

Sequence alignments (1)

Why create them?

How are they stored?

How can they be manipulated?

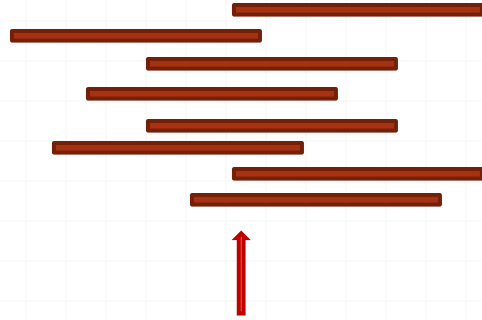
This presentation

- o - is intended to :*
 - o give a brief overview of types of sequence alignment file formats*
 - o provide brief background useful for the SAMTOOLS tutorial run in the practical sessions*
 - o Course notes for tutorial : http://biobits.org/samtools_primer.html*
- o There will be more detailed presentations / sessions on both the above topics*

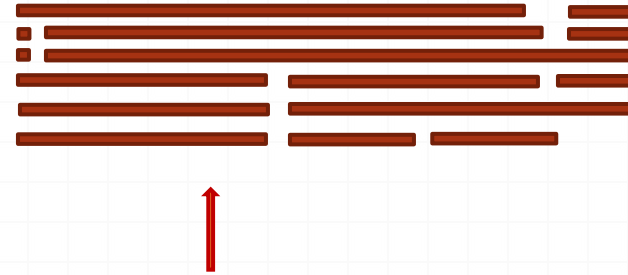
Sequence alignment data

- What is the purpose of creating and storing a sequence alignment?
- There are multiple purposes
- Different aims
- These aims are associated with different ways of representing the alignment
- And thus different **alignment file formats**

- Related but different aims, meanings and file formats
- Sequence read alignment (“assembly”)
- Multiple protein or nucleotide sequence alignment



- Each nucleotide position (column) represents multiple copies of the same base of an original sequence (e.g. genome sequence)



- Each position (column) represents a **homologous** nucleotide (or amino acid).
- Sequences are evolutionarily related (homologous) sequences, typically from different organisms, and/or multiple members of a gene family
- Gaps represent insertions/ deletions

“Traditional” alignment of 2 or more sequences

- Pairwise alignment and multiple alignment
- From an algorithmic point of view:
- There are some significant differences between methods used to:
 - compare and align only 2 sequences (**pairwise alignment**)
 - compare and align 3 or more sequences (**multiple alignment**)
- Some pairwise alignment tools produce their own particular alignment formats
 - E.g. some programs in the versatile EMBOSS software package

- o a **global alignment**, i.e. complete lengths of both sequences are aligned

P0A1L1	0	-----	0
Q0PBF7_CAMJE	1	MRLLVLFLLILPLYSVELISYNIYDRNDRVDLMLSFDNAYNGKISQKKEK	50
P0A1L1	0	-----	0
Q0PBF7_CAMJE	51	NLTLLTFSDLTYSKDELKELNSQLVDKISISSKNNTYIMLQNKKQNINLE	100
P0A1L1	1	-----MMKTEATV-SQPTAPAGS--PLM--QVSGALIG--- ::: .: . .:.: : .	28
Q0PBF7_CAMJE	101	LSSINDKFGVRIRAIIEQGKANIESAPTTTANNSQEELMPKPKSTSLEGYDY	150
P0A1L1	29	-----IIALILAAAWVIKRMGFAPKGNSVRGLKVSASASLGPRERVV ::: :.....:.....:	70
Q0PBF7_CAMJE	151	TNYILVMLILVILLIVLWWFKKTMVYKNNNVSRDFTMIQRFLDKNNQLV	200
P0A1L1	71	IVEVENARLVLGVTASQINLLHTLPPEANDTEAPVPAPPADFQNMMSLLK ::::.. .::.....: :.....	120
Q0PBF7_CAMJE	201	VFDHANKRYTMIIGNSNVLLESIEIPEEQTIKHTEKTEKNFDSFFEENKK	250
P0A1L1	121	RSGRS----- 125 	
Q0PBF7_CAMJE	251	RIQNLIQRQKGKKS	265

