# Package 'Arothron'

# February 1, 2023

Type Package  Title Geometric Morphometric Methods and Virtual Anthropology Tools  Version 2.0.5  Author Antonio Profico, Costantino Buzi, Silvia Castiglione, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano  Maintainer Antonio Profico <antonio.profico@gmail.com>  Description Tools for geometric morphometric analysis. The package includes tools of virtual anthropology to align two not articulated parts belonging to the same specimen, to build virtual cavities as endocast (Profico et al. 2021 <doi:10.1002 ajpa.24340="">).  Depends R (&gt;= 3.5.0)  Imports abind (&gt;= 1.4), alphashape3d (&gt;= 1.3), compositions (&gt;= 1.40-1), doParallel (&gt;= 1.0.11), foreach (&gt;= 1.44), geometry (&gt;= 0.3-6), graphics(&gt;= 3.4.0), grDevices(&gt;= 3.4.0), methods (&gt;= 3.5), Morpho (&gt;= 2.5.0), parallel (&gt;= 1.0), rgl (&gt;= 1.0.1), Rvcg (&gt;= 0.17), stats (&gt;= 3.4.0), stats4 (&gt;= 4.0), stringr (&gt;= 1.3.0), utils (&gt;= 3.4.0), vegan (&gt;= 2.4)  License GPL-2  Encoding UTF-8  LazyLoad yes  RoxygenNote 7.2.3  NeedsCompilation no  Repository CRAN  Date/Publication 2023-02-01 12:40:08 UTC  R topics documented:  Arothron-package  Altapic  aro.clo.points  arraytolist  bary.mesh</doi:10.1002></antonio.profico@gmail.com>	
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Arothron-package 3 41 **42** Index Arothron-package eometric Morphometric Methods and Virtual Anthropology Tools **Description** Tools for geometric morphometric analysis. The package includes tools of virtual anthropology to align two not articulated parts belonging to the same specimen, to build virtual cavities as endocast (Profico et al, 2021 <doi:10.1002/ajpa.24340>). Author(s) Antonio Profico, Costantino Buzi, Silvia Castiglione, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano Altapic example dataset

# Description

2D image of the Altamura man fossil

# Usage

data(Altapic)

# Author(s)

Antonio Profico

4 arraytolist

aro.clo.points

aro.clo.points

# **Description**

Find the closest matches between a reference (2D or 3D matrix) and a target matrix (2D/3D) or mesh returning row indices and distances

#### Usage

```
aro.clo.points(target, reference)
```

# **Arguments**

target kxm matrix or object of class "mesh3d" reference numeric: a kxm matrix (coordinates)

#### Value

position numeric: a vector of the row indices

distances numeric: a vector of the coordinates distances

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Examples**

```
#load an example: mesh, and L set
data(yoda_sur)
data(yoda_set)
sur<-yoda_sur
set<-yoda_set
ver_pos<-aro.clo.points(target=sur,reference=set)</pre>
```

arraytolist

arraytolist

# **Description**

converts an array in a list storing each element of the third dimension of the array (specimen) as element of the list

#### Usage

```
arraytolist(array)
```

bary.mesh 5

# **Arguments**

array

a kx3xn array with landmark coordinates

#### Value

a list containing the landmark configurations stored as separated elements

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

bary.mesh

bary.mesh

# Description

This function calculates the barycenter of a matrix or a 3D mesh

# Usage

```
bary.mesh(mesh)
```

#### **Arguments**

mesh

matrix mesh vertex

### Value

barycenter numeric: x,y,z coordinates of the barycenter of the mesh

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Examples**

```
#load an example: mesh, and L set
data(SCP1.mesh)
sur<-SCP1.mesh
bary<-bary.mesh(mesh=sur)</pre>
```

6 compare\_check.set

# **Description**

This function applyes the Digital Alignment Tool (DTA) on a disarticulated model using a reference landmark configuration

#### Usage

```
compare_check.set(RM_set_1, RM_set_2, DM_set_1, DM_set_2, DM_mesh_1, DM_mesh_2)
```

#### **Arguments**

RM_set_1	matrix: 3D landmark set of the first module acquired on the reference model
RM_set_2	matrix: 3D landmark set of the second module acquired on the reference model
DM_set_1	matrix: 3D landmark set of the first module acquired on the disarticulated model
DM_set_2	matrix: 3D landmark set of the second module acquired on the disarticulated model
DM_mesh_1	mesh3d: mesh of the disarticulated model (first module)
DM_mesh_2	mesh3d: mesh of the disarticulated model (second module)

#### Value

SF1 numeric: scale factor used to scale the reference set (first module)

SF2 numeric: scale factor used to scale the reference set (second module)

RM\_set\_1\_sc matrix: scaled 3D reference set (first module)

RM\_set\_2\_sc matrix: scaled 3D reference set (second module)

AM\_model list: output of the Morpho::rotmesh.onto function

dist\_from\_mesh numeric: mesh distance between the aligned model and the scaled reference set

eucl\_dist\_1 numeric: euclidean distance between the landmark configuration of the disarticulated and reference model (first module)

eucl\_dist\_2 numeric: euclidean distance between the landmark configuration of the disarticulated and reference model (second module)

procr\_dist numeric: procrustes distance between the landmark configuration of the aligned and reference model

procr\_dist\_1 numeric: procrustes distance between the landmark configuration of the disarticulated and reference model (first module)

procr\_dist\_2 numeric: procrustes distance between the landmark configuration of the disarticulated and reference model (second module)

eucl\_dist numeric: euclidean distance between the landmark configuration of the aligned and reference model

