Package 'msco'

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Type Package
Title Multi-Species Co-Occurrence Analyses
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Description Co-occurrence patterns are typically explored between species pairs. However, doing so inevitably ignore the possible higher-order interference and interactions among three or more species in a community. To address this, the multi-species co-occurrence (msco) index, which calculates the joint occupancy, was recently proposed (<doi: added="" be="" to="">). By reckoning with possible higher-order species interactions, the msco index can encapsulate nine possible archetypes of species co-occurrence patterns and avert the statistical type II error by examining patterns that would otherwise not be detected by any pairwise co-occurrence metrics. To dissect the effects of neutral encounter versus trait-based processes on multi-species co-occurrence and related higher-order interactions, generalised B-spline model has also been proposed (<doi: added="" be="" to="">). The R package "msco" implements the multi-species co-occurrence index and includes a list of functions for calculating this index for different orders, as well as testing and visualizing species co-occurrence archetypes. This package also implements the generalized B-spline model and related analyses.</doi:></doi:>
Depends R (>= $3.5.0$)
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Imports scales, car, ggplot2, dplyr, magick, minpack.lm, cowplot, gtools, tibble, tidyr, caret, pagenum, ape, import, taxize, glm2, MASS, splines2, phylogram, phytools
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Arch_schem

A schematic figure of the archetypes

Description

A schematic diagram illustrating nine possible archetypes (from the null model test) of the patterns of species co-occurrences in ecological communities. The archetypes are denoted $\{Ai: i \in (1:9)\}$. See the details below.

Usage

Arch_schem()

Value

The Arch_schem function returns a **schematic diagram** of the archetypes of species co-occurrence patterns (denoted by $\{Ai: i\in (1:9)\}$), with the following components:

Archetype	Description/Interpretation
A1	The joint occupancy value of the observed community matrix (observed; dark solid line) is above the null model. This means the null hypothesis (i.e. a statement that imply any change in the observed patterns do not reflect any community assembly process as underlying cause) should be rejected, confirming the presence of a mechanism of interest being tested (Lagat <i>et al.</i> , 2021a). It is typical of a community whose species are positively associated (or aggregated) more often than would be expected by chance. Such patterns of community structure may arise from a number of ecological mechanisms including environmental filtering or shared habitat requirements (Cordero and Jackson, 2019).
A2	The observed is greater than null expectation for $i = 2$ but within null expectation for $i \geq 3$. This implies a pairwise metric detects a non-random pattern of the community structure, but when higher order species are considered, a random pattern is produced. This is typical of a community whose species are aggregated more often than by chance in sites with few species than in sites with many species (Lagat <i>et al.</i> , 2021a).
A3	The observed is greater than null expectation for lower orders, within null expectation for medium orders, and less than null expectation for higher orders. This means species co-occur more often than by chance in sites with few species, but are segregated more often than by chance in sites with many species, depicting

et al., 2021a).

a community structured by two different community assembly processes (Lagat

Arch_schem 3

The observed is within null expectation for i = 2 but greater than null expectation for $i \geq 3$. This means when use pairwise co-occurrence is used, the null hypothesis is not rejected, but when joint occupancy is used, the same null hypothesis is rejected. I.e., pairwise co-occurrence fails at detecting patterns of aggregation for sites with many species, i.e. a type II error or false negative (Lagat *et al.*, 2021a).

The observed is within the null expectation for all orders $i \geq 2$, implying the test is not statistically significant. This has been ecologically inferred to mean ecological communities are random and that no community assembly processes or mechanisms influence their structure (Lagat *et al.*, 2021a; Cordero and Jackson, 2019; Gotelli and Sounding, 2001).

The observed is within the null expectation for i=2 but less than the null expectation for $i\geq 3$. This means when use pairwise co-occurrence is used, the null hypothesis is not rejected, but when joint occupancy is used, the same null hypothesis is rejected. I.e., pairwise co-occurrence fails at detecting patterns of segregation for sites with many species, i.e. a type II error or false negative (Lagat $et\ al.$, 2021a).

The observed is less than null expectation for lower orders, within null expectation for medium orders, and greater than null expectation for higher orders. Implying species are segregated more often than would be expected by chance in sites with few species, but co-occur more often than by chance in sites with many species, depicting a community structured by two different community assembly processes (Lagat *et al.*, 2021a).

The observed is less than null expectation for i = 2 but within null expectation for $i \geq 3$. This means a pairwise metric detects a non-random pattern of the community structure, but when higher order species are considered, a random pattern is produced. This is typical of a community whose species are segregated more often than by chance in sites with few species than in sites with many species (Lagat *et al.*, 2021a).

The joint occupancy value of the observed community matrix (dark solid line) is below the null model. This means the null hypothesis should be rejected, confirming the presence of a mechanism of interest being tested (Lagat *et al.*, 2021a). It is typical of a community structured by inter-specific competition or limiting similarity, though predation might also generate similar patterns (Hein et al. 2014).

Note

Α5

Α6

Α7

Α8

Α9

Arch_schem is not a generic function which can take in any dataset and give the outputs, but a path to a schematic diagram saved in this package. A representational figure from empirical, simulated or any known .csv binary data matrices can be accessed with Jo.plots function.

References

- 1. Cordero, R.D. and Jackson, D.A. (2019). Species-pair associations, null models, and tests of mechanisms structuring ecological communities. *Ecosphere* **10.** https://doi.org/10.1002/ecs2.2797
- 2. Gotelli, N. J. and Sounding, E. (2001). Research frontiers in null model analysis. *Glob. Ecol. Biogeogr.* **10**, 337-343. https://doi.org/10.1046/j.1466-822X.2001.00249.x
- 3. Hein et al. (2014). Fish introductions reveal the temperature dependence of species interactions. *Proc. R. Soc. B Biol. Sci.* **281**. https://doi.org/10.1098/rspb.2013.2641

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4. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

cross_valid

Cross validation of the generalised B-spline model

Description

This function implements four different cross-validation techniques to evaluate the predictive ability of the generalised B-spline model (*sensu* Lagat *et al.*, 2021b). The four different techniques implemented are:

- · validation set approach;
- k -fold;
- · Leave-one-out-cross-validation (LOOCV), and
- · Repeated k-fold.

Usage

```
cross_valid(gbsm_obj, type = "k-fold", p, k, k_fold.repeats)
```

Arguments

gbsm_obj	An object of class "gbsm" (i.e., assigned to gbsm function).
type	The type of the cross-validation approach used. It must be \in {"validation.set", "k-fold", "LOOCV", "repeated.k-fold" }
p	The percentage (in decimal form) of data used in training the model. The value is used if the cross-validation approach implemented is "validation.set".
k	The value of k used in both "k-fold" and "repeated.k-fold" types of cross-validation. This value represents the number of subsets or groups that a given sample of data is to be split into. A value of 5 or 10 is used in practice, as it leads to an ideal bias-variance trade-off (Lagat <i>et al.</i> , 2021b).

k_fold.repeats The number of replicates used in "repeated.k-fold" type of cross-validation.

Details

The k-fold cross-validation approach is highly recommended due to its computational efficiency and an acceptable bias-variance trade-off, subject to the value of k chosen to be either 5 or 10 (Lagat *et al.*, 2021b). For more details on the other cross-validation approaches, see Lagat *et al.* (2021c).

Value

Depending on the type of cross-validation approach implemented, the cross_valid function returns:

- a data.frame with the following test errors (for "validation.set"):
 - RMSE: A root mean squared error;
 - R_squared: the Pearson's r^2 , and
 - MAE: the mean absolute error.
- an array with test errors as above including the type of the regression model used, size of the samples, number of predictors, type of cross-validation performed, and summary of sample sizes (for "k-fold", "LOOCV", and "repeated.k-fold").