“Traditional” alignment of 2 or more sequences

- Typically, DNA sequences representing (complete) genes
- Or protein sequences
- A principle purpose is to identify the parts of the sequence which are the same and which are different
- I.e. identify **conservation** and **divergence**
- This can highlight regions of potential functional importance
- Can be especially informative if you are dealing with several/many sequences (i.e. a **multiple alignment**)


```
>MATK_ROSCA/1-330
MEEFQGYLELYRSQQHDFLYPLIFREYIYALAHDRGLNR....SVLLDNVGYDK.KSSL
LIKRLIS...RMYQQNHFLISVNDNSQNKFF...GYNKNLYS..Q..IISEGFAVIV
EIPFSLRLVSSL...KE.TETVKSYN..LRSIHSIPEPFDEKFPHLNYASDVLIPYPIHL
EILVQTLRYVCVKDPPSLHLRLFLPHEYYNMNTLIT..PKKSIF...AKSNQR..LFLLLY
NSYVCEYESILLFLRNQSNHLRLTSSGILFERIRFYEKIKYPVEEVFANDFPATLWFFKD
PFIQYVRYQGSILASKDTPLLNKWKYYLVNFWQCHFVWSQPGRIHINQLSKHSDFDL
G
>MATK_HORVD/1-336
MEKFEYGESEKSRQQYFVYPLLFQREYIYAFADHYGLNG....SEPVEIVSWNNKFFSS
LLVKRLII...RMYQQNFDLNSVNHNPQDRLLDYKIFFYSEFYS..Q..ILSEGAIVV
EIPFSLRELSCP...KE.KEIPKFN..LRSIHSIPEPFLEKFLHLDYLSHIEIPYPIHL
EILVQLLQYRIQDVPSLHLRFFLNYSNMNSFIT..SMKSILFF.QKENKR..LVKFLY
NSYVSEYEFLLFLRKQSSCLPLAYSGTFLERIHFSRKMEHFG..IMYPGFSRKTILWFEMD
PLIHVYRYQGSILASKGSFFLKKKWKCYLINFWQYFYFFFTQPRRIHINQLNSCDFDM
G
>O46990_9MAGN/1-323
MEKSQGYLELDKSWRHDFLYPLIFQREYIYALAHQGLNR....SILLENTDHDN.KYSS
LIVKRLIT...RMHQNHFLIFDNDNSQNPFW...KHNHNLYS..Q..TISEGEVIV
EIPFSPRFVDSL...EKKKIVKSN..LRSIHSIPEPFLEQFLHLNFVSNLIPYPIHL
EIVVQSLRYRVKDASSLHLRFFLFT.....LNKSISSE.SKRNRR..FFLFLY
NSHVYESTFIFLRLNKTSLRSTSSGAFLERIFFYGIKHLI..EVFANDFQAILWLEKD
PFMHYVRYQGSILASKRTSLRMNKWKYYLVNFWQCFVYVWSQPGRVISINQLSNHSLDFL
G
>MATK_SAXIN/1-335
MEEFQGYLELNKFRONDFLYPLIFQREYIYALAHQILKK....CILSDNLSYDN.KSSS
LIVKRLIT...QMSQLNHLIISDNDNSQNTFL...GHTNKLIDY..QNKMISEGFAVVV
EIQFSLRLVSSL...ER.REIVKSN..LRSIHSIPEPFLEDNFLHLNYVSDILIPHPIHL
EILVQTLRYVWKDASSLHLRFFLYEYQNRSTLITSTPKKAISIV.SKGNHR..LFLILY
NSYLCEYESIFIFIQNSSHLRSISSTGLFERIYFYGIKHLV..EVFYDPTVLWLEKA
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G
>MATK_SULSU/1-333
MGEPQGYLELDKFRQHHFLYPLIFQREYIYALAHDHVNLNR....SILLDNFGYDN.KSSS
IIVKRLIT...RMGQQNHLLISANSYNNKKNFL...GHNKNFDS..Q..MISEGFAVIV
EIPFSLRLVSSL...ER.KEIVKSN..LRSIHSIPEPFLEDNFLHLNYVSDILIPHPIHL
EILVQTLRYVWKDASSLHLRFFLYEYQNSNLSITPTPKKSISIV.SQRNQR..LFLFLY
NSYVCEYESTFIFLNQSSHLQSTSYGTLFERIYFYGIKHLV..EVFSNDPPTAPWLEKD
PFMHYVRYQGSILASKGTPLLLNKWKYYLVNFWQCHYVWSQSGRIHINQLSNHSLDFL
G
>MATK_SINAL/1-329
MEKFGYLELDGARQQSFYPLFFREYIYVLSYDHGLNRLNRNRSIFLENADYDK.KYSS
LIVKRLIL...RMYEQNRLIIPTKDLNTNL.....GHTNLFYY..Q..MISVLEAVIV
EIPFSLRLGSSP...EG.KNLKSYN..LQSIHSIPEPFLEKFSHFNYVLVDLIPYPIHL
EILVQTLRYRVKDASSLHFFRCLVEYCNMKNFDS..KKK.....SILNPR..FFLFLY
NSHVCEYESIFFFLRKQSSHLRSTSYEVFFERILFYGKIQHFL.KVFNMFAPAILGLLKD
PFLHYVRYHGKYLATKDTPLMHNKWKYYFVNFWQCFYVWSQSKININQLSKDNFEFL
G
>MATK_ACTCH/1-331
MEEFKRYLELDRSQQHDFVYTLFQREYIYVLADHGLNRLNRNRSIFLENADYDK.KYSS
LIVKRLIT...QMDQQNHFLISPDNSQNPFL...GHNTNLYS..Q..MILEGFAVVV
EIPFSLRLISSL...EG.KETVSKH..LRSIHSIPEPFLEAKFSRLNYVLDILIPYPIHL
EILVQTLRYRVKDASSLHFFRCLVEYCNMKNFDS..KKK.....SILNPR..FFLFLY
NSHVCEYESIFFFLRKQSSHLRSTSYEVFFERILFYGKIQHFL.KVFNMFAPAILGLLKD
PFLHYVRYHGKYLATKDTPLMHNKWKYYFVNFWQCFYVWSQSKININQLSKDNFEFL
G
```

