## Package 'pdi'

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Type Package

**Title** Phenotypic Index Measures for Oak Decline Severity

Version 0.4.2

#### **Description**

Oak declines are complex disease syndromes and consist of many visual indicators that include aspects of tree size, crown condition and trunk condition. This can cause difficulty in the manual classification of symptomatic and non-symptomatic trees from what is in reality a broad spectrum of oak tree health condition. Two phenotypic oak decline indexes have been developed to quantitatively describe and differentiate oak decline syndromes in Quercus robur. This package provides a toolkit to generate these decline indexes from phenotypic descriptors using the machine learning algorithm random forest. The methodology for generating these indexes is outlined in Finch et al. (2121) <doi:10.1016/j.foreco.2021.118948>.

URL https://jasenfinch.github.io/pdi

BugReports https://github.com/jasenfinch/pdi/issues

**License** GPL-3 **Encoding** UTF-8

**Imports** dplyr, magrittr, purrr, randomForest, readxl, stringr, tibble, tidyr, tidyselect

LazyData true

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Suggests testthat, covr, knitr, rmarkdown, ggplot2

VignetteBuilder knitr

NeedsCompilation no

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agrilus Exit Hole Density

*Agrilus exit hole density (m*^-2)

## Description

Calculate Agrilus biguttatus exit hole density.

## Usage

```
agrilusExitHoleDensity(n, d, s = 2)
```

#### **Arguments**

```
n number of Agrilus exit holes
d diameter at breast height (m)
```

s height to which stem surveyed from the tree base (m)

```
agrilusExitHoleDensity(2,1.02,1.3)
```

bleedPrevalence 3

|--|

## Description

Calculate estimated bleed prevalence.

## Usage

```
bleedPrevalence(a, A, b, B, d, s = 3)
```

## Arguments

a	average active bleed size (mm)
A	number of active bleeds
b	average black stain size (mm)
В	number of black stains
d	diameter at breast height (m)
S	height to which stem surveyed from the tree base (m)

## **Examples**

```
bleedPrevalence(30,10,40,5,1,1.3)
```

calcDIs	Calculate Decline Indexes	

## Description

Calculate Phenotypic Decline Index (PDI) and Decline Acuteness Index (DAI).

## Usage

```
calcDIs(rfModels, PDI = TRUE, DAI = TRUE, invertPDI = TRUE, invertDAI = TRUE)
```

## **Arguments**

rfModels	list containing random forest models as returned by rf()
PDI	TRUE/FALSE, calculate PDI?
DAI	TRUE/FALSE, calculate DAI?
invertPDI	invert the PDI scale? TRUE/FALSE. Ignored if argument PDI is FALSE
invertDAI	invert the DAI scale? TRUE/FALSE. Ignored if argument DAI is FALSE

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#### **Examples**

```
library(dplyr)
## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',</pre>
  package = 'pdi'),full.names = TRUE)
## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
   mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
     `Lower crown height (m)`),
     `Crown condition (%)` = crownCondition(`Missing crown (%)`,
                                `Crown transparency (%)`),
     'Crown volume (m<sup>3</sup>)' = crownVolume('Crown radius (m)',
                                `Total height (m)`,
                                `Lower crown height (m)`,
                                `Crown condition (%)`),
     'Bleed prevalence (%)' = bleedPrevalence('Active bleed length (mm)',
                                'Active bleeds',
                                'Black staining length (mm)',
                                `Black staining`,
                                'Diameter at breast height (m)'),
     `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
                                `Diameter at breast height (m)`)
)
t <- makeAnalysisTable(d)</pre>
## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)</pre>
## Calculate decline indexese
DIs <- calcDIs(m,DAI = FALSE,invertPDI = FALSE) %>%
  bind_cols(d %>%
    select(Location, ID, Status))
```

crownCondition

Crown condition

## Description

Calculate crown condition (%).

#### Usage

```
crownCondition(m, t)
```

## Arguments

```
m missing crown (%)
t crown transparency (%)
```

## Examples

```
crownCondition(50,60)
```

```
crownProductionEfficiency
```

Crown production efficiency

## Description

Calculate the crown production efficiency

## Usage

```
crownProductionEfficiency(crown_surface_area, crown_volume)
```

## Arguments

```
crown_surface_area crown surface area (m^2) crown_volume crown volume (m^3)
```

## **Examples**

```
crownProductionEfficiency(34,35)
```

crownSurfaceArea

Crown surface area (m^2)

## Description

Calculate the crown surface area

#### Usage

```
crownSurfaceArea(r, h, 1, c)
```

6 crown Volume

## Arguments

r	crown radius	(m)

h total height (m)

lower crown height (m)

c crown condition (%)

## **Examples**

```
crownSurfaceArea(3,15,10,50)
```

crownVolume

Estimated crown volume (m^3)

## Description

Calculate estimated crown volume.

