

# Package ‘SNPtools’

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**Title** S4 Tools for Reading and Organizing Genetic Data

**Version** 0.1.0

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**Description** SNPtools provides an integrated suite of tools for handling SNP genotype data in large-scale genetic studies. The package supports importing and merging genotype files, performing quality control on SNP markers and samples, and preparing data for downstream analyses using popular software such as FImpute and PLINK. It offers S4 classes and methods to efficiently encapsulate SNP data, along with utilities for generating genotype summary statistics and visualization. Additional functionalities in the current version include anticlustering approaches for batch effect control, automated script generation for external software, and streamlined workflows for large datasets commonly encountered in animal and plant breeding programs. The package is designed to facilitate reproducible and scalable SNP data analyses in quantitative and statistical genetics.

**Logo** man/figures/logo.png

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snpStats,  
tidyverse,  
dplyr

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data.table,  
Rcpp,  
stringi

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**VignetteBuilder** knitr

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---

cbind_SnpMatrix	<i>Safe cbind for SnpMatrix preserving dimnames</i>
-----------------	---

---

**Description**

This function performs a column-wise binding of multiple `SnpMatrix` objects, explicitly preserving row names and column names, avoiding unexpected "object has no names" warnings.

**Usage**

```
cbind_SnpMatrix(...)
```

**Arguments**

...                      `SnpMatrix` objects to combine (must have identical row names).

**Value**

A single combined `SnpMatrix` with preserved row and column names.

**Examples**

```
## Not run:  
cbind_SnpMatrix(matrix1, matrix2)  
  
## End(Not run)
```

---

check.call.rate	<i>Check SNP call rate</i>
-----------------	----------------------------

---

**Description**

Identifies SNPs with call rates below a minimum threshold.

**Usage**

```
check.call.rate(summary, min.call.rate)
```

**Arguments**

summary                      A data frame with SNP summary statistics (must contain 'Call.rate' column).  
min.call.rate                Numeric value specifying the minimum acceptable call rate.

**Value**

Character vector with SNP names below threshold. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(Call.rate = c(0.85, 0.95), row.names = c("SNP1", "SNP2"))
check.call.rate(df, 0.9)
```

---

check.ibs

*Check Identity-By-State (IBS) for a genotype pair*


---

**Description**

Checks IBS status for two genotypes.

**Usage**

```
check.ibs(gen)
```

**Arguments**

gen                      Numeric vector of length two with genotype codes.

**Value**

Integer: 2 if identical non-heterozygotes, 0 if opposite homozygotes, -1 otherwise.

**Author(s)**

Roberto Higa

**Examples**

```
check.ibs(c(1, 1))
check.ibs(c(1, 3))
```

---

check.identical.samples

*Check identical samples based on distance*


---

**Description**

Identifies sample pairs considered identical based on genotype distances.

**Usage**

```
check.identical.samples(genotypes, threshold = 0)
```

**Arguments**

genotypes                Genotype matrix (samples x SNPs).  
threshold                Numeric distance threshold. Default 0.

**Value**

List of identical sample pairs.

**Author(s)**

Roberto Higa

**Examples**

```
mat <- matrix(sample(0:2, 20, TRUE), nrow = 5)
rownames(mat) <- paste0("S", 1:5)
check.identical.samples(mat, 0.5)
```

---

```
check.identical.samples.by.block
```

*Check identical samples by block*

---

**Description**

Identifies identical samples within SNP blocks.

**Usage**

```
check.identical.samples.by.block(genotypes, blcsize, threshold = 0)
```

**Arguments**

genotypes	Genotype matrix.
blcsize	Block size (number of SNPs).
threshold	Distance threshold. Default 0.

**Value**

List of identical sample pairs.

**Author(s)**

Roberto Higa

**Examples**

```
# See check.identical.samples example
```

---

```
check.mendelian.inconsistencies
```

*Check Mendelian inconsistencies*

---

**Description**

Identifies Mendelian inconsistencies between father-child pairs.

**Usage**

```
check.mendelian.inconsistencies(genotypes, father, child)
```

**Arguments**

genotypes	Genotype matrix.
father	Vector of father sample IDs.
child	Vector of child sample IDs.

**Value**

Data frame summarizing inconsistencies per pair.

**Author(s)**

Roberto Higa

**Examples**