FASTA

MSF

```
# STOCKHOLM 1.0
#=> ID      MatK_JN
#=> AC      PF01824.13
#=> DR      MatK/Tmk amino terminal region
#=> AU      Bateman A
#=> SE      Pfam-B_30 (release 4.2)
#=> GA      21:70 21:70;
#=> TC      21:70 21:70;
#=> NC      21:10 19:80;
#=> GH      hmmbuild HMM.ann SEED.ann
#=> SH      hmmssearch -Z 23193494 -E 1000 --cpu 4 HMM pfamseq
#=> TP      Family
#=> RN      [1]
#=> RI      8255751
#=> RT      Evolutionary relationships among group II intron-encoded
#=> RT      proteins and identification of a conserved domain that may be
#=> RT      related to maturase function.
#=> RA      Mohr G, Perlman PS, Lambowitz AH;
#=> RL      Nucleic Acids Res 1993;21:4991-4997.
#=> DR      INTERPRO: IPR024942;
#=> CC      The function of this region is unknown.
#=> SQ      15
#=> GS      MATK_ROSCA/1-330      AC 0782552.1
#=> GS      MATK_HORVD/1-336      AC 0781802.1
#=> GS      O46990_9MAGN/1-323      AC 046990.1
#=> GS      MATK_SAXIN/1-335      AC P36435.1
#=> GS      MATK_SULSU/1-333      AC P36436.1
#=> GS      MATK_SINAL/1-329      AC P09364.2
#=> GS      MATK_ACTCH/1-331      AC 047141.1
#=> GS      078408_9ERIC/1-333      AC 078408.1
#=> GS      062994_ELLPA/1-333      AC 062994.1
#=> GS      O47144_BEFFRA/1-333      AC 047144.1
#=> GS      078246_9ASTR/1-333      AC 078246.1
#=> GS      MATK_SOLU/1-336      AC P32088.2
#=> GS      O03561_9GENT/1-332      AC O03561.2
#=> GS      O63139_9GENT/1-338      AC O63139.1
#=> GS      O20105_9GENT/1-334      AC O20105.1
MATK_ROSCA/1-330
MEEFQGYLELYRSQQHDFLYPLIFREYIYALAHDRGLNR....SVLLDNVGYDK.KSSLLIKRLIS...RMYQQNHFLISVNDNSQNKFF...GYNKNLYS..Q..IISEGFAVIVEIPFSLRLVSSL...KE.TETVKSYN..LRSIHSIPEPFDEKFPHLNYAS
MATK_HORVD/1-336
MEKFEYGESEKSRQQYFVYPLLFQREYIYAFADHYGLNG....SEPVEIVSWNNKFFSSLLVKRLII...RMYQQNFDLNSVNHNPQDRLLDYKIFFYSEFYS..Q..ILSEGAIVVEIPFSLRELSCP...KE.KEIPKFN..LRSIHSIPEPFLEKFLHLDYLSHIEIPYPIHL
O46990_9MAGN/1-323
MEKSQGYLELDKSWRHDFLYPLIFQREYIYALAHQGLNR....SILLENTDHDN.KYSSLIVKRLIT...RMHQNHFLIFDNDNSQNPFW...KHNHNLYS..Q..TISEGEVIVEIPFSPRFVDSL...EKKKIVKSN..LRSIHSIPEPFLEQFLHLNFVSNLIPYPIHL
MATK_SAXIN/1-335
MEEFQGYLELNKFRONDFLYPLIFQREYIYALAHQILKK....CILSDNLSYDN.KSSS
MATK_SULSU/1-333
MGEPQGYLELDKFRQHHFLYPLIFQREYIYALAHDHVNLNR....SILLDNFGYDN.KSSS
MATK_SINAL/1-329
MEKFGYLELDGARQQSFYPLFFREYIYVLSYDHGLNRLNRNRSIFLENADYDK.KYSSLIVKRLIL...RMYEQNRLIIPTKDLNTNL.....GHTNLFYY..Q..MISVLEAVIVEIPFSLRLGSSP...EG.KNLKSYN..LQSIHSIPEPFLEKFSHFNYVLVDLIPYPIHL
MATK_ACTCH/1-331
MEEFKRYLELDRSQQHDFVYTLFQREYIYVLADHGLNRLNRNRSIFLENADYDK.KYSSLIVKRLIT...QMDQQNHFLISPDNSQNPFL...GHNTNLYS..Q..MILEGFAVVVEIPFSLRLISSL...EG.KETVSKH..LRSIHSIPEPFLEAKFSRLNYVLDILIPYPIHL
```

```
!!AA_MULTIPLE_ALIGNMENT 1.0

