

Polarisation-resolved super-resolution microscopy

by

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Cover image: artefacts of an old actin-phalloidin stained sample.



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Abstract

This project aims to implement polarisation microscopy on the Tegenfeldt STED (stimulated emission depletion) microscope. STED microscopy is a targeted optical super-resolution method that can attain sub-diffraction resolution using visible light. This is now complemented with polarisation microscopy, which can measure orientation of a fluorophore and, by extension, the molecule it is bound to. The current setup is one of the first systems in the world to combine STED and polarisation, but its polarisation microscopy capability has never been tested.

In this thesis, the effect of the various optical components in the microscope on the polarisation state of light were characterised and the system was calibrated to perform several variations of polarisation microscopy. In the process, a new method was developed that applies the operating principle of STED to increase resolution in the polarisation domain, called pSTED for polarisation-dependent stimulated emission depletion. The preliminary results are promising, but more work is required to demonstrate its potential as a new and innovative microscopy method.

All of the above methods have been applied to biological samples of human cell lines in which the actin cytoskeleton was fluorescently stained. The cells in these samples have been exposed to Yersinia bacteria. Pathogenic members of the Yersinia genus, of which the plague-causing *Y. pestis* is a member, destroy the actin cytoskeleton. In the absence of large actin fibres, it has been found that actin can form self-organising patterns on the microscale. This discovery opened up an exciting line of research where polarisation microscopy will be highly applicable.

(find all references to I/me/we/us)

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Nomenclature

APD	Avalanche photodetector
AOM	Acousto-optical modulator
au	Arbitrary units
CCD	Charge-coupled device
DAPI	4',6-diamidino-2-phenylindole
DNA	Deoxyribonucleic acid
FLIM	Fluorescence lifetime imaging microscopy
FOV	Field of view
FWHM	Full width at half maximum
GFP	Green fluorescent protein
HSV	Hue, saturation, value
HWP	Half-wave plate
MPE	Maximum permissible exposure
NA	Numerical aperture
OD	Optical density (an OD2 filter reduces the light intensity by a factor of 10^2)
PBS	Polarising beam splitter
PMT	Photomultiplier tube
PSF	Point spread function
pSTED	Polarisation-dependent stimulated emission depletion microscopy
QWP	Quarter-wave plate
ROI	Region of interest
SARS-CoV2	Severe acute respiratory syndrome coronavirus 2
SiR	Silicon-rhodamine
SLM	Spatial light modulator
SNR	Signal-to-noise ratio
STED	Stimulated emission depletion microscopy
TCSPC	Time-correlated single photon counter

1

Introduction

Between 1900 and 2000, advances in public health have increased life expectancy by 25 years [1]. The Centers for Disease Control and Prevention (CDC) credits advances in molecular biology such as vaccinations, better prevention of infectious diseases, better treatment of coronary heart disease and stroke, and more [2]. Another pillar of molecular biology is the rapid development of SARS-CoV2 vaccines in the current pandemic [3, 4]. There are still many incurable diseases in the world, but by studying how they work, one can learn how cells and organisms function, and propose new treatments. Studying one disease can also guide treatment of another, as was the case when research into amyloidoses (a group of diseases including Alzheimer's disease, Parkinson's disease and diabetes type II, characterised by toxic amyloid protein aggregation), spun off a new direction of research into cancer treatment [5].

This thesis is part of a research programme that studies Yersinia. Yersinia is a genus of bacteria that includes *Y. pestis*, which causes the plague, estimated to have killed millions of people in the Byzantine empire in the 6th century CE, and again 50 million in Europe in the 14th century, which was at that point about one third of the entire population [6]. Vaccinations are now available and the incidence is low, less than 1000 cases per year worldwide [7]. While there is no direct clinical need for more research into Yersinia, it is still a fascinating subject of fundamental importance, since it has an interesting mode of action. Yersinia bacteria secrete toxins into the cell that break down the actin network [8]. The actin cytoskeleton works like the bones and muscles of a cell and is therefore vital to all kinds of processes, including embryonic development, cancer metastasis, and the immune system [9–13]. Furthermore, it has been discovered that the absence of large actin fibres reveals self-organising actin microstructures [14]. Their presence was quite a surprise and studying them may lead to new fundamental discoveries about actin and its role in healthy cell function.

The goal of this thesis is to set up a polarisation-resolved super-resolution fluorescence microscope to study these actin microstructures. Microscopy –

imaging structures at microscopic scales – is a wide and varied field. Microscopy was born in the sixteen hundreds, with Antonmevan Leeuwenhoek’s discovery of bacteria and other single-celled organisms [15]. Over time, the resolution of microscopes has strongly improved, and as the design and performance of lens systems got better and better, they stopped being the resolution bottleneck. In 1873, Ernst Abbe determined that the best possible focus that a microscope can reach is limited by the wavelength of the light used [16]. This meant that microscopes of the time were limited to a resolution of roughly 200 nm (assuming focused blue light). It was long believed that the Abbe limit was a fundamental limit of nature, and that the only way around it was to use light of a different wavelength. This is one of the reasons for the development of electron microscopes, as quantum mechanics explains that accelerated electrons have a much shorter wavelength than visible light [17].

However, using visible light for microscopy has some major advantages that electron microscopy cannot provide. Unlike with other methods, it is possible to tag specific protein species and other relevant molecules in the cell with a fluorescent label. For that reason, fluorescence microscopy has been an invaluable tool in modern biology [18]. There are thousands of small organic fluorescent labels available, and more are still being developed [19, 20]. The introduction of GFP and other fluorescent proteins was a remarkable development in the field, as labels can now be genetically fused to proteins of interest [21, 22]. In the year 2020 alone, over twenty thousand papers indexed in the PubMed database mention fluorescence in their title or abstract.

Fluorescence microscopy is not only capable of reporting on the location of a fluorophore, but also on its orientation. This is because fluorophores have a transition dipole moment; they will not absorb light that is polarised perpendicular to that dipole, and will only emit radiation polarised parallel to it. Therefore, fluorescence emission anisotropy is related to the dipole moment, which is determined by the orientation of the molecule that the fluorophore is bound to.

The group in which this project was conducted, led by Jonas Tegenfeldt, already owned a stimulated emission depletion (STED) microscope, capable of performing fluorescence microscopy below the Abbe limit. It was fitted with some extra optics to enable polarisation microscopy by the supplier (Abberior GmbH, Germany), but as they had never done STED and polarisation microscopy at the same time, these capabilities were untested at the start of this thesis project. Therefore, the main goal of this thesis was to test the setup and properly calibrate it for polarisation microscopy experiments. Most of my learnings and contributions will hence be found in the Background and Methods sections of this report. Along the way, a new microscopy protocol was developed that applies the working principle of STED to the polarisation domain to increase polarisation resolution: polarisation-dependent STED (pSTED). This will be treated in the Methods section.

2

Background

This section introduces concepts and techniques essential to this thesis. First, the emergence of fluorescence from discrete molecular energy states will be discussed. Then, the resolution-limiting diffraction limit will be introduced, as well as two methods to get around that limit. The first method, STED microscopy, uses targeted quenching of fluorescent molecules (also called dyes or fluorophores) at the edge of the focal spot to effectively narrow down the spot size. The second method exploits the polarisation state of excitation or emission light to measure the orientation of certain structures in a sample, even if their size is below the resolution limit.

2.1. Fluorescence microscopy

In short, fluorescence is a phenomenon in which a molecule absorbs a photon of one wavelength, spends a short amount of time (a couple of nanoseconds) in an excited energy state, and then emits a photon of a wavelength longer than the first one. The difference in wavelength, called the Stokes shift, is essential to perform microscopy, and the fact that molecules have discrete energy states is key to generate that difference.

Consider a molecule with two electronic energy states, each of which has a number of vibrational energy levels as shown in Figure 2.1. When the molecule is in its ground state (the lowest available energy level, S_0), it can absorb photons only if the energy they carry matches the difference between the ground state and another energy level. From an excited state (S_1), the system can relax into lower vibrational levels without emitting radiation, after which it emits a photon and relaxes back to the ground state. Two possible paths are shown in the figure. Because some energy is lost in a non-radiative way, the emitted photons will carry less energy than the absorbed photon, resulting in the wavelength difference mentioned before. This is called the Stokes shift.

This can be made more rigorous by considering what happens to a simplified two-level quantum system such as a hydrogen atom subject to a Hamiltonian H_0 . Denote the energy states of the electron with $|1\rangle$ and $|2\rangle$. These are

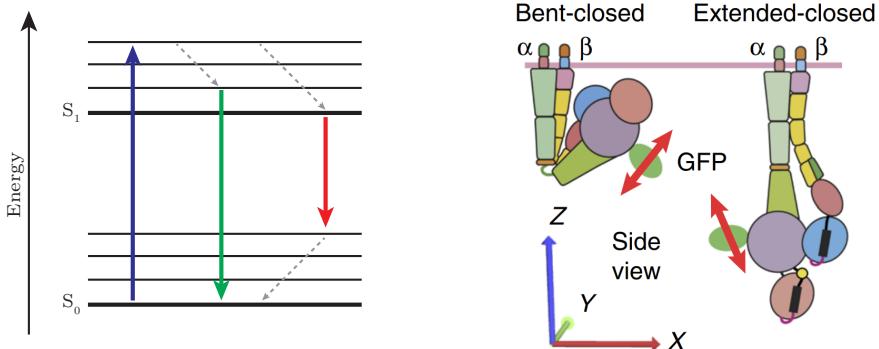


Figure 2.1: **Left:** Illustration of the discrete energy states of a molecule and some possible transitions between them. The difference between energy levels determines the wavelength and colour of the absorbed or emitted photon. S_1 and S_2 are electronic energy levels that each contain a set of vibrational states. **Right:** Illustration of how a fluorophore's dipole moment (red arrow) can report on the orientation of the molecule it is bound to. In this case, the fluorophore is GFP (green fluorescent protein) and the target is a transmembrane integrin protein with subunits α and β . Image by Nordenfelt et al. [23].

eigenstates of the Hamiltonian with energy $\epsilon_m e$. When exposed to an electric field $\mathbf{E} = \mathbf{E}_0 \cos \omega t$, the Hamiltonian is perturbed: $H' = H_0 + \boldsymbol{\mu} \cdot \mathbf{E}(t)$, where $\boldsymbol{\mu} = -e\mathbf{r}$. If the system is in the ground state at time $t = 0$, the probability to find it in the excited state at time t is given by

$$|c_2(t)|^2 = \left| \langle 2 | e^{-iHt/\hbar} | 1 \rangle \right|^2. \quad (2.1)$$

For weak fields, a first-order approximation is

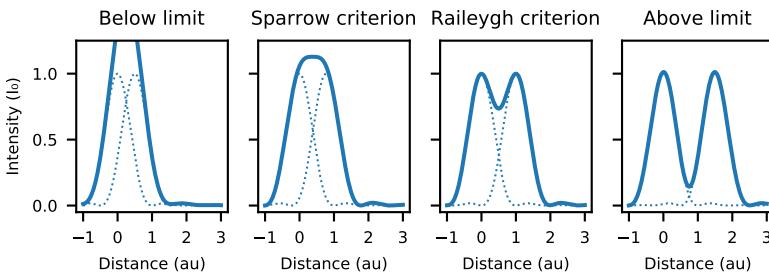
$$|c_2(t)|^2 = \left| \frac{\boldsymbol{\mu}_{21} \cdot \mathbf{E}_0 \sin((\omega_0 - \omega)t/2)}{\hbar(\omega_0 - \omega)} \right|^2, \quad (2.2)$$

where $\boldsymbol{\mu}_{21} = \langle 2 | \boldsymbol{\mu} | 1 \rangle$ and $\omega_0 = (\epsilon_2 - \epsilon_1)/\hbar$. More detail can be found in C.J. Foot [24]. This equation determines that there are two conditions for a transition. Firstly, the frequency of the electric field ω must match the energy difference between the states ω_0 . (Note that the sign of ω_0 is unspecified. This process can represent absorption as well as stimulated emission.) Secondly, the electric field is most effective at driving the transition when it is polarised along the transition dipole moment $\boldsymbol{\mu}_{21}$. For example, if an electron is confined to a chemical bond, it will not interact with electric fields polarised orthogonal to that bond. The precise energy levels available depend on the molecule and its environment. Therefore, every fluorescent molecule has a unique absorption and emission spectrum, which needs to be considered when planning a fluorescence experiment.

2.2. The diffraction limit

The resolution of fluorescence microscopy used to be limited by light diffraction. Consider the diffraction limit in one of the simplest possible setups:

	Criterion	Definition
Rayleigh	The first minimum of one point's Airy function coincides with the maximum of another	$d_{xy} = .61\lambda/NA$
FWHM	The width of the Airy function at half of the peak height	$d_{xy} = .51\lambda/NA$
Abbe	Resolving all groups of the 19-group Nobert plate [16, 25]	$d_{xy} = .5\lambda/NA$
Sparrow	The distance between fluorophores at which the central maximum splits	$d_{xy} = .47\lambda/NA$

Table 2.1: Some common definitions of the resolution limit d_{xy} .**Figure 2.2:** Illustration of the diffraction limit. Below the resolution limit, two fluorophores (with PSF dashed) appear under a microscope as a single peak (solid, sum of the intensities of the two fluorophores). Distance is measured in multiples of the Rayleigh limit (.61 λ/NA).

an epifluorescence microscope. Epifluorescence microscopy is fairly similar to bright field microscopy; it just uses excitation light of a well-defined wavelength instead of white light. The sample, which is stained with a fluorescent dye, is illuminated at a wavelength that dye can absorb. The dye molecules will then emit photons of longer wavelengths in their emission spectrum, which are collected by the microscope objective. Before imaging by a CCD camera, scattered laser light can be filtered away using a dichroic mirror that passes emission light and reflects excitation light.

It used to be the case that microscopes were limited in resolution by the quality of their lenses and the pixel density of the CCD, but there is a much more fundamental limit, first described by Ernst Abbe. This limit is caused by the presence of an aperture. When a beam of light goes through an aperture of finite size, such as a lens, it cannot be focused onto an infinitesimally small point. Instead, it will generate a spot with a radius approximately equal to $\lambda/2NA$, where λ is the wavelength of the light and $NA = n \sin \theta$ is the numerical aperture, the product of the index of refraction and the sine of the half-cone angle θ of the acceptance cone.

