AAM-IPL-Wk-5-SVM-ProjectName-Full-Code-V4

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1 AAM-IPL Week-4 SVM - Breast Cancer Diagnosis

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Project Implementation Details: As published in the project announcement in AAM-IPL Online Classroom

This project implements a Support Vector Machine (SVM) classifier to predict the class of breast cancer from the provided dataset. The dataset is loaded - load_breast_cancer()- from sklearn. The breast cancer dataset consists of 569 samples, each representing a patient with a set of features.

The dataset is used for training a SVM classifier and evaluate its performance using various metrics such as accuracy, precision, recall, F1-score. Additionally, various other classifiers, such as Logistic Regression, Naive Bayes, and K-Nearest Neighbors are also implemented and evaluated for comparison.

AAM-IPL of GPREC is brought to you by Brillium Technologies.

```
[1]: # Importing necessary libraries for breast cancer SVM classifier
     from sklearn import datasets
     from sklearn.model_selection import train_test_split
     from sklearn.preprocessing import StandardScaler
     from sklearn.metrics import accuracy_score
     from sklearn.svm import SVC
     import pandas as pd # for dataframe manipulation
     from pandas.plotting import parallel_coordinates # for parallel coordinates_
      ⇒plot of breast cancer data set
     import matplotlib.pyplot as plt # for plotting graphs
     from sklearn.manifold import TSNE # for t-SNE plot
     from sklearn.ensemble import RandomForestClassifier
     from sklearn.metrics import auc, confusion_matrix, precision_score, __
      recall_score, roc_curve
     from sklearn.model_selection import cross_val_score
     from sklearn.metrics import f1 score
     from sklearn.linear_model import LogisticRegression
     from sklearn.naive_bayes import GaussianNB
     from sklearn.neighbors import KNeighborsClassifier
```

```
from sklearn.tree import DecisionTreeClassifier
     import seaborn as sns
     from collections import Counter
     import numpy as np
     from sklearn.metrics import accuracy_score, f1_score, roc_curve, auc, u
      →precision_recall_curve, confusion_matrix, ConfusionMatrixDisplay
     from matplotlib.colors import ListedColormap
     import seaborn as sns
     from matplotlib.offsetbox import TextArea, DrawingArea, OffsetImage, u
      \hookrightarrowAnnotationBbox
     import matplotlib.image as mpimg
     from matplotlib.transforms import Affine2D
     from matplotlib.offsetbox import OffsetImage, AnnotationBbox
     from scipy.ndimage import rotate
     from PIL import Image
[2]: # Create virtual environment
     # !python -m venv venv
     # Activate virtual environment
     # !source venv/bin/activate
[2]: # Define name, email, plot watermarks etc.
     name = "Venkateswar Reddy Melachervu"
     email = "venkat.reddy.gf@gprec.ac.in"
     # Load your watermark image (assuming it's in the same directory)
     aam_ipl_wama_image = plt.imread('AAM-IPL-Watermark-for-Plots.png')
     # Function to add a diagonal watermark
     def add_aam_ipl_wama_revised(ax, watermark_image, zoom, alpha=0.3,_
      →rotation_angle=45):
         Adds an image watermark diagonally to the provided axis.
         Parameters:
         - ax: The axis to add the watermark to.
         - watermark_image: The image to use as the watermark.
         - alpha: The transparency level for the watermark.
         - zoom: The scale of the watermark image.
         - rotation_angle: The angle to rotate the watermark (default is 45 degrees).
         imagebox = OffsetImage(watermark_image, alpha=alpha, zoom=zoom)
         # Create a transformation for diagonal placement
         trans_data = Affine2D().rotate_deg(rotation_angle) + ax.transData
         ab = AnnotationBbox(imagebox, (0.5, 0.5), frameon=False, xycoords='axes⊔
      fraction', boxcoords="axes fraction", pad=0, transform=trans_data)
```

```
# Add watermark image to the axis
   ax.add_artist(ab)
# Load the breast cancer dataset from sklearn
bcancer = datasets.load_breast_cancer()
X = bcancer.data
Y = bcancer.target
# Print the description/meta data
print(bcancer.DESCR)
# Scale the data using standard scaler
scaler = StandardScaler()
standard_scaled_X = scaler.fit_transform(X)
# Display column/feature names in the breast cancer dataset
df = pd.DataFrame(data=X, columns=bcancer.feature_names)
df['target'] = Y
# Set display option to show all columns
pd.set_option('display.max_columns', None)
pd.set option('display.width', 1000)
print('Breast Cancer Data Set Details:')
print(f'\tComprises of {df.shape[0]} data sample vectors')
print(f'\tEach vector is of {df.shape[1]-1} features/dimensions and a target ∪
 ⇔scalar response.')
# Count the data samples for each target value
target_counts = df['target'].value_counts()
# Print the counts separately
count 0 = target counts[0]
count_1 = target_counts[1]
print(f"\tNumber of samples with target value 1 - \"Benign/Non-cancerous⊔
 →Tumors\": {count_1}")
print(f"\tNumber of samples with target value 0 - \"Malignant/Cancerous⊔
 →Tumors\": {count_0}")
print(f"\tTotal number of data samples - \"Breast Cancer\" data set: {count_0}")
# Capitalize the first letter of each word in the feature names
df.columns = [col.title() for col in df.columns]
# Filter out the 'Target' column after capitalizing
feature_columns = [col for col in df.columns if col != 'Target']
```

```
# Convert to list
column_names = feature_columns
print(f"\tThe feature names are:")
print(f'\t{column_names}')
```

