Package 'microsynth'

June 30, 2023
Title Synthetic Control Methods with Micro- And Meso-Level Data
Version 2.0.44
Description A generalization of the 'Synth' package that is designed for data at a more granular level (e.g., micro-level). Provides functions to construct weights (including propensity score-type weights) and run analyses for synthetic control methods with micro- and meso-level data; see Robbins, Saunders, and Kilmer (2017) <doi:10.1080 01621459.2016.1213634=""> and Robbins and Davenport (2021) <doi:10.18637 jss.v097.i02="">.</doi:10.18637></doi:10.1080>
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microsynth

Synthetic control methods for disaggregated, micro-level data.

Description

Implements the synthetic control method for micro-level data as outlined in Robbins, Saunders, and Kilmer (2017). microsynth is designed for use in assessment of the effect of an intervention using longitudinal data. However, it may also be used to calculate propensity score-type weights in cross-sectional data. microsynth is a generalization of Synth (see Abadie and Gardeazabal (2003) and Abadie, Diamond, Hainmueller (2010, 2011, 2014)) that is designed for data at a more granular level (e.g., micro-level). For more details see the help vignette: vignette('microsynth', package = 'microsynth').

microsynth develops a synthetic control group by searching for weights that exactly match a treatment group to a synthetic control group across a number of variables while also minimizing the discrepancy between the synthetic control group and the treatment group across a set second set of variables. microsynth works in two primary steps: 1) calculation of weights and 2) calculation of results. Time series plots of treatment vs. synthetic control for pertinent outcomes may be performed using the function plot.microsynth().

The time range over which data are observed is segmented into pre- and post-intervention periods. Treatment is matched to synthetic control across the pre-intervention period, and the effect of the intervention is assessed across the post-intervention (or evaluation) period. The input end.pre (which gives the last pre-intervention time period) is used to delineate between pre- and post-intervention. Note that if the intervention is not believed to have an instantaneous effect, end.pre should indicate the time of the intervention.

Variables are categorized as outcomes (which are time-variant) and covariates (which are time-invariant). Using the respective inputs match.covar and match.out, the user specifies across which covariates and outcomes (and which pre-intervention time points of the outcomes) treatment is to be exactly matched to synthetic control. The inputs match.covar.min and match.out.min are similar but instead specify variables across which treatment is to be matched to synthetic control as closely as possible. If there are no variables specified in match.covar.min and match.out.min, the function calibrate() from the survey package is used to calculate weights. Otherwise, the function LowRankQP() from the package of the same name is used, if it is available on the user's machine (it is now in the CRAN archive, so would need to be installed by other means). If the LowRankQP package is unavailable, it will use ipop() from the kernlab package. In the event that the model specified by match.covar and match.out is not feasible (i.e., weights do not exist that exactly match treatment and synthetic control subject to the given constraints), a less restrictive backup model is used.

microsynth has the capability to perform statistical inference using Taylor series linearization, a jackknife and permutation methods. Several sets of weights are calculated. A set of main weights is calculated that is used to determine a point estimate of the intervention effect. The main weights can also be used to perform inferences on the point estimator via Taylor series linearization. If a jackknife is to be used, one set of weights is calculated for each jackknife replication group, and if permutation methods are to be used, one set of weights is calculated for each permutation group. If treatment and synthetic control are not easily matched based upon the model outlined

in match.covar and match.out (i.e., an exact solution is infeasible or nearly infeasible), it is recommended that the jackknife not be used for inference.

The software provides the user the option to output overall findings in an Excel file. For each outcome variable, the results list the estimated treatment effect, as well as confidence intervals of the effect and p-values of a hypothesis test that assesses whether the effect is zero. Such results are produced as needed for each of the three methods of statistical inference noted above. microsynth can also apply an omnibus test that examines the presence of a treatment effect jointly across several outcomes.