In particular, a circular aperture will convert point sources in the sample plane to Airy patterns in the image plane. Mathematically, this can be described by a convolution product between the point spread function (PSF)



Figure 2.3: Working principle of STED microscopy. Fluorophores at the edge of the excitation PSF are quenched by stimulated emission, effectively resulting in a narrower PSF. Both beams are circularly polarised.

h_{det} and the fluorophore distribution ρ ,

$$I(\mathbf{x}) = (h_{det} * \rho)(\mathbf{x}) := \int d\mathbf{x}' h_{det}(\mathbf{x} - \mathbf{x}') \rho(\mathbf{x}'), \quad (2.3)$$

where the triple integration over the three components of \mathbf{x}' is implied, resulting in an image $I(\mathbf{x})$. When two fluorophores are close enough together that their PSFs overlap, they appear as a single object instead of two separate ones. This happens when the distance between them is around $\lambda/2NA$, and that is what determines the resolution of a microscope. The exact resolution depends on your definition of this minimum resolvable distance. There are several definitions, but they are all proportional to λ/NA . Some of them are listed in Figure 2.1 and visualised in Figure 2.2.

For a long time, physicists thought this limit was practically unavoidable [26], but in the next sections, two ways will be discussed in which one can get information from a system below the resolution limit. The first method (STED microscopy) directly increases the image resolution. The excitation light is still subject to the Abbe limit, but the extra laser effectively improves the size of the focal spot. The second is indirect and requires playing with a new aspect of light (polarisation) that allows you to get orientational information about structures that are otherwise below the resolution limit.

2.3. Super-resolution microscopy

There are several ways to increase image resolution directly, many of which use the photophysics of individual dyes to turn off a subset of them during imaging. Some methods do this stochastically, such as STORM (stochastic optical reconstruction microscopy) and PALM (photoactivated localisation microscopy) [27, 28]. These localisation-based methods activate a random subset of fluorophores, such that the average distance between them is above the resolution limit. Then they are imaged with a camera and the centres of their PSFs are estimated. The precision of that estimate scales with $1/\sqrt{N}$ where N is the numbers of photons collected.

On the other hand, there are targeted techniques such as STED (stimulated emission depletion), GSD (ground state depletion), RESOLFT (Reversible saturable optical fluorescence transitions), and more [29–31]. Generally, they

work by turning off fluorophores on the outer edge of the focal spot (depletion). The resolution achieved is dependent on the intensity of the depletion laser. This is a case of targeted instead of stochastic photoswitching. The Tegenfeldt group own a STED microscope, so that is what we will focus on in this section.

In essence, a STED microscope is a scanning confocal microscope with an extra laser that can selectively deplete fluorescence by stimulated emission. Its working principle is shown in Figures 2.3 and 2.4. In a scanning confocal microscope, the excitation laser does not illuminate the whole sample at once, but is scanned over it. This means only fluorophores in an Airy disk around the focus will be excited. Furthermore, the detector is now comprised of a pinhole and a photodetector (not a camera), which filters out most of the out-of-focus light [32]. Therefore, the PSF of a confocal microscope is the product of the laser PSF h_{ex} and the detection probability h_{det}

$$h_{conf}(\mathbf{x}) = h_{ex}(\mathbf{x}) \cdot h_{det}(\mathbf{x}), \quad (2.4)$$

where the most important characteristic of $h_{det}(\mathbf{x})$ is that it is quite narrow in the z direction. The smaller the detection pinhole, the stronger this effect. However, a smaller detection pinhole also means that even a fraction of the light from the focal plane will be rejected from the photodetector. Therefore, there is a trade-off between z resolution and the signal-to-noise ratio (SNR).

Even though both of these PSFs are diffraction-limited, a STED microscope can reach an arbitrarily small resolution [33]. It does so by illuminating the sample with a donut-shaped laser at a wavelength longer than the emission wavelength. Referring back to Figure 2.1, the blue laser excites the fluorophores that emit in green, but a red transition is also allowed. Under illumination with a red laser, this transition is made more favourable by the process of stimulated emission, first postulated by Einstein in 1926 [34]. This way, fluorophores at the edge of the excitation PSF can be prevented from emitting green light, while fluorophores at the centre do not experience stimulated emission, which reduces the width of the effective point spread function. An illustration of this effect is given in Figure 2.3.

Mathematically, the STED beam depletes fluorescence according to

$$\eta(\mathbf{x}) = \exp(-\sigma_{dep} h_{dep}(\mathbf{x})), \quad (2.5)$$

which relates the fraction of fluorescence remaining as a function of the depletion intensity h_{dep} . Therefore, the STED point spread function can be expressed as

$$h_{STED} = h_{ex}(\mathbf{x}) \cdot \eta(\mathbf{x}) \cdot h_{det}(\mathbf{x}), \quad (2.6)$$

which can be read as “the probability for a dye to be excited, times the probability for it not to be quenched by stimulated emission, times the probability for fluorescence emission to be detected by the CCD”. Even though every individual PSF ($h_{ex}(\mathbf{x})$, $h_{dep}(\mathbf{x})$ and $h_{det}(\mathbf{x})$) is diffraction-limited, the fact that you need to multiply them results in a narrowing of the effective PSF below the Abbe limit. This means that the extent of the resolution improvement depends on the intensity of the depletion laser I_{dep} . Harke et al. approximate

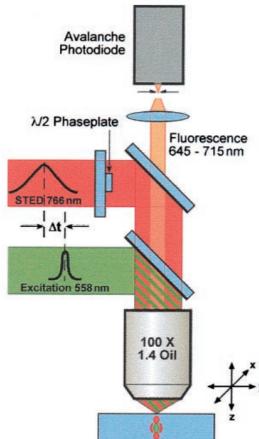


Figure 2.4: The rough layout of a STED microscope. The excitation laser is focused on the sample, and the STED laser forms a donut pattern around the focal spot. Emitted light is collected by the objective and focused on a pinhole before it is detected by an APD. Figure from Klar et al. [29].

the STED resolution as

$$d_{STED} = \frac{d_c}{\sqrt{1 + d_c^2 a^2 \frac{I_{dep}}{I_{sat}}}}, \quad (2.7)$$

where d_c is the resolution limit of a confocal microscope, a the steepness of the donut pattern (i.e. high a means very tight central minimum in the donut-shaped intensity), and I_{sat} is the depletion intensity at which fluorophore brightness is reduced by half [35].

Although STED can reach an arbitrary resolution under optimal circumstances, the resolution reached in biological specimens is more usually around 50 nm [33, 36]. If information is desired at an even smaller scale using fluorescence microscopy, then other methods are required. There are several reasons the resolution of STED in biological samples has not been able to reach the single-nanometre scale. One is that these samples consist of cells in a water-based solution, which causes a strong mismatch in the index of refraction between the sample and the optics used (commonly glass and oil). Another is the limited photostability of the dyes. Every time a fluorophore absorbs a photon, there is a probability that a photochemical event occurs that damages (bleaches) the fluorophore. Good fluorophores can typically absorb 10 000 to 40 000 photons before this happens [37]. Therefore, high excitation powers result in high bleaching rates. Photobleaching is mainly due to interactions with reactive oxygen species and is therefore not only determined by the properties of the fluorophore, but also the environment. Photobleaching can also be induced by other causes, such as the absorption of multiple photons. Refer to Diaspro et al. for an overview of photobleaching [38]. When imaging live cells, phototoxicity also needs to be taken into account. Because molecules

other than the targeted fluorophores can also absorb laser light (possibly resulting in autofluorescence), incident radiation can be toxic for the cell, for the same reasons as it can bleach a fluorophore [37]. Therefore, photobleaching and phototoxicity both limit the laser power that can be applied.

2.4. Polarisation microscopy

Polarisation microscopy uses the vectorial nature of light – the fact that light consists of electromagnetic waves that with a certain polarisation – to get orientational information of fluorophores below the resolution limit. Other methods to get information on distances below the resolution limit include fluorescence resonance energy transfer (FRET), for example, which is useful for measuring distances between two fluorophores on the order of nanometres [39].

Among others, light polarisation microscopy has been used to measure how the structure of DNA changes when it is subject to a strong stretching force, how integrin proteins respond to an applied force and the order of molecules embedded in the cell membrane [23, 40–42]. In this section, we will first introduce the concept of light polarisation, then discuss how it can be used in a microscope, and finally mention some optical components that affect the light polarisation, which are crucial to conducting a polarisation microscopy experiment. For a more exhaustive introduction to polarisation microscopy, there are several good resources available [43–45].

2.4.1. The polarisation ellipse.

Light is a transverse electromagnetic wave, meaning that there are oscillations of the electric and magnetic field along the path of a light ray, and that these oscillations are orthogonal to the propagation direction. In other words, if the light propagates along \mathbf{k} , the electric and magnetic fields \mathbf{E} and \mathbf{B} must satisfy $\mathbf{E} \cdot \mathbf{k} = \mathbf{B} \cdot \mathbf{k} = 0$. (The fields themselves are also orthogonal to each other, so \mathbf{B} can be neglected without compromising this analysis.)

For the sake of simplicity, consider a ray propagating in the z direction. The electric field at any point in space and time can be written as

$$E_x(z, t) = E_{0x} \cos(kz - \omega t + \phi_x), \quad (2.8)$$

$$E_y(z, t) = E_{0y} \cos(kz - \omega t + \phi_y). \quad (2.9)$$

where \mathbf{E}_0 is the amplitude of the oscillation, k is the wavenumber (the length of \mathbf{k}), ω is the radial frequency and ϕ is an arbitrary phase. Note that the wavenumber and the frequency are related to each other through the speed of light c , since $\omega = kc = \hbar c / \lambda$. Note also that the x and y components can have a phase difference.

Letting $\delta = \phi_y - \phi_x$, it can be shown that

$$\left(\frac{E_x}{E_{0x}} \right)^2 - 2 \cos \delta \frac{E_x}{E_{0x}} \frac{E_y}{E_{0y}} + \left(\frac{E_y}{E_{0y}} \right)^2 = \sin^2 \delta, \quad (2.10)$$

which is the equation for an ellipse. This means that at any point in time, the point (E_x, E_y) lies on the ellipse defined by the equation above, which is

Polarisation state	E_{0x}	E_{0y}	δ	Jones vector
Linear along x	1	0	any	(1, 0)
Linear along y	0	1	any	(0, 1)
Linear at $\pm 45^\circ$	1	± 1	0	(1, ± 1)
Circular (right-handed)	1	1	$\pi/4$	(1, i)
Circular (left-handed)	1	1	$-\pi/4$	(1, $-i$)

Table 2.2: List of a number of polarisation states (not normalised).

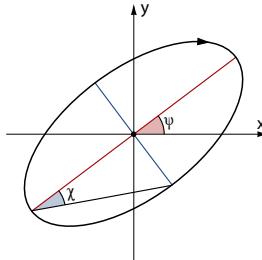


Figure 2.5: The meaning of ψ and χ in the context of the polarisation ellipse. (Figure by Wikipedia user Inductiveload)

called the polarisation ellipse and is completely determined by E_{0x} , E_{0y} and δ . Together, these values determine whether the polarisation is linear, circular, or something in between. Refer to Table 2.2 for an overview.

In general, the polarisation ellipse can be defined by means of two angles: the orientation ψ and ellipticity χ , as shown in Figure 2.5. They can be calculated from $\alpha = \arctan(E_{0y}/E_{0x})$ and the phase difference δ using

$$\tan 2\psi = \tan 2\alpha \cos \delta, \quad (2.11)$$

$$\sin 2\chi = \sin 2\alpha \sin \delta \quad (2.12)$$

This definition of χ is dependent on the ratio of the maximum and minimum amplitude of the electric field as is presented in Figure 2.5. One can define a similar value based on the corresponding intensities. Since $I = |\mathbf{E}_0|^2$,

$$\chi_m e = \arctan I_{max}/I_{min} = \arctan \left[(E_{max}/E_{min})^2 \right]. \quad (2.13)$$

To distinguish between them, we will be using χ_E (χ as defined in Equation 2.12) and $\chi_m e$ (as defined in Equation 2.13) from now on. These are useful concepts because they are relatively easy to calculate from intensity measurements. Therefore, we will use them when we characterise laser beams in the setup.

2.4.2. Microscopy.

Why is polarisation relevant to microscopy? Well, the probability of excitation is dependent on the polarisation of incoming light. Since a fluorophore can be

considered a dipole, it will not interact with radiation that is orthogonal to the transition dipole moment of the fluorophore (see discussion surrounding Equation 2.2). If the excitation laser is polarised along an angle ψ and the dipole is oriented along θ , the intensity of light emitted by that fluorophore will satisfy

$$I(\psi, \theta) \propto \cos^2(\psi - \theta). \quad (2.14)$$

This is Malus's law. Analogously, light emitted from a fluorophore is always linearly polarised parallel to the transition dipole moment. One can place a linearly polarising filter in front of the detector to measure a fluorophore's orientation. If the polariser emits light polarised at an angle ψ , then the intensity measured at the detector also follows Malus's law, meaning that these two setups are analogous (not taking into account depolarisation effects in an experimental setup). As an example, see Figure 4.4. Dipole excitation by circularly polarised light is usually not dependent on θ , but there are fluorophores that are sensitive to the handedness of circularly polarised light [46].

Common polarisation microscopy protocols include: rotating a linearly polarised excitation laser, rotating a polarising filter in the excitation beam path, or doing both. Historically, polarisation measurements are performed by excitation of the sample with a linearly polarised laser and measuring the fractions of emission intensity that were parallel and orthogonal to the excitation light. With those, one can calculate the anisotropy of a sample [47]. Other polarisation microscopy methods will be presented in Section 3.2.

2.4.3. Jones calculus.

Lasers are usually linearly polarised, but it is possible to manipulate their polarisation state to perform different variations of polarisation microscopy. There are optical elements such as waveplates that can do this for us, and the best way to understand them is through Jones calculus. This is an incredibly useful way to model light polarisation, but it does require expressing the electric field as a complex function:

$$\mathbf{E}(z, t) = \mathbf{E}_0 e^{i(kz - \omega t)}. \quad (2.15)$$

In the following analysis, \mathbf{E} will be treated as a two-dimensional vector with only an x and y component, since $E_z = 0$. The phase difference between the two components is now contained in \mathbf{E}_0 , which can be expressed as

$$\mathbf{E}_0 = \begin{pmatrix} E_{0x} \\ E_{0y} e^{i\delta} \end{pmatrix}. \quad (2.16)$$

Therefore, \mathbf{E}_0 , the Jones vector, contains all information about the polarisation state of a beam. The Jones vectors for some special polarisation states are listed in Table 2.2.

The usefulness of Jones calculus lies in its ability to represent optical components as matrices acting on this vector. For example, a polariser that transmits x -polarised light has the following matrix form:

$$S_p = \begin{pmatrix} 1 & 0 \\ 0 & 0 \end{pmatrix}. \quad (2.17)$$

It is easy to verify that $S_p \mathbf{E}_0 = E_{0x}$.