stdout MSF: 361 Type: P 28/05/14 CompCheck: 7924 ..

Name: MATK_ROSCA/1-330      Len: 361      Check: 6381 Weight: 1.00
Name: MATK_HORVD/1-336      Len: 361      Check: 534 Weight: 1.00
Name: O46990_9MAGN/1-323      Len: 361      Check: 5814 Weight: 1.00
Name: MATK_SAXIN/1-335      Len: 361      Check: 4324 Weight: 1.00
Name: MATK_SULSU/1-333      Len: 361      Check: 3164 Weight: 1.00
Name: MATK_SINAL/1-329      Len: 361      Check: 2893 Weight: 1.00
Name: MATK_ACTCH/1-331      Len: 361      Check: 7535 Weight: 1.00
Name: O78408_9ERIC/1-333      Len: 361      Check: 4909 Weight: 1.00
Name: O62994_ELLPA/1-333      Len: 361      Check: 4516 Weight: 1.00
Name: O47144_BEFFRA/1-333      Len: 361      Check: 3986 Weight: 1.00
Name: O78246_9ASTR/1-333      Len: 361      Check: 9032 Weight: 1.00
Name: MATK_SOLU/1-336      Len: 361      Check: 97 Weight: 1.00
Name: O03561_9GENT/1-332      Len: 361      Check: 7541 Weight: 1.00
Name: O63139_9GENT/1-338      Len: 361      Check: 3336 Weight: 1.00
Name: O20105_9GENT/1-334      Len: 361      Check: 3862 Weight: 1.00

//

1                                     50
MATK_ROSCA/1-330      MEEFQGYLELYRSQQHDFLYPLIFREYIYALAHDRGLNR....SVLLDN
MATK_HORVD/1-336      MEKFEYGESEKSRQQYFVYPLLFQREYIYAFADHYGLNG....SEPVEI
O46990_9MAGN/1-323      MEKSQGYLELDKSWRHDFLYPLIFQREYIYALAHQGLNR....SILLEN
MATK_SAXIN/1-335      MEEYQGYLELNKFRONDFLYPLIFQREYIYALAHQILKK....CILSDN
MATK_SULSU/1-333      MGEPQGYLELDKFRQHHFLYPLIFQREYIYALAHDHVNLNR....SILLDN
MATK_SINAL/1-329      MEKFGYLELFDGARQQSFYPLFFREYIYVLSYDHGLNRLNRNRSIFLEN
MATK_ACTCH/1-331      MEEFKRYLELDRSQQHDFVYTLFQREYIYVLADHGLNR....SILLEN
O78408_9ERIC/1-333      MEEFKRYLELDRSQQHDFVYTLFQREYIYALAHDRDLNR....SFFQFS
O62994_ELLPA/1-333      MEEFKRYLELDRSQQHDFVYTLFQREYIYALAHDRGLNK....SIFLEN
O47144_BEFFRA/1-333      MEELKRYLELDSSQQHDFIYTLFQREYIYALAHDRGLNR....SIFSEN
O78246_9ASTR/1-333      MEKFSQSYLGLDRSQQHDFYPLIFQREYIYVLADHGLNR....SILLEN
MATK_SOLU/1-336      MEIIRHYLQPDSSQQHNFYPLIFQREYIYALAQDHGLNRN....RSILLEN
O03561_9GENT/1-332      MEEIQRYLQDRSQQHGFYPLIFQREYIYALAHDSHLNR....SILLEN
O63139_9GENT/1-338      MEEIQRYLQDRSQQHGFYPLIFQREYIYVLADHSHLNR....FILLN
O20105_9GENT/1-334      MEEIPRYLQDRSQQLSFYPLIFQREYIYALAHDSHLNR....AILLEN

51                                     100
MATK_ROSCA/1-330      VGYDK.KSSLLIKRLIS...RMYQQNHFLISVNDNSQNKFF...GYN
MATK_HORVD/1-336      VSWNNKKFSLLVKRLII...RMYQQNFDLNSVNHNPQDRLLDYKIFFY
O46990_9MAGN/1-323      TDHDN.KYSSLIVKRLIT...RMHQNHFLIFDNDNSQNPFW...KHN
MATK_SAXIN/1-335      LSYDN.KSSSLLIKRLIT...QMSQLNHLIISDNDNSQNTFL...GHT
MATK_SULSU/1-333      FGYDN.KSSSIIIVKRLIT...RMGQQNHLLISANSYNNKKNFL...GHN
MATK_SINAL/1-329      ADYDK.KYSSLIVKRLIL...RMYEQNRLIIPTKDLNTNL.....GHT
```

