

# Package ‘algaeClassify’

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**Title** Determine Phytoplankton Functional Groups Based on Functional Traits

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**Description** Functions that facilitate the use of accepted taxonomic nomenclature, collection of functional trait data, and assignment of functional group classifications to phytoplankton species. Possible classifications include Morpho-functional group (MFG; Salmaso et al. 2015 <doi:10.1111/fw.b.12520>) and CSR (Reynolds 1988; Functional morphology and the adaptive strategies of phytoplankton. In C.D. Sandgren (ed). Growth and reproductive strategies of freshwater phytoplankton, 388-433. Cambridge University Press, New York). Versions 1.3.0 and later no longer include the `algae_search()` function for querying the algaebase online taxonomic database ([www.algaebase.org](http://www.algaebase.org)). Users are advised to verify taxonomic names directly using algaebase and cite the database in resulting publications. Note that none of the algaeClassify authors are affiliated with algaebase in any way. However, species lists can be checked against a variety of taxonomic databases using the geographic name resolution service (GNRS) via wrapper functions for the `taxize` package, with convenient output format and unlikely names for phytoplankton taxa removed. Currently accepted and outdated synonyms, and higher taxonomy, can be extracted for lists of species from the ITIS database using wrapper functions for the `ritis` package. The algaeClassify package is a product of the GEISHA (Global Evaluation of the Impacts of Storms on freshwater Habitat and Structure of phytoplankton Assemblages), funded by CESAB (Centre for Synthesis and Analysis of Biodiversity) and the USGS John Wesley Powell Center for Synthesis and Analysis, with data and other support provided by members of GLEON (Global Lake Ecology Observation Network). This software is preliminary or provisional and is subject to revision. It is being provided to meet the need for timely best science. The software has not received final approval by the U.S. Geological Survey (USGS). No warranty, expressed or implied, is made by the USGS or the U.S. Government as to the functionality of the software and related material nor shall the fact of release constitute any such warranty. The software is provided on the condition that neither the USGS nor the U.S. Government shall be held liable for any damages resulting from the authorized or unauthorized use of the software.

**Depends** R (>= 3.4.0)

**Imports** lubridate, stats, taxize, ritis

**License** GPL-2 | GPL-3

**Encoding** UTF-8

**LazyData** true

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**GithubRef** working

**GithubSHA1** f7188b9197ddcd69980141d69a92d5f4b341837c

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accum	<i>Split a dataframe column with binomial name into genus and species columns. Plots change in species richness over time, generates species accumulation curve, and compares SAC against simulated idealized curve assuming all unique taxa have equal probability of being sampled at any point in the time series. (author Dietmar Straile)</i>
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### Description

Split a dataframe column with binomial name into genus and species columns. Plots change in species richness over time, generates species accumulation curve, and compares SAC against simulated idealized curve assuming all unique taxa have equal probability of being sampled at any point in the time series. (author Dietmar Straile)

### Usage

```
accum(  
  b_data,  
  phyto_name = "phyto_name",  
  column = NA,  
  n = 100,  
  save.pdf = FALSE,  
  lakename = "",  
  datename = "date_dd_mm_yy",  
  dateformat = "%d-%m-%y"  
)
```

### Arguments

b_data	Name of data.frame object
phyto_name	Character string: field containing phytoplankton id (species, genus, etc.)
column	column name or number for field containing abundance (biomass,biovol, etc.). Can be NA if the dataset only contains a species list for each sampling date.
n	number of simulations for randomized ideal species accumulation curve
save.pdf	TRUE/FALSE- should plots be displayed or saved to a pdf?
lakename	optional character string for adding lake name to pdf output
datename	character string name of b_data field containing date
dateformat	character string: posix format for datename column

### Value

a two panel plot with trends in richness on top, and cumulative richness vs. simulated accumulation curve on bottom

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows
head(lakegeneva)

accum(b_data=lakegeneva,column='biovol_um3_ml',n=10,save.pdf=FALSE)
```

---

bestmatch	<i>fuzzy partial matching between a scientific name and a list of possible matches</i>
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---

**Description**

fuzzy partial matching between a scientific name and a list of possible matches

**Usage**

