. Cyclin E1 overexpression can lead to cell transformation as a result of an increase in cyclin E1 activity^{200,220}.

GENE

MYC

ALTERATION

amplification

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no available therapies that directly target MYC. However, preclinical data indicate that MYC overexpression may predict sensitivity to investigational agents targeting CDK1²²¹⁻²²², CDK2²²³, Aurora kinase A²²⁴⁻²³¹, Aurora kinase B²³²⁻²³⁵, glutaminase²³⁶⁻²³⁹, or BET bromodomain-containing proteins²⁴⁰⁻²⁴³, as well as agents targeting both HDAC and PI3K²⁴⁴⁻²⁴⁶. A Phase 2 study reported a PFS benefit associated with a combination of the Aurora A kinase inhibitor alisertib and paclitaxel as second-line therapy for patients with MYC-overexpressed small cell lung

cancer but not for patients without MYC overexpression²⁴⁷. A patient with MYC-amplified invasive ductal breast carcinoma experienced a PR to an Aurora kinase inhibitor²⁴⁸. The glutaminase inhibitor CB-839, in combination with either everolimus or cabozantinib, has demonstrated encouraging efficacy in Phase 1 and 2 studies enrolling patients with pretreated advanced renal cell carcinoma²⁴⁹⁻²⁵⁰.

— Nontargeted Approaches —

MYC amplification has also been suggested to predict response to chemotherapy in patients with breast cancer in some studies²⁵¹⁻²⁵². Preclinical evidence suggests that colon cancer cells with MYC amplification may be more sensitive to 5-fluorouracil and paclitaxel²⁵³⁻²⁵⁴.

FREQUENCY & PROGNOSIS

MYC amplification has been reported in 10-50% of non-small cell lung cancer (NSCLC) samples,

including adenocarcinoma and/or squamous cell carcinoma subtypes²⁵⁵⁻²⁵⁹. In the Lung Adenocarcinoma TCGA and Lung Squamous Cell Carcinoma TCGA datasets, putative MYC amplification has been reported in 9% and 4.5% of cases, respectively¹²²⁻¹²³. MYC amplification has been associated with metastasis in NSCLC, as well as with poor prognosis in early stage lung adenocarcinoma specifically²⁵⁵⁻²⁵⁸.

FINDING SUMMARY

MYC (c-MYC) encodes a transcription factor that regulates many genes related to cell cycle regulation and cell growth. It is an oncogene and may be activated in as many as 20% of cancers²⁶⁰. MYC dysregulation (amplification, overexpression, translocation) has been identified in a number of different cancer types²⁶¹. MYC amplification has been significantly linked with increased mRNA and protein levels and results in the dysregulation of a large number of target genes^{260,262-263}.

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

RNF43

ALTERATION

R145*

TRANSCRIPT ID

NM_017763

CODING SEQUENCE EFFECT

433C>T

VARIANT ALLELE FREQUENCY (% VAF)

16.6%

particularly Porcupine inhibitors, in multiple tumor types²⁶⁴⁻²⁶⁸. In a Phase 1 basket study for the Porcupine inhibitor RXCoo4, 1 of 2 patients with tumors harboring an RNF43 mutation achieved SD²⁶⁹. Of the patients with WNT-ligand-dependent tumors, including those with RNF43 mutations, RSPO fusions, or those with biliary tract or thymus cancer, 71% (5/7) experienced SD²⁶⁹. Therefore, patients whose tumors harbor inactivating alterations in RNF43 may benefit from WNT pathway inhibitors, which are under investigation in clinical trials.

associated with mismatch repair deficiency and microsatellite instability (MSI) in colorectal²⁷¹, endometrial²⁷¹, and gastric cancers²⁷⁵⁻²⁷⁶; one study reported RNF43 alterations in more than 50% of MSI gastric carcinomas²⁷⁵.

FINDING SUMMARY

RNF43 encodes a ubiquitin ligase²⁷⁷ that was discovered because it is overexpressed in colon cancer²⁷⁸. RNF43 and the homologous E3 ubiquitin ligase ZNRF3 are tumor suppressors that function as negative regulators of WNT signaling²⁶⁴⁻²⁶⁸. An additional tumor-suppressor-like role for RNF43 in colon cancer is hypothesized to occur via its interaction with the ubiquitin-protein ligase NEDL1, which is predicted to enhance the pro-apoptotic effects of p53²⁷⁹.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

Preclinical studies have reported that RNF43 is a negative regulator of WNT signaling, and RNF43 loss or inactivation leads to WNT activation and confers sensitivity to WNT pathway inhibitors,

FREQUENCY & PROGNOSIS

Mutations in RNF43 have been reported in 18-27% of endometrial cancers²⁷⁰⁻²⁷¹, 3-5% of pancreatic cancers²⁷², 21% of ovarian mucinous carcinomas²⁷³, 9% of liver fluke-associated cholangiocarcinomas²⁷⁴, and up to 18% of colorectal cancers^{61,271}. RNF43 mutations are

GENE

RAD21

ALTERATION

amplification

and hereditary BRCA-wild-type but not hereditary BRCA1-mutant cancers²⁸¹. Furthermore, SNPs in or near RAD21 have been linked with risk of breast cancer development²⁸³⁻²⁸⁴. RAD21 overexpression has also been correlated with poor prognosis in endometrial cancer²⁸⁵ and in colorectal cancer (CRC), especially in KRAS-mutant CRC²⁸⁶. Heterogeneity of RAD21 expression also correlated with aggressive tumor behavior and shorter survival in endometrial cancer²⁸⁷. RAD21 amplification has been more frequently reported in hormone-refractory than in treatment-naïve prostate cancer, but RAD21 amplification did not correlate with expression²⁸⁸. In the context of ovarian cancer, both RAD21 overexpression and downregulation have been observed, but RAD21 expression was not prognostic²⁸⁹. Downregulation of RAD21 expression resulted in sensitization of cultured breast^{282,290} and CRC²⁸⁶ cells to chemotherapy, thereby suggesting that RAD21 overexpression confers resistance to chemotherapy.

FINDING SUMMARY

RAD21 encodes a protein involved in DNA double-strand break repair and sister chromatid cohesion as a part of the cohesin complex²⁹¹⁻²⁹⁴. In preclinical studies, downregulation of RAD21 or other cohesin components leads to loss of expression from amplified genes, as well as amplifications themselves upon cell passaging²⁹⁵, but also leads to an increase in deletions, insertions, and other rearrangements²⁹⁶. High RAD21 expression has also been associated with increased genomic instability²⁸¹. Cohesin complex also organizes chromatin domains and regulates gene expression²⁹⁷⁻²⁹⁸. Both overexpression and reduction of expression of RAD21 has been reported to alter gene expression²⁹⁹. RAD21 amplification has been correlated with increased expression in breast^{281-282,300} and endometrial²⁸⁵ cancers. Other RAD21 alterations, including truncating and point mutations, have been reported in the context of cancer, but the majority have not been characterized.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no therapies to target alterations in this gene.

FREQUENCY & PROGNOSIS

RAD21 amplifications, point mutations, and truncating mutations have been reported in various cancers²⁸⁰. In the context of breast cancer, increased RAD21 expression has been correlated with poor prognosis in multiple subtypes²⁸¹⁻²⁸², including sporadic Grade 3 but not Grade 1 cancers²⁸¹, as well as hereditary BRCA2-mutant

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

TP53

ALTERATION

P151T

TRANSCRIPT ID

NM_000546

CODING SEQUENCE EFFECT

451C>A

VARIANT ALLELE FREQUENCY (% VAF)

36.9%

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor adavosertib³⁰¹⁻³⁰⁴, or p53 gene therapy and immunotherapeutics such as SGT-53³⁰⁵⁻³⁰⁹ and ALT-801³¹⁰. In a Phase 1 study, adavosertib in combination with gemcitabine, cisplatin, or carboplatin elicited PRs in 9.7% and SDs in 53% of patients with solid tumors; the response rate was 21% (4/19) for patients with TP53 mutations versus 12% (4/33) for patients who were TP53 wildtype³¹¹. A Phase 2 trial of adavosertib in combination with chemotherapy (gemcitabine, carboplatin, paclitaxel, or doxorubicin) reported a 32% (30/94, 3 CR) ORR and a 73% (69/94) DCR for patients with platinum-refractory TP53-mutated ovarian, Fallopian tube, or peritoneal cancer³¹². A smaller Phase 2 trial of adavosertib in combination with carboplatin achieved a 43% (9/21, 1 CR) ORR and a 76% (16/21) DCR for patients with platinum-refractory TP53-mutated ovarian cancer³¹³. The combination of adavosertib with paclitaxel and carboplatin for patients with TP53-mutated ovarian cancer also significantly increased PFS compared with paclitaxel and carboplatin alone³¹⁴. In the Phase 2 VIKTORY trial, patients with TP53-mutated metastatic and/or recurrent gastric cancer experienced a 24% (6/25) ORR with adavosertib combined with paclitaxel¹⁶². A Phase 1 trial of neoadjuvant adavosertib in combination with cisplatin and docetaxel for head and neck squamous cell carcinoma (HNSCC) elicited a 71% (5/7) response rate for patients with TP53

alterations³¹⁵. The Phase 2 FOCUS4-C trial for patients with TP53- and RAS-mutated colorectal cancer reported improvement in PFS (3.61 vs. 1.87 months, HR=0.35, p=0.0022), but not OS (14.0 vs 12.8 months, p=0.93), following adavosertib treatment compared with active monitoring³¹⁶. In a Phase 1b clinical trial of SGT-53 in combination with docetaxel for patients with solid tumors, 75% (9/12) of evaluable patients experienced clinical benefit, including 2 confirmed and 1 unconfirmed PRs and 2 instances of SD with significant tumor shrinkage³⁰⁹. Missense mutations leading to TP53 inactivation may also be sensitive to therapies that reactivate mutated p53 such as APR-246³¹⁷⁻³¹⁹. In a Phase 1b trial for patients with p53-positive high-grade serous ovarian cancer, APR-246 combined with carboplatin and pegylated liposomal doxorubicin achieved a 52% (11/21) response rate and 100% DCR³²⁰. ATR inhibitor treatment of chronic lymphocytic leukemia (CLL) cells with biallelic inactivation of TP53 suppressed cell viability, promoted DNA damage, and attenuated xenograft growth in preclinical studies³²¹⁻³²²; however, ATR inhibitors as monotherapy had little effect on these parameters in solid tumor models in other preclinical studies³²³⁻³²⁴. Therefore, it is unclear whether TP53 inactivation predicts sensitivity to ATR inhibition.

FREQUENCY & PROGNOSIS

TP53 is one of the most commonly mutated genes in lung cancer; mutations have been reported in 43-80% of non-small cell lung cancers (NSCLCs)^{122-123,325-330}, including 42-52% of lung adenocarcinomas and 58-83% of lung squamous cell carcinomas (cBioPortal, COSMIC, Feb 2022)^{48-49,122-123}. TP53 homozygous deletion has been observed in 1.4% of lung adenocarcinoma and <1% of lung squamous cell carcinoma cases (cBioPortal, Feb 2022)^{191,331}. In one study of 55 patients with lung adenocarcinoma, TP53 alterations correlated with immunogenic features including PD-L1 expression, tumor mutation burden and neoantigen presentation; likely as a consequence of this association TP53 mutations correlated with improved clinical outcomes to PD-1 inhibitors pembrolizumab and nivolumab in this study³³². Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma³³³.

FINDING SUMMARY

Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers³³⁴. Alterations such as seen here may disrupt TP53 function or expression³³⁵⁻³³⁹.

POTENTIAL GERMLINE IMPLICATIONS

One or more of the TP53 variants observed here has been described in the ClinVar database as a likely pathogenic or pathogenic germline mutation (by an expert panel or multiple submitters) associated with Li-Fraumeni syndrome (ClinVar, Sep 2021)³⁴⁰. Follow-up germline testing would be needed to distinguish whether the finding in this patient is somatic or germline. Germline mutations in TP53 are associated with the very rare autosomal dominant disorder Li-Fraumeni syndrome and the early onset of many cancers³⁴¹⁻³⁴³, including sarcomas³⁴⁴⁻³⁴⁵. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000³⁴⁶ to 1:20,000³⁴⁵. For pathogenic TP53 mutations identified during tumor sequencing, the rate of germline mutations was 1% in the overall population and 6% in tumors arising before age 30³⁴⁷. In the appropriate clinical context, germline testing of TP53 is recommended.

POTENTIAL CLONAL HEMATOPOIESIS IMPLICATIONS

Variants seen in this gene have been reported to occur in clonal hematopoiesis (CH), an age-related process in which hematopoietic stem cells acquire somatic mutations that allow for clonal expansion³⁴⁸⁻³⁵³. CH in this gene has been associated with increased mortality, risk of coronary heart disease, risk of ischemic stroke, and risk of secondary hematologic malignancy³⁴⁸⁻³⁴⁹. Clinical management of patients with CH in this gene may include monitoring for hematologic changes and reduction of controllable risk factors for cardiovascular disease³⁵⁴. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH^{352,355-356}. Patient-matched peripheral blood mononuclear cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH.

ORDERED TEST # ORD-1306820-01

GENOMIC FINDINGS

GENE

ZNF217

ALTERATION

amplification

has been suggested as a potential biomarker for treatment with the DNA synthesis inhibitor and AKT inhibitor triciribine in breast cancer based on preclinical findings in cultured cells and xenografts expressing high levels of ZNF217; triciribine treatment also restored sensitivity to doxorubicin in these cells³⁶¹.

may contribute to tumorigenesis³⁷³⁻³⁷⁵, and increased expression or activation of ERBB3^{362,376}, FAK³⁶², Aurora kinase A³⁵⁹, AKT³⁶⁰, and TGF-beta/SMAD signaling³⁶² has been demonstrated in ZNF217-expressing tumors or cells.

POTENTIAL TREATMENT STRATEGIES

— Targeted Therapies —

There are no available targeted therapies to address genomic alterations in ZNF217. Expression of ZNF217 may predict relapse of estrogen receptor (ER)-positive breast cancer under hormone therapy through its direct interaction with ER-alpha³⁵⁷⁻³⁵⁸. ZNF217 overexpression has also been associated with resistance to paclitaxel³⁵⁹ and doxorubicin³⁶⁰ in breast cancer cell lines. ZNF217

FREQUENCY & PROGNOSIS

Amplification and/or overexpression of ZNF217 has been reported in breast³⁶², ovarian³⁶³⁻³⁶⁴, gastric³⁶⁵⁻³⁶⁶, colon³⁶⁷, prostate³⁶⁸, esophageal³⁶⁹, and urothelial carcinomas³⁷⁰, glioblastoma³⁷¹, and ovarian carcinosarcomas³⁷². Overexpression in these tumors has generally been linked with aggressive tumor behavior and poor clinical prognosis. High levels of ZNF217 expression result in dysregulation of a broad range of genes that

FINDING SUMMARY

ZNF217 encodes a candidate oncogene that has likely roles in histone modification and transcriptional repression^{360,377}. ZNF217 amplification has been correlated with protein overexpression in breast carcinoma tumors and cell lines³⁷⁸. The role of ZNF217 in promoting tumorigenesis was established in preclinical studies demonstrating that expression of ZNF217 results in the immortalization of both human mammary epithelial cells and ovarian surface epithelial cells in culture³⁷⁹⁻³⁸⁰.