CScorreffect 7

single\_l\_1 numeric: euclidean distance between the landmark configuration of the disarticulated and reference model (first module)

single\_1\_2 numeric: euclidean distance between the landmark configuration of the disarticulated and reference model (second module)

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

CScorreffect CScorreffect Plot showing the correlation in the shape space between original and combined dataset omitting or including the normalization factors calculated with Arothron and MLECScorrection

#### **Description**

CScorreffect Plot showing the correlation in the shape space between original and combined dataset omitting or including the normalization factors calculated with Arothron and MLECScorrection

#### Usage

```
CScorreffect(
    array1,
    array2,
    nPCs = c(1:3),
    from = 0.02,
    to = 0.9,
    length.out = 100
)
```

### **Arguments**

array1 array: first set of landmark configuration
array2 array: second set of landmark configuration

nPCs numeric vector: specify which PC scores will be selected in the correlation test

from numeric: the lower interval of the normalization factor distribution to numeric: the lower interval of the normalization factor distribution

length.out numeric: number of values ranged between from and to

#### Value

PCscores PCscores matrix of the combined dataset applying the normalization factor calculated by using the maximum likelihood estimation

PCs PCs matrix of the combined dataset applying the normalization factor calculated by using the maximum likelihood estimation

corr mean correlation between original and combined dataset

CSratios normalization factor calculated by using the maximum likelihood estimation

8 dec.curve

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

### **Examples**

```
## Not run:
# Femora case study
data(femsets)
all_pois<-matrix(1:(200*61),nrow=61,ncol=200,byrow = FALSE)
set_ext_100<-femsets[all_pois[,1:100],,]
set_int_100<-femsets[all_pois[,101:200],,]
set_int_50<-set_int_100[c(matrix(1:6100,ncol=61)[seq(1,100,2),]),,]
set_int_20<-set_int_100[c(matrix(1:6100,ncol=61)[seq(1,100,5),]),,]
set.seed(123)
sel<-sample(1:100,10)
set_int_10r<-set_int_100[c(matrix(1:6100,ncol=61)[sel,]),,]

CScorreffect(set_ext_100,set_int_50,nPCs=1:3)
CScorreffect(set_ext_100,set_int_20,nPCs=1:3)

CScorreffect(set_ext_100,set_int_10r,nPCs=1:3)

## End(Not run)</pre>
```

dec.curve

dec.curve

#### **Description**

This function computes the order of points on a open 3D curve and finds intermediate points

# Usage

```
dec.curve(mat_input, mag, plot = TRUE)
```

#### **Arguments**

mat\_input numeric: a kx3 matrix

numeric: how many times will be divided by the number of initial points plot logical: if TRUE will be plotted the starting and final point matrices

#### Value

matt numeric: a kx3 matrix with points coordinates

#### Author(s)

DM\_base\_sur

#### **Examples**

```
## Not run:
## Create and plot a 3D curve
require(compositions)
require(rgl)
curve_3D<-cbind(1:10,seq(1,5,length=10),rnorm(10,sd = 0.2))
plot3D(curve_3D,bbox=FALSE)
close3d()
## Create and plot the new 3D curve (with intermediate points)
dec_curve_3D<-dec.curve(curve_3D, 2, plot = TRUE)
## End(Not run)</pre>
```

DM\_base\_sur

example dataset

# Description

3D mesh of the first part of the Homo sapiens disarticulated model

#### Usage

```
data(DM_base_sur)
```

### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

DM\_face\_sur

example dataset

#### **Description**

3D mesh of the second part of the Homo sapiens disarticulated model

# Usage

```
data(DM_face_sur)
```

#### Author(s)

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DM\_set example dataset

# Description

Landmark configurations of the two part of the disarticulated model

# Usage

```
data(DM_set)
```

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

dta dta

# Description

This function applyes the Digital Alignment Tool (DTA) on a disarticulated model using a reference sample

# Usage

```
dta(
   RM_sample,
   mod_1,
   mod_2,
   pairs_1,
   pairs_2,
   DM_mesh_1,
   DM_mesh_2,
   DM_set_1,
   DM_set_2,
   method = c("euclidean")
)
```

# Arguments

RM\_sample 3D array: 3D landmark configurations of the reference sample

mod\_1 numeric vector: vector containing the position of which landmarks belong to the
first module

mod\_2 numeric vector: vector containing the position of which landmarks belong to the
second module

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pairs_1	matrix: a X x 2 matrix containing the indices of right and left landmarks of the first module
pairs_2	matrix: a X x 2 matrix containing the indices of right and left landmarks of the second module
DM_mesh_1	mesh3d: mesh of the disarticulated model (first module)
DM_mesh_2	mesh3d: mesh of the disarticulated model (second module)
DM_set_1	matrix: 3D landmark set of the first module acquired on the disarticulated model
DM_set_2	matrix: 3D landmark set of the second module acquired on the disarticulated model
method	character: specify method to be used to individuate the best DTA ("euclidean" or "procrustes")

#### Value

AM\_mesh mesh3d: mesh of the aligned model

AM\_set matrix: landmark configuration of the aligned model

AM\_id character: name of the item of the reference sample resulted as best DTA

AM\_SF\_1 numeric: scale factor used to scale the reference set (first module)

AM\_SF\_2 numeric: scale factor used to scale the reference set (second module)

distance numeric: distance between the landmark configuration of the aligned and the reference model

tot\_proc numeric vector: procrustes distances between aligned and reference models (all DTAs)

tot eucl numeric vector: euclidean distances between aligned and reference models (all DTAs)

setarray 3D array: landmark configurations of the disarticulated model aligned on each item of the reference sample

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

#### References

Profico, A., Buzi, C., Davis, C., Melchionna, M., Veneziano, A., Raia, P., & Manzi, G. (2019). A new tool for digital alignment in Virtual Anthropology. The Anatomical Record, 302(7), 1104-1115.

