# Package 'BALLI'

October 12, 2022

Type Package
Title Expression RNA-Seq Data Analysis Based on Linear Mixed Model
Version 0.2.0
Author Kyungtaek Park <qkrrudxor147@snu.ac.kr></qkrrudxor147@snu.ac.kr>
Maintainer Kyungtaek Park <qkrrudxor147@snu.ac.kr></qkrrudxor147@snu.ac.kr>
<b>Description</b> Analysis of gene expression RNA-seq data using Bartlett-Adjusted Likelihood-based LInear model (BALLI). Based on likelihood ratio test, it provides comparisons for effect of one or more variables. See Kyungtaek Park (2018) <doi:10.1101 344929=""> for more information.</doi:10.1101>
<b>Depends</b> R (>= 2.15.0), edgeR, limma, MASS, parallel, stats, methods
License GPL
Encoding UTF-8
LazyData true
RoxygenNote 6.1.1
Suggests knitr, rmarkdown
VignetteBuilder knitr
NeedsCompilation no
Repository CRAN
<b>Date/Publication</b> 2019-04-25 10:40:07 UTC
balli       2         Balli-class       3         balliFit       3         LargeDataObject-class       4         tecVarEstim       4         TecVarList-class       5
Index 6

2 balli

balli BALLI	
-------------	--

#### **Description**

DEG analysis using BALLI algorithm

#### Usage

```
balli(object, intV = 2, logcpm = NULL, tecVar = NULL,
  design = NULL, numCores = NULL, threshold = 1e-06, maxiter = 200)
```

#### **Arguments**

object	a TecVarList object
intV	numeric vector designating interest variable(s) which is (are) column number(s) of design matrix
logcpm	logcpm values for each gene and each sample
tecVar	estimated technical variance values for each gene and each sample
design	design matrix with samples in row and covariable(s) to be estimated in column
numCores	number of cores to be used for multithreding. If NULL, a single core is used
threshold	threshold for convergence
maxiter	maximum number of iteration to converge of estimated biological variance. If not, biological variance is estimated by using Brent method

#### Value

an Balli object including Result and topGenes list. Following components are shown by Result (same order of genes with input data) and topGenes (ordered by pBALLI in Result):

```
log2FClog2 fold changes of interest variable(s)lLLIlog-likelihoods estimated by LLIlBALLIlog-likelihoods estimated by BALLIpLLIp-values estimated by LLIpBALLIp-values estimated by BALLIBCFBartlett's correction factor
```

expr <- data.frame(t(sapply(1:1000,function(x)rnbinom(20,mu=500,size=50)))) group <- c(rep("A",10),rep("B",10)) design <- model.matrix(~group, data = expr) dge <- DGEList(counts=expr, group=group) dge <- calcNormFactors(dge) tV <- tecVarEstim(dge,design) balli(tV,intV=2)

Balli-class 3

Balli-class	Class Balli Class Balli holds results from BALLI	

## Description

Class Balli Class Balli holds results from BALLI

|--|--|

## Description

Estimates likelihood and Bartlett correction factor using BALLI algorithm of each gene

## Usage

```
balliFit(y_mat, x_mat, tecVar, intVar = 2, full = T, cfault = 0,
  miter = 200, conv = 1e-06)
```

## Arguments

y_mat	numeric vector containing log-cpm values of each gene and each sample
x_mat	design matrix with samples in row and covariable(s) to be estimated in column
tecVar	numeric vector containing estimated technical variance of a gene of each sample
intVar	numeric vector designating interest variable(s) which is (are) column number(s) of x_mat
full	logical value designating full model (TRUE) or reduced model (FALSE).
cfault	initial value of index showing whether converged (0) or not (1).
miter	maximum number of iteration to converge.
conv	threshold for convergence

#### Value

following components are estimated

11	log-likelihoods
beta	coefficients of interested variable(s) $\\$
alpha	coefficients of nuisance variable(s)
BCF	Bartlett's correction factor
cfault	index whether converged or not

4 tec VarEstim

#### **Examples**

```
expr <- data.frame(t(sapply(1:1000,function(x)rnbinom(20,mu=500,size=50))))
group <- c(rep("A",10),rep("B",10))
design <- model.matrix(~group, data = expr)
dge <- DGEList(counts=expr, group=group)
dge <- calcNormFactors(dge)
tV <- tecVarEstim(dge,design)
gtv <- tV$tecVar[1,]
gdat <- data.frame(logcpm=tV$logcpm[1,],design,tecVar=gtv)
gy <- matrix(unlist(gdat[,1]),ncol=1)
gx <- matrix(unlist(gdat[,2:(ncol(gdat)-1)]),ncol=ncol(gdat)-2)
balliFit(y_mat=gy,x_mat=gx,tecVar=gtv,intVar=2,full=TRUE,cfault=0,miter=200,conv=1e-6)</pre>
```

LargeDataObject-class Class LargeDataObject Class LargeDataObject holds large data such as technical variance and results from BALLI fit

#### Description

Class LargeDataObject Class LargeDataObject holds large data such as technical variance and results from BALLI fit

tecVarEstim Technical Variance Estimation

#### **Description**

Estimate technical variance by using voom-trend. The code is derived from voom function in limma package

#### Usage

```
tecVarEstim(counts, design = NULL, lib.size = NULL, span = 0.5, ...)
```

## Arguments

counts	a DGEList object
design	design matrix with samples in row and coefficient(s) to be estimated in column
lib.size	numeric vector containing total library sizes for each sample
span	width of the lowess smoothing window as a proportion
	other arguments are passed to lmFit.

TecVarList-class 5

#### Value

an TecVarList object with the following components:

targets matrix containing covariables, library sizes and normalization foctors of each

sample

design matrix with samples in row and covariable(s) to be estimated in column

logcpm values of each gene and each sample

tecVar estimated techical variance of each gene and each sample

#### **Examples**

```
expr <- data.frame(t(sapply(1:1000,function(x)rnbinom(20,mu=500,size=50))))
group <- c(rep("A",10),rep("B",10))
design <- model.matrix(~group, data = expr)
dge <- DGEList(counts=expr, group=group)
dge <- calcNormFactors(dge)
tecVarEstim(dge,design)</pre>
```

TecVarList-class

Class TecVarList Class TecVarList holds technical variance

## Description

Class TecVarList Class TecVarList holds technical variance

## **Index**

```
balli, 2
Balli-class, 3
balliFit, 3

LargeDataObject-class, 4
tecVarEstim, 4
TecVarList-class, 5
```