```
bestmatch(enteredName, possibleNames, maxErr = 3, trunc = TRUE)
```

**Arguments**

enteredName	Character string with name to check
possibleNames	Character vector of possible matches
maxErr	maximum number of different bits allowed for a partial match
trunc	TRUE/FALSE. if true and no match, retry with last three letters truncated

**Value**

a character string with the best match, or 'multiplePartialMatches'

**Examples**

```
possibleMatches=c('Viburnum edule','Viburnum acerifolia')
bestmatch(enteredName='Viburnum edulus',possibleNames=possibleMatches)
```

---

csrTraits	<i>Database of functional traits for MFG classification, derived from Rimet et al. 2019</i>
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---

**Description**

Database of functional traits for MFG classification, derived from Rimet et al. 2019

**Usage**

```
data(mfgTraits)
```

## Format

A data frame with columns:

**phyto\_name** binomial scientific name

**genus** genus name

**species** species name

**SAV** surface area:volume ratio

**MLD** maximum linear dimension (micrometers)

**MSV** product of SAV and MLD; unitless

**volume.um3** cell or colony biovolume

**surface.area.um2** biological unit (cell or colony) surface area accounting for mucilage

**Colonial** 1/0 indicates colonial growth form

**Number.of.cells.per.colony** literature-based average colony abundance

**Geometrical.shape.of.the.colony** Shape descriptions. See Rimet et al. 2019 for abbreviations

**traitCSR** CSR classification using traits\_to\_CSR function and criteria from Reynolds 2006

---

date_mat	<i>Transform a phytoplankton timeseries into a matrix of abundances for ordination</i>
----------	--

---

## Description

Transform a phytoplankton timeseries into a matrix of abundances for ordination

## Usage

```
date_mat(
  phyto.df,
  abundance.var = "biovol_um3_ml",
  summary.type = "abundance",
  taxa.name = "phyto_name",
  date.name = "date_dd_mm_yy",
  format = "%d-%m-%y",
  time.agg = c("day", "month", "year", "monthyear"),
  fun = mean_naomit
)
```

## Arguments

phyto.df	Name of data.frame object
abundance.var	Character string: field containing abundance data. Can be NA if the dataset only contains a species list for each sampling date.
summary.type	'abundance' for a matrix of aggregated abundance, 'presence.absence' for 1 (present) and 0 (absent).
taxa.name	Character string: field containing taxonomic identifiers.
date.name	Character string: field containing date.
format	Character string: POSIX format string for formatting date column.
time.agg	Character string: time interval for aggregating abundance. default is day.
fun	function for aggregation. default is mean, excluding NA's.

**Value**

A matrix of phytoplankton abundance, with taxa in rows and time in columns. If `time.agg = 'month'`, returns a 3dimensional matrix (taxa,month,year). If `abundance.var = NA`, matrix cells will be 1 for present, 0 for absent

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

geneva.mat1<-date_mat(lakegeneva,time.agg='month',summary.type='presence.absence')
geneva.mat2<-date_mat(lakegeneva,time.agg='month',summary.type='abundance')

geneva.mat1
geneva.mat2
```

---

<code>genus_search_itis</code>	<i>Wrapper function for several functions in ritis:: Searches ITIS database for matches to a genus name outputs matches, current accepted names, synonyms, and higher taxonomy</i>
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---

**Description**

Wrapper function for several functions in ritis:: Searches ITIS database for matches to a genus name outputs matches, current accepted names, synonyms, and higher taxonomy

**Usage**

```
genus_search_itis(genus, higher = FALSE)
```

**Arguments**

<code>genus</code>	Character string. genus name to search for in ITIS
<code>higher</code>	Boolean. If TRUE, add higher taxonomic classifications to output

**Value**

input data.frame with a new character column of MFG classifications and diagnostic information

**Examples**

```
genus='Anabaena'
genus_search_itis(genus,higher=TRUE)
```

---

genus_species_extract	<i>Split a dataframe column with binomial name into genus and species columns.</i>
-----------------------	--

---

**Description**

Split a dataframe column with binomial name into genus and species columns.

**Usage**

```
genus_species_extract(phyto.df, phyto.name)
```

**Arguments**

phyto.df	Name of data.frame object
phyto.name	Character string: field in phyto.df containing species name.