Stockholm

Some multiple sequence alignment formats – these are all plain text files (“flatfiles”)

Numerous other formats exist

CLUSTAL format is widely used

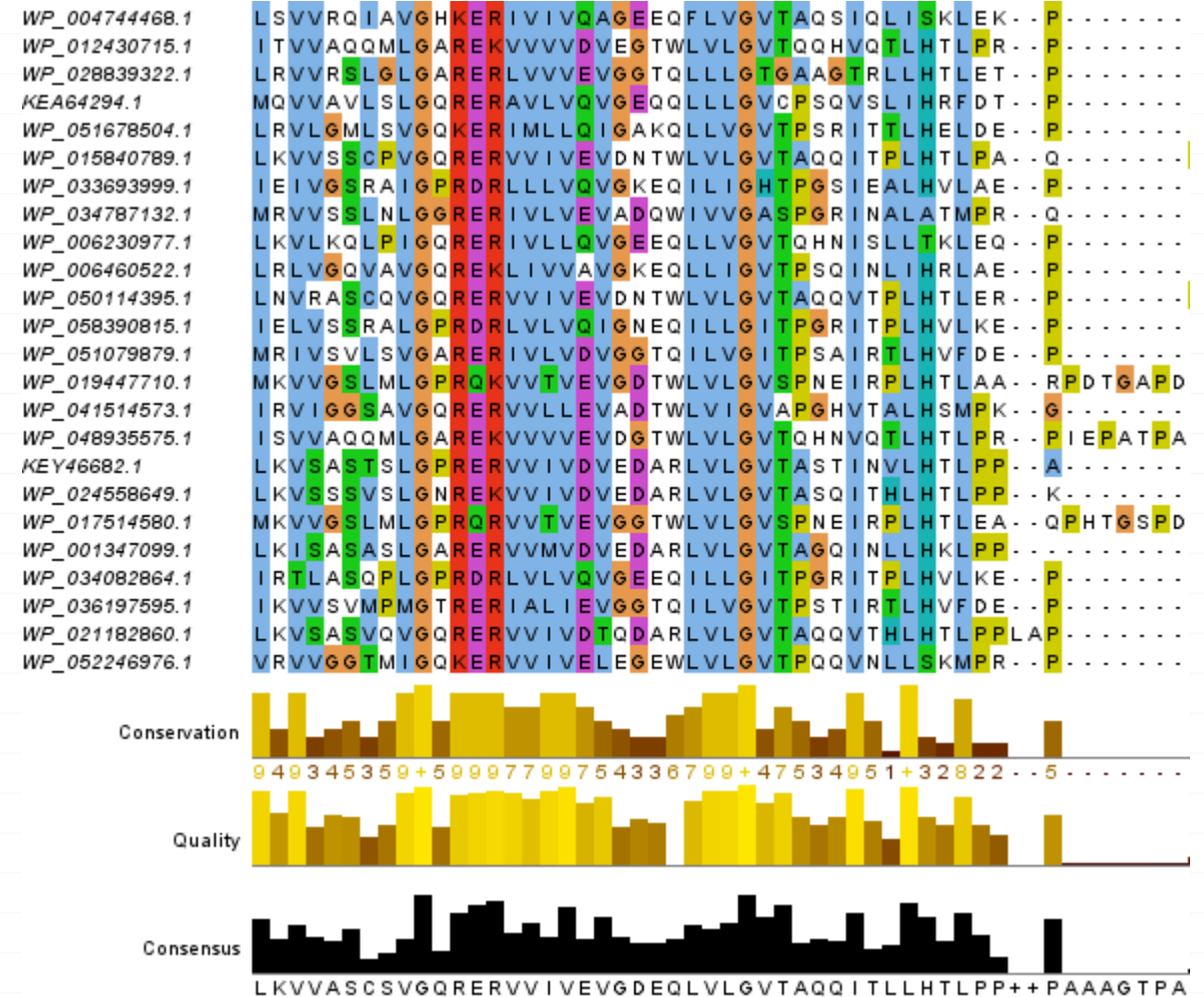
26/10/2016



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Viewing multiple alignments

- Example display from a program (Jalview) which reads the flatfiles and creates a formatted/annotated view



- Example BLAST output
 - A list of **pairwise** alignments
 - One sequence (the Query) is always the same
 - In each case, the other sequence is the 'hit' (Subject)
 - N.B. these are **local** alignments (i.e. only of matching segments, not necessarily the whole sequence)
- More compact BLAST output formats are available
- These are **flatfiles** (plain text)
- Again, when interpreted and displayed by other software they may appear very different
- E.g. on a website which provides a BLAST service

```
> tr|A0A031GPC8|A0A031GPC8_9BURK Flagellar biosynthesis protein
FliO OS=Janthinobacterium lividum GN=fliO PE=4 SV=1
Length=196
```

```
Score = 45.8 bits (107), Expect = 0.009, Method: Composition-based stats.
Identities = 16/114 (14%), Positives = 40/114 (35%), Gaps = 5/114 (4%)
```

```
Query 143 TSLEGYDYTNYLIVMLILVILLIVLWWFKKTMVYKNNNVS RDFTMIFQRFLDKNNQLVVF 202
      +      I ++ ++ LLI L WF K K + ++ L ++V+