```
# Requires proper parent-child genotype data
```

---

```
check.mendelian.inconsistencies.pair
```

*Check Mendelian inconsistencies for a pair*

---

**Description**

Calculates number of inconsistencies and total comparable SNPs for a parent-child pair.

**Usage**

```
check.mendelian.inconsistencies.pair(g1, g2)
```

**Arguments**

g1	Genotype vector for parent.
g2	Genotype vector for child.

**Value**

Numeric vector: [# inconsistencies, # comparable SNPs].

**Author(s)**

Roberto Higa

**Examples**

```
# Used internally by check.mendelian.inconsistencies
```

---

```
check.sample.call.rate
```

*Check Sample Call Rate*

---

**Description**

Identifies samples with call rate below a given threshold.

**Usage**

```
check.sample.call.rate(sample.summary, min.call.rate)
```

**Arguments**

`sample.summary` A data frame with a "Call.rate" column for each sample.

`min.call.rate` Minimum acceptable call rate (between 0 and 1).

**Value**

A character vector with the names of samples to remove.

---

```
check.sample.heterozygosity
```

*Check sample heterozygosity*

---

**Description**

Identifies samples with heterozygosity values deviating beyond a specified threshold.

**Usage**

```
check.sample.heterozygosity(sample.summary, max.dev)
```

**Arguments**

`sample.summary` Data frame containing sample summary (must have 'Heterozygosity' column).

`max.dev` Maximum number of standard deviations allowed from mean.

**Value**

Character vector with sample names considered outliers. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
ss <- data.frame(Heterozygosity = c(0.2, 0.5, 0.7))
rownames(ss) <- c("Ind1", "Ind2", "Ind3")
check.sample.heterozygosity(ss, 1)
```

---

check.snp.chromo	<i>Check SNP by chromosome</i>
------------------	--------------------------------

---

**Description**

Filters SNP names belonging to specified chromosomes.

**Usage**

```
check.snp.chromo(snpmap, chromosomes)
```

**Arguments**

snpmap	Data frame with SNP map info (must contain columns 'Chromosome' and 'Name').
chromosomes	Vector of chromosome identifiers to filter.

**Value**

Character vector with SNP names.

**Author(s)**

Roberto Higa

**Examples**

```
snpmap <- data.frame(Chromosome = c(1, 1, 2), Name = c("SNP1", "SNP2", "SNP3"))
check.snp.chromo(snpmap, 1)
```



---

check.snp.hwe	<i>Check SNP Hardy-Weinberg equilibrium deviation</i>
---------------	---

---

**Description**

Identifies SNPs deviating from HWE beyond a z-score threshold.

**Usage**

```
check.snp.hwe(snp.summary, max.dev)
```

**Arguments**

snp.summary	Data frame with SNP summary (must contain 'z.HWE' column).
max.dev	Maximum z-score allowed.

**Value**

Character vector with SNP names deviating from HWE. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(z.HWE = c(2, 5), row.names = c("SNP1", "SNP2"))
check.snp.hwe(df, 3)
```

---

check.snp.maf	<i>Check SNP minor allele frequency</i>
---------------	---

---

**Description**

Identifies SNPs with minor allele frequency below a minimum threshold.

**Usage**

```
check.snp.maf(snp.summary, min.maf)
```

**Arguments**

snp.summary	Data frame with SNP summary (must contain 'MAF' column).
min.maf	Minimum MAF allowed.

**Value**

Character vector with SNP names below threshold. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(MAF = c(0.01, 0.2), row.names = c("SNP1", "SNP2"))
check.snp.maf(df, 0.05)
```

---

check.snp.mgf

*Check SNP missing genotype frequencies*

---

**Description**

Identifies SNPs with genotype frequencies below a minimum threshold.

**Usage**

```
check.snp.mgf(snp.summary, min.mgf)
```

**Arguments**

snp.summary	Data frame with columns 'P.AA', 'P.AB', 'P.BB'.
min.mgf	Minimum genotype frequency allowed.

