

# Package ‘blastar’

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**Title** BLAST and Sequence Analysis Tools

**Version** 0.1.1

**Description** Description: Provides streamlined tools for retrieving sequences from NCBI, performing sequence alignments (pairwise and multiple), and building phylogenetic trees. Implements the Needleman-Wunsch algorithm for global alignment (Needleman & Wunsch (1970) <[doi:10.1016/0022-2836\(70\)90057-4](https://doi.org/10.1016/0022-2836(70)90057-4)>), Smith-Waterman for local alignment (Smith & Waterman (1981) <[doi:10.1016/0022-2836\(81\)90087-5](https://doi.org/10.1016/0022-2836(81)90087-5)>), and Neighbor-Joining for tree construction (Saitou & Nei (1987) <[doi:10.1093/oxfordjournals.molbev.a040454](https://doi.org/10.1093/oxfordjournals.molbev.a040454)>).

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**Imports** rentrez, ape, Biostrings, dplyr, tibble

**Suggests** msa, pwalgn, testthat (>= 3.0.0)

**Config/testthat/edition** 3

**URL** <https://github.com/loukesio/blastar>

**BugReports** <https://github.com/loukesio/blastar/issues>

**NeedsCompilation** no

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align\_sequences      *Align DNA Sequences (Pairwise or Multiple)*


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### Description

This function takes a tibble with a "sequence" column (and optional "accession" names) and performs either a pairwise alignment between two sequences or a multiple sequence alignment (MSA) across all.

### Usage

```
align_sequences(
  df,
  method = c("pairwise", "msa"),
  pairwise_type = "global",
  msa_method = "ClustalOmega",
  seq_indices = c(1, 2)
)
```

### Arguments

df	A tibble or data.frame containing at least: <ul style="list-style-type: none"> <li>sequence: character vector of DNA sequences</li> <li>accession (optional): names for each sequence; if present, they will be used as identifiers in the alignment object.</li> </ul>
method	One of: <ul style="list-style-type: none"> <li>"pairwise": perform a pairwise alignment between two sequences</li> <li>"msa": perform a multiple sequence alignment on all sequences</li> </ul>
pairwise_type	For pairwise only, alignment type: "global" (Needleman–Wunsch), "local" (Smith–Waterman), or "overlap".
msa_method	For MSA only, method name: "ClustalOmega", "ClustalW", or "Muscle".
seq_indices	Integer vector of length 2; indices of the two sequences to align when method = "pairwise". Defaults to c(1, 2).

### Value

If method="pairwise", a list with:

- alignment: a PairwiseAlignmentsSingleSubject object
- pid: percent identity (numeric) If method="msa", an object of class MsaDNAMultipleAlignment or similar.

**Examples**

```
# Pairwise alignment example (requires pwalgn package)
if (requireNamespace("pwalgn", quietly = TRUE)) {
  data <- data.frame(
    accession = c("seq1", "seq2"),
    sequence = c("ACGTACGTACGT", "ACGTACGTTTGT"),
    stringsAsFactors = FALSE
  )

  res_pw <- align_sequences(
    df = data,
    method = "pairwise",
    pairwise_type = "global"
  )
  res_pw$pid
}

# Multiple sequence alignment (requires msa package)
if (requireNamespace("msa", quietly = TRUE)) {
  data_msa <- data.frame(
    accession = c("seq1", "seq2", "seq3"),
    sequence = c("ATGCATGC", "ATGCTAGC", "ATGGATGC")
  )
  res_msa <- align_sequences(data_msa, method = "msa", msa_method = "ClustalOmega")
  print(res_msa)
}
```

build\_nj\_tree

*Build a Neighbor-Joining tree from a multiple sequence alignment***Description**

This function takes a Multiple Sequence Alignment (MSA) object (e.g., output of `align_sequences(method = "msa")`) and generates a Neighbor-Joining (NJ) tree.

**Usage**

```
build_nj_tree(msa, model = "raw", pairwise.deletion = TRUE)
```

**Arguments**

<code>msa</code>	A multiple alignment object (class <code>MsaDNAMultipleAlignment</code> or similar)
<code>model</code>	Evolutionary model for distance calculation passed to <code>ape::dist.dna</code> (e.g., "raw", "JC69", "K80", etc.)
<code>pairwise.deletion</code>	Logical. If TRUE, compute distances with pairwise deletion

**Value**

An object of class phylo (NJ tree)

**Examples**

```
# Build NJ tree from multiple sequence alignment (requires msa package)
if (requireNamespace("msa", quietly = TRUE)) {
  # Create example sequences
  df <- data.frame(
    accession = c("seq1", "seq2", "seq3"),
    sequence = c("ATGCATGC", "ATGCTAGC", "ATGGATGC")
  )

  # Generate MSA
  msa_result <- align_sequences(df, method = "msa", msa_method = "ClustalOmega")

  # Build NJ tree
  tree <- build_nj_tree(msa_result, model = "raw")
  print(tree)
}
```

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fetch\_metadata

*Fetch Metadata (and optionally sequence ranges) from NCBI*

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**Description**

Fetch Metadata (and optionally sequence ranges) from NCBI

**Usage**

```
fetch_metadata(accessions, db = c("nuccore", "protein"), seq_range = NULL)
```

**Arguments**

accessions	Character vector of accession numbers.
db	Either "nuccore" or "protein".
seq_range	Either: <ul style="list-style-type: none"> <li>• NULL (default): fetch full sequence for every accession</li> <li>• numeric(2): fetch that same start–end for <i>all</i> accessions</li> <li>• named list: each element is a numeric(2) vector, names are accessions; will fetch only that slice for the named accession, full sequence for others.</li> </ul>

**Value**

A tibble with columns accession, accession\_version, title, organism, sequence

**Examples**

```
# Fetch metadata for a nucleotide sequence
result <- fetch_metadata("NM_000546", db = "nucore")

# Fetch specific sequence range (positions 1-100)
result_range <- fetch_metadata("NM_000546", db = "nucore", seq_range = c(1, 100))

# Fetch multiple accessions
result_multi <- fetch_metadata(c("NM_000546", "NM_001126"), db = "nucore")
```

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