Sbjct 79 PASSAGSLLQTIFALMFVLLALLIGLAWFMKRYGPKVMGGNNKMRVSSSLNLGGRERIVLV 138

Query 203 DHANKRYTMIIGNSNV-LLESIEIPEEQTIKHTTE----KTEKNFDSFFEENKKR 251
      + A++ + + L ++ E + NF ++ ++
Sbjct 139 EVADQWIVVGASPGRINALATMPRQEGDLPQLATAQNGPAAANFSEWLKQTIEK 192
```

```
> tr|A0A0B1REZ0|A0A0B1REZ0_9ENTR Flagellar assembly protein FliO
OS=Pantoea rodasii GN=QU24_01715 PE=4 SV=1
Length=131
```

```
Score = 44.6 bits (104), Expect = 0.009, Method: Composition-based stats.
Identities = 15/98 (15%), Positives = 40/98 (41%), Gaps = 2/98 (2%)
```

```
Query 156 VMLILVILLIVL-WWFKKTMVYKNNNVS RDFTMIFQRFLDKNNQLVVF DHANKRYTMIIG 214
      V+ ++V+L++ W K+ ++ + + ++V+ D A+ R + +
Sbjct 29 VLAVIVLLILACGWLAKRLGFAPKTVNTQALKISASVQVGRQERVVIVDTADARLVLGVT 88

Query 215 NSNVL-LESIEIPEEQTIKHTTEKTEKNFDSFFEENKKR 251
      + L S+ + + ++F F+ KR
Sbjct 89 AQQITHLHSLPPVPPEELASNSVAPQDFRQLFQNLVKR 126
```

```
> tr|A0A090U6M1|A0A090U6M1_9ENTR Flagellar biosynthesis protein
FliO OS=Citrobacter farmeri GTC 1319 GN=fliO PE=4 SV=1
Length=124
```

```
Score = 44.2 bits (103), Expect = 0.010, Method: Composition-based stats.
Identities = 14/84 (17%), Positives = 30/84 (36%), Gaps = 0/84 (0%)
```

```
Query 168 WWFKKTMVYKNNNVS RDFTMIFQRFLDKNNQLVVF DHANKRYTMIIGNSNVLLLESIEIPE 227
      W K+ + +R + L ++V+ D + R + + SN+ + P
Sbjct 37 WVIKRLCFSPKCSHTRCLKVSASTSLGPRERVVIVDVEDARLVLCVTASNISVLHTLPPA 96
```