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References

1. Fushiki, T. (2011). Estimation of prediction error by using K-fold cross-validation. *Stat. Comput.* **21**, 137-146. https://doi.org/10.1007/s11222-009-9153-8

- 2. Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.
- 3. Lagat, V. K., Latombe, G. and Hui, C. (2021c). msco: an R software package for null model testing of multi-species interactions and interference with covariates. DOI: <To be added>.
- 4. Pearson, K. (1895) VII. Note on regression and inheritance in the case of two parents. *proceedings of the royal society of London*, **58**:240-242. https://doi.org/10.1098/rspl. 1895.0041

Examples

```
## Not run:
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
setwd(my.path)
s.data <- get(load("s.data.csv")) ## Species-by-site matrix</pre>
t.data <- get(load("t.data.csv")) ## Species-by-trait matrix</pre>
\verb|p.d.mat| <- get(load("p.d.mat.csv"))| \textit{ ## Species-by-species phylogenetic distance matrix}
gbsm_obj <- msco::gbsm(s.data, t.data, p.d.mat, metric= "Simpson_eqn", d.f=4,</pre>
 order.jo=3, degree=3, n=1000, b.plots=FALSE, scat.plot=FALSE,
  response.curves=FALSE, leg=1, max.vif, max.vif2, start.range=c(-0.1,0))
val.set <- msco::cross_valid(gbsm_obj, type="validation.set", p=0.8)</pre>
val.set
kfold <- msco::cross_valid(gbsm_obj, type="k-fold", k=5)</pre>
kfold
loocv <- msco::cross_valid(gbsm_obj, type="LOOCV")</pre>
loocv
repeated.kfold <- msco::cross_valid(gbsm_obj, type="repeated.k-fold", k=5, k_fold.repeats=100)
repeated.kfold
## End(Not run)
```

gbsm

A generalised B-spline modelling for a set of neutral and trait-based variables

Description

This function implements the generalised B-spline model (*sensu* Lagat *et al.*, 2021b) for dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference. Generalized linear model (*sensu* Hastie and Tibshirani, 1986) with binomial variance distribution and log link functions employed, with predictors transformed using a linear combination of B-splines (*sensu* Curry and Schoenberg, 1988).

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Usage

```
gbsm(
  s.data,
  t.data,
 p.d.mat,
 metric = "Simpson_eqn",
  d.f = 4,
 order.jo = 3,
  degree = 3,
 n = 1000,
  b.plots = TRUE,
  gbsm.model,
  scat.plot = TRUE,
  response.curves = TRUE,
 vlabel = TRUE,
  leg = 1,
 max.vif = 20,
 max.vif2 = 10,
  start.range = c(-0.1, 0)
)
```

Arguments

metric

n

gbsm.model

s.data A species-by-site presence/absence data.frame with entries indicating occurrence (1) and non-occurrence (0) of species in a site.

t.data A data.frame with traits as columns and species as rows. The species must be the same as in s.data.

p.d.mat A symmetric matrix with dimension names as species and entries indicating the phylogenetic distance between any two of them (species).

The type of rescaling applied to the joint occupancy metric. Available options are: Simpson_eqn for Simpson equivalent, Sorensen_eqn for Sorensen equivalent, raw_prop for the raw form of the metric rescaled by dividing by the total number of sites, N, and raw for the raw form of the metric without rescaling.

d. f Degrees of freedom for B-splines.

order.jo Specific number of species for which the joint occupancy is computed. To implement generalised B-spline modelling for multiple orders, see gbsm_m.orders function.

degree Degree of the B-splines.

Number of samples for which the joint occupancy is computed. These samples are non-overlapping. I.e., sampling is done without replacement. If the total number of combinations of i species chosen from the total species pool m, i.e. choose(m,i), is less than this value (n), choose(m,i) is used as the (maximum) number of samples one can set. Otherwise sampling without replacement is performed to select just the n samples.

b.plots Boolean value indicating if B-spline basis functions (of the first predictor) should be plotted.

The model used if the raw form of the metric is choosen. Availbale options are "quasipoisson" for quasipoisson GLM or "nb" for negative binomial GLM. Other metric types strictly uses binomial GLM.

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scat.plot Boolean value indicating if scatter plots between joint occupancy and its pre-

dicted values should be plotted.

response.curves

A boolean value indicating if all response curves should be plotted.

ylabel Boolean value indicating if the y label should be included in the response curves.

This parameter is added to help control the appearance of plots in gbsm_m.orders

function.

leg Boolean value indicating if the legend of the gbsm outputs should be included

in the plots. This parameter is added to help control the appearance of plots in

gbsm_m.orders function.

max.vif This parameter is used to detect and avoid multi-collinearity among covariates.

Its value can be varied to have an intermediate GBSM model (based on GLM) with certain VIF values. Any predictor variable (from the original model) with VIF greater than this value is removed. This can be repeated until an ideal VIF

of less or equal to a desired value is achieved.

max.vif2 Like max.vif, this parameter is used to detect and avoid multi-collinearity among

covariates. Its value can be varied to have a final GBSM model (based on GLM) with certain VIF values much less than max.vif. Any predictor variable (from the intermediate model) with VIF greater than this value is removed. This can be repeated until an ideal VIF of less or equal to a desired value is achieved.

start.range Range of starting values for glm regression.

Value

gbsm function returns a list containing the following outputs:

order.jo Order of joint occupancy

Predictors Predictor variables used in GLM regression with binomial variance distribution

function and log link function.

Responses Response variables from GLM regression with binomial variance distribution

function and log link function.

coeff Coefficients of the generalized linear model used.

glm_obj Generalized linear model used.

j.occs Rescaled observed joint occupancies. See metricabove.

 $bs_pred \qquad \qquad B-spline-transformed\ Predictors.$

var.expld Amount of variation in joint occupancy explained by the Predictors. I.e., it is

the Pearson's r^2 between the observed and predicted values of joint occupancy.

Original.VIFs Variance inflation factors from the original GBSM model (before removing co-

variates exceeding max.vif).

Intermediate.VIFs

Variance inflation factors from the intermediate GBSM model (after removing

the 1st set of covariates exceeding max.vif).

Final.VIFs Variance inflation factors from the final GBSM model (after removing the 2nd

set of covariates exceeding $\max.vif2$).

summary summary of the regression results

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References

1. Curry, H. B., and Schoenberg, I. J. (1988). On Pólya frequency functions IV: the fundamental spline functions and their limits. In *IJ Schoenberg Selected Papers* (pp. 347-383). Birkhäuser, Boston, MA. https://doi.org/10.1007/978-1-4899-0433-1_17

- 2. Hastie, T., and Tibshirani, R. (1986). Generalized additive models. *Stat. Sci.* 1(3), 297-310. https://doi.org/10.1214/ss/1177013604
- 3. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.
- 4. Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.

Examples

```
## Not run:
 my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
 setwd(my.path)
 s.data <- get(load("s.data.csv")) ## Species-by-site matrix</pre>
 t.data <- get(load("t.data.csv")) ## Species-by-trait matrix</pre>
 \verb|p.d.mat <- get(load("p.d.mat.csv"))| ## Species-by-species phylogenetic distance matrix|\\
RNGkind(sample.kind = "Rejection")
 set.seed(0)
 my.gbsm <- msco::gbsm(s.data, t.data, p.d.mat, metric = "Simpson_eqn", gbsm.model,</pre>
 d.f=4, order.jo=3, degree=3, n=1000, b.plots=TRUE, scat.plot=TRUE,
   response.curves=TRUE, leg=1, max.vif = 10, max.vif2 = 3,
    start.range=c(-0.1,0))
 head(my.gbsm$bs_pred)
 head(my.gbsm$Predictors)
head(my.gbsm$Responses)
 my.gbsm$order.jo
 my.gbsm$var.expld
my.gbsm$Original.VIFs
my.gbsm$Intermediate.VIFs ## Resulting covariate VIFs after removing covariates with VIF > max.vif
my.gbsm$Final.VIFs ## Resulting covariate VIFs after removing covariates with VIF > max.vif2
## End(Not run)
```

gbsm.res

Results on generalised B-spline modelling (presented in Lagat et al., 2021b)

Description

This function allows the replication of the results on generalised B-spline modelling, presented in Lagat *et al.* (2021b). Executing gbsm.res() therefore gives these outputs that are saved as .RDS files in msco. If the codes that produced these (saved) outcomes are desired, the codes below are made available.

