```
#set working directory
     setwd("~/04 Champion Lab/02 N-terminal Acetylation/OnePot/NAT5")
 3
 4
     #import libraries
 5
     library(ggplot2)
 6
     library(dplyr)
 7
    library(stringr)
8
    library(tidyr)
9
    library(ggrepel)
10 library (QFeatures)
11
    library(corrplot)
12
    library(limma)
13
     library(tibble)
14
15
     #set colors
16
     red <- hue pal()(3)[1]
17
     green <- hue_pal()(3)[2]
18
     blue <- hue pal()(3)[3]
19
20
     #import protein quant data and filter contaminants
21
     proteins <- read.csv("lfq.proteins.csv")</pre>
22
     proteins <- proteins[!grepl("CONTAM", proteins$Accession),]</pre>
23
24
     #parse accession and gene info, save as dictionary
25
     proteins$Accession2 <- str remove(str extract(proteins$Accession, "^[^\\|]*\\|"), "\\|")</pre>
26
     proteins$gene <- str remove(str remove(str extract(proteins$Accession,
     "^[^\\|]*\\|[^\\|]*\\|"),
27
                                                      "^[^\\|]*\\|"), "\\|")
28
     geneDict <- as.list(proteins$gene)</pre>
29
     names(geneDict) <- proteins$Accession2</pre>
30
31
     #clean column names
32
     names(proteins)[which(grep1("tr[0-9].Area", names(proteins)))] <- str remove(str extract(</pre>
     names(proteins)[which(grep1("tr[0-9].Area", names(proteins)))],
33
     " [^{\circ}] + br[0-9] tr[0-9]"), "^ ")
34
     #set missing values to NA
35
     proteins[proteins == 0] <- NA</pre>
36
37
     #import to Qfeatures and set metadata
38
     Qprot <- readQFeatures(table = proteins,</pre>
39
                              ecol = which(grepl("tr[0-9]", names(proteins))),
40
                              fnames = "Accession2",
41
                             name = "raw proteins")
42
     colnames(Qprot[[1]])
43
     Qprot$bioRep <- str extract(colnames(Qprot[[1]]), "br[0-9]+")</pre>
     Qprot$techRep <- str_extract(colnames(Qprot[[1]]), "tr[0-9]+$")</pre>
44
45
     Qprot$condition <- str extract(colnames(Qprot[[1]]), "^[^ ]*")</pre>
46
     Qprot$Inj <- colnames(Qprot[[1]])</pre>
47
     colData(Qprot)
48
49
50
     #log transform, base 2 so that later FC is in base 2
51
     Qprot <- addAssay(Qprot,</pre>
52
                        logTransform((Qprot[["raw proteins"]]),
53
                                      base = 2),
54
                        name = "log proteins")
55
     #normalize
56
     Qprot <- addAssay(Qprot,</pre>
57
                        normalize(Qprot[["log_proteins"]],
58
                                   method = "center.median"),
59
                        name = "norm proteins")
60
     #save figure of log and normaliztions
61
     png ("LogNormalization.png", units = "px", width = 1500, height = 1000)
62
     par(mfrow = c(1,3))
63
     limma::plotDensities(assay(Qprot[[1]]), legend = F)
64
     limma::plotDensities(assay(Qprot[[2]]), legend = F)
65
     limma::plotDensities(assay(Qprot[[3]]), legend = F)
66
     dev.off()
```

```
68
 69
      #add in old column data to normalized data
 70
      colData(Qprot[["norm proteins"]]) <- colData(Qprot)</pre>
 71
      #save as a summary dataframe
 73
      brSummaryWide <- Qprot[["norm proteins"]] %>% assay() %>% data.frame() %>%
      rownames to column ("Accession")
 74
      proteinsBR <- Qprot[["norm proteins"]]</pre>
 75
      #set variables as factor, and clarify the reference as WT
 76
      proteinsBR$condition <- factor(Qprot$condition)</pre>
 77
      proteinsBR$condition <- relevel(proteinsBR$condition, ref = "WT")</pre>
 78
 79
 80
      #create model based on values in condition
 81
      model design <- model.matrix(~proteinsBR$condition)
 82
      \#fit proteins to linear model with lmFit and the matrix we just created
 83
      fitted lm <- proteinsBR \%>%
 84
 85
        assay() %>%
 86
        lmFit(design = model design) %>%
 87
        eBayes()
 88
      #pull WT vs deletion
 89
 90
      DE DEL <- topTable(fit = fitted lm,
                          adjust.method = "BH",
 91
 92
                          number = Inf,
 93
                          coef = "proteinsBR$conditiond1839",
 94
                          sort.by = "p",
 9.5
                          confint = T)
 96
 97
      DE DEL$Accession <- rownames (DE DEL)
 98
 99
100
101
      #calculate corrected z score based on B-H adujsted p-value. Altman,
      https://doi.org/10.1136/bmj.d2090
102
      DE DEL$corrected z \leftarrow sqrt(0.743 - (2.404*log(DE DEL$adj.P.Val))) - 0.862
103
      #calculate the one directional 95% confidence interval
104
      DE DEL$error <- abs(DE DEL$logFC /DE DEL$corrected z) * 1.96
105
106
107
      #pull only values with non-missing expression data
108