.. _breast_cancer_dataset:

Breast cancer wisconsin (diagnostic) dataset

Data Set Characteristics:

:Number of Instances: 569

:Number of Attributes: 30 numeric, predictive attributes and the class

:Attribute Information:

- radius (mean of distances from center to points on the perimeter)
- texture (standard deviation of gray-scale values)
- perimeter
- area
- smoothness (local variation in radius lengths)
- compactness (perimeter^2 / area 1.0)
- concavity (severity of concave portions of the contour)
- concave points (number of concave portions of the contour)
- symmetry
- fractal dimension ("coastline approximation" 1)

The mean, standard error, and "worst" or largest (mean of the three worst/largest values) of these features were computed for each image, resulting in 30 features. For instance, field 0 is Mean Radius, field 10 is Radius SE, field 20 is Worst Radius.

- class:
 - WDBC-Malignant
 - WDBC-Benign

:Summary Statistics:

Min	Max
=====	=====
6.981	28.11
9.71	39.28
43.79	188.5
143.5	2501.0
0.053	0.163
	6.981 9.71 43.79 143.5

_____ ____

```
compactness (mean):
                                   0.019 0.345
concavity (mean):
                                   0.0
                                         0.427
concave points (mean):
                                   0.0
                                         0.201
symmetry (mean):
                                   0.106 0.304
fractal dimension (mean):
                                   0.05
                                         0.097
radius (standard error):
                                   0.112 2.873
texture (standard error):
                                   0.36
                                         4.885
perimeter (standard error):
                                   0.757 21.98
area (standard error):
                                   6.802 542.2
smoothness (standard error):
                                   0.002 0.031
compactness (standard error):
                                   0.002 0.135
concavity (standard error):
                                         0.396
                                   0.0
concave points (standard error):
                                   0.0
                                          0.053
symmetry (standard error):
                                   0.008 0.079
fractal dimension (standard error):
                                   0.001 0.03
radius (worst):
                                   7.93
                                         36.04
texture (worst):
                                   12.02 49.54
perimeter (worst):
                                   50.41 251.2
area (worst):
                                   185.2 4254.0
smoothness (worst):
                                   0.071 0.223
compactness (worst):
                                   0.027 1.058
concavity (worst):
                                   0.0
                                          1.252
concave points (worst):
                                   0.0
                                         0.291
symmetry (worst):
                                   0.156 0.664
fractal dimension (worst):
                                   0.055 0.208
```

:Missing Attribute Values: None

:Class Distribution: 212 - Malignant, 357 - Benign

:Creator: Dr. William H. Wolberg, W. Nick Street, Olvi L. Mangasarian

:Donor: Nick Street

:Date: November, 1995

This is a copy of UCI ML Breast Cancer Wisconsin (Diagnostic) datasets. https://goo.gl/U2Uwz2

Features are computed from a digitized image of a fine needle aspirate (FNA) of a breast mass. They describe characteristics of the cell nuclei present in the image.

Separating plane described above was obtained using Multisurface Method-Tree (MSM-T) [K. P. Bennett, "Decision Tree Construction Via Linear Programming." Proceedings of the 4th Midwest Artificial Intelligence and Cognitive Science Society,

pp. 97-101, 1992], a classification method which uses linear programming to construct a decision tree. Relevant features were selected using an exhaustive search in the space of 1-4 features and 1-3 separating planes.

The actual linear program used to obtain the separating plane in the 3-dimensional space is that described in:
[K. P. Bennett and O. L. Mangasarian: "Robust Linear Programming Discrimination of Two Linearly Inseparable Sets", Optimization Methods and Software 1, 1992, 23-34].

This database is also available through the UW CS ftp server:

ftp ftp.cs.wisc.edu
cd math-prog/cpo-dataset/machine-learn/WDBC/

- .. dropdown:: References
 - W.N. Street, W.H. Wolberg and O.L. Mangasarian. Nuclear feature extraction for breast tumor diagnosis. IS&T/SPIE 1993 International Symposium on Electronic Imaging: Science and Technology, volume 1905, pages 861-870, San Jose, CA, 1993.
 - O.L. Mangasarian, W.N. Street and W.H. Wolberg. Breast cancer diagnosis and prognosis via linear programming. Operations Research, 43(4), pages 570-577, July-August 1995.
- W.H. Wolberg, W.N. Street, and O.L. Mangasarian. Machine learning techniques to diagnose breast cancer from fine-needle aspirates. Cancer Letters 77 (1994)

163-171.