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Usage

```
gbsm.res()
```

Value

Returns all the results presented in Lagat et al. (2021b). To replicate

• Figs. 1, 3, 4, 5, and Tables 1 and S1, execute the following code:

```
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
setwd(my.path)
s.data <- get(load("s.data.csv")) ##Species-by-site matrix</pre>
t.data <- get(load("t.data.csv")) ##Species-by-trait matrix</pre>
p.d.mat <- get(load("p.d.mat.csv")) ##Species-by-species phylogenetic distance matrix</pre>
RNGkind(sample.kind = "Rejection")
set.seed(1)
gb.res <- msco::gbsm_m.orders(s.data,</pre>
             t.data,
             p.d.mat,
             metric = "Simpson_eqn",
             gbsm.model,
             orders = c(2:5, 8, 10, 15),
             d.f = 4,
             degree = 3,
             n = 1000,
             k = 5,
             p = 0.8,
             type = "k-fold",
             scat.plots = TRUE,
             response.curves = TRUE,
             j.occs.distrbn = TRUE,
             mp.plots = TRUE,
             max.vif = 10,
            \max.vif2 = 3,
             start.range=c(-0.1,0)
          )
gb.res$contbn_table$`order 3` ## Table 1
gb.res$model.validation.table ## Table S1
gb.res$Original.VIFs$`order 3`
gb.res$Intermediate.VIFs$`order 3` ## Resulting covariate VIFs after removing
                                        ## covariates with VIF > max.vif
gb.res$Final.VIFs$`order 3` ## Resulting covariate VIFs after removing
                                         ## covariates with VIF > max.vif2
```

• Figs. S1, 2, and S2, execute the following codes:

```
- Fig. S1:
```

```
remotes::install_github("jinyizju/V.PhyloMaker", force = TRUE)
library(V.PhyloMaker)
my.path <- system.file("extdata/gsmdat", package = "msco")
setwd(my.path)</pre>
```

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```
s.data <- get(load("s.data.csv")) ##Species-by-site matrix</pre>
taxa <- get(load("taxa.levels.csv")) ##Species taxa</pre>
my.phylo.plot <- msco::s.phylo(s.data,</pre>
                            p.d.mat = NULL,
                            database = "ncbi",
                            obs.taxa = FALSE,
                            taxa.levels = taxa,
                            Obs.data = FALSE,
                            phy.d.mat = FALSE,
                            phylo.plot = TRUE)
- Fig. 2:
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
setwd(my.path)
s.data <- get(load("s.data.csv")) ##Species-by-site matrix</pre>
t.data <- get(load("t.data.csv")) ##Species-by-trait matrix</pre>
p.d.mat <- get(load("p.d.mat.csv")) ##Species-by-species phylogenetic distance matrix</pre>
RNGkind(sample.kind = "Rejection")
set.seed(1)
my.gbsm <- msco::gbsm(s.data,</pre>
                    t.data,
                    p.d.mat,
                    metric = "Simpson_eqn",
                    gbsm.model,
                    d.f = 4,
                    order.jo = 3,
                    degree = 3,
                    n = 1000,
                    b.plots = TRUE,
                    scat.plot = FALSE,
                    response.curves = FALSE,
                    leg = 1,
                   max.vif = 10,
                   \max.vif2 = 3,
                    start.range=c(-0.1,0)
                  )
- Fig. 6:
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
setwd(my.path)
s.data <- get(load("s.data.csv")) ##Species-by-site matrix</pre>
t.data <- get(load("t.data.csv")) ##Species-by-trait matrix</pre>
p.d.mat <- get(load("p.d.mat.csv")) ##Species-by-species phylogenetic distance matrix</pre>
RNGkind(sample.kind = "Rejection")
set.seed(1)
pe <- msco::pred.error.bands(s.data,</pre>
                      t.data,
                      p.d.mat,
                      metric = "Simpson_eqn",
                      gbsm.model,
```

```
d.f = 4,
  simm = 10,
  orders = c(2:5, 8, 10, 15),
  degree = 3,
  n = 1000,
  start.range = c(-0.2, 0)
```

Caveat: The above codes can collectively take approximately 7 minutes to execute (with prediction uncertainty plot taking 6 minutes alone). It took 7.3895 minutes to run (and output results) on a 64 bit system with 8 GB RAM and 3.60 GHz CPU.

Note

The function gbsm.res is not for general use. We included it in this package to help the readers of Lagat *et al.* (2021b) paper, who may want to get a deeper understanding of how the results presented in this paper were arrived at. It also allows deeper scrutiny of Lagat *et al.* (2021b)'s findings.

References

Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.

Examples

```
## Not run:
gbs.res <- msco::gbsm.res()
gbs.res$contbn_table$`order 3`
gbs.res$model.validation.table
gbs.res$Original.VIFs$`order 3`
gbs.res$Intermediate.VIFs$`order 3`
gbs.res$Final.VIFs$`order 3`</pre>
## End(Not run)
```

 ${\tt gbsm_m.orders}$

Predictor's contribution and model performance assessment from the results on multiple orders of joint occupancy

Description

This function implements the generalised B-spline model (gbsm; *sensu* Lagat et al., 2021b) for dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference. Unlike gbsm that performs gbsm for a single order of species, gbsm_m.orders takes into account multiple orders of joint occupancy. In particular: for multiple joint occupancy orders, this function computes:

- each predictor's contribution to the explained variation in joint occupancy,
- the goodness-of-fit and model performance from cross-validation, and

- plots the:
 - response curves,
 - scatter plots (between the observed and predicted joint occupancy values),
 - histograms of the joint occupancy frequency distribution, and
 - model performance plots.

Usage

```
gbsm_m.orders(
  s.data,
  t.data,
  p.d.mat,
  metric = "Simpson_eqn",
  orders,
  d.f = 4,
  degree = 3,
  n = 1000,
  k = 5,
  p = 0.8,
  type = "k-fold",
  gbsm.model,
  scat.plots = FALSE,
  response.curves = TRUE,
  j.occs.distrbn = FALSE,
  mp.plots = FALSE,
  max.vif = 20,
  max.vif2 = 10,
  start.range = c(-0.1, 0)
)
```

Arguments

s.data	A species-by-site presence/absence data.frame with entries indicating occurrence (1) and non-occurrence (0) of species in a site.
t.data	A data.frame with traits as columns and species as rows. The species must be the same as in ${\tt s.data}$.
p.d.mat	A symmetric matrix with dimnames as species and entries indicating the phylogenetic distance between any two of them (species).
metric	As for gbsm.
orders	Specific number of species for which the joint occupancy is computed.
d.f	As for gbsm.
degree	As for gbsm.
n	As for gbsm.
k	As for cross_valid.
р	As for cross_valid.
type	As for cross_valid.
gbsm.model	As for gbsm.
scat.plots	Boolean value indicating if scatter plots between joint occupancy and its predicted values should be plotted.

response.curves

A boolean value indicating if all response curves for all joint occupancy orders (jo.orders) should be plotted.

j.occs.distrbn A boolean value indicating if the histograms of the frequency distribution of

observed joint occupancy should be output.

mp.plots A boolean value indicating if the model performance plots should be output.

max.vif As for gbsm.
max.vif2 As for gbsm.
start.range As for gbsm.

Value

gbsm_m.orders function returns a list containing the following outputs:

- jo.orders: A set of joint occupancy orders.
- contrbn_table: A list of data.frames consisting of:
 - predictor: A column of predictors.
 - var.expld_M1: A column of goodness-of-fit (I.e., the Pearson's r^2 between the observed and predicted values of joint occupancy when all predictors are used in the model.
 - var.expld_M2: The Pearson's r^2 between the observed and the predicted values of joint occupancy when all predictors except the predictor whose contribution is to be determined, are used in the model.
 - contribution: Each predictor's proportion of contribution in explaining joint occupancy. This value is given by: contribution = $\frac{var.expld_{M1}-var.expld_{M2}}{var.expld_{M1}}$
- model.validation.table: A data.frame with:
 - orders: Orders of joint occupancy used.
 - Rsquared_gf: Goodness-of-fit of the model. I.e., it is the Pearson's r^2 between the observed and predicted values of joint occupancy, for different orders.
 - Rsquared_cv: Model performance from cross-validation.
- metric: As for gbsm.
- d.f: As for gbsm.
- n: As for gbsm.
- degree: As for gbsm.
- jo.orders: Orders of joint occupancy used.
- Original.VIFs As for gbsm (different orders).
- Intermediate.VIFs As for gbsm (different orders).
- Final.VIFs As for gbsm (different orders).

References

- 1. Curry, H. B., and Schoenberg, I. J. (1988). On Pólya frequency functions IV: the fundamental spline functions and their limits. In *IJ Schoenberg Selected Papers* (pp. 347-383). Birkhäuser, Boston, MA. https://doi.org/10.1007/978-1-4899-0433-1_17
- 2. Hastie, T., and Tibshirani, R. (1986). Generalized additive models. *Stat. Sci.* 1(3), 297-310. https://doi.org/10.1214/ss/1177013604

3. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

4. Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.

Examples