Usage

```
microsynth(
  data,
  idvar,
  intvar,
  timevar = NULL,
  start.pre = NULL,
  end.pre = NULL,
  end.post = NULL,
  match.out = TRUE,
 match.covar = TRUE,
  match.out.min = NULL,
  match.covar.min = NULL,
  result.var = TRUE,
  omnibus.var = result.var,
  period = 1,
  scale.var = "Intercept",
  confidence = 0.9,
  test = "twosided",
  perm = 0,
  jack = 0,
  use.survey = TRUE,
  cut.mse = Inf,
  check.feas = FALSE,
  use.backup = FALSE,
  w = NULL,
  max.mse = 0.01,
  maxit = 250,
  cal.epsilon = 1e-04,
  calfun = "linear",
  bounds = c(0, Inf),
  result.file = NULL,
  printFlag = TRUE,
  n.cores = TRUE,
  ret.stats = FALSE
)
```

Arguments

data

A data frame. If longitudinal, the data must be entered in tall format (e.g., at the case/time-level with one row for each time period for each case). Missingness is not allowed. All individuals must have non-NA values of all variables at all time points.

idvar

A character string that gives the variable in data that identifies multiple records from the same case.

intvar

A character string that gives the variable in data that corresponds to the intervention variable. The intervention variable indicates which cases (and times) have received the intervention. The variable should be binary, with a 1 indicating treated and 0 indicating untreated. If end.pre is specified, a case is considered treated if there is 1 or more non-zero entries in the column indicated by intvar for that case (at any time point). If end.pre is not specified, an attempt will be made to use intvar to determine which time periods will be considered post-intervention (i.e., the times contained in the evaluation period). In this case, the evaluation period is considered to begin at the time of the first non-zero entry in intvar).

timevar

A character string that gives the variable in data that differentiates multiple records from the same case. Can be set to NULL only when used with cross-sectional data (i.e., with one observation per entry in idvar).

start.pre

An integer indicating the time point that corresponds to the beginning of the pre-intervention period used for matching. When start.pre = NULL (default), it is reset to the minimum time appearing in the column given by timevar. If match.out (and match.out.min) are given in list format, start.pre is ignored except for plotting.

end.pre

An integer that gives the final time point of the pre-intervention period. That is, end.pre is the last time at which treatment and synthetic control will be matched to one another. All time points following end.pre are considered to be post-intervention and the behavior of outcomes will be compared between the treatment and synthetic control groups across those time periods. Setting end.pre = NULL will begin the post-intervention period at the time that corresponds to the first non-zero entry in the column indicated by intvar.

end.post

An integer that gives the maximum post-intervention time that is taken into when compiling results. That is, the treatment and synthetic control groups are compared across the outcomes listed in result.var from the first time following the intervention up to end.post. Can be a vector (ordered, increasing) giving multiple values of end.post. In this case, the results will be compiled for each entry in end.post. When end.post = NULL (the default), it is reset to the maximum time that appears in the column given by timevar.

match.out

Either A) logical, B) a vector of variable names that indicates across which timevarying variables treatment is to be exactly matched to synthetic control preintervention, or C) a list consisting of variable names and timespans over which variables should be aggregated before matching. Note that outcome variables and time-varying covariates should be included in match.out.

If match.out = TRUE (the default), it is set equal to result.var; if match.out = NULL or match.out = FALSE, no outcome variables are factored into the calculation of weights. If match.out is passed a vector of variable names, then

weights are calculated to match treatment and synthetic control for the value of each variable that appears in match.out at each time point from start.pre to end.pre. Otherwise, to allow more flexibility, match.out may also be a list that gives an outcome-based model outlining more specific constraints that are to be exactly satisfied within calibration weighting. In this case, each entry of match.out is a vector of integers, and the names of entries of match.out are the outcome variables to which the vectors correspond. Each element of the vectors gives a number of time points that are to be aggregated for the respective outcome, with the first element indicating time points immediately prior the beginning of the post-intervention period. The sum of the elements in each vector should not exceed the number of pre-intervention time periods in the data.