Breast Cancer Data Set Details:

Comprises of 569 data sample vectors

Each vector is of 30 features/dimensions and a target scalar response. Number of samples with target value 1 - "Benign/Non-cancerous Tumors":

357

212

Number of samples with target value 0 - "Malignant/Cancerous Tumors":

Total number of data samples - "Breast Cancer" data set: 212
The feature names are:

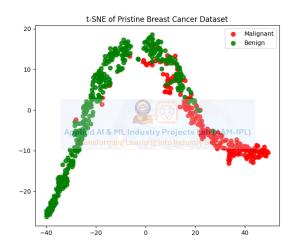
['Mean Radius', 'Mean Texture', 'Mean Perimeter', 'Mean Area', 'Mean Smoothness', 'Mean Compactness', 'Mean Concavity', 'Mean Concave Points', 'Mean Symmetry', 'Mean Fractal Dimension', 'Radius Error', 'Texture Error', 'Perimeter Error', 'Area Error', 'Smoothness Error', 'Compactness Error', 'Concavity Error', 'Concave Points Error', 'Symmetry Error', 'Fractal Dimension Error', 'Worst Radius', 'Worst Texture', 'Worst Perimeter', 'Worst Area', 'Worst Smoothness', 'Worst Compactness', 'Worst Concavity', 'Worst Concave Points', 'Worst Symmetry', 'Worst Fractal Dimension']

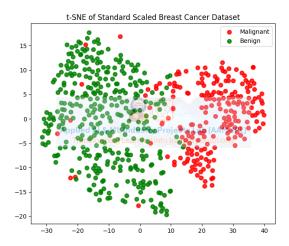
```
[3]: # Display the first row of the data set
     print('First row of standard scaled data sample in the Breast Cancer data set⊔

is:')

     first sample = df.iloc[0]
     for idx, (feature, value) in enumerate(first_sample.items(), start=1):
         print(f"{idx}. {feature:<30} {value:.6f}")</pre>
    First row of standard scaled data sample in the Breast Cancer data set is:
                                      17.990000
    1. Mean Radius
    2. Mean Texture
                                      10.380000
    3. Mean Perimeter
                                      122.800000
    4. Mean Area
                                      1001.000000
    5. Mean Smoothness
                                      0.118400
    6. Mean Compactness
                                      0.277600
                                      0.300100
    7. Mean Concavity
    8. Mean Concave Points
                                     0.147100
    9. Mean Symmetry
                                     0.241900
    10. Mean Fractal Dimension
                                      0.078710
    11. Radius Error
                                       1.095000
    12. Texture Error
                                      0.905300
    13. Perimeter Error
                                       8.589000
    14. Area Error
                                       153.400000
    15. Smoothness Error
                                       0.006399
    16. Compactness Error
                                       0.049040
    17. Concavity Error
                                       0.053730
    18. Concave Points Error
                                       0.015870
    19. Symmetry Error
                                       0.030030
    20. Fractal Dimension Error
                                       0.006193
    21. Worst Radius
                                       25.380000
    22. Worst Texture
                                      17.330000
    23. Worst Perimeter
                                      184.600000
    24. Worst Area
                                      2019.000000
    25. Worst Smoothness
                                      0.162200
    26. Worst Compactness
                                     0.665600
    27. Worst Concavity
                                      0.711900
    28. Worst Concave Points
                                     0.265400
    29. Worst Symmetry
                                       0.460100
    30. Worst Fractal Dimension
                                       0.118900
    31. Target
                                       0.000000
[5]: # Lets plot t-SNE - t-distributed stochastic neighbor embedding plot of the
     ⇒breast cancer data set - visual dimensionality reduction technique
     import os
     logical_cores = os.cpu_count()
     os.environ['LOKY_MAX_CPU_COUNT'] = str(logical_cores)
     # Perform t-SNE on pristine data
```

