# Package 'nplr'

October 13, 2022

Type Package

Title N-Parameter Logistic Regression

Version 0.1-7
<b>Date</b> 2016-12-25
Maintainer Frederic Commo <fredcommo@gmail.com></fredcommo@gmail.com>
<b>Depends</b> methods
Imports stats, graphics, utils
Suggests RUnit,knitr
VignetteBuilder knitr
<b>Description</b> Performing drug response analyses and IC50 estimations using n-Parameter logistic regression. Can also be applied to proliferation analyses.
License GPL
<pre>URL https://github.com/fredcommo/nplr</pre>
NeedsCompilation no
Author Frederic Commo [aut, cre], Brian M. Bot [aut]
Repository CRAN
<b>Date/Publication</b> 2016-12-28 16:38:22
R topics documented:  convertToProp
nplr
nplrAccessors
overlay
plot.nplr
Index 12

2 convertToProp

#### **Description**

Convert a vector of values to proportions, given a minimun and a maximun value (optional). See Details and Examples.

## Usage

```
convertToProp(y, T0 = NULL, Ctrl = NULL)
```

## Arguments

y : a vector of values (responses to x).

T0 : the minimal value to consider. If NULL (default), min(y, na.rm=TRUE) will

be used. See Details and Warning.

ctrl : the maximal value to consider. If NULL (default), max(y, na.rm=TRUE) will

be used. See Details and Warning.

#### **Details**

In typical cell viability experiments, responses to drug concentrations (inhibition rate) may be estimated with respect to a time zero (T0) and an untreated condition values (Ctrl), as described in [1]:

If none of the TO and Ctrl values are provided, min(y, na.rm=TRUE) and max(y, na.rm=TRUE) will be used, respectively. See Warning.

## Value

a vector of values.

*Warning:* Note that, for drug response analyses, rescaling the responses between 0 to 1 using to the min and max of y, would lead to estimate a EC50 (the half effect between the maximum and the minimum of the observed effects), rather than a IC50.

#### Note

The data used as examples come from the NCI-60 Growth Inhibition Data: https://wiki.nci.nih.gov/display/NCIDTPdata/NCI-60+Growth+Inhibition+Data, except for multicell.tsv which are simulated data.

## Author(s)

Frederic Commo, Brian M. Bot

getEstimates 3

#### References

1 - https://dtp.nci.nih.gov/branches/btb/ivclsp.html

#### See Also

nplr

#### **Examples**

```
## Using the MDA-N data
  op <- par(no.readonly=TRUE)</pre>
                                      # save default parameters
  require(nplr)
  path <- system.file("extdata", "mdan.txt", package = "nplr")</pre>
  mdan <- read.delim(path)</pre>
# fit a model on the original responses (proportions of control):
  conc <- mdan$CONC</pre>
  y0 <- mdan$GIPROP
  model0 <- nplr(conc, y0)</pre>
# Adjust the data between 0 to 1, then fit a new model:
  y1 <- convertToProp(y0)</pre>
  model1 <- nplr(conc, y1)</pre>
  par(mfrow=c(1, 2))
  plot(model0, ylim = range(0, 1), main = "Original y values")
  plot(model1, ylim = range(0, 1), main = "Rescaled y values")
  par(op)
```

getEstimates

Function to Estimate x Given y.

## **Description**

This function takes as its first argument a model returned by nplr(). By inverting the logistic model, it estimates the x values corresponding to one (or a vector of) y target(s) provided. The standard error of the model, defined as the mean squared error on the fitted values, is used to estimate a confidence interval on the predicted x values, according to the specified conf.level. see Details.

## Usage

```
## S4 method for signature 'nplr'
getEstimates(object, targets = seq(.9, .1, by = -.1), B = 1e4, conf.level = .95)
```

4 getEstimates

#### **Arguments**

object : an object of class nplr.

targets : one, of a vector of, numerical value(s) for which the corresponding x has to be

estimated. Default are target values from .9 to .1.

B : the length of the y distribution from which the x confidence interval is esti-

mated.

conf.level : the estimated x confidence interval, bounded by (1-conf.level)/2 and 1 - (1-

conf.level)/2 (by default .95, which gives x.025 and x.975).

#### **Details**

In n-parameter logistic regressions, none of the parameters follow any particular distribution from which confidence intervals can be estimated. To overcome this issue, the standard error is used to generate a normal distribution of the target(s) passed to the function. The quantiles of that distribution are used in order to provide estimated bounds for the corresponding x value, with respect to conf.level. See also Warning.

