# Package 'crso'

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Title Cancer Rule Set Optimization ('crso')

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Description An algorithm for identifying candidate driver combinations in cancer. CRSO is based on a theoretical model of cancer in which a cancer rule is defined to be a collection of two or more events (i.e., alterations) that are minimally sufficient to cause cancer. A cancer rule set is a set of cancer rules that collectively are assumed to account for all of ways to cause cancer in the population. In CRSO every event is designated explicitly as a passenger or driver within each patient. Each event is associated with a patient-specific, event-specific passenger penalty, reflecting how unlikely the event would have happened by chance, i.e., as a passenger. CRSO evaluates each rule set by assigning all samples to a rule in the rule set, or to the null rule, and then calculating the total statistical penalty from all unassigned event. CRSO uses a three phase procedure find the best rule set of fixed size K for a range of Ks. A core rule set is then identified from among the best rule sets of size K as the rule set that best balances rule set size and

Users should consult the 'crso' vignette for an example walk through of a full CRSO run.

The full description, of the CRSO algorithm is presented in:

Klein MI, Cannataro V, Townsend J, Stern DF and Zhao H. ``Identifying combinations of cancer driver in individual patients."

BioRxiv 674234 [Preprint]. June 19, 2019. <doi:10.1101/674234>.

Please cite this article if you use 'crso'.

**Depends** R (>= 3.5.0), foreach

statistical penalty.

Imports stats, utils

License GPL-2

**Encoding UTF-8** 

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RoxygenNote 6.1.1

Suggests knitr, rmarkdown

VignetteBuilder knitr

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## NeedsCompilation no

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## **R** topics documented:

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## Description

Make full rule library of all rules that satisfy minimum coverage threshold.

## Usage

```
buildRuleLibrary(D, rule.thresh, min.epr)
```

## **Arguments**

D	Binary matrix of N events and M samples
rule.thresh	Minimum fraction of rules covered. Default is .03
min.epr	minimum events per rule. Default is 2.

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## **Examples**

```
library(crso)
data(skcm)
list2env(skcm.list,envir=globalenv())
rm.full <- buildRuleLibrary(D,rule.thresh = 0.05) # build rule library
dim(rm.full) # Should be matrix with dimension 60 x 71</pre>
```

evaluateListOfIMs

Evaluate list of rule set matrices

## Description

Evaluate list of rule set matrices

#### Usage

```
evaluateListOfIMs(D, Q, rm, im.list)
```

## **Arguments**

D binary matrix of events by samples
Q penalty matrix of events by samples
rm matrix of rules ordered by phase one
im.list list of rule set matrices

#### Value

list of Js for each rule set matrix

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getBestRs	el 10†

Get list of best rule sets of size K for all K

#### **Description**

Get list of best rule sets of size K for all K

## Usage

```
getBestRsList(rm, tpl, til)
```

#### **Arguments**

rm	binary rule matrix
tpl	list of top performances

til list of top rule set index matrices

## **Examples**

getCoreK

Determine core K from phase 3 tpl and til

## **Description**

Determine core K from phase 3 tpl and til

#### Usage

```
getCoreK(D, rm, tpl, til, cov.thresh, perf.thresh)
```

#### **Arguments**

D	input matrix D
rm	binary rule matrix
tpl	list of top performances
til	list of top rule set index matrices

cov.thresh core coverage threshold, defaults is 95 core performance threshold, default is 90

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#### **Examples**

getCoreRS

Get core rules from phase 3 tpl and til

#### **Description**

Get core rules from phase 3 tpl and til

#### **Usage**

```
getCoreRS(D, rm, tpl, til, cov.thresh, perf.thresh)
```

#### **Arguments**

D	input matrix D
rm	binary rule matrix
tpl	list of top performances
til	list of top rule set index matrices
cov.thresh	core coverage threshold, defaults is 95
perf.thresh	core performance threshold, default is 90

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getGCDs

Get Generalized Core Duos

## Description

Get Generalized Core Duos

#### Usage

```
getGCDs(list.subset.cores)
```

#### **Arguments**

```
list.subset.cores
```

list of subset cores

## **Examples**

```
list.subset.cores <- list(c("A.B.C","D.E","A.D"),c("A.C","B.C.D","D.E"), c("A.B.C","D.E"),c("A.B.C","D.E","B.C.D")) getGCDs(list.subset.cores) # Confidence column should be 100, 100, 100, 75, 50, 25, 25
```

getGCEs

Get Generalized Core Events

#### **Description**

Get Generalized Core Events

## Usage

```
getGCEs(list.subset.cores)
```

## Arguments

```
list.subset.cores
```

list of subset cores

```
list.subset.cores <- list(c("A.B.C","D.E","A.D"), c("A.C","B.C.D","D.E"),c("A.B.C","D.E"),c("A.B.C","D.E","B.C.D")) getGCEs(list.subset.cores) # Confidence column should be 100, 100, 100, 100, 100
```