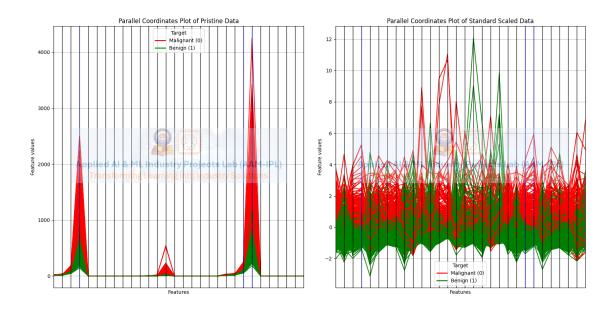
```
tsne_pristine = TSNE(n_components=2, random_state=42)
X_tsne_pristine = tsne_pristine.fit_transform(X)
# Perform t-SNE on standard scaled data
tsne_scaled = TSNE(n_components=2, random_state=42)
X_tsne_scaled = tsne_scaled.fit_transform(standard_scaled_X)
# Print the derived X and Y values
print("t-SNE on Pristine Data (first 5 points):")
print(X_tsne_pristine[:5])
print("\nt-SNE on Standard Scaled Data (first 5 points):")
print(X_tsne_scaled[:5])
# Define colors and line width for the plot
colors = ['red', 'green']
lw = 2
# Plot t-SNE results
fig, axes = plt.subplots(1, 2, figsize=(16, 6))
# t-SNE on pristine data
axes[0].set title('t-SNE of Pristine Breast Cancer Dataset')
for color, i, target_name in zip(colors, [0, 1], bcancer.target_names):
    axes[0].scatter(X tsne pristine[Y == i, 0], X tsne pristine[Y == i, 1],
 Golor=color, alpha=.8, lw=lw, label=target_name.capitalize())
axes[0].legend(loc='best', shadow=False, scatterpoints=1)
# t-SNE on standard scaled data
axes[1].set_title('t-SNE of Standard Scaled Breast Cancer Dataset')
for color, i, target_name in zip(colors, [0, 1], bcancer.target_names):
    axes[1].scatter(X_tsne_scaled[Y == i, 0], X_tsne_scaled[Y == i, 1],_u
 Golor=color, alpha=.8, lw=lw, label=target_name.capitalize())
axes[1].legend(loc='best', shadow=False, scatterpoints=1)
# Add watermark with the loaded image
add_aam_ipl_wama_revised(axes[0], aam_ipl_wama_image, 0.3)
add_aam_ipl_wama_revised(axes[1], aam_ipl_wama_image, 0.3)
plt.show()
t-SNE on Pristine Data (first 5 points):
[[ 42.337463 -12.640097 ]
 [ 42.50024
             -9.92803 ]
 [ 37.894382 -10.039126 ]
 [-17.654438 1.4511237]
 [ 36.764866 -7.9306464]]
```





```
[6]: # Parallel coordinates plot of the breast cancer data set for pristine and
      ⇔standard scaled data
     # Create DataFrames from the data
     df_pristine = pd.DataFrame(data=X, columns=bcancer.feature_names)
     df_pristine['target'] = Y
     df_scaled = pd.DataFrame(data=standard_scaled_X, columns=bcancer.feature_names)
     df scaled['target'] = Y
     # Convert target to string for color coding
     df_pristine['target'] = df_pristine['target'].astype(str)
     df_scaled['target'] = df_scaled['target'].astype(str)
     # Define colors for each class explicitly
     color_dict = {'0': 'red', '1': 'green'}
     # Create a colormap from the color dictionary
     cmap = plt.cm.colors.ListedColormap([color_dict['0'], color_dict['1']])
     # Identify features with mean value above 100 - Exclude the 'target' column
     mean_values = df_pristine.drop('target', axis=1).mean()
     features_above_100 = mean_values[mean_values > 100].index
     # Create subplots for parallel coordinates
```

```
fig, axes = plt.subplots(1, 2, figsize=(16, 8))
# Parallel coordinates plot for pristine data
parallel_coordinates(df_pristine, 'target', colormap=cmap, ax=axes[0])
axes[0].set_title('Parallel Coordinates Plot of Pristine Data')
axes[0].set_xlabel('Features')
axes[0].set_ylabel('Feature values')
# Highlight the dominant features
for feature in features_above_100:
    axes[0].axvline(df pristine.columns.get loc(feature), color='blue',
 →linestyle='--', linewidth=1)
# Parallel coordinates plot for standard scaled data
parallel_coordinates(df_scaled, 'target', colormap=cmap, ax=axes[1])
axes[1].set_title('Parallel Coordinates Plot of Standard Scaled Data')
axes[1].set xlabel('Features')
axes[1].set_ylabel('Feature values')
# Highlight the dominant features
for feature in features above 100:
    axes[1].axvline(df_scaled.columns.get_loc(feature), color='blue',_
 →linestyle='--', linewidth=1)
# Add custom legend
handles = [plt.Line2D([0], [0], color=color_dict['0'], lw=2), plt.Line2D([0], __
\hookrightarrow [0], color=color dict['1'], lw=2)]
axes[0].legend(handles, ['Malignant (0)', 'Benign (1)'], title='Target')
axes[1].legend(handles, ['Malignant (0)', 'Benign (1)'], title='Target')
# Remove x-axis labels
axes[0].set_xticks([])
axes[1].set_xticks([])
# Add watermark to both plots
add_aam_ipl_wama_revised(axes[0], aam_ipl_wama_image, 0.4)
add_aam_ipl_wama_revised(axes[1], aam_ipl_wama_image, 0.4)
plt.tight_layout()
plt.show()
```



```
[7]: # Plot curves for all classifiers - accuracy, confusion matrix, ROC curve, PR
     ⇔curve, feature importance, F1 score
     # Define the formulae as strings
     precision_formula = "Precision = True Positives / (True Positives + False ∪
      →Positives)"
     recall_formula = "Recall = True Positives / (True Positives + False Negatives)"
     f1_score_formula = "F1 Score = 2 * (Precision * Recall) / (Precision + Recall)"
     # Print the formulae
     print(precision_formula)
     print(recall_formula)
     print(f1_score_formula, '\n')
     # Define a dictionary of models
     models = {
         'Linear SVM': SVC(kernel='linear', random_state=0, probability=True),
         'RBF Kernel SVM': SVC(kernel='rbf', random_state=0, probability=True),
         'Poly Kernel SVM': SVC(kernel='poly', degree=3, random_state=0,_
      ⇔probability=True),
         'Sigmoid Kernel SVM': SVC(kernel='sigmoid', random_state=0,__
      ⇒probability=True),
         'Logistic Regression': LogisticRegression(max_iter=10000, random_state=0),
         'Decision Tree': DecisionTreeClassifier(criterion='entropy',
      →random state=0),
         'K-Nearest Neighbors': KNeighborsClassifier(n_neighbors=5,_
      →metric='minkowski', p=2),
         'Naive Bayes': GaussianNB(),
```

```
'Random Forest': RandomForestClassifier(n_estimators=10,__
 ⇔criterion='entropy', random_state=0)
}
# Initialize plot
fig, axs = plt.subplots(len(models), 6, figsize=(24, len(models) * 5))
fig.subplots_adjust(hspace=0.5)
# Split the data
X train, X test, Y train, Y test = train test_split(standard_scaled_X, Y, 
→test_size=0.2, random_state=0)
# Loop through models
for idx, (name, model) in enumerate(models.items()):
   # Train model
   model.fit(X_train, Y_train)
   # Predict results
   Y_pred = model.predict(X_test)
   Y_proba = model.predict_proba(X_test)[:, 1] if hasattr(model,_

¬'predict_proba') else None

   accuracy = accuracy_score(Y_test, Y_pred)
   print(f"{name} Accuracy Score: {100 * accuracy:.2f}%")
   # Compute metrics
   cm = confusion_matrix(Y_test, Y_pred)
   fpr, tpr, _ = roc_curve(Y_test, Y_proba) if Y_proba is not None else ([], _
 →[], [])
   roc_auc = auc(fpr, tpr) if Y_proba is not None else None
   precision, recall, _ = precision_recall_curve(Y_test, Y_proba) if Y_proba⊔
 →is not None else ([], [], [])
   f1 = f1_score(Y_test, Y_pred)
    # Plot Accuracy Score curve
   if Y_proba is not None:
        thresholds = np.linspace(0, 1, 100)
        accuracy_scores = [accuracy_score(Y_test, (Y_proba > threshold).
 →astype(int)) for threshold in thresholds]
        axs[idx, 5].plot(thresholds, accuracy_scores, color='green', lw=2)
        axs[idx, 5].set_xlim([0.0, 1.0])
        axs[idx, 5].set_ylim([0.0, 1.05])
        axs[idx, 5].set_xlabel('Decision Threshold')
        axs[idx, 5].set_ylabel('Accuracy Score')
        axs[idx, 5].set_title(f'{name} Accuracy Score Curve')
```