ORDERED TEST # ORD-1306820-01

THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Afatinib

Assay findings association

EGFR

exon 19 deletion (E746_A750del)

AREAS OF THERAPEUTIC USE

Afatinib is an irreversible kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4. It is FDA approved for the first-line treatment of patients with metastatic non-small cell lung cancer (NSCLC) and nonresistant EGFR mutations and for the treatment of patients with metastatic, squamous NSCLC after progression on platinum-based chemotherapy. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer^{66-67,381-382}, whereas data for patients with other tumor types are limited^{69-74,383}. On the basis of limited clinical data, MET amplification may be associated with reduced efficacy of EGFR inhibitors such as afatinib and dacomitinib. Several studies have reported patients with EGFR-mutated non-small cell lung cancer (NSCLC) who experienced emergence of MET amplification in association with acquired resistance to afatinib or dacomitinib^{97,110-113}.

SUPPORTING DATA

Afatinib has shown significant clinical activity for patients with NSCLC and the EGFR common sensitizing mutations L858R or exon 19 deletions, based on extensive clinical evidence^{66,381,384-387}. Two randomized Phase 3 trials reported significantly improved median PFS from afatinib compared with chemotherapy for patients with EGFR common sensitizing mutations (LUX-Lung 3, 13.6 vs. 6.9 months, HR 0.47, $p < 0.001$; LUX-Lung 6, 11.0 vs. 6.6 months, HR 0.28, $p < 0.0001$)^{66,381}. However, while afatinib significantly increased OS relative to chemotherapy for patients with EGFR exon 19 alterations in these two trials (LUX-Lung 3, 33.3 vs. 21.1 months, HR=0.54; LUX-Lung 6, 31.4 vs. 18.4 months, HR=0.64), no significant OS differences were observed in treatment for patients with L858R mutation¹⁴⁰. A similar alteration-specific difference was observed for EGFR-mutated treatment-naive NSCLC in a retrospective analysis, which reported numerically longer median OS from second- versus first-generation EGFR TKIs (48.8 vs. 26.4 months, HR=0.59) for patients with exon 19 deletions, but no substantial difference for patients with L858R (25.4 vs. 20.6 months, HR=0.90)³⁸⁴. A Phase 2b study of first-line afatinib compared with gefitinib, also for NSCLC with exon 19 deletions or L858R, reported similar median OS for the

two therapies (27.9 vs. 24.5 months, HR=0.86) but significantly longer time-to-treatment-failure (13.7 vs. 11.5 months, HR=0.75) and higher ORR (73% vs. 56%, $p=0.0018$) with afatinib³⁸⁵. Patients with metastatic NSCLC and common EGFR mutations who progressed on prior chemotherapy experienced an ORR of 50.0% (30/60) from afatinib in a Phase 4 trial³⁸⁶. As first-line therapy for NSCLC with EGFR exon 19 deletions or L858R, prospective or randomized Phase 2 trials have reported a median PFS of 10.2 months and OS of 24.8 months for patients unfit for chemotherapy³⁸⁷ and an ORR of 72.5% ($n=40$, 1 CR), DCR of 100% (40/40), and median PFS and OS of 15.2 and 30.0 months, respectively, for elderly patients ≥ 70 years old³⁸⁸. A retrospective study of afatinib administered to Asian patients with NSCLC, 99% of whom were previously treated with erlotinib and/or gefitinib, reported an ORR of 27.4% (63/230) for patients with common sensitizing EGFR mutations and an ORR of 24.4% (105/431) for the entire cohort³⁸⁹. In a case report, a patient with NSCLC with exon 19 deletion and leptomeningeal metastases experienced an ongoing 16-month PR from afatinib in extracranial, brain, and leptomeningeal lesions³⁹⁰. For patients with erlotinib- or gefitinib-resistant NSCLC and EGFR mutations, Phase 2/3 studies of afatinib treatment have generally reported ORRs of only 7 to 9%³⁹¹⁻³⁹⁶; however, DCRs of more than 50% have been observed³⁹⁵. In a Phase 1b or observational study, patients with EGFR-mutated NSCLC who progressed on afatinib experienced further clinical benefit from subsequent treatment with afatinib and cetuximab³⁹⁷ or osimertinib³⁹⁸, respectively. Extensive clinical data have demonstrated that afatinib is effective for patients with EGFR-mutated advanced NSCLC, including exon 19 deletions and L858 mutations, as well as uncommon sensitizing mutations in exons 18 or 20^{66,140,381,385,387,389,399}. Afatinib has also shown activity for patients with advanced NSCLC and ERBB2 mutations, most of which were exon 20 insertions^{395,400-410}. The randomized Phase 3 LUX-Lung 8 trial comparing afatinib with erlotinib as second-line therapy for advanced lung squamous cell carcinoma (SCC) reported significantly longer median OS (7.9 vs. 6.8 months, HR=0.81), significantly longer median PFS (2.6 vs. 1.9 months, HR=0.81), and higher DCR (51% vs. 40%, $p=0.002$) for patients treated with afatinib³⁹⁹. For patients who progressed on afatinib monotherapy, additional clinical benefit has been reported from afatinib combined with paclitaxel⁴¹¹.

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THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Capmatinib

Assay findings association

MET
amplification - equivocal

AREAS OF THERAPEUTIC USE

Capmatinib is a selective MET tyrosine kinase inhibitor that is FDA approved to treat patients with non-small cell lung cancer harboring MET exon 14 skipping-associated alterations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data in non-small cell lung cancer^{152,157-160,412}, hepatocellular carcinoma¹⁵⁶, renal cell carcinoma¹⁶¹, and gastric cancer¹⁶², MET amplification may predict sensitivity to selective MET inhibitors.

SUPPORTING DATA

In the Phase 2 GEOMETRY mono-1 study for patients with advanced NSCLC and MET gene copy number (GCN) ≥ 10 , capmatinib elicited ORRs of 29–40%, median PFS of 4.1–4.2 months, and median OS of 9.6–10.6 months

across treatment-naïve and previously treated cohorts⁴¹³. A Phase 1 study of capmatinib monotherapy for advanced EGFR- and ALK-wild-type NSCLC reported ORRs of 46.7% (7/15) for patients with MET GCN ≥ 6 , 25% (3/12) for patients with MET GCN 4–6, and 5.9% (1/17) for patients with MET GCN < 4 ; median PFS was 3.7 months overall, and 7.9 months for patients with MET GCN ≥ 6 ⁴¹⁴. Phase 1b/2 trial of capmatinib and nazartinib for patients with EGFR-mutated, EGFR-TKI-resistant NSCLC and unknown MET status reported a 42% (14/33, 2 CRs) ORR, with no correlation observed between responses and T790M status⁴¹⁵. Multiple Phase 1 and 2 clinical studies have reported limited efficacy for capmatinib monotherapy in non-NSCLC indications, with no responses observed for patients with glioblastoma (n=10)⁴¹⁶, gastric cancer (n=9), or other advanced solid tumors (n=24)⁴¹⁷⁻⁴¹⁸.

Crizotinib

Assay findings association

MET
amplification - equivocal

AREAS OF THERAPEUTIC USE

Crizotinib is an inhibitor of the kinases MET, ALK, ROS1, and RON. It is FDA approved to treat patients with ALK rearrangement- or ROS1 rearrangement-positive non-small cell lung cancer (NSCLC), and to treat pediatric and young adult patients with ALK rearrangement-positive anaplastic large cell lymphoma (ALCL). Please see the drug label for full prescribing information.

GENE ASSOCIATION

Sensitivity of MET alterations to crizotinib is suggested by extensive clinical data in patients with MET-amplified cancers, including non-small cell lung cancer (NSCLC)^{145-147,419-420}, gastric cancer¹⁹⁶, gastroesophageal cancer¹⁴⁹, glioblastoma¹⁵⁰, and carcinoma of unknown primary¹⁵¹, as well as in patients with MET-mutated cancers, including NSCLC⁴²¹⁻⁴²⁶, renal cell carcinoma (RCC)⁴²⁷, and histiocytic sarcoma⁴²¹. Crizotinib has also benefited patients with NSCLC or histiocytic sarcoma tumors harboring various alterations associated with MET exon 14 skipping^{188,421-422,424-426}.

SUPPORTING DATA

In a small study for patients with NSCLC and MET

overexpression with or without gene amplification, crizotinib elicited 11 PRs and 3 SDs in 19 evaluable patients⁴²⁰. Crizotinib has demonstrated efficacy in patients with NSCLC and ALK rearrangements⁴²⁸⁻⁴³², ROS1 rearrangements⁴³³⁻⁴³⁷, an NTRK1 fusion⁴³⁸, or MET activation^{145-147,419-420,422-426,439-445}. The Phase 2 METROS and AcSe trials have reported ORRs of 31.3% to 32.0%, median PFS of 3.2 to 5.0 months, and median OS of 5.4 to 7.7 months for patients with MET amplified advanced non-small cell lung cancer (NSCLC); a higher level of amplification was predictive of better response in the AcSe trial ($P=0.04$)^{433,446}. Additional patients with MET amplified NSCLC have been reported to experience clinical benefit from crizotinib in several case studies^{145-147,442,445,447}. A patient with lung adenocarcinoma harboring K860I and L858R EGFR mutations, who acquired both EGFR T790M and MET amplification upon various treatments, experienced clinical benefit from subsequent combination treatment of osimertinib and crizotinib¹⁰⁶. Two patients with ALK-positive NSCLC and acquired MET amplification experienced benefit from crizotinib monotherapy and crizotinib in combination with lorlatinib⁴⁴⁸.

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THERAPIES WITH CLINICAL BENEFIT

IN PATIENT'S TUMOR TYPE

Dacomitinib

Assay findings association

EGFR

exon 19 deletion (E746_A750del)

AREAS OF THERAPEUTIC USE

Dacomitinib is a second generation irreversible tyrosine kinase inhibitor that targets the kinase domains of EGFR, ERBB2/HER2, and ERBB4/HER4. It is FDA approved for the first-line treatment of patients with metastatic non-small cell lung cancer (NSCLC) with EGFR exon 19 deletion or exon 21 L858R substitution mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR activating mutations may indicate sensitivity to afatinib or dacomitinib for patients with non-small cell lung cancer^{66-67,381-382}, whereas data for patients with other tumor types are limited^{69-74,383}. Patients with untreated advanced NSCLC and EGFR exon 19 deletions achieved an ORR of 76%¹⁴² and a median OS of 34.1 months with dacomitinib⁶⁷. On the basis of limited clinical data, MET amplification may be associated with reduced efficacy of EGFR inhibitors such as afatinib and dacomitinib. Several studies have reported patients with EGFR-mutated non-small cell lung cancer (NSCLC) who experienced emergence of MET amplification in association with acquired resistance to afatinib or dacomitinib^{97,110-113}.

SUPPORTING DATA

A randomized Phase 3 trial in patients with NSCLC with

activating EGFR mutations (primarily L858R or exon 19 deletions) reported improved clinical benefit with first-line dacomitinib compared with gefitinib (median OS, 34.1 vs. 26.8 months, HR=0.760; median PFS, 14.7 vs. 9.2 months, HR=0.59)^{142,449}; median OS was 34.1 to 36.7 months and ORR was 74.9% to 79.3%, depending on the dosing regimen⁴⁵⁰. A pooled subgroup analysis of patients with NSCLC with activating EGFR mutations reported improved clinical efficacy with dacomitinib treatment compared with erlotinib (median PFS, 14.6 vs. 9.6 months, HR=0.717; median OS, 26.6 vs. 23.2 months, HR=0.737)⁴⁵¹. Reduced efficacy of dacomitinib treatment in patients with NSCLC harboring the EGFR T790M mutation has been reported in multiple studies⁴⁵²⁻⁴⁵⁴. A Phase 1 trial of combination dacomitinib and a MEK1/2 inhibitor for patients with KRAS-mutated CRC, NSCLC, or pancreatic cancer reported 20/36 SDs and 16 PDs, however toxicity from this combination prevented long-term treatment in this patient population⁴⁵⁵. A Phase 2 study of dacomitinib in patients with NSCLC who had been previously treated with chemotherapy or erlotinib and were not selected for EGFR mutations reported an ORR of 4.5% (3/66)⁴⁵³. In one study, the combination of dacomitinib and crizotinib was ineffective and associated with high toxicity in patients with NSCLC⁴⁵⁶.

Tepotinib

Assay findings association

MET

amplification - equivocal

AREAS OF THERAPEUTIC USE

Tepotinib is a selective MET tyrosine kinase inhibitor that is FDA approved to treat patients with metastatic non-small cell lung cancer harboring MET exon 14 skipping alterations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

On the basis of clinical data in non-small cell lung cancer^{152,157-160,412}, hepatocellular carcinoma¹⁵⁶, renal cell carcinoma¹⁶¹, and gastric cancer¹⁶², MET amplification may predict sensitivity to selective MET inhibitors.

SUPPORTING DATA

For patients with NSCLC and concurrent MET amplification and EGFR mutation, the Phase 2 INSIGHT study showed that tepotinib plus gefitinib improved ORR (66.7% [8/12] vs. 42.9% [3/7]), median PFS (16.6 vs. 4.2

months, HR=0.13), and median OS (37.3 vs. 13.1 months, HR=0.08) compared with chemotherapy¹⁵⁹⁻¹⁶⁰. The Phase 2 VISION study of tepotinib reported an ORR of 42% (10/24) and an mPFS of 4.2 months for patients with MET-amplified advanced non-small cell lung cancer, with responses observed in the first-, second-, and third-line settings¹⁵⁷. Tepotinib has primarily been investigated in non-small cell lung cancer (NSCLC) and has demonstrated efficacy as a single agent for patients with MET amplification¹⁵⁷ and MET exon 14-skipping alterations⁴⁵⁷⁻⁴⁵⁸. Tepotinib has also been shown to be efficacious in combination with gefitinib for patients with concurrent EGFR mutation and MET amplification or MET overexpression in Phase 2 studies¹⁵⁹⁻¹⁶⁰. A case study reported 1 PR lasting 9 months for a patient with HLA-DRB1-MET fusion-positive NSCLC metastatic to the brain⁴⁵⁹.

