

# Package ‘SNPtools’

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**Title** S4 Tools for Reading and Organizing Genetic Data

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**Maintainer** Vinícius Junqueira <junqueiravinicius@hotmail.com>

**Description** Provides S4-based structures to encapsulate the import, organization, and processing of genetic data from PLINK and FImpute files, with customizable arguments. Includes tools for combining SNP panels, summarizing genotype data, and facilitating downstream quality control and analysis workflows.

**Depends** R (>= 4.1.0),  
snpStats,  
tidyverse

**Imports** methods,  
data.table,  
fQC

**Suggests** knitr,  
rmarkdown

**VignetteBuilder** knitr

**Encoding** UTF-8

**License** GPL-3

**RoxygenNote** 7.3.2

## Contents

check.sample.call.rate . . . . .	2
combinarSNPData . . . . .	2
FImputeRunner . . . . .	3
genoToDF . . . . .	4
getGeno . . . . .	5
importAllGenos . . . . .	5
importFImputeResults . . . . .	6
import_genos_list . . . . .	6
plotPCAgroups . . . . .	7
qcSamples . . . . .	8

qcSNPs . . . . .	9
qc_header . . . . .	10
rbindSnpFlexible . . . . .	10
read.fimpute . . . . .	11
runAnticlusteringPCA . . . . .	12
runFImpute . . . . .	12
saveFImpute . . . . .	13
saveFImputeRaw . . . . .	14
savePlink . . . . .	15
summary,SNPDataLong-method . . . . .	16
<b>Index</b>	<b>17</b>

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check.sample.call.rate	<i>Check Sample Call Rate</i>
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**Description**

Identifies samples with call rate below a given threshold.

**Usage**

check.sample.call.rate(sample.summary, min.call.rate)

**Arguments**

- sample.summary    A data frame with a "Call.rate" column for each sample.
- min.call.rate    Minimum acceptable call rate (between 0 and 1).

**Value**

A character vector with the names of samples to remove.

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combinarSNPData	<i>Combine multiple SNPDataLong objects</i>
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**Description**

This function merges a list of SNPDataLong objects, typically representing different SNP panels or datasets, into a single unified SNPDataLong object. It ensures that all genotype matrices have the same set of SNPs (filling missing SNPs with NA), and merges the marker map information while removing duplicate SNP entries.

**Usage**

combinarSNPData(lista)

**Arguments**

`lista`                      A list of SNPDataLong objects to be combined.

**Details**

The function performs the following steps internally:

1. Computes the union of all SNPs across input objects.
2. Fills missing SNP columns in each genotype matrix with NA-coded columns.
3. Combines genotype matrices by rows (individuals).
4. Merges marker maps, removing duplicates (retaining the first occurrence).
5. Creates and returns a new SNPDataLong object with the combined data.

**Value**

A single SNPDataLong object containing the combined genotype matrix, merged map, and a concatenated path string.

**Examples**

```
## Not run:
# Example usage:
combined_data <- combinarSNPData(list(snp_data1, snp_data2, snp_data3))

## End(Not run)
```

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FImputeRunner	<i>Build FImputeRunner object</i>
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**Description**

A convenience function to construct a ‘FImputeRunner’ object from basic inputs.

**Usage**

```
FImputeRunner(object, path, exec_path = "FImpute3", name = "data")
```

**Arguments**

<code>path</code>	A character string indicating the directory to save FImpute files.
<code>exec_path</code>	Path to the FImpute executable (default = "FImpute3").
<code>name</code>	Name for the dataset (used internally, default = "gen_data").
<code>geno</code>	A SnpMatrix object.
<code>map</code>	A data.frame with SNP metadata (columns: Name, Chromosome, Position).

**Value**

An object of class 'FImputeRunner'.

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genoToDF

*Convert geno slot from SNPDataLong to a data.frame*

---

**Description**

Converts the genotype matrix (geno slot) of a SNPDataLong object to a data.frame, with optional centering and scaling per SNP (column).