## Usage

```
crownVolume(r, h, 1, c)
```

## Arguments

```
r crown radius (m)
```

h total height (m)

lower crown height (m)

c crown condition (%)

```
crownVolume(3,15,10,50)
```

descriptorContributions

descriptorContributions

Descriptor contributions

#### **Description**

Calculate average descriptor contributions to random forest models.

#### Usage

```
descriptorContributions(rfModels)
```

#### **Arguments**

rfModels

list containing random forest models as returned by rf()

#### **Details**

See see ?randomForest::importance for details on random forest importance metrics.

```
library(dplyr)
## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',</pre>
  package = 'pdi'),full.names = TRUE)
## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
   mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
     `Lower crown height (m)`),
     `Crown condition (%)` = crownCondition(`Missing crown (%)`,
                                `Crown transparency (%)`),
     'Crown volume (m<sup>3</sup>)' = crownVolume('Crown radius (m)',
                                `Total height (m)`,
                                `Lower crown height (m)`,
                                `Crown condition (%)`),
     'Bleed prevalence (%)' = bleedPrevalence('Active bleed length (mm)',
                                'Active bleeds',
                                'Black staining length (mm)',
                                'Black staining',
                                'Diameter at breast height (m)'),
     `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
                                `Diameter at breast height (m)`)
)
```

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```
t <- makeAnalysisTable(d)
## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)
descriptor_contributions <- m %>%
    descriptorContributions()
```

liveCrownRatio

Live crown ratio

## Description

Calculate the live crown ratio

## Usage

```
liveCrownRatio(h, 1)
```

## Arguments

h total height (m)

lower crown height (m)

#### **Examples**

liveCrownRatio(15,10)

makeAnalysisTable

Make analysis table

#### Description

prepare data table ready for random forest analysis

## Usage

```
makeAnalysisTable(phenoData)
```

## Arguments

phenoData

tibble containing phenotype data

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#### **Examples**

```
library(dplyr)
## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',</pre>
  package = 'pdi'),full.names = TRUE)
## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
   mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
     `Lower crown height (m)`),
     `Crown condition (%)` = crownCondition(`Missing crown (%)`,
                                `Crown transparency (%)`),
     'Crown volume (m<sup>3</sup>)' = crownVolume('Crown radius (m)',
                                `Total height (m)`,
                                `Lower crown height (m)`,
                                `Crown condition (%)`),
     `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
                                `Active bleeds`,
                                `Black staining length (mm)`,
                                `Black staining`,
                                'Diameter at breast height (m)'),
     `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
                                'Diameter at breast height (m)')
)
t <- makeAnalysisTable(d)</pre>
```

mds

Multidimensional scaling

#### **Description**

perform multidimensional scaling of random forest proximities

## Usage

```
mds(rfModels, dimensions = 2)
```

#### Arguments

rfModels list containing random forest models as returned by rf()

dimensions number of dimensions to scale to

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#### **Examples**

```
library(dplyr)
## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',</pre>
  package = 'pdi'),full.names = TRUE)
## Prepare data
d <- map(files,readPhenotypeSheet) %>%
  map(preparePhenotypeData) %>%
  bind_rows() %>%
  siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
     `Lower crown height (m)`),
     `Crown condition (%)` = crownCondition(`Missing crown (%)`,
                                `Crown transparency (%)`),
     'Crown volume (m<sup>3</sup>)' = crownVolume('Crown radius (m)',
                                `Total height (m)`,
                                `Lower crown height (m)`,
                                `Crown condition (%)`),
     `Bleed prevalence (%)` = bleedPrevalence(`Active bleed length (mm)`,
                                'Active bleeds',
                                'Black staining length (mm)',
                                `Black staining`,
                                'Diameter at breast height (m)'),
     `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
                                `Diameter at breast height (m)`)
)
t <- makeAnalysisTable(d)</pre>
## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)</pre>
mds_data <- mds(m,2)</pre>
```

minMaxScale

Min-max scaling

#### **Description**

Variable min-max scaling.

## Usage

minMaxScale(vec)

#### **Arguments**

vec

vector of numbers to scale

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#### **Examples**