ORDERED TEST # ORD-1306820-01

THERAPIES ASSOCIATED WITH RESISTANCE

IN PATIENT'S TUMOR TYPE

Erlotinib

✖ Resistance of variant(s) to associated therapy is likely

Assay findings association

EGFR

exon 19 deletion (E746_A750del)

MET

amplification - equivocal

AREAS OF THERAPEUTIC USE

Erlotinib is a small-molecule inhibitor of EGFR. It is FDA approved as a monotherapy or in combination with ramucirumab for patients with metastatic non-small cell lung cancer (NSCLC) harboring EGFR exon 19 deletions or exon 21 (L858R) mutations. Erlotinib is also FDA approved in combination with gemcitabine as a first-line treatment for advanced pancreatic cancer. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Amplification or activation of EGFR may predict sensitivity to therapies such as erlotinib. For patients with activating mutations in EGFR, treatment with erlotinib has been associated with improved response and lengthened time to progression^{64,460-462}. On the basis of extensive clinical data, MET amplification may predict resistance to first-generation EGFR inhibitors, including erlotinib and gefitinib (NCCN NSCLC Guidelines, v2.2021). A retrospective study of patients with EGFR-mutated lung adenocarcinoma treated with gefitinib reported that increased MET copy number was associated with significantly shorter PFS (7.6 vs. 15.9 months, HR=3.83, p=0.0008) and OS (16.8 vs. 33.0 months, HR=2.25, p=0.03), compared with cases without MET copy number increase⁸³. Additionally, studies have reported >30 patients with EGFR-mutated non-small cell lung cancer (NSCLC) who experienced emergence of MET amplification in association with acquired resistance to erlotinib or gefitinib^{84-100,116,463}.

SUPPORTING DATA

For patients with EGFR-mutated NSCLC, the Phase 3 EURLAC trial reported improved PFS with first-line

erlotinib relative to platinum-based chemotherapy (9.7 vs. 5.2 months, HR=0.37)⁶⁴. A Phase 3 study reported similar efficacy of erlotinib and gefitinib for patients with EGFR-mutated NSCLC⁴⁶⁴. Meta-analysis of studies comparing erlotinib or gefitinib versus chemotherapy in the first-line setting reported no significant improvement in OS for patients with EGFR-mutated NSCLC; however, the lack of improved OS was attributed to the effectiveness of postprogression salvage therapy⁴⁶⁵. In the maintenance setting, the placebo-controlled Phase 3 SATURN trial reported significantly improved PFS with maintenance erlotinib following first-line platinum-based chemotherapy irrespective of EGFR status; however, the largest effect was seen for patients with EGFR mutations (HR=0.10)⁴⁶⁰. In the neoadjuvant setting, a Phase 2 trial reported a numerically improved ORR and significantly longer PFS with erlotinib compared with chemotherapy for patients with advanced EGFR-mutated NSCLC⁴⁶¹. In the placebo-controlled Phase 3 RELAY trial, the addition of ramucirumab to erlotinib improved PFS for previously untreated patients with NSCLC harboring EGFR L858R or exon 19 deletion (19.4 vs. 12.4 months, HR=0.59)⁴⁶⁶. In a Phase 2 trial, no clinical benefit was observed from the addition of bevacizumab to erlotinib for patients with NSCLC harboring EGFR exon 19 deletion or L858R mutation⁴⁶⁷. In one study, median PFS (4.1 vs. 11.7 months, HR=9.7) and median OS (14.1 vs. 47.0 months, HR=10.2) were significantly shorter for patients with NSCLC harboring EGFR L747_A750>P (n=6) relative to those with deletions affecting EGFR E746_A750 (n=24) treated with first-line erlotinib⁴⁶⁸. The Phase 3 BR.21 trial demonstrated prolonged OS for genomically unselected patients with NSCLC treated with erlotinib compared with those treated with standard chemotherapy⁴⁶⁹.

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THERAPIES ASSOCIATED WITH RESISTANCE

IN PATIENT'S TUMOR TYPE

Gefitinib

✗ Resistance of variant(s) to associated therapy is likely

Assay findings association

EGFR

exon 19 deletion (E746_A750del)

MET

amplification - equivocal

AREAS OF THERAPEUTIC USE

Gefitinib targets the tyrosine kinase EGFR and is FDA approved to treat non-small cell lung cancer (NSCLC) harboring exon 19 deletions or exon 21 (L858R) substitution mutations in EGFR. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Activation of EGFR may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and PFS for patients with EGFR-mutated non-small cell lung cancer (NSCLC) treated with gefitinib compared with chemotherapy^{462,470-475}, and responses have been reported for patients with EGFR-rearranged NSCLC⁴⁷⁶⁻⁴⁷⁷. On the basis of extensive clinical data, MET amplification may predict resistance to first-generation EGFR inhibitors, including erlotinib and gefitinib (NCCN NSCLC Guidelines, v2.2021). A retrospective study of patients with EGFR-mutated lung adenocarcinoma treated with gefitinib reported that increased MET copy number was associated with significantly shorter PFS (7.6 vs. 15.9 months, HR=3.83, p=0.0008) and OS (16.8 vs. 33.0 months, HR=2.25, p=0.03), compared with cases without MET copy number increase⁸³. Additionally, studies have reported >30 patients with EGFR-mutated non-small cell lung cancer (NSCLC) who experienced emergence of MET amplification in association with acquired resistance to erlotinib or gefitinib^{84-100,116,463}.

SUPPORTING DATA

A Phase 3 trial of first-line gefitinib therapy for patients with NSCLC and EGFR exon 19 deletions or L858R mutations reported a longer PFS (9.2 months vs. 6.3 months)⁴⁷² but no change in median OS (34.9 months vs. 37.2 months) compared with patients treated with cisplatin plus docetaxel (median OS of 37.2 months)⁴⁷⁸. In patients with EGFR-mutated NSCLC who progressed on 1st or 2nd generation EGFR TKIs, combination of gefitinib with the MET inhibitor capmatinib achieved

ORRs of 32-47% and DCRs of 74-75% in cohorts with MET amplification or overexpression¹¹⁴. In this same setting, gefitinib in combination with the MET inhibitor tepotinib elicited the largest benefit in patients with MET amplification or high-level MET overexpression⁴⁷⁹; in the cohort with MET amplification, gefitinib with tepotinib significantly improved ORR (75.0% vs. 42.9%, OR = 4.00) and median PFS (19.8 vs. 5.5 months, HR = 0.25) as compared with pemetrexed and platinum chemotherapy⁴⁸⁰. Gefitinib achieved an ORR of 69.8% and an OS of 19.2 months as first-line treatment for Caucasian patients with non-small cell lung carcinoma (NSCLC) and EGFR sensitizing mutations⁶⁵. In the retrospective analysis of a Phase 3 study for East Asian patients, gefitinib was reported to have a longer PFS for patients with EGFR mutation-positive NSCLC compared with carboplatin/paclitaxel doublet chemotherapy^{473,481}. Two Phase 3 trials of gefitinib plus pemetrexed and carboplatin compared with gefitinib alone for patients with advanced NSCLC harboring EGFR activating mutations reported significantly higher ORRs (75.3% and 84% vs. 62.5% and 67%), longer median PFSs (16 and 20.9 months vs. 8 and 11.9 months), and longer median OSs (50.9 months and not reached vs. 17 and 38.8 months) with combination treatment; however, combination treatment was associated with increased Grade 3 or higher adverse events⁴⁸²⁻⁴⁸³. Retrospective analysis of East Asian patients with advanced NSCLC receiving first-line gefitinib therapy reported that patients with EGFR exon 19 mutations experienced a longer median PFS (10.9 months) compared with patients with EGFR mutations in exon 18 (7.9 months), exon 20 (1.2 months), exon 21 (7.7 months), or double mutations (5.7 months); however, no differences in OS were seen between EGFR mutations⁴⁸⁴. In a Phase 1 study for treatment-naïve patients with NSCLC, best ORRs of 78% (7/9) were observed in patients treated with combination gefitinib and the PD-L1 inhibitor durvalumab as first-line treatment and of 80% (8/10) in those treated with the combination after gefitinib monotherapy⁴⁸⁵.

ORDERED TEST # ORD-1306820-01

THERAPIES ASSOCIATED WITH RESISTANCE

IN PATIENT'S TUMOR TYPE

Osimertinib

✖ Resistance of variant(s) to associated therapy is likely

Assay findings association

EGFR

exon 19 deletion (E746_A750del)

MET

amplification - equivocal

AREAS OF THERAPEUTIC USE

Osimertinib is an irreversible EGFR TKI that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is FDA approved in various treatment settings for patients with non-small cell lung cancer (NSCLC) whose tumors have EGFR exon 19 deletions, exon 21 L858R mutations, or T790M mutations. Please see the drug label for full prescribing information.

GENE ASSOCIATION

EGFR TKI-sensitizing mutations or rearrangements and/or the EGFR T790M mutation may predict sensitivity to osimertinib in non-small cell lung cancer^{68,141,476,486-487}. Patients with untreated advanced NSCLC and EGFR exon 19 deletions or L858R mutations achieved an ORR of 80% and a median PFS of 21.4 and 14.4 months, respectively¹⁴¹. On the basis of extensive clinical data, MET amplification may predict resistance to third-generation EGFR inhibitors, including osimertinib (NCCN NSCLC Guidelines, v2.2021). Studies have reported >30 patients with EGFR-mutated non-small cell lung cancer (NSCLC) who experienced emergence of MET amplification in association with acquired resistance to osimertinib^{91,101-109,488-491}.

SUPPORTING DATA

In EGFR-mutated, MET-positive NSCLC, combination of osimertinib with the MET inhibitor savolitinib elicited an ORR of 64.4% (56/87) with a median PFS of 9.0 to 9.1 months for T790M-negative patients with progression on first- or second-generation TKIs, and an ORR of 66.7% (12/18) with a median PFS of 11.0 months for T790M-positive patients with progression on first- or second-generation TKIs⁴⁹². Patients with progression on third-generation TKIs achieved an ORR of 30.4% (21/69) and a median PFS of 5.4 months⁴⁹². The Phase 3 FLAURA study reported that, relative to erlotinib or gefitinib, first-line osimertinib significantly increased both median PFS (18.9

vs. 10.2 months, HR=0.46) and median OS (38.6 vs. 31.8 months; HR=0.80) for patients with advanced NSCLC and activating, sensitizing EGFR mutations (specifically, exon 19 deletion or L858R)^{141,493}. In the Phase 3 ADAURA study, patients with early Stage (IB/II/IIIA) EGFR-mutated NSCLC experienced longer PFSs on osimertinib compared to placebo in the adjuvant setting (not reached vs. 28.1 months; HR=0.21)⁴⁹⁴. A Phase 1 study reported that T790M-negative patients with acquired EGFR TKI resistance experienced an ORR of 21% and a median PFS of 2.8 months⁶⁸. A Phase 1b/2 study evaluating osimertinib in combination with the CD73 inhibitor oleclumab for patients with advanced EGFR-mutated, T790M-negative NSCLC reported an ORR of 19% (4/19), a DCR of 81%, and mPFS of 11 months (Kim et al., 2021 AACR Abstract CT163). A Phase 2 trial of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with untreated advanced non-small cell lung cancer (NSCLC) harboring EGFR del19 or L858R reported no difference in ORR (82% vs 86%) and median PFS (22.1 vs 20.2 months, HR 0.862 p=0.213)⁴⁹⁵. The Phase 2 BOOSTER study of osimertinib in combination with bevacizumab versus osimertinib monotherapy for patients with advanced NSCLC with EGFR-sensitizing mutations (exon 19 del or L858R) and L790M at progression on prior EGFR TKI reported no difference in ORR (55% vs 55%), median OS (24.0 vs 24.3 months, HR 1.03 p=0.91), or median PFS (15.4 vs 12.3 months, HR 0.96 p=0.83), although improved PFS was observed for the combination in the subgroup of current or former smokers (16.5 vs 8.4, HR 0.52) while nonsmokers had no benefit (HR 1.47)⁴⁹⁶. The Phase 1b TATTON study of osimertinib in combination with selumetinib, savolitinib, or durvalumab for patients with previously treated EGFR-mutated NSCLC reported ORRs of 42% (15/36), 44% (8/18), and 44% (10/23), respectively⁴⁹⁷.

ORDERED TEST # ORD-1306820-01

THERAPIES WITH CLINICAL BENEFIT

IN OTHER TUMOR TYPE

Cabozantinib

Assay findings association
MET
amplification - equivocal

AREAS OF THERAPEUTIC USE

Cabozantinib inhibits multiple tyrosine kinases, including MET, RET, VEGFRs, and ROS1. It is FDA approved as monotherapy to treat patients with renal cell carcinoma (RCC), hepatocellular carcinoma (HCC), medullary thyroid cancer (MTC), and differentiated thyroid cancer (DTC). It is also approved in combination with nivolumab to treat RCC. Please see the drug label for full prescribing information.

GENE ASSOCIATION

Sensitivity of MET alterations to cabozantinib is suggested by clinical responses in patients with non-small cell lung cancer (NSCLC) harboring MET mutations associated with MET exon 14 skipping, with or without concurrent MET amplification^{422,498}, as well as by extensive preclinical data⁴⁹⁹⁻⁵⁰⁵.

SUPPORTING DATA

Cabozantinib elicited a CR in a patient with lung adenocarcinoma harboring a MET amplification and a mutation affecting MET exon 14 splicing⁴²². A Phase 2 randomized discontinuation trial of cabozantinib reported a 10.0% (6/60) ORR and a 58.3% (35/60) DCR, with median PFS of 4.2 months, for patients with genomically unselected, heavily pretreated NSCLC⁵⁰⁶. Patients with EGFR wild-type non-squamous NSCLC who had progressed after previous treatment experienced longer median PFS with cabozantinib alone or combined with erlotinib (4.3 and 4.7 months, HR=0.39 and 0.37, respectively) compared with single agent erlotinib (1.8 months) in a randomized Phase 2 trial⁵⁰⁷. A Phase 1 study of cabozantinib for advanced solid tumors reported an ORR of 20.0% (4/20; 4 PRs, all in EGFR-mutated tumors) and DCR of 100% (20/20) in the expansion cohort for Japanese patients with NSCLC⁵⁰⁸.

NOTE Genomic alterations detected may be associated with activity of certain FDA approved drugs, however, the agents listed in this report may have varied evidence in the patient's tumor type.

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS

NOTE Clinical trials are ordered by gene and prioritized by: age range inclusion criteria for pediatric patients, proximity to ordering medical facility, later trial phase, and verification of trial information within the last two months. While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and

should be investigated by the physician or research staff. This is not a comprehensive list of all available clinical trials. Foundation Medicine displays a subset of trial options and ranks them in this order of descending priority: Qualification for pediatric trial → Geographical proximity → Later trial phase. Clinical trials listed here may have additional enrollment criteria that may require

medical screening to determine final eligibility. For additional information about listed clinical trials or to conduct a search for additional trials, please see [clinicaltrials.gov](https://www.foundationmedicine.com/genomic-testing#support-services). Or visit <https://www.foundationmedicine.com/genomic-testing#support-services>.