**Usage**

```
genoToDF(object, center = FALSE, scale = FALSE)
```

**Arguments**

object	An object of class SNPDataLong.
center	Logical or numeric. If TRUE (default FALSE), center columns to mean zero.
scale	Logical or numeric. If TRUE (default FALSE), scale columns to standard deviation one.

**Value**

A data.frame with individuals as rows and SNPs as columns (numeric 0/1/2, or centered/scaled values).

**Examples**

```
## Not run:  
df <- genoToDF(nelore_imputed, center = TRUE, scale = TRUE)  
head(df[, 1:5])  
  
## End(Not run)
```

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getGeno	<i>Flexible and efficient genotype file reading with autodetection using fread</i>
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### Description

This generic and method allow flexible import of SNP genotype data from Illumina FinalReport files, supporting fast initial column detection using `data.table::fread`, followed by full genotype matrix construction via `snpStats::read.snps.long`.

### Usage

```
getGeno(...)
```

### Arguments

path	Path to the directory containing FinalReport.txt
fields	A list specifying column indices for sample, SNP, allele1, allele2, and confidence
codes	A character vector with allele codes (e.g., <code>c("A", "B")</code> )
threshold	Confidence threshold for genotype calling
sep	Field separator used in the files
skip	Number of lines to skip at the start of the file
verbose	Logical; if TRUE, displays progress messages
every	Frequency of progress update (number of SNPs)

### Value

An `SNPDataLong` object containing the genotype matrix and map, or NULL if an error occurs

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importAllGenos	<i>Import and combine multiple genotype configurations</i>
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### Description

This generic and method import genotype data from multiple configurations defined in an `SNPImportList` object, then combine them into a single unified `SNPDataLong` object.

### Usage

```
importAllGenos(object)
```

**Arguments**

object                    An object of class SNPImportList containing import configurations

**Value**

A single combined SNPDataLong object

---

importFImputeResults    *Import imputed FImpute results from disk*

---

**Description**

Reads existing imputed results from a given path and returns an object of class SNPDataLong.

**Usage**

```
importFImputeResults(path)
```

**Arguments**

path                    Character. Path to the folder containing 'output\_fimpute' (e.g., "fimpute\_run\_nelore").

**Value**

An object of class SNPDataLong containing the imputed genotypes and SNP map.

**Examples**

```
## Not run:
imputed_obj <- importFImputeResults("fimpute_run_nelore")
head(imputed_obj@map)

## End(Not run)
```

---

import\_geno\_list        *Import multiple genotype datasets from a list of configurations*

---

**Description**

This function iterates over a list of configuration lists (each specifying parameters such as path, fields, separators, etc.), imports each genotype dataset using getGeno(), and then combines them into a single SNPDataLong object.

**Usage**

```
import_geno_list(config_list)
```

**Arguments**

`config_list` A list of configuration lists. Each element must include at least `path` and `fields`. Optional elements include `codes`, `threshold`, `sep`, `skip`, and `verbose`.

**Value**

A unified `SNPDataLong` object containing combined genotype data from all configurations.

**Examples**

```
## Not run:
configs <- list(
  list(path = "panel1", fields = list(sample = 2, snp = 1, allele1 = 7, allele2 = 8, confidence = 9)),
  list(path = "panel2", fields = list(sample = 2, snp = 1, allele1 = 7, allele2 = 8, confidence = 9), threshold = 0.10)
)
combined_data <- import_genotype_list(configs)

## End(Not run)
```

---

plotPCAgroups

---

*Plot PCA groups from anticlustering result*


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**Description**

Plot PCA groups from anticlustering result

**Usage**

```
plotPCAgroups(pca_res, groups, pcs = c(1, 2), filename = NULL)
```

**Arguments**

`pca_res` A `prcomp` object.  
`groups` A factor or vector of group assignments.  
`pcs` Vector of length 2 indicating which PCs to plot (default: `c(1, 2)`).  
`filename` Optional. If provided, saves plot to this file (e.g., "antic.png").