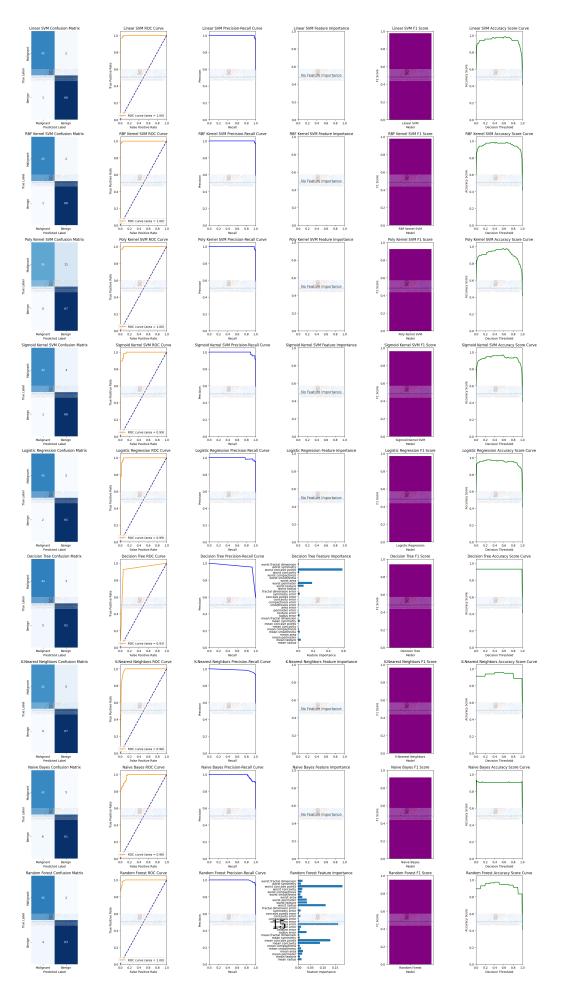
```
axs[idx, 5].text(0.5, 0.5, 'No Accuracy Curve',
→horizontalalignment='center', verticalalignment='center', fontsize=12)
      axs[idx, 5].set_title(f'{name} Accuracy Score Curve')
  # Plot confusion matrix
  sns.heatmap(cm, annot=True, fmt='d', cmap='Blues', ax=axs[idx, 0],
⇔cbar=False, xticklabels=['Malignant', 'Benign'], yticklabels=['Malignant', u
axs[idx, 0].set_xlabel('Predicted Label')
  axs[idx, 0].set_ylabel('True Label')
  axs[idx, 0].set_title(f'{name} Confusion Matrix')
  # Plot ROC curve
  if Y_proba is not None:
      axs[idx, 1].plot(fpr, tpr, color='darkorange', lw=2, label=f'ROC curve_u
\Rightarrow(area = {roc_auc:.2f})')
      axs[idx, 1].plot([0, 1], [0, 1], color='navy', lw=2, linestyle='--')
      axs[idx, 1].set_xlim([0.0, 1.0])
      axs[idx, 1].set_ylim([0.0, 1.05])
      axs[idx, 1].set_xlabel('False Positive Rate')
      axs[idx, 1].set_ylabel('True Positive Rate')
      axs[idx, 1].set_title(f'{name} ROC Curve')
      axs[idx, 1].legend(loc='lower right')
      axs[idx, 1].text(0.5, 0.5, 'No ROC Curve', __
→horizontalalignment='center', verticalalignment='center', fontsize=12)
      axs[idx, 1].set_title(f'{name} ROC Curve')
  # Plot Precision-Recall curve
  if Y_proba is not None:
      axs[idx, 2].plot(recall, precision, color='blue', lw=2)
      axs[idx, 2].set_xlim([0.0, 1.0])
      axs[idx, 2].set_ylim([0.0, 1.05])
      axs[idx, 2].set_xlabel('Recall')
      axs[idx, 2].set ylabel('Precision')
      axs[idx, 2].set_title(f'{name} Precision-Recall Curve')
  else:
      axs[idx, 2].text(0.5, 0.5, 'No PR Curve', horizontalalignment='center', u
⇔verticalalignment='center', fontsize=12)
      axs[idx, 2].set_title(f'{name} Precision-Recall Curve')
  # Plot feature importance for models that have it
  if hasattr(model, 'feature_importances_'):
      importances = model.feature_importances_
      axs[idx, 3].barh(range(len(importances)), importances, align='center')
      axs[idx, 3].set_yticks(range(len(importances)))
      axs[idx, 3].set_yticklabels(bcancer.feature_names)
```

```
axs[idx, 3].set_xlabel('Feature Importance')
        axs[idx, 3].set_title(f'{name} Feature Importance')
    else:
        axs[idx, 3].text(0.5, 0.5, 'No Feature Importance',
  whorizontalalignment='center', verticalalignment='center', fontsize=12)
        axs[idx, 3].set title(f'{name} Feature Importance')
    # Plot F1 Score
    axs[idx, 4].bar(name, f1, color='purple')
    axs[idx, 4].set_ylim([0, 1])
    axs[idx, 4].set_xlabel('Model')
    axs[idx, 4].set_ylabel('F1 Score')
    axs[idx, 4].set_title(f'{name} F1 Score')
   # Add watermark to all subplots
    for ax in axs[idx, :]:
        add_aam_ipl_wama_revised(ax, aam_ipl_wama_image, 0.15)
plt.tight_layout()
plt.show()
Precision = True Positives / (True Positives + False Positives)
Recall = True Positives / (True Positives + False Negatives)
F1 Score = 2 * (Precision * Recall) / (Precision + Recall)
Linear SVM Accuracy Score: 97.37%
```