#### **Examples**

```
## Load and plot the disarticulated model of the Homo sapiens case study
library(compositions)
library(rgl)
data(DM_base_sur)
data(DM_face_sur)
open3d()
wire3d(DM_base_sur,col="white")
wire3d(DM_face_sur,col="white")
```

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```
## Load the landmark configurations associated to the DM
data(DM_set)
## Load the reference sample
data(RMs_sets)
## Define the landmarks belonging to the first and second module
mod_1<-c(1:17) #cranial base
mod_2<-c(18:32) #facial complex
## Define the paired landmarks for each module (optional symmetrization process)
pairs_1<-cbind(c(4,6,8,10,12,14,16),c(5,7,9,11,13,15,17))
pairs_2<-cbind(c(23,25,27,29,31),c(24,26,28,30,32))
## Run DTA
ex.dta<-dta(RM_sample=RMs_sets, mod_1=mod_1, mod_2=mod_2, pairs_1=pairs_1, pairs_2=pairs_2,
DM_mesh_1=DM_base_sur,DM_mesh_2=DM_face_sur, DM_set_1= DM_set[mod_1,], DM_set_2=DM_set[mod_2,])
## Print the name of the best RM
ex.dta$AM_id
## Save the mesh and the landmark set of the AM
AM_mesh<-ex.dta$AM_mesh
AM_set<-ex.dta$AM_set
## Plot the aligned 3D model
library(compositions)
library(rgl)
open3d()
wire3d(AM_mesh,col="white")
plot3D(AM_set,bbox=FALSE,add=TRUE)
```

endomaker

endomaker

# **Description**

Build endocast from a skull 3D mesh

### Usage

```
endomaker(
  mesh = NULL,
  path_in = NULL,
  param1_endo = 1,
  npovs = 50,
  volume = TRUE,
  alpha_vol = 100,
  nVoxels = 1e+05,
  decmesh = 20000,
  alpha_ext = 30,
  ncells = 50000,
  npovs_calse = 50,
  param1_calse = 2,
  param1_ast = 1.3,
  decendo = 20000,
```

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```
scalendo = 0.5,
alpha_end = 100,
mpovdist = 10,
plot = FALSE,
colmesh = "orange",
save = FALSE,
outpath = tempdir(),
num.cores = NULL
)
```

#### **Arguments**

mesh mesh3d: 3D model of the skull

path\_in character: path of the skull where is stored param1\_endo numeric: parameter for spherical flipping

npovs numeric: number of Points of View used in the endocast construction

volume logical: if TRUE the calculation of the volume (expressed in cc) through con-

cave is returned

alpha\_vol numeric: alpha shape for volume calculation

nVoxels numeric: number of voxels for estimation endocranial volume

decmesh numeric: decmesh

alpha\_ext numeric: alpha shape for construction external cranial mesh numeric: approximative number of cell for 3D grid construction numeric: number of Points of View for construction of skull shell

param1\_calse numeric: parameter for calse (construction shell)

param1\_ast numeric: parameter for ast3d (construction row endocast)

decendo numeric: desired number of triangles (row endocast)

scalendo numeric: scale factor row endocast (for definition of POVs)
alpha\_end numeric: alpha shape value for concave hull (row endocast)

mpovdist numeric: mean value between POVs and mesh

plot logical: if TRUE the endocast is plotted colmesh character: color of the mesh to be plotted

save logical: if TRUE the mesh of the endocast is saved

outpath character: path where save the endocast

num. cores numeric: numbers of cores to be used in parallel elaboration

#### Value

endocast mesh3d: mesh of the endocast

volume numeric: volume of the endocast expressed in cc

14 endomaker\_dir

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

#### References

Profico, A., Buzi, C., Melchionna, M., Veneziano, A., & Raia, P. (2020). Endomaker, a new algorithm for fully automatic extraction of cranial endocasts and the calculation of their volumes. American Journal of Physical Anthropology.

# Examples

# Description

Build library of endocasts from skull 3D meshes

#### Usage

```
endomaker_dir(
 dir_path,
 param1\_endo = 1.5,
 npovs = 50,
  volume = TRUE,
  alpha_vol = 50,
 nVoxels = 1e+05,
  decmesh = 20000,
  alpha_ext = 30,
  ncells = 50000,
  npovs\_calse = 50,
  param1\_calse = 3,
  param1_ast = 1.3,
  decendo = 20000,
  scalendo = 0.5,
  alpha_end = 100,
 mpovdist = 10,
 plotall = FALSE,
```

endomaker\_dir 15

```
colmesh = "orange",
  save = FALSE,
  outpath = tempdir(),
  num.cores = NULL
)
```