Mirrors also affect the polarisation of incident light. In this case, it is useful to define a new coordinate system. Let k be the propagation direction, p the direction that is orthogonal to k , but in the plane of incidence (the plane formed by k and the normal to the mirror surface, i.e. p is parallel to the plane of incidence). Finally, let s be orthogonal to k and p (s stands for the German word senkrecht). Mirrors flip the s component of the field polarisation, while keeping p unchanged. So, a mirror whose surface is parallel to the x -axis has a Jones matrix of the form

$$S_{mx} = \begin{pmatrix} 1 & 0 \\ 0 & -1 \end{pmatrix}. \quad (2.18)$$

Another important type of optical component in the setup is a waveplate. Waveplates or phase retarders are made out of birefringent crystals such as quartz, in which the index of refraction a ray of light experiences is dependent on its polarisation. This happens when a crystal structure lacks cubic symmetry. When the coordinate system is aligned with the crystal axes, it could be the case that $k_x = k_z = k_o$ (the ordinary axes), while $k_y = k_e$ (the extraordinary axis). k_e may both be greater or less than k_o . In these crystals, Equation 2.15 is no longer valid and should be replaced by

$$\mathbf{E}(z, t) = \begin{pmatrix} E_{0x} e^{i(k_o z - \omega t)} \\ E_{0y} e^{i(k_e z - \omega t + \delta)} \end{pmatrix}. \quad (2.19)$$

This can also be written in the form

$$\mathbf{E}(z, t) = \begin{pmatrix} E_{0x} \\ E_{0y} e^{i(\Gamma(z) + \delta)} \end{pmatrix} e^{i(k_o z - \omega t)}, \text{ where } \Gamma(z) = (k_e - k_o)z. \quad (2.20)$$

As one can see, a waveplate only imparts a delay on the y -component of a beam, depending on its thickness z and its birefringence. The phase factor common to both components can be neglected in order to represent the action of a waveplate by the following Jones matrix in the local coordinate system of the waveplate with

$$S_\Gamma = \begin{pmatrix} 1 & 0 \\ 0 & e^{i\Gamma} \end{pmatrix}. \quad (2.21)$$

The axes in this coordinate system can be called the fast and slow axes, where the field component along the slow axis is the one that incurs the phase delay.

Generally, waveplates are characterised by the relative delay they impart on polarisation components. Quarter-wave plates delay it by a quarter of a wavelength compared to the fast propagating ray, corresponding to $\Gamma = (2n + 1/2)\pi$ (for any integer n). Therefore, the Jones matrix of a quarter-wave plate satisfies

$$S_{\lambda/4} = \begin{pmatrix} 1 & 0 \\ 0 & e^{i\pi/2} \end{pmatrix} = \begin{pmatrix} 1 & 0 \\ 0 & i \end{pmatrix}. \quad (2.22)$$

Let's consider what happens to some specific cases. If x or y polarised light passes through a quarter-wave plate, its polarisation will not change. But light

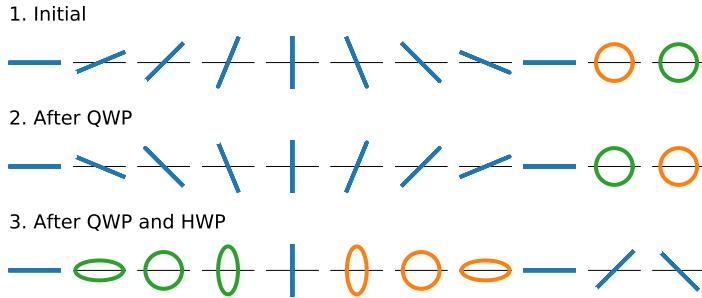


Figure 2.6: The effect of quarter-wave plates (QWP) and half-wave plates (HWP) on the polarisation ellipse. The fast axis is horizontal and indicated in the figure. The slow axis is vertical. Orange: left-handed polarisation. Green: right-handed polarisation. The ray is propagating into the page.

polarised along $+45^\circ$ (-45°) will be turned into left-handed (right-handed) light, and vice versa. Therefore, a quarter-wave plate can convert between linearly and circularly polarised light, as shown in Figure 2.6.

The second type of waveplate is a half-wave plate. It features a delay of $\Gamma = (2n + 1)\pi$, and its Jones matrix looks like

$$S_{\lambda/2} = \begin{pmatrix} 1 & 0 \\ 0 & e^{i\pi} \end{pmatrix} = \begin{pmatrix} 1 & 0 \\ 0 & -1 \end{pmatrix}, \quad (2.23)$$

which corresponds to mirroring the polarisation state about the x -axis. Another way to think about it is that a ray polarised along an angle ψ will be rotated by an angle -2ψ , and the handedness of circularly polarised light will be reversed, see Figure 2.6.

The power of Jones calculus lies in its ability to model the behaviour of a sequence of optical elements at arbitrary rotations. First, the Jones matrix needs to be defined for a rotated component. This is simply

$$S(\theta) = R(\theta) \cdot S \cdot R(-\theta), \quad \text{where} \quad R(\theta) = \begin{pmatrix} \cos \theta & -\sin \theta \\ \sin \theta & \cos \theta \end{pmatrix} \quad (2.24)$$

and θ is the angle of the component's fast axis with the lab coordinate system's x axis. As an example, x polarised light can be sent through a polarising filter at an angle θ , which results in

$$I(\theta) \propto \left| S_p(\theta) \cdot \begin{pmatrix} 1 \\ 0 \end{pmatrix} \right|^2 = \cos^2 \theta. \quad (2.25)$$

That is Malus's law, as defined before. The same behaviour can also be recovered using a fixed polarising filter and a half-wave plate at an angle $\theta/2$:

$$I(\theta) \propto \left| S_p(0) \cdot S_{\lambda/2}(\theta/2) \cdot \begin{pmatrix} 1 \\ 0 \end{pmatrix} \right|^2 = \cos^2 \theta. \quad (2.26)$$

Jones calculus has its limitations, the main one being that it cannot represent unpolarised light (an incoherent sum of different polarisations). Müller calculus, based on real-valued 4D Stokes vectors instead of the complex 2D Jones vectors, can handle it. As such, Müller calculus can also quantify depolarisation induced by non-ideal microscope optics. In this thesis, Jones calculus was used to develop a theoretical understanding of the optical elements in the STED microscope, for which Müller matrices are not necessary. In the next sections, the theory presented here will be used to implement polarisation microscopy on the Tegenfeldt STED microscope and will even be developed further to extend the concept of the optical point spread function into polarisation space.

3

Methods

This section discusses the optical setup, describes a number of ways in which one can perform polarisation microscopy and introduces pSTED, a method to increase the microscope resolution in the polarisation domain. Finally, the samples used in this thesis will be discussed.

3.1. Microscope setup

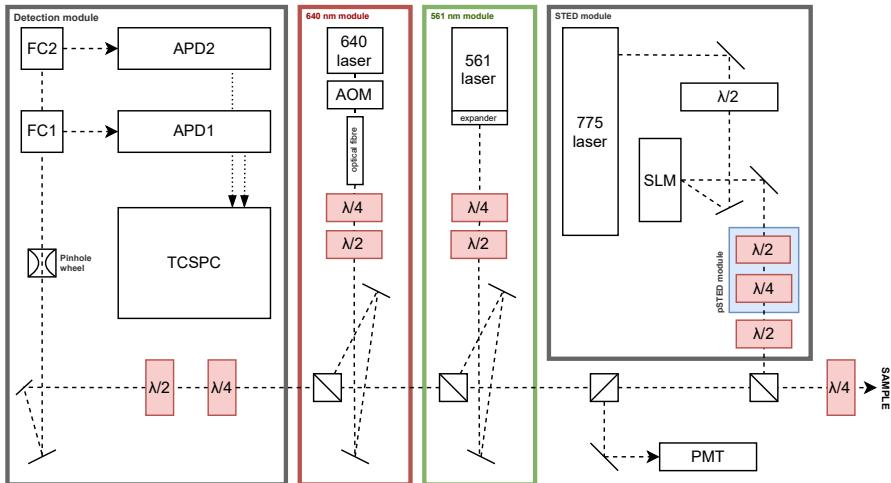


Figure 3.1: Schematic overview of the Tegenfeldt microscope. The sample is located in a microscope housing on the bottom right (outside this figure). FC1 and FC2 are swapable filter cubes. Half-wave and quarter-wave plates are denoted $\lambda/2$ and $\lambda/4$, respectively. Dashed lines represent the light path, except those that go to the TCSPC (those are digital connections). Red components are specifically designed to alter light polarisation. The components marked in blue (in the pSTED module) were installed during the project.

In short, the Tegenfeldt STED microscope is a scanning confocal fluorescence microscope constructed by Abberior Instruments GmbH (Germany). It

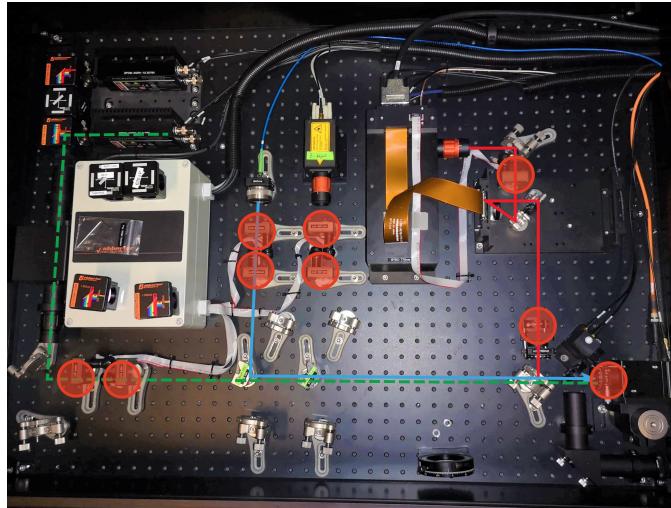


Figure 3.2: Photo of the optical enclosure, in the same orientation as Figure 3.1. The pSTED module is not shown. Waveplates are highlighted in red, and the beam paths of the 640 nm laser, the 775 nm laser, and the emission light are highlighted in blue, red, and green, respectively.

features two excitation lasers (at 561 nm and 640 nm) and one depletion laser at 775 nm for two-channel confocal or STED microscopy. It also contains a time-correlated single photon counter (TCSPC) for fluorescence lifetime imaging microscopy (FLIM) and a highly sensitive photomultiplier tube (PMT). Refer to Figures 3.1 and 3.2 for the layout of these optical elements. Samples are located inside an inverted Nikon microscope body (Ti-E, not shown in the figure) equipped with a piezo stage (M-687 PILine XY-stage system and P-736 PIano (Physik Instrumente) Z Microscope Scanner), a 60x, 1.4 NA oil immersion objective (Nikon Plan Apo) and a QUADScan beam scanner (Cambridge Technology).

The microscope has a number of waveplates that are responsible for manipulating the polarisation of laser light and fluorescence light (highlighted in red in Figure 3.1). In normal operation, all lasers are circularly polarised at the sample plane, but the waveplates in the excitation modules (640 and 561 nm) can be controlled through software to excite the sample with linearly polarised light. The waveplates in the STED module cannot, and therefore the depletion beam is always circularly polarised. Part of this project involved working with a linearly polarised depletion beam, so two waveplates were added in the STED module to achieve that result. Finally, a polarising beam splitter can be inserted into one of the filter cube housings in front of the APDs to measure the polarisation of photons emitted by fluorescence in the sample. In order for this measurement to make any sense, the waveplates in the detection module had to be set up correctly. More information about the laser and detection modules can be found in Appendix D.

3.2. Conventional polarisation microscopy: acquisition and analysis

Because the current setup offers so much control over the light polarisation on both the excitation and detection ends, one can perform polarisation microscopy in several different ways:

1. Measuring the intensity of emission components parallel and orthogonal to linearly polarised excitation (I_{\parallel} and I_{\perp}). This is a very established method of polarisation microscopy, and allows for making anisotropy images, where every pixel is calculated according to

$$r = \frac{I_{\parallel} - I_{\perp}}{I_{\parallel} + 2I_{\perp}}. \quad (3.1)$$

One advantage of this method is that it requires only one acquisition, whereas the following require multiple.

2. Detection of emission polarised at different angles after circularly polarised (or non-polarised) excitation.
3. Detection of total emission intensity as a function of the polarisation angle of excitation light.

Methods 1 and 2 require the POL cube to be placed in one of the filter cube housings. When placed in FC1, the POL cube reflects *s*-polarised light (vertical) into APD1 and transmits *p*-polarised light (horizontal) to be collected by APD2. Unfortunately, we cannot use these methods yet, since they rely on the action of the detection waveplates, even if they are not actively used during the acquisition. Once we do, however, the waveplates can be used to sample more than two angles during an acquisition. This is necessary to distinguish between light polarised along $\pm 45^\circ$, since these two angles give exactly the same intensity when sampling at 0° and 90° degrees.

The last method is not subject to those constraints, and is already achievable on the Tegenfeldt microscope. I have carried out some measurements and wrote code to analyse that data. The process of acquiring and analysing these images goes as follows:

1. Acquire a stack of images at different excitation polarisation angles θ_n , resulting in a three-dimensional array of intensity values I_{nxy} .
2. Align images in this stack with each other, as the excitation beam seems to move as a function of polarisation angle.
3. Compensate for photobleaching.
4. For every pixel, calculate the Fourier coefficient corresponding to a 180° -periodic signal.
5. Based on that information, construct a new image in the HSV (hue, saturation, value) colour space where pixel colour depends on the polarisation direction, the saturation shows the degree of polarisation and the brightness (value) shows the total intensity of a pixel.

For more information, refer to Section D.3. The results of this method are presented in Section 4.1

3.3. Polarisation-dependent STED microscopy (pSTED)

Since polarisation is important to both excitation and emission of a fluorophore, it stands to reason that stimulated emission should also be polarisation dependent. In fact, that is what the calculation in Dyba et al.'s calculation of STED resolution is based on [35, 48]. Moreover, excitation and stimulated emission are described by the same quantum mechanical process [24].

Would it then be possible to adapt the principle of conventional STED microscopy to increase the polarisation resolution of a microscope, instead of its spatial resolution? In this section, I will introduce a definition of polarisation resolution, a mathematical description of how this might be improved using pSTED, and describe how I adapted the microscope setup to perform pSTED.