**Value**

Character vector with SNP names below threshold. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(P.AA = c(0.01, 0.5), P.AB = c(0.02, 0.4), P.BB = c(0.01, 0.1))
rownames(df) <- c("SNP1", "SNP2")
check.snp.mgf(df, 0.05)
```

---

check.snp.monomorf	<i>Check SNP monomorphic status</i>
--------------------	-------------------------------------

---

**Description**

Identifies SNPs considered monomorphic.

**Usage**

```
check.snp.monomorf(snp.summary)
```

**Arguments**

snp.summary      Data frame with columns 'P.AA', 'P.AB', 'P.BB'.

**Value**

Character vector with monomorphic SNP names. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(P.AA = c(1, 0.5), P.AB = c(0, 0.5), P.BB = c(0, 0))
rownames(df) <- c("SNP1", "SNP2")
check.snp.monomorf(df)
```

---

check.snp.no.position	<i>Check SNP no position</i>
-----------------------	------------------------------

---

**Description**

Identifies SNPs with position equal to zero in the SNP map.

**Usage**

```
check.snp.no.position(snpmap)
```

**Arguments**

snpmap              Data frame with columns 'Position' and 'Name'.

**Value**

Character vector with SNP names without position. Returns 'NULL' if none.

**Author(s)**

Roberto Higa

### Examples

```
df <- data.frame(Position = c(0, 100), Name = c("SNP1", "SNP2"))
check.snp.no.position(df)
```

---

```
check.snp.same.position
```

*Check SNPs mapped to the same position*

---

### Description

Identifies groups of SNPs that are mapped to the exact same genomic position on each chromosome. Returns a list where each element corresponds to one group of overlapping SNPs.

Identifies SNPs that share the same position on the same chromosome.

### Usage

```
check.snp.same.position(snpmap)
```

```
check.snp.same.position(snpmap)
```

### Arguments

snpmap                      Data frame with columns 'Chromosome', 'Position', and 'Name'.

### Value

A list of character vectors, each with names of SNPs found at the same position.

List of SNP groups sharing positions.

### Author(s)

Roberto Higa

### Examples

```
df <- data.frame(Chromosome = c(1, 1, 2), Position = c(100, 100, 200), Name = c("SNP1", "SNP2", "SNP3"))
check.snp.same.position(df)
```

---

combinarSNPData	<i>Combine multiple SNPDataLong objects</i>
-----------------	---

---

**Description**

This function merges a list of SNPDataLong objects, typically representing different SNP panels or datasets, into a single unified SNPDataLong object. It ensures that all genotype matrices have the same set of SNPs (filling missing SNPs with NA), and merges the marker map information while removing duplicate SNP entries.

**Usage**

```
combinarSNPData(lista)
```

**Arguments**

lista                      A list of SNPDataLong objects to be combined.

**Value**

A single SNPDataLong object containing the combined genotype matrix, merged map, and a concatenated path string.

**Examples**

```
## Not run:
combined <- combinarSNPData(list(snp_obj1, snp_obj2, snp_obj3))

## End(Not run)
```

---

doPCA	<i>Do genome relationship matrix PCA</i>
-------	--

---

**Description**

Performs PCA using the genome relationship matrix (GRM).

**Usage**

```
doPCA(genotypes)
```

**Arguments**

genotypes                Genotype matrix.

**Value**

List containing 'pcs' (principal components) and 'eigen' (eigenvalues).

**Author(s)**

Roberto Higa

**Examples**

```
# Requires matrix of numeric genotypes
```

---

exploratory.plots

*Exploratory plots for SNP and sample summary*

---

**Description**

Generates exploratory plots: MAF histograms, HWE plots, heterozygosity scatter, MDS, and dendrogram.

**Usage**

```
exploratory.plots(  
  snp.summary,  
  snps.plot,  
  sample.summary,  
  samples.plot,  
  distm,  
  glabels,  
  mds.plot,  
  hierq.plot  
)
```

**Arguments**

snp.summary	Data frame with SNP summary.
snps.plot	Filename for SNP histogram plot.
sample.summary	Data frame with sample summary.
samples.plot	Filename for heterozygosity plot.
distm	Distance matrix for samples.
glabels	Sample labels for plots.
mds.plot	Filename for MDS plot.
hierq.plot	Filename for hierarchical cluster plot.