```
## Not run:
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
 setwd(my.path)
 s.data <- get(load("s.data.csv")) ## Species-by-site matrix</pre>
 t.data <- get(load("t.data.csv")) ## Species-by-Trait matrix</pre>
 p.d.mat <- get(load("p.d.mat.csv")) ## Species-by-species phylogenetic distance matrix</pre>
 RNGkind(sample.kind = "Rejection")
 set.seed(1)
 jp <- msco::gbsm_m.orders(s.data, t.data, p.d.mat, gbsm.model,</pre>
 metric="Simpson\_eqn", orders = c(3:5, 8, 10, 15, 20), d.f=4,
  degree=3, n=1000, k=5, p=0.8, type="k-fold", scat.plots=TRUE,
   response.curves=TRUE, j.occs.distrbn=TRUE, mp.plots=TRUE,
     max.vif = 10, max.vif2 = 4, start.range=c(-0.2,0))
 jp$contbn_table[[1]]
 jp$model.validation.table
 jp$jo.orders
 jp$Original.VIFs$`order 3`
 jp$Intermediate.VIFs$`order 3`
 jp$Final.VIFs$`order 3`
 ## Close the open plots.gbsm.pdf file before running the 2nd example
 RNGkind(sample.kind = "Rejection")
 set.seed(1)
 jp2 <- msco::gbsm_m.orders(s.data, t.data, p.d.mat, gbsm.model,</pre>
 metric="Sorensen_eqn", orders = c(3:5, 8, 10, 15, 20), d.f=4,
  degree=3, n=1000, k=5, p=0.8, type="k-fold", scat.plots=TRUE,
   response.curves=TRUE, j.occs.distrbn=TRUE, mp.plots=TRUE,
     max.vif = 10, max.vif2 = 4, start.range=c(-0.2,0))
 jp2$contbn_table[[1]]
 jp2$model.validation.table
 jp2$jo.orders
 jp2$Original.VIFs$`order 3`
 jp2$Intermediate.VIFs$`order 3`
 jp2$Final.VIFs$`order 3`
## Close the open plots.gbsm.pdf file before running the 3rd example
RNGkind(sample.kind = "Rejection")
 set.seed(1)
 jp3 <- msco::gbsm_m.orders(s.data, t.data, p.d.mat, gbsm.model,</pre>
 metric="raw_prop", orders = c(3:5, 8, 10, 15, 20), d.f=4,
  degree=3, n=1000, k=5, p=0.8, type="k-fold", scat.plots=TRUE,
   response.curves=TRUE, j.occs.distrbn=TRUE, mp.plots=TRUE,
   max.vif = 10, max.vif2 = 4, start.range=c(-0.2,0))
 jp3$contbn_table[[1]]
```

*j.*occ 15

```
jp3$model.validation.table
 jp3$jo.orders
 jp3$Original.VIFs$`order 3`
 jp3$Intermediate.VIFs$`order 3`
 jp3$Final.VIFs$`order 3`
## Close the open plots.gbsm.pdf file before running the 4th example
RNGkind(sample.kind = "Rejection")
 set.seed(1)
 jp4 <- msco::gbsm_m.orders(s.data, t.data, p.d.mat, gbsm.model="nb",</pre>
 metric="raw", orders = c(3:5, 8, 10, 15, 20), d.f=4,
   degree=3, n=1000, k=5, p=0.8, type="k-fold", scat.plots=TRUE,
   response.curves=TRUE, j.occs.distrbn=TRUE, mp.plots=TRUE,
   max.vif = 10, max.vif2 = 4, start.range=c(-0.2,0))
 jp4$contbn_table[[1]]
 jp4$model.validation.table
 jp4$jo.orders
 jp4$Original.VIFs$`order 3`
 ip4$Intermediate.VIFs$`order 3`
 jp4$Final.VIFs$`order 3`
## Close the open plots.gbsm.pdf file before running the 5th example
RNGkind(sample.kind = "Rejection")
 set.seed(1)
 jp5 <- msco::gbsm_m.orders(s.data, t.data, p.d.mat, gbsm.model="quasipoisson",</pre>
 metric="raw", orders = c(3:5, 8, 10, 15, 20), d.f=4,
   degree=3, n=1000, k=5, p=0.8, type="k-fold", scat.plots=TRUE,
   response.curves=TRUE, j.occs.distrbn=TRUE, mp.plots=TRUE,
   max.vif = 10, max.vif2 = 4, start.range=c(-0.2,0))
 jp5$contbn_table[[1]]
 jp5$model.validation.table
 jp5$jo.orders
 jp5$Original.VIFs$`order 3`
 jp5$Intermediate.VIFs$`order 3`
 jp5$Final.VIFs$`order 3`
## End(Not run)
```

j.occ

Expected value of joint occupancy for order i and its standard deviation

Description

This function computes joint occupancy (the average number of sites harbouring a given number of i species simultaneously), and its standard deviation.

Usage

```
j.occ(s.data, order, metric = "raw")
```

j.occs

Arguments

s.data	A species-by-site presence/absence matrix with entries indicating occurrence (1) and non-occurrence (0) of species in a site.
order	Specific number of species for which joint occupancy and its standard deviation is computed.
metric	The type of rescaling applied to the joint occupancy metric. Available options are: Simpson_eqn for Simpson equivalent, Sorensen_eqn for Sorensen equivalent, and raw for the raw form of index without rescaling.

Value

Returns a list with the following outputs:

jo.val	Joint occupancy value.
jo.sd	The standard deviation of jo.val.

References

Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

Examples

```
ex.data <- read.csv(system.file("extdata", "274.csv", package = "msco"))
jo <- msco::j.occ(ex.data, order = 3, metric = "raw")
jo

ex.data2 <- read.csv(system.file("extdata", "65.csv", package = "msco"))
jo2 <- msco::j.occ(ex.data2, order = 3, metric = "raw")
jo2</pre>
```

j.occs Expected value of joint occupancy and its standard deviation for a range of orders

Description

This function computes joint occupancy (the average number of sites harbouring a given number of i species simultaneously) and its standard deviation for a range of orders (number of species).

Usage

```
j.occs(s.data, orders = 1:nrow(s.data), metric = "raw")
```

Arguments

s.data	A species-by-site presence/absence matrix with entries indicating occurrence (1) and non-occurrence (0) of species in a site.
orders	Range number of species for which joint occupancy and its standard deviation is computed.
metric	The type of rescaling applied to the joint occupancy metric. Available options are: Simpson_eqn for Simpson equivalent, Sorensen_eqn for Sorensen equivalent, and raw for the raw form of index without rescaling.

Value

Returns a list with the following outputs:

```
jo.vals A vector of joint occupancy values for a range number of species (in orders).

A vector of standard deviations of jo.vals.
```

References

Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

Examples

```
ex.data <- read.csv(system.file("extdata", "274.csv", package = "msco"))
jos <- msco::j.occs(ex.data, orders = 1:nrow(ex.data), metric = "raw")
jos

ex.data2 <- read.csv(system.file("extdata", "65.csv", package = "msco"))
jos2 <- msco::j.occs(ex.data2, orders = 1:nrow(ex.data), metric = "raw")
jos2</pre>
```

Jo.eng

Joint occupancy model engine

Description

This function is the engine behind the null model testing of species co-occurrence patterns, and analyses of the joint occupancy decline and the parametric forms of this decline, for one particular community. In particular, Jo.eng:

- computes the joint occupancy (i.e. the number of sites or assemblages harbouring multiple species simultaneously);
- performs a null model test using the same index;
- fits the three regression models (exponential, power law and exponential-power law) to joint occupancy decline (*sensu* Lagat *et al.*, 2021a) with order (number of species);
- estimates the parameter values of these models;
- determines the best model among the three using AIC values;
- quantifies the performance of the fitted models using the Pearson's r^2 ;
- plots the joint occupancy decline regression and null models, and
- ascertains the archetypes of the patterns of species co-occurrences (from null model test) from which inferences on the type of drivers structuralising ecological communities can be made.

Usage

```
Jo.eng(
    s.data,
    algo = "sim2",
    metric = "raw",
    nReps = 999,
    dig = 3,
```