The general idea is that one can define a vectorial PSF (although a better term is photon fluence) that takes light polarisation into account. In a conventional polarisation microscopy setup, the probability of excitation of a fluorophore is proportional to $\cos^2 \Delta$, where Δ is the angle between the excitation polarisation and the transition dipole moment. This results in a FWHM resolution for conventional polarisation microscopy of

$$d_\theta = 90^\circ. \quad (3.2)$$

By illuminating the sample with a depletion beam that is orthogonally polarised to the excitation laser, we can suppress fluorescence of fluorophores on the edge of this PSF, just like in conventional STED. This is illustrated in Figure 3.3.

What follows is strongly inspired by Dyba et al. [48]. Let $\mathbf{x} = (x, y, z)$ be the spatial coordinates, θ be an angle with the x axis in the xy plane (ranging from 0 to 2π) and ϕ be an angle with the z axis in the xz plane (ranging from $-\pi$ to $+\pi$). We can also introduce a generalised coordinate $\mathbf{y} = (\mathbf{x}, \theta, \phi)$ for a more convenient notation. Let the fluorophore population be described by the density $\rho(\mathbf{y})$. This function is normalised such that its integral is equal to the number of fluorophores. For example, a single fluorophore at the origin, whose transition dipole points in the x direction with no out-of-plane tilt would be described by the Dirac distribution $\rho_0 = \delta(\mathbf{y})$, defined as follows:

$$\delta(\mathbf{y}) = 0 \quad \text{if } \mathbf{y} \neq 0, \quad (3.3)$$

$$\int f(\mathbf{y})\delta(\mathbf{y} - \mathbf{y}_0) d\mathbf{y} = f(\mathbf{y}_0). \quad (3.4)$$

The excitation beam is described by the electric field amplitude \mathbf{E}_{ex} , polarised along θ_{ex} , and analogous for the depletion beam. Like Equation 2.3, the emission intensity measured when the laser is focused on \mathbf{x} and polarised along θ , ϕ is now a convolution integral where we also have to integrate over the angles θ and ϕ like

$$\begin{aligned} I_{em} &= A \int \sigma_{ex} |\hat{\mathbf{n}}_{\theta' \phi'} \cdot \mathbf{E}_{ex}(\mathbf{x} - \mathbf{x}')|^2 \rho(\mathbf{y}') d\mathbf{y}' \\ &= \int \sigma_{ex} I_{ex}(\mathbf{x}' - \mathbf{x}) |\hat{\mathbf{n}}_{\theta' \phi'} \hat{\mathbf{n}}_{\theta \phi}|^2 \rho(\mathbf{y}') d\mathbf{y}' \end{aligned} \quad (3.5)$$

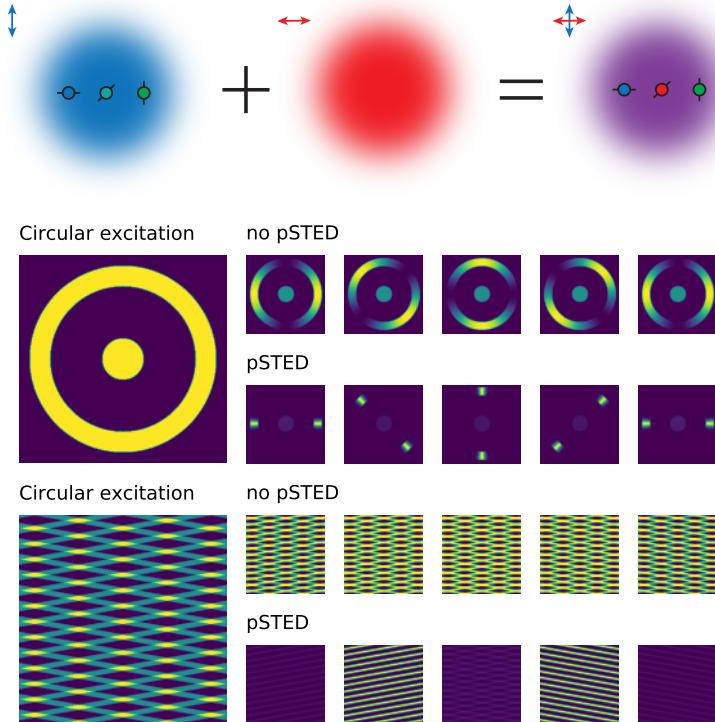


Figure 3.3: **Top:** Illustration of the working principle of pSTED. The arrows outside indicate the polarisation of the laser light. The circles represent fluorophores with their transition dipole moment indicated by a line. The orthogonal polarisation of the depletion beam suppresses fluorescence of the fluorophore oriented at 45° . **Middle:** A simulation of pSTED compared to other excitation schemes on a sample consisting of a non-polarised disk and a radially polarised ring, where the excitation polarisation goes from 0° to 180° . **Bottom:** Simulation of a grid of superimposed fibres, where the fluorophores are aligned with the fibre. If the fibre spacing is below the spatial resolution, then pSTED can show the presence of two different orientations. The excitation polarisation in this figure goes from -20° to 20° .

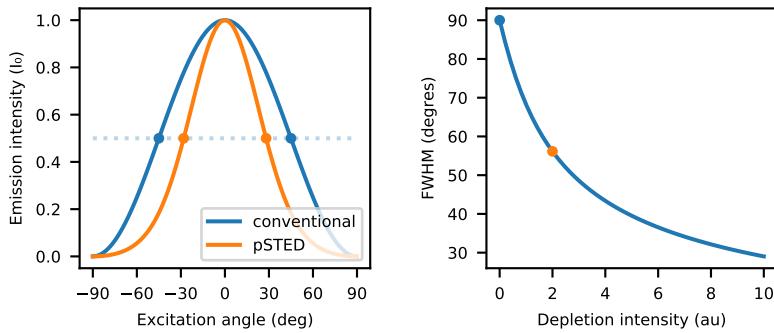


Figure 3.4: **Left:** Narrowing of the fluorophore response as a result of a depletion field (of intensity $I_{dep} = 2$, as compared to the right pane). **Right:** The FWHM angular resolution d_θ as a function of depletion intensity.

where $\hat{\mathbf{n}}_{\theta\phi}$ is a unit vector pointing in the direction of (θ, ϕ) and $A = \epsilon_0 c / 2$. ϵ_0 is a constant denoting the vacuum permittivity. The dot product can be expressed as

$$|\hat{\mathbf{n}}_{\theta'\phi'} \cdot \hat{\mathbf{n}}_{\theta\phi}|^2 = \cos^2(\theta - \theta') \cos^2(\phi - \phi'), \quad (3.6)$$

so we can express the emission intensity as a convolution product like

$$I_{em} = (\rho * h_{ex})(\mathbf{y}), \quad (3.7)$$

$$\text{where } h_{ex}(\mathbf{y}) = \sigma_{ex} I_{ex}(\mathbf{x}) \cos^2 \theta \cos^2 \phi. \quad (3.8)$$

If we have a constant laser intensity I_{ex} that is polarised along θ_{ex} without any out-of-plane polarisation, this reduces to

$$I_{em} = \sigma_{ex} I_{ex} \cos^2 \theta_{ex} \quad (3.9)$$

for the distribution ρ_0 . I_{em} is maximal when the excitation polarisation is in the x direction, as expected. From this equation, we can derive the same angular resolution as before, but with more rigour.

Now, we need to include the effect of the polarised depletion field, which amounts to adding an exponential factor under the integral

$$I_{em} = A \int \sigma_{ex} |\hat{\mathbf{n}}_{\theta'\phi'} \cdot \mathbf{E}_{ex}(\mathbf{x} - \mathbf{x}')|^2 \exp(-\sigma_{dep} |\hat{\mathbf{n}}_{\theta'\phi'} \cdot \mathbf{E}_{dep}(\mathbf{x} - \mathbf{x}')|^2) \rho(\mathbf{y}') d\mathbf{y}'. \quad (3.10)$$

In essence, this is equivalent to multiplying the kernel h_{ex} with the function η

$$h(\mathbf{y}) = h_{ex}(\mathbf{y}) \cdot \eta(\mathbf{y}) \quad , \text{ where} \quad (3.11)$$

$$\eta(\mathbf{y}) = \exp(-\sigma_{dep} I_{dep}(\mathbf{x}) \sin^2 \theta), \quad (3.12)$$

which assumes both electric fields are orthogonally polarised and confined to the xy plane.

Now, we can calculate the angular resolution of this method. With a bit of algebra, it can be shown that the FWHM resolution equals

$$d_\theta = 2 \arccos \sqrt{\frac{\mathcal{W}(ke^k/2)}{k}}, \quad (3.13)$$

where $k = \sigma_{dep} I_{dep}$ and $\mathcal{W}(\cdot)$ denotes the Lambert W-function, the inverse of the function $f(x) = xe^x$. As expected, $d_\theta = 90^\circ$ for $I_d = 0$. For other values of I_d , the resolution is plotted in Figure 3.4.

In an experiment, one could account for depolarising effects such as rotational diffusion, energy transfer, et cetera. That is not necessary at this point, but could be easily modelled by convolving η with a depolarisation kernel δ . This kernel would be a probability distribution that quantifies how much a dipole can move and rotate between absorption and interaction with the depletion beam. It is also possible to model polarisation-sensitive detection with

$$h = h_{ex} \cdot (\eta * \delta_1) \cdot (h_{det} * \delta_2). \quad (3.14)$$

3.4. Samples

During the project, samples were very generously provided by research groups led by Pontus Nordenfelt and Vinay Swaminathan (Faculty of Medicine, Lund University).

Results of the cell sample shown in the experimental section contain a human cell line infected with bacteria of the *Yersinia* genus. Stainings present are: DAPI (tags DNA in the nucleus), GFP (tags the bacteria) and SiR-actin (tags the actin cytoskeleton). SiR-actin is an organic molecule (silicon-rhodamine) rigidly linked to an actin monomer. This dye is perfect for the setup, as SiR can be excited at 664 nm and has a sufficient absorption cross-section at 775 nm to perform STED. In addition, the fact that it is rigidly bound to the actin cytoskeleton means the light it emits is strongly polarised and reports on the actin filament orientation.

I also used two control samples for calibration measurements. The first is a sample of reflective gold colloids, which were used to image the laser point spread functions. The second contains larger Tetraspec fluorescent beads (Invitrogen).

4

Results

4.1. Conventional polarisation microscopy

SiR has been reported to be suitable for STED microscopy with excitation at 640 nm and depletion at 775 nm [49], so one would expect the SiR-actin stained sample to behave well in the setup too. The effect of the depletion laser (in conventional, i.e. donut mode) is presented in Figures 4.1 and 4.2. When zoomed in (Figure 4.2), it becomes visible that the fibres are not uniformly stained as individual fluorophores become apparent.

Even though using polarisation on the detection side is still impossible, the setup is able to obtain polarisation-resolved images by varying the angle of excitation polarisation. As a proof of concept, the sample described in Section 3.4 was imaged with linearly polarised excitation light from 0° to 170° in steps of 10° . Applying the algorithm detailed in Section 3.2 to three different ROIs resulted in Figure 4.4. As expected, vertically oriented fibres are excited by horizontal polarisation, because the transition dipole of SiR-actin is more or less orthogonal to the fibre [50]. A power law scaling of the saturation (degree of polarisation) and value (total brightness) in the algorithm can be used to adjust the brightness and contrast of the figure, as shown in Figure 4.5.

Polarisation images at STED resolution are unfortunately difficult to acquire due to strong photobleaching, which limits the number of points at which the excitation polarisation can be sampled (see Figure 4.3). There is a qualitative difference between the intensity profile of two ROIs that contain orthogonally oriented actin fibres. In particular, the maximum of one is located at 45° (after subtracting an exponential photobleaching response), while the other shows a maximum around 135° . The quality of this particular image is unfortunately not sufficient to colour it as in Figure 4.4.

In short, this section demonstrates successful conventional polarisation microscopy on the microscope setup and that SiR-actin is a suitable fluorophore to conduct research in Yersinia samples. There are two main limitations at the moment: the detection waveplates are not calibrated correctly, so using polarisation in the detection pathway is impossible, and the trade-off between

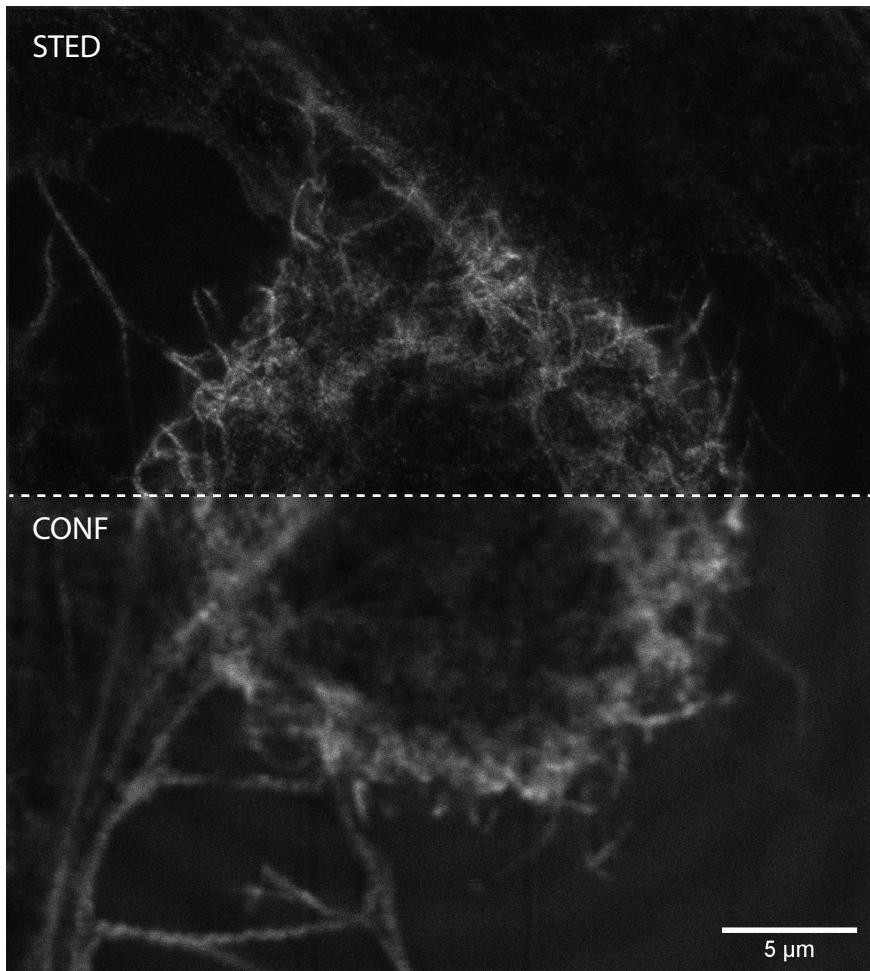


Figure 4.1: stained with SiR-actin, in confocal mode (bottom) and with the STED laser at 15% power (top). More FOVs are displayed in Figure 4.2.