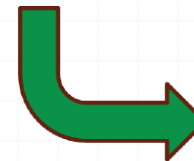
“Traditional” alignment of 2 or more sequences

- Strictly speaking, an alignment is only meaningful if the sequences are homologous (related by descent from a common ancestor)
- This common descent applies to the individual nucleotide (or amino acid) positions –
 - i.e., equivalent bases/ amino acids are lined up
- This also enables inferences of evolutionary events:
 - Substitutions
 - Insertions
 - Deletions

“Traditional” alignment of 2 or more sequences

- o But the process of alignment can help to **determine whether the sequences are homologous or not**
- o This principle applies to **sequence similarity search** methods
 - o E.g. **BLAST**
- o - in which a single query sequence is compared, to many sequences in a database, one at a time
- o i.e. **many pairwise alignments** (all involving the query sequence)
- o Each alignment has a score
- o Only those with a sufficiently high score are treated as **hits**
- o The rest are ignored

- One way of very briefly summarising an alignment is to quote a single metric such as:
 - 'percentage identity'
 - 'percentage similarity'
- However, these are not absolutes, and depend on **how** the sequences are aligned
 - I.e., which method and scoring parameters are used
- These, and other aspects of alignment will be described further in a future session



Principles
of sequence
alignment

Other reasons for pairwise alignment

- Comparing 2 DNA sequences (or 1 RNA sequence and 1 DNA sequence) –
- where one is expected to be a (often very small) fragment of the other
- E.g.:
- Genome **re**sequencing
 - - and *de novo* genome sequencing
- RNASeq mapping

—
?

Genome resequencing

- You already have a **reference genome sequence** of organism X
- The new project is to sequence the genome of organism Y
 - Might be a closely related species
 - Or a different strain of the same species
- Assumption is that the 2 genome sequences are the same or very similar along most of their length
- — Each sequence read from genome sequence Y can be “mapped” to the equivalent position in sequence X

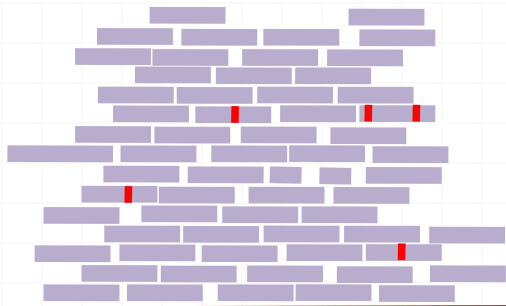
Genome resequencing

- N.B. depending on the circumstances, it may be just as good (or preferable) to do *de novo* assembly of genome Y
 - And then compare the whole of Y to X
 - This “comparative genomics” can be rather more complex than just a pairwise alignment of X and Y
- Resequencing of many genomes – or particular parts of genomes – helps to identify variations between strains in a species
- And variation among individuals in a population
- E.g. identification of SNPs
 - → *variant calling*

RNASeq

- mRNA transcripts are sampled and sequenced
- The object is to map each “read” to a reference genome sequence
- Some reads won’t match perfectly, due to sequencing errors
 - - and even some biological errors
- This is more complicated in eukaryotes than prokaryotes
 - Due to the presence of introns (present in reference genome, absent from reads) and splice variants
- One approach to “community RNASeq” (metatranscriptomics) involves mapping reads to several/many reference genomes
 - Some reads will originate from genomes which are absent from the reference set

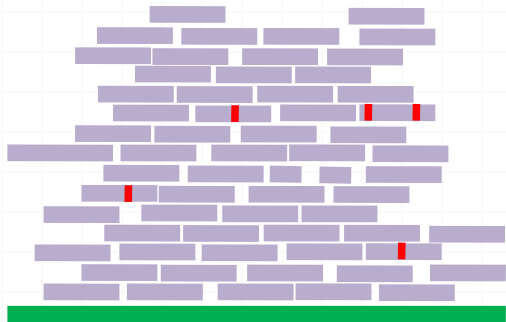
Storing information on mapped reads - economically



- *Is there any point in explicitly recording all the bases which are the **same** as the reference?*

- Each read can be recorded by specifying:
- Its start position relative to the reference genome sequence
- Its length
- All the differences between the read and the reference

De novo genome sequence assembly



- *Again, it should be necessary to record only positions and differences of each read, relative to the consensus*

- Similar principles
- But no reference genome sequence to compare the reads with
- Various algorithms, which in essence compare the reads with each other
- Produces “contigs”
- A consensus sequence can be produced from each contig

How can mapping locations and differences be encoded?