**Value**

A data.frame with new character fields 'genus' and 'species'

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

head(lakegeneva) #need to split the phyto_name column
new.lakegeneva=genus_species_extract(lakegeneva,'phyto_name')

head(new.lakegeneva)
```

---

gnr_simple	<i>Wrapper function for taxize::gnr_resolve() checks species names against a variety of online databases supports fuzzy partial matching</i>
------------	--

---

**Description**

Provides convenient output with a single result, using a variety of criteria for the best match

**Usage**

```
gnr_simple(
  name,
  sourceid = NULL,
  topscore = TRUE,
  numhits = TRUE,
  canonical = TRUE,
  with_context = TRUE,
  ...
)
```

**Arguments**

name	character string binomial scientific name to resolve
sourceid	integer with data source id from <code>taxize::gnr_datasources()</code>
topscore	boolean. Should the best match be returned based on score?
numhits	boolean. Should the best match be returned based on the number of sources with a match?
canonical	If TRUE, names do not include authorship or date
with_context	If TRUE, Match scores are weighted for taxonomic consistency
...	Other parameters passed to <code>taxize::gnr_resolve()</code>

**Value**

new data.frame with name matches, column indicating an exact match, scores, and number of hits (matches) from different data sources in `gnr_resolve()`

**Examples**

```
#use taxize::gnr_datasources() to see all possible data sources for name checking.
name<-"Aphanazomenon flos-aquae"
#sourceid=3 for ITIS database,195 for Algaebase
gnr_simple(name,sourceid=3)
```

---

gnr_simple_df	<i>Wrapper function to apply gnr_simple across a data.frame or list of species names</i>
---------------	--

---

**Description**

Provides convenient output with a row per name. To streamline merging with original data.

**Usage**

```
gnr_simple_df(
  df,
  name.column,
  sourceid = NA,
  topscore = TRUE,
  numhits = TRUE,
  canonical = TRUE,
  with_context = TRUE,
  ...
)
```

**Arguments**

df	data.frame containing names to check
name.column	integer or character string with column name containing species names
sourceid	integer with data source id from <code>taxize::gnr_datasources()</code>
topscore	boolean. Should the best match be returned based on score?



numhits	boolean. Should the best match be returned based on the number of sources with a match?
canonical	If TRUE, names do not include authorship or date
with_context	If TRUE, Match scores are weighted for taxonomic consistency
...	Other parameters passed to taxize::gnr_resolve()

### Value

new data.frame original names (orig.name), 1/0 flag for an exact match, the best match (matched.name), and other output from gnr\_simple(). scores, and number of hits (matches) from different data sources in gnr\_resolve()

### Examples

```
data(lakegeneva)
#example dataset with 50 rows

new.lakegeneva <- genus_species_extract(lakegeneva, 'phyto_name')
new.lakegeneva$genus_species <- trimws(paste(new.lakegeneva$genus,
new.lakegeneva$species))

#checking for matches from all GNRS sources:
lakegeneva.namematches <- gnr_simple_df(new.lakegeneva, "genus_species")
head(lakegeneva.namematches)
```

---

itis_search_df	<i>Wrapper function for applying genus_search_itis and species_search_itis to a whole data.frame containing scientific names</i>
----------------	--

---

### Description

Wrapper function for applying genus\_search\_itis and species\_search\_itis to a whole data.frame containing scientific names

### Usage

```
itis_search_df(df, namecol = NA, higher = FALSE, genus.only = FALSE)
```

### Arguments

df	data.frame containing names to check
namecol	integer or character string with column name containing species or genus names
higher	Boolean. If TRUE, add higher taxonomic classifications to output
genus.only	boolean If TRUE, search for matches with just the genus name using genus_search_itis

### Value

data.frame with submitted names (orig.name), matched names (matched.name), 1/0 flag indicating that original name is currently accepted (orig.name.accepted), 1/0 flag indicating if search was genus\_only (for distinguishing genus\_search\_itis and species\_search\_itis results), synonyms if any, and higher taxonomy (if higher=TRUE)

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

new.lakegeneva <- genus_species_extract(lakegeneva, 'phyto_name')

#checking for genus-only name matches in ITIS, and extracting higher taxonomy
#flagging names with imperfect or no matches

lakegeneva.genus.itischeck <- itis_search_df(new.lakegeneva, "genus", genus.only=TRUE)
head(lakegeneva.genus.itischeck)
```

