

Package ‘tlsR’

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Title Detection and Spatial Analysis of Tertiary Lymphoid Structures

Version 0.1.2

Description Fast, reproducible detection and quantitative analysis of tertiary lymphoid structures (TLS) in multiplexed tissue imaging. Implements Independent Component Analysis Trace (ICAT) index, local Ripley's K scanning, automated K Nearest Neighbor (KNN)-based TLS detection, and T-cell clusters identification as described in Amiryousefi et al. (2025) <[doi:10.1101/2025.09.21.677465](https://doi.org/10.1101/2025.09.21.677465)>.

Note TLS, ICAT, KNN are correct technical acronyms.

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Encoding UTF-8

LazyData true

RoxygenNote 7.3.2

Imports fastICA, spatstat.geom, spatstat.explore, dbscan, RANN,
grDevices, graphics, stats, utils

Suggests testthat (>= 3.0.0)

Config/testthat.edition 3

Depends R (>= 3.5)

NeedsCompilation no

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calc_icat*Calculate ICAT (Immune Cell Arrangement Trace) Index*

Description

Quantifies linear/organized arrangement of cells within a TLS using FastICA.

Arguments

- patientID* Character. Name of the sample in *ldata*.
- tlsID* Numeric/integer. TLS identifier.
- ldata* Named list of data frames (optional; defaults to global *ldata*).

Value

Numeric ICAT value.

Examples

```
data(toy_ldata)
ldata <- detect_TLS("ToySample", k = 30, ldata = toy_ldata) # First detect TLS
if (max(ldata[["ToySample"]]$tls_id_knn) > 0) {
  icat <- calc_icat("ToySample", tlsID = 1, ldata = ldata)
  icat
}
```

detect_tic*Detect Tumor-Infiltrating T-cell Clusters (TIC)*

Description

Uses HDBSCAN approximation on T cells outside TLS regions to find TIC.

Arguments

- sample* Character. Sample name.
- ldata* Optional list.

Value

Modified data frame for the sample (invisibly).

Examples

```
data(toy_pdata)
ldata <- detect_TLS("ToySample", k = 30, ldata = toy_pdata) # Need TLS first
ldata <- detect_tic("ToySample", ldata = ldata)
table(ldata[["ToySample"]]$tcell_cluster_hdbscan)
```

detect_TLS

Detect Tertiary Lymphoid Structures using a KNN-density approach

Description

This function identifies TLS candidates germinated with B cells (BIC) by: 1. Finding regions of high local B-cell density (KNN-based). 2. Requiring a minimum number of neighbouring T cells (T-B cell co-localisation). 3. Applying sensible size and shape filters.

Arguments

LSP	Character. Sample name in the global ldata list.
k	Integer. Number of nearest neighbours to consider (default 30 - works very well on 0.325 um/px imaging).
bcell_density_threshold	Numeric. Minimum average 1/k-distance for B cells to be considered "dense" (default 15 um).
min_B_cells	Integer. Minimum number of B cells in a candidate TLS (default 50).
min_T_cells_nearby	Integer. Minimum T cells within 50 um of the B-cell cluster centre (default 30).
max_distance_T	Numeric. Radius (um) to search for surrounding T cells (default 50).
ldata	Optional. Named list of data frames. If NULL, uses global ldata.

Value

The original data frame with two new columns:

tls_id_knn	0 = non-TLS, positive integer = TLS cluster ID
tls_center_x, tls_center_y	Coordinates of detected TLS centres (only for TLS cells)

Examples

```
data(toy_pdata)
ldata <- detect_TLS("ToySample", k = 30, ldata = toy_pdata)
table(ldata[["ToySample"]]$tls_id_knn)
plot(ldata[["ToySample"]]$x, ldata[["ToySample"]]$y,
      col = ifelse(ldata[["ToySample"]]$tls_id_knn > 0, "red", "gray"),
      pch = 19, cex = 0.5, main = "Detected TLS in toy data")
```

<code>scan_clustering</code>	<i>Scan Tissue for Local Immune Cell Clustering (K-integral)</i>
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Description

Sliding-window Centerel L-Function (CLF) version of the Ripley's K analysis with whole tissue pseudo-plots.

Arguments

<code>ws</code>	Window size in microns.
<code>sample</code>	Character. Sample name in <code>ldata</code> .
<code>phenotype</code>	One of "T cells", "B cells", or "Both".
<code>plot</code>	Logical. Show diagnostic plot?
<code>creep</code>	Integer. Grid density factor.
<code>ldata</code>	Optional list (defaults to global <code>ldata</code>).

Value

List of `Lest` objects for significant windows.

Examples

```
data(toy_ldata)
# This one may produce plots and take ~10 sec
models <- scan_clustering(ws = 500, sample = "ToySample",
                           phenotype = "B cells", plot = FALSE, ldata = toy_ldata)
length(models)
```

<code>toy_ldata</code>	<i>Toy multiplexed imaging data for examples</i>
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Description

A small synthetic dataset mimicking multiplexed tissue imaging data. It contains one sample named "ToySample" with columns required by `tlsR` functions.

Usage

`toy_ldata`

Format

A named list with one element:

ToySample A data frame with columns:

- x: x-coordinate in microns
- y: y-coordinate in microns
- coarse_phen_vec: Cell phenotype ("B cells", "T cells", or "Other")
- row_index: Integer row index (1 to nrow)
- cflag: Integer flag column (0 for all cells)

Examples

```
data(toy_ldata)
str(toy_ldata[["ToySample"]])
plot(toy_ldata[["ToySample"]]$x, toy_ldata[["ToySample"]]$y,
     col = as.factor(toy_ldata[["ToySample"]]$coarse_phen_vec),
     pch = 19, cex = 0.5, main = "Toy sample cells")
```

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