GENE
CCNE1
RATIONALE
Strong preclinical and clinical data suggest that CCNE1 amplification may predict sensitivity to

WEE1 inhibitors.

ALTERATION
amplification

NCT02513563
PHASE 2

AZD1775 Plus Carboplatin-Paclitaxel in Squamous Cell Lung Cancer

TARGETS
WEE1
LOCATIONS: Florida, Ohio

NCT04768868
PHASE 1

The Safety and Pharmacokinetics Preliminary Efficacy of IMP7068 in Patients With Advanced Solid Tumors

TARGETS
WEE1
LOCATIONS: Georgia, Texas, Kentucky, Kansas, Beijing (China), Taipei (Taiwan), Taoyuan (Taiwan), Wuhan (China), Taichung (Taiwan)

NCT03968653
PHASE 1

Study of Oral Debio 0123 in Combination With Carboplatin in Participants With Advanced Solid Tumors

TARGETS
WEE1
LOCATIONS: Barcelona (Spain), Leiden (Netherlands), Nijmegen (Netherlands), Groningen (Netherlands)

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS

| | | | |
|---|--|---|---|
| GENE EGFR | | RATIONALE EGFR activating mutations, rearrangements, or amplification may predict sensitivity to EGFR-targeted therapies. Strategies to overcome resistance to current agents include next-generation EGFR inhibitors and combination | therapies. On the basis of extensive clinical evidence, MET amplification may predict resistance to first-, second-, and third-generation EGFR TKIs in EGFR-mutated non-small cell lung cancer (NSCLC). |
| ALTERATION exon 19 deletion (E746_A750del) | | | |
| NCT03778229 | | PHASE 2 | |
| Osimertinib Plus Savolitinib in EGFRm+/MET+ NSCLC Following Prior Osimertinib | | TARGETS EGFR, MET | |
| LOCATIONS: Santiago (Chile), Barretos (Brazil), Porto Alegre (Brazil), São Paulo (Brazil), Sao Paulo (Brazil), Hato Rey (Puerto Rico), Rio de Janeiro (Brazil), Salvador (Brazil), Florida, District of Columbia | | | |
| NCT04606771 | | PHASE 2 | |
| A Study Comparing Savolitinib Plus Osimertinib vs Savolitinib Plus Placebo in Patients With EGFRm+ and MET Amplified Advanced NSCLC | | TARGETS MET, EGFR | |
| LOCATIONS: Caba (Argentina), Buenos Aires (Argentina), California, Mumbai (India), Rohini (India), Bangalore (India), Taipei 112 (Taiwan), Taipei (Taiwan), Taoyuan City (Taiwan) | | | |
| NCT03940703 | | PHASE 2 | |
| A Study of Tepotinib Plus Osimertinib in Epidermal Growth Factor Receptor (EGFR) Tyrosine Kinase Inhibitor (TKI) Relapsed Mesenchymal-epithelial Transition Factor (MET) Amplified Non-small Cell Lung Cancer (NSCLC) | | TARGETS MET, EGFR | |
| LOCATIONS: Florida, Texas, Tennessee, Kentucky, Maryland, New York, Massachusetts, Illinois, California | | | |
| NCT04077463 | | PHASE 1 | |
| A Study of Lazertinib as Monotherapy or in Combination With JNJ-61186372 in Japanese Participants With Advanced Non-small Cell Lung Cancer | | TARGETS EGFR, MET | |
| LOCATIONS: Rio Piedras (Puerto Rico), Florida, Virginia, Pennsylvania, Missouri, New York, Massachusetts, Michigan, California | | | |
| NCT02795156 | | PHASE 2 | |
| Study to Assess the Activity of Molecularly Matched Targeted Therapies in Select Tumor Types Based on Genomic Alterations | | TARGETS BRAF, VEGFRs, RET, KIT, EGFR, ERBB4, ERBB2, MET, ROS1 | |
| LOCATIONS: Florida, Tennessee, Missouri, Wisconsin, Colorado | | | |
| NCT02609776 | | PHASE 1 | |
| A Dose Escalation Study of JNJ-61186372 in Participants With Advanced Non-Small Cell Lung Cancer | | TARGETS MET, EGFR | |
| LOCATIONS: Florida, Texas, Virginia, Maryland, Pennsylvania, Missouri, New York, Massachusetts, Michigan, Illinois | | | |

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS
NCT02716116
PHASE 1/2

A Trial of AP32788 in Non-Small Cell Lung Cancer

TARGETS
EGFR, ERBB2

LOCATIONS: Florida, Georgia, North Carolina, Virginia, Arizona, California

NCT03755102
PHASE NULL

A Study of Dacomitinib in Patients With Metastatic EGFR Mutant Lung Cancer Previously Treated With Osimertinib

TARGETS
ERBB4, EGFR, ERBB2

LOCATIONS: New Jersey, New York

NCT03944772
PHASE 2

Phase 2 Platform Study in Patients With Advanced Non-Small Lung Cancer Who Progressed on First-Line Osimertinib Therapy (ORCHARD)

TARGETS
EGFR, PD-L1, RET, MET, ALK

LOCATIONS: Texas, Maryland, New York, Connecticut, Massachusetts, Illinois, California

NCT03392246
PHASE 2

A Phase 2 Study of Osimertinib in Combination With Selumetinib in EGFR Inhibitor naïve Advanced EGFR Mutant Lung Cancer

TARGETS
MEK, EGFR

LOCATIONS: North Carolina, Massachusetts

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS

GENE
MET
ALTERATION
amplification - equivocal

RATIONALE

Activating MET alterations may confer sensitivity to MET inhibitors. On the basis of extensive clinical evidence, MET amplification may predict

resistance to first-, second-, and third-generation EGFR TKIs in EGFR-mutated non-small cell lung cancer (NSCLC).

NCT03778229
PHASE 2

Osimertinib Plus Savolitinib in EGFRm+/MET+ NSCLC Following Prior Osimertinib

TARGETS
EGFR, MET

LOCATIONS: Santiago (Chile), Barretos (Brazil), Porto Alegre (Brazil), São Paulo (Brazil), Sao Paulo (Brazil), Hato Rey (Puerto Rico), Rio de Janeiro (Brazil), Salvador (Brazil), Florida, District of Columbia

NCT04606771
PHASE 2

A Study Comparing Savolitinib Plus Osimertinib vs Savolitinib Plus Placebo in Patients With EGFRm+ and MET Amplified Advanced NSCLC

TARGETS
MET, EGFR

LOCATIONS: Caba (Argentina), Buenos Aires (Argentina), California, Mumbai (India), Rohini (India), Bangalore (India), Taipei 112 (Taiwan), Taipei (Taiwan), Taoyuan City (Taiwan)

NCT03940703
PHASE 2

A Study of Tepotinib Plus Osimertinib in Epidermal Growth Factor Receptor (EGFR) Tyrosine Kinase Inhibitor (TKI) Relapsed Mesenchymal-epithelial Transition Factor (MET) Amplified Non-small Cell Lung Cancer (NSCLC)

TARGETS
MET, EGFR

LOCATIONS: Florida, Texas, Tennessee, Kentucky, Maryland, New York, Massachusetts, Illinois, California

NCT03539536
PHASE 2

Study of Telisotuzumab Vedotin (ABBV-399) in Subjects With Previously Treated c-Met+ Non-Small Cell Lung Cancer

TARGETS
MET

LOCATIONS: Florida, Alabama, Mississippi, Texas, Craiova (Romania), Tennessee

NCT03175224
PHASE 1/2

CBT-101 Study for Advanced Solid Tumors and c-Met Dysregulation

TARGETS
MET

LOCATIONS: Rio Piedras (Puerto Rico), Florida, Louisiana, South Carolina

NCT04077099
PHASE 1/2

REGN5093 in Patients With MET-Altered Advanced Non-Small Cell Lung Cancer

TARGETS
MET

LOCATIONS: Bordeaux Cedex 9 (France), Montpellier (France), Florida, Texas, Alabama, Kentucky, District of Columbia, Pennsylvania, Missouri

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS
NCT02795156
PHASE 2

Study to Assess the Activity of Molecularly Matched Targeted Therapies in Select Tumor Types Based on Genomic Alterations

TARGETS
BRAF, VEGFRs, RET, KIT, EGFR, ERBB4,
ERBB2, MET, ROS1

LOCATIONS: Florida, Tennessee, Missouri, Wisconsin, Colorado

NCT03170960
PHASE 1/2

Study of Cabozantinib in Combination With Atezolizumab to Subjects With Locally Advanced or Metastatic Solid Tumors

TARGETS
PD-L1, MET, ROS1, RET, VEGFRs

LOCATIONS: Florida, Louisiana, South Carolina, Texas, Georgia, Virginia

NCT02609776
PHASE 1

A Dose Escalation Study of JNJ-61186372 in Participants With Advanced Non-Small Cell Lung Cancer

TARGETS
MET, EGFR

LOCATIONS: Florida, Texas, Virginia, Maryland, Pennsylvania, Missouri, New York, Massachusetts, Michigan, Illinois

NCT03297606
PHASE 2

Canadian Profiling and Targeted Agent Utilization Trial (CAPTUR)

TARGETS
VEGFRs, ABL, SRC, ALK, ROS1, AXL,
TRKA, MET, TRKC, DDR2, KIT, EGFR,
PD-1, CTLA-4, PARP, CDK4, CDK6,
FLT3, CSF1R, RET, mTOR, ERBB2, MEK,
BRAF, SMO

LOCATIONS: London (Canada), Toronto (Canada), Kingston (Canada), Ottawa (Canada), Montreal (Canada), Regina (Canada), Saskatoon (Canada), Edmonton (Canada), Vancouver (Canada)

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS

GENE
MYC
ALTERATION
amplification

RATIONALE

MYC overexpression may predict sensitivity to inhibition of CDKs, especially CDK1 and CDK2, of Aurora kinases, including Aurora kinase A and B, and of BET domain proteins, which are reported to downregulate MYC expression and MYC-dependent transcriptional programs.

NCT04553133
PHASE 1/2

PF-07104091 as a Single Agent and in Combination Therapy

TARGETS

CDK6, Aromatase, CDK4, CDK2

LOCATIONS: Texas, Massachusetts, Michigan

NCT04555837
PHASE 1/2

Alisertib and Pembrolizumab for the Treatment of Patients With Rb-deficient Head and Neck Squamous Cell Cancer

TARGETS

Aurora kinase A, PD-1

LOCATIONS: Texas

NCT01434316
PHASE 1

Veliparib and Dinaciclib in Treating Patients With Advanced Solid Tumors

TARGETS

PARP, CDK1, CDK9, CDK5, CDK2

LOCATIONS: Massachusetts

NCT03220347
PHASE 1

A Study to Assess the Safety, Tolerability, Pharmacokinetics and Preliminary Efficacy of CC-90010 in Subjects With Advanced Solid Tumors and Relapsed/Refractory Non-Hodgkin's Lymphomas

TARGETS

BRD2, BRD3, BRD4, BRDT

LOCATIONS: Madrid (Spain), Kashiwa (Japan), Koto-ku (Japan), Chikusa-ku (Japan)

ORDERED TEST # ORD-1306820-01

CLINICAL TRIALS
GENE
RNF43
RATIONALE
Based on preclinical evidence, tumors with loss or inactivation of RNF43 may be sensitive to inhibitors of the WNT signaling pathway.

ALTERATION
R145*

NCT02521844
PHASE 1

A Study to Evaluate the Safety and Tolerability of ETC-1922159 in Advanced Solid Tumours

TARGETS
PORCN
LOCATIONS: Texas, North Carolina, Colorado, Singapore (Singapore)

NCT01351103
PHASE 1

A Study of LGK974 in Patients With Malignancies Dependent on Wnt Ligands

TARGETS
PORCN, PD-1
LOCATIONS: Texas, Maryland, New York, California, Madrid (Spain), Valencia (Spain), Hospitalet de Llobregat (Spain), Barcelona (Spain), Rotterdam (Netherlands), Utrecht (Netherlands)

NCT03447470
PHASE 1

Study to Evaluate the Safety and Tolerability of RXC004 in Advanced Malignancies

TARGETS
PORCN
LOCATIONS: Manchester (United Kingdom), Oxford (United Kingdom), Sutton (United Kingdom), London (United Kingdom), Newcastle (United Kingdom)

ORDERED TEST # ORD-1306820-01

APPENDIX
Variants of Unknown Significance

NOTE One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued, and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

ASXL1
L1213F

ATR
amplification

CARD11
S794T

EGFR
T259M

EPHB1
amplification

ERG
amplification

FANCA
L1038V

MED12
F1221fs*74

MLL2
G2493E

PDGFRB
A1099V

PIK3CB
amplification

PRKCI
amplification

SGK1
E84K and amplification

TEK
L549V

TERC
amplification

TIPARP
amplification

TNFAIP3
amplification

U2AF1
amplification

ORDERED TEST # ORD-1306820-01

APPENDIX
Genes Assayed in FoundationOne®CDx

FoundationOne CDx is designed to include genes known to be somatically altered in human solid tumors that are validated targets for therapy, either approved or in clinical trials, and/or that are unambiguous drivers of oncogenesis based on current knowledge. The current assay interrogates 324 genes as well as introns of 36 genes involved in rearrangements. The assay will be updated periodically to reflect new knowledge about cancer biology.