```
s.dplot = FALSE,
All.plots = TRUE,
Jo.coeff = TRUE,
my.AIC = TRUE,
my.rsq = TRUE,
Exp_Reg = TRUE,
P.law_Reg = TRUE,
Exp_p.l_Reg = TRUE,
Obs.data = FALSE,
Sim.data = FALSE,
Jo_val.sim = FALSE,
C.I_Jo_val.sim = FALSE,
Jo_val.obs = TRUE,
Metric = TRUE,
Algorithm = TRUE,
S.order = TRUE,
nmod_stats = TRUE,
Pt_Arch_Vals = TRUE,
Atype = TRUE,
p.n.plot = FALSE,
trans = FALSE,
lab = FALSE,
leg = FALSE,
m.n.plot = FALSE
```

Arguments

s.data	A species-by-site presence/absence matrix with entries indicating occurrence (1) and non-occurrence (0) of species in a site.
algo	Randomisation algorithm used for the comparison with the null model. The possible options to choose from are: sim1, sim2, sim3, sim4, sim5, sim6, sim7, sim8, and sim9, all from Gotelli (2000). sim2 is highly recommended (see Lagat <i>et al.</i> , 2021a).
metric	The type of rescaling applied to the joint occupancy metric. Available options are: Simpson_eqn for Simpson equivalent, Sorensen_eqn for Sorensen equivalent, and raw for the raw form of index without rescaling.
nReps	Number of simulations used in the null model test.
dig	The number of decimal places of the joint occupancy values (y axis) in the plots. The default is 3.
s.dplot	A Boolean indicating whether the standard deviation of multi-species co-occurrence index should be included in the plots of joint occupancy decline or not.
All.plots	A Boolean indicating whether joint occupancy decline regression and null model plots should be output.
Jo.coeff	A Boolean indicating if coefficient estimates of the joint occupancy decline regression models should be printed.
my.AIC	A Boolean indicating whether Akaike Information Criterion of the joint occupancy decline regression models should be output or not.
my.rsq	A Boolean indicating whether square of correlation coefficient between the ob-

served and predicted values of joint occupancy should be output.

Exp_Reg	A Boolean indicating if exponential regression parametric model should be printed.
P.law_Reg	A Boolean indicating if power law regression parametric model should be printed.
Exp_p.l_Reg	A Boolean indicating if exponential-power law regression parametric model should be printed.
Obs.data	A Boolean indicating if observed/empirical data should be output.
Sim.data	A Boolean indicating if simulated/random data produced using any of the simulation algorithms should be output.
Jo_val.sim	A Boolean indicating if joint occupancy values of the simulated species-by-site presence/absence matrices should be output.
C.I_Jo_val.sim	A Boolean indicating if 95% confidence interval of the joint occupancy values of the simulated data should be printed. This interval is the area under the null model.
Jo_val.obs	A Boolean indicating if joint occupancy values of the observed species-by-site presence/absence matrices should be output.
Metric	A Boolean indicating if metric used should be printed.
Algorithm	A Boolean indicating if simulation algorithm used should be printed.
S.order	A Boolean indicating if the number of species whose joint occupancy is computed should be printed.
nmod_stats	A Boolean indicating whether the summary statistics for the null model test should be output.
Pt_Arch_Vals	A Boolean indicating if character strings indicating the location of joint occupancy value of the observed data relative to the critical values of the 95% closed confidence interval for every order (number of species), should be printed.
Atype	A Boolean indicating if a character string indicating the overall archetype of joint occupancy decline should be printed. This value must be \in {"A1", "A2", "A3", "A4", "A5", "A6", "A7", "A8", "A9"} or "NA". "NA" could be the combinations of two or more of the nine expected archetypes.
p.n.plot	A Boolean indicating whether null model plot produced using the pairwise natural metric should be output.
trans	A Boolean indicating if the observed and simulated values used in p.n.plot should be transformed by raising them to $(1/100)$. This can be done to compare p.n.plot with All.plots at a point where the order, $i = 2$.
lab	A Boolean indicating if the plot labels should be added to the m.n.plot. This parameter helps to control the appearance of plots in this function.
leg	A Boolean indicating if the legend should be added to the m.n.plot. This parameter helps to control the appearance of plots in this function.
m.n.plot	A Boolean indicating whether null model plot produced using joint occupancy metrics should be output. The default is FALSE.

Value

Jo. eng function returns a list containing the following outputs:

all.plots Joint occupancy decline regression and null model plots.

jo.coeff Coefficient estimates of the joint occupancy decline regression models.

AIC Akaike information criterion of the joint occupancy decline regression models.

r2 Square of correlation coefficient between the observed and predicted values of joint occupancy. Exponential regression parametric model. Exp_reg Power law regression parametric model. P.law_reg Exponential-power law regression parametric model. Exp_p.l_reg Obs.data Observed/empirical data. Simulated/random data produced using any of the simulation algorithms. Sim.data Joint occupancy value of the simulated species-by-site presence/absence matrijo.val.sim C.I_Jo_val.sim 95% confidence interval of the joint occupancy value of the simulated data. joint occupancy value of the observed species-by-site presence/absence matrijo.val.obs Metric Metric used. It must be "j.occ". Algorithm Simulation algorithm used. nReps Number of simulations performed. This value together with the joint occupancy value of the observed data, constitutes the sampling distribution. s.order Number of species whose joint occupancy is computed. Character strings indicating the location of joint occupancy value of the ob-Pt_Arch_vals served data relative to the critical values of the 95% closed confidence interval of the simulated data, for every order (number of species). Archetype A character string indicating the overall archetype from Pt_Arch_vals. It must $be \in \{\text{"A1"}, \text{"A2"}, \text{"A3"}, \text{"A4"}, \text{"A5"}, \text{"A6"}, \text{"A7"}, \text{"A8"}, \text{"A9"}\} \text{ or "NA"}. \text{"NA"}$

References

Arch schem).

1. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

could be the combinations of two or more of the nine expected archetypes (see

2. Gotelli, N. J. (2000). Null model analysis of species co-occurrence patterns. *Ecology*, 81(9), 2606-2621. https://doi.org/10.1890/0012-9658(2000)081[2606:NMAOSC]2.0.C0;2

Examples

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Jo.plots

Joint occupancy parametric and null model plots

Description

Plots the null model and joint occupancy decline with order (number of species) and fits the decline to exponential, power law and exponential-power law parametric models, respectively.

Usage

```
Jo.plots(jo_Obj)
```

Arguments

jo_Obj

A joint occupancy model object returned by the function Jo.eng.

Details

This function provides a visualization of the forms of joint occupancy decline and null model test. It offers information on:

- the outcomes of the null model test (through the appended archetype value on (e) plot) and
- the comparisons between the forms of joint occupancy decline (through the affixed AIC and rsq values on (b), (c) and (d) plots, respectively).

Value

Produces a figure consisting of the following plots:

- (a) Joint occupancy decline.
- (b) Exponential regression of the joint occupancy decline.
- (c) Power law regression of the joint occupancy decline.
- (d) Exponential-power law regression of the joint occupancy decline.
- (e) Null model test.

References

Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.

Examples

```
## Not run:
ex.data <- read.csv(system.file("extdata", "274.csv", package = "msco"))
jo_Obj <- msco::Jo.eng(ex.data, nReps = 999, All.plots = TRUE, s.dplot = FALSE, dig = 3)
jplots <- msco::Jo.plots(jo_Obj)
jplots

ex.data2 <- read.csv(system.file("extdata", "22.csv", package = "msco"))
jo_Obj2 <- msco::Jo.eng(ex.data2, nReps = 999, All.plots = TRUE, s.dplot = FALSE, dig = 3)
jplots2 <- msco::Jo.plots(jo_Obj2)</pre>
```

Jo.res

```
piplots2

ex.data3 <- read.csv(system.file("extdata", "78.csv", package = "msco"))
jo_Obj3 <- msco::Jo.eng(ex.data3, nReps = 999, All.plots = TRUE, s.dplot = FALSE, dig = 3)
jplots3 <- msco::Jo.plots(jo_Obj3)
jplots3

ex.data4 <- read.csv(system.file("extdata", "65.csv", package = "msco"))
jo_Obj4 <- msco::Jo.eng(ex.data4, nReps = 999, All.plots = TRUE, s.dplot = FALSE, dig = 3)
jplots4 <- msco::Jo.plots(jo_Obj4)
jplots4

## End(Not run)</pre>
```

Jo.res

Results on joint occupancy index (presented in Lagat et al., 2021a)

Description

This function allows the replication of the results presented in Lagat *et al.* (2021a). Executing Jo.res() therefore gives these outputs that are saved as .RDS files in msco. If the codes that produced these (saved) outcomes are desired, the codes below are made available.

Usage

```
Jo.res()
```

Value

Returns all the results presented in Lagat et al. (2021a). To replicate these results, execute the following code:

```
RNGkind(sample.kind = "Rejection")
set.seed(39)
my.path <- system.file("extdata/myCSVs", package = "msco")</pre>
setwd(my.path)
my.files <- gtools::mixedsort(list.files(path = my.path, pattern = ".csv"))</pre>
Lag.res <- msco::mJo.eng(my.files,</pre>
                  algo = "sim2",
                  metric = "raw",
                  Archetypes = FALSE,
                  AICs = FALSE,
                  Delta_AIC = FALSE,
                  datf.Delta_AIC = TRUE,
                  param_hist = TRUE,
                  params = FALSE,
                  my.r2 = FALSE,
                  my.r2.s = TRUE,
                  best.mod2 = TRUE,
                  best.mod3 = TRUE,
                  params_c.i = TRUE
```

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```
);Lag.res
graphics::hist(as.numeric(Lag.res$datf.Delta_AIc$Classes), labels = TRUE,
           col = c("blue", "red", "black"), breaks=seq(0, 3, 1), xaxt = "n",
           ylab = "No. of Communities", xlab = " ",
           main = "Three parametric models compared")
h <- graphics::hist(as.numeric(Lag.res$datf.Delta_AIc$Classes), plot = FALSE)</pre>
graphics::axis(1, at = c(0.5,1.5,2.5), labels = c("Exponential-P.law",
           "Exponential", "Power_law"),
           tick = FALSE, padj= -1.5)
graphics::hist(as.numeric(Lag.res$datf.Delta_AIc$Exp.pl.D_AIC), col = "blue",
           ylab = "No. of communities", xlab = "Delta_AIC", labels = FALSE,
           main = "Exponential-Power law", border = "blue",
      breaks = seq(range(as.numeric(Lag.res$datf.Delta_AIc$Exp.pl.D_AIC))[1],
               ceiling(range(as.numeric(Lag.res$datf.Delta_AIc$Exp.pl.D_AIC))[2]),1))
graphics::hist(as.numeric(Lag.res$datf.Delta_AIc$Exp.D_AIC), col = "red",
           ylab = "No. of communities", xlab = "Delta_AIC", labels = FALSE,
           main = "Exponential", border = "red",
         breaks = seq(range(as.numeric(Lag.res$datf.Delta_AIc$Exp.D_AIC))[1],
               ceiling(range(as.numeric(Lag.res$datf.Delta_AIc$Exp.D_AIC))[2]), 1))
graphics::hist(as.numeric(Lag.res$datf.Delta_AIc$Pl.D_AIC), col = "black",
           ylab = "No. of communities", xlab = "Delta_AIC", labels = FALSE,
           main = "Power law", border = "black",
          breaks = seq(range(as.numeric(Lag.res$datf.Delta_AIc$Pl.D_AIC))[1],
               ceiling(range(as.numeric(Lag.res$datf.Delta_AIc$Pl.D_AIC))[2]), 1))
```

Caveat: The above code can take approximately 8 minutes to execute. It took 8.210296 minutes to run (and output results) on a 64 bit system with 32 GB RAM and i7-11800H @ 2.30GHz processor.

• Fig. 3 can be replicated using:

```
grDevices::dev.new()
msco:::nullmod_archs()
```

• Fig. S1 can be replicated using:

```
my.path <- system.file("extdata/myCSVs", package = "msco")
setwd(my.path)
my.files <- gtools::mixedsort(list.files(path = my.path, pattern = ".csv"))
grDevices::dev.new()
msco:::richness.variances(my.files)</pre>
```

Note

The function Jo.res is not for general use. We included it in this package to help the readers of Lagat *et al.* (2021a) paper, who may want to get a deeper understanding of how the results presented in this paper were arrived at. It also allows deeper scrutiny of Lagat *et al.* (2021a)'s findings.

References

Lagat, V. K., Latombe, G. and Hui, C. (2021a). A multi-species co-occurrence index to avoid type II errors in null model testing. DOI: <To be added>.

Examples

```
## Not run:
ms.res <- msco::Jo.res()
ms.res$r2.s
ms.res$best.mod2
ms.res$best.mod3
ms.res$params_c.i
## End(Not run)</pre>
```

mJo.eng

Joint occupancy model engine for multiple communities

Description

This function is the engine behind the null model testing of species co-occurrence patterns, and analyses of the joint occupancy decline and the parametric forms of this decline, for multiple communities. In particular:

- It performs the null model testing of species co-occurrence patterns and generates the archetypes
 depicting how joint occupancy declines with the number of species (the order of msco) based
 on species-by-site presence/absence .csv data matrices. From these archetypes, inferences
 can be made according to the implemented null models;
- Determines the robustness of the exponential, power law and exponential-power law forms of joint occupancy decline by computing the Pearson's r^2 between the joint occupancy values of the observed data and predicted data, for all orders of species;
- Gives a summary of the total number of communities (under each and for all archetypes) whose forms of joint occupancy decline have $r^2 > 0.95$;
- Computes the AIC and Delta AIC of joint occupancy decline regression models for all communities;
- Computes the total number of communities:
 - with exponential as the best form of joint occupancy decline than power law and vice versa;
 - with either of the three regression models (exponential, power law and exponential-power law) having the best form of the joint occupancy decline;
- Estimates the parameters of:

```
1. exponential: j^{\{i\}} = a \times exp(b \times i);
2. power law: j^{\{i\}} = a \times i^b; and
3. exponential-power law: j^{\{i\}} = a \times exp(b \times i) \times i^c
```

forms of joint occupancy decline, respectively, and their 95% confidence interval.

Usage

```
mJo.eng(
  my.files,
  algo = "sim2",
  metric = "raw",
  nReps = 999,
```

```
Archetypes = FALSE,
AICs = FALSE,
Delta_AIC = FALSE,
datf.Delta_AIC = FALSE,
param_hist = FALSE,
params = FALSE,
best.mod2 = FALSE,
best.mod3 = FALSE,
params_c.i = FALSE,
my.r2 = FALSE,
my.r2.s = FALSE
```

Arguments

my.r2.s

Ĕ	guinents	
	my.files	A vector containing names of species-by-site presence/absence .csv data matrices. The data matrices should be saved in the working directory.
	algo	Simulation algorithm used. The possible options to choose from are: $sim1$, $sim2$, $sim3$, $sim4$, $sim5$, $sim6$, $sim7$, $sim8$, and $sim9$, all from Gotelli (2000). $sim2$ is highly recommended (see Lagat $et\ al.$, 2021a).
	metric	The type of rescaling applied to the joint occupancy metric. Available options are: Simpson_eqn for Simpson equivalent, Sorensen_eqn for Sorensen equivalent, and raw for the raw form of index without rescaling.
	nReps	Number of simulations used in the null model test.
	Archetypes	A Boolean indicating if the archetypes of the patterns of species co-occurrences in multiple communities should be included in the output.
	AICs	A Boolean indicating whether the akaike information criterion (AIC) and Delta AIC of joint occupancy decline regression models for all communities should be included in the output.
	Delta_AIC	A Boolean indicating whether Delta AIC (excluding AIC) should be output.
	datf.Delta_AIC	A Boolean indicating whether a data.frame with Classes and Param.mods as columns, where the former has 1, 2 and 3 values categorizing the three parametric models that has $Delta_AIC=0$ for each communities.
	param_hist	A Boolean indicating whether a histogram of the number of communities where the three parametric forms (exponential, power law and exponential-power law) of joint occupancy decline had the lowest AIC values.
	params	A Boolean indicating whether parameter estimates of the joint occupancy decline regression models should be included in the output.
	best.mod2	A Boolean indicating if exponential and power law regression model comparisons should be included in the output.
	best.mod3	A Boolean indicating if exponential, power law and exponential-power law regression model comparisons should be included in the output.
	params_c.i	A Boolean indicating if 95% C.I of the parameter estimates of the joint occupancy decline regression models should be included in the output.
	my.r2	A Boolean indicating if the robustness of joint occupancy decline regression

models should be computed and output.

cline regression models should be computed and output.

A Boolean indicating if the robustness summary values of joint occupancy de-

Details

mJo. eng function is useful when analyzing multiple species-by-site presence/absence data matrices at once. If one community matrix is analyzed, the outputs of the function Jo.eng should suffice.

Value

mJo. eng function returns a list containing the following outputs:

\$Archs

For every community, a list consisting of:

- \$nmod_stats: A data frame with the summary statistics for the null model test; and
- \$Archetype: Archetypes of the patterns of species co-occurrences in ecological communities/matrices (my.files). These archetypes must be ∈ {"A1", "A2", "A3", "A4", "A5", "A6", "A7", "A8", "A9"} or "NA". "NA" could be the combinations of two or more of the nine expected archetypes.

\$all.AICs

A list of data. frames containing the following components:

df	The number of parameters in each of the three (exponential, power law and exponential-power law) joint occupancy decline regression models.
aic	The aic values for each of the three joint occupancy decline regression models.
delta_aic3	The delta_aic values for each of the three joint occupancy decline regression models.
delta_aic2	The delta_aic values for exponential and power law forms of joint occupancy decline regression models.

\$params

A data.frame consisting of:

arch	The archetypes of the patterns of species co-occurrences in each of the species-by-site presence/absence .csv data matrices.
a.ex	The a parameter estimate of the exponential form of joint occupancy decline.
b.ex	The b parameter estimate of the exponential form of joint occupancy decline.
a.pl	The a parameter estimate of the power law form of joint occupancy decline.
b.pl	The b parameter estimate of the power law form of joint occupancy decline.
a.expl	The a parameter estimate of the exponential-power law form of joint occupancy decline.
b.expl	The b parameter estimate of the exponential-power law form of joint occupancy decline.
c.expl	The c parameter estimate of the exponential-power law form of joint occupancy decline.