**Value**

None. Plots are saved as JPEG files.

**Author(s)**

Roberto Higa

**Examples**

```
# Requires proper SNP and sample summary data frames
```

---

FImputeRunner	<i>Build FImputeRunner object</i>
---------------	-----------------------------------

---

**Description**

A convenience function to construct a ‘FImputeRunner’ object from basic inputs.

**Usage**

```
FImputeRunner(object, path, exec_path = "FImpute3", name = "data")
```

**Arguments**

path	A character string indicating the directory to save FImpute files.
exec_path	Path to the FImpute executable (default = "FImpute3").
name	Name for the dataset (used internally, default = "gen_data").
geno	A SnpMatrix object.
map	A data.frame with SNP metadata (columns: Name, Chromosome, Position).

**Value**

An object of class ‘FImputeRunner’.

---

genoToDF	<i>Convert geno slot from SNPDataLong to a data.frame</i>
----------	---

---

**Description**

Converts the genotype matrix (geno slot) of a SNPDataLong object to a data.frame, with optional centering and scaling per SNP (column).

**Usage**

```
genoToDF(object, center = FALSE, scale = FALSE)
```

**Arguments**

object	An object of class SNPDataLong.
center	Logical or numeric. If TRUE (default FALSE), center columns to mean zero.
scale	Logical or numeric. If TRUE (default FALSE), scale columns to standard deviation one.

**Value**

A data.frame with individuals as rows and SNPs as columns (numeric 0/1/2, or centered/scaled values).

**Examples**

```
## Not run:
df <- genoToDF(nelore_imputed, center = TRUE, scale = TRUE)
head(df[, 1:5])

## End(Not run)
```

---

get.correl.fc	<i>Get correlation (fc method)</i>
---------------	------------------------------------

---

**Description**

Calculates genotype correlation using a fast check (fc) method.

**Usage**

```
get.correl.fc(g1, g2)
```

**Arguments**

g1	Genotype vector.
g2	Genotype vector.

**Value**

Numeric value of correlation.

**Author(s)**

Roberto Higa

**Examples**

```
g1 <- sample(0:2, 10, TRUE)
g2 <- sample(0:2, 10, TRUE)
get.correl.fc(g1, g2)
```

---

get.gender	<i>Get gender based on heterozygosity</i>
------------	---

---

**Description**

Infers gender using heterozygosity thresholds.

**Usage**

```
get.gender(sample.summary, threshM, threshF)
```



**Arguments**

sample.summary    Data frame with 'Heterozygosity' column.  
threshM            Numeric threshold for males.  
threshF            Numeric threshold for females.

**Value**

Data frame with columns 'heterozygosity' and 'sex'.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(Heterozygosity = c(0.1, 0.3, 0.6))  
rownames(df) <- c("A", "B", "C")  
get.gender(df, 0.2, 0.5)
```

---

get.hwe.chi2

*Get HWE chi-square p-values*

---

**Description**

Calculates Hardy-Weinberg equilibrium chi-square p-values for SNPs.

**Usage**

```
get.hwe.chi2(snp.summary)
```

**Arguments**

snp.summary        Data frame with columns 'Calls', 'P.AA', 'P.AB', 'P.BB'.

**Value**

Numeric vector with p-values.

**Author(s)**

Roberto Higa

**Examples**

```
df <- data.frame(Calls = c(100, 100), P.AA = c(0.6, 0.4), P.AB = c(0.3, 0.4), P.BB = c(0.1, 0.2))  
get.hwe.chi2(df)
```

---

getGeno	<i>Flexible and efficient genotype file reading with autodetection using fread</i>
---------	--

---

### Description

This generic method allows flexible import of SNP genotype data from Illumina FinalReport files, supporting fast initial column detection using `data.table::fread`, followed by full genotype matrix construction via `snpStats::read.snps.long`.

### Usage