DNA GENE LIST: ENTIRE CODING SEQUENCE FOR THE DETECTION OF BASE SUBSTITUTIONS, INSERTION/DELETIONS, AND COPY NUMBER ALTERATIONS

| | | | | | | | | |
|-------------|-----------------|-----------------|---------|---------------|---------|---------------|------------------|--------|
| ABL1 | ACVR1B | AKT1 | AKT2 | AKT3 | ALK | ALOX12B | AMER1 (FAM123B) | APC |
| AR | ARAF | ARFRP1 | ARID1A | ASXL1 | ATM | ATR | ATRX | AURKA |
| AURKB | AXIN1 | AXL | BAP1 | BARD1 | BCL2 | BCL2L1 | BCL2L2 | BCL6 |
| BCOR | BCORL1 | BRAF | BRCA1 | BRCA2 | BRD4 | BRIP1 | BTG1 | BTG2 |
| BTB | C11orf30 (EMSY) | C17orf39 (GID4) | CALR | CARD11 | CASP8 | CBFB | CBL | CCND1 |
| CCND2 | CCND3 | CCNE1 | CD22 | CD274 (PD-L1) | CD70 | CD79A | CD79B | CDC73 |
| CDH1 | CDK12 | CDK4 | CDK6 | CDK8 | CDKN1A | CDKN1B | CDKN2A | CDKN2B |
| CDKN2C | CEBPA | CHEK1 | CHEK2 | CIC | CREBBP | CRKL | CSF1R | CSF3R |
| CTCF | CTNNA1 | CTNNB1 | CUL3 | CUL4A | CXCR4 | CYP17A1 | DAXX | DDR1 |
| DDR2 | DIS3 | DNMT3A | DOT1L | EED | EGFR | EP300 | EPHA3 | EPHB1 |
| EPHB4 | ERBB2 | ERBB3 | ERBB4 | ERCC4 | ERG | ERRF1 | ESR1 | EZH2 |
| FAM46C | FANCA | FANCC | FANCG | FANCL | FAS | FBXW7 | FGF10 | FGF12 |
| FGF14 | FGF19 | FGF23 | FGF3 | FGF4 | FGF6 | FGFR1 | FGFR2 | FGFR3 |
| FGFR4 | FH | FLCN | FLT1 | FLT3 | FOXO2 | FUBP1 | GABRA6 | GATA3 |
| GATA4 | GATA6 | GNAI1 | GNAI3 | GNAQ | GNAS | GRM3 | GSK3B | H3F3A |
| HDAC1 | HGF | HNF1A | HRAS | HSD3B1 | ID3 | IDH1 | IDH2 | IGF1R |
| IKBKE | IKZF1 | INPP4B | IRF2 | IRF4 | IRS2 | JAK1 | JAK2 | JAK3 |
| JUN | KDMSA | KDMS5C | KDM6A | KDR | KEAP1 | KEL | KIT | KLHL6 |
| KMT2A (MLL) | KMT2D (MLL2) | KRAS | LTK | LYN | MAF | MAP2K1 (MEK1) | MAP2K2 (MEK2) | MAP2K4 |
| MAP3K1 | MAP3K13 | MAPK1 | MCL1 | MDM2 | MDM4 | MED12 | MEF2B | MEN1 |
| MERTK | MET | MITF | MKNK1 | MLH1 | MPL | MRE11A | MSH2 | MSH3 |
| MSH6 | MST1R | MTAP | MTOR | MUTYH | MYC | MYCL (MYCL1) | MYCN | MYD88 |
| NBN | NF1 | NF2 | NFE2L2 | NFKB1A | NKX2-1 | NOTCH1 | NOTCH2 | NOTCH3 |
| NPM1 | NRAS | NSD3 (WHSC1L1) | NT5C2 | NTRK1 | NTRK2 | NTRK3 | P2RY8 | PALB2 |
| PARK2 | PARP1 | PARP2 | PARP3 | PAX5 | PBRM1 | PDCD1 (PD-1) | PDCD1LG2 (PD-L2) | PDGFRA |
| PDGFRB | PDK1 | PIK3C2B | PIK3C2G | PIK3CA | PIK3CB | PIK3R1 | PIM1 | PMS2 |
| POLD1 | POLE | PPARG | PPP2R1A | PPP2R2A | PRDM1 | PRKAR1A | PRKCI | PTCH1 |
| PTEN | PTPN11 | PTPRO | QKI | RAC1 | RAD21 | RAD51 | RAD51B | RAD51C |
| RAD51D | RAD52 | RAD54L | RAF1 | RARA | RB1 | RBM10 | REL | RET |
| RICTOR | RNF43 | ROS1 | RPTOR | SDHA | SDHB | SDHC | SDHD | SETD2 |
| SF3B1 | SGK1 | SMAD2 | SMAD4 | SMARCA4 | SMARCB1 | SMO | SNCAIP | SOC3 |
| SOX2 | SOX9 | SPEN | SPOP | SRC | STAG2 | STAT3 | STK11 | SUFU |
| SYK | TBX3 | TEK | TET2 | TGFBR2 | TIPARP | TNFAIP3 | TNFRSF14 | TP53 |
| TSC1 | TSC2 | TYRO3 | U2AF1 | VEGFA | VHL | WHSC1 | WT1 | XPO1 |
| XRCC2 | ZNF217 | ZNF703 | | | | | | |

DNA GENE LIST: FOR THE DETECTION OF SELECT REARRANGEMENTS

| | | | | | | | | |
|------|------|-------|--------|-------|---------|-------|--------|-------------|
| ALK | BCL2 | BCR | BRAF | BRCA1 | BRCA2 | CD74 | EGFR | ETV4 |
| ETV5 | ETV6 | EWSR1 | EZR | FGFR1 | FGFR2 | FGFR3 | KIT | KMT2A (MLL) |
| MSH2 | MYB | MYC | NOTCH2 | NTRK1 | NTRK2 | NUTM1 | PDGFRA | RAF1 |
| RARA | RET | ROS1 | RSP02 | SDC4 | SLC34A2 | TERC* | TERT** | TPRSS2 |

*TERC is an NCRNA

**Promoter region of TERT is interrogated

ADDITIONAL ASSAYS: FOR THE DETECTION OF SELECT CANCER BIOMARKERS

Loss of Heterozygosity (LOH) score

Microsatellite (MS) status

Tumor Mutational Burden (TMB)

ORDERED TEST # ORD-1306820-01

APPENDIX

About FoundationOne®CDx

FoundationOne CDx fulfills the requirements of the European Directive 98/79 EC for in vitro diagnostic medical devices and is registered as a CE-IVD product by Foundation Medicine's EU Authorized Representative, Qarad b.v.b.a, Cipalstraat 3, 2440 Geel, Belgium.



ABOUT FOUNDATIONONE CDx

FoundationOne CDx was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationOne CDx may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratories are qualified to perform high-complexity clinical testing.

Please refer to technical information for performance specification details:
www.rochefoundationmedicine.com/f1cdxtech.

INTENDED USE

FoundationOne®CDx (F1CDx) is a next generation sequencing based in vitro diagnostic device for detection of substitutions, insertion and deletion alterations (indels), and copy number alterations (CNAs) in 324 genes and select gene rearrangements, as well as genomic signatures including microsatellite instability (MSI), tumor mutational burden (TMB), and for selected forms of ovarian cancer, loss of heterozygosity (LOH) score, using DNA isolated from formalin-fixed, paraffin-embedded (FFPE) tumor tissue specimens. The test is intended as a companion diagnostic to identify patients who may benefit from treatment with therapies in accordance with approved therapeutic product labeling. Additionally, F1CDx is intended to provide tumor mutation profiling to be used by qualified health care professionals in accordance with professional guidelines in oncology for patients with solid malignant neoplasms.

TEST PRINCIPLES

FoundationOne CDx will be performed exclusively as a laboratory service using DNA extracted from formalin-fixed, paraffin-embedded (FFPE) tumor samples. The proposed assay will employ a single DNA extraction method from routine FFPE biopsy or surgical resection specimens, 50-1000 ng of which will undergo whole-genome shotgun library construction and hybridization-based capture of all coding exons from 309 cancer-related genes, one promoter region, one non-coding (ncRNA), and select intronic regions from 34 commonly rearranged genes, 21 of which also include the coding exons. The assay therefore includes detection of alterations in a total of 324 genes.

Using an Illumina® HiSeq platform, hybrid capture-selected libraries will be sequenced to high uniform depth (targeting >500X median coverage with >99% of exons at coverage >100X). Sequence data will be processed using a customized analysis pipeline designed to accurately detect all classes of genomic alterations, including base substitutions, indels, focal copy number amplifications, homozygous gene deletions, and selected genomic rearrangements (e.g., gene fusions). Additionally, genomic signatures including loss of heterozygosity (LOH), microsatellite instability (MSI) and tumor mutational burden (TMB) will be reported.

THE REPORT

Incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research. The F1CDx report may be used as an aid to inform molecular eligibility for clinical trials. Note: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Diagnostic Significance

FoundationOne CDx identifies alterations to select cancer-associated genes or portions of genes (biomarkers). In some cases, the Report also highlights selected negative test results regarding biomarkers of clinical significance.

Qualified Alteration Calls (Equivocal and Subclonal)

An alteration denoted as "amplification – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence that the copy number of a gene exceeds the threshold for identifying copy number amplification. The threshold used in FoundationOne CDx for identifying a copy number amplification is four (4) for ERBB2 and six (6) for all other genes. Conversely, an alteration denoted as "loss – equivocal" implies that the FoundationOne CDx assay data provide some, but not unambiguous, evidence for homozygous deletion of the gene in question. An alteration denoted as "subclonal" is one that the FoundationOne CDx analytical methodology has identified as being present in <10% of the assayed tumor DNA.

Ranking of Therapies and Clinical Trials

Ranking of Therapies in Summary Table

Therapies are ranked based on the following criteria: Therapies with clinical benefit (ranked alphabetically within each evidence category), followed by therapies associated with resistance (when applicable).

Ranking of Clinical Trials

Pediatric trial qualification → Geographical proximity → Later trial phase.

NATIONAL COMPREHENSIVE CANCER NETWORK® (NCCN®) CATEGORIZATION

Biomarker and genomic findings detected may be associated with certain entries within the NCCN Drugs & Biologics Compendium® (NCCN Compendium®) (www.nccn.org). The NCCN Categories of Evidence and Consensus indicated reflect the highest possible category for a given therapy in association with each biomarker or genomic finding. Please note, however, that the accuracy and applicability of these NCCN categories within a report may be impacted by the patient's clinical history, additional biomarker information, age, and/or co-occurring alterations. For additional information on the NCCN categories, please refer to the NCCN Compendium®. Referenced with permission from the NCCN Clinical Practice Guidelines in Oncology (NCCN Guidelines®). © National Comprehensive Cancer Network, Inc. 2022. All rights reserved. To view the most recent and complete version of the guidelines, go online to NCCN.org. NCCN makes no warranties of any kind whatsoever regarding their content, use or application and disclaims any responsibility for their application or use in any way.

Limitations

1. In the fractional-based MSI algorithm, a tumor specimen will be categorized as MSI-H, MSS, or MS-Equivocal according to the fraction of microsatellite loci determined to be altered or unstable (i.e., the fraction unstable loci score). In the F1CDx assay, MSI is evaluated based on a genome-wide analysis across >2000 microsatellite loci. For a given microsatellite locus, non-somatic alleles are discarded, and the microsatellite is categorized as unstable if remaining alleles differ from the reference genome. The final fraction unstable loci score is calculated as the number of unstable microsatellite loci divided by the number of evaluable microsatellite loci. The MSI-H and MSS cut-off thresholds were determined by analytical concordance to a PCR comparator assay using a pan-tumor FFPE tissue sample set. Patients with results categorized as "MS-

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- Stable" with median exon coverage <300X, "MS-Equivocal," or "Cannot Be Determined" should receive confirmatory testing using a validated orthogonal (alternative) method.
2. TMB by F1CDx is determined by counting all synonymous and non-synonymous variants present at 5% allele frequency or greater (after filtering) and the total number is reported as mutations per megabase (mut/Mb) unit. Observed TMB is dependent on characteristics of the specific tumor focus tested for a patient (e.g., primary vs. metastatic, tumor content) and the testing platform used for the detection; therefore, observed TMB results may vary between different specimens for the same patient and between detection methodologies employed on the same sample. The TMB calculation may differ from TMB calculations used by other assays depending on variables such as the amount of genome interrogated, percentage of tumor, assay limit of detection (LoD), filtering of alterations included in the score, and the read depth and other bioinformatic test specifications. Refer to the SSED for a detailed description of these variables in FMI's TMB calculation https://www.accessdata.fda.gov/cdrh_docs/pdf17/P170019B.pdf. The clinical validity of TMB defined by this panel has been established for TMB as a qualitative output for a cut-off of 10 mutations per megabase but has not been established for TMB as a quantitative score.
 3. The LOH score is determined by analyzing SNPs spaced at 1Mb intervals across the genome on the FoundationOne CDx test and extrapolating an LOH profile, excluding arm- and chromosome-wide LOH segments. Detection of LOH has been verified only for ovarian cancer patients, and the LOH score result may be reported for epithelial ovarian, peritoneal, or Fallopian tube carcinomas. The LOH score will be reported as "Cannot Be Determined" if the sample is not of sufficient quality to confidently determine LOH. Performance of the LOH classification has not been established for samples below 35% tumor content. There may be potential interference of ethanol with LOH detection. The interfering effects of xylene, hemoglobin, and triglycerides on the LOH score have not been demonstrated.
 4. Alterations reported may include somatic (not inherited) or germline (inherited) alterations; however, the test does not distinguish between germline and somatic alterations. The test does not provide information about susceptibility.
 5. Biopsy may pose a risk to the patient when archival tissue is not available for use with the assay. The patient's physician should determine

whether the patient is a candidate for biopsy.

6. Reflex testing to an alternative FDA approved companion diagnostic should be performed for patients who have an *ERBB2* amplification result detected with copy number equal to 4 (baseline ploidy of tumor +2) for confirmatory testing. While this result is considered negative by FoundationOne®CDx (F1CDx), in a clinical concordance study with an FDA approved FISH test, 70% (7 out of 10 samples) were positive, and 30% (3 out of 10 samples) were negative by the FISH test with an average ratio of 2.3. The frequency of *ERBB2* copy number 4 in breast cancer is estimated to be approximately 2%. Multiple references listed in <https://www.mycancergenome.org/content/disease/breast-cancer/ERBB2/238/> report the frequency of *HER2* overexpression as 20% in breast cancer. Based on the F1CDx *HER2* CDx concordance study, approximately 10% of *HER2* amplified samples had copy number 4. Thus, total frequency is conservatively estimated to be approximately 2%.

REPORT HIGHLIGHTS

The Report Highlights includes select genomic and therapeutic information with potential impact on patient care and treatment that is specific to the genomics and tumor type of the sample analyzed. This section may highlight information including targeted therapies with potential sensitivity or resistance; evidence-matched clinical trials; and variants with potential diagnostic, prognostic, nontargeted treatment, germline, or clonal hematopoiesis implications. Information included in the Report Highlights is expected to evolve with advances in scientific and clinical research. Findings included in the Report Highlights should be considered in the context of all other information in this report and other relevant patient information. Decisions on patient care and treatment are the responsibility of the treating physician.

VARIANT ALLELE FREQUENCY

Variant Allele Frequency (VAF) represents the fraction of sequencing reads in which the variant is observed. This attribute is not taken into account for therapy inclusion, clinical trial matching, or interpretive content. Caution is recommended in interpreting VAF to indicate the potential germline or somatic origin of an alteration, recognizing that tumor fraction and tumor ploidy of samples may vary.