\$best.mod2

Atable containing the following components:

n	The number of ecological communities represented by species-by-site presence/absence .csv data matrices.
n.lwst_aic	The number of communities with exponential as the best form of joint occupancy decline than power law.

n.delta_aic	The number of communities whose exponential and power law forms of joint occupancy decline have $delta_aic = 0$, respectively. This number must be equal to n.lwst_aic.
%	The percentage of $n.lwst_aic$ (or $n.delta_aic$) relative to the total number of communities (n) analyzed.

\$best.mod3

A table containig the following components:

n	The number of ecological communities represented by species-by-site presence/absence .csv data matrices.
n.lwst_aic	The number of communities with exponential or power law or exponential-power law as the best form of joint occupancy decline among the three (exponential, power law and exponential-power law) regression models.
n.delta_aic	The number of communities whose exponential, power law and exponential-power law forms of joint occupancy decline, respectively, have delta_aic = 0. This number must be equal to n.lwst_aic.
%	The percentage of n.lwst_aic (or n.delta_aic) relative to the total number of communities (n) analyzed.

\$params_c.i

A data.frame consisting of:

arch	The archetypes of the patterns of species co-occurrences in each of the species-by-site presence/absence .csv data matrices.
n	The number of communities under every archetype.
ex_%	The percentages of the number of communities (under every archetype) where exponential form of joint occupancy decline fitted better than power law.
a.ex	The 95% closed confidence interval of the a parameter estimates of the exponential form of joint occupancy decline, under every archetype.
b.ex	The 95% closed confidence interval of the b parameter estimates of the exponential form of joint occupancy decline, under every archetype.
p.l_%	The percentages of the number of communities (under every archetype) where power law form of joint occupancy decline fitted better than exponential.
a.pl	The 95% closed confidence interval of the a parameter estimates of the power law form of joint occupancy decline, under every archetype.
b.pl	The 95% closed confidence interval of the b parameter estimates of the power law form of joint occupancy decline, under every archetype.
ex.pl_%	The percentages of the number of communities (under every archetype) where exponential-power law form of joint occupancy decline fitted better than both the exponential and power law forms.
a.expl	The 95% closed confidence interval of the a parameter estimates of the exponential-power law form of joint occupancy decline, under every archetype.
b.expl	The 95% closed confidence interval of the b parameter estimates of the exponential-power law form of joint occupancy decline, under every archetype.
c.expl	The 95% closed confidence interval of the c parameter estimates of the exponential-power law form of joint occupancy decline, under every archetype.

\$r2

A list of data. frames containig the following components:

rsq. ex r^2 for the exponential form of joint occupancy decline.

rsq.pl r^2 for the power law form of joint occupancy decline.

rsq.ex.pl r^2 for the exponential-power law form of joint occupancy decline.

\$r2.s

• A list containing the following components:

\$rsq.per.Archs

- Archs: Archetypes of the patterns of species co-occurrences in each of the species-by-site presence/absence .csv data matrices.
- n.a: Number of communities under each archetype.
- rsq. ex: Number of communities under each archetype whose exponential forms of joint occupancy decline have $r^2>0.95$.
- rsq.pl: Number of communities under each archetype whose power law forms of joint occupancy decline have $r^2>0.95.$
- rsq.ex-pl: Number of communities under each archetype whose exponential-power law forms of joint occupancy decline have $r^2 > 0.95$.

\$rsq.all.Communities

- n: Number of all communities analyzed
- ex: Number of communities whose exponential forms of joint occupancy decline have $\ensuremath{r^2}\xspace > 0.95$
- pl: Number of communities whose power law forms of joint occupancy decline have $r^2>0.95$
- ex.pl: Number of communities whose exponential-power law forms of joint occupancy decline have $r^2>0.95$

\$m.Jo.plots

Produces a .pdf file with multiple figures each consisting of the following plots:

- (a) as for Jo.plots
- (b) as for Jo.plots
- (c) as for Jo.plots
- (d) as for Jo.plots
- (e) as for Jo.plots

References

- 1. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.
- 2. Gotelli, N. J. (2000). Null model analysis of species co-occurrence patterns. *Ecology*, 81(9), 2606-2621. https://doi.org/10.1890/0012-9658(2000)081[2606:NMAOSC]2.0.C0;2
- 3. Pearson, K. (1895) VII. Note on regression and inheritance in the case of two parents. *proceedings of the royal society of London*, **58**:240-242. https://doi.org/10.1098/rspl. 1895.0041
- 4. Petrossian, G.A., Maxfield, M (2018). An information theory approach to hypothesis testing in criminological research. *crime science*, 7(1), 2. https://doi.org/10.1186/s40163-018-0077-5

msco.res 29

Examples

```
## Not run:
my.path <- system.file("extdata", package = "msco")</pre>
setwd(my.path)
my.files <- gtools::mixedsort(list.files(path = my.path, pattern = "*.csv"))</pre>
my.res <- msco::mJo.eng(my.files = my.files, algo = "sim2", Archetypes = TRUE,</pre>
             metric = "raw", nReps = 999, AICs = FALSE, params = FALSE,
             best.mod2 = FALSE, best.mod3 = FALSE, params_c.i = FALSE,
             my.r2 = FALSE, my.r2.s = FALSE)
my.res$Archs$`252.csv`
my.path2 <- system.file("extdata/myCSVs", package = "msco")</pre>
setwd(my.path2)
my.files2 <- gtools::mixedsort(list.files(path = my.path2, pattern = "*.csv"))</pre>
my.res2 <- msco::mJo.eng(my.files = my.files2[250:255], algo = "sim2", Archetypes = FALSE,</pre>
              metric = "raw", nReps = 999, AICs = FALSE, params = TRUE,
              best.mod2 = FALSE, best.mod3 = FALSE, params_c.i = FALSE,
              my.r2 = FALSE, my.r2.s = FALSE)
my.res2
my.path2 <- system.file("extdata/myCSVs", package = "msco")</pre>
setwd(my.path2)
my.files2 <- gtools::mixedsort(list.files(path = my.path2, pattern = "*.csv"))</pre>
my.res3 <- msco::mJo.eng(my.files = my.files2[250:255], algo = "sim2", Archetypes = FALSE,</pre>
              metric = "raw", nReps = 999, AICs = FALSE, params = FALSE,
              best.mod2 = FALSE, best.mod3 = FALSE, params_c.i = TRUE,
              my.r2 = FALSE, my.r2.s = FALSE)
my.res3
## End(Not run)
```

msco.res

Results on msco illustration (presented in Lagat et al., 2021c)

Description

This function allows the replication of the results on msco R package illustration paper presented in Lagat *et al.* (2021c). Executing msco.res() therefore gives these outputs that are saved as .RDS files in msco. If the codes that produced these (saved) outcomes are desired, the codes below are made available.

Usage

```
msco.res()
```

Value

Returns all the results presented in Lagat et al. (2021c). To replicate

• Figs. 1, 2 and Table 2, execute the following code:

30 msco.res

```
RNGkind(sample.kind = "Rejection")
  set.seed(14)
  ex.data <- read.csv(system.file("extdata", "251.csv", package = "msco"))</pre>
  j.en <- msco::Jo.eng(ex.data,</pre>
               algo = "sim2",
               metric = "raw",
               nReps = 999,
               dig = 3,
               s.dplot = FALSE,
               All.plots = TRUE,
               Jo.coeff = TRUE,
               my.AIC = TRUE,
               my.rsq = TRUE,
               Exp_Reg = TRUE,
               P.law_Reg = TRUE,
               Exp_p.l_Reg = TRUE,
               Obs.data = FALSE,
               Sim.data = FALSE,
               Jo_val.sim = FALSE,
               lab = FALSE,
               leg = FALSE,
               C.I_Jo_val.sim = FALSE,
               Jo_val.obs = TRUE,
               Metric = TRUE,
               Algorithm = TRUE,
               S.order = TRUE,
               nmod_stats = TRUE,
               Pt_Arch_Vals = TRUE,
               Atype = TRUE,
               p.n.plot = TRUE,
               trans = FALSE,
               m.n.plot = FALSE)
  j.en$jo.coeff ## Table 1
  j.en$AIC; j.en$r2 ## Table 2
  j.en$nmod_stats ## Table 3
  grDevices::dev.new()
  j.en$all.plots
• Fig. 4, execute the following code:
  RNGkind(sample.kind = "Rejection")
  set.seed(14)
  grDevices::dev.new()
  msco:::nullmod_archs2()
• Fig. 5, execute the following code:
  my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
  setwd(my.path)
  s.data <- get(load("s.data.csv")) #Species-by-site matrix</pre>
  t.data <- get(load("t.data.csv")) #Species-by-trait matrix</pre>
  p.d.mat <- get(load("p.d.mat.csv")) #Species-by-species phylogenetic distance matrix</pre>
```