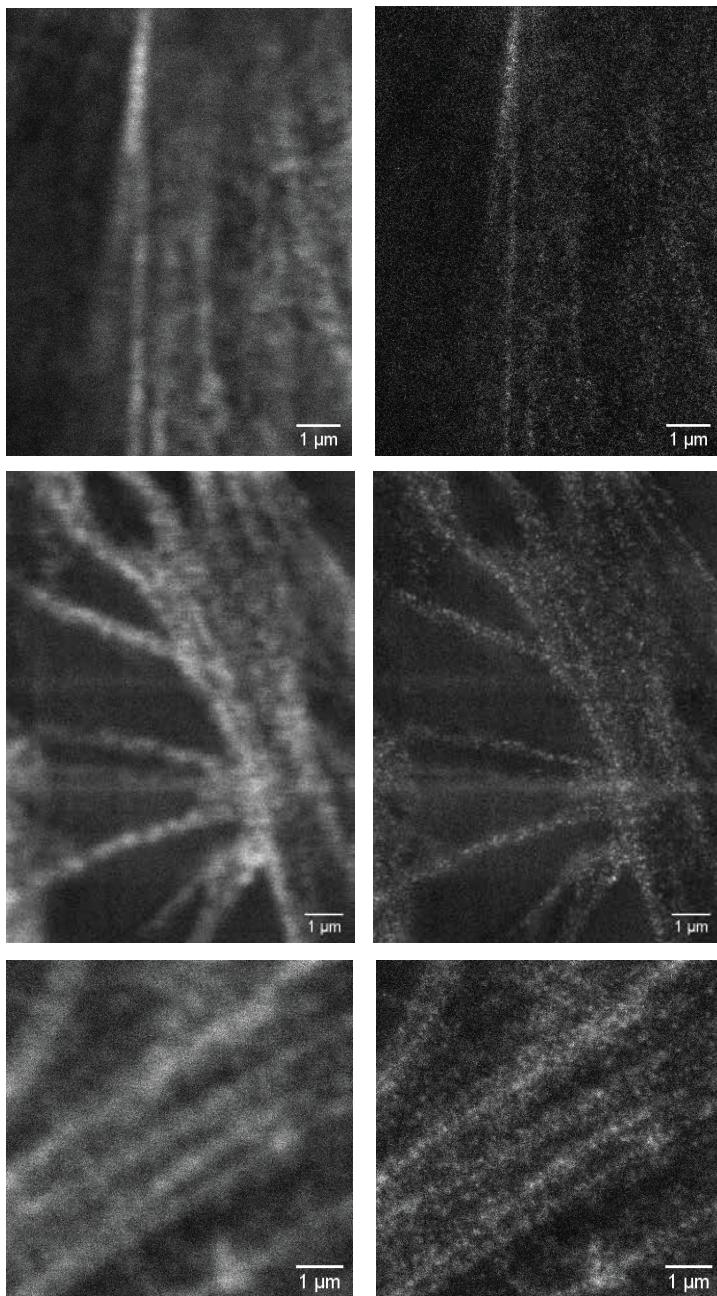


Figure 4.2: Confocal (left) and STED (right) acquisitions of various fields of view.

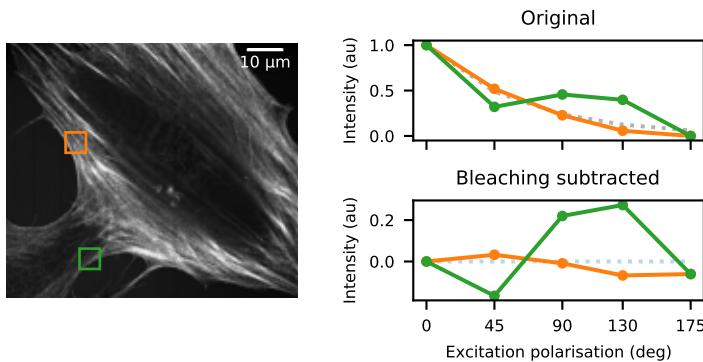


Figure 4.3: **Left:** First frame of a polarisation acquisition series, with two ROIs indicated that contain fibres of roughly perpendicular orientation. **Top right:** Integrated intensity of those ROIs at different sample points, normalised to the intensity present in the first frame. Dashed line: best-fit exponential curve. **Bottom right:** Same as top right, but with the exponential subtracted.

spatial resolution and polarisation information requires special care.

4.2. pSTED

Because the microscope was not set up to offer control over the polarisation of the depletion laser, a set of two new waveplates had to be installed to achieve polarisation-dependent depletion at the sample. A QWP was installed to negate the effects of the HWP and the QWP at the microscope end of the optical enclosure, and a HWP to control the polarisation angle. More details are presented in Section D.4.

After setting up the pSTED optics, these are the results so far: to verify that stimulated emission is polarisation-dependent, isotropic fluorescent beads were exposed two different conditions. In the first condition, the beads were excited with vertically polarised light and depleted by a laser polarised along different angles and at different power levels. Regardless of the angle, the fluorescence signal drops as the depletion power is increased, but the rate at which depends on the depletion polarisation. The depletion rate is highest when the lasers are aligned and lowest when they are orthogonal. This result can be explained by photoselection. When an isotropic sample is excited by linearly polarised light, fluorophores with a dipole moment orthogonal to the excitation beam will not be excited. As a result, the average excited fluorophore's transition dipole is oriented parallel to the excitation polarisation. Since depletion is most effective when the depletion polarisation is aligned with the fluorophore (as predicted by Equation 3.10) in this case that also means it is most effective when aligned with the excitation laser. This was indeed observed, see Figure 4.6 (Left).

On the other hand, if the sample is excited with circularly polarised light, the rate of depletion should not depend on the depletion polarisation. This measurement was performed twice. The first time by imaging an ROMEat

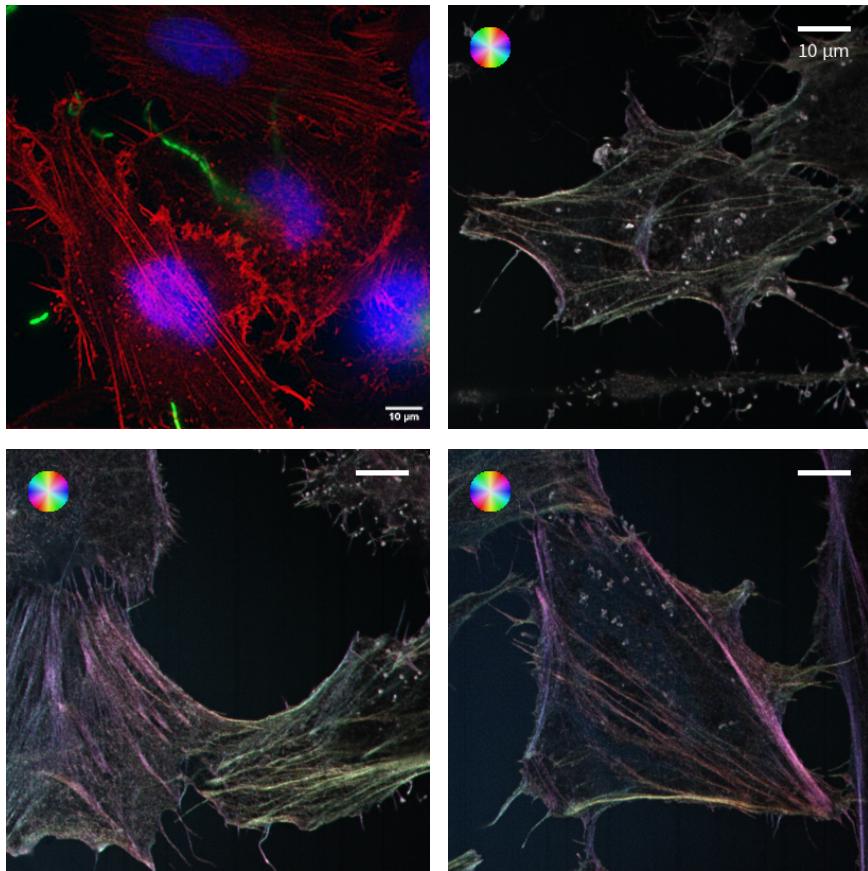


Figure 4.4: **Top left:** Composite view of a *Yersinia* sample, labelled with phalloidin instead of actin. Blue: DNA (DAPI). Green: bacteria (GFP). Red: actin (phalloidin). **Others:** Polarisation microscopy images of three different cells. The colour wheel indicates the direction of polarised light corresponding to a certain colour. Scale bars 10 μm .

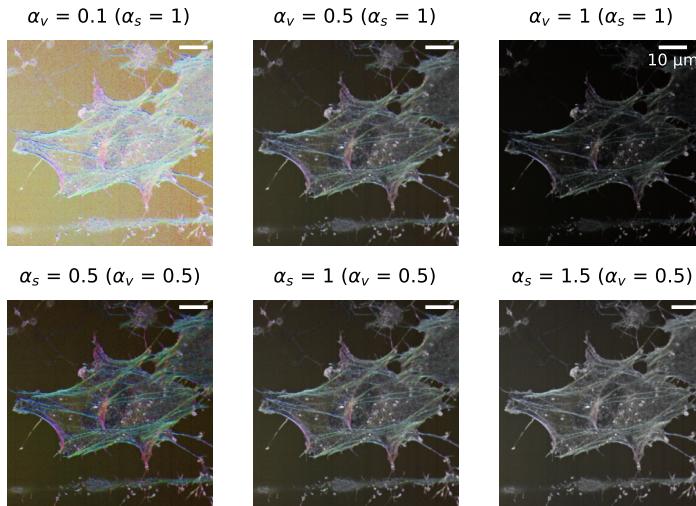


Figure 4.5: The effect of the exponents α_s (tuning saturation) and α_v (scaling brightness) on an image.

different depletion powers before changing the depletion polarisation and the second time by imaging ROIs at different depletion polarisations before changing the depletion power. The average of those measurements is shown in Figure 4.6 (Right). While the results are substantially different from the linear polarisation case, it has not been calculated whether the difference between them is statistically significant. This would be an important calculation to do. (Note that it was much easier to automate the acquisition of the linear polarisation data. As such, the figure shown is an average of five runs, while the other only shows data of a single run.)

Even though they could be improved, the results do show that depletion efficiency seems to be dependent on the polarisation of the depletion laser and the orientation of the fluorophore, and that pSTED is likely to work. Another experiment was performed on a cell sample, but since the actin fibres are not isotropic, it was unnecessary to use linearly polarised excitation to select a subset of fluorophores. Instead, several fibres were imaged with circularly polarised excitation light and manipulated the depletion angle to assess whether it was possible to perform pSTED on a biological sample, see Figure 4.7. Without a depletion laser, photobleaching is visible (blue). With a depletion laser, the photobleaching response disappears and is taken over by a signal that depends on the depletion polarisation (orange). Although more research is needed, this shows that pSTED could be a valuable new tool in the polarisation microscopy field.

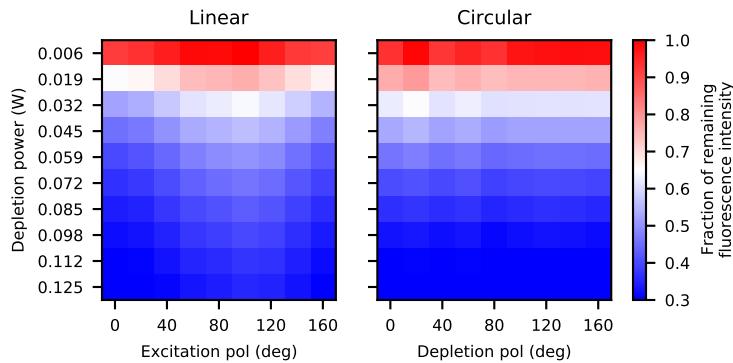


Figure 4.6: Dependency of surviving fluorescence on intensity and polarisation of depletion beam. **Left:** linearly polarised excitation at 0° (vertical). **Right:** circularly polarised excitation.

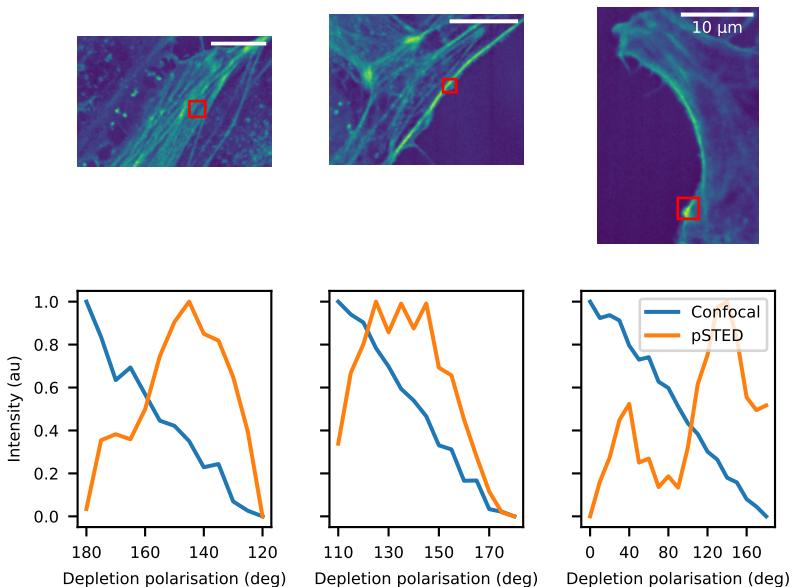


Figure 4.7: When samples are excited with circularly polarised light, the polarisation of the depletion beam can modulate the fluorescence intensity. **Top:** FOVs with a marked ROI in red. **Bottom:** Intensity profile, as a function of depletion polarisation. Orange: depletion beam turned on. Blue: depletion beam turned off.

5

Outlook

Firstly, polarisation microscopy on the Tegenfeldt STED microscope has been achieved, and with that the main goal of this project. It is now possible to perform polarisation microscopy at confocal resolution and overlay the structure found by non-polarised STED micrographs taken on the same setup. This is already a great result. It was also confirmed that SiR-actin is a probe that is suitable for the setup: it is excited at 640 nm, it is efficiently depleted by the 775 nm laser and its bond to actin is sufficiently rigid for polarisation microscopy.