---

lakegeneva	<i>example dataset from lake Geneva, Switzerland</i>
------------	--

---

**Description**

example dataset from lake Geneva, Switzerland

**Usage**

```
data(lakegeneva)
```

**Format**

A data frame with columns:

**lake** lake name  
**phyto\_name** phytoplankton species name  
**month** month of sampling  
**year** year of sampling  
**date\_dd\_mm\_yy** date of sampling  
**biovol\_um3\_ml** biovolume

---

mean_naomit	<i>Compute mean value while ignoring NA's</i>
-------------	---

---

**Description**

Compute mean value while ignoring NA's

**Usage**

```
mean_naomit(x)
```

**Arguments**

x A numeric vector that may contain NA's

**Value**

the mean value

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

mean_naomit(lakegeneva$biovol_um3_ml)
```

---

mfgTraits

*Functional Trait Database derived from Rimet et al.*


---

**Description**

Functional Trait Database derived from Rimet et al.

**Usage**

```
data(mfgTraits)
```

**Format**

A data frame with columns:

**phyto\_name** binomial scientific name

**genus** genus name

**species** species name

**Mobility.apparatus** 1/0 indicates presence/absence of flagella or motility

**Size** character values 'large' or 'small'; based on 35 micrometer max linear dimension

**Colonial** 1/0 indicates typical colonial growth form or not

**Filament** 1/0 indicates filamentous growth form or not

**Centric** 1/0 indicates diatoms with centric growth form

**Gelatinous** 1/0 indicates presence/absence of mucilage

**Aerotopes** 1/0 indicates presence/absence of aerotopes

**Class** Taxonomic class

**Order** Taxonomic order

**MFG.fromtraits** MFG classification using traits\_to\_mfg function

---

mfg_csr_convert	<i>Returns a CSR classification based on Morphofunctional group (MFG). Correspondence based on Salmaso et al. 2015 and Reynolds et al. 1988</i>
-----------------	---

---

### Description

Returns a CSR classification based on Morphofunctional group (MFG). Correspondence based on Salmaso et al. 2015 and Reynolds et al. 1988

### Usage

```
mfg_csr_convert(mfg)
```

### Arguments

mfg                      Character string with MFG name, following Salmaso et al. 2015

### Value

A character string with values 'C','S','R','CR','SC','SR', or NA

### Examples

```
mfg_csr_convert("11a-NakeChlor")
```

---

mfg_csr_convert_df	<i>Returns a CSR classification based on Morphofunctional group (MFG). Correspondence based on Salmaso et al. 2015 and Reynolds et al. 1988</i>
--------------------	---

---

### Description

Returns a CSR classification based on Morphofunctional group (MFG). Correspondence based on Salmaso et al. 2015 and Reynolds et al. 1988

### Usage

```
mfg_csr_convert_df(phyto.df, mfg)
```

### Arguments

phyto.df                dataframe containing a character field containing MFG classifications  
mfg                      Character string with MFG name, following Salmaso et al. 2015

### Value

A dataframe with an additional field named CSR, containing CSR classifications or NA

**Examples**

```
data(lakegeneva)
lakegeneva<-genus_species_extract(lakegeneva,'phyto_name')
lakegeneva<-species_to_mfg_df(lakegeneva)
lakegeneva<-mfg_csr_convert_df(lakegeneva,mfg='MFG')
head(lakegeneva)
```

---

mfg_csr_library	<i>MFG-CSR correspondence based on CSR-trait relationships in Reynolds et al. 1988 and MFG-trait relationships in Salmaso et al. 2015</i>
-----------------	---

---

**Description**

MFG-CSR correspondence based on CSR-trait relationships in Reynolds et al. 1988 and MFG-trait relationships in Salmaso et al. 2015

**Usage**

```
data(mfg_csr_library)
```

**Format**

A data frame with columns:

**MFG** full MFG name from Salmaso et al. 2015

**CSR** CSR classification including intermediate classes

---

phyto_ts_aggregate	<i>Aggregate phytoplankton timeseries based on abundance. Up to 3 grouping variables can be given: e.g. genus, species, stationid, depth range. If no abundance var is given, will aggregate to presence/absence of grouping vars.</i>
--------------------	--

---

**Description**

Aggregate phytoplankton timeseries based on abundance. Up to 3 grouping variables can be given: e.g. genus, species, stationid, depth range. If no abundance var is given, will aggregate to presence/absence of grouping vars.

**Usage**