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```
RNGkind(sample.kind = "Rejection")
set.seed(1)
gb.res <- msco::gbsm_m.orders(s.data,</pre>
            t.data,
            p.d.mat,
            metric = "Simpson_eqn",
            gbsm.model,
            orders = c(3:5, 8, 10, 15, 20),
            d.f = 4,
            degree = 3,
            n = 1000,
            k = 5,
            p = 0.8
            type = "k-fold",
            scat.plots = FALSE,
            response.curves = TRUE,
            j.occs.distrbn = FALSE,
            mp.plots = FALSE,
            max.vif = 10,
            \max.vif2 = 3,
            start.range=c(-0.1,0)
          )
gb.res$Original.VIFs$`order 3`
gb.res$Intermediate.VIFs$`order 3` ## Resulting covariate VIFs after removing
                                       ## covariates with VIF > max.vif
gb.res$Final.VIFs$`order 3` ## Resulting covariate VIFs after removing
                                 ## covariates with VIF > max.vif2
```

Note

The function msco.res is not for general use. We included it in this package to help the readers of Lagat *et al.* (2021c) paper, who may want to get a deeper understanding of how the results presented in this paper were arrived at. It also allows deeper scrutiny of Lagat *et al.* (2021c)'s findings, and broader understanding of the main functionalities of msco R package.

References

- 1. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.
- 2. Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.
- 3. Lagat, V. K., Latombe, G. and Hui, C. (2021c). msco: an R software package for null model testing of multi-species interactions and interference with covariates. DOI: <To be added>.

Examples

```
## Not run:
ms.res <- msco::msco.res()
ms.res$nmod_stats ## Table 2</pre>
```

32 pred.error.bands

```
## End(Not run)
```

pred.error.bands

Prediction uncertainty

Description

This function plots the response curves showing the effect of the predictors (i.e. trait-based and neutral forces) on joint occupancy as the response variable, with prediction error bands (as the standard deviation from the mean of the response variable) for all orders of joint occupancy.

Usage

```
pred.error.bands(
    s.data,
    t.data,
    p.d.mat,
    metric = "Simpson_eqn",
    gbsm.model,
    d.f = 4,
    simm = 10,
    orders,
    degree = 3,
    n = 1000,
    max.vif = 40,
    max.vif2 = 30,
    start.range = c(-0.1, 0)
)
```

Arguments

start.range

As for gbsm_m.orders.

s.data	A species-by-site presence/absence data.frame with entries indicating occurrence (1) and non-occurrence (0) of species in a site.
t.data	A data. frame with traits as columns and species as rows. The species must be the same as in ${\tt s.data}$.
p.d.mat	A symmetric matrix with dimnames as species and entries indicating the phylogenetic distance between any two of them (species).
metric	As for gbsm_m.orders.
gbsm.model	As for gbsm_m.orders.
d.f	As for gbsm_m.orders.
simm	Number of Monte Carlo simulations performed
orders	As for gbsm_m.orders
degree	As for gbsm_m.orders.
n	As for gbsm_m.orders.
max.vif	As for gbsm.
max.vif2	As for gbsm.

pred.error.bands 33

Value

• the response curves with prediction error bands for all orders of joint occupancy

References

- 1. Lagat, V. K., Latombe, G. and Hui, C. (2021a). *A multi-species co-occurrence index to avoid type II errors in null model testing*. DOI: <To be added>.
- 2. Lagat, V. K., Latombe, G. and Hui, C. (2021b). Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.

Examples

```
## Not run:
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
 setwd(my.path)
 s.data <- get(load("s.data.csv")) ## Species-by-site matrix</pre>
 t.data <- get(load("t.data.csv")) ## Species-by-Trait matrix</pre>
p.d.mat <- get(load("p.d.mat.csv")) ## Species-by-species phylogenetic distance matrix</pre>
RNGkind(sample.kind = "Rejection")
 set.seed(1)
pe <- msco::pred.error.bands(s.data, t.data, p.d.mat, metric="Simpson_eqn", d.f=4, simm=10,</pre>
 orders = c(2:5, 8, 10, 15), degree=3, n=1000, gbsm.model, start.range=c(-0.2, 0))
 pe$predictors$`order 2`
 pe$responses$`order 2`
pe$responses.sim_stats$`order 2`
 pe$predictors$`order 3`
 pe$responses$`order 3`
 pe$responses.sim_stats$`order 3`
 pe$predictors$`order 10`
pe$responses$`order 10`
pe$responses.sim_stats$`order 10`
## End(Not run)
```

34 s.phylo

s.phylo	Species phylogeny generator	

Description

This function generates the phylogeny of species and plots the phylogenetic tree. In particular, given a species-by-site matrix (community), s.phylo:

- uses the tax_name function to obtain (from the NCBI or ITIS online databases) the genus and family taxa levels of species in the community. If NCBI is used, getting an API key is recommended. See tax_name for more information. NCBI is used as default in this function;
- uses the phylo.maker function to obtain the phylogeny (an object of class: "phylo") of species in the community using taxa obtained above;
- computes the phylogenetic distance matrix using the cophenetic.phylo function and the phylogeny obtained above as input;
- plots the phylogenetic tree using the phylogeny obtained above.

Usage

```
s.phylo(
    s.data,
    p.d.mat,
    database = "ncbi",
    obs.taxa = FALSE,
    taxa.levels = NULL,
    Obs.data = FALSE,
    phy.d.mat = TRUE,
    phylo.plot = TRUE
```

Arguments

s.data	A species-by-site presence/absence data.frame with entries indicating occurrence (1) and non-occurrence (0) of species in a site. The rows should have species' scientific names following binomial nomenclature, with no initials.
p.d.mat	As for gbsm.
database	The online database used to obtain the taxonomic names (species, genus and family) for a given rank (species list in this function). The options are "ncbi" (default) or "itis".
obs.taxa	A Boolean indicating if taxa.levels should be included in the returned list.
taxa.levels	Species taxa (i.e. a data.frame with species, genus, and family as colnames) used in extracting phylogenetic distance matrix between species. If supplied, taxa.levels won't be computed from online repositories. Taxa provision is highly recommended.
Obs.data	A Boolean indicating if s.data should be included in the returned list.
phy.d.mat	A Boolean indicating if phylogenetic distance matrix should be in the returned list.
phylo.plot	Boolean value indicating if the phylogenetic tree (cluster dendrogram) should be plotted.

s.phylo 35

Value

Returns a list with the following outputs:

- s.data: A data.frame with sites as columns and species as rows.
- taxa.levels: A data.frame with the following columns:
 - species: Species names in s.data.
 - genus: Genus names of species in s.data.
 - family: Family names of species in s.data.
- p.d.matrix: A symmetric matrix with dimension names as species and entries indicating the phylogenetic distance between any two of them (species).
- phylo.plot: A phylogenetic tree (cluster dendrogram) of species in s.data

References

- 1. Binomial nomenclature' (2020) Wikipedia. Available at: https://en.wikipedia.org/wiki/Binomial_nomenclature (Accessed: 09 November 2020).
- 2. Lagat, V. K., Latombe, G. and Hui, C., 2021b. Dissecting the effects of random encounter versus functional trait mismatching on multi-species co-occurrence and interference with generalised B-spline modelling. DOI: <To be added>.

Examples

```
## Not run:
remotes::install_github("jinyizju/V.PhyloMaker", force = TRUE)
library(V.PhyloMaker)
my.path <- system.file("extdata/gsmdat", package = "msco")</pre>
setwd(my.path)
s.data <- get(load("s.data.csv"))</pre>
taxa <- get(load("taxa.levels.csv"))</pre>
my.s.phylo <- msco::s.phylo(s.data, p.d.mat = NULL, database = "ncbi", obs.taxa=TRUE,</pre>
 taxa.levels = taxa, Obs.data=TRUE, phy.d.mat=TRUE, phylo.plot = TRUE)
my.s.data <- my.s.phylo$s.data</pre>
my.s.data
my.taxa <- my.s.phylo$taxa.levels</pre>
my.taxa
my.p.d.mat <- my.s.phylo$phylogenetic.distance.matrix</pre>
my.p.d.mat
## End(Not run)
```

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