#### Value

A data set containing:

y : the target value.

x.05 : the lower bound of the estimated 95% confidence interval (default). If another

value is passed to conf.level, x will be labelled as x.(1-conf.level)/2.

x : the estimated value.

x.95 : the upper bound of the estimated 95% confidence interval (default). If another

value is passed to conf.level, x will be labelled as x.1-(1-conf.level)/2.

Warning: Notice that, if any  $target \le B$  or  $target \ge T$ , in other words outside the 2 asymptotes, the maximal (or minimal) possible value the model can estimates is returned.

#### Note

The data used in the examples are samples from the NCI-60 Growth Inhibition Data: https://wiki.nci.nih.gov/display/NCIDTPdata/NCI-60+Growth+Inhibition+Data, except for multicell.tsv which are simulated data.

#### Author(s)

Frederic Commo, Brian M. Bot

#### See Also

```
nplr, plot.nplr, , nplrAccessors
```

nplr 5

#### **Examples**

```
# Using the PC-3 data
require(nplr)
path <- system.file("extdata", "pc3.txt", package="nplr")
pc3 <- read.delim(path)
model <- nplr(x = pc3$CONC, y = pc3$GIPROP)
getEstimates(model)
getEstimates(model, c(.3, .6), conf.level = .9)</pre>
```

nplr

Function to Fit n-Parameter Logistic Regressions.

## **Description**

This function computes a weighted n-parameters logistic regression, given x (typically compound concentrations) and y values (responses: optic densities, fluorescence, cell counts,...). See Details.

## Usage

## **Arguments**

x : a vector of numeric values, e.g. a vector of drug concentrations.

y : a vector of numeric values, e.g. a vector of responses, typically provided as

proportions of control.

useLog : Logical. Should x-values be Log10-transformed. Default to TRUE, set to FALSE

if x is already in Log10.

LPweight: a coefficient to adjust the weights. LPweight = 0 will compute a non-

weighted np-logistic regression.

npars : a numeric value (or "all") to specify the number of parameters to use in the

model. If "all" the logistic model will be tested with 2 to 5 parameters, and the

best option will be returned. See Details

method : a character string to specify what weight method to use. Options are "res" (Default),

"sdw", "gw". See Details

silent : Logical. Specify whether warnings ad/or messages has to be silenced. De-

fault to FALSE.

#### **Details**

The 5-parameter logistic regression is of the form:

$$y = B + (T - B)/[1 + 10^{(b)} * (xmid - x)]^{s}$$

6 nplr

where B and T are the bottom and top asymptotes, respectively, b and xmid are the Hill slope and the x-coordinate at the inflexion point, respectively, and s is an asymetric coefficient. This equation is sometimes referred to as the Richards' equation [1,2].

When specifying npars = 4, the s parameter is forced to be 1, and the corresponding model is a 4-parameter logistic regression, symetrical around its inflexion point. When specifying npars = 3 or npars = 2, add 2 more constraints and force B and T to be 0 and 1, respectively.

#### Weight methods:

The model parameters are optimized, simultaneously, using nlm, given a sum of squared errors function, sse(Y), to minimize:

$$sse(Y) = \Sigma [W.(Yobs - Yfit)^{2}]$$

where Yobs, Yfit and W are the vectors of observed values, fitted values and weights, respectively. In order to reduce the effect of possible outliers, the weights can be computed in different ways, specified in nplr:

• residual weights, "res":

$$W = (1/residuals)^L Pweight$$

where residuals and LPweight are the squared error between the observed and fitted values, and a tuning parameter, respectively. Best results are generally obtained by setting LPweight=0.25 (default value), while setting LPweight=0 results in computing a non-weighted sum of squared errors.

• standard weights, "sdw":

$$W = 1/Var(Yobs_r)$$

where Var(Yobs\_r) is the vector of the within-replicates variances.

• general weights, "gw":

$$W = 1/Y fit^L Pweight$$

where Yfit are the fitted values. As for the residuals-weights method, setting LPweight = 0 results in computing a non-weighted sum of squared errors.

The standard weights and general weights methods are describes in [3].

#### Value

An object of class nplr.