```
getGeno(...)
```

### Arguments

path	Path to the directory containing <code>FinalReport.txt</code>
fields	A list specifying column indices for sample, SNP, allele1, allele2, and confidence
codes	A character vector with allele codes (e.g., <code>c("A", "B")</code> )
threshold	Confidence threshold for genotype calling
sep	Field separator used in the files
skip	Number of lines to skip at the start of the file
verbose	Logical; if TRUE, displays progress messages
every	Frequency of progress update (number of SNPs)

### Value

An `SNPDataLong` object containing the genotype matrix and map, or NULL if an error occurs

---

ibs.pair	<i>IBS pair statistics</i>
----------	----------------------------

---

### Description

Calculates IBS mean and standard deviation between two samples.

### Usage

```
ibs.pair(g1, g2)
```

### Arguments

g1	Genotype vector for first sample.
g2	Genotype vector for second sample.

**Value**

Numeric vector: [mean IBS, standard deviation].

**Author(s)**

Roberto Higa

**Examples**

```
g1 <- sample(0:2, 10, TRUE)
g2 <- sample(0:2, 10, TRUE)
ibs.pair(g1, g2)
```

---

importAllGenos

---

*Import and combine multiple genotype configurations*


---

**Description**

This generic and method import genotype data from multiple configurations defined in an `SNPImportList` object, then combine them into a single unified `SNPDataLong` object.

**Usage**

```
importAllGenos(object)
```

**Arguments**

`object`                      An object of class `SNPImportList` containing import configurations

**Value**

A single combined `SNPDataLong` object

---

importFImputeResults

---

*Import imputed FImpute results from disk*


---

**Description**

Reads existing imputed results from a given path and returns an object of class `SNPDataLong`.

**Usage**

```
importFImputeResults(path, method = "R")
```

**Arguments**

`path`                      Character. Path to the folder containing 'output\_fimpute' (e.g., "fimpute\_run\_nelore").  
`method`                    Character. "R" (default) or "Rcpp". Passed to `read.fimpute()`.

**Value**

An object of class `SNPDataLong` containing the imputed genotypes and SNP map.

---

import_geno_list	<i>Import multiple genotype datasets from a list of configurations</i>
------------------	--

---

### Description

Reads and imports multiple genotype datasets specified in a list of configurations. Each configuration must include the path to the genotype data and information on field mapping. Optionally, you can also specify codes, quality threshold, separator, lines to skip, and a subset of IDs to retain. The function automatically fills the 'xref\_path' slot per individual and combines maps into a single data.frame, adding a 'SourcePath' column indicating their origin and removing duplicated SNP rows (by Name). Prints progress messages indicating the current path being loaded (with counter).

### Usage

```
import_geno_list(config_list)
```

### Arguments

config_list	A list of configuration lists. Each element should contain: - 'path' (character): Path to the genotype file or folder. - 'fields' (list): Named list defining the columns (e.g., SNP ID, sample ID, alleles, confidence). - 'codes' (character vector, optional): Allele codes (default is c("A", "B")). - 'threshold' (numeric, optional): Maximum allowed missingness or confidence threshold (default 0.15). - 'sep' (character, optional): Field separator in the input file (default "tab-delimited"). - 'skip' (integer, optional): Number of lines to skip at the beginning of the file (default 0). - 'verbose' (logical, optional): Whether to print detailed messages (default TRUE). - 'subset' (character vector, optional): Vector of sample IDs to retain after import.
-------------	---

### Value

An object of class 'SNPDataLong' containing: - Combined genotype matrix ('geno'). - Combined map ('map') as a single data.frame with 'SourcePath' column and without duplicated rows. - Combined 'xref\_path' vector (one entry per individual). - 'path' slot as a semicolon-separated string of all input dataset paths.

---

pairs2sets	<i>Convert pairs to sets</i>
------------	------------------------------

---

### Description

Groups sample pairs into sets of related samples.

### Usage

```
pairs2sets(pairs)
```

### Arguments

pairs	Matrix or list of sample pairs.
-------	---------------------------------

**Value**

List of sets of samples.

**Author(s)**

Roberto Higa

**Examples**

```
pairs <- matrix(c("A", "B", "B", "C", "D", "E"), ncol = 2, byrow = TRUE)
pairs2sets(pairs)
```

---

plotPCAgroups

---

*Plot PCA groups from anticlustering result*


---

**Description**

Plot PCA groups from anticlustering result

**Usage**