      DE DEL <- DE DEL[!is.na(DE DEL$AveExpr),]
109
      DE DEL$CountWT <- NA
110
      DE DEL$CountExp <- NA
111
112
      #count number of valid measurments for each strain
113
      for (i in 1:nrow(DE DEL)) {
114
        CountDF <- brSummaryWide[brSummaryWide$Accession == DE DEL$Accession[i],]</pre>
115
        DE DEL$CountWT[i] <- sum(!is.na(CountDF[,grepl("WT", names(CountDF))][1,]))</pre>
116
        DE DEL$CountExp[i] <- sum(!is.na(CountDF[,grepl("d1839", names(CountDF))][1,]))
117
118
119
      #set appropriate negative and positive infinite changes for totally missing values
120
      DE DEL$logFC[DE DEL$CountWT == 0] <- Inf</pre>
121
      DE DEL$logFC[DE DEL$CountExp == 0] <- -Inf
122
123
      #set filters
124
      adjSigCutoff <- 0.05
125
      NonAdjSigCutoff <- 0.05
126
      LFCcutoff <- 1
127
128
129
      #option to plot either adj or non-adj p values
130
     adj <- T
131
132
      if (adj) {
133
        proteins sig <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff]))</pre>
```

67

```
134
        proteins sig2 <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
135
                                                              abs(DE DEL$logFC) > LFCcutoff]))
136
        proteins label pos <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
137
                                                                   DE DEL$logFC > LFCcutoff]))
138
        proteins label neg <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
139
                                                                   DE DEL$logFC < -LFCcutoff]))</pre>
140
      } else {
141
       proteins sig <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff]))
142
        proteins sig2 <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
143
                                                              abs(DE DEL$logFC) > LFCcutoff]))
144
        proteins label pos <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
                                                                   DE DEL$logFC > LFCcutoff]))
145
        proteins label neg <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
146
147
                                                                   DE DEL$logFC < -LFCcutoff]))</pre>
148
149
150
151
152
153
154
      #add in gene and protein name info from saved dictionaries
155
      DE DEL$gene <- sapply(DE DEL$Accession, function(Accession) geneDict[[Accession]])
156
157
      #set colors
158
      colorsUPDOWN <- c(blue, red)</pre>
159
      names(colorsUPDOWN) <- c(T, F)</pre>
160
161
      #add extra area above and below plot for error bars
162
      plotBufferFactor <- 1.5</pre>
163
      maxY <- max(DE DEL$logFC[is.finite(DE DEL$logFC)]) * plotBufferFactor</pre>
164
      minY <- min(DE DEL$logFC[is.finite(DE DEL$logFC)]) * plotBufferFactor</pre>
165
166
      #plot MA
167
      ggplot() +
168
        theme bw (base size = 25) +
169
        theme (panel.grid = element blank(),
              legend.position = "none") +
170
171
        labs(y = "Log2 Fold Change", x = "Normalized Average Expression (Log)",
172
             title = expression("WT vs " * Delta * italic(emp1))) +
173
        geom point(data = DE DEL[!DE DEL$Accession %in% proteins sig &
174
                                     is.finite(DE DEL$logFC),],
175
                   aes(x = AveExpr, y = logFC), color = "grey50",
176
                   alpha = 0.8, size = 0.8) +
177
        geom hline(yintercept = 0, linetype = 1) + #add line at LFC = 0
178
        geom hline(yintercept = c(-LFCcutoff, LFCcutoff), linetype = 2) +
179
        geom point(data = DE DEL[DE DEL$Accession %in% proteins sig,],
180
                   aes(x = AveExpr, y = logFC, color = logFC > 0), size = 0.8) +
181
        scale color manual(values = colorsUPDOWN) +
182
        scale fill manual(values = colorsUPDOWN) +
183
        geom segment (data = DE DEL[DE DEL$Accession %in% proteins sig2,],
184
                     aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = 1) +
                      #add 95% CI lines
185
        geom segment (data = DE DEL[DE DEL$Accession %in% proteins sig2,],
186
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error,
187
                          color = logFC > 0), lwd = 0.5) +
188
        geom point (data = DE DEL[DE DEL$Accession %in% proteins sig2 &
189
                                     is.finite(DE DEL$logFC),],
190
                   aes(x = AveExpr, y = logFC, fill = logFC > 0), size = 3,
191
                   shape = 21, stroke = 1.5) +
192
        geom point(data = DE DEL[DE DEL$logFC > 0 & !is.finite(DE DEL$logFC),],
193