The following examples show the proper formatting of match.out as a list. Assume that there are two outcomes, Y1 and Y2 (across which treatment is to be matched to synthetic control), and end.pre = 10 (i.e., the post-intervention period begins at time 11). Let match.out = list('Y1' = c(1, 3, 3), 'Y2'=c(2,5,1)). According to this specification, treatment is to be matched to synthetic control across: a) The value of Y1 at time 10; b) the sum of Y1 across times 7, 8 and 9; c) the sum of Y1 across times 4, 5 and 6; e) The sum of Y2 across times time 9 and 10; e) the sum of Y2 across times 4, 5, 6, 7, and 8; f) the value of Y2 at time 3. Likewise, if match.out = list('Y1' = 10, 'Y2' = rep(1,10)), Y1 is matched to synthetic control the entire aggregated pre-intervention time range, and Y2 is matched at each pre-intervention time point individually.

match.covar

Either a logical or a vector of variable names that indicates which time invariant covariates are to be used for weighting. Weights are calculated so that treatment and synthetic control exactly match across these variables. If match.covar = TRUE, it is set equal to a vector of variable names corresponding to the time invariant variables that appear in data. If match.covar = FALSE, it is set to NULL (in which case no time-invariant variables are used for matching when calculating weights).

match.out.min

A vector or list of the same format as match.out that is used to specify additional time-varying variables to match on, but which need not be matched exactly. Weights are calculated so the distance is minimized between treatment and synthetic control across these variables. If match.out.min = NULL, no outcome-based constraints beyond those indicated by match.out are imposed (i.e., all outcome variables will be matched on exactly).

match.covar.min

A vector of variable names that indicates supplemental time invariant variables that are to be used for weighting, for which exact matches are not required. Weights are calculated so the distance is minimized between treatment and synthetic control across these variables.

result.var

A vector of variable names giving the outcome variables for which results will be reported. Time-varying covariates should be excluded from result.var. If result.var = TRUE (the default), result.var is set as being equal to all time-varying variables that appear in data. If result.var = NULL or result.var = FALSE, results are not tabulated.

omnibus.var

A vector of variable names that indicates the outcome variables that are to be

used within the calculation of the omnibus statistic. Can also be a logical indicator. When omnibus.var = TRUE, it is reset as being equal to result.var. When omnibus.var = NULL or omnibus = FALSE, no omnibus statistic is calculated. omnibus.var should not contain elements not in result.var.

period

An integer that gives the granularity of the data that will be used for plotting and compiling results. If match.out and match.out.min are provided a vector of variable names, it will also affect the calculation of weights used for matching. In this case, matching of treatment and synthetic control is performed at a temporal granularity defined by period. For instance, if monthly data are provided and period = 3, data are aggregated to quarters for plots and results (and weighting unless otherwise specified). If match.out and match.out.min are provided a list, period only affects plots and how results are displayed.

Note that plotting is performed with plot.microsynth(); however, a microsynth object is required as input for that function and period should be specified in the creation of that object.

scale.var

A variable name. When comparing the treatment group to all cases, the latter is scaled to the size of the former with respect to the variable indicated by scale.var. Defaults to the number of units receiving treatment (i.e., the intercept).

confidence

The level of confidence for confidence intervals.

test

The type of hypothesis test (one-sided lower, one-sided upper, or two-sided) that is used when calculating p-values. Entries of 'lower', 'upper', and 'twosided' are recognized.

perm

An integer giving the number of permutation groups that are used. If perm = 0, no permutation groups are generated, permutation weights are not calculated, and permutations do not factor into the reported results. perm is set to the number of possible permutation groups if the former exceeds the latter.

jack

An integer giving the number of replication groups that are used for the jack-knife. jack can also be a logical indicator. If jack = 0 or jack = FALSE, no jackknife replication groups are generated, jackknife weights are not calculated, and the jackknife is not considered when reporting results. If jack = TRUE, it is reset to being equal to the minimum between the number of total cases in the treatment group and the total number of cases in the control group. jack is also reset to that minimum if it, as entered, exceeds that minimum.

use.survey

If use.survey = TRUE, Taylor series linearization is applied to the estimated treatment effect within each permutation group. Setting use.survey = TRUE makes for better inference but increases computation time substantially. Confidence intervals for permutation groups are calculated only when use.survey = TRUE.

cut.mse

The maximum error (given as mean-squared error) permissible for permutation groups. Permutation groups with a larger than permissible error are dropped when calculating results. The mean-squared error is only calculated over constraints that are to be exactly satisfied.