```
phyto_ts_aggregate(
  phyto.data,
  DateVar = "date_dd_mm_yy",
  SummaryType = c("abundance", "presence.absence"),
  AbundanceVar = "biovol_um3_ml",
  GroupingVar1 = "phyto_name",
  GroupingVar2 = NA,
  GroupingVar3 = NA,
```

```

    remove.rare = FALSE,
    fun = sum,
    format = "%d-%m-%y"
  )

```

### Arguments

phyto.data	data.frame
DateVar	character string: field name for date variable. character or POSIX data.
SummaryType	'abundance' for a matrix of aggregated abundance, 'presence.absence' for 1 (present) and 0 (absent).
AbundanceVar	character string with field name containing abundance data Can be NA if data is only a species list and aggregated presence/absence is desired.
GroupingVar1	character string: field name for first grouping variable. defaults to spp.
GroupingVar2	character string: name of additional grouping var field
GroupingVar3	character string: name of additional grouping var field
remove.rare	TRUE/FALSE. If TRUE, removes all instances of GroupingVar1 that occur < 5 of time periods.
fun	function used to aggregate abundance based on grouping variables
format	character string: format for DateVar POSIXct conversion

### Value

a data.frame with grouping vars, date\_dd\_mm\_yy, and abundance or presence/absence

### Examples

```

data(lakegeneva)
lakegeneva<-genus_species_extract(lakegeneva, 'phyto_name')
lg.genera=phyto_ts_aggregate(lakegeneva, SummaryType='presence.absence',
                             GroupingVar1='genus')
head(lg.genera)

```

---

sampeff

*Visually assess change in sampling effort over time (author: Dietmar Straile)*

---

### Description

Visually assess change in sampling effort over time (author: Dietmar Straile)

### Usage

```

sampeff(
  b_data,
  column,
  save.pdf = F,
  lakename = "",
  datecolumn = "date_dd_mm_yy",
  dateformat = "%d-%m-%y"
)

```

**Arguments**

<code>b_data</code>	Name of data.frame object
<code>column</code>	column name or number for field containing abundance (biomass,biovol, etc.) can be NA for presence absence
<code>save.pdf</code>	TRUE/FALSE Should the output plot be saved to a file? defaults to FALSE
<code>lakename</code>	Character string for labeling output plot
<code>datecolumn</code>	Character String or number specifying dataframe field with date information
<code>dateformat</code>	Character string specifying POSIX data format

**Value**

a time-series plot of minimum relative abundance over time. This should change systematically with counting effort.

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

sampeff(lakegeneva,column=6) #column 6 contains biovolume
```

---

<code>species_mfg_library</code>	<i>Trait-based MFG classifications for common Eurasion/North American phytoplankton species. See accompanying manuscript for sources</i>
----------------------------------	--

---

**Description**

Trait-based MFG classifications for common Eurasion/North American phytoplankton species. See accompanying manuscript for sources

**Usage**

```
data(species_mfg_library)
```

**Format**

A data frame with columns:

**genus** genus name

**species** species name

**MFG** corresponding MFG classification based on Salmaso et al. 2015

**source** literature or online source for MFG classification

## References

Algaebase <https://www.algaebase.org>

Phycokey <http://www.cfb.unh.edu/phycokey/phycokey.htm>

Western Diatoms of North America <https://diatoms.org>

CyanoDB 2 <http://www.cyanodb.cz/>

Nordic Microalgae <http://nordicmicroalgae.org>

Phytopedia <https://www.eoas.ubc.ca/research/phytoplankton/>

Kapustin, D., Sterlyagova, I. and Patova, E., 2019. Morphology of *Chrysastrella paradoxa* stomatocysts from the Subpolar Urals (Russia) with comments on related morphotypes. *Phytotaxa*, 402(6), pp.295-300.