                    shape = 23, aes(x = AveExpr, size = CountExp, fill = logFC > 0), y = maxY,
                   stroke = 1, alpha = 0.6) +
194
        geom point(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC),],</pre>
195
                   shape = 23, aes(x = AveExpr, size = CountWT, fill = logFC > 0), y = minY,
                   stroke = 1, alpha = 0.6) +
196
        coord cartesian(ylim = c(minY, maxY)) +
        scale size (range = c(1,3)) +
197
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteins label pos,],
198
199
                          aes(x = AveExpr, y = logFC, label = gene), nudge y = \frac{1}{1}, nudge x = \frac{-1.2}{1}
```

```
200
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteins label neg,],
201
                          aes(x = AveExpr, y = logFC, label = gene), nudge y = -1, nudge x = -1
                          1.2) +
202
        geom label repel(data = DE DEL[DE DEL$logFC > 0 & !is.finite(DE DEL$logFC) & DE DEL$
        CountExp > 1, ],
203
                          aes(x = AveExpr, label = gene), y = maxY, nudge y = -1, nudge x = -1)
2.04
        geom label repel(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC) & DE DEL$
        CountWT > 1, ],
205
                          aes(x = AveExpr, label = gene), y = minY, nudge y = \frac{1}{1}, nudge x = \frac{1}{1})
206
207
208
209
      ggsave ("WTvsDEL MAplot.png", width = 15, height = 12)
210
211
      write.table(DE DEL, "WTvsDELprotein.tsv", sep = "\t", row.names = F)
212
213
214
215
      #add in Acetylation changers
216
      acetylation <- read.csv("1839 acetylation reported.tsv", sep = "\t")</pre>
217
218
      proteinList <- acetylation$Accession2</pre>
219
220
221
      ggplot() +
222
        theme bw (base size = 25) +
223
        theme (panel.grid = element blank(),
224
               legend.position = "none") +
225
        labs(y = "Log2 Fold Change", x = "Normalized Average Expression (Log)",
             title = expression("WT vs " * Delta * italic(emp1))) +
226
227
        geom point (data = DE DEL,
228
                    aes(x = AveExpr, y = logFC), color = "grey50",
229
                    alpha = 0.8, size = 0.8) +
230
        geom hline(yintercept = 0, linetype = 1) + #add line at LFC = 0
231
        geom hline(yintercept = c(-LFCcutoff, LFCcutoff), linetype = 2) +
232
        geom segment(data = DE DEL[DE DEL$Accession %in% proteinList,],
233
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = \frac{1}{2}) +
                      #add 95% CI lines
234
        geom segment(data = DE DEL[DE DEL$Accession %in% proteinList,],
235
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = 0.5,
236
                      color = green) +
237
        geom point(data = DE DEL[DE DEL$Accession %in% proteinList &
238
                                     is.finite(DE DEL$logFC),],
239
                    aes(x = AveExpr, y = logFC), size = 3,
240
                    fill = green,
241
                    shape = 21, stroke = 1.5) +
242
        geom point(data = DE DEL[DE DEL$logFC > 0 & !is.finite(DE DEL$logFC),],
243
                    shape = 23, aes(x = AveExpr), y = maxY, stroke = 1, alpha = 0.6) +
244
        geom point(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC),],</pre>
245
                    shape = 23, aes(x = AveExpr), y = minY, stroke = 1, alpha = 0.6) +
246
        coord cartesian(ylim = c(minY, maxY)) +
247
        scale size (range = c(1,3)) +
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteinList,],
248
249
                          aes(x = AveExpr, y = logFC, label = gene), nudge y = \frac{3}{2}, nudge x = \frac{1.2}{2}
250
                          max.overlaps = 40)
251
252
253
      ggsave("WTvsDEL MAplot acetylationChangers.png", width = 15, height = 12)
254
255
256
      #log transform, base 10 for galaxy plots
257
      Qprot <- addAssay(Qprot,</pre>
                         logTransform((Qprot[["raw proteins"]]),
258
259
                                       base = 10),
260
                         name = "log proteins 10")
261
      Qprot <- addAssay(Qprot,</pre>
```

```
262
                         normalize(Qprot[["log proteins 10"]],
263
                                   method = "center.median"),
264
                         name = "norm_proteins_10")
265
266
267
268
      #add in old column data to normalized data
269
      colData(Qprot[["norm proteins 10"]]) <- colData(Qprot)</pre>
270
271
      #pull log 10 data
272
      loglog <- Qprot[["norm proteins 10"]] %>% assay() %>% data.frame() %>% rownames to column
      ("Accession")
273
274
      #pivot longer
275
      loglogLong <- loglog %>% pivot longer(2:ncol(loglog))
276