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getGCRs

Get Generalized Core Rules

## Description

Get Generalized Core Rules

#### Usage

```
getGCRs(list.subset.cores)
```

#### **Arguments**

```
list.subset.cores
```

list of subset cores

#### **Examples**

```
list.subset.cores <- list(c("A.B.C","D.E","A.D"),c("A.C","B.C.D","D.E"), c("A.B.C","D.E"),c("A.B.C","D.E","B.C.D")) getGCRs(list.subset.cores) # Confidence column should be 100, 75, 50, 25, 25
```

getPoolSizes

Get pool sizes for phase 2

## Description

Get pool sizes for phase 2

#### Usage

```
getPoolSizes(rm.ordered, k.max, max.nrs.ee, max.compute)
```

#### **Arguments**

rm. ordered binary rule matrix ordered from phase 1

k.max maximum rule set size

max.nrs.ee max number of rule sets per k

max.compute maximum raw rule sets considered per k

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#### **Examples**

```
library(crso)
data(skcm)
list2env(skcm.list,envir=globalenv())
rm.full <- buildRuleLibrary(D,rule.thresh = 0.05) # Rule library matrix, dimension: 60 x 71
rm.ordered <- rm.full # Skip phase one in this example
getPoolSizes(rm.ordered,k.max = 7,max.nrs.ee = 10000)
# [1] 60 60 40 23 18 16 15</pre>
```

getRulesAsStrings

Represent binary rule matrix as strings

#### **Description**

Represent binary rule matrix as strings

#### Usage

```
getRulesAsStrings(rm)
```

#### **Arguments**

rm

binary rule matrix

#### Value

vector or rules represented as strings

```
library(crso)
data(skcm)
list2env(skcm.list,envir=globalenv())
rm.full <- buildRuleLibrary(D,rule.thresh = 0.1) # Small rule library matrix, dimension: 5 x 71
getRulesAsStrings(rm.full)
# output should be: "BRAF-M.CDKN2A-MD" "CDKN2A-MD.NRAS-M"
# "BRAF-M.PTEN-MD" "ADAM18-M.BRAF-M" "ADAM18-M.CDKN2A-MD"</pre>
```

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makeFilteredImList Make filtered im list from phase 3 im list

#### **Description**

Make filtered im list from phase 3 im list

## Usage

```
makeFilteredImList(D, Q, rm, til, filter.thresh)
```

#### **Arguments**

D binary matrix of events by samples
Q penalty matrix of events by samples
rm matrix of rules ordered by phase one
til im list from phase 3

filter.thresh minimum percentage of samples assigned to each rule in rs

#### Value

filtered top im list

#### **Examples**

makePhaseOneOrderedRM Order rules according to phase one importance ranking

## **Description**

Order rules according to phase one importance ranking

## Usage

```
makePhaseOneOrderedRM(D, rm.start, spr, Q, trn, n.splits, shouldPrint)
```

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#### **Arguments**

D	Binary matrix of N events and M samples
rm.start	Starting binary rule matrix (i.e., rule library)

spr Random rule sets per rule in each phase one iteration. Default is 40.

Q Penalty matrix, negative log of passenger probability matrix.trn Target rule number for stopping iterating. Default is 16.

n.splits number of splits for parallelization. Default is all available cpus.

shouldPrint Print progress updates? Default is TRUE

#### Value

binary rule matrix ordered by phase one importance ranking

#### **Examples**

```
data(skcm)
list2env(skcm.list,envir=globalenv())
Q <- log10(P)
rm.full <- buildRuleLibrary(D,rule.thresh = 0.06) # Rule library matrix, dimension: 36s x 71
rm.ordered <- makePhaseOneOrderedRM(D,rm.full,spr = 1,Q,trn = 34,shouldPrint = TRUE)
# note, for real applications, spr should be at least 40.</pre>
```

makePhaseThreeImList Make phase 3 im list from phase 2 im list

#### **Description**

Make phase 3 im list from phase 2 im list

#### Usage

```
makePhaseThreeImList(D, Q, rm.ordered, til.ee, pool.sizes, max.stored,
  max.nrs.borrow, shouldPrint)
```

## Arguments

D	binary matrix of events by samples
Q	penalty matrix of events by samples
rm.ordered	matrix of rules ordered by phase one

til.ee list of rule set matrices (im list) from phase two

pool.sizes pool sizes for phase two
max.stored max number of rule sets saved

max.nrs.borrow max number of new rule sets per k, default is 10<sup>5</sup>

shouldPrint Print progress updates? Default is TRUE

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#### Value

```
phase 3 top im list
```

#### **Examples**

 ${\tt makePhaseTwoImList}$ 

Output list of top rule sets for each k in 1:k.max

#### **Description**

Output list of top rule sets for each k in 1:k.max

## Usage

```
makePhaseTwoImList(D, Q, rm.ordered, k.max, pool.sizes, max.stored,
    shouldPrint)
```

## Arguments

D	binary matrix of events by samples
Q	penalty matrix of events by samples
rm.ordered	matrix of rules ordered by phase one
k.max	max k
pool.sizes	vector of the number of top rules evaluated for each $\boldsymbol{k}$
max.stored	max number of rule sets saved
shouldPrint	Print progress updates? Default is TRUE

#### Value

largest n such that n choose k < max.num.rs

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#### **Examples**

makeSubCoreList

Get list of core rules from random subsets of samples

#### **Description**

Get list of core rules from random subsets of samples

#### **Usage**

```
makeSubCoreList(D, Q, rm, til, num.subsets, num.evaluated, shouldPrint)
```

#### Arguments

D	input matrix D
Q	input matrix Q
rm	binary rule matrix
til	list of top rule set index matrices
num.subsets	number of subset iterations, default is 100
num.evaluated	number of top rs considered per k per iteration, default is 1000
shouldPrint	Print progress updates? Default is TRUE

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	skcm.list	Example data set derived from TCGA skin cutaneous melanoma (SKCM) data.
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## Description

A dataset containing the processed inputs used in the melanoma analysis within the CRSO publication.

## Usage

skcm.list

#### **Format**

A list with 3 items

- **D** Binary alteration matrix. Rows are candidate driver events, columns are samples.
- **P** Passenger probability matrix corresponding to D. **cnv.dictionary** Data frame containing copy number genes. ...

#### **Source**

Dataset derived from data generated by the TCGA Research Network: https://www.cancer.gov/tcga

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