There is some work left before results on biological specimens are ready to be published, however. Firstly, the calibrations of the microscope can be improved. The 640 nm laser is not as well polarised as the 561 nm one is, despite going through a similar set of optics. The circularity of the STED polarisation can also be improved. Improving the quality of the laser polarisations would make polarisation measurements easier and more accurate. Secondly, the waveplates of both excitation lasers rotate more than necessary when the laser polarisation is rotated through the software. Improving this point should increase the speed of acquisition and the polarisation quality. Thirdly, a quantitative understanding of the depolarising effects of different components is needed, instead of simply a qualitative one. This knowledge may then be used for correcting and interpreting the raw data. For instance, a sample could be imaged in this setup and in another lab and compare the results quantitatively. If they agree, the setup is ready to produce publication-quality polarisation microscopy results.

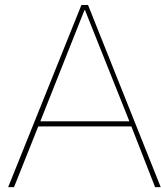
Secondly, there is some other ground to be gained on the conventional polarisation microscopy front: in the detection waveplates. Once these are properly aligned, it becomes possible to use a polarising beam splitter in the detection pathway, which gives twice the amount of orientational information in a single acquisition. This is great for experiments that are limited by the photon budget: very sensitive samples can then be imaged with polarisation at confocal resolution, or less sensitive ones could be imaged at STED resolution. Therefore, the calibration of the detection waveplates should be tuned and

measurement protocols that include them or a PBS can be developed. And if it is impossible to find exact alignment, then maybe it is possible to correct for that fact after acquisition.

Thirdly, the pSTED effort should be continued. The results so far are promising, but have not yet demonstrated the full potential of this method. This is due in part to finding the right balance between angular resolution and photodamage due to the increased depletion power. That problem is exacerbated by the fact that having a higher angular resolution requires acquiring more frames on top of the higher depletion power in every frame. Given the promising preliminary evidence, imaging a fresh sample could allow for polarisation images of higher quality and reveal the polarisation signature of actin micropatterns. On the more practical side of things, there are two obvious improvements that can be made: running the cabling to the HWP rotator stage through the back of the optical box instead of crushing under the lid of the optical enclosure and integrating the HWP into the microscope control software to automate pSTED acquisitions.

(discuss why dropping the donut can be worth it.)

Finally, the setup offers some possibilities that have not yet been discussed in this thesis. There are many methods in which fluorescence polarisation can be added. Using the fluorescence lifetime imaging (FLIM) module, it should be possible to quantify the rotational diffusion of a fluorophore by assessing how its polarisation signal changes on a timescale of nanoseconds. Likewise, it should be possible to quantify fluorescence resonant energy transfer (FRET) between fluorophores by measuring the depolarisation effect the sample has on incoming light. Another opportunity is to apply these results to other research programmes in the Tegenfeldt group: polarisation microscopy should be able to quantify the degree of polarisation of DNA flowing through a microfluidic device, such as a DLD (deterministic lateral displacement) chip.



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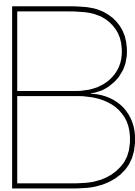
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I want to thank my friends and family for the continued support they gave me during this project, and simply for being part of my journey. Finally, I want to thank the Swedish public in general. I am extremely grateful for the opportunity to live in and discover your country for the past two years. It's been a blast!

Jonas, I promised you I'd share my star recipes, so check out the next page.

Misir Wat

Ingredients

- 4 tablespoons butter (or niter kibber, that's even better)
- 1 large yellow onion, very finely diced
- 3 cloves garlic, finely minced
- 1 tomato, very finely chopped
- 3 tablespoons tomato paste
- 2 tablespoons berbere , divided (you can get this at African Daily Market in Lund)
- 1 cup red lentils, rinsed
- 2.5 cups of vegetable stock
- 1 teaspoon salt

Preparation Melt 3 tablespoons of the butter in a medium stock pot. Add the onions and cook over medium-high heat for 8-10 minutes until golden brown. Add the garlic, tomatoes, tomato paste and 1 tablespoon of the berbere and cook for 5-7 minutes. Reduce the heat if needed to prevent burning. Add the broth and salt, bring it to a boil, reduce the heat to low and cover and simmer the lentils, stirring occasionally, for 40 minutes (adding more broth if needed) or until the lentils are soft. Stir in the remaining tablespoon of butter and berbere. Simmer for a couple more minutes. Add salt to taste. Serve with Ethiopian injera, sourdough bread, or rice.

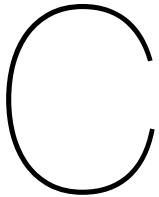
Source: daringgourmet.com

Ginger Beer

To start a ginger bug, combine 0.5 litres of water, 20 g of ginger (chopped or grated, but not peeled), and 28 g sugar in a glass pot. Cover it with a clean towel and keep it at room temperature. Until it becomes fizzy, add 20 g of ginger and 30 g of sugar every day.

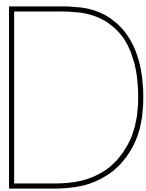
Then, combine 2 L of water, 200 g of sugar, and 75 g of ginger in a pot. Bring it to a boil, then reduce the heat and simmer for 5 to 8 minutes. Take it off the heat and let it cool down to room temperature. Add 100 g of strained ginger bug, and optionally some spices and the juice of three lemons. Divide over some airtight bottles and keep those at room temperature. Burp them daily by opening the cap to release built-up gas for about a week until the flavour is right and they are very carbonated. Then store them in the fridge for at least a couple of hours and enjoy!

Source: joshuaweissman.com



Code and data availability

I've spent quite a bit of effort ensuring this entire thesis easily reproducible from the raw data. To that end, the source code and data necessary to produce this file and the figures in it are published on GitHub, at <https://github.com/wduverger/msc-thesis>. If you experience any trouble getting the code to run, have any questions regarding this thesis or just want to chat about polarisation microscopy in general, please reach out! I'm always happy to help.



Characterisation and operation of the microscope

This appendix gives more details on the optics of the microscope, with a focus on the polarisation state of light at different points in the beam paths. These are important for operating the setup, but less so for understanding the project as a whole.

D.1. The laser modules

The fast-switching 561 nm laser emits horizontally polarised light, but the polarisation can be tuned by three waveplates in its path. The last one, a quarter-wave plate, is fixed in its rotation angle. In the standard mode of operation, excitation laser light should be circularly polarised to minimise resolution reduction due to lens distortion etc. [35]. In that mode, the (fast or slow) axis of the first quarter-wave plate is aligned with the laser, such that it does not affect polarisation and the half-wave plate is set such that it rotates the (linearly polarised) light to 45° with respect to the second quarter-wave plate. See Table D.1 for the polarisation characteristics in the calibration provided by Abberior. The 561 nm laser is quite well-calibrated. In circular polarisation, it reaches a χ very close to perfectly circular (45°).

The 640 nm laser does not have fast-switching built in, so instead, the light is fed into the microscope through a polarisation-preserving optical fibre by an acousto-optical modulator (AOM). The AOM is a crystal in the beam path, in which sound waves can be generated by a piezo element. The standing sound wave creates a diffraction grating with a variable pitch. That pitch is determined by the sound frequency, which can be modulated such that the first-order diffracted beam is either sent into the fibre aperture or not. The rest of the beam path is very similar to the 561 module, but the calibration of the waveplates in this pathway is not as accurate. Refer to Table D.1.

The depletion laser travels through an entirely different set of optics than the excitation lasers to generate a donut beam. First, it travels through a half-

Laser source	I_{max}/I_{min}	χ_I (deg)	χ_E (deg)	ψ (deg)
561 nm, linear (0°)	23.6	2.42	11.6	0
561 nm, circular	1.14	41.2	43.1	20
640 nm, linear (0°)	6.13	9.25	22.0	0
640 nm, circular	1.59	32.2	38.5	120
775 nm, circular	1.61	31.8	39.2	40
775 nm, linear	5.47	10.6	23.1	0

Table D.1: Polarisation characteristics of the lasers. Shown are linearity I_{max}/I_{min} , ellipticity χ_E (χ_I) of the electric field (intensity), and ellipse orientation ψ (anti-clockwise angle between ellipse orientation and vertical axis in sample plane). This data is based on Figure D.1, except the linear setting of the 775 nm laser. This required fitting new waveplates, as explained later (Section D.4).

wave plate that aligns the polarisation to the SLM (spatial light modulator). This HWP is necessary because an SLM adds an arbitrary spatially patterned phase delay to incident light, but only to the component polarised along its active axis. It will not alter the phase of the orthogonally polarised component. Using the proper phase delay patterns, one can create any (diffraction-limited) image in the sample plane. In this case, that would be a donut shape. Then – ignoring the pSTED module for now, since that was not included in the original setup – the beam travels through a half-wave plate to ensure circular polarisation in the sample plane (after going through a quarter-wave plate at 45° to the QWP axes), just like the excitation lasers. This is done to ensure that the depletion efficiency does not depend on sample orientation and to avoid polarisation-dependent PSF distortion by lenses and other optics. The quality of the STED polarisation is similar to that of the 640 nm laser. Note that this is actually a significant result, since it means that the donut beam is not isotropically polarised. It will be far more effective at quenching fluorophores oriented along 40° than those at 130° .

One more thing we can derive from this data (see Figure D.1), is that the 640 laser needs to ramp up every time it is powered on, due to the lack of fast switching. This can be somewhat remedied in software by turning the laser “on”, but at 0% power. The effect of this is that the laser is on but the AOM directs laser light away from the fiber aperture, thereby shortening the time to ramp up the laser power. Furthermore, at low powers, the laser intensity may not be proportional to the power setting in software. Their actual relationship is shown in Figure D.2. If experiments need to be done at low power, a neutral density filter is required. This is luckily not the case for biological specimens with a low density of fluorophores.

Looking at the calibrations of the waveplates in the excitation modules (Figure D.3), one can see that they move quite erratically, but they do work, as shown in Figure D.1. This could be an effect of the automated calibration performed by Abberior. In the 561 nm calibration, for example, one can see that the QWP constantly flips between about 30° and 120° , which are 90° apart. The calibration works, but the eccentricity of the polarisation ellipse depends

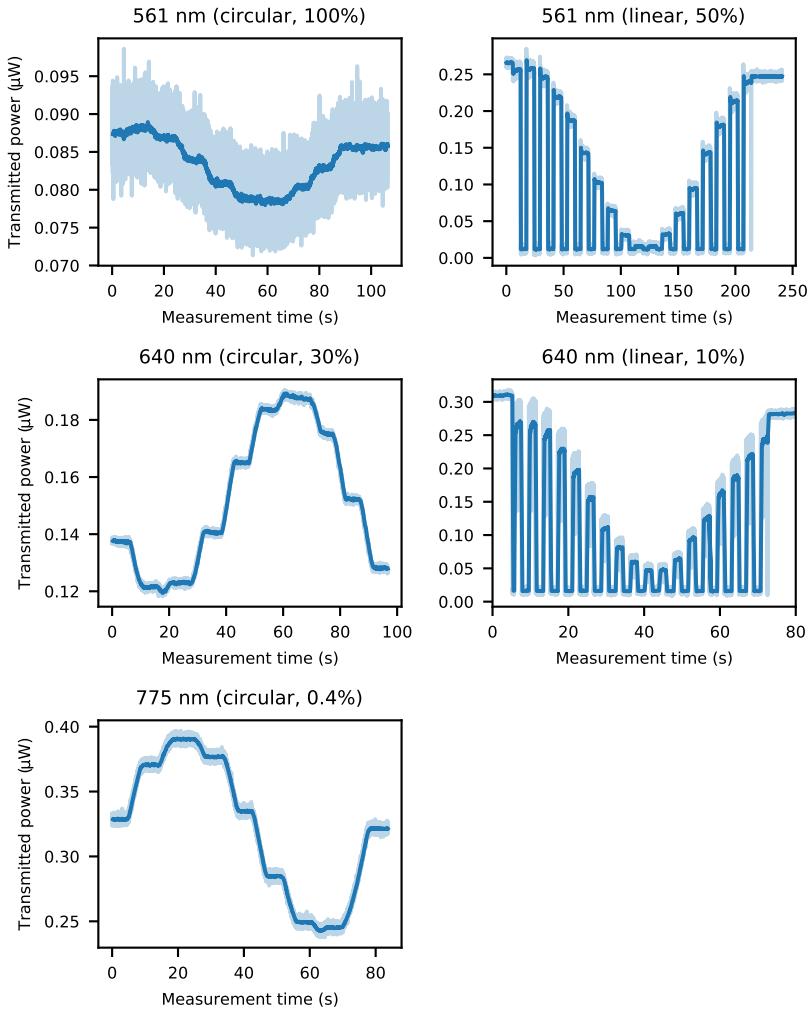


Figure D.1: Polarisation characterisation of the different lasers. To characterise circular polarisation, a polariser was placed in the sample and scanned from 0° to 180° in steps of 20° (note that a sample is lacking at 260° in the 775 nm data). Linear polarisation was characterised by fixing the polariser in place at 0° and scanning the laser itself from 0° to 180° in steps of 10° . Dark blue: average over 200 samples, light blue: raw data.

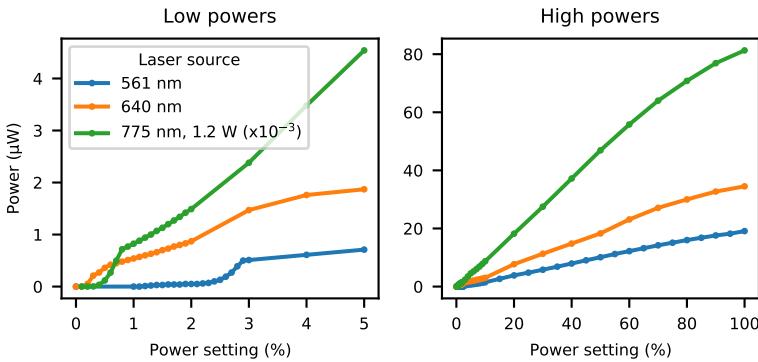


Figure D.2: The power of each of the three lasers as measured through a 10x objective. Notice the non-linearity at low power settings. The 775 nm data was scaled down by three orders of magnitude.

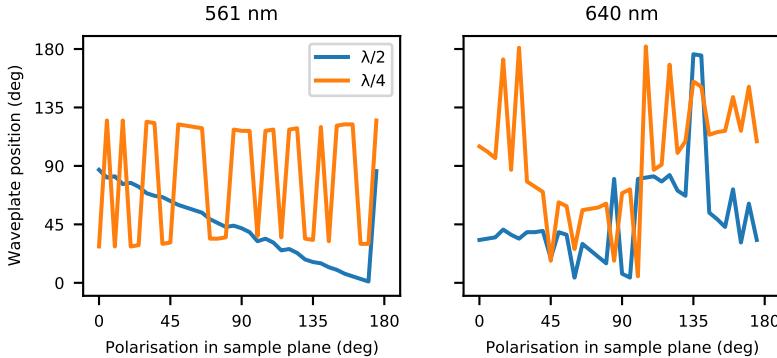


Figure D.3: Calibrations of the excitation waveplates, as supplied by the microscope manufacturer.

on the orientation. In other words, for a more rigorous characterisation of the microscope, Table D.1 should have an entry for every possible calibration setting.