Linear SVM Accuracy Score: 97.37%
RBF Kernel SVM Accuracy Score: 97.37%
Poly Kernel SVM Accuracy Score: 90.35%
Sigmoid Kernel SVM Accuracy Score: 95.61%
Logistic Regression Accuracy Score: 96.49%
Decision Tree Accuracy Score: 92.98%

K-Nearest Neighbors Accuracy Score: 95.61%

Naive Bayes Accuracy Score: 90.35% Random Forest Accuracy Score: 94.74%



```
[8]: # Plot relative performance plots of all models - accuracy, ROC curve, PR
     ⇔curve, F1 score
     # Define a dictionary of models
     models = {
         'Linear SVM': SVC(kernel='linear', random_state=0, probability=True),
         'RBF Kernel SVM': SVC(kernel='rbf', random_state=0, probability=True),
         'Poly Kernel SVM': SVC(kernel='poly', degree=3, random_state=0,_
      →probability=True),
         'Sigmoid Kernel SVM': SVC(kernel='sigmoid', random_state=0,_
      ⇔probability=True),
         'Logistic Regression': LogisticRegression(max_iter=10000, random_state=0),
         'Decision Tree': DecisionTreeClassifier(criterion='entropy',
      →random_state=0),
         'K-Nearest Neighbors': KNeighborsClassifier(n_neighbors=5,_

→metric='minkowski', p=2),
         'Naive Bayes': GaussianNB(),
         'Random Forest': RandomForestClassifier(n_estimators=10,__
     ⇔criterion='entropy', random_state=0)
     }
     # Split the data
     X_train, X_test, Y_train, Y_test = train_test_split(standard_scaled_X, Y,__
     →test_size=0.2, random_state=0)
     # Initialize dictionaries to store metrics
     accuracy scores = {}
     f1_scores = {}
     roc_data = {}
     prc_data = {}
     # Calculate metrics
     for name, model in models.items():
         model.fit(X_train, Y_train)
         Y_pred = model.predict(X_test)
         Y_proba = model.predict_proba(X_test)[:, 1] if hasattr(model,_

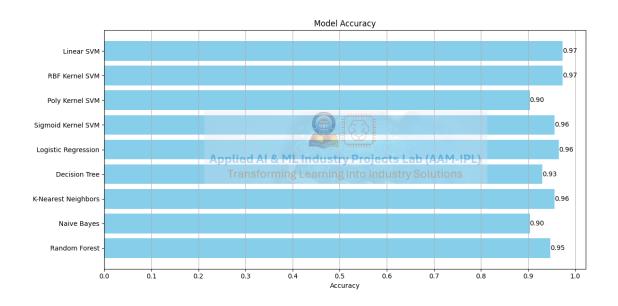
¬'predict_proba') else None

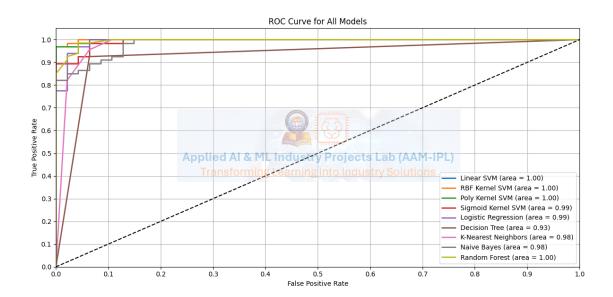
         # Accuracy
         accuracy = accuracy_score(Y_test, Y_pred)
         accuracy_scores[name] = accuracy
         # F1 Score
         f1 = f1_score(Y_test, Y_pred)
```