**Value**

A `ggplot` object (also prints to screen).

**Examples**

```
## Not run:
res <- runAnticlusteringPCA(nelore_imputed, K = 2, n_pcs = 20)
plotPCAgroups(res$pca, res$groups)

## End(Not run)
```

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qcSamples	<i>Quality control on samples</i>
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## Description

Applies quality control (QC) procedures to samples in a ‘SNPDataLong’ object, based on heterozygosity and call rate thresholds.

## Usage

```
qcSamples(x, ...)

## S4 method for signature 'SNPDataLong'
qcSamples(
  x,
  heterozygosity = NULL,
  smp_cr = NULL,
  action = c("report", "filter", "both")
)
```

## Arguments

x	An object of class ‘SNPDataLong’.
heterozygosity	A numeric threshold or range for heterozygosity. Samples outside this threshold are removed.
smp_cr	Minimum acceptable sample call rate (between 0 and 1). Samples below this value are removed.
action	Character string indicating the action to perform. One of: - “report”: only returns a list of samples to remove and those kept; - “filter”: returns a filtered object without reporting; - “both”: performs filtering and returns the filtered object.

## Value

Depending on the ‘action’ argument: - “report”: returns a list with removed and kept samples; - “filter”: returns a new ‘SNPDataLong’ object with filtered genotypes; - “both”: returns a list with: - ‘filtered’: the filtered ‘SNPDataLong’ object; - ‘report’: a list of removed and kept samples.



**Description**

Allows applying genotypic quality filters with user-defined criteria, including call rate, MAF, HWE, monomorphism, chromosome filtering, and removal of SNPs at the same genomic position.

**Usage**

```
qcSNPs(x, ...)
```

**Arguments**

x	An object of class SNPDataLong.
missing_ind	Maximum allowed proportion of missing data per individual (optional). *[Currently not implemented in this function]*
missing_snp	Maximum allowed proportion of missing data per SNP (optional). *[Currently not implemented in this function]*
min_snp_cr	Minimum acceptable call rate for SNPs.
min_maf	Minimum minor allele frequency allowed for SNPs (optional).
hwe	p-value threshold for Hardy-Weinberg equilibrium test (optional).
snp_position	Logical. If TRUE, removes SNPs mapped to the same position, keeping the one with the highest MAF.
snp_mono	Logical. If TRUE, identifies and removes monomorphic SNPs.
remove_chr	Character vector of chromosomes to exclude (optional).
action	One of "report" (returns a list of removed SNPs), "filter" (returns a filtered SNPDataLong object), or "both" (returns both).

**Value**

Depending on the action argument: - "report": list with SNPs removed by each criterion and SNPs kept. - "filter": filtered SNPDataLong object. - "both": list with the filtered object and detailed report.

**Examples**

```
## Not run:
set.seed(123)
mat <- matrix(sample(c(0, 1, 2, NA), 100, replace = TRUE, prob = c(0.4, 0.4, 0.15, 0.05)),
              nrow = 10, ncol = 10)
colnames(mat) <- paste0("snp", 1:10)
rownames(mat) <- paste0("ind", 1:10)
map <- data.frame(Name = colnames(mat), Chrom = 1, Position = 1:10)
x <- new("SNPDataLong", geno = mat, map = map)
```

```
qcSNPs(x, min_snp_cr = 0.8, min_maf = 0.05, snp_mono = TRUE, snp_position = TRUE, action = "report")

## End(Not run)
```

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qc_header	<i>Formatted header message</i>
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### Description

Prints a formatted message with a border for section titles in the console.

### Usage

```
qc_header(title)
```

### Arguments

title                      Character string to be printed inside the header box.

### Value

No return value. Used for side effects (message).

### Examples

```
qc_header("Quality Control on Samples")
```

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rbindSnpFlexible	<i>Faster row-bind for SnpMatrix objects with differing columns</i>
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### Description

Combines multiple SnpMatrix objects by rows, automatically handling differing SNP columns, optimized for large matrices.