check.feas

A logical indicator of whether or not the feasibility of the model specified by match.out is evaluated prior to calculation of weights. If check.feas = TRUE, feasibility is assessed. If match.out is found to not specify a feasible model,

a less restrictive feasible backup model will be applied to calculate the main weights and for jackknife and permutation methods. use.backup A logical variable that, when true, indicates whether a backup model should be used whenever the model specified by match. out yields unsatisfactory weights. Weights are deemed to be unsatisfactory if they do not sufficiently satisfy the constraints imposed by match.out and match.covar. Different backup models may be used for each of the main, jackknife or permutation weights as needed. A microsynth object or a list of the form as returned by a prior application w of microsynth. If w = NULL, weights are calculated from scratch. Entering a non-NULL value affords the user the ability to use previously calculated weights. max.mse The maximum error (given as mean-squared error) permissible for constraints that are to be exactly satisfied. If max.mse is not satisfied by these constraints, and either check. feas = TRUE or use. backup = TRUE, then back-up models are used. The maximum number of iterations used within the calibration routine (calibrate() maxit from the survey package) for calculating weights. The tolerance used within the calibration routine (calibrate() from the survey cal.epsilon package) for calculating weights. calfun The calibration function used within the calibration routine (calibrate() from the survey package) for calculating weights. bounds Bounds for calibration weighting (fed into the calibrate() from the survey package). result.file A character string giving the name of a file that will be created in the home directory containing results. If result.file = NULL (the default), no file is created. If end.post has length 1, a .csv file is created. If end.post has length greater than one, a formatted .xlsx file is created with one tab for each element of end.post. If result.file has a .xlsx (or .xls) extension (e.g., the last five characters of result.file are '.xlsx'), an .xlsx file is created regardless of the length of end.post. printFlag If TRUE, microsynth will print history on console. Use printFlag = FALSE for silent computation. n.cores The number of CPU cores to use for parallelization. If n.cores is not specified by the user, it is guessed using the detectCores function in the parallel package. If TRUE (the default), it is set as detectCores(). If NULL, it is set as detectCores() - 1. If FALSE, it is set as 1, in which case parallelization is not invoked. Note that the documentation for detectCores makes clear that it is not failsafe and could return a spurious number of available cores.

Details

ret.stats

delta.out.

microsynth requires specification of the following inputs: data, idvar, intvar. data is a longitudinal data frame; idvar and intvar are character strings that specific pertinent columns of data. In longitudinal data, timevar should be specified. Furthermore, specification of match.out and match.covar is recommended.

if set to TRUE, returns four additional elements: stats, stats1, stats2 and

microsynth can also be used to calculate propensity score-type weights in cross sectional data (in which case timevar does not need to be specified) as proposed by Hainmueller (2012).

microsynth calculates weights using survey::calibrate() from the survey package in circumstances where a feasible solution exists for all constraints, whereas LowRankQP::LowRankQP() is used to assess feasibility and to calculate weights in the event that a feasible solution to all constraints does not exist. The LowRankQP routine is memory-intensive and can run quite slowly in data that have a large number of cases. To prevent LowRankQP from being used, set match.out.min = NULL, match.covar.min= NULL, check.feas = FALSE, and use.backup = FALSE.

Value

microsynth returns a list with up to five elements: a) w, b) Results, c) svyglm.stats, and d) Plot.Stats, and e) info.

w is a list with six elements: a) Weights, b) Intervention, c) MSE, d) Model, e) Summary, and f) keep.groups. Assume there are C total sets of weights calculated, where C = 1 + jack + perm, and there are N total cases across the treatment and control groups. w\$Weights is an N x C matrix, where each column provides a set of weights. w\$Intervention is an N x C matrix made of logical indicators that indicate whether or not the case in the respective row is considered treated (at any point in time) for the respective column. Entries of NA are to be dropped for the respective jackknife replication group (NAs only appear in jackknife weights). w\$MSE is a 6 x C matrix that give the MSEs for each set of weights. MSEs are listed for the primary and secondary constraints for the first, second, and third models. Note that the primary constraints differ for each model (see Robbins and Davenport, 2021). w\$Mode1 is a length-C vector that indicates whether backup models were used in the calculation of each set of weights. w\$keep.groups is a logical vector indicating which groups are to be used in analysis (groups that are not used have pre-intervention MSE greater than cut.mse. w\$Summary is a three-column matrix that (for treatment, synthetic control, and the full dataset), shows aggregate values of the variables across which treatment and synthetic control are matched. The summary, which is tabulated only for the primary weights, is also printed by microsynth while weights are being calculated.

