SpGPCW: Spatially Varying Gaussian Process Model for Critical Window Estimation

SpGPCW_Example

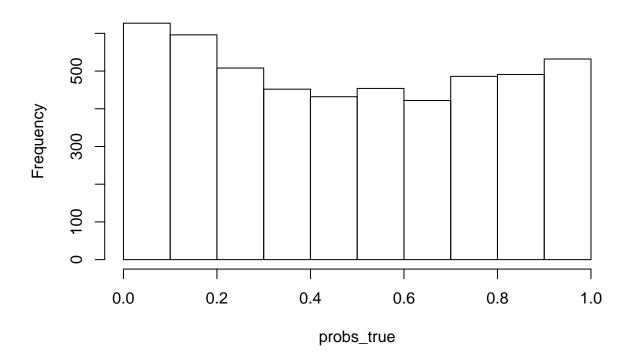
set.seed(2365)

- [1] Simulate data from the proposed model:
 - Setting the reproducibility seed and initializing packages for data simulation:

```
library(SpGPCW)
library(mnormt) #Multivariate normal distribution
library(boot) #Inverse logit transformation
library(spdep) #Creating a grid
## Loading required package: sp
## Loading required package: Matrix
## Loading required package: spData
## To access larger datasets in this package, install the spDataLarge
## package with: `install.packages('spDataLarge',
## repos='https://nowosad.github.io/drat/', type='source'))`
  • Setting the global data values:
n<-5000 #Sample size
m<-25 #Number of exposure time periods
g<-4 #Size of square spatial grid
s<-g^2 #Number of spatial locations
grid<-cell2nb(nrow=g,
              ncol=g,
              type="rook",
              torus=FALSE) #Evenly spaced grid
neighbors<-nb2mat(grid,
                  zero.policy=TRUE,
                  style="B") #Adjacency matrix
MCAR<-diag(rowSums(neighbors)) -</pre>
      neighbors
site_id<-rep(s, times=n)</pre>
for(j in 1:s){
   site_id[(1 + floor(n/s)*(j-1)):(floor(n/s)*j)] < -j
z<-matrix(0, nrow=n, ncol=m)</pre>
for(j in 1:s){
   z[(site_id == j),]<-matrix(rnorm(n=sum(site_id == j)),</pre>
                              nrow=sum(site_id == j),
                               ncol=m,
                               byrow=TRUE) #Exposure design matrices
for(j in 1:m){
  z[,j] <- z[,j] / IQR(z[,j]) #Data standardization (interquartile range)
  }
```

```
x<-matrix(1,</pre>
                                  nrow=n,
                                  ncol=2) #Covariate design matrix
x[,2] < -rnorm(n)
beta_true<- c(-0.10, 0.20)
sigma2_theta_true<-0.25
sigma2_eta_true<-0.05
phi_true<-0.20
Sigma_true<-sigma2_theta_true*chol2inv(chol(temporal_corr_fun(m, phi_true)[[1]]))
eta_true<-rmnorm(n=1,
                                                         mean=rep(0, times=m),
                                                          varcov=Sigma true)
eta_true<-eta_true -
                                 mean(eta_true)
rho_true<-0.45
theta_true<-rmnorm(n=1,</pre>
                                                                mean=rep(eta_true, times=s),
                                                                varcov=chol2inv(chol(kronecker((rho_true*MCAR + (1 - rho_true)*diag(s)),
                                                                                                                                                                           chol2inv(chol(Sigma_true))))))
for(j in 1:s){
          theta_true[(1 + (j-1)*m):(m*j)] < -theta_true[(1 + (j-1)*m):(m*j)] - -theta_true[(1 + (j-1)*m]:(m*j)] - -theta_true[(1 
                                                                                                                           mean(theta_true[(1 + (j-1)*m):(m*j)])
          }
logit_p_true<-rep(0, times=n)</pre>
for(j in 1:s){
          logit_p_true[site_id == j]<-x[(site_id == j),]%*%beta_true +</pre>
                                                                                                          z[(site_id == j),]%*%theta_true[(1 + (j-1)*m):(j*m)]
probs_true<-inv.logit(logit_p_true)</pre>
hist(probs_true)
```

Histogram of probs_true



• Simulating the analysis dataset:

[2] Fit SpGPCW to estimate spatially varying critical windows of susceptibility:

```
## Progress: 5%

## rho Acceptance: 25%

## phi Acceptance: 24%

## **************

## Progress: 10%

## rho Acceptance: 26%

## phi Acceptance: 27%

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## Progress: 15%

## rho Acceptance: 25%

## phi Acceptance: 27%

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## Progress: 20%

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## phi Acceptance: 26%

## phi Acceptance: 26%
```

```
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## Progress: 30%
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## *********
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## Progress: 90%
```

```
## rho Acceptance: 27%
## phi Acceptance: 26%
## **********
## Progress: 95%
## rho Acceptance: 27%
## phi Acceptance: 26%
## *********
## Progress: 100%
## rho Acceptance: 27%
## phi Acceptance: 26%
## *********
[3] Analyzing Output:
par(mfrow=c(2,2))
plot(results$beta[1, 1001:10000],
     type="1",
     ylab="beta0",
     xlab="Sample")
abline(h=beta_true[1],
       col="red".
       lwd=2) #True value
plot(results$beta[2, 1001:10000],
     type="1",
     ylab="beta1",
     xlab="Sample")
abline(h=beta_true[2],
       col="red",
       lwd=2) #True value
plot(rowMeans(results$eta[,1001:10000]),
     eta_true)
abline(0, 1)
theta<-simplify2array(results$theta)</pre>
theta_post_means<-rep(0, times=(s*m))</pre>
counter<-0
for(j in 1:s){
   for(k in 1:m){
      counter<-counter + 1</pre>
      theta_post_means[counter] <-mean(theta[j,k,1001:10000])</pre>
      }
    }
plot(theta_post_means,
    theta_true)
abline(0, 1)
```

