Additional File 1

Table S1. List of genes, sequences of primers, cycling conditions for PCR assays, and amplicon sizes.

Assay	Gene	Primer sequence	PCR program ^a	Product size (bp)	Reference
E. coli	uspA	F: 5'-CCGATACGCTGCCAATCAGT-3'	94°C/ 120 s, 70°C/ 60 s,	884	1
confirmation	R: 5'-ACGCAGACCGTAGGCCAGAT-3'	72°C/ 60 s (30 cycles)			
Phylogenetic	chuA _	F: 5'-GACGAACCAACGGTCAGGAT-3' R: 5'-TGCCGCCAGTACCAAAGACA-3'	94°C/ 30 s,	279	
typing (Triplex	ујаА	F: 5'-TGAAGTGTCAGGAGACGCTG-3' 55°C/ 30 s, R: 5'-ATGGAGAATGCGTTCCTCAAC-3' 72°C/ 30 s	211	2	
method)	TspE4.C2	F: 5'-GAGTAATGTCGGGGCATTCA-3' R: 5'-CGCGCCAACAAAGTATTACG-3'	(30 cycles)	152	
Phylogenetic typing	chuA	F: 5'-ATGGTACCGGACGAACCAAC-3' R: 5'-TGCCGCCAGTACCAAAGACA-3'	94°C/ 5 s,	288	
(Quadruplex method)	ујаА	F: 5'-CAAACGTGAAGTGTCAGGAG-3' R: 5'-AATGCGTTCCTCAACCTGTG-3'	57°C/ 20 s (30 cycles)	211	3
metriouj	TspE4.C2	F: 5'-CACTATTCGTAAGGTCATCC-3'		152	

		R: 5'-AGTTTATCGCTGCGGGTCGC-3'			
	arpA	arpA F: 5'-AACGCTATTCGCCAGCTTGC-3'		400	
	(Group F)	R: 5'-TCTCCCCATACCGTACGCTA-3'		400	
	arpA	F: 5'-GATTCCATCTTGTCAAAATATGCC-3'		301	
	(Group E)	R: 5'-GAAAAGAAAAGAATTCCCAAGAG-3'		301	
	trpA	F: 5'-AGTTTTATGCCCAGTGCGAG-3'	94°C/ 5 s,		
	(Group C)	D. F/ TOTOCOCCOCTOA COCCO 2/	59°C/ 20 s	219	
	(Group c)	R: 5'-TCTGCGCCGGTCACGCCC-3'	(30 cycles)		
	trpA	F: 5'-CGGCGATAAAGACATCTTCAC-3'	94°C/ 5 s,		
	(Internal	D. F./ COAA CCCCCCCCCCAAC 2/	57°C/ 20 s	489	
	control)	R: 5'-GCAACGCGGCCTGGCGGAAG-3'	(30 cycles)		
		F: 5'-GACCCGGCACAAGCATAAGC-3'	95°C/ 30 s,		
	eae	1.5 UNCCCOUCHCHAUCHTAACC 5	54°C/ 90 s,	384	
		R: 5'-CCACCTGCAGCAACAAGAGG-3'	72°C/ 90 s	304	
DECb		N. 5 CCACCTGCAGCAACAAGAGG	(30 cycles)		4
DLC	stx1	F: 5'-ATAAATCGCCATTCGTTGACTAC-3'	95°C/ 30 s,	180	7
	31/1	R: 5'-AGAACGCCCACTGAGATCATC-3'	52°C/ 60 s,	100	
	stx2	F: 5'-GGCACTGTCTGAAACTGCTCC-3'	72°C/ 60 s	255	
	31,72	R: 5'-TCGCCAGTTATCTGACATTCTG-3'	(35 cycles)	233	

	fyuA	F: 5'-GTAAACAATCTTCCCGCTCGGCAT-3'	72°C/ 90 s	850	
ExPEC ^c		R: 5'-GGCCAGAACATTTGCTCCCTTGTT-3'	63°C/ 90 s,		5
	vat	F: 5'-TCAGGACACGTTCAGGCATTCAGT-3'	94°C/ 30 s,	1100	
			(35 cycles)		
	aggA	R: 5'-TCAACCTTGACACTTGCC-3'	72°C/ 120 s	414	
		1.5 /1100/11/100011/1/10 5	50°C/ 60 s,	414	
	bfpA	R: 5'-ATAGCAGTCGATTTAGCAGCC-3' F: 5'-ATGCATTACTTTGGGTTTAG-3'	94°C/ 60 s,		
			(30 cycles)		
			72°C/ 40 s	461	
		F: 5'-ATTGAATCTGCAATGGTGC-3'	55°C/ 40 s,		
			95°C/ 40 s,		
		R: 5'-GCCGGTCAGCCACCCTCTGAGAGTAC-3'	(35 cycles)		
	іраН	D. E. C.C.C.C.T.O.A.C.C.T.C.O.A.C.T.C.O.A.C.C.T.C.O.A.C.T.C.O.A.C.T.C.O.A.C.T.C.O.A.C.T.C.O.A.C.T.C.O.A.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.O.A.C.C.T.C.T.C.T.C.O.A.C.C.T.C.T.C.T.C.T.C.O.A.C.C.T.C.T.C.T.C.O.A.C.T.C.T.C.T.C.O.A.C.T.C.T.C.T.C.T.C.T.C.T.C.T.C.T.C.T.C	72°C/ 60 s	620	
		F: 5'-GTTCCTTGACCGCCTTTCCGATACCGTC-3'	60°C/ 60 s,		
			94°C/ 40 s,		
	st1A	R: 5'-ATAACATCCAGCACAGGC-3'	(35 cycles)	186	
	ItA	F: 5'-TCTGTATTATCTTTCCCCTC-3'	72°C/ 120 s		
		R: 5'-CCGAATTCTGTTATATATGTC-3'	50°C/ 60 s,	696	
	4.0	F: 5'-GGCGACAGATTATACCGTGC-3'	94°C/ 60 s,	666	

