Package 'copcor'

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LazyLoad yes

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Title Corre	elates of Protection and Correlates of Risk Functions
Depends R	R (>= 3.6), kyotil
Imports m	nethods
Suggests R	RUnit, R.rsp, survival
ers as	n Correlates of protection (CoP) and correlates of risk (CoR) study the immune biomark-sociated with an infectious disease outcome, e.g. COVID or HIV-1 infection. This pack-ontains shared functions for analyzing CoP and CoR, including bootstrapping proces, competing risk estimation, and bootstrapping marginalized risks.
VignetteBu	nilder R.rsp
License GI	PL (>= 2)
NeedsCom	pilation no
Yiwe Chen Bhav	ouyi Fong [cre], on He [aut], chen Yu [aut], esh Borate [aut], Gilbert [aut]
Maintainer	r Youyi Fong <youyifong@gmail.com></youyifong@gmail.com>
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copcor

copcor

Description

Functions used in the study of correlates of protection and correlates of risk.

See the Index link below for a list of available functions.

cove.boost.collapse.strata

Collapse sample strata for COVE boost correlates study

Description

Collapse sample strata for COVE boost correlates study

Usage

```
cove.boost.collapse.strata (dat.b, n.demo)
```

Arguments

dat.b data frame

n. demo number of demographics strata, e.g. 6 for COVE correlates

Details

This function is used by both correlates_processing repo and correlates_reporting3 repo

Value

dat.b, whose Wstratum has been updated

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plotting

Plotting Helper Functions

Description

Functions for plotting.

Usage

```
draw.x.axis.cor(xlim, llox, llox.label, for.ggplot=FALSE)
get.xlim(dat, marker, lloxs)
```

Arguments

xlim xlim
llox lower limit

llox.label label for lower limit

for.ggplot Boolean dat data frame

marker name of the biomarker variable

lloxs list of lloxs

Details

draw.x.axis.cor is used by both cor_coxph and cor_threshold

Value

real

predictCompetingRisk

Cumulative Incidence Function (CIF) Under Competing Risk

Description

Offers two approaches (Approach 2 is recommended, pcr2 is just an alias for predictCompetingRisk2). Weights are allowed in the optional arguments.

Usage

```
predictCompetingRisk2(formula.list, data, t0, newdata = data, ...)
pcr2(formula.list, data, t0, newdata=data, ...)
predictCompetingRisk(formula, formula.all, data, t0, newdata=data, stype=2, ctype=2, ...)
pcr(formula, formula.all, data, t0, newdata=data, stype=2, ctype=2, ...)
```

Arguments

formula.list list of formulae for cause-specific failures. Assume the first cause to be the cause

of interest

formula for the cause-specific failure

formula all formula for all-cause failure

data data frame

the time till which cumulative incidence function is computed

newdata new data for making prediction, default to the data for fitting the models

stype computation of the survival curve, 1=direct, 2= exponenial of the cumulative

hazard. Default 2, which is the default of basehaz and predict.coxph

ctype whether the cumulative hazard computation should have a correction for ties,

1=no, 2=yes. Default 2, which is the default of basehaz and predict.coxph

When there is only one cause, CIF is conceptually 1 - surival prob.(https://www.publichealth.columbia.edu/research/population

... optional arguments that are passed to coxph, the most import of which is weights

Details

Approach 2, predictCompetingRisk2, fits cause-specific Cox models to each cause to compute cumulative incidence function for the cause of interest under competing risk.

health-methods/competing-risk-analysis)

The function is implemented in R with matrix operation. Because looping through time points and

The function is implemented in R with matrix operation. Because looping through time points and subjects is vectorized, it is quite fast (faster than riskRegression in limited testing, which implements in C, but per uses more memory.)

One way to check the implementation of this function is to compare its results with the results of predict.coxph when there is only one cause. The tests in the examples code below show that when the risk is small (e.g. shorter followup time), the CIF computed by this function and the 1-survival estimated via 1-exp(-H) by predict.coxph, where H is cumulative hazard, are close to each other. But when the risk is high, the difference between the two are more noticeable. These results make sense because, e.g.,

If t0 = the first time failure point, CIF = h1 = $H1 \sim 1$ -exp(-H1) If t0 = the second time point,

CIF = H1 + exp(-H1) * h2 (by def)

 $\sim H1 + \exp(-H1)(1 - \exp(-h2))$

 $= H1 + \exp(-H1) - \exp(-H2)$

 $\sim 1 - \exp(-H2)$

Approach 1, predictCompetingRisk, fits a cause-specific Cox model and a all-cause Cox model to compute cumulative incidence function for the cause of interest under competing risk.