---

species_search_itis	<i>Wrapper function for several functions in ritis:: Searches ITIS database for matches to a binomial scientific name outputs matches, current accepted names, synonyms, and higher taxonomy</i>
---------------------	--

---

## Description

Wrapper function for several functions in ritis:: Searches ITIS database for matches to a binomial scientific name outputs matches, current accepted names, synonyms, and higher taxonomy

## Usage

```
species_search_itis(genspp, higher = FALSE)
```

## Arguments

genspp	Character string. Binomial scientific name with space between genus and species.
higher	Boolean. If TRUE, add higher taxonomic classifications to output

## Value

data.frame with submitted name (orig.name), matched name (matched.name), 1/0 flag indicating that original name is currently accepted (orig.name.accepted), 1/0 flag indicating if search was genus\_only (for distinguishing genus\_search\_itis and species\_search\_itis results), synonyms if any, and higher taxonomy (if higher=TRUE)

## Examples

```
species="Aphanizomenon flosaquae"
species_search_itis(species,higher=TRUE)
```



---

species_to_mfg	<i>Conversion of a single genus and species name to a single MFG. Uses species.mfg.library</i>
----------------	--

---

**Description**

Conversion of a single genus and species name to a single MFG. Uses species.mfg.library

**Usage**

```
species_to_mfg(genus, species = "", flag = 1, mfgDbase = NA)
```

**Arguments**

genus	Character string: genus name
species	Character string: species name
flag	Resolve ambiguous mfg: 1 = return(NA), 2 = manual selection
mfgDbase	data.frame of species MFG classifications. Defaults to the supplied species.mfg.library data object

**Value**

a data frame with MFG classification and diagnostic information. ambiguous.mfg=1 if multiple possible mfg matches genus.classification=1 if no exact match was found with genus + species name partial.match=1 if mfg was based on fuzzy matching of taxonomic name.

**Examples**

```
species_to_mfg('Scenedesmus', 'bijuga')
#returns "11a-NakeChlor"
```

---

species_to_mfg_df	<i>Wrapper function to apply species_phyto_convert() across a data.frame</i>
-------------------	--

---

**Description**

Wrapper function to apply species\_phyto\_convert() across a data.frame

**Usage**

```
species_to_mfg_df(phyto.df, flag = 1, mfgDbase = NA)
```

**Arguments**

phyto.df	Name of data.frame. Must have character fields named 'genus' and 'species'
flag	Resolve ambiguous MFG: 1 = return(NA), 2 = manual selection
mfgDbase	specify library of species to MFG associations.

**Value**

input data.frame with a new character column of MFG classifications and diagnostic information

**Examples**

```
data(lakegeneva)
#example dataset with 50 rows

new.lakegeneva <- genus_species_extract(lakegeneva, 'phyto_name')
new.lakegeneva <- species_to_mfg_df(new.lakegeneva)
head(new.lakegeneva)
```

---

traitranges	<i>surface/volume ratio and max linear dimension criteria for CSR From Reynolds 1988 and Reynolds 2006</i>
-------------	--

---

**Description**

surface/volume ratio and max linear dimension criteria for CSR From Reynolds 1988 and Reynolds 2006

**Usage**

```
data(traitranges)
```

**Format**

A data frame with columns:

**Measurement** measurement type

**C.min** minimum value for C

**S.min** minimum value for S

**R.min** minimum value for R

**C.max** maximum value for C

**S.max** maximum value for S

**R.max** maximum value for R

**units** units of measurement

**source** source for criteria

---

traits_to_csr	<i>Assign phytoplankton species to CSR functional groups, based on surface to volume ratio and maximum linear dimension ranges proposed by Reynolds et al. 1988;2006</i>
---------------	--

---

## Description

Assign phytoplankton species to CSR functional groups, based on surface to volume ratio and maximum linear dimension ranges proposed by Reynolds et al. 1988;2006

## Usage

```
traits_to_csr(
  sav,
  msv,
  msv.source = "Reynolds 2006",
  traitrange = algaeClassify::traitranges
)
```

## Arguments

sav	numeric estimate of cell or colony surface area /volume ratio
msv	numeric product of surface area/volume ratio and maximum linear dimension
msv.source	character string with reference source for distinguishing criteria
traitrange	data frame with trait criteria for c,s,r groups. The included table can be replaced with user-defined criteria if desired. Measurements are: Surface area/volume ratio (sav), maximum linear dimension (mld) and mld*sav (msv).