#### **Arguments**

dir\_path character: path of the folder where the skull meshes are stored

param1\_endo numeric vector: parameter for spherical flipping

npovs numeric: number of Points of View used in the endocast construction volume logical: if TRUE the volume of the endocast (ECV) is estimated

alpha\_vol numeric: alpha shape for volume calculation

nVoxels numeric: number of voxels for estimation endocranial volume

decmesh numeric: decmesh

alpha\_ext numeric: alpha shape for construction external cranial mesh numerls numeric: approximative number of cell for 3D grid construction npovs\_calse numeric: number of Points of View for construction of skull shell

param1\_calse numeric: parameter for calse (construction shell)

param1\_ast numeric: parameter for ast3d (construction row endocast) decendo numeric: desired number of triangles (row endocast)

scalendo numeric: scale factor row endocast (for definition of POVs)
alpha\_end numeric: alpha shape value for concave hull (row endocast)
mpovdist numeric vector: mean value between POVs and mesh

plotall logical: if TRUE the endocasts are plotted colmesh character: color of the mesh to be plotted

save logical: if TRUE the mesh of the endocast is saved

outpath character: path where save the endocast

num. cores numeric: number of cores to be used in parallel elaboration

#### Value

endocasts mesh3d: list of meshes of the extracted endocasts volumes numeric: volumes of the endocasts expressed in cc

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

#### References

Profico, A., Buzi, C., Melchionna, M., Veneziano, A., & Raia, P. (2020). Endomaker, a new algorithm for fully automatic extraction of cranial endocasts and the calculation of their volumes. American Journal of Physical Anthropology.

16 export\_amira

endo\_set

example dataset

# Description

POVs defined inside the endocranial cavity

# Usage

```
data(endo_set)
```

### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

export\_amira

export\_amira

# Description

This function exports a list of 3D landmark set in separate files (format landmarkAscii)

# Usage

```
export_amira(lista, path)
```

# Arguments

list containing 3D landmark sets

path character: path of the folder where saving the Amira landmark sets

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# Examples

```
x<-c(1:20)
y<-seq(1,3,length=20)
z<-rnorm(20,0.01)
vertices<-cbind(x,y,z)
set<-list(vertices)
example<-export_amira(set,path=tempdir())</pre>
```

export\_amira.path 17

export\_amira.path export\_amira.path

# **Description**

Convert and save a 3D matrix into a AmiraMesh ASCII Lineset (.am) object

# Usage

```
export_amira.path(
  vertices,
  filename,
  Lines = c(1:(dim(vertices)[1] - 1) - 1, -1),
  path
)
```

# Arguments

vertices numeric: a kx3 matrix

filename character: name of the requested output

Lines numeric: sequence of the vertices that defines the line

path character: folder path

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Examples**

```
x<-c(1:20)
y<-seq(1,3,length=20)
z<-rnorm(20,0.01)
vertices<-cbind(x,y,z)
export_amira.path(vertices=vertices,filename="example_line",path=tempdir())</pre>
```

ext.int.mesh ext.int.mesh

# **Description**

This function finds the vertices visible from a set of points of view

18 ext.int.mesh

#### Usage

```
ext.int.mesh(
  mesh,
  views = 20,
  dist.sphere = 3,
  param1 = 2.5,
  param2 = 10,
  default = TRUE,
  import_pov,
  matrix_pov,
  expand = 1,
  scale.factor,
  method = "ast3d",
  start.points = 250,
  num.cores = NULL
)
```

### **Arguments**

mesh	object of class	mesh3d

views numeric: number of points of view

dist.sphere numeric: scale factor. This parameter the distance betweem the barycenter of

the mesh and the radius of the sphere used to define set of points of view

param1 numeric: first parameter for spherical flipping (usually ranged from 0.5 to 5,

try!)

param2 numeric second paramter for spherical flipping (don't change it!)

default logical: if TRUE the points of views are defined automatically, if FALSE define

the matrix\_pov

import\_pov logical: if NULL an interactive 3D plot for the definition of the points of view

is returned

matrix\_pov matrix: external set of points of view

expand numeric: scale factor for the grid for the interactive 3D plot scale.factor numeric: scale factor for sphere inscribed into the mesh

method character: select "a" or "c"

start.points numeric: number of POVs available

num.cores numeric: number of cores

#### Value

position numeric: a vector with vertex number nearest the landmark set

#### Author(s)

ext.mesh.rai 19

#### References

Profico, A., Schlager, S., Valoriani, V., Buzi, C., Melchionna, M., Veneziano, A., ... & Manzi, G. (2018). Reproducing the internal and external anatomy of fossil bones: Two new automatic digital tools. American Journal of Physical Anthropology, 166(4), 979-986.

ext.mesh.rai

ext.mesh.rai

# **Description**

This function returns a 3D mesh with colours based on the vertices visibile from each point of view

#### Usage

```
ext.mesh.rai(scans, mesh)
```

# **Arguments**

scans an ext.int.mesh

mesh matrix mesh vertex (the same of the ext.int.mesh object)

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

femsets example dataset

# **Description**

3D semilandmark configurations of 21 human femora

# Usage

data(femsets)

#### Author(s)

20 human\_skull

grid\_pov grid\_pov

# Description

This function creates a grid for an interactive way to define the set of the points of view

# Usage

```
grid_pov(mesh, expand = 1)
```

# **Arguments**

mesh object of class mesh3d

expand numeric: scale factor for the grid for the interactive 3D plot

# Value

matrice matrix: matrix with the x,y,z coordinates of the points of view

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

human\_skull example dataset

# **Description**

3D mesh of a human skull

# Usage

```
data(human_skull)
```

#### Author(s)

image2palettes 21

image2palettes

image 2 palettes

# Description

Create palettes from an image

# Usage

```
image2palettes(
  array,
  resize = 4,
  unique = FALSE,
  scale = F,
  k = 3,
  lcols = 7,
  plsaxis = 1,
  cex = 5,
  cext = 0.5
)
```