The difference between predictCompetingRisk and predictCompetingRisk2 is that instead of fitting a model to the overall failure, a model is fit for each cause, including the cause of interest. The overall survival is computed by adding together the cumulative hazard from individual causes.

The second approach is recommended because it is more stable.

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Value

A vector of real numbers as the risk till t0 for each subject in newdata

References

riskRegression: Predicting the Risk of an Event using Cox Regression Models by Brice Ozenne, Anne Lyngholm Sorensen, Thomas Scheike, Christian Torp-Pedersen, Thomas Alexander Gerds https://journal.r-project.org/archive/2017/RJ-2017-062/RJ-2017-062.pdf Thanks to Professor Gerds for helpful discussion.

Competing Risk Analysis Columbia Public Health https://www.publichealth.columbia.edu/research/population-health-methods/competing-risk-analysis

Introduction to the Analysis of Survival Data in the Presence of Competing Risks Peter C Austin, Douglas S Lee, Jason P Fine https://www.ahajournals.org/doi/full/10.1161/CIRCULATIONAHA.115.017719

See Also

predictCompetingRisk.

Examples

```
library(survival)
# prepare a dataset with competing risk
lung1=lung[order(lung$time),]
lung1$status=lung1$status-1
lung1$status[1:50]=2
with(lung1, table(status))
lung1$status.1=ifelse(lung1$status==1,1,0)
lung1$status.2=ifelse(lung1$status==2,1,0)
lung1$status.a=ifelse(lung1$status==0,0,1)
lung1$wt=rep(1, nrow(lung1))
# predictCompetingRisk2
t0=1000
formula.list=list(
   Surv(time, status.1) ~ age,
   Surv(time, status.2) ~ age
)
cif.2=pcr2(formula.list, lung1, t0)
fit=coxph(formula.list[[1]], lung1)
newdata=lung1
newdata$time=t0
coxpred = 1 - exp(-predict(fit, newdata=newdata, type="expected"))
```

```
plot(cif.2, coxpred)
# dealing with weights
lung1$wt=c(rep(2,50), rep(1, nrow(lung1)-50))
cif=predictCompetingRisk2(formula.list, lung1, t0, weights=lung1$wt)
# predictCompetingRisk
t0=1000
form =Surv(time, status.1) ~ age
form.a=Surv(time, status.a) ~ age
cif=predictCompetingRisk(form, form.a, lung1, t0, newdata=lung1, weights=lung1$wt, stype=2,ctype=2)
# more validation code
# when there is no covariate and one cause, CIF = 1 - KM estimate of survival prob
lung1=lung[order(lung$time),]
lung1$status=lung1$status-1
with(lung1, table(status))
lung1$status.1=ifelse(lung1$status==1,1,0)
lung1$status.a=ifelse(lung1$status==0,0,1)
lung1$wt=rep(1, nrow(lung1))
# stype=2 is surv=prod limit
fitKM <- survfit(Surv(time, status.1) ~ 1, data=lung1, stype=1, ctype=2)</pre>
#[1,] 0.004385965 0.9956236 0.9956140
#[2,] 0.017660474 0.9824946 0.9824561
cif=predictCompetingRisk(Surv(time, status.1) ~ 1, Surv(time, status.1) ~ 1, lung1, t0=11,
   newdata=lung1[1,,drop=FALSE], weights=lung1$wt, stype=1, ctype=2)
cif # 0.01754386
1-cif # 0.9824561 = summary(fitKM)$surv at t=11
# when there are covariates and one cause, CIF and 1-exp(-H) are close to each other
# when H is small but not close when H is large
form =Surv(time, status.1) ~ age
form.a=Surv(time, status.a) ~ age
oldpar <- par(mfrow = c(1,2))
for (t0 in c(12,1000)) {
   fit=coxph(form, lung1, weights=lung1$wt)
   lung2=lung1; lung2$time=t0
   r=predict(fit, type="expected", newdata=lung2)
   message(head(basehaz(fit, centered=TRUE)))
  cif=predictCompetingRisk(form, form.a, lung1, t0, newdata=lung1, weights=lung1$wt, stype=2,
       ctype=2)
   plot(cif, 1-exp(-r), xlab="Cumulative incidence function",
       ylab="Expected number of events from predict.coxph", main=paste0("t0: ", t0))
```

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```
abline(0,1)
  message(head(cbind(cif, "1-exp(-r)"=1-exp(-r))))
}
par(oldpar)
```

utils

Utility Functions

Description

Helpful functions.

Usage

```
marker.name.to.assay (a)
```

Arguments

а

assay name

Details

This function ...

Value

string

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