### Usage

```
rbindSnpFlexible(...)
```

### Arguments

...                      One or more SnpMatrix objects.

**Value**

A single SnpMatrix object with all rows combined.

**Examples**

```
## Not run:
combined <- rbindSnpFlexible(brangus_geno, batch_BM@geno)

## End(Not run)
```

---

read.fimpute	<i>Read imputed genotypes from FImpute output and return SNPData-Long object</i>
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---

**Description**

Reads imputed genotypes and SNP information from FImpute output, builds a SnpMatrix and a corresponding map, and returns an SNPDataLong object.

**Usage**

```
read.fimpute(file)
```

**Arguments**

file                      Character. Path to the FImpute output directory (usually "output\_fimpute").

**Value**

An object of class SNPDataLong containing the imputed genotypes and SNP map.

**Examples**

```
## Not run:
snp_long <- read.fimpute("output_fimpute")

## End(Not run)
```

---

runAnticlusteringPCA     *Run PCA and Anticlustering on SNPDataLong*

---

### Description

Converts a SNPDataLong object to a data.frame, runs PCA, and performs anticlustering grouping.

### Usage

```
runAnticlusteringPCA(object, K = 2, n_pcs = 20, center = TRUE, scale = TRUE)
```

### Arguments

object	An object of class SNPDataLong.
K	Number of groups for anticlustering.
n_pcs	Number of top principal components to use (default: 20).
center	Logical or numeric. Center columns before PCA (default: TRUE).
scale	Logical or numeric. Scale columns before PCA (default: TRUE).

### Value

A list with: - groups: vector with group assignments. - pca: the PCA result object (prcomp). - pcs: matrix of top PCs used in anticlustering.

### Examples

```
## Not run:
res <- runAnticlusteringPCA(nelore_imputed, K = 2, n_pcs = 20)
table(res$groups)

## End(Not run)
```

---

runFImpute     *Run FImpute from a FImputeRunner object*

---

### Description

This function runs the external FImpute software using a 'FImputeRunner' object, ensuring that all required input files are present and the results are imported.

### Usage

```
runFImpute(object, verbose = TRUE)

## S4 method for signature 'FImputeRunner'
runFImpute(object, verbose = TRUE)
```

**Arguments**

object            An object of class 'FImputeRunner'.  
 verbose          Logical. If TRUE (default), FImpute output will be printed to the console.

**Value**

An updated 'FImputeRunner' object with the 'results' slot populated (SnpMatrix).

**Examples**

```
## Not run:
# Example: Running FImpute from a FImputeRunner object

path_fimpute <- "fimpute_run_example"
param_file <- file.path(path_fimpute, "fimpute.par")
fimpute_exec <- "FImpute3" # assuming it is in PATH

export_obj <- new("FImputeExport",
                  geno = geno_obj@geno,
                  map = geno_obj@map,
                  path = path_fimpute)

runner <- new("FImputeRunner",
              export = export_obj,
              par_file = param_file,
              exec_path = fimpute_exec)

runner <- runFImpute(runner, verbose = TRUE)
head(runner@results)

## End(Not run)
```

---

saveFImpute	<i>Save genotype and map files in FImpute format</i>
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---

**Description**

S4 method to export genotype ('.gen'), map ('.map'), and parameter ('data.par') files compatible with the [FImpute](<https://www.aps.uoguelph.ca/~msargol/fimpute/>) software.

**Usage**

```
saveFImpute(object, ...)
```

## S4 method for signature 'FImputeExport'

```
saveFImpute(object)
```

## S4 method for signature 'SNPDataLong'

```
saveFImpute(object, path = NULL)
```

**Arguments**

object	An object of class 'FImputeExport' or 'SNPDataLong'.
...	Further arguments passed to methods.
path	Character. Output directory where files will be written (only for 'SNPDataLong' method; default = "fimpute_run").