Precision of VAF for base substitutions and indels

| BASE SUBSTITUTIONS | %CV* |
|--------------------|--------------|
| Repeatability | 5.11 - 10.40 |
| Reproducibility | 5.95 - 12.31 |
| INDELS | %CV* |
| Repeatability | 6.29 - 10.00 |
| Reproducibility | 7.33 - 11.71 |

*Interquartile Range = 1st Quartile to 3rd Quartile

VARIANTS TO CONSIDER FOR FOLLOW-UP GERMLINE TESTING

The variants indicated for consideration of follow-up germline testing are 1) limited to reportable short variants with a protein effect listed in the ClinVar genomic database (Landrum et al., 2018; 29165669) as Pathogenic, Pathogenic/Likely Pathogenic, or Likely Pathogenic (by an expert panel or multiple submitters), 2) associated with hereditary cancer-predisposing disorder(s), 3) detected at an allele frequency of >10%, and 4) in select genes reported by the ESMO Precision Medicine Working Group (Mandelker et al., 2019; 31050713) to have a greater than 10% probability of germline origin if identified during tumor sequencing. The selected genes are *ATM*, *BAP1*, *BRCA1*, *BRCA2*, *BRIP1*, *CHEK2*, *FH*, *FLCN*, *MLH1*, *MSH2*, *MSH6*, *MUTYH*, *PALB2*, *PMS2*, *POLE*, *RAD51C*, *RAD51D*, *RET*, *SDHA*, *SDHB*, *SDHC*, *SDHD*, *TSC2*, and *VHL*, and are not inclusive of all cancer susceptibility genes. The content in this report should not substitute for genetic counseling or follow-up germline testing, which is needed to distinguish whether a finding in this patient's tumor sequencing is germline or somatic. Interpretation should be based on clinical context.

VARIANTS THAT MAY REPRESENT CLONAL HEMATOPOIESIS

Variants that may represent clonal hematopoiesis (CH) are limited to select reportable short variants in defined genes identified in solid tumors only. Variant selection was determined based on gene tumor-suppressor or oncogene status, known role in solid tumors versus hematological malignancies, and literature prevalence. The defined genes are *ASXL1*, *CBL*, *DNMT3A*, *IDH2*, *JAK2*, *KMT2D* (*MLL2*), *MPL*, *MYD88*, *SF3B1*, *TET2*, and *U2AF1* and are not inclusive of all CH genes. The content in this report should not substitute for dedicated hematological workup. Comprehensive genomic profiling of solid tumors detects nontumor alterations that are due to CH. Patient-matched peripheral blood mononuclear

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cell sequencing is required to conclusively determine if this alteration is present in tumor or is secondary to CH. Interpretation should be based on clinical context.

LEVEL OF EVIDENCE NOT PROVIDED

Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

NO GUARANTEE OF CLINICAL BENEFIT

This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

NO GUARANTEE OF REIMBURSEMENT

Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationOne CDx.

TREATMENT DECISIONS ARE RESPONSIBILITY OF PHYSICIAN

Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment. Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report. Certain sample or variant characteristics may result in reduced sensitivity. FoundationOne CDx is performed using DNA derived from tumor, and as such germline events may not be reported.

SELECT ABBREVIATIONS

| ABBREVIATION | DEFINITION |
|--------------|-----------------------------|
| CR | Complete response |
| DCR | Disease control rate |
| DNMT | DNA methyltransferase |
| HR | Hazard ratio |
| ITD | Internal tandem duplication |
| MMR | Mismatch repair |
| mut/Mb | Mutations per megabase |
| NOS | Not otherwise specified |
| ORR | Objective response rate |
| OS | Overall survival |
| PD | Progressive disease |
| PFS | Progression-free survival |
| PR | Partial response |
| SD | Stable disease |
| TKI | Tyrosine kinase inhibitor |

MR Suite Version 6.0.0

The median exon coverage for this sample is 910x

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References

1. Gatalica Z, et al. Cancer Epidemiol. Biomarkers Prev. (2014) PMID: 25392179
2. Kroemer G, et al. Oncoimmunology (2015) PMID: 26140250
3. Lal N, et al. Oncoimmunology (2015) PMID: 25949894
4. Le DT, et al. N. Engl. J. Med. (2015) PMID: 26028255
5. Ayers et al., 2016; ASCO-SITC Abstract P60
6. Warth A, et al. Virchows Arch. (2016) PMID: 26637197
7. Ninomiya H, et al. Br. J. Cancer (2006) PMID: 16641899
8. Vanderwalde A, et al. Cancer Med (2018) PMID: 29436178
9. Zang YS, et al. Cancer Med (2019) PMID: 31270941
10. Dudley JC, et al. Clin. Cancer Res. (2016) PMID: 26880610
11. Takamochi K, et al. Lung Cancer (2017) PMID: 28676214
12. Pytkänen L, et al. Environ. Mol. Mutagen. (1997) PMID: 9329646
13. Gonzalez R, et al. Ann. Oncol. (2000) PMID: 11061602
14. Chen XQ, et al. Nat. Med. (1996) PMID: 8782463
15. Merlo A, et al. Cancer Res. (1994) PMID: 8174113
16. Kocarnik JM, et al. Gastroenterol Rep (Oxf) (2015) PMID: 26337942
17. You JF, et al. Br. J. Cancer (2010) PMID: 21081928
18. Bairwa NK, et al. Methods Mol. Biol. (2014) PMID: 24623249
19. Boland CR, et al. Cancer Res. (1998) PMID: 9823339
20. Pawlik TM, et al. Dis. Markers (2004) PMID: 15528785
21. Boland CR, et al. Gastroenterology (2010) PMID: 20420947
22. Samstein RM, et al. Nat. Genet. (2019) PMID: 30643254
23. Goodman AM, et al. Mol. Cancer Ther. (2017) PMID: 28835386
24. Goodman AM, et al. Cancer Immunol Res (2019) PMID: 31405947
25. Cristescu R, et al. Science (2018) PMID: 30309915
26. Ready N, et al. J. Clin. Oncol. (2019) PMID: 30785829
27. Hellmann MD, et al. N. Engl. J. Med. (2018) PMID: 29658845
28. Hellmann MD, et al. Cancer Cell (2018) PMID: 29657128
29. Hellmann MD, et al. Cancer Cell (2018) PMID: 29731394
30. Rozeman EA, et al. Nat Med (2021) PMID: 33558721
31. Sharma P, et al. Cancer Cell (2020) PMID: 32916128
32. Rizvi NA, et al. Science (2015) PMID: 25765070
33. Colli LM, et al. Cancer Res. (2016) PMID: 27197178
34. Wang VE, et al. J Immunother Cancer (2017) PMID: 28923100
35. Carbone DP, et al. N. Engl. J. Med. (2017) PMID: 28636851
36. Rizvi H, et al. J. Clin. Oncol. (2018) PMID: 29337640
37. Forde PM, et al. N. Engl. J. Med. (2018) PMID: 29658848
38. Miao D, et al. Nat. Genet. (2018) PMID: 30150660
39. Chae YK, et al. Clin Lung Cancer (2019) PMID: 30425022
40. Paz-Ares et al., 2019; ESMO Abstract LBA80
41. Hellmann MD, et al. N. Engl. J. Med. (2019) PMID: 31562796
42. Chalmers ZR, et al. Genome Med (2017) PMID: 28420421
43. Spigel et al., 2016; ASCO Abstract 9017
44. Xiao D, et al. Oncotarget (2016) PMID: 27009843
45. Shim HS, et al. J Thorac Oncol (2015) PMID: 26200269
46. Govindan R, et al. Cell (2012) PMID: 22980976
47. Ding L, et al. Nature (2008) PMID: 18948947
48. Imielinski M, et al. Cell (2012) PMID: 22980975
49. Kim Y, et al. J. Clin. Oncol. (2014) PMID: 24323028
50. Stein et al., 2019; DOI: 10.1200/PO.18.00376
51. Chen Y, et al. J. Exp. Clin. Cancer Res. (2019) PMID: 31088500
52. Yu H, et al. J Thorac Oncol (2019) PMID: 30253973
53. Pfeifer GP, et al. Mutat. Res. (2005) PMID: 15748635
54. Hill VK, et al. Annu Rev Genomics Hum Genet (2013) PMID: 23875803
55. Pfeifer GP, et al. Oncogene (2002) PMID: 12379884
56. Johnson BE, et al. Science (2014) PMID: 24336570
57. Choi S, et al. Neuro-oncology (2018) PMID: 29452419
58. Cancer Genome Atlas Research Network, et al. Nature (2013) PMID: 23636398
59. Briggs S, et al. J. Pathol. (2013) PMID: 23447401
60. Heitzner E, et al. Curr. Opin. Genet. Dev. (2014) PMID: 24583393
61. Nature (2012) PMID: 22810696
62. Roberts SA, et al. Nat. Rev. Cancer (2014) PMID: 25568919
63. Marabelle A, et al. Lancet Oncol. (2020) PMID: 32919526
64. Rosell R, et al. Lancet Oncol. (2012) PMID: 22285168
65. Douillard JY, et al. Br. J. Cancer (2014) PMID: 24263064
66. Sequist LV, et al. J. Clin. Oncol. (2013) PMID: 23816960
67. Mok TS, et al. J. Clin. Oncol. (2018) PMID: 29864379
68. Jänne PA, et al. N. Engl. J. Med. (2015) PMID: 25923549
69. Hong MH, et al. Cancer (2020) PMID: 32749686
70. Kim HS, et al. Oncotarget (2015) PMID: 26462025
71. Kim HS, et al. Clin. Cancer Res. (2015) PMID: 25424851
72. Mondal G, et al. Acta Neuropathol (2020) PMID: 32303840
73. Cavallieri S, et al. Eur. J. Cancer (2018) PMID: 29734047
74. Chi AS, et al. JCO Precis Oncol (2020) PMID: 32923886
75. Leigh et al., 2021; ESMO Abstract 1192MO
76. Cho et al., 2020; ESMO Abstract 1258O
77. Bauml et al., 2021; ASCO Abstract 9006
78. Shu et al., 2021; ESMO Abstract 1193MO
79. Jänne PA, et al. Cancer Discov (2021) PMID: 34548309
80. Ahn MJ, et al. Lancet Respir Med (2017) PMID: 29056570
81. Yang Z, et al. Sci Transl Med (2016) PMID: 27928026
82. Ahn MJ, et al. Lancet Oncol (2019) PMID: 31587882
83. Noro R, et al. BMC Cancer (2015) PMID: 25886066
84. Turke AB, et al. Cancer Cell (2010) PMID: 20129249
85. Sequist LV, et al. Sci Transl Med (2011) PMID: 21430269
86. Yanagita M, et al. Clin Cancer Res (2016) PMID: 27281561
87. Li A, et al. Clin. Cancer Res. (2017) PMID: 28396313
88. Yu HA, et al. Clin. Cancer Res. (2013) PMID: 23470965
89. Miyoshi S, et al. Onco Targets Ther (2015) PMID: 25914548
90. Bahcall M, et al. Cancer Discov (2016) PMID: 27694386
91. Wang Y, et al. Lung Cancer (2020) PMID: 32540560
92. Gainer JF, et al. J Thorac Oncol (2016) PMID: 26988570
93. Yoshimura K, et al. Respir Med Case Rep (2017) PMID: 28271038
94. Zhu VW, et al. Lung Cancer (Auckl) (2019) PMID: 30881166
95. Seki N, et al. Case Rep Oncol () PMID: 30792648
96. Li YQ, et al. Ann Oncol (2017) PMID: 28961830
97. Wagener-Rydzek S, et al. BMC Cancer (2020) PMID: 32397977
98. Takezawa K, et al. Cancer Discov (2012) PMID: 22956644
99. Womack JP, et al. J Thorac Oncol (2015) PMID: 26709484
100. Cardona AF, et al. Target Oncol (2017) PMID: 28620690
101. Planchard D, et al. Ann. Oncol. (2015) PMID: 26269204
102. Ortiz-Cuaran S, et al. Clin Cancer Res (2016) PMID: 27252416
103. Ou SI, et al. Lung Cancer (2016) PMID: 27393507
104. Wang Y, et al. Lung Cancer (2018) PMID: 29571987
105. Ramalingam SS, et al. J. Clin. Oncol. (2018) PMID: 28841389
106. York ER, et al. J Thorac Oncol (2017) PMID: 28274743
107. Nishiyama A, et al. Cancer Sci (2020) PMID: 32735723
108. Martinez-Marti A, et al. Ann Oncol (2017) PMID: 28961841
109. Piotrowska Z, et al. Cancer Discov (2018) PMID: 30257958
110. Long Y, et al. Ann Palliat Med (2020) PMID: 32648452
111. Liang SK, et al. Oncotarget (2017) PMID: 29163842
112. He Q, et al. Ann Oncol (2020) PMID: 32122695
113. Choudhury NJ, et al. JCO Precis Oncol (2021) PMID: 34250398
114. Wu YL, et al. J. Clin. Oncol. (2018) PMID: 30156984
115. Scheffler M, et al. J Thorac Oncol (2015) PMID: 26001148
116. He Q, et al. Transl Lung Cancer Res (2020) PMID: 32676342
117. Reck M, et al. Lancet Respir Med (2019) PMID: 30922878
118. Socinski MA, et al. J Thorac Oncol (2021) PMID: 34311108
119. Socinski MA, et al. N. Engl. J. Med. (2018) PMID: 29863955
120. Lu et al., 2021; ESMO Abstract VP9-2021
121. Vallee A, et al. Int. J. Oncol. (2013) PMID: 23934203
122. Nature (2014) PMID: 25079552
123. Nature (2012) PMID: 22960745
124. Watzka SB, et al. Eur J Cardiothorac Surg (2010) PMID: 20353893
125. Liang Z, et al. BMC Cancer (2010) PMID: 20637128
126. Grob TJ, et al. Lung Cancer (2013) PMID: 23238037
127. Park S, et al. Histol. Histopathol. (2012) PMID: 22207554
128. Dobashi Y, et al. Hum. Pathol. (2011) PMID: 21040950
129. Ludovini V, et al. Cancer Chemother. Pharmacol. (2013) PMID: 23314677
130. Skrzypski M, et al. Clin Lung Cancer (2013) PMID: 23870818
131. Kim SH, et al. Histol. Histopathol. (2012) PMID: 22419022
132. Lee JS, et al. Ann. Surg. Oncol. (2013) PMID: 23525704
133. Oakley GJ, et al. J Thorac Oncol (2011) PMID: 21587084
134. Marks JL, et al. J Thorac Oncol (2008) PMID: 18303429
135. Izar B, et al. Ann. Thorac. Surg. (2013) PMID: 23932319
136. Ciardiello F, et al. N. Engl. J. Med. (2008) PMID: 18337605
137. Lynch TJ, et al. N. Engl. J. Med. (2004) PMID: 15118073
138. Paez JG, et al. Science (2004) PMID: 15118125
139. Pao W, et al. Proc. Natl. Acad. Sci. U.S.A. (2004) PMID: 15329413
140. Yang JC, et al. Lancet Oncol. (2015) PMID: 25589191
141. Soria JG, et al. N. Engl. J. Med. (2018) PMID: 29151359
142. Wu YL, et al. Lancet Oncol. (2017) PMID: 28958502
143. Gilmer TM, et al. Cancer Res. (2008) PMID: 18199554
144. Foster SA, et al. Cancer Cell (2016) PMID: 26996308
145. Ou SH, et al. J Thorac Oncol (2011) PMID: 21623265
146. Schwab R, et al. Lung Cancer (2014) PMID: 24192513
147. Le X, et al. Clin Lung Cancer (2015) PMID: 25922291
148. Schrock AB, et al. J Thorac Oncol (2017) PMID: 28315738
149. Lennerz JK, et al. J. Clin. Oncol. (2011) PMID: 22042947
150. Chi AS, et al. J. Clin. Oncol. (2012) PMID: 22162573
151. Palma NA, et al. Case Rep Oncol (2014) PMID: 25232318
152. Schuler et al., 2016; ASCO Abstract 9067