# Arguments

array	array: rgb array
resize	numeric: desidered resize factor
unique	logical: if TRUE each color is counted once
scale	logical: if TRUE (color) variables are scaled
k	numeric: desidered number of clusters (i.e., number of palettes)
lcols	numeric: length of the color vector of each palette
plsaxis	numeric: desidered PLS axis
cex	numeric: size of colored squares
cext	numeric: size of color names

# Value

paletteslist list: color palettes arranged in a list

# Author(s)

Antonio Profico

22 landmark\_frm2amira

#### **Examples**

```
## Not run:
require(jpeg)
require(Morpho)
data("Altapic")
image2palettes(Altapic,resize=1,unique=T,scale=T,k=3,lcols=5,plsaxis=1,cext=0.5)
## End(Not run)
```

krd1\_tooth

example dataset

#### **Description**

3D mesh of a decidous Neanderthal tooth

#### Usage

```
data(krd1_tooth)
```

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

landmark\_frm2amira

landmark\_frm2amira

#### **Description**

This function converts the .frm files, from Evan Toolbox, stored in a folder into the format land-markAscii

#### Usage

```
landmark_frm2amira(path_folder_frm, path_amira_folder)
```

# **Arguments**

### Author(s)

listtoarray 23

listtoarray

listtoarray convert a list into an array

# Description

listtoarray convert a list into an array

# Usage

```
listtoarray(mylist)
```

# **Arguments**

```
mylist a list
```

#### Value

a kx3xn array with landmark coordinates

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

localmeshdiff

localmeshdiff Calculate and Visualize local differences between two meshes

# **Description**

localmeshdiff Calculate and Visualize local differences between two meshes

# Usage

```
localmeshdiff(
  mesh1,
  mesh2,
  ploton = 1,
  diffarea = ((area_shape1 - area_shape2)/area_shape2) * 100,
  paltot = rainbow(200),
  from = NULL,
  to = NULL,
  n.int = 200,
  out.rem = TRUE,
  fact = 1.5,
  visual = 1,
  scale01 = TRUE,
  colwire = "pink"
)
```

24 localmeshdiff

#### **Arguments**

mesh1	reference mesh: object of class "mesh3d"
mesh2	target mesh: object of class "mesh3d"
ploton	numeric: define which mesh will be used to visualize local differences
diffarea	formula: define how calculating differences in area. area_shape1 refers to mesh1, area_shape2 refers to mesh2 $$
paltot	character vector: specify the colors which are used to create a color palette
from	numeric: minimum distance to be colorised
to	numeric: maximum distance to be colorised
n.int	numeric: determines break points for color palette
out.rem	logical: if TRUE outliers will be removed
fact	numeric: factor k of the interquartile range
visual	numeric: if equals to 1 the mesh is plotted without a wireframe, if set on 2 a wireframe is added
scale01	logical: if TRUE the vector of distances is scaled from 0 to 1
colwire	character: color of the wireframe

#### Value

vect numeric vector containing local differeces in area between the reference and the target mesh

#### Author(s)

Antonio Profico, Costantino Buzi, Silvia Castiglione, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

### References

Melchionna, M., Profico, A., Castiglione, S., Sansalone, G., Serio, C., Mondanaro, A., ... & Manzi, G. (2020). From smart apes to human brain boxes. A uniquely derived brain shape in late hominins clade. Frontiers in Earth Science, 8, 273.

# **Examples**

```
## Not run:
library(Arothron)
library(rgl)
data("primendoR")
neaset<-primendoR$sets[,,11]
sapset<-primendoR$sets[,,14]
#defining a mesh for the neanderthal right hemisphere
neasur<-list("vb"=t(cbind(neaset,1)),"it"=primendoR$sur$it)
class(neasur)<-"mesh3d"
#defining a mesh for the modern human right hemisphere
sapsur<-list("vb"=t(cbind(sapset,1)),"it"=primendoR$sur$it)
class(neasur)<-"mesh3d"</pre>
```

Lset2D\_list 25

```
layout3d(t(c(1,2)),sharedMouse = TRUE)
localmeshdiff(sapsur,neasur,1,scale01 = TRUE,
paltot=c("darkred","red","orange","white","lightblue","blue","darkblue"))
next3d()
localmeshdiff(neasur,sapsur,1,scale01 = TRUE,
paltot=c("darkred","red","orange","white","lightblue","blue","darkblue"))
## End(Not run)
```

Lset2D\_list

example dataset

# Description

List containing five 2D-landmark configurations acquired along five different anatomical views

#### Usage

```
data(Lset2D_list)
```

### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

Lset3D\_array

example dataset

#### **Description**

Array containing a cranial 3D-landmark configuration acquired on a Primate sample

# Usage

```
data(Lset3D_array)
```

# Author(s)

26 MLECScorrection

malleus\_bone

example dataset

# Description

3D mesh of a human malleus

# Usage

data(malleus\_bone)

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

MAs\_sets

example dataset

# Description

Landmark configurations of the manual alignments

#### Usage

data(MAs\_sets)

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

MLECScorrection

MLECScorrection Maximum Likelihood Estimation of the normalization factor to be applied to optimize the correlation between two landmark configurations to be combined by using twodviews and

# Description

MLECScorrection Maximum Likelihood Estimation of the normalization factor to be applied to optimize the correlation between two landmark configurations to be combined by using twodviews and