One can measure the PSFs of the lasers by scanning over a reflective gold bead and acquiring an image on the photomultiplier tube (PMT). These are shown in Figure D.4.

D.2. The detection module

The main detectors of the microscope are a set of avalanche photodiodes (APDs), but there is also a highly sensitive PMT right after the QWP on the microscope end. The PMT is usually used to measure the point spread functions of the lasers and to align them with each other. In normal operation, the light travels on to the detection waveplates, then through a pinhole wheel, passes filters and dichroic mirrors in the filter cube housings, and is finally

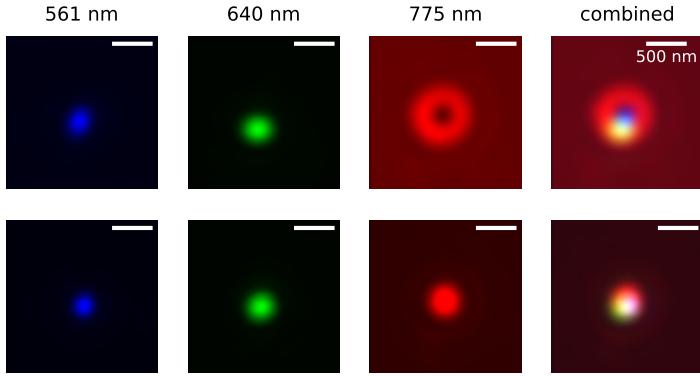


Figure D.4: Point spread functions of the different lasers (in donut and Gaussian modes) at different SLM configurations, by measuring the reflection from 100 nm wide gold beads.

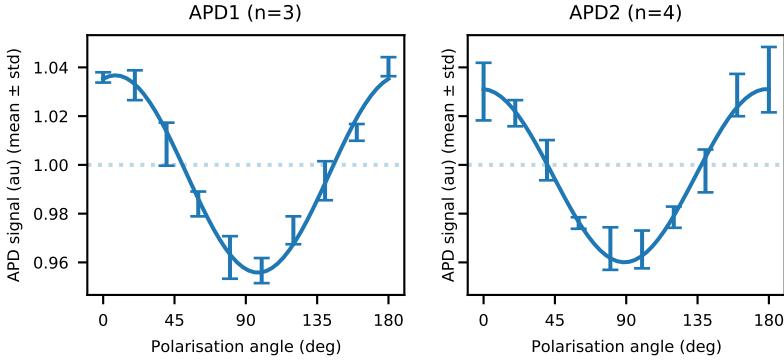


Figure D.5: Dependence of the signal from APD1 on the angle of polarisation of incoming light.

reflected onto the APDs. The wheel contains pinholes of different sizes, which allows for choosing the trade-off between light collection and z resolution, depending on the quality of the sample. Different filter cubes are available with various bandpass filters, dichroic mirrors, and/or a polarising beam splitter (PBS).

The APDs show a slight polarisation sensitivity of about 10% (see Figure D.5). I measured this by exciting Tetraspec beads with the 561 laser set to circular excitation, such that the emission light is non-polarised. Then I placed a linear polariser behind the detection waveplates and measured their signal as a function of the polariser angle. It seemed like the beam moved depending on the incident polarisation, as aligning the APDs when the signal was minimal did help a little, but the imperfect circularity of the 561 laser may also play a role, as it is on the same order of magnitude.

D.2.1. Calibrating the detection waveplates

We did have some problems with the waveplates in the detection module. They can be controlled through Abberior's software suite (Imspector), but it is not clear if they are set up correctly. The calibration is based on a control angle that I will denote θ . It would theoretically be possible for this setup to rotate polarised light of any orientation, since

$$S_{\lambda/2}(\theta/2)S_{\lambda/4}(0)S_{\lambda/4}(0) = \begin{pmatrix} \cos \theta & -\sin \theta \\ \sin \theta & \cos \theta \end{pmatrix}, \quad (\text{D.1})$$

which is simply the rotation matrix $R(\theta)$. The exact position of the quarter-wave plates does not matter, as long as they are aligned with each other. If the waveplates are at an angle ϕ instead of 0, then this set of waveplates rotates the polarisation by an angle $(\theta - 2\phi)$ instead.

It is unclear what goal the original calibration (see Figure D.6) should serve. If we approximate the default calibration with a QWP at θ and a HWP at 0, then the action of these waveplates would be

$$S_{\lambda/2}(0) \cdot S_{\lambda/4}(\theta) \cdot S_{\lambda/4}(0) = \begin{pmatrix} \cos^2 \theta + i \sin^2 \theta & (1+i) \cos \theta \sin \theta \\ (-1+i) \cos \theta \sin \theta & \cos^2 \theta - i \sin^2 \theta \end{pmatrix}. \quad (\text{D.2})$$

Therefore, I developed a new calibration. To do so, I first needed to establish at what angle the second quarter-wave plate would be aligned to the first one. I did this by placing a polariser in the sample holder (P1) and illuminating it with the top lamp, such that the light incident on the first quarter-wave plate was linearly polarised. Then I placed another polariser (P2) after that waveplate (i.e. before the last two waveplates) that I rotated to assess the linearity of the polarisation there. I found the angle of P1 at which light was most circular after the first QWP and subsequently placed P2 after all waveplates to find the angle of the second QWP that would make the polarisation linear again. At this point, the QWPs should be aligned. They could also be at 90° to one another. To distinguish between these setups, compare a measurement such as Figure D.8 with simulations in Figure D.7.

Second, I assessed both calibrations. As presented in Figure D.6, the new calibration works rather well, unlike the original one. However, its performance seems to depend on the precise angle of P1, see Figure D.8. This needs to be fixed before we can confidently use polarisation-affecting elements in the detection path. Luckily, simulations show that this effect can be explained by misalignment between the QWPs. Therefore, a more accurate calibration should fix this problem.

Finally, I checked the POL cube, which contains a polarising beam splitter (PBS) and some wavelength filters. It seems to work as expected (see Figure D.9). The transmitted ray has an extinction ratio of about 0.2, meaning that 20% of the power of rays that are orthogonally polarised to the transmission axis (and hence should be reflected by the PBS) is actually transmitted. On the other hand, it has an extinction ratio of .01 for the reflected wave.

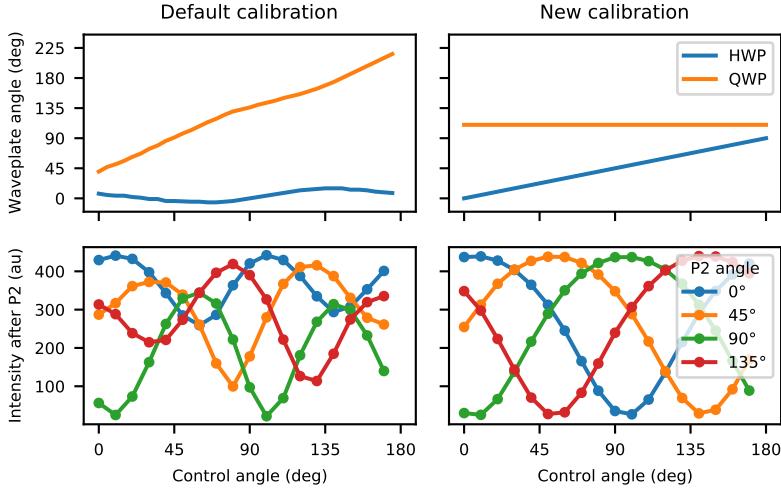


Figure D.6: A comparison of the default detection waveplate calibration and the new one. The new calibration seems to actually rotate incident polarisation by an arbitrary angle.

D.3. Protocol for polarisation microscopy

After acquisition, a stack of images I_{nxy} (with excitation light polarised along θ_n) must go through a number of steps to be converted into a false-colour polarisation image. Those steps are detailed in this section.

D.3.1. Stack alignment.

Images in a stack are aligned using an ECC (enhanced correlation coefficient) optimisation algorithm implemented in OpenCV, an open source computer vision library, which optimises an adjusted version of the correlation between two images, subject to certain constraints, e.g. translation, without shearing or rotation [51]. When the correlation between two images is maximised, they are aligned. The innovation of the ECC algorithm lies in the definition of an enhanced correlation coefficient which requires less computational power and is more robust than other algorithms [51].

If the goal is to generate a new image stack I'_{nxy} , corrected for sample or beam drift, we can take the first frame as reference, setting

$$I'_{0xy} = I_{0xy}. \quad (\text{D.3})$$

For every other frame I_{nxy} , we can calculate a warp matrix using OpenCV with the `findTransformECC()` method, which maximises the correlation between I_{nxy} and $I'_{(n-1)xy}$. Since we only observe drift in the images, we only allow translation (no shearing or rotation). Finally, we transform the original image using that warp matrix and `warpAffine()` and save the result as I'_{nxy} . In other words:

$$I'_{nxy} = \text{warpAffine}\left(I_{nxy}, \text{findTransformECC}\left(I'_{(n-1)xy}, I_{nxy}\right)\right), \quad (\text{D.4})$$

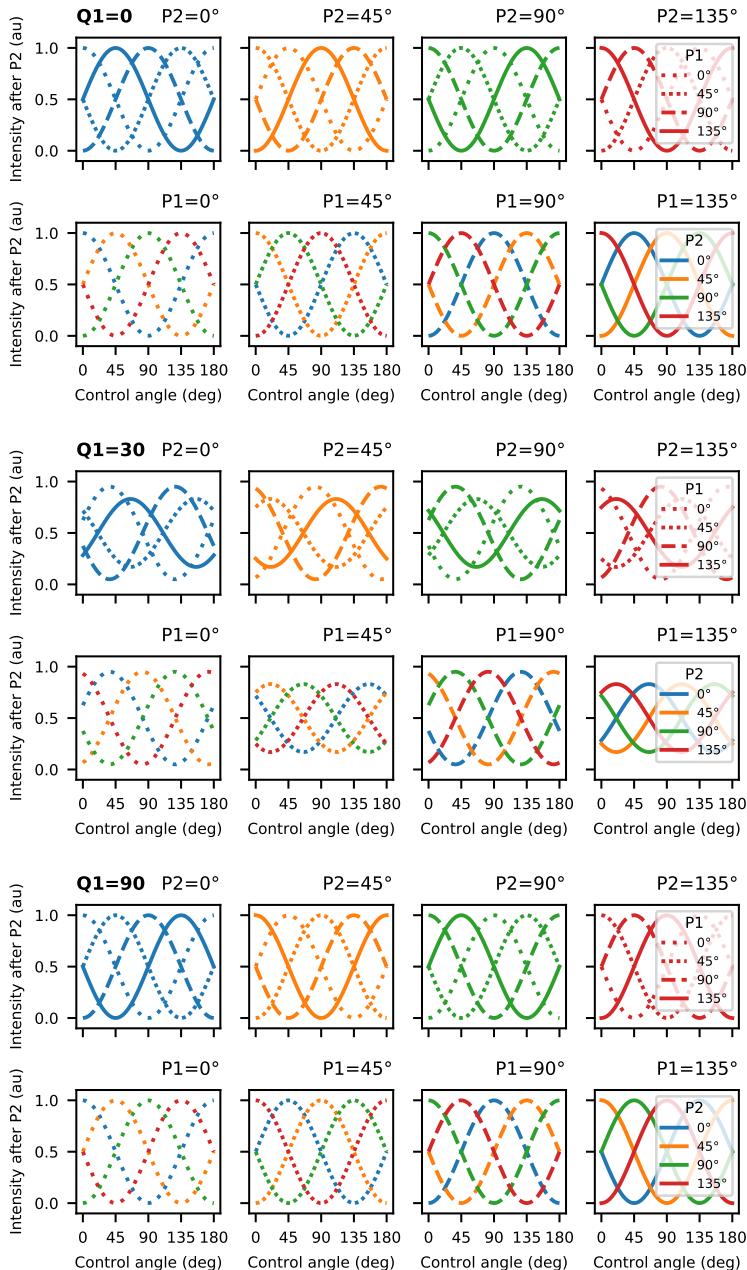


Figure D.7: Simulation of the effect of QWP alignment on the action of the detection pathway. Shown are three simulations in which the first QWP is rotated by 0, 30, and 90 degrees, respectively. The second QWP is always rotated by 0, and the HWP is rotated to half the rotation angle (x-axis). When the QWPs have an offset of 90 degrees (bottom), the upper and lower panels (with either P1 or P2 constant) are identical, but when they are aligned (top), there is a phase difference. When they are not aligned at all (middle), intensity variations are prominent. This simulation is based on Equation D.1.

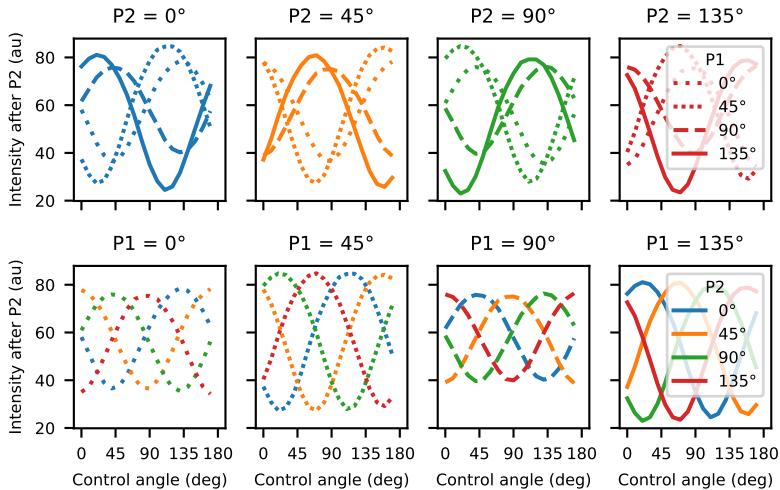


Figure D.8: The effect of rotating the incoming polarisation (P1). The detection waveplates seem to rotate the polarisation for P1 at 45° or 135°, but seem to circularise incoming light at 0° and 90° somewhat. (Note that the angle of P1 does not have a proper reference since I forgot to home the rotator.)

for all $n > 0$.

D.3.2. Bleaching compensation.