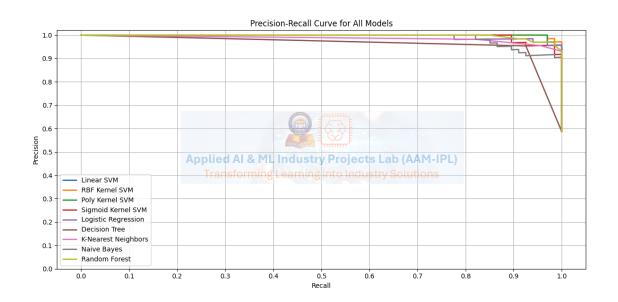
```
f1_scores[name] = f1
    # ROC Curve
    if Y_proba is not None:
        fpr, tpr, _ = roc_curve(Y_test, Y_proba)
        roc_auc = auc(fpr, tpr)
        roc_data[name] = (fpr, tpr, roc_auc)
        # Precision-Recall Curve
        precision, recall, _ = precision_recall_curve(Y_test, Y_proba)
        prc_data[name] = (precision, recall)
# Plot Accuracy Scores
plt.figure(figsize=(12, 6))
bars = plt.barh(list(accuracy_scores.keys()), list(accuracy_scores.values()),__
 ⇔color='skyblue')
plt.xlabel('Accuracy')
plt.title('Model Accuracy')
plt.gca().invert yaxis()
plt.grid(axis='x')
# Set x-axis ticks
plt.xticks(np.arange(0, 1.1, 0.1))
add_aam_ipl_wama_revised(plt.gca(), aam_ipl_wama_image, 0.4)
# Add value labels to each bar
for bar in bars:
    plt.text(bar.get_width(), bar.get_y() + bar.get_height()/2,
             f'{bar.get_width():.2f}', va='center', ha='left')
plt.tight_layout()
plt.show()
# Plot ROC Curves
plt.figure(figsize=(12, 6))
for name, (fpr, tpr, roc auc) in roc data.items():
    plt.plot(fpr, tpr, lw=2, label=f'{name} (area = {roc_auc:.2f})')
plt.plot([0, 1], [0, 1], 'k--')
plt.xlim([0.0, 1.0])
plt.ylim([0.0, 1.05])
plt.xlabel('False Positive Rate')
plt.ylabel('True Positive Rate')
plt.title('ROC Curve for All Models')
plt.legend(loc='lower right')
plt.grid()
# Set x-axis ticks
plt.xticks(np.arange(0, 1.1, 0.1))
# Set y-axis ticks
plt.yticks(np.arange(0, 1.1, 0.1))
```

```
add_aam_ipl_wama_revised(plt.gca(), aam_ipl_wama_image, 0.4)
plt.tight_layout()
plt.show()
# Plot Precision-Recall Curves
plt.figure(figsize=(12, 6))
for name, (precision, recall) in prc_data.items():
    plt.plot(recall, precision, lw=2, label=name)
plt.xlabel('Recall')
plt.ylabel('Precision')
plt.title('Precision-Recall Curve for All Models')
plt.legend(loc='best')
plt.grid()
# Set x-axis ticks
plt.xticks(np.arange(0, 1.1, 0.1))
# Set y-axis ticks
plt.yticks(np.arange(0, 1.1, 0.1))
add_aam_ipl_wama_revised(plt.gca(), aam_ipl_wama_image, 0.4)
plt.tight_layout()
plt.show()
# Plot F1 Scores
plt.figure(figsize=(12, 6))
bars = plt.barh(list(f1_scores.keys()), list(f1_scores.values()),__

¬color='orange')
plt.xlabel('F1 Score')
plt.title('Model F1 Scores')
plt.gca().invert_yaxis()
plt.grid(axis='x')
# Set x-axis ticks
plt.xticks(np.arange(0, 1.1, 0.1))
add_aam_ipl_wama_revised(plt.gca(), aam_ipl_wama_image, 0.4)
# Add value labels to each bar
for bar in bars:
    plt.text(bar.get_width(), bar.get_y() + bar.get_height()/2,
             f'{bar.get_width():.2f}', va='center', ha='left')
plt.tight_layout()
plt.show()
```