Further, Results is a list where each element gives the final results for each value of end.post. Each element of Results is itself a matrix with each row corresponding to an outcome variable (and a row for the omnibus test, if used) and each column denotes estimates of the intervention effects and p-values, upper, and lower bounds of confidence intervals as found using Taylor series linearization (Linear), jackknife (jack), and permutation (perm) methods where needed.

In addition, svyglm.stats is a list where each element is a matrix that includes the output from the regression models run using the svyglm() function to estimate the treatment effect. The list has one element for each value of end.post, and the matrices each have one row per variable in result.var.

Next, Plot.Stats contains the data that are displayed in the plots which may be generated using plot.microsynth(). Plot.Stats is a list with four elements (Treatment, Control, All, Difference). The first three elements are matrices with one row per outcome variable and one column per time point. The last element (which gives the treatment minus control values) is an array that contains data for each permutation group in addition to the true treatment area. Specifically, Plot.Stats\$Difference[,,1] contains the time series of treatment minus control for the true intervention group; Plot.Stats\$Difference[,,i+1] contains the time series of treatment minus control for the i^th permutation group.

Next, info documents some input parameters for display by print(). A summary of weighted matching variables and of results can be viewed using summary

Lastly, if ret.stats is set to TRUE, four additional elements are returned: stats, stats1, stats2 and delta.out. stats contains elements with the basic statistics that are the same as the main microsynth output: outcomes in treatment, control and percentage change. stats1 are the estimates of svyglm() adjusted by their standard errors. stats2 is the percent change in the observed value from each outcome from the hypothetical outcome absent intervention. delta.out is a Taylor series linearization used to approximate the variance of the estimator.

References

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Robbins MW, Saunders J, Kilmer B (2017). A framework for synthetic control methods with high-dimensional, micro-level data: Evaluating a neighborhood-specific crime intervention,? *Journal of the American Statistical Association*, 112(517), 109-126.

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Examples

```
start.pre=1, end.pre=12, end.post=16,
                  match.out=match.out, match.covar=cov.var,
                  result.var=match.out, omnibus.var=match.out,
                  test='lower',
                  n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea1)
plot_microsynth(sea1)
## Not run:
# Repeat matching and estimation, with permutations and jackknife
# Set permutations and jack-knife to very few groups (2) for
# quick demonstration only.
# runtime: ~30 min
sea2 <- microsynth(seattledmi,</pre>
                     idvar='ID', timevar='time', intvar='Intervention',
                     start.pre=1, end.pre=12, end.post=c(14, 16),
                     match.out=match.out, match.covar=cov.var,
                     result.var=match.out, omnibus.var=match.out,
                     test='lower',
                     perm=250, jack=TRUE,
                     result.file=file.path(tempdir(), 'ExResults2.xlsx'),
                     n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea2)
plot_microsynth(sea2)
# Specify additional outcome variables for matching, which makes
# matching harder.
match.out <- c('i_robbery','i_aggassau','i_burglary','i_larceny',</pre>
       'i_felony','i_misdemea','i_drugsale','i_drugposs','any_crime')
# Perform matching, setting check.feas = T and use.backup = T
# to ensure model feasibility
# runtime: ~40 minutes
sea3 <- microsynth(seattledmi,</pre>
                   idvar='ID', timevar='time', intvar='Intervention',
                   end.pre=12,
                   match.out=match.out, match.covar=cov.var,
                   result.var=match.out, perm=250, jack=0,
                   test='lower', check.feas=TRUE, use.backup = TRUE,
                   result.file=file.path(tempdir(), 'ExResults3.xlsx'),
                   n.cores = min(parallel::detectCores(), 2))
# Aggregate outcome variables before matching, to boost model feasibility
match.out <- list( 'i_robbery'=rep(2, 6), 'i_aggassau'=rep(2, 6),</pre>
         'i_burglary'=rep(1, 12), 'i_larceny'=rep(1, 12),
         'i_felony'=rep(2, 6), 'i_misdemea'=rep(2, 6),
         'i_drugsale'=rep(4, 3), 'i_drugposs'=rep(4, 3),
```

```
'any_crime'=rep(1, 12))
# After aggregation, use.backup and cheack.feas no longer needed
# runtime: ~40 minutes
sea4 <- microsynth(seattledmi, idvar='ID', timevar='time',</pre>
         intvar='Intervention', match.out=match.out, match.covar=cov.var,
         start.pre=1, end.pre=12, end.post=16,
         result.var=names(match.out), omnibus.var=names(match.out),
         perm=250, jack = TRUE, test='lower',
         result.file=file.path(tempdir(), 'ExResults4.xlsx'),
         n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea4)
plot_microsynth(sea4)
# Generate weights only (for four variables)
match.out <- c('i_felony', 'i_misdemea', 'i_drugs', 'any_crime')</pre>
# runtime: ~ 20 minutes
sea5 <- microsynth(seattledmi, idvar='ID', timevar='time',</pre>
         intvar='Intervention', match.out=match.out, match.covar=cov.var,
         start.pre=1, end.pre=12, end.post=16,
         result.var=FALSE, perm=250, jack=TRUE,
         n.cores = min(parallel::detectCores(), 2))
# View weights
summary(sea5)
# Generate results only
sea6 <- microsynth(seattledmi, idvar='ID', timevar='time',</pre>
          intvar='Intervention',
          start.pre=1, end.pre=12, end.post=c(14, 16),
          result.var=match.out, test='lower',
          w=sea5, result.file=file.path(tempdir(), 'ExResults6.xlsx'),
          n.cores = min(parallel::detectCores(), 2))
# View results (including previously-found weights)
summary(sea6)
# Generate plots only
plot_microsynth(sea6, plot.var=match.out[1:2])
# Apply microsynth in the traditional setting of Synth
# Create macro-level (small n) data, with 1 treatment unit
set.seed(86879)
ids.t <- names(table(seattledmi$ID[seattledmi$Intervention==1]))</pre>
ids.c <- setdiff(names(table(seattledmi$ID)), ids.t)</pre>
ids.synth <- c(base::sample(ids.t, 1), base::sample(ids.c, 100))</pre>
seattledmi.one <- seattledmi[is.element(seattledmi$ID,</pre>
           as.numeric(ids.synth)), ]
```