		R: 5'-TGACGATTAACGAACCGGAAGGGA-3'	(30 cycles)		
	chuA	F: 5'-CTGAAACCATGACCGTTACG-3'		652	
		R: 5'-TTGTAGTAACGCACTAAACC-3'		032	
	yfcV _	F: 5'-ACATGGAGACCACGTTCACC-3'	292		
	y)cv _	R: 5'-GTAATCTGGAATGTGGTCAGG-3'		292	
	kpsM K1	F: 5'-TAGCAAACGTTCTATTGGTGC-3'	94°C/ 30 s,	153	
			63°C/ 30 s,		6
			68°C/ 3 min		
			(25 cycles)		
	F: 5'-GCGCATTTGCTGATAC	F: 5'-GCGCATTTGCTGATACTGTTG-3'	94°C/ 30 s,	577	
	kpsM II _		58°C/ 30 s,		7
	рэлг	R: 5'-AGGTAGTTCAGACTCACACCT-3'	72°C/ 3 min	3,,	
			(25 cycles)		

^aBefore starting the PCR cycle, DNA was first denatured at 95°C/15 min. After completion

³ of the cycle, there was a final primer extension at 72°C/8 min.

⁴ bDiarrheagenic *E. coli*

^{5 &}lt;sup>c</sup>Extra-intestinal pathogenic *E. coli*

6	References in Table 1
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Table S2. Sample collection dates and locations, and phylogenetic groupings of *E. coli* by

51 triplex and quadruplex PCR assays.

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Date of sample collection	Location (no. of confirmed <i>E. coli</i>) J ^a (1) Z ^b (2)	Phylogenetic grouping by triplex PCR (no. of isolates) B1 (2) D (1)	Phylogenetic grouping by quadruplex PCR (no. of isolates) B1 (2) D (1)
June 2018	J (7) Z (2) H ^c (5)	A (6) B1 (3) D (5)	A (6) B1 (3) D (4) E (1)
July 2018	J (1) Z (3) H (2)	A (3) B1 (1) B2 (1) D (1)	A (3) B1 (1) B2 (1) D (1)
August 2018	Ј (6) Z (6) H (7)	A (9) B1 (5) D (2)	A (9) B1 (5) D (1) E (1)
September 2018	J (2) Z (1) H (9)	A (8) B1 (3) B2 (1)	A (7) B1 (3) B2 (1) C (1)
October 2018	Ј (4) Z (1) H (4)	A (4) B1 (3) D (2)	A (4) B1 (3) D (1) F (1)

1.70		
	A (7)	A (7)
	B1 (2)	B1 (2)
H (4)		
J (3)	A (3)	A (3)
		B1 (3)
		D (2)
11 (3)	J (4)	F (2)
		A (3)
1 (4)	A (3)	B1 (3)
	B1 (3)	B2 (2)
	B2 (2)	D (3)
H (5)	D (6)	F (1)
		E (2)
J (4)	A (3)	A (3)
Z (2)	B1 (3)	B1 (3)
H (2)	D (2)	F (2)
1 (2)	A (6)	A (6)
	B1 (1)	B1 (1)
	B2 (4)	B2 (4)
п (3)	D (1)	D (1)
		A (14)
J (7)	A (16)	B1 (5)
Z (7)	B1 (5)	D (1)
H (10)	D (3)	F (2)
		C (2)
	Z (4) H (3) J (4) Z (5) H (5) J (4) Z (2) H (2) J (3) Z (6) H (3) J (7) Z (7)	Z (1) H (4) J (3) Z (4) B1 (3) H (3) D (4) A (3) B1 (3) B1 (3) B2 (2) D (6) J (4) A (3) Z (2) B1 (3) H (2) D (2) A (6) H (3) D (1) J (7) A (16) Z (7) B1 (5)

^{52 &}lt;sup>a</sup>Jabriya.

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^{53 &}lt;sup>b</sup>Zahraa.

^{54 &}lt;sup>c</sup>Hateen.

Table S3. Distribution of ExPEC according to location and date of collection of samples.

Month/Year	No. of ExPEC	Location			
Worting real	isolates	Jabriya	Zahraa	Hateen	
May/2018	1	0	1	0	
June/2018	7	2	1	4	
July/2018	2	0	1	1	
August/2018	8	1	2	5	
September/2018	2	0	0	2	
October/2018	4	1	0	3	
November/2018	3	2	1	0	
December/2018	5	2	2	1	
January/2019	7	2	3	2	
February/2019	4	0	2	2	
March/2019	7	2	3	2	
April/2019	7	2	0	5	