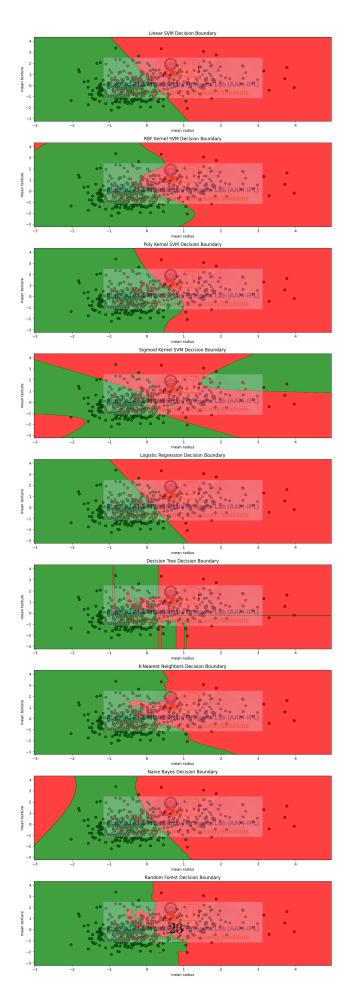
```
'K-Nearest Neighbors': KNeighborsClassifier(n_neighbors=5,__
 →metric='minkowski', p=2),
    'Naive Bayes': GaussianNB(),
    'Random Forest': RandomForestClassifier(n estimators=10,__
 ⇔criterion='entropy', random_state=0)
}
# Initialize plot
fig, axs = plt.subplots(len(models), 1, figsize=(12, len(models) * 4))
fig.subplots_adjust(hspace=0.5)
# Split the data
X_train, X_test, Y_train, Y_test = train_test_split(standard_scaled_X, Y,_
# Select two features for decision boundary plot
feature1, feature2 = 0, 1 # example feature indices, change as needed
# Loop through models
for idx, (name, model) in enumerate(models.items()):
    # Train and plot decision boundary on two features
   X_train_2d = X_train[:, [feature1, feature2]]
   X test 2d = X test[:, [feature1, feature2]]
   model_2d = model.__class__(**model.get_params()) # Create a new instance_
 ⇔with the same parameters
   model_2d.fit(X_train_2d, Y_train)
   X_set, y_set = X_train_2d, Y_train
   X1, X2 = np.meshgrid(np.arange(start=X_set[:, 0].min() - 1, stop=X_set[:, __
 0].max() + 1, step=0.01),
                        np.arange(start=X_set[:, 1].min() - 1, stop=X_set[:, __
 41].max() + 1, step=0.01))
   Z = model_2d.predict(np.array([X1.ravel(), X2.ravel()]).T).reshape(X1.shape)
    # Plot the decision boundary
   axs[idx].contourf(X1, X2, Z, alpha=0.75, cmap=ListedColormap(('red', L

¬'green')))
   axs[idx].scatter(X_set[:, 0], X_set[:, 1], c=y_set,__

¬cmap=ListedColormap(('red', 'green')), edgecolor='k')

   axs[idx].set xlim(X1.min(), X1.max())
   axs[idx].set_ylim(X2.min(), X2.max())
   axs[idx].set_xlabel(bcancer.feature_names[feature1])
   axs[idx].set_ylabel(bcancer.feature_names[feature2])
   axs[idx].set_title(f'{name} Decision Boundary')
    # Add watermark to the current subplot
```

```
add_aam_ipl_wama_revised(axs[idx], aam_ipl_wama_image, 0.4)
plt.tight_layout()
plt.show()
```



[11]: | jupyter nbconvert --to pdf AAM-IPL-Wk-5-SVM-ProjectName-Full-Code-V4.ipynb C:\Program Files\Python313\Scripts\jupyter-nbconvert.EXE__main__.py:4: DeprecationWarning: Parsing dates involving a day of month without a year specified is ambiguious and fails to parse leap day. The default behavior will change in Python 3.15 to either always raise an exception or to use a different default year (TBD). To avoid trouble, add a specific year to the input & format. See https://github.com/python/cpython/issues/70647. [NbConvertApp] Converting notebook AAM-IPL-Wk-5-SVM-ProjectName-Full-Code-V4.ipynb to pdf [NbConvertApp] Support files will be in AAM-IPL-Wk-5-SVM-ProjectName-Full-Code-V4_files\ [NbConvertApp] Making directory .\AAM-IPL-Wk-5-SVM-ProjectName-Full-Code-V4_files [NbConvertApp] Writing 106321 bytes to notebook.tex [NbConvertApp] Building PDF [NbConvertApp] Running xelatex 3 times: ['xelatex', 'notebook.tex', '-quiet'] [NbConvertApp] Running bibtex 1 time: ['bibtex', 'notebook'] [NbConvertApp] WARNING | b had problems, most likely because there were no citations

[NbConvertApp] Writing 3496329 bytes to AAM-IPL-Wk-5-SVM-ProjectName-Full-

[NbConvertApp] PDF successfully created

Code-V4.pdf