Package 'AnanseSeurat'

November 11, 2023

```
Title Construct ANANSE GRN-Analysis Seurat
Version 1.2.0
Description Enables gene regulatory network (GRN) analysis on single cell clusters,
     using the GRN analysis software 'ANANSE', Xu et al.(2021) <doi:10.1093/nar/gkab598>.
     Export data from 'Seurat' objects, for GRN analysis by 'ANANSE'
     implemented in 'snakemake'. Finally, incorporate results for visualization
     and interpretation.
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```

2 config_scANANSE

R topics documented:

conf	ig_scANANSE config_scANANSE	
Index		11
	per_cluster_df	10
	Maelstrom_Motif2TF	
	import_seurat_scANANSE	8
	import_seurat_maelstrom	7
	Factor_Motif_Plot	6
	export_CPM_scANANSE	5
	export_ATAC_scANANSE	
	export_ATAC_maelstrom	
	DEGS_scANANSE	
	config_scANANSE	2

Description

This functions generates a sample file and config file for running Anansnake based on the seurat object

Usage

```
config_scANANSE(
   seurat_object,
   output_dir,
   min_cells = 50,
   cluster_id = "seurat_clusters",
   genome = "./scANANSE/data/hg38",
   additional_contrasts = c()
)
```

Arguments

```
seurat_object seurat object
output_dir directory where the files are outputted
min_cells minimum of cells a cluster needs to be exported
cluster_id ID used for finding clusters of cells
genome genomepy name or location of the genome fastq file
additional_contrasts
additional contrasts to add between clusters within cluster_ID
```

Value

None, outputs snakemake config file in the output directory

DEGS_scANANSE 3

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
config_scANANSE(sce_small, min_cells = 2, output_dir = tempdir())</pre>
```

DEGS_scANANSE

DEGS_scANANSE

Description

Calculate the differential genes needed for ananse influence

Usage

```
DEGS_scANANSE(
    seurat_object,
    output_dir,
    min_cells = 50,
    cluster_id = "seurat_clusters",
    genome = "./scANANSE/data/hg38",
    RNA_count_assay = "RNA",
    additional_contrasts = "None"
)
```

Arguments

Value

None, outputs DEG files in the output directory

```
sce_small <- readRDS(system.file("extdata","sce_obj_tiny.Rds",package = 'AnanseSeurat'))
DEGS_scANANSE(sce_small, min_cells = 2, output_dir = tempdir())</pre>
```

```
export_ATAC_maelstrom export_seurat_Maelstrom
```

Description

normalize and export the peak table of a seurat object based on clusters

Usage

```
export_ATAC_maelstrom(
   seurat_object,
   output_dir,
   min_cells = 50,
   ATAC_peak_assay = "peaks",
   cluster_id = "seurat_clusters",
   select_top_rows = TRUE,
   n_top_rows = 1e+05
)
```

Arguments

```
seurat_object object
output_dir directory where the files are outputted
min_cells minimum of cells a cluster needs to be exported
ATAC_peak_assay
assay of the seurat object containing the peaks and peakcounts
cluster_id ID used for finding clusters of cells
select_top_rows
only output the top variable rows, or all rows if false
n_top_rows amount of variable rows to export
```

Value

None, outputs maelstrom peak counts table in the output directory

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
config_scANANSE(sce_small, min_cells = 2, output_dir = tempdir())</pre>
```

```
export_ATAC_scANANSE
export_ATAC_scANANSE
```

Description

This functions exports ATAC values from a seurat object

Usage

```
export_ATAC_scANANSE(
   seurat_object,
   output_dir,
   min_cells = 50,
   ATAC_peak_assay = "peaks",
   cluster_id = "seurat_clusters"
)
```

Arguments

```
seurat_object object
output_dir directory where the files are outputted
min_cells minimum of cells a cluster needs to be exported
ATAC_peak_assay
assay of the seurat object containing the peaks and peakcounts
cluster_id ID used for finding clusters of cells
```

Value

None, outputs ATAC peak count file in the output directory

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
export_ATAC_scANANSE(sce_small, min_cells = 2, output_dir = tempdir())</pre>
```

```
export\_CPM\_scANANSE export\_CPM\_scANANSE
```

Description

This functions exports CPM values from a seurat object

Factor_Motif_Plot

Usage

```
export_CPM_scaNANSE(
   seurat_object,
   output_dir,
   min_cells = 50,
   RNA_count_assay = "RNA",
   cluster_id = "seurat_clusters"
)
```

Arguments

```
seurat_object the seurat object used to export the CPM values from
output_dir directory where the files are outputted
min_cells minimum of cells a cluster needs to be exported
RNA_count_assay
assay of the seurat object containing the RNA count data
cluster_id ID used for finding clusters of cells
```

Value

None, outputs CPM and counts files in the output directory

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
export_CPM_scANANSE(sce_small, min_cells = 2, output_dir = tempdir())</pre>
```

```
Factor_Motif_Plot Factor_Motif_Plot
```

Description

plot both expression of a TF, and the motif accessibility of the associated motif. Finally, fetch the motif logo from the Maelstrom directory.

Usage

```
Factor_Motif_Plot(
    seurat_object,
    TF_list,
    assay_RNA = "RNA",
    assay_maelstrom = "MotifTFanticor",
    logo_dir = "~/maelstrom/logos",
    col = c("darkred", "white", "darkgrey"),
    dim_reduction = "umap"
)
```

Arguments