```
set.seed(1234)
d <- runif(20,1,10)</pre>
minMaxScale(d)
```

phenotypingTemplate

Phenotyping template

## Description

Export a copy of the oak phenotyping data collection spreadsheet.

## Usage

```
phenotypingTemplate(path = ".")
```

## Arguments

path

directory path for export output

#### **Examples**

```
## Not run:
phenotypingTemplate()
## End(Not run)
```

preparePhenotypeData Prepare phenotype data

## Description

Process parsed phenotype data sheets into a tibble suitable for random forest analysis.

#### Usage

```
preparePhenotypeData(phenotypeData)
```

## **Arguments**

phenotypeData parsed phenotype data collection sheet returned from readPhenotypeSheet

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#### **Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
    package = 'pdi'),full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
    map(preparePhenotypeData)
```

readPhenotypeSheet

Read phenptyping sheet

## Description

Parse .xlsx phenotype data collection sheets.

#### Usage

```
readPhenotypeSheet(file)
```

#### **Arguments**

file

file path to excel file to parse

## **Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
    package = 'pdi'),full.names = TRUE)

## Prepare data
d <- readPhenotypeSheet(files[1])</pre>
```

rf

Random forest analysis

#### **Description**

Perform random forest repetitions.

#### Usage

```
rf(analysisTable, cls, params = list(), nreps = 100, seed = 1234)
```

siteAdjustment 13

#### **Arguments**

 $analysis {\tt Table} \quad tibble \ of \ phenotype \ data \ suitable \ for \ random \ forest \ analysis \ as \ returned \ by \ prepare {\tt PhenotypeData}$ 

cls analysisTable column to use as response vector. NULL for unsupervised analy-

ses.

params additional arguments to pass to randomForest::randomForest

nreps number of repetitions seed random number seed

## Examples

```
library(dplyr)
## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',</pre>
 package = 'pdi'),full.names = TRUE)
## Prepare data
d <- map(files,readPhenotypeSheet) %>%
 map(preparePhenotypeData) %>%
 bind_rows() %>%
 siteAdjustment() %>%
  mutate(`Live crown ratio (%)` = liveCrownRatio(`Total height (m)`,
     `Lower crown height (m)`),
     `Crown condition (%)` = crownCondition(`Missing crown (%)`,
                                `Crown transparency (%)`),
     'Crown volume (m<sup>3</sup>)' = crownVolume('Crown radius (m)',
                                `Total height (m)`,
                                `Lower crown height (m)`,
                                `Crown condition (%)`),
     'Bleed prevalence (%)' = bleedPrevalence('Active bleed length (mm)',
                                'Active bleeds',
                                'Black staining length (mm)',
                                `Black staining`,
                                `Diameter at breast height (m)`),
     `Agrilus exit hole density (m^-2)` = agrilusExitHoleDensity(`Agrilus exit holes`,
                                `Diameter at breast height (m)`)
)
t <- makeAnalysisTable(d)</pre>
## Generate random forest models
m <- rf(t,cls = NULL,nreps = 10)</pre>
```

siteAdjustment

Site adjustment

#### **Description**

Perform a site adjustment of selected descriptors.

#### Usage

```
siteAdjustment(
  phenoData,
  descriptors = c("Diameter at breast height (m)", "Lower crown height (m)",
    "Timber height (m)", "Total height (m)", "Crown radius (m)")
)
```

#### **Arguments**

phenoData phenoData tibble containing phenotype data

descriptors columns of phenoData on which to perform site correction

#### **Examples**

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
    package = 'pdi'), full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
    map(preparePhenotypeData) %>%
    bind_rows() %>%
    siteAdjustment()
```

siteAdjustmentFactors Site adjustment factors

#### **Description**

Return site adjustment factors of selected phenotypic descriptors.

#### Usage

```
siteAdjustmentFactors(
  phenoData,
  descriptors = c("Diameter at breast height (m)", "Lower crown height (m)",
    "Timber height (m)", "Total height (m)", "Crown radius (m)")
)
```

#### **Arguments**

phenoData phenoData tibble containing phenotype data
descriptors columns of phenoData on which calculate site correction factors

siteAdjustmentFactors 15

```
library(dplyr)

## Retrieve file paths for example data
files <- list.files(system.file('phenotypeDataCollectionSheets',
    package = 'pdi'), full.names = TRUE)

## Prepare data
d <- map(files,readPhenotypeSheet) %>%
    map(preparePhenotypeData) %>%
    bind_rows() %>%
    siteAdjustment()

sa_factors <- siteAdjustmentFactors(d)</pre>
```

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