#### Usage

```
MLECScorrection(array1, array2, scale = TRUE, nPCs = 1:5)
```

noise.mesh 27

# Arguments

array1	array: first set of landmark configuration
array2	array: second set of landmark configuration
scale	logical: if FALSE the analysis is performed in the shape space, if TRUE the analysis is performed in the size and shape space (gpa without scaling)

nPCs numeric vector: specify which PC scores will be selected in the correlation test

#### Value

PCscores PCscores matrix of the combined dataset applying the normalization factor calculated by using the maximum likelihood estimation

PCs PCs matrix of the combined dataset applying the normalization factor calculated by using the maximum likelihood estimation

corr mean correlation between original and combined dataset

CSratios normalization factor calculated by using the maximum likelihood estimation

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Description**

This function adds noise to a mesh

# Usage

```
noise.mesh(mesh, noise = 0.025, seed = 123)
```

# **Arguments**

mesh	triangular mesh stored as object	et of class "mesh3d"

noise sd deviation to define vertex noise seed for random number generator

#### Value

```
mesh_n a 3D model of class "mesh3d" with noise
```

#### Author(s)

28 out.inn.mesh

#### **Examples**

```
#load mesh
library(compositions)
library(rgl)
data("SCP1.mesh")
mesh<-SCP1.mesh
#add noise
noised<-noise.mesh(mesh,noise=0.05)
#plot original and mesh with noise added
open3d()
shade3d(mesh,col=3)
shade3d(noised,col=2,add=TRUE)</pre>
```

out.inn.mesh

out.inn.mesh

#### **Description**

This function separates a 3D mesh subjected to the ext.int.mesh into two 3D models: the visible mesh and the not visible one

#### Usage

```
out.inn.mesh(scans, mesh, plot = TRUE)
```

# Arguments

scans an ext.int.mesh

mesh matrix mesh vertex (the same of the ext.int.mesh object)

plot logical: if TRUE the wireframe of the mesh with the visible vertices is plotted

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Examples**

```
## Not run:
#CA-LSE tool on Neanderthal tooth
#load a mesh
data(krd1_tooth)
library(rgl)
library(compositions)
ca_lse_krd1<-ext.int.mesh(mesh= krd1_tooth, views=50, param1=3, default=TRUE,
import_pov = NULL,expand=1, scale.factor=1,num.cores = NULL)
vis_inv_krd1<-out.inn.mesh(ca_lse_krd1, krd1_tooth, plot=TRUE)
inv_mesh<-vcgIsolated(vis_inv_krd1$invisible)</pre>
```

patches\_frm2amira 29

```
open3d()
shade3d(inv_mesh,col=2)
open3d()
shade3d(vis_inv_krd1$visible, col=3)
#CA-LSE tool on human malleus
#load a mesh
data(malleus_bone)
ca_lse_malleus<-ext.int.mesh(mesh= malleus_bone, views=50, param1=3,</pre>
default=TRUE, import_pov = NULL, expand=1, scale.factor=1)
vis_inv_malleus<-out.inn.mesh(ca_lse_malleus, malleus_bone, plot=TRUE)</pre>
inv_mesh<- vis_inv_malleus$invisible</pre>
inv_mesh<-ca_lse_malleus$invisible
#AST-3D tool
#load a mesh
data(human_skull)
data(endo_set)
ast3d_endocast<-ext.int.mesh(mesh=human_skull, views=50, param1=0.6, default=FALSE,
import_pov = TRUE,expand=1, matrix_pov =endo_set, scale.factor=1,num.cores = NULL)
vis_inv_endo<-out.inn.mesh(ast3d_endocast,human_skull,plot=TRUE)</pre>
vis_mesh<-vcgIsolated(vis_inv_endo$visible)</pre>
open3d()
shade3d(vis_mesh,col=3)
open3d()
shade3d(vis_inv_endo$invisible, col=2)
## End(Not run)
```

patches\_frm2amira

patches\_frm2amira

#### **Description**

This function converts the .frm files, from Evan Toolbox, stored in a folder into the format land-markAscii (semilandmark patches)

# Usage

```
patches_frm2amira(path_folder_frm, path_amira_folder)
```

### Arguments

```
path_folder_frm

character: path of the folder where the .frm files are stored

path_amira_folder

character: path folder to store the landmarkAscii configurations
```

#### Author(s)

30 permutangle

PCscoresCorr	PCscoresCorr Perform a correlation test between two matrices of PC-scores

# **Description**

PCscoresCorr Perform a correlation test between two matrices of PCscores

#### Usage

```
PCscoresCorr(matrix1, matrix2, nPCs = 1:5)
```

# **Arguments**

```
matrix1 matrix: first set of PC scores
matrix2 matrix: second set of PC scores
```

nPCs numeric vector: specify which PC scores will be selected in the correlation test

#### Value

```
corr the correlation values associated to each pair of PC scores p.values p-values associated to the correlation test
```

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# Description

Create palettes from an image

# Usage

```
permutangle(
  mat,
  var,
  group1,
  group2,
  scale = FALSE,
  iter = 100,
  cex1 = range01(var[group1] + 1),
  cex2 = range01(var[group2] + 1),
```

permutangle 31

```
cex3 = 0.7,
cex4 = 1.2,
labels = c("stgr1", "stgr2", "endgr1", "endgr2"),
pch1 = 19,
pch2 = 19,
pch3 = 19,
col1 = "red",
col2 = "blue"
)
```