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```
# Apply microsynth to the new macro-level data
# runtime: < 5 minutes</pre>
sea8 <- microsynth(seattledmi.one, idvar='ID', timevar='time',</pre>
           intvar='Intervention',
           start.pre=1, end.pre=12, end.post=16,
           match.out=match.out[4],
           match.covar=cov.var, result.var=match.out[4],
           test='lower', perm=250, jack=FALSE,
           check.feas=TRUE, use.backup=TRUE,
           n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea8)
plot_microsynth(sea8)
# Use microsynth to calculate propensity score-type weights
# Prepare cross-sectional data at time of intervention
seattledmi.cross <- seattledmi[seattledmi$time==16, colnames(seattledmi)!='time']</pre>
# Apply microsynth to find propensity score-type weights
# runtime: ~5 minutes
sea9 <- microsynth(seattledmi.cross, idvar='ID', intvar='Intervention',</pre>
             match.out=FALSE, match.covar=cov.var, result.var=match.out,
             test='lower', perm=250, jack=TRUE,
             n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea9)
## End(Not run)
```

plot_microsynth

Plotting for microsynth objects.

Description

Using a microsynth object as an input, this function gives time series plots of selected outcomes.

Usage

```
plot_microsynth(
   ms,
   plot.var = NULL,
   start.pre = NULL,
   end.pre = NULL,
   end.post = NULL,
   file = NULL,
   sep = TRUE,
```

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```
plot.first = NULL,
  legend.spot = "bottomleft",
  height = NULL,
  width = NULL,
  at = NULL,
  labels = NULL,
  all = "cases",
  main.tc = NULL,
  main.diff = NULL,
  xlab.tc = NULL,
  ylab.diff = NULL,
  ylab.diff = NULL
```

Arguments

ms	A microsynth object
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plot.var A vector of variable names giving the outcome variables that are shown in plots.

