

Dr. Vladimir O. Talibov

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Neversvägen 43
SE 22479
Lund, Sweden

+46728453329
vladimir.talibov@maxiv.lu.se
vladimir.o.talibov@gmail.com

Summary	A protein biochemist with an interest in early stage drug discovery. Experienced in biophysical and kinetic methods.	
Skills	<p><u>Experimental</u>: Biophysical methods (SPR biosensors, TSA, MST), enzymology, protein techniques, expression&purification, macromolecular crystallography</p> <p><u>Computer</u>: Linux, markup, RDKit, KNIME</p> <p><u>Languages</u>: English, Russian, Swedish (basic)</p> <p><u>Expertise</u>: Biophysical methods, enzymology, small molecules, biosensors.</p>	
Experience	<i>Scientist</i> MAX IV Laboratory, Lund, Sweden	2019 – current
	<ul style="list-style-type: none">• Development of operation protocols for MAX IV fragment screening facility; design, curation and maintenance of in-house fragment library• BioMAX user support as a beamline scientist.	
	<i>PhD student, Researcher</i> Uppsala University, Uppsala, Sweden	2014 – 2019
	<ul style="list-style-type: none">• Design&development of biophysical (SPR, MST, TSA) and enzymatic assays• Maintenance of biosensors and chromatographic equipment.	
	<i>Laboratory Assistant</i> OOO "Biochip-IMB", Moscow, Russia	2012 – 2014
	<ul style="list-style-type: none">• Development and validation of protein microarray-based clinical diagnostics assays• QC of proteins, synthetic oligonucleotides and reactive small molecules.	
Education	<i>PhD in Biochemistry</i> Uppsala University, Uppsala, Sweden Advisor: U. Helena Danielson Thesis: "Interaction kinetic analysis in drug design, enzymology and protein research" .	2014 – 2019
	<i>BSc&MSc in Chemistry</i> Moscow State University, Moscow, Russia Specialisation in bioorganic chemistry.	2008 – 2013
Interests	Molecular recognition, early stage lead discovery, biophysical methods, interaction kinetics.	

Publications

Research articles: 7; details are available at [GScholar](#).

Other: reviews - 2, book chapters - 1, patents - 1.

- [1] G. M. Lima, V. O. Talibov, E. Jagudin, C. Sele, M. Nyblom, W. Knecht, D. T. Logan, T. Sjögren, and U. Mueller. “FragMAX: the fragment-screening platform at the MAX IV Laboratory”. In: *Acta Crystallogr., Sect. D: Struct. Biol.* 76.8 (2020), pp. 771–777.
- [2] T. Ursby, K. Åhnberg, R. Appio, O. Aurelius, A. Barczyk, A. Bartalesi, M. Bjelčić, F. Bolmsten, Y. Cerenius, R. B. Doak, et al. “BioMAX—the first macromolecular crystallography beamline at MAX IV Laboratory”. In: *J. Synchrotron Radiat.* 27.5 (2020).
- [3] J. Yang*, V. O. Talibov*, S. Peintner, C. Rhee, V. Poongavanam, M. Geitmann, M. R. Sebastiano, B. Simon, J. Hennig, D. Dobritzsch, U. H. Danielson, and J. Kihlberg. “Macrocyclic Peptides Uncover a Novel Binding Mode for Reversible Inhibitors of LSD1”. In: *ACS Omega* 8.5 (2020), pp. 3979–3995.
- [4] E. Fabini*, V. O. Talibov*, F. Mihalic, M. Naldi, M. Bartolini, C. Bertucci, A. Del Rio, and U. H. Danielson. “Unveiling the biochemistry of the epigenetic regulator SMYD3”. In: *Biochemistry* 58.35 (2019), pp. 3634–3645.
- [5] V. O. Talibov, V. Linkuvienė, U. H. Danielson, and D. Matulis. “Kinetic Analysis of Carbonic Anhydrase–Sulfonamide Inhibitor Interactions”. In: *Carbonic Anhydrase as Drug Target*. Springer, Cham, 2019, pp. 125–140.
- [6] V. Linkuviene*, V. O. Talibov*, U. H. Danielson, and D. Matulis. “Introduction of intrinsic kinetics of protein–ligand interactions and their implications for drug design”. In: *J. Med. Chem.* 61.6 (2018), pp. 2292–2302.
- [7] C. Seeger, V. O. Talibov, and U. H. Danielson. “Biophysical analysis of the dynamics of calmodulin interactions with neurogranin and Ca²⁺/calmodulin-dependent kinase II”. In: *J. Mol. Recognit.* 30.8 (2017), e2621.
- [8] V. O. Talibov, V. Linkuvienė, D. Matulis, and U. H. Danielson. “Kinetically selective inhibitors of human carbonic anhydrase isozymes I, II, VII, IX, XII, and XIII”. In: *J. Med. Chem.* 59.5 (2016), pp. 2083–2093.
- [9] V. I. Butvilovskaya, M. V. Tsybulskaya, A. A. Tikhonov, V. O. Talibov, P. V. Belousov, A. Y. Sazykin, A. M. Schwartz, S. A. Surzhikov, A. A. Stomakhin, O. N. Solopova, et al. “Preparation of recombinant serpins B3 and B4 and investigation of their specific interactions with antibodies using hydrogel-based microarrays”. In: *Mol. Biol.* 49.5 (2015), pp. 705–713.
- [10] B. Koos, G. Cane, K. Grannas, L. Löf, L. Arngården, J. Heldin, C.-M. Clausson, A. Klaesson, M. K. Hirvonen, F. M. De Oliveira, et al. “Proximity-dependent initiation of hybridization chain reaction”. In: *Nat. Commun.* 6 (2015), p. 7294.
- [11] G. U. Feyzkhanova, M. A. Filippova, V. O. Talibov, E. I. Dementieva, V. V. Maslennikov, Y. P. Reznikov, N. Offermann, A. S. Zasedatelev, A. Y. Rubina, and M. Fooke-Achterrath. “Development of hydrogel biochip for in vitro allergy diagnostics”. In: *J. Immunol. Methods* 406 (2014), pp. 51–57.
- [12] A. Y. Rubina, G. U. Feizkhanova, M. A. Filippova, V. O. Talibov, M. Fooke-Achterrath, and A. S. Zasedatelev. “Multiplex assay of allergen-specific and total immunoglobulins of E and G classes in the biochip format”. In: *Dokl. Biochem. Biophys.* 447.1 (2012), p. 289.