# **Arguments**

mat	array: rgb array
var	numeric: desidered resize factor
group1	logical: if TRUE each color is counted once
group2	logical: if TRUE (color) variables are scaled
scale	numeric: desidered number of clusters (i.e., number of palettes)
iter	numeric: length of the color vector of each palette
cex1	numeric: desidered PLS axis
cex2	numeric: size of colored squares
cex3	numeric: size of color names
cex4	numeric: size of color names
labels	numeric: size of color names
pch1	numeric: size of color names
pch2	numeric: size of color names
pch3	numeric: size of color names
col1	numeric: size of color names
col2	numeric: size of color names

# Value

```
angle list: color palettes arranged in a list permangles list: color palettes arranged in a list angle list: color palettes arranged in a list iterangles list: color palettes arranged in a list p-value list: color palettes arranged in a list PCA_angle list: color palettes arranged in a list PCA_interangles list: color palettes arranged in a list PCA_p-value list: color palettes arranged in a list
```

# Author(s)

Antonio Profico

pov\_selecter

# **Examples**

```
## Not run:
require(shapes)
require(Morpho)
data("gorf.dat")
data("gorm.dat")
Array<-bindArr(gorf.dat,gorm.dat,along=3)</pre>
CS<-apply(Array,3,cSize)</pre>
Sex<-c(rep("F",dim(gorf.dat)[3]),rep("M",dim(gorm.dat)[3]))</pre>
#Shape and size space
AllTrajFB<-permutangle(procSym(Array,scale=FALSE,CSinit = FALSE)$PCscores,
var=CS, group1=which(Sex=="F"), group2=which(Sex=="M"), scale=FALSE, iter=50)
hist(AllTrajFB$iterangles,breaks = 100,xlim=c(0,90))
abline(v=AllTrajFB$angle,lwd=2,col="red")
hist(AllTrajFB$PCA_iterangles,breaks = 100,xlim=c(0,90))
abline(v=AllTrajFB$PCA_angle,lwd=2,col="red")
#Shape space
AllTrajFB<-permutangle(procSym(Array)$PCscores,
var=CS,group1=which(Sex=="F"),group2=which(Sex=="M"),scale=FALSE,iter=50)
hist(AllTrajFB$iterangles,breaks = 100,xlim=c(0,90))
abline(v=AllTrajFB$angle,lwd=2,col="red")
hist(AllTrajFB$PCA_iterangles,breaks = 100,xlim=c(0,90))
abline(v=AllTrajFB$PCA_angle,lwd=2,col="red")
## End(Not run)
```

pov\_selecter

pov\_selecter

# **Description**

Internal function to define the points of view

#### Usage

```
pov_selecter(mesh, grid, start.points = 250, method = "ast3d")
```

# **Arguments**

mesh object of class mesh3d grid matrix: a 3D grid

start.points numeric: number of center to be found

method character: select "a" or "c" for respectively AST-3D and CA-LSE method

# Value

selection numeric: positioning vector of the selected points of the grid

primendoR 33

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

primendoR

example dataset

#### **Description**

right brain hemisphere of 19 primate species

# Usage

data(primendoR)

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

read.amira.dir

read.amira.dir

# Description

This function reads and stores in an array the coordinated allocated in a folder in separate files (format landmarkAscii)

#### Usage

```
read.amira.dir(path.dir, nland)
```

# Arguments

path.dir

character: path of the folder

nland

numeric: number of landmark sampled in Amira

#### Value

array.set numeric: a kx3xn array with landmark coordinates

# Author(s)

34 read.path.amira

read.amira.set

read.amira.set

# **Description**

This function converts a landmarkAscii file set in a kx3x1 array

#### Usage

```
read.amira.set(name.file, nland)
```

# Arguments

name.file character: path of a landmarkAscii file

nland numeric: number of landmark sampled in Amira, if is set on "auto" it will be

automatically recognized

#### Value

array.set numeric: a kx3x1 array with landmark coordinates

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

read.path.amira

read.path.amira

### **Description**

This function extracts and orders the coordinate matrix from a surface path file from Amira

# Usage

```
read.path.amira(path.name)
```

#### **Arguments**

path.name

character: path of surface path .ascii extension file

#### Value

data numeric: a kxd matrix with xyz coordinates

# Author(s)

repmat 35

# Description

This function repeats copies of a matrix

# Usage

```
repmat(X, m, n)
```

#### **Arguments**

X numeric: a matrix

m numeric: number of times to repeat the X matrix in row and column dimension

n numeric: repetition factor for each dimesion

#### Value

matrice: repeated matrix

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

RMs_sets	example dataset	

# **Description**

Array containing the landmark coordinates of the reference sample for Digital Alignment Tool example

# Usage

```
data(RMs_sets)
```

# Author(s)

SM\_set

SCP1.mesh

example dataset

# Description

Mesh of the Saccopastore 1 Neanderthal skull

# Usage

```
data(SCP1.mesh)
```

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

sinus\_set

example dataset

# **Description**

POVs sampled inside the maxillary sinus cavity

#### Usage

```
data(sinus_set)
```

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

SM\_set

example dataset

# Description

Landmark configuration associated to the starting model

# Usage

```
data(SM_set)
```

#### Author(s)

spherical.flipping 37

spherical.flipping spherical.flipping

#### **Description**

Internal spherical flippping function

# Usage

```
spherical.flipping(C, mesh, param1, param2)
```

# **Arguments**

C numeric: coordinates of the point of view

mesh object of class mesh3d

param1 numeric: first parameter for spherical flipping (usually ranged from 0.1 to 3,

try!)

param2 numeric second paramter for spherical flipping (don't change it!)