If plot.var = NULL, all outcome variables that are included in ms are plotted. Only variables contained in the input result.var as used in the creation of ms

can be plotted using plot().

start.pre An integer indicating the time point that corresponds to the earliest time period

that will be plotted. When start.pre = NULL, it is reset to the minimum time

appearing in ms.

end.pre An integer that gives the final time point of the pre-intervention period. That

is, end.pre is the last time at which treatment and synthetic control will were matched to one another. All time points following end.pre are considered to be post-intervention and the behavior of outcomes will be compared between the treatment and synthetic control groups across those time periods. If end.pre = NULL the end of the pre-intervention period will be determined from the object

ms.

end.post An integer that gives final time point that will be plotted. When end.post =

NULL (the default), it is reset to the maximum time that appears in ms.

file A character string giving the name of file that will be created in the home direc-

tory containing plots. The name should have a .pdf extension.

sep If sep = TRUE, separate plots will be generated for each outcome. Applicable

only if plots are saved to file (plot.file is non-NULL). To change display of

plots produced as output, use par.

plot. first The number of permutation groups to plot.

legend. spot The location of the legend in the plots.

height The height of the graphics region (in inches) when a pdf is created.

width The width of the graphics region (in inches) when a pdf is created.

at A vector that gives the location of user-specified x-axis labels. at should be a

(numeric) subset of the named time points contained in ms (e.g., colnames (ms\$Plot.Stats\$Treatment)

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labels	A vector of the same length as at that gives the names of the labels that will be marked at the times indicated by at in the plots.
all	A scalar character string giving the unit name for cases. If NULL, a third curve showing the overall outcome levels is not plotted.
main.tc	A scalar (or a vector of the same length as plot.var) character string giving the title to be used for the first plots (that show treatment and control). Defaults to plot.var.
main.diff	A scalar (or a vector of the same length as plot.var) character string giving the title to be used for the second plots (that show differences between treatment and control). Defaults to plot.var.
xlab.tc	A scalar (or a vector of the same length as plot.var) character string giving the x-axis labels to be used for the first plots (that show treatment and control). Defaults to ''.
xlab.diff	A scalar (or a vector of the same length as plot.var) character string giving the x-axis labels to be used for the second plots (that show differences between treatment and control). Defaults to ''.
ylab.tc	A scalar (or a vector of the same length as plot.var) character string giving the y-axis labels to be used for the first plots (that show treatment and control). Defaults to plot.var.
ylab.diff	A scalar (or a vector of the same length as plot.var) character string giving the y-axis labels to be used for the second plots (that show differences between treatment and control). Defaults to 'Treatment - Control'.

Details

Plots are given over both pre- and intervention time periods and shown in terms of raw outcome values or treatment/control differences. Time series of permutation groups may be overlaid to help illustrate statistical uncertainty.

Only required input is a parameter ms which is a microsynth object.

Value

No return value, called for side effects (i.e., to produce plots of outcome values and treatment/control differences, with the option to write to file).

Examples

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```
# Perform matching and estimation, without permutations or jackknife
# runtime: <1 min</pre>
sea1 <- microsynth(seattledmi,</pre>
                  idvar='ID', timevar='time', intvar='Intervention',
                  start.pre=1, end.pre=12, end.post=16,
                  match.out=match.out, match.covar=cov.var,
                  result.var=match.out, omnibus.var=match.out,
                  test='lower',
                  n.cores = min(parallel::detectCores(), 2))
# Plot with default settings in the GUI.
plot_microsynth(sea1)
# Make plots, display, and save to a single file (plots.pdf).
plot_microsynth(sea1, file = file.path(tempdir(), 'plots.pdf'), sep = FALSE)
# Make plots for only one outcome, display, and save to a single file.
plot_microsynth(sea1, plot.var = 'any_crime',
     file = file.path(tempdir(), 'plots.pdf'), sep = FALSE)
```

print.microsynth

Displaying microsynth Fits and Results

Description

Print method for class 'microsynth'.

Usage

```
## S3 method for class 'microsynth'
print(x, ...)
```

Arguments

x A microsynth object produced by microsynth()

... further arguments passed to or from other methods.

Value

The functions print.microsynth and summary.microsynth display information about the microsynth fit and estimation results, if available.

The output includes two parts: 1) a display of key input parameters; and 2) estimated results, in a similar format as they appear when saved to .csv or .xlsx., once for each specified post-intervention evaluation time.

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Examples