#### slots

- x: the x values as they are used in the model. It can be Log10(x) if useLog was set to TRUE.
- y: the y values.
- useLog: logical.
- npars: the best number of parameters if npars="all", the specified number of parameters, otherwise.
- LPweight: the weights tuning parameter.

nplr 7

- yFit: the y fitted values.
- xCurve: the x values generated to draw the curve. 200 points between the min and max of x.
- yCurve : the fitted values used to draw the curve. the fitted values corresponding to xCurve.
- inflPoint : the inflexion point x and y coordinates.
- goodness: the goodness-of-fit. The correlation between the fitted and the observed y values
- stdErr: the mean squared error between the fitted and the observed y values
- pars: the model parameters.
- AUC : the area under the curve estimated using both the trapezoid method and the Simpson's rule.

#### Note

The data used in the examples are samples from the NCI-60 Growth Inhibition Data: https://wiki.nci.nih.gov/display/NCIDTPdata/NCI-60+Growth+Inhibition+Data, except for multicell.tsv which are simulated data.

#### Author(s)

Frederic Commo, Brian M. Bot

#### References

- 1- Richards, F. J. (1959). A flexible growth function for empirical use. J Exp Bot 10, 290-300.
- 2- Giraldo J, Vivas NM, Vila E, Badia A. Assessing the (a)symmetry of concentration-effect curves: empirical versus mechanistic models. Pharmacol Ther. 2002 Jul;95(1):21-45.
- 3- Motulsky HJ, Brown RE. Detecting outliers when fitting data with nonlinear regression a new method based on robust nonlinear regression and the false discovery rate. BMC Bioinformatics. 2006 Mar 9;7:123.

#### See Also

convertToProp, getEstimates, plot.nplr, nplrAccessors

#### **Examples**

```
# Using the PC-3 data
  require(nplr)
  path <- system.file("extdata", "pc3.txt", package = "nplr")
  pc3 <- read.delim(path)
  model <- nplr(x = pc3$CONC, y = pc3$GIPROP)
  plot(model)</pre>
```

8 overlay

nplrAccessors

nplr accessor functions

## **Description**

Methods for extracting information from an object of class nplr. Each of the below methods are simply convenience functions which extract the corresponding slots (as the name of each method suggests) from the object of class nplr.

#### Methods

```
signature(object = "nplr")  • getX(object)
```

- getY(object)
- getXcurve(object)
- getYcurve(object)
- getFitValues(object)
- getInflexion(object)
- getPar(object)
- getAUC(object)
- getGoodness(object)
- getStdErr(object)
- getWeights(object)

## See Also

nplr, getEstimates

overlay

Plotting Multiple nplr Objects

## **Description**

To superimpose multiple logistic models fitted using nplr.

#### Usage

```
overlay(modelList = NULL, showLegend = TRUE, Cols = NULL, ...)
```

## **Arguments**

```
modelList : list. A list of objects of class nplr.
```

showLegend : logical. Whether the legend has to be displayed.

Cols : character. A vector of colors to use. If NULL (default), greys will be used.

... : Other graphical parameters. See par.

plot.nplr 9

## **Details**

None

#### **Source**

None

#### References

None

#### See Also

```
plot.nplr
```

## **Examples**

```
path <- system.file("extdata", "multicell.tsv", package="nplr")
multicell <- read.delim(path)

# Computing models (to store in a list)
cellsList <- split(multicell, multicell$cell)
Models <- lapply(cellsList, function(tmp){
   nplr(tmp$conc, tmp$resp, silent = TRUE)
   })

# Visualizing
overlay(Models, xlab = expression(Log[10](Conc.)), ylab = "Resp.",
   main="Superimposing multiple curves", cex.main=1.5)</pre>
```

plot.nplr

Plotting nplr Objects

## Description

This function allows visualizing logistic models fitted using nplr.

## Usage

```
## S3 method for class 'nplr'
plot(x, pcol = "aquamarine1", lcol = "red3",
    showEstim = FALSE, showCI = TRUE, showGOF = TRUE, showInfl = FALSE,
    showPoints = TRUE, showSDerr = FALSE, B = 1e4, conf.level = .95, unit = "", ...)
```

10 plot.nplr

#### **Arguments**

x : an object of class nplr

pcol : the points color.
lcol : the line color.

showEstim : logical/numeric. If a numerical value is passed (a y value to reach), the esti-

mated x value, and interval, is displayed on the plot. Default is FALSE

showCI : logical. show the estimated confidence interval showGOF : logical. show the estimated goodness-of-fit. showInf1 : logical. add the inflexion point on the plot.

showPoints : logical. add the points on the plot.

showSDerr : logical. add the standard errors on the plot (maybe useful in case of experiment

with replicates).