**Value**

No return value. Files are saved to disk.

**Examples**

```
## Not run:
if (requireNamespace("snpStats", quietly = TRUE)) {
  mat <- matrix(sample(c(0L, 1L, 2L), 50, replace = TRUE), nrow = 5)
  colnames(mat) <- paste0("snp", 1:10)
  rownames(mat) <- paste0("ind", 1:5)

  sm <- new("SnpMatrix", data = as.raw(mat))
  map <- data.frame(Name = colnames(mat), Chromosome = 1, Position = 1:10)
  x <- new("SNPDataLong", geno = sm, map = map)

  saveFImpute(x, path = tempdir())
}

## End(Not run)
```

---

saveFImputeRaw

*Export genotypes and map using basic arguments*


---

**Description**

Convenience function to export FImpute files directly from a 'SnpMatrix' and map 'data.frame'.

**Usage**

```
saveFImputeRaw(geno, map, path)
```

**Arguments**

geno	A 'SnpMatrix' object (from the 'snpStats' package).
map	A 'data.frame' with columns 'Name', 'Chromosome', and 'Position'.
path	Path where the files will be saved.

**Value**

No return value. Files are saved to disk.

---

savePlink	<i>Save SNPDataLong object to PLINK format</i>
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---

## Description

Saves genotype and map data from an SNPDataLong object in PLINK format (.ped/.map and optionally binary files).

## Usage

```
savePlink(  
  object,  
  path = "plink_out",  
  name = "plink_data",  
  run_plink = TRUE,  
  chunk_size = 1000  
)
```

## Arguments

object	An object of class SNPDataLong.
path	Character. Directory where files will be saved.
name	Character. Base name for PLINK output files.
run_plink	Logical. If TRUE (default), runs PLINK1 to convert to binary files. If FALSE, only .ped and .map files are saved.
chunk_size	Integer. Number of individuals per chunk for writing .ped file (default: 1000).

## Value

No return value. Files are saved to disk.

## Examples

```
## Not run:  
savePlink(genotypes_qc, path = "plink_out", name = "nelore_qc", run_plink = TRUE, chunk_size = 2000)  
  
## End(Not run)
```

---

summary,SNPDataLong-method

*Summary for SNPDataLong objects*

---

### Description

Provides a detailed summary of an SNPDataLong object, including sample and SNP counts, proportion of missing data, and SNP distribution by chromosome if mapping information is available.

### Usage

```
## S4 method for signature 'SNPDataLong'  
summary(object, ...)
```

### Arguments

object            An object of class SNPDataLong.

### Value

Prints a summary to the console. Returns NULL (invisible).



# Index

`check.sample.call.rate`, [2](#)  
`combinarSNPData`, [2](#)  
  
`FImputeRunner`, [3](#)  
  
`genoToDF`, [4](#)  
`getGeno`, [5](#)  
  
`import_geno_list`, [6](#)  
`importAllGenos`, [5](#)  
`importFImputeResults`, [6](#)  
  
`plotPCAgroups`, [7](#)  
  
`qc_header`, [10](#)  
`qcSamples`, [8](#)  
`qcSamples`, `SNPDataLong`-method  
    (`qcSamples`), [8](#)  
`qcSNPs`, [9](#)  
  
`rbindSnpFlexible`, [10](#)  
`read.fimpute`, [11](#)  
`runAnticlusteringPCA`, [12](#)  
`runFImpute`, [12](#)  
`runFImpute`, `FImputeRunner`-method  
    (`runFImpute`), [12](#)  
  
`saveFImpute`, [13](#)  
`saveFImpute`, `FImputeExport`-method  
    (`saveFImpute`), [13](#)  
`saveFImpute`, `SNPDataLong`-method  
    (`saveFImpute`), [13](#)  
`saveFImputeRaw`, [14](#)  
`savePlink`, [15](#)  
`summary`, `SNPDataLong`-method, [16](#)