- Various approaches
- Some have 'evolved' beyond their original purpose
- **CIGAR** format (“Compact Idiosyncratic Gapped Alignment Report”)
 - (and it *is* idiosyncratic)
- CIGAR is used in **SAM** format (“Sequence Alignment/Map”) files
- SAM, has a “binary” (compressed) equivalent, BAM
 - **SAM and BAM are still very widely used**
- A more recent development is **CRAM** format
 - Stores only differences compared to reference – **minimal storage space**
 - CRAM files are **not** plain text; you need utilities (in the CRAMTOOLS software) to view them – displayed in e.g. FASTQ or SAM format

What do you need to know?

- **So how much do you need to know about these formats?**
 - A little awareness of the concepts will be useful.
 - Do not be concerned with understanding all the details.
 - (Probably not many people do understand them all...
 - the full SAM specification has many details, some more documented than others)
 - There have been many approaches to **compressing** this kind of sequence data, due to the ever-increasing sizes of data sets
 - Compression of SAM/BAM in particular
 - Compression of FASTQ data in general
 - Beyond the scope of this topic

CIGAR and SAM in brief

A quick overview for now.

There are some more details in another presentation



CIGAR format

- One CIGAR 'string' per alignment (read ↔ reference):
 - Example: '6=1X7=2X6='
- Original CIGAR specification was designed to specify for each alignment:
 - How long the alignment is
 - Where the insertions and deletions (indels) are
 - And some other attributes like 'clipping', 'padding' (ignore this now)
- Later additions to CIGAR also permitted specification of:
 - Where mismatching bases occur
 - But **NOT** what those mismatches actually are
 - -it was simply not the original purpose of the tools which used CIGAR

SAM format

- Each read is represented on one line (1 line = 1 ‘record’)
- Each line has a field specifying the position on the reference sequence
- And a field in CIGAR format
- Then it gets complicated...
- ***Also additional optional fields*** – these can be used to state:
 - explicit base differences (the optional **MD field**)
 - (and many other things)
- SAM format **can** explicitly state the sequence of each read
 - And often does
 - But does **not** have to

That all seems a bit messy....?

So why is it like this?

Evolution of bioinformatics formats

- This is an example of bioinformatics format specifications which have become revised over the years
- The original design may have been to achieve something very specific
- Additional functionality has been enabled by additional information included in the data files
- New fields “bolted on” to a simpler, earlier spec
- Complete revisions are generally uncommon because of the need to retain backward compatibility

Some context

- Example: You want to know **how many reads** are aligned to each part of the genome



Some context

- Example: You want to know **how many reads** are aligned to each part of the genome
- E.g. transcriptomics (RNAseq) – thus, gene expression levels
 - Which genes are most represented?
 - You don't care if there are a small number of mismatches due to sequence errors
 - You don't even care what the sequences of the alignments in each region actually *are*
 - **All you want to know is where the alignment/mapping program (whatever that may be) mapped the reads**
 - **Only minimal CIGAR/SAM information is required for this**
 - (N.B. SAM format can optionally store scores produced by the alignment program)

Summary (1)

- There are many different file formats to store alignment data
- These can be read and created by many different software tools
- Nearly all of these formats are plain text (“flatfile”)
 - BAM is not, but is a compressed version of SAM
 - Various other compressed, “binary” formats exist, e.g. CRAM
- These file formats are used for a variety of purposes
 - Which data types are stored in any one file can depend on the purpose
- It is **not** necessary to understand the detailed specifications of these data file formats
- But it is important to know:
 - ***what kind of data can/cannot be contained in these files***

Summary (2)

- SAM and BAM data files are principally manipulated with the SAMTOOLS software package
- Various other software can read SAM/BAM files
 - E.g. visual browsers, like the Integrated Genome Viewer
- External link to SAMTOOLS tutorial at BIOBITS:
 - http://biobits.org/samtools_primer.html
- If you are interested in learning more details of SAM, CIGAR etc., these are provided in a further presentation