Since photobleaching will influence the calculation of the Fourier coefficients, it needs to be separated from the polarisation response. We can estimate the bleaching rate from the ratio between two frames at identical polarisations. Unfortunately, the Inspector software suite does not allow for rotating the excitation polarisation by more than 175°, so we cannot do this exactly. It would be possible if we were able to use the POL cube, but that is not an option, as explained before. Let \bar{I}_0 be the mean intensity of the first image,

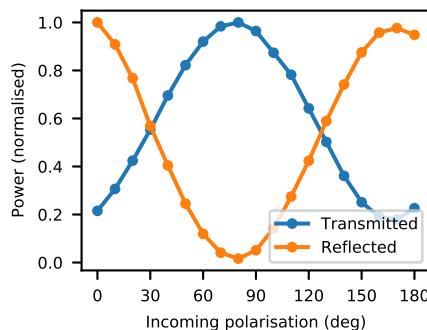


Figure D.9: Performance of the polarising beam splitter.

and \bar{I}_N the mean intensity of the last one. For now, we simply estimate the bleaching per frame as

$$r = \left(\frac{\bar{I}_N}{\bar{I}_0} \right)^{1/N}, \quad (\text{D.5})$$

given $N + 1$ number of frames were acquired. Then we can compensate for photobleaching by multiplying every frame with a correction factor as

$$I''_{nxy} = r^{-n} I'_{nxy}. \quad (\text{D.6})$$

D.3.3. Colouring the image.

For every pixel, calculate a complex-valued Fourier coefficient corresponding to a period of 180° (at which we should see polarisation dependence) using

$$F_{xy} = \sum_n I''_{nxy} e^{i2\theta_n}. \quad (\text{D.7})$$

Then construct an image in HSV colour space where

$$h_{xy} = \arg(F_{xy}), \quad (\text{D.8})$$

$$s_{xy} = |F_{xy}| / v_{xy}, \quad (\text{D.9})$$

$$v_{xy} = \sum_n I_{nxy}. \quad (\text{D.10})$$

Finally, normalise every channel c to be in the range $(0, 1)$ and optionally apply a power law with a manually chosen coefficient α_c for optimal visualisation,

$$c'_{xy} = \left(\frac{c_{xy}}{\max c_{xy}} \right)^{\alpha_c}. \quad (\text{D.11})$$

The effect of α_c is presented in Section 4.1. If necessary, the Fourier image F_{xy} can be blurred before constructing a false colour image. This can be useful when the image is too noisy for a decent Fourier calculation.

D.4. pSTED optics

The system was not set up to offer control over the depletion polarisation. Instead, the STED polarisation is always circular, and this is ensured by a set of two fixed waveplates: a QWP and a HWP (refer to Figure 3.1 and the description of the STED module in that section).

To control the polarisation of the depletion beam for pSTED, I first had to linearise the polarisation by compensating for the QWP in the beam path. That can be done by placing a QWP in the pSTED module in Figure 3.1, since

$$S_{\lambda/4}(\phi) S_{\lambda/2}(0) S_{\lambda/4}(-\phi) = R(2\phi). \quad (\text{D.12})$$

When the quarter-wave plates are aligned like that (when they are each others mirror image with respect to the HWP fast axis), this system simply rotates the polarisation by a fixed angle. This means that adding a HWP before this

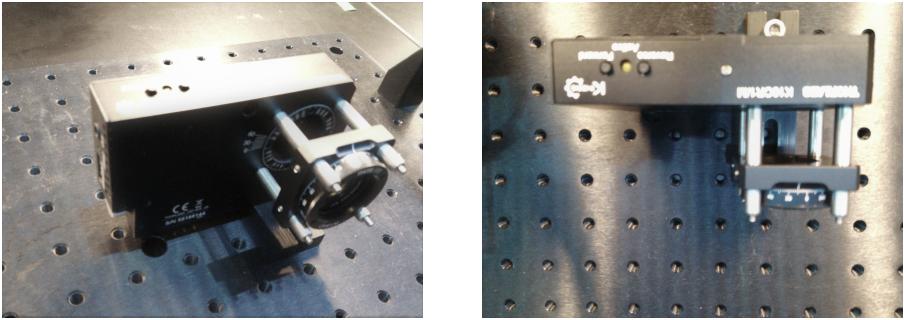


Figure D.10: The new pSTED optics. The rotational stage holds a HWP and a stationary QWP is mounted on a cage system. To take these optics out of the microscope, only the screw that holds the foot needs to be removed.

setup suffices to get full control over the polarisation angle of the depletion beam.

In practice, the first step was to mount these two new waveplates in the beam path. They were mounted right after the SLM on an easily removable foot, see Figure D.10. This way, conventional STED and polarisation-dependent STED are both easily performed. The HWP was mounted inside a rotational stage and the QWP in a cage system attached to the rotational stage, such that it is always aligned with the laser beam, but that its rotational angle is fixed. The first step was to find the angle of the QWP that maximised the linearity of the light polarisation at the sample. This was done by placing a polariser and a power meter at the sample and rotating them to characterise the polarisation of the depletion beam. I did this a couple of times, from which we concluded that the optimal angle for the QWP was 15° . Next, I had to find the angle of the HWP at which the depletion beam is vertically polarised. In that position, the depletion beam has a polarisation parallel to the excitation lasers (at 0°). This turned out to be 38.5° (or equivalently $38.5^\circ + 180^\circ = 218.5^\circ$). See Figure D.12. At a linearity of $I_{max}/I_{min} = 5.47$, the polarisation of the linearised pSTED beam is comparable in quality to the 640 laser (see Table D.1). Note that the HWP angle $\theta_{\lambda/2}$ must satisfy

$$\theta_{\lambda/2} = 218.5^\circ - \theta/2 \quad (\text{D.13})$$

to achieve a linear polarisation along θ in the sample plane.

Now that we have control over the beam polarisation, we can check how the new waveplates influence the PSF. In Figure D.11, the PSF is shown for different polarisation directions. There are several conclusions we can draw from this data. Firstly, the PSF is not radially symmetric any longer, even though the SLM was set to Gaussian mode. (This is easy to do. It only requires removing the donut-generating pattern on the SLM.) In particular, the ellipse orientation is parallel to the light polarisation, so we see the PSF rotating as we change the polarisation (see also Figure D.13). This can be explained by the fact that lenses interact differently with light polarised along different directions [52]. This is the reason one should only use linearly polarised

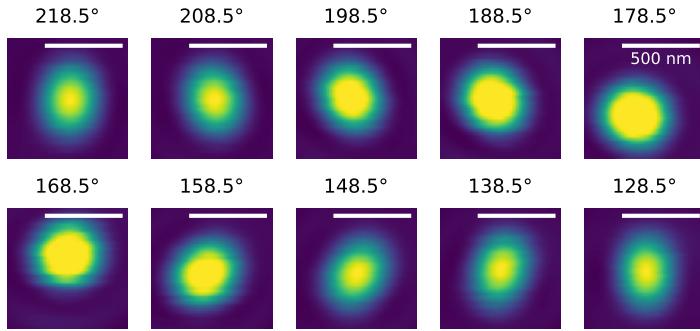


Figure D.11: Depletion PSF as a function of beam polarisation. The figure titles indicate the angle of the new HWP. The shown images are centred on the pixel with the highest intensity to correct for displacement.

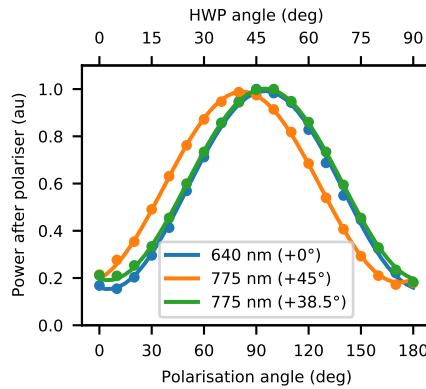


Figure D.12: The new rotating HWP in the pSTED module controls the depletion beam polarisation. With an offset of 38.5°, the depletion beam is parallel to the 640 laser (set to vertical linear polarisation).

excitation light when performing polarisation microscopy.

Other effects of the polarisation include the following: the intensity of the depletion beam varies as a function of the polarisation angle (see Figure D.13). This is probably due to linear dichroism present in the optical elements between the waveplates and the sample. This is not a surprising result, but should be accounted for during either image acquisition or analysis. Furthermore, the maximum of the PSF moves a little: up to roughly 100 nm from its mean position (Figure D.14). Fortunately, this is quite a bit below the diffraction limit for 775 nm light, i.e. around 400 nm, so the effect will be rather small.

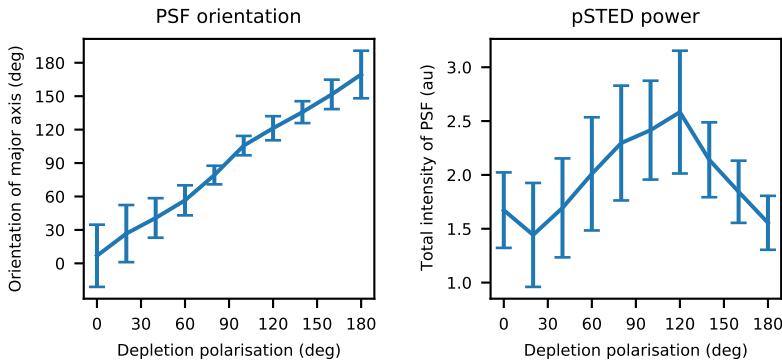


Figure D.13: Left: The pSTED PSF is elliptical, and its orientation matches the light polarisation. Right: pSTED intensity depends on polarisation. Error bars show standard deviation, $n=10$.

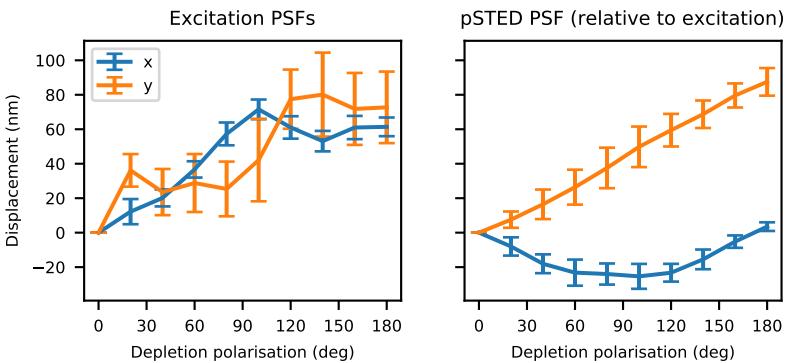


Figure D.14: The displacement of the laser PSFs as a function of depletion polarisation. Left: The displacement of the 561 nm and 640 nm PSFs (representing drift in the image). Right: Displacement of the pSTED PSF relative to the excitation PSFs.

D.5. A note on laser safety

The 775 nm line is a class 4 laser source. Under normal operation, the user is protected from it. However, when calibrating the STED beam or placing new components in the beam path, it is theoretically possible for the collimated laser beam to be reflected into the user's eyes. A high-powered laser beam can do permanent damage to the skin and retina, so we have to make sure we stay below the limits imposed by the Work Environment Agency's (Arbetsmiljöverkets) limits [53]. These regulations set forth three main conditions to calculate the Maximum Permissible Exposure (MPE) of a pulsed laser, see Table 2.6 of the regulations. Important values and formulas about our setup, as well as the limits provided by the Work Environment Agency are provided in Table D.2 and in the text below.

Quantity	Symbol	Value
Laser operating characteristics		
Beam radius	r	0.5 mm
Pulse width (FWHM)	τ	1.3 ns
Pulse repetition frequency	f	40 MHz
Pulse energy	E_{pulse}	31 nJ
Average power	P_{avg}	1.25 W
Safety parameters		
Thermal correction time	T_{min}	18 μ s
Correction factor a	C_a	1.41
Correction factor c	C_c	1
Correction factor e	C_e	1

Table D.2: Operating characteristics of the 775 laser line and relevant safety parameters.

Rule 1: The dose of a single pulse must not exceed the single-pulse MPE. The pulse dose H_{pulse} of the 775 nm laser at full power is

$$H_{pulse} = \frac{E_{pulse}}{2\pi r^2} \approx 39 \text{ mJ/m}^2, \quad (\text{D.14})$$

whereas the MPE equals

$$H_{pulse}^{MPE} = 5 \times 10^{-3} C_a C_e = 7.1 \text{ mJ/m}^2. \quad (\text{D.15})$$

This formula is found in table 2.2 of the regulations.

Rule 2: The dose of a single pulse may not exceed the thermally-corrected MPE. This weighs the pulse MPE with the amount of pulses in an interval T_{min} . The number of pulses in such an interval is $n = f \cdot T_{min}$, so

$$H_{thermal}^{MPE} = n^{-1/4} H_{pulse}^{MPE} = 1.3 \text{ mJ/m}^2. \quad (\text{D.16})$$

Rule 2 is therefore more strict than the rule 1. For safe operation, the laser must be ran at a power below 3.3% ($= H_{thermal}^{MPE}/H_{pulse}$).

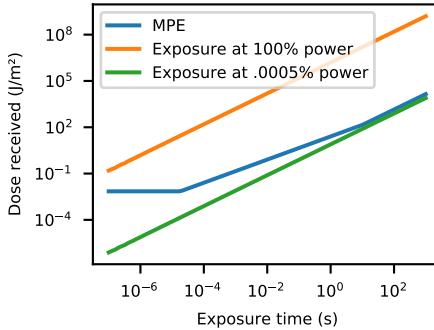


Figure D.15: Maximum permissible and actual exposure to the collimated STED beam as a function of exposure time.

Rule 3: The cumulative dose for a group of pulses in an interval of time t must not exceed the MPE for a single pulse of that time. Taking the necessary values from tables 2.2 and 2.3 of the regulations, the cumulative MPE is defined as

$$H_{tot}^{MPE}(t) = \begin{cases} 5 \times 10^{-3} C_a C_e & t < 18 \mu\text{s}, \\ 18t^{0.75} C_a C_e & 18 \mu\text{s} < t < 10 \text{ s}, \\ 10t C_a C_c & t > 10 \text{ s}. \end{cases} \quad (\text{D.17})$$

The actual dose, on the other hand, is

$$H_{tot}(t) = \lfloor ft \rfloor H_{pulse}, \quad (\text{D.18})$$

where $\lfloor \cdot \rfloor$ is the flooring function. This function is plotted in Figure D.15, from which it can be seen that the laser is only safe to use at 0.0009% capacity:

$$P_{safe} = \frac{10C_a C_c}{f H_{pulse}} \approx 0.0009\%. \quad (\text{D.19})$$

Since the minimum laser power offered by the software is .05%, OD2 goggles should be worn to guarantee safe operation of the 775 nm laser.