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

#### References

Profico, A., Schlager, S., Valoriani, V., Buzi, C., Melchionna, M., Veneziano, A., ... & Manzi, G. (2018). Reproducing the internal and external anatomy of fossil bones: Two new automatic digital tools. American Journal of Physical Anthropology, 166(4), 979-986.#' @export

Katz, S., Tal, A., & Basri, R. (2007). Direct visibility of point sets. In ACM SIGGRAPH 2007 papers (pp. 24-es).

trasf.mesh trasf.mesh

#### **Description**

This function centers a mesh on the barycenter coordinates

# Usage

```
trasf.mesh(mesh, barycenter)
```

#### **Arguments**

mesh a 3D mesh of class "mesh3d" barycenter numeric: coordinates of the center 38 twodvarshape

#### Value

mesh a 3D mesh of class "mesh3d"

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

twodvarshape twodvarshape Calculates the shape variation associated to a value of PC scores associated to a specific combined landmark configuration or view	,
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# **Description**

twodvarshape Calculates the shape variation associated to a value of PC scores associated to a specific combined landmark configuration or view

# Usage

```
twodvarshape(twodviews_ob, scores, PC, view)
```

# **Arguments**

twodviews\_ob object from twodviews()

scores numeric: the values of the PC scores for which the visualization is called

PC PC chosen

view numeric: which landmark configuration will be used to build the shape variation

#### Value

mat matrix of coordinates associated to the called shape variation

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# References

Profico, A., Piras, P., Buzi, C., Del Bove, A., Melchionna, M., Senczuk, G., ... & Manzi, G. (2019). Seeing the wood through the trees. Combining shape information from different landmark configurations. Hystrix, 157-165.

twodviews 39

#### **Examples**

```
library(Arothron)
#load the 2D primate dataset
data("Lset2D_list")
#combine the 2D datasets and PCA
combin2D<-twodviews(Lset2D_list,scale=TRUE,vector=c(1:5))
#calculate the shape variation associated to the negative extreme value of PC1
min_PC1<-twodvarshape(combin2D,min(combin2D$PCscores[,1]),1,5)
plot(min_PC1,asp=1)
#calculate the shape variation associated to the positive extreme value of PC1
max_PC1<-twodvarshape(combin2D,max(combin2D$PCscores[,1]),1,5)
plot(max_PC1,asp=1)</pre>
```

twodviews Combine and calculate the PCscores matrix from a list of different landmark configurations to be combined

# Description

twodviews Combine and calculate the PCscores matrix from a list of different landmark configurations to be combined

# Usage

```
twodviews(twodlist, scale = TRUE, vector = NULL)
```

#### **Arguments**

twodlist a list containing the landmark configurations of each anatomical view stored as

separated lists

scale logical: TRUE for shape-space, FALSE for form-space vector numeric vector: defines which views are to be used

#### Value

PCscores PC scores

PCs Pricipal Components (eigenvector matrix)

Variance table of the explained variance by the PCs

size vector containing the Centroid Size of each configuration

mshapes a list containing the mean shape of each landmark configuration

dims number of landmarks of each configuration

dimm dimension (2D or 3D) of each combined landmark configuration

twodlist the list used as input

40 volendo

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

#### References

Profico, A., Piras, P., Buzi, C., Del Bove, A., Melchionna, M., Senczuk, G., ... & Manzi, G. (2019). Seeing the wood through the trees. Combining shape information from different landmark configurations. Hystrix, 157-165.

# **Examples**

```
library(Morpho)
#load the 2D primate dataset
data("Lset2D_list")
length(Lset2D_list)
#combine the 2D datasets and PCA
combin2D<-twodviews(Lset2D_list,scale=TRUE,vector=c(1:5))</pre>
combin2D$size
#plot of the first two Principal Components
plot(combin2D$PCscores)
text(combin2D$PCscores,labels=rownames(combin2D$PCscores))
#load the 3D primate dataset
data("Lset3D_array")
#GPA and PCA
GPA_3D<-procSym(Lset3D_array)</pre>
#plot of the first two Principal Components
plot(GPA_3D$PCscores)
text(GPA_3D$PCscores,labels=rownames(GPA_3D$PCscores))
```

volendo

volendo

#### **Description**

Calculate the volume of a mesh by using a voxel-based method

#### Usage

```
volendo(mesh, alpha_vol = 100, ncells = 1e+05)
```

# **Arguments**

mesh object of class mesh3d

alpha\_vol numeric: alpha shape for construction external concave hull numeric: approximative number of cell for 3D grid construction

#### Value

vol numeric: volume of the mesh expressed in cc

yoda\_set 41

#### Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

# **Examples**

```
## Not run:
#load the human skull
library(rgl)
data(human_skull)
sapendo<-endomaker(human_skull,param1_endo = 1.0,vol=FALSE, num.cores=NULL)
volsap<-volendo(sapendo$endocast)
## End(Not run)</pre>
```

yoda\_set

example dataset

# **Description**

Landmark set on Yoda

# Usage

```
data(yoda_set)
```

# Author(s)

Antonio Profico, Costantino Buzi, Marina Melchionna, Paolo Piras, Pasquale Raia, Alessio Veneziano

yoda\_sur

example dataset

# Description

Mesh of Yoda

# Usage

```
data(yoda_sur)
```

# Author(s)

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