```
seurat_object seurat object
```

TF_list list of TFs to plot the expression and linked motif Z-score for

assay_RNA RNA_count_assay assay containing the RNA data

assay_maelstrom

maelstrom assay used for zscore vizualization, often either TFcor or TFanticor

logo_dir directory containing motif logos generated by gimme maelstrom

col colours used for zscore vizualization
dim_reduction dimensionality reduction method to use

Value

patchwork plot containing a expression dimreduction plot, a maelstrom motif score dimreduction plot, and a png image of the motif

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
logos_dir_path <- system.file("extdata","maelstrom","logos",package = 'AnanseSeurat')
sce_small <- Factor_Motif_Plot(sce_small,
    c('gene1', 'gene2'),
    dim_reduction = 'pca',
    logo_dir = logos_dir_path)</pre>
```

import_seurat_maelstrom

import seurat Maelstrom

Description

load Maelstrom enriched motifs

Usage

```
import_seurat_maelstrom(
  seurat_object,
  cluster_id = "seurat_clusters",
  maelstrom_file = "~/final.out.txt",
  return_df = FALSE
)
```

Arguments

seurat_object object

cluster_id ID used for finding clusters of cells

maelstrom_file maelstrom final.out.txt file

return_df return both the seurat object and a dataframe with maelstrom scores as a list

Value

seurat object with the maelstrom motif scores addes as an assay

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
maelstromfile_path <- system.file("extdata","maelstrom","final.out.txt",package = 'AnanseSeurat')
sce_small <- import_seurat_maelstrom(sce_small, maelstrom_file = maelstromfile_path)</pre>
```

import_seurat_scANANSE

import_seurat_scANANSE

Description

import the influences from a anansnake directory into a seurat object

Usage

```
import_seurat_scANANSE(
   seurat_object,
   cluster_id = "seurat_clusters",
   anansnake_inf_dir = "None",
   return_df = FALSE
)
```

Arguments

```
seurat_object seurat object

cluster_id ID used for finding clusters of cells

anansnake_inf_dir

influence directory generated by anansnake

return_df return both the seurat object and a dataframe with influence scores as a list
```

Value

seurat object with the influence scores addes as an assay

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
infdir <- system.file("extdata","influence",package = 'AnanseSeurat')
sce_small <- import_seurat_scANANSE(sce_small, anansnake_inf_dir = infdir)</pre>
```

Maelstrom_Motif2TF

Maelstrom_Motif2TF

Description

create motif-factor links & export tables for printing motif score alongside its binding factor

Usage

```
Maelstrom_Motif2TF(
   seurat_object,
   mot_mat = NULL,
   m2f_df = NULL,
   cluster_id = "seurat_clusters",
   maelstrom_dir = "./maelstrom/",
   combine_motifs = "means",
   RNA_expression_assay = "RNA",
   RNA_expression_slot = "data",
   expr_tresh = 10,
   cor_tresh = 0.3,
   curated_motifs = FALSE,
   cor_method = "pearson",
   return_df = FALSE
)
```

Arguments

seurat_object object mot_mat motif_matrix, if not provided extracts one from the single cell object from the maelstrom assay m2f_df motif to factor dataframe, if not provided extracts from the maelstrom directory cluster_id ID used for finding clusters of cells maelstrom_dir directory where the GimmeMotifs m2f table is stored combine_motifs means (take mean multiple motifscores), max_var (take motif with highest variance), or max_cor (take motif with best correlation to gene expression) RNA_expression_assay Seurat assay containing factor expression info RNA_expression_slot slot within assay used for calculating mean factor expression per cluster minimum sum of gene counts over all cells in RNA_expression_assay to filter expr_tresh genes by cor_tresh minimum value of to filter the cor() output by curated_motifs use only curated motifs (T), or all motifs in the database (F) cor_method specify one of the cor() methods return_df return both the seurat object and two dataframes with maelstrom scores and

expression values as a list

per_cluster_df

Value

seurat object with two assays added, MotifTFcor for TFs with positive correlation to the linked motif, and MotifTFanticor for TFs with positive correlation to the linked motif

Examples

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
maelstrom_dir_path <- system.file("extdata","maelstrom",package = 'AnanseSeurat')
sce_small <- Maelstrom_Motif2TF(sce_small, maelstrom_dir = maelstrom_dir_path)</pre>
```

per_cluster_df

per_cluster_df

Description

generate a table of the assay score averages per cluster identifier cell

Usage

```
per_cluster_df(
   seurat_object,
   assay = "influence",
   cluster_id = "seurat_clusters"
)
```

Arguments

seurat_object seurat object

assay assay containing influence or motif scores generated from cluster pseudobulk

cluster_id ID used for finding clusters of cells

Value

dataframe with assay scores, concatinating cells from each per cluster

```
sce_small <- readRDS(system.file("extdata","sce_small.Rds",package = 'AnanseSeurat'))
df <- per_cluster_df(sce_small)</pre>
```

Index

```
config_scananse, 2

DEGS_scananse, 3

export_ATAC_maelstrom, 4
export_ATAC_scananse, 5
export_CPM_scananse, 5

Factor_Motif_Plot, 6

import_seurat_maelstrom, 7
import_seurat_scananse, 8

Maelstrom_Motif2TF, 9

per_cluster_df, 10
```