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APPENDIX

References

153. Wu et al., 2018; WCLC Abstract P1.01-97
154. Gainor JF, et al. J Thorac Oncol (2020) PMID: 31864558
155. Gautschi O, et al. J Thorac Oncol (2020) PMID: 31864554
156. Faivre et al., 2021; ASCO GI Abstract 329
157. Le et al., 2021; ASCO Abstract 9021
158. Yang et al., 2019; AACR Abstract CT193
159. Park et al., 2019; ESMO Abstract 4770
160. Wu et al., 2019; IASLC Abstract MA09.09
161. Gan HK, et al. Clin. Cancer Res. (2019) PMID: 30952639
162. Lee J, et al. Cancer Discov (2019) PMID: 31315834
163. Kim ST, et al. Transl Oncol (2019) PMID: 30695737
164. Kwak et al., 2015; ASCO GI Abstract 01
165. Spigel DR, et al. J. Clin. Oncol. (2013) PMID: 24101053
166. Catenacci DV, et al. Cancer Discov (2011) PMID: 22389872
167. Harding JJ, et al. Clin. Cancer Res. (2019) PMID: 31142504
168. Strickler JH, et al. J. Clin. Oncol. (2018) PMID: 30285518
169. Camidge et al., 2021; AACR Abstract CT179
170. Yang et al., 2020; AACR Abstract CT127
171. Dziadziuszko R, et al. J Thorac Oncol (2012) PMID: 22237262
172. Murray S, et al. J. Exp. Clin. Cancer Res. (2012) PMID: 22992338
173. Xia N, et al. Exp. Lung Res. (2013) PMID: 23919423
174. Nakamura Y, et al. Cancer Sci. (2007) PMID: 17459054
175. Cappuzzo F, et al. J. Clin. Oncol. (2009) PMID: 19255323
176. Zucali PA, et al. Ann. Oncol. (2008) PMID: 18467317
177. An SJ, et al. PLoS ONE (2012) PMID: 22768234
178. Le et al., 2020; AACR Abstract 3385
179. Chen JJ, et al. Lung Cancer (2013) PMID: 23079155
180. Chen YT, et al. J Thorac Oncol (2011) PMID: 22052229
181. Kanteti R, et al. J. Environ. Pathol. Toxicol. Oncol. (2009) PMID: 19817696
182. To C, et al. Exp. Cell Res. (2002) PMID: 11795945
183. Tsuta K, et al. J Thorac Oncol (2012) PMID: 22198430
184. Tanaka A, et al. Lung Cancer (2012) PMID: 21733594
185. Tong JH, et al. Clin. Cancer Res. (2016) PMID: 26847053
186. Lee GD, et al. J Thorac Oncol (2017) PMID: 28502721
187. Gow CH, et al. Lung Cancer (2017) PMID: 28024701
188. Awad et al., 2017; ASCO Abstract 8511
189. J. Clin. Oncol. (2011) PMID: 22042966
190. Jung KH, et al. Arch. Pharm. Res. (2012) PMID: 22553051
191. Gao J, et al. Sci Signal (2013) PMID: 23550210
192. Ang CS, et al. Anticancer Res. (2013) PMID: 23898085
193. Abou-Bakr AA, et al. Gulf J Oncolog (2013) PMID: 23996864
194. Ho JC, et al. Semin Respir Crit Care Med (2013) PMID: 24258573
195. Madoz-Gürpide J, et al. J Transl Med (2015) PMID: 26319934
196. Ali SM, et al. Oncologist (2015) PMID: 25882375
197. Kwak EL, et al. Cancer Discov (2015) PMID: 26432108
198. Lin AB, et al. Clin. Cancer Res. (2017) PMID: 28331049
199. Chen X, et al. Clin. Cancer Res. (2018) PMID: 30181387
200. Mörry T, et al. Int. J. Biochem. Cell Biol. (2004) PMID: 15147722
201. Lee JM, et al. Lancet Oncol. (2018) PMID: 29361470
202. Lheureux S, et al. Lancet (2021) PMID: 33485453
203. Oza AM, et al. Clin. Cancer Res. (2020) PMID: 32611648
204. Fu et al., 2021; AACR abstract 974
205. Toledo LI, et al. Nat. Struct. Mol. Biol. (2011) PMID: 21552262
206. Buisson R, et al. Mol. Cell (2015) PMID: 26365377
207. Yang L, et al. Oncotarget (2015) PMID: 26204491
208. Kok YP, et al. Oncogenesis (2020) PMID: 33028815
209. Taylor-Harding B, et al. Oncotarget (2015) PMID: 25557169
210. Etemadmoghadam D, et al. Clin. Cancer Res. (2013) PMID: 24004674
211. Scaltriti M, et al. Proc. Natl. Acad. Sci. U.S.A. (2011) PMID: 21321214
212. Nanos-Webb A, et al. Breast Cancer Res. Treat. (2012) PMID: 21695458
213. Ma T, et al. Mol. Cancer Ther. (2013) PMID: 23686769
214. Blons H, et al. BMC Med Genomics (2008) PMID: 18549475
215. Koutsami MK, et al. J. Pathol. (2006) PMID: 16739112
216. Leung SY, et al. Mod. Pathol. (2006) PMID: 16575401
217. Lin L, et al. Cancer Res. (2000) PMID: 11156406
218. Mayr D, et al. Am. J. Clin. Pathol. (2006) PMID: 16753589
219. Nakayama N, et al. Cancer (2010) PMID: 20336784
220. Stamatakis M, et al. World J Surg Oncol (2010) PMID: 21176227
221. Horiuchi D, et al. J. Exp. Med. (2012) PMID: 22430491
222. Goga A, et al. Nat. Med. (2007) PMID: 17589519
223. Molenaar JJ, et al. Proc. Natl. Acad. Sci. U.S.A. (2009) PMID: 19525400
224. Dammert MA, et al. Nat Commun (2019) PMID: 31375684
225. Mollaoglu G, et al. Cancer Cell (2017) PMID: 28089889
226. Cardnell RJ, et al. Oncotarget (2017) PMID: 29088717
227. Wang L, et al. Mol Oncol (2017) PMID: 28417568
228. Takahashi Y, et al. Ann. Oncol. (2015) PMID: 25632068
229. Li Y, et al. Thyroid (2018) PMID: 30226440
230. Mahadevan D, et al. PLoS ONE (2014) PMID: 24893165
231. Park SJ, et al. Target Oncol (2019) PMID: 31429028
232. Helfrich BA, et al. Mol. Cancer Ther. (2016) PMID: 27496133
233. Hook KE, et al. Mol. Cancer Ther. (2012) PMID: 22222631
234. Yang D, et al. Proc. Natl. Acad. Sci. U.S.A. (2010) PMID: 20643922
235. He J, et al. Anticancer Drugs (2019) PMID: 30540594
236. Shroff EH, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 25964345
237. Effenberger M, et al. Oncotarget (2017) PMID: 29156762
238. Qu X, et al. Biochem. Biophys. Res. Commun. (2018) PMID: 30103944
239. Xiang Y, et al. J. Clin. Invest. (2015) PMID: 25915584
240. Delmore JE, et al. Cell (2011) PMID: 21889194
241. Bandopadhyay P, et al. Clin. Cancer Res. (2014) PMID: 24297863
242. Lovén J, et al. Cell (2013) PMID: 23582323
243. Otto C, et al. Neoplasia (2019) PMID: 31734632
244. Dong LH, et al. J Hematol Oncol (2013) PMID: 23866964
245. Pei Y, et al. Cancer Cell (2016) PMID: 26977882
246. Fu XH, et al. Acta Pharmacol. Sin. (2019) PMID: 30224636
247. Owonikoko TK, et al. J Thorac Oncol (2020) PMID: 31655296
248. Ganesan P, et al. Mol. Cancer Ther. (2014) PMID: 25253784
249. Tannir et al., 2018; ASCO GU Abstract 603
250. Motzer et al., 2019; ESMO Abstract LBA54
251. Pereira CB, et al. PLoS ONE (2013) PMID: 23555992
252. Yasojima H, et al. Eur. J. Cancer (2011) PMID: 21741827
253. Arango D, et al. Cancer Res. (2001) PMID: 11406570
254. Bottone MG, et al. Exp. Cell Res. (2003) PMID: 14516787
255. Lockwood WW, et al. Oncogene (2008) PMID: 18391978
256. Kubokura H, et al. Ann Thorac Cardiovasc Surg (2001) PMID: 11578259
257. Iwakawa R, et al. Clin. Cancer Res. (2011) PMID: 21148746
258. Boelens MC, et al. Lung Cancer (2009) PMID: 19324446
259. Yokota J, et al. Oncogene (1988) PMID: 2838790
260. Dang CV, et al. Semin. Cancer Biol. (2006) PMID: 16904903
261. Nesbit CE, et al. Oncogene (1999) PMID: 10378696
262. Blancato J, et al. Br. J. Cancer (2004) PMID: 15083194
263. Fromont G, et al. Hum. Pathol. (2013) PMID: 23574779
264. Hao HX, et al. Nature (2012) PMID: 22575959
265. Koo BK, et al. Nature (2012) PMID: 22895187
266. Jiang X, et al. Proc. Natl. Acad. Sci. U.S.A. (2013) PMID: 23847203
267. Koo BK, et al. Proc. Natl. Acad. Sci. U.S.A. (2015) PMID: 26023187
268. Tsukiyama T, et al. Mol. Cell. Biol. (2015) PMID: 25825523
269. Cook et al., 2021; ESMO Abstract 517MO
270. Kinde I, et al. Sci Transl Med (2013) PMID: 23303603
271. Giannakis M, et al. Nat. Genet. (2014) PMID: 25344691
272. Madan B, et al. Mol. Cancer Ther. (2015) PMID: 25901018
273. Ryland GL, et al. J. Pathol. (2013) PMID: 23096461
274. Ong CK, et al. Nat. Genet. (2012) PMID: 22561520
275. Wang K, et al. Nat. Genet. (2014) PMID: 24816253
276. Nature (2014) PMID: 25079317
277. Sugiyama T, et al. Exp. Cell Res. (2008) PMID: 18313049
278. Yagyu R, et al. Int. J. Oncol. (2004) PMID: 15492824
279. Shinada K, et al. Biochem. Biophys. Res. Commun. (2011) PMID: 21108931
280. Zehir A, et al. Nat. Med. (2017) PMID: 28481359
281. Yan M, et al. Breast Cancer Res. (2012) PMID: 22537934
282. Xu H, et al. Breast Cancer Res. (2011) PMID: 21255398
283. Stevens KN, et al. Breast Cancer Res. Treat. (2011) PMID: 21607584
284. Sehl ME, et al. Clin. Cancer Res. (2009) PMID: 19276285
285. Supernat A, et al. Oncol Lett (2012) PMID: 23205091
286. Deb S, et al. Br. J. Cancer (2014) PMID: 24548858
287. Supernat A, et al. Transl Oncol (2014) PMID: 25048628
288. Porkka KP, et al. Genes Chromosomes Cancer (2004) PMID: 14603436
289. Davis SJ, et al. Mol. Cancer Ther. (2015) PMID: 25852062
290. Atienza JM, et al. Mol. Cancer Ther. (2005) PMID: 15767545
291. Xu H, et al. Nat. Rev. Cancer (2011) PMID: 21326324
292. Hill VK, et al. Biochim. Biophys. Acta (2016) PMID: 27207471
293. Solomon DA, et al. BMB Rep (2014) PMID: 24856830
294. Bauerschmidt C, et al. Nucleic Acids Res. (2010) PMID: 19906707
295. Yun J, et al. Nucleic Acids Res. (2016) PMID: 26420833
296. Gelot C, et al. Nucleus (2016) PMID: 27326661
297. Sofueva S, et al. EMBO J. (2013) PMID: 24185899
298. Deng Z, et al. EMBO J. (2012) PMID: 23010778
299. Yun J, et al. EMBO Rep. (2016) PMID: 27466323
300. Mahmood SF, et al. Carcinogenesis (2014) PMID: 24148822
301. Hirai H, et al. Cancer Biol. Ther. (2010) PMID: 20107315
302. Bridges KA, et al. Clin. Cancer Res. (2011) PMID: 21799033
303. Rajeshkumar NV, et al. Clin. Cancer Res. (2011) PMID: 21389100
304. Osman AA, et al. Mol. Cancer Ther. (2015) PMID: 25504633