```
plotPCAgroups(pca_res, groups, pcs = c(1, 2), filename = NULL)
```

**Arguments**

pca_res	A prcomp object.
groups	A factor or vector of group assignments.
pcs	Vector of length 2 indicating which PCs to plot (default: c(1, 2)).
filename	Optional. If provided, saves plot to this file (e.g., "antic.png").

**Value**

A ggplot object (also prints to screen).

**Examples**

```
## Not run:
res <- runAnticlusteringPCA(nelore_imputed, K = 2, n_pcs = 20)
plotPCAgroups(res$pca, res$groups)

## End(Not run)
```

qcSamples

*Quality control on samples***Description**

Applies quality control (QC) procedures to samples in a ‘SNPDataLong’ object, based on heterozygosity and call rate thresholds.

**Usage**

```
qcSamples(x, ...)

## S4 method for signature 'SNPDataLong'
qcSamples(
  x,
  heterozygosity = NULL,
  smp_cr = NULL,
  action = c("report", "filter", "both")
)
```

**Arguments**

x	An object of class ‘SNPDataLong’.
heterozygosity	A numeric threshold or range for heterozygosity. Samples outside this threshold are removed.
smp_cr	Minimum acceptable sample call rate (between 0 and 1). Samples below this value are removed.
action	Character string indicating the action to perform. One of: - “report”: only returns a list of samples to remove and those kept; - “filter”: returns a filtered object without reporting; - “both”: performs filtering and returns the filtered object.

**Value**

Depending on the ‘action’ argument: - “report”: returns a list with removed and kept samples; - “filter”: returns a new ‘SNPDataLong’ object with filtered genotypes; - “both”: returns a list with: - ‘filtered’: the filtered ‘SNPDataLong’ object; - ‘report’: a list of removed and kept samples.

qcSNPs

*Quality Control for SNPDataLong with optional criteria***Description**

Applies flexible quality control filters on an object of class SNPDataLong. Supports call rate filtering, minor allele frequency (MAF), Hardy-Weinberg equilibrium (HWE), removal of monomorphic SNPs, exclusion of specific chromosomes, optionally removing SNPs without positions, and optionally removing SNPs at the same genomic position (keeping the one with highest MAF).

**Usage**

```
qcSNPs(x, ...)
```

**Arguments**

<code>x</code>	An object of class <code>SNPDataLong</code> .
<code>missing_ind</code>	Maximum allowed proportion of missing data per individual (currently not implemented).
<code>missing_snp</code>	Maximum allowed proportion of missing data per SNP (currently not implemented).
<code>min_snp_cr</code>	Minimum acceptable call rate for SNPs (e.g., 0.95). SNPs below this threshold are removed.
<code>min_maf</code>	Minimum minor allele frequency allowed for SNPs (e.g., 0.05). SNPs with lower MAF are removed.
<code>hwe</code>	p-value threshold for Hardy-Weinberg equilibrium test (e.g., 1e-6). SNPs violating this are removed.
<code>snp_position</code>	Logical. If TRUE, removes SNPs mapped to the same position, retaining only the one with highest MAF.
<code>no_position</code>	Logical. If TRUE, removes SNPs without defined genomic positions.
<code>snp_mono</code>	Logical. If TRUE, removes monomorphic SNPs (with no variation).
<code>remove_chr</code>	Character vector of chromosomes to exclude (e.g., <code>c("X", "Y")</code> ).
<code>action</code>	One of "report" (returns a list of removed SNPs), "filter" (returns filtered <code>SNPDataLong</code> ), or "both" (returns both).

**Value**

Depending on the `action` argument: - "report": list of SNPs removed by each filter and SNPs retained. - "filter": filtered `SNPDataLong` object. - "both": list containing the filtered object and detailed report.

**Examples**