## Value

a character string with one of 5 return values: C,CR,S,R, or SR. CR and SR groups reflect overlap between criteria for the 3 main groups.

## See Also

[/urlhttps://powellcenter.usgs.gov/geisha](https://powellcenter.usgs.gov/geisha) for project information

## Examples

```
traits_to_csr(sav=0.2,msv=10,msv.source='Reynolds 2006',traitrange=traitranges)
```

---

traits_to_csr_df	<i>Add CSR functional group classifications to a dataframe of phytoplankton species, based on surface to volume ratio and maximum linear dimension ranges proposed by Reynolds et al. 1988;2006</i>
------------------	---

---

## Description

Add CSR functional group classifications to a dataframe of phytoplankton species, based on surface to volume ratio and maximum linear dimension ranges proposed by Reynolds et al. 1988;2006

## Usage

```
traits_to_csr_df(
  df,
  sav,
  msv,
  msv.source = "Reynolds 2006",
  traitrange = algaeClassify::traitranges
)
```

## Arguments

df	name of dataframe
sav	character string with name of column that contains surface to volume ratio values
msv	character string with name of column that contains maximum linear dimension * surface to volume ratio values
msv.source	character string with reference source for distinguishing criteria
traitrange	data frame with trait criteria for c,s,r groups. The included table can be replaced with user-defined criteria if desired. Measurements are: Surface area/volume ratio (sav), maximum linear dimension (mld) and mld*sav (msv).

## Value

a character string with one of 5 return values: C,CR,S,SR, or R

## Examples

```
csr.df<-data.frame(msv=10,sav=1)

csr.df$CSR<-traits_to_csr_df(csr.df, 'msv', 'sav')

print(csr.df)
```

---

traits_to_mfg	<i>Assign MFG based on binary functional traits and taxonomy (Class and Order)</i>
---------------	--

---

## Description

Assign MFG based on binary functional traits and taxonomy (Class and Order)

## Usage

```
traits_to_mfg(
  flagella = NA,
  size = NA,
  colonial = NA,
  filament = NA,
  centric = NA,
  gelatinous = NA,
  aerotopes = NA,
  class = NA,
  order = NA
)
```

## Arguments

flagella	1 if flagella are present, 0 if they are absent.
size	Character string: 'large' or 'small'. Classification criteria is left to the user.
colonial	1 if typically colonial growth form, 0 if typically unicellular.
filament	1 if dominant growth form is filamentous, 0 if not.
centric	1 if diatom with centric growth form, 0 if not. NA for non-diatoms.
gelatinous	1 mucilagenous sheath is typically present, 0 if not.
aerotopes	1 if aerotopes allowing buoyancy regulation are typically present, 0 if not.
class	Character string: The taxonomic class of the species
order	Character string: The taxonomic order of the species

## Value

A character string of the species' morphofunctional group

## Examples

```
traits_to_mfg(flagella = 1, size = "large", colonial = 1, filament = 0, centric = NA, gelatinous = 0,
  aerotopes = 0, class = "Euglenophyceae", order = "Euglenales")
```

---

traits_to_mfg_df	<i>Assign morphofunctional groups to a dataframe of functional traits and higher taxonomy</i>
------------------	---

---

### Description

Assign morphofunctional groups to a dataframe of functional traits and higher taxonomy

### Usage

```
traits_to_mfg_df(
  dframe,
  arg.names = c("flagella", "size", "colonial", "filament", "centric", "gelatinous",
    "aerotopes", "class", "order")
)
```

### Arguments

dframe	An R dataframe containing functional trait information and higher taxonomy
arg.names	Character string of column names corresponding to arguments for traits_to_mfg()

### Value

A character vector containing morpho-functional group (MFG) designations

### Examples

```
#create a two-row example dataframe of functional traits
func.dframe=data.frame(flagella=1,size=c("large","small"),colonial=0,filament=0,centric=NA,
  gelatinous=0,aerotopes=0,class="Euglenophyceae",order="Euglenales",
  stringsAsFactors=FALSE)

#check the dataframe
print(func.dframe)

#run the function to produce a two-element character vector
func.dframe$MFG<-traits_to_mfg_df(func.dframe,c("flagella","size","colonial",
  "filament","centric","gelatinous",
  "aerotopes","class","order"))

print(func.dframe)
```

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