B : the length of simulated y values. Used to estimate the confidence interval

conf.level : the confidence level. See getEstimates

unit : the unit to specify when showEstim is TRUE. Default is an empty string.

... : other graphical parameters. See par.

#### **Details**

None

#### Note

The data used in the examples are samples from the NCI-60 Growth Inhibition Data: https://wiki.nci.nih.gov/display/NCIDTPdata/NCI-60+Growth+Inhibition+Data, except for multicell.tsy which are simulated data.

#### Source

None

#### References

None

#### See Also

overlay

## **Examples**

```
# Using the PC-3 data
  require(nplr)
path <- system.file("extdata", "pc3.txt", package = "nplr")
pc3 <- read.delim(path)
model <- nplr(x = pc3$CONC, y = pc3$GIPROP)
plot(model, showEstim = 0.5, unit = "nM")</pre>
```

summary.nplr 11

summary.nplr

summaryzing nplr Objects

## **Description**

A S3 method to visualize a model summary as a table.

## Usage

```
## S3 method for class 'nplr'
summary(object, ...)
```

## Arguments

object : an object of class nplr

... : other optional parameters (not used).

## **Details**

None

#### Note

The data used in the examples are samples from the NCI-60 Growth Inhibition Data: https://wiki.nci.nih.gov/display/NCIDTPdata/NCI-60+Growth+Inhibition+Data, except for multicell.tsv which are simulated data.

#### **Source**

None

## References

None

## See Also

```
plot.nplr
```

#### **Examples**

```
# Using the PC-3 data
require(nplr)
path <- system.file("extdata", "pc3.txt", package = "nplr")
pc3 <- read.delim(path)
model <- nplr(x = pc3$CONC, y = pc3$GIPROP)
summary(model)</pre>
```

## **Index**

```
* datasets
                                                getXcurve,nplr-method(nplrAccessors), 8
    overlay, 8
                                                getXcurve-methods (nplrAccessors), 8
    plot.nplr, 9
                                                getY (nplrAccessors), 8
    summary.nplr, 11
                                                getY, nplr-method (nplrAccessors), 8
                                                getY-methods (nplrAccessors), 8
convertToProp, 2, 7
                                                getYcurve (nplrAccessors), 8
                                                getYcurve,nplr-method(nplrAccessors), 8
getAUC (nplrAccessors), 8
                                                getYcurve-methods (nplrAccessors), 8
getAUC, nplr-method (nplrAccessors), 8
getAUC-methods (nplrAccessors), 8
                                                nplr, 3, 4, 5, 8–11
getEstimates, 3, 7, 8, 10
                                                nplr-class (nplr), 5
getEstimates,nplr-method
                                                nplrAccessors, 4, 7, 8
        (getEstimates), 3
                                                nplrAccessors, nplr-method
getEstimates-methods (getEstimates), 3
                                                         (nplrAccessors), 8
getFitValues (nplrAccessors), 8
                                                nplrAccessors-methods (nplrAccessors), 8
getFitValues,nplr-method
                                                overlay, 8, 10
        (nplrAccessors), 8
getFitValues-methods (nplrAccessors), 8
                                                 par, 8, 10
getGoodness (nplrAccessors), 8
                                                plot.nplr, 4, 7, 9, 9, 11
getGoodness,nplr-method
        (nplrAccessors), 8
                                                 summary.nplr, 11
getGoodness-methods (nplrAccessors), 8
getInflexion(nplrAccessors), 8
getInflexion,nplr-method
        (nplrAccessors), 8
getInflexion-methods (nplrAccessors), 8
getPar (nplrAccessors), 8
getPar,nplr-method(nplrAccessors), 8
getPar-methods (nplrAccessors), 8
getStdErr (nplrAccessors), 8
getStdErr,nplr-method(nplrAccessors), 8
getStdErr-methods (nplrAccessors), 8
getWeights (nplrAccessors), 8
getWeights,nplr-method(nplrAccessors),
getWeights-methods (nplrAccessors), 8
getX (nplrAccessors), 8
getX, nplr-method (nplrAccessors), 8
getX-methods (nplrAccessors), 8
getXcurve (nplrAccessors), 8
```