```
## Not run:
set.seed(123)
mat <- matrix(sample(c(0, 1, 2, NA), 100, replace = TRUE, prob = c(0.4, 0.4, 0.15, 0.05)),
              nrow = 10, ncol = 10)
colnames(mat) <- paste0("snp", 1:10)
rownames(mat) <- paste0("ind", 1:10)
map <- data.frame(Name = colnames(mat), Chromosome = 1, Position = 1:10)
x <- new("SNPDataLong", geno = mat, map = map, path = "dummy_path", xref_path = rep("chip1", 10))

# Example using multiple filters
qcSNPs(x, min_snp_cr = 0.8, min_maf = 0.05, snp_mono = TRUE, no_position = TRUE, snp_position = TRUE, action = "both")

## End(Not run)
```

---

qc_header	<i>Formatted header message</i>
-----------	---------------------------------

---

**Description**

Prints a formatted message with a border for section titles in the console.

**Usage**

```
qc_header(title)
```

**Arguments**

title                      Character string to be printed inside the header box.

**Value**

No return value. Used for side effects (message).

**Examples**

```
qc_header("Quality Control on Samples")
```

---

rbindSnpFlexible	<i>Faster row-bind for SnpMatrix objects with differing columns</i>
------------------	---

---

**Description**

Combines multiple SnpMatrix objects by rows, automatically handling differing SNP columns, optimized for large matrices.

**Usage**

```
rbindSnpFlexible(...)
```

**Arguments**

...                      One or more SnpMatrix objects.

**Value**

A single SnpMatrix object with all rows combined.

**Examples**

```
## Not run:
combined <- rbindSnpFlexible(brangus_geno, batch_BM@geno)

## End(Not run)
```



---

rbind_SnpMatrix	<i>Safe rbind for SnpMatrix preserving dimnames</i>
-----------------	---

---

**Description**

This function performs a row-wise binding of multiple SnpMatrix objects, explicitly preserving row names and column names, avoiding unexpected "object has no names" warnings.

**Usage**

```
rbind_SnpMatrix(...)
```

**Arguments**

... SnpMatrix objects to combine (must have identical column names).

**Value**

A single combined SnpMatrix with preserved row and column names.

**Examples**

```
## Not run:
rbind_SnpMatrix(matrix1, matrix2)

## End(Not run)
```

---

read.fimpute	<i>Read imputed genotypes from FImpute output and return SNPData-Long object</i>
--------------	--

---

**Description**

Reads imputed genotypes and SNP information from FImpute output, builds a SnpMatrix and a corresponding map, and returns an SNPDataLong object.

**Usage**

```
read.fimpute(file, method = c("R", "Rcpp"))
```

**Arguments**

file	Character. Path to the FImpute output directory (usually "output_fimpute").
method	Character. "R" (default) for vectorized R implementation, or "Rcpp" for compiled C++ implementation.

**Value**

An object of class SNPDataLong containing the imputed genotypes and SNP map.

**Examples**

```
## Not run:
snp_long <- read.fimpute("output_fimpute", method = "R")

## End(Not run)
```

---

runAnticlusteringPCA    *Run PCA and Anticlustering on SNPDataLong*

---

**Description**

Converts a SNPDataLong object to a data.frame, runs PCA, and performs anticlustering grouping.

**Usage**

```
runAnticlusteringPCA(object, K = 2, n_pcs = 20, center = TRUE, scale = TRUE)
```

**Arguments**

object	An object of class SNPDataLong.
K	Number of groups for anticlustering.
n_pcs	Number of top principal components to use (default: 20).
center	Logical or numeric. Center columns before PCA (default: TRUE).
scale	Logical or numeric. Scale columns before PCA (default: TRUE).

**Value**

A list with: - groups: vector with group assignments. - pca: the PCA result object (prcomp). - pcs: matrix of top PCs used in anticlustering.

**Examples**

```
## Not run:
res <- runAnticlusteringPCA(nelore_imputed, K = 2, n_pcs = 20)
table(res$groups)

## End(Not run)
```

---

`runFImpute`*Run FImpute from a FImputeRunner object*

---

### Description

This function runs the external FImpute software using a 'FImputeRunner' object, ensuring that all required input files are present and the results are imported.

### Usage

```
runFImpute(object, verbose = TRUE)