```
# Use seattledmi, block-level panel data, to evaluate a crime intervention.
# Declare time-variant (outcome) and time-invariant variables for matching
cov.var <- c('TotalPop', 'BLACK', 'HISPANIC', 'Males_1521',</pre>
       'HOUSEHOLDS', 'FAMILYHOUS', 'FEMALE_HOU', 'RENTER_HOU', 'VACANT_HOU')
match.out <- c('i_felony', 'i_misdemea', 'i_drugs', 'any_crime')</pre>
set.seed(99199) # for reproducibility
# Perform matching and estimation, without permutations or jackknife
# runtime: < 1 min</pre>
sea1 <- microsynth(seattledmi,</pre>
                  idvar='ID', timevar='time', intvar='Intervention',
                  start.pre=1, end.pre=12, end.post=16,
                  match.out=match.out, match.covar=cov.var,
                  result.var=match.out, omnibus.var=match.out,
                  test='lower',
                  n.cores = min(parallel::detectCores(), 2))
# View results
print(sea1)
```

seattledmi

Data for a crime intervention in Seattle, Washington

Description

The dataset contains information used to evaluate a Drug Market Intervention (DMI) occurring in parts of Seattle, Washington in 2013. The data include 2010 block-level Census data and counts of crime reported by the Seattle Police, by crime type. Crime data are available for one year prior to the intervention and two years after. DMIs are an intervention intended to disrupt drug markets by targeting enforcement priorities at specific market participants. The intervention was applied to 39 blocks in Seattle's International District.

Usage

seattledmi

Format

A data frame with 154,272 rows and 22 columns, consisting of 9,642 unique blocks with 16 (quarterly) observations each. It contains the following variables:

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ID unique Census block ID

time time unit (in quarters)

Intervention time-variant binary indicator; all treated units receive 0 pre-intervention and 1 from the start of the intervention onward, while untreated cases receive 0s throughout

i_robbery number of robberies reported in that block-quarter (time-variant)

i_aggassau number of aggravated assaults reported

i_burglary number of burglaries reported

i_larceny number of larcenies reported

i_felony number of felony crimes reported

i_misdemea number of misdemeanor crimes reported

i_drugsale number of drug sales reported

i_drugposs number of drug possession incidents reported

i_drugs number of drug sale or possession incidents reported

any_crime number of all crimes reported

TotalPop number of residents

BLACK number of African American residents

HISPANIC number of Hispanic residents

Males_1521 number of male residents aged 15-21

HOUSEHOLDS number of households

FAMILYHOUS number of family households

FEMALE_HOU number of female-headed households

RENTER_HOU number of households occupied by renters

VACANT_HOU number of vacant housing units

Source

Demographic data obtained from the 2010 Census, and administrative crime data from the Seattle Police Department.

summary.microsynth

Summarizing microsynth Fits and Results

Description

Summary method for class 'microsynth'.

Usage

```
## S3 method for class 'microsynth'
summary(object, ...)
```

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Arguments

object A microsynth object produced by microsynth()
... further arguments passed to or from other methods.

Value

The functions print.microsynth and summary.microsynth displays information about the microsynth fit and estimation results, if available.

The output includes two parts: 1) a matching summary that compares characteristics of the treatment to the synthetic control and the population; and 2) estimated results, in a similar format as they appear when saved to .csv or .xlsx., once for each specified post-intervention evaluation time.

Examples

```
# Use seattledmi, block-level panel data, to evaluate a crime intervention.
# Declare time-variant (outcome) and time-invariant variables for matching
cov.var <- c('TotalPop', 'BLACK', 'HISPANIC', 'Males_1521',</pre>
       'HOUSEHOLDS', 'FAMILYHOUS', 'FEMALE_HOU', 'RENTER_HOU', 'VACANT_HOU')
match.out <- c('i_felony', 'i_misdemea', 'i_drugs', 'any_crime')</pre>
set.seed(99199) # for reproducibility
# Perform matching and estimation, without permutations or jackknife
# runtime: < 1 min</pre>
sea1 <- microsynth(seattledmi,</pre>
                  idvar='ID', timevar='time', intvar='Intervention',
                  start.pre=1, end.pre=12, end.post=16,
                  match.out=match.out, match.covar=cov.var,
                  result.var=match.out, omnibus.var=match.out,
                  test='lower',
                  n.cores = min(parallel::detectCores(), 2))
# View results
summary(sea1)
```

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