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APPENDIX

References

305. Xu L, et al. Mol. Cancer Ther. (2002) PMID: 12489850
306. Xu L, et al. Mol. Med. (2001) PMID: 11713371
307. Camp ER, et al. Cancer Gene Ther. (2013) PMID: 23470564
308. Kim SS, et al. Nanomedicine (2015) PMID: 25240597
309. Pirolo KF, et al. Mol. Ther. (2016) PMID: 27357628
310. Hajdenberg et al., 2012; ASCO Abstract e15010
311. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27601554
312. Moore et al., 2019; ASCO Abstract 5513
313. Leijen S, et al. J. Clin. Oncol. (2016) PMID: 27998224
314. Oza et al., 2015; ASCO Abstract 5506
315. Méndez E, et al. Clin. Cancer Res. (2018) PMID: 29535125
316. Seligmann JF, et al. J Clin Oncol (2021) PMID: 34538072
317. Lehmann S, et al. J. Clin. Oncol. (2012) PMID: 22965953
318. Mohell N, et al. Cell Death Dis (2015) PMID: 26086967
319. Fransson Å, et al. J Ovarian Res (2016) PMID: 27179933
320. Gourley et al., 2016; ASCO Abstract 5571
321. Kwok M, et al. Blood (2016) PMID: 26563132
322. Boudry M, et al. Haematologica (2019) PMID: 30975914
323. Dillon MT, et al. Mol. Cancer Ther. (2017) PMID: 28062704
324. Middleton FK, et al. Cancers (Basel) (2018) PMID: 30127241
325. Mogi A, et al. J. Biomed. Biotechnol. (2011) PMID: 21331359
326. Tekpli X, et al. Int. J. Cancer (2013) PMID: 23011884
327. Vignot S, et al. J. Clin. Oncol. (2013) PMID: 23630207
328. Maeng CH, et al. Anticancer Res. (2013) PMID: 24222160
329. Cortot AB, et al. Clin Lung Cancer (2014) PMID: 24169260
330. Itakura M, et al. Br. J. Cancer (2013) PMID: 23922113
331. Cerami E, et al. Cancer Discov (2012) PMID: 22588877
332. Dong ZY, et al. Clin. Cancer Res. (2017) PMID: 28039262
333. Seo JS, et al. Genome Res. (2012) PMID: 22975805
334. Brown CJ, et al. Nat. Rev. Cancer (2009) PMID: 19935675
335. Joerger AC, et al. Annu. Rev. Biochem. (2008) PMID: 18410249
336. Kato S, et al. Proc. Natl. Acad. Sci. U.S.A. (2003) PMID: 12826609
337. Kamada R, et al. J. Biol. Chem. (2011) PMID: 20978130
338. Zerdoumi Y, et al. Hum. Mol. Genet. (2017) PMID: 28472496
339. Yamada H, et al. Carcinogenesis (2007) PMID: 17690113
340. Landrum MJ, et al. Nucleic Acids Res. (2018) PMID: 29165669
341. Bougeard G, et al. J. Clin. Oncol. (2015) PMID: 26014290
342. Sorrell AD, et al. Mol Diagn Ther (2013) PMID: 23355100
343. Nichols KE, et al. Cancer Epidemiol. Biomarkers Prev. (2001) PMID: 11219776
344. Kleihues P, et al. Am. J. Pathol. (1997) PMID: 9006316
345. Gonzalez KD, et al. J. Clin. Oncol. (2009) PMID: 19204208
346. Laloo F, et al. Lancet (2003) PMID: 12672316
347. Mandelker D, et al. Ann. Oncol. (2019) PMID: 31050713
348. Jaiswal S, et al. N. Engl. J. Med. (2014) PMID: 25426837
349. Genovese G, et al. N. Engl. J. Med. (2014) PMID: 25426838
350. Xie M, et al. Nat. Med. (2014) PMID: 25326804
351. Acuna-Hidalgo R, et al. Am. J. Hum. Genet. (2017) PMID: 28669404
352. Severson EA, et al. Blood (2018) PMID: 29678827
353. Fuster JJ, et al. Circ. Res. (2018) PMID: 29420212
354. Hematology Am Soc Hematol Educ Program (2018) PMID: 30504320
355. Chabon JJ, et al. Nature (2020) PMID: 32269342
356. Razavi P, et al. Nat. Med. (2019) PMID: 31768066
357. Nguyen NT, et al. Mol Oncol (2014) PMID: 24973012
358. Frierie S, et al. BMC Genomics (2014) PMID: 24962896
359. Thollet A, et al. Mol. Cancer (2010) PMID: 21059223
360. Huang G, et al. Hum. Mol. Genet. (2005) PMID: 16203743
361. Littlepage LE, et al. Cancer Discov (2012) PMID: 22728437
362. Vendrell JA, et al. Cancer Res. (2012) PMID: 22593193
363. Li J, et al. Int J Clin Exp Pathol (2014) PMID: 25031722
364. Rahman MT, et al. Anticancer Res. (2012) PMID: 22843878
365. Yang SH, et al. Clin. Cancer Res. (2005) PMID: 15701848
366. Shida A, et al. Anticancer Res. (2014) PMID: 25202062
367. Rooney PH, et al. J. Pathol. (2004) PMID: 15476264
368. Szczyrba J, et al. Int. J. Cancer (2013) PMID: 22815235
369. Geppert CI, et al. Br. J. Cancer (2014) PMID: 24853183
370. Toncheva D, et al. Tumour Biol. () PMID: 15897688
371. Mao XG, et al. Lab. Invest. (2011) PMID: 21483406
372. Schipf A, et al. Virchows Arch. (2008) PMID: 18193277
373. Quinlan KG, et al. Biochim. Biophys. Acta (2007) PMID: 17572303
374. Krig SR, et al. J. Biol. Chem. (2007) PMID: 17259635
375. Cowger JJ, et al. Oncogene (2007) PMID: 17130829
376. Krig SR, et al. Oncogene (2010) PMID: 20661224
377. Banck MS, et al. Epigenetics (2009) PMID: 19242095
378. Collins C, et al. Proc. Natl. Acad. Sci. U.S.A. (1998) PMID: 9671742
379. Nonet GH, et al. Cancer Res. (2001) PMID: 11245413
380. Li P, et al. Int. J. Cancer (2007) PMID: 17266044
381. Wu YL, et al. Lancet Oncol. (2014) PMID: 24439929
382. Passaro et al., 2019; ELCC Abstract 1150
383. Audet et al., 2013; ASCO Abstract 6041
384. Lau SC, et al. Clin Lung Cancer (2019) PMID: 31178389
385. Paz-Ares L, et al. Ann. Oncol. (2017) PMID: 28426106
386. Thongprasert S, et al. Lung Cancer Manag (2019) PMID: 31807143
387. Januszewski et al., 2018; IASLC WCLC Abstract P1.13-17
388. Suzuki et al., 2018; IASLC WCLC Abstract P1.01-92
389. Chang et al., 2018; IASLC WCLC Abstract P1.01-11
390. Llinás-Quintero N, et al. Case Rep Oncol Med (2019) PMID: 31637072
391. Miller VA, et al. Lancet Oncol. (2012) PMID: 22452896
392. Chen X, et al. Lung Cancer (2013) PMID: 23664448
393. Katakami N, et al. J. Clin. Oncol. (2013) PMID: 23816963
394. Landi L, et al. Clin Lung Cancer (2014) PMID: 25242668
395. De Grève J, et al. Lung Cancer (2015) PMID: 25682316
396. Yang JC, et al. Lancet Oncol. (2015) PMID: 26051236
397. Horn L, et al. Lung Cancer (2017) PMID: 29110849
398. Yamamoto N, et al. Adv Ther (2020) PMID: 31863283
399. Soria JC, et al. Lancet Oncol. (2015) PMID: 26156651
400. Dziadziuszko R, et al. J Thorac Oncol (2019) PMID: 30825613
401. Lai WV, et al. Eur. J. Cancer (2019) PMID: 30685684
402. Greulich H, et al. Proc. Natl. Acad. Sci. U.S.A. (2012) PMID: 22908275
403. Gow CH, et al. J Thorac Oncol (2015) PMID: 26134234
404. Mazieres J, et al. Ann. Oncol. (2016) PMID: 26598547
405. Mazieres J, et al. J. Clin. Oncol. (2013) PMID: 23610105
406. De Grève J, et al. Lung Cancer (2012) PMID: 22325357
407. Li BT, et al. Lung Cancer (2015) PMID: 26559459
408. Costa DB, et al. J Thorac Oncol (2016) PMID: 26964772
409. Yuan B, et al. Front Oncol (2020) PMID: 32477948
410. Fang W, et al. Oncologist (2019) PMID: 31748336
411. Schuler M, et al. Ann. Oncol. (2016) PMID: 26646759
412. Wu et al., 2018; WCLC Abstract P1.01-97
413. Wolf et al., 2020; ASCO Abstract 9509
414. Schuler M, et al. Ann. Oncol. (2020) PMID: 32240796
415. Tan et al., 2017; IASLC Abstract P3.02b-117
416. van den Bent M, et al. J. Neurooncol. (2020) PMID: 31776899
417. Bang YJ, et al. Cancer Sci. (2020) PMID: 31778267
418. Esaki T, et al. Cancer Sci. (2019) PMID: 30724423
419. Vassal et al., 2015; ASCO Abstract 2595
420. Li et al., 2015; ASCO Abstract 8090
421. Frampton GM, et al. Cancer Discov (2015) PMID: 25971938
422. Paik PK, et al. Cancer Discov (2015) PMID: 25971939
423. Bendorra MA, et al. J Thorac Oncol (2016) PMID: 26845121
424. Waqar SN, et al. J Thorac Oncol (2015) PMID: 25898962
425. Mendenhall MA, et al. J Thorac Oncol (2015) PMID: 25898965
426. Jenkins RW, et al. Clin Lung Cancer (2015) PMID: 25769807
427. Stein MN, et al. Eur. Urol. (2015) PMID: 25457019
428. Shaw et al., 2016; ASCO Abstract 9066
429. Lu et al., 2016; ASCO Abstract 9058
430. Yoshida T, et al. J. Clin. Oncol. (2016) PMID: 27354483
431. Solomon BJ, et al. N. Engl. J. Med. (2014) PMID: 25470694
432. Shaw AT, et al. N. Engl. J. Med. (2013) PMID: 23724913
433. Moro-Sibilot D, et al. Ann. Oncol. (2019) PMID: 31584608
434. Goto et al., 2016; ASCO Abstract 9022
435. Shaw AT, et al. N. Engl. J. Med. (2014) PMID: 25264305
436. Mazieres J, et al. J. Clin. Oncol. (2015) PMID: 25667280
437. Scheffler M, et al. Oncotarget (2015) PMID: 25868855
438. Vaishnavi A, et al. Nat. Med. (2013) PMID: 24162815
439. Drilon et al., 2016; ASCO Abstract 108
440. Camidge et al., 2014; ASCO Abstract 8001
441. Schrock AB, et al. J Thorac Oncol (2016) PMID: 27343443
442. Jorge SE, et al. Lung Cancer (2015) PMID: 26791794
443. Mahjoubi L, et al. Invest New Drugs (2016) PMID: 26892698
444. Awad MM, et al. J. Clin. Oncol. (2016) PMID: 26729443
445. Zhang Y, et al. J Thorac Oncol (2016) PMID: 26724472
446. Landi L, et al. Clin. Cancer Res. (2019) PMID: 31416808
447. Caparica R, et al. J Thorac Oncol (2017) PMID: 27664533
448. Dagogo-Jack I, et al. Clin Cancer Res (2020) PMID: 32086345
449. Opsomer RJ, et al. Acta Urol Belg (1985) PMID: 2986437
450. Wu et al., 2018; WCLC abstract MA26.11
451. Ramalingam SS, et al. Ann. Oncol. (2016) PMID: 26768165
452. Yu HA, et al. Lung Cancer (2017) PMID: 29191595
453. Reckamp KL, et al. Cancer (2014) PMID: 24501009
454. Jänne PA, et al. Clin. Cancer Res. (2011) PMID: 21220471
455. van Geel RMJM, et al. Br. J. Cancer (2020) PMID: 32147669
456. Jänne PA, et al. J Thorac Oncol (2016) PMID: 26899759
457. Paik PK, et al. N. Engl. J. Med. (2020) PMID: 32469185
458. Mazieres et al., 2020; ESMO Abstract 1283P
459. Blanc-Durand F, et al. Oncologist (2020) PMID: 32716573
460. Cappuzzo F, et al. Lancet Oncol. (2010) PMID: 20493771
461. Zhong WZ, et al. J. Clin. Oncol. (2019) PMID: 31194613

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References

462. Petrelli F, et al. Clin Lung Cancer (2012) pmid: 22056888
463. Li JW, et al. Cancer Biol Ther (2019) pmid: 31131689
464. Yang JJ, et al. Br. J. Cancer (2017) pmid: 28103612
465. Lee CK, et al. J. Natl. Cancer Inst. (2017) pmid: 28376144
466. Nakagawa K, et al. Lancet Oncol. (2019) pmid: 31591063
467. Stinchcombe TE, et al. JAMA Oncol (2019) pmid: 31393548
468. Truini A, et al. Clin. Cancer Res. (2019) pmid: 31182434
469. Shepherd FA, et al. N. Engl. J. Med. (2005) pmid: 16014882
470. Han JY, et al. J. Clin. Oncol. (2012) pmid: 22370314
471. Maemondo M, et al. N. Engl. J. Med. (2010) pmid: 20573926
472. Mitsudomi T, et al. Lancet Oncol. (2010) pmid: 20022809
473. Mok TS, et al. N. Engl. J. Med. (2009) pmid: 19692680
474. Qi WX, et al. Curr Med Res Opin (2015) pmid: 25329826
475. Zhao H, et al. J Thorac Oncol (2015) pmid: 25546556
476. Wang J, et al. Int. J. Cancer (2019) pmid: 30255937
477. Baik CS, et al. J Thorac Oncol (2015) pmid: 26398831
478. Yoshioka H, et al. Ann. Oncol. (2019) pmid: 31553438
479. Cheng et al., 2018; ESMO Abstract 13770
480. Yang et al., 2019; AACR Abstract CT193/7
481. Fukuoka M, et al. J. Clin. Oncol. (2011) pmid: 21670455
482. Noronha V, et al. J. Clin. Oncol. (2019) pmid: 31411950
483. Hosomi Y, et al. J. Clin. Oncol. (2020) pmid: 31682542
484. Sutiman N, et al. J Thorac Oncol (2017) pmid: 27908825
485. Gibbons DL, et al. J Thorac Oncol (2016) pmid: 27198414
486. Alanazi A, et al. Lung Cancer Manag (2020) pmid: 33318755
487. Kim et al., 2021; DOI: 10.1200/PO.20.00296
488. Piotrowska et al., 2017; ASCO Abstract 90
489. Ramalingam et al., 2018; ESMO Abstract LBA50
490. Papadimitrakopoulou et al., 2018; ESMO Abstract LBA51
491. Hu et al., 2018; ASCO Abstract 9077
492. Sequist LV, et al. Lancet Oncol. (2020) pmid: 32027846
493. Ramalingam SS, et al. N. Engl. J. Med. (2019) pmid: 31751012
494. Herbst et al., 2020; ASCO Abstract LBA5
495. Kenmotsu et al., 2021; ESMO Abstract LBA44
496. Soo et al., 2021; ESMO Abstract VP3-2021
497. Oxnard GR, et al. Ann. Oncol. (2020) pmid: 32139298
498. Klemperer SJ, et al. J Thorac Oncol (2017) pmid: 27693535
499. Yakes FM, et al. Mol. Cancer Ther. (2011) pmid: 21926191
500. Weber H, et al. J Biomol Screen (2014) pmid: 25260782
501. Navis AC, et al. PLoS ONE (2013) pmid: 23484006
502. Yeh I, et al. Nat Commun (2015) pmid: 26013381
503. Lee YH, et al. Cancers (Basel) (2014) pmid: 25534569
504. Torres KE, et al. Clin. Cancer Res. (2011) pmid: 21540237
505. Sameni M, et al. Clin. Cancer Res. (2016) pmid: 26432786
506. Hellerstedt BA, et al. Clin Lung Cancer (2019) pmid: 30528315
507. Neal JW, et al. Lancet Oncol. (2016) pmid: 27825638
508. Nokihara H, et al. Clin Lung Cancer (2019) pmid: 30718102