## S4 method for signature 'FImputeRunner'
runFImpute(object, verbose = TRUE)
```

### Arguments

<code>object</code>	An object of class 'FImputeRunner'.
<code>verbose</code>	Logical. If TRUE (default), FImpute output will be printed to the console.

### Value

An updated 'FImputeRunner' object with the 'results' slot populated (SnpMatrix).

### Examples

```
## Not run:
# Example: Running FImpute from a FImputeRunner object

path_fimpute <- "fimpute_run_example"
param_file <- file.path(path_fimpute, "fimpute.par")
fimpute_exec <- "FImpute3" # assuming it is in PATH

export_obj <- new("FImputeExport",
                  geno = geno_obj@geno,
                  map = geno_obj@map,
                  path = path_fimpute)

runner <- new("FImputeRunner",
             export = export_obj,
             par_file = param_file,
             exec_path = fimpute_exec)

runner <- runFImpute(runner, verbose = TRUE)
head(runner@results)

## End(Not run)
```

---

saveFImpute	<i>Save genotype and map files in FImpute format</i>
-------------	--

---

### Description

S4 method to export genotype (.gen), map (.map), and parameter (fimpute.par) files compatible with [FImpute](<https://www.aps.uoguelph.ca/~msargol/fimpute/>).

### Usage

```
saveFImpute(object, ...)

## S4 method for signature 'FImputeExport'
saveFImpute(object)

## S4 method for signature 'SNPDataLong'
saveFImpute(object, path = NULL)
```

### Arguments

object	An object of class 'FImputeExport' or 'SNPDataLong'.
...	Additional arguments passed to methods.
path	Output directory (default: "fimpute_run" for SNPDataLong).

### Value

No return value. Files are saved to disk.

---

saveFImputeRaw	<i>Export genotypes and map using basic arguments</i>
----------------	---

---

### Description

Convenience function to export FImpute files directly from a 'SnpMatrix' and map 'data.frame'.

### Usage

```
saveFImputeRaw(geno, map, path, xref = NULL)
```

### Arguments

geno	A 'SnpMatrix' object.
map	A data.frame with columns 'Name', 'Chromosome', 'Position', and 'SourcePath'.
path	Output directory.
xref	Optional vector of identifiers per individual (used to assign numeric chip IDs).

---

savePlink	<i>Save SNPDataLong object to PLINK format</i>
-----------	--

---

### Description

Saves genotype and map data from an SNPDataLong object in PLINK format (.ped/.map and optionally binary files).

### Usage

```
savePlink(
  object,
  path = "plink_out",
  name = "plink_data",
  run_plink = TRUE,
  chunk_size = 1000
)
```

### Arguments

object	An object of class SNPDataLong.
path	Character. Directory where files will be saved.
name	Character. Base name for PLINK output files.
run_plink	Logical. If TRUE (default), runs PLINK1 to convert to binary files. If FALSE, only .ped and .map files are saved.
chunk_size	Integer. Number of individuals per chunk for writing .ped file (default: 1000).

### Value

No return value. Files are saved to disk.

### Examples

```
## Not run:
savePlink(genotypes_qc, path = "plink_out", name = "nelore_qc", run_plink = TRUE, chunk_size = 2000)

## End(Not run)
```

---

Subset	<i>Subset method for SNPDataLong</i>
--------	--------------------------------------

---

### Description

Subset method for SNPDataLong

### Usage

```
Subset(object, index, margin = 1, keep = TRUE)
```

**Arguments**

object	A SNPDataLong object.
index	Character vector with row (individual) or column (SNP) names to filter.
margin	Integer: 1 = rows (individuals), 2 = columns (SNPs).
keep	Logical: TRUE to keep specified levels, FALSE to discard them.

**Value**

A new SNPDataLong object, subsetting accordingly.

---

summary,SNPDataLong-method

*Summary for SNPDataLong objects*

---

**Description**

Provides a detailed summary of an SNPDataLong object, including sample and SNP counts, proportion of missing data, and SNP distribution by chromosome if mapping information is available.

**Usage**

```
## S4 method for signature 'SNPDataLong'  
summary(object, ...)
```

**Arguments**

object	An object of class SNPDataLong.
--------	---------------------------------

**Value**

Prints a summary to the console. Returns NULL (invisible).

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