Protocol related to slide coating

Summarized by YAN Jie lab;

Slide cleaning:

1. Clean of the staining jar by detergent. And place the #1.5 coverslips (7 pieces each jar to a " $|\cdot|\cdot|\cdot|\cdot|$ " shape). Each channel need 1 piece of functionalized cover glass for the bottom (22×40 mm) and 1 piece of clean cover glass for the top (20×20 mm).



- 2. Ultrasonic to clean the cover glasses
- 2a. Detergent (20% Decon90 in DIwater) wash 30 min. flush 20 times with di-water.



- 2b. Methanol (100%) wash 30 min.
- 2c. Acetone (100%) wash 30 min.
- 2d. Dry in oven at about 110 °C for 20 min. The 20×20 mm glasses can be stored in a sealed container, and ready to used to make the channels. The 22×40 mm glasses need more steps to clean.
- 3. For 22×40 mm glasses, fill H₂SO₄:H₂O₂:DIwater = 1:1:5 = 4ml: 4ml: 20 ml in each staining jar. Put the jars in oven/heater at 120° C for 30 minutes (Use a secondary container if put the jars in oven). Then wash with DIwater for 30 times or more.
- 4. Dry in oven at ~ 110 °C for 20 min or blow with N₂.
- 5. Plasma to clean the cover glasses for 20 min.

Function by Glutaralderhyde

- 1. Fill APTES (1% in methanol) for 1 hour. Wash with methanol for 3 times. Dry in oven at about $110\,^{\circ}\text{C}$ for 20 min.
- 2. Fill Glutaralderhyde (1% in DIwater) for 1 hour. Ultrasonicate 2 times in DIwater. Blow the cover glasses to dry with N_2 .

Function by Streptavidin in channels.

1. Make channels with double side tape and KE-445 silicone glue.



- 2. For each channel, 50 μ L of Streptavidin diluted in PBS buffer, ~20 ng/ μ l (or less), overnight in a sealed dish. Excessive water in needed in the dish to prevent the channel from getting dry.
- 3. For each channel, 50 μ L of 100 mM Tris-HCl pH 7.4 for 2 hours.
- 4. For each channel, $50~\mu L$ of BSA buffer (20 mM Tris-HCl, 50~mM NaCl/KCl, 2% BSA), store in 4 degree in a sealed dish. Excessive water in need to prevent the channel from getting dry. Fill in more 2% BSA buffer and water each for 3 days.