      loglogLong$strain <- str extract(loglogLong$name, "^[^]+")</pre>
277
278
      #average area and count of non na values for each
279
      loglogGrouped <- loglogLong %>% group by(Accession, strain) %>%
280
        summarise(Area = mean(value, na.rm = T),
281
                   count = sum(!is.na(value)))
282
283
      #pivot wider for plotting
284
      loglogWide <- loglogGrouped %>% pivot wider(id cols = Accession,
285
                                                     values from = Area,
286
                                                    names from = strain)
287
      #plot
288
      ggplot(loglogWide) +
289
        geom point (aes (x = WT, y = d1839), color = "grey50",
290
                    size = 0.8, alpha = 0.8) +
291
        theme bw (base size = 25) +
292
        theme (panel.grid = element blank(),
               legend.position = "none") +
293
        geom abline(slope = \frac{1}{2}, intercept = \frac{1}{2}, linetype = \frac{1}{2}, size = \frac{1}{2}) +
294
295
        geom point(data = loglogWide[loglogWide$Accession %in% proteins sig2,],
296
                    aes(x = WT, y = d1839, fill = WT < d1839), size = 3,
297
                    shape = 21, stroke = 1.5) +
298
        scale fill manual(values = colorsUPDOWN) +
299
        labs(title = expression("WT vs " * Delta * italic(emp1)),
300
              x = "Normalized WT expression (log)",
301
             y = expression("Normalized " * Delta * italic(emp1) * " expression (log)"))
302
303
      ggsave("galaxyWTvsDelProteinDE.png", width = 10, height = 10)
304
305
306
      ggplot(loglogWide) +
307
        geom_point(aes(x = WT, y = d1839), color = "grey50",
308
                    size = 0.8, alpha = 0.8) +
309
        theme bw (base size = 25) +
310
        theme (panel.grid = element blank(),
311
               legend.position = "none") +
312
        geom abline(slope = 1, intercept = 0, linetype = 2, size = 1) +
313
        geom point (data = loglogWide[loglogWide$Accession %in% proteinList,],
314
                    aes(x = WT, y = d1839), fill = green, size = 3,
315
                    shape = 21, stroke = 1.5) +
316
        scale fill manual(values = colorsUPDOWN) +
317
        labs(title = expression("WT vs " * Delta * italic(emp1)),
318
              x = "Normalized WT expression (log)",
              y = expression("Normalized " * Delta * italic(emp1) * " expression (log)")) +
319
320
        geom label repel(data = loglogWide[loglogWide$Accession %in% proteinList,],
321
                          aes(x = WT, y = d1839, label = Accession), nudge y = \frac{1}{1}, nudge x = \frac{-1}{1},
322
                          max.overlaps = 40)
323
324
325
      ggsave("galaxyWTvsDelAcetylationChangers labs.png", width = 10, height = 10)
326
327
      ggplot(loglogWide) +
328
        geom point (aes (x = WT, y = d1839), color = "grey50",
329
                    size = 0.8, alpha = 0.8) +
```

```
330
        theme bw (base size = 25) +
331
        theme (panel.grid = element blank(),
332
              legend.position = "none") +
333
        geom abline(slope = 1, intercept = 0, linetype = 2, size = 1) +
334
        geom point (data = loglogWide[loglogWide$Accession %in% proteinList,],
335
                   aes(x = WT, y = d1839), fill = green, size = 3,
336
                   shape = 21, stroke = 1.5) +
337
        geom point(data = loglogWide[loglogWide$Accession %in% proteins sig2,],
                   aes(x = WT, y = d1839, fill = WT < d1839), size = 3,
338
339
                   shape = 21, stroke = 1.5) +
340
        scale fill manual(values = colorsUPDOWN) +
341
        labs(title = expression("WT vs " * Delta * italic(emp1)),
342
             x = "Normalized WT expression (log)",
             y = expression("Normalized " * Delta * italic(emp1) * " expression (log)"))
343
344
345
      ggsave ("galaxyWTvsDelAcetylationChangers andDE.png", width = 10, height = 10)
346
347
348
      #SAME as above but for complement instea of Del
349
      #fit proteins to linear model with lmFit and the matrix we just created
350
      fitted lm <- proteinsBR %>%
351
        assay() %>%
        lmFit(design = model design) %>%
352
353
        eBayes ()
354
355
      DE DEL <- topTable(fit = fitted lm,
356
                          adjust.method = "BH",
357
                          number = Inf,
358
                          coef = "proteinsBR$conditionc1839",
359
                          sort.by = "p",
360
                          confint = T)
361
362
363
      DE DEL$Accession <- rownames (DE DEL)
364
365
366
367
      #calculate corrected z score based on B-H adujsted p-value. Altman,
      https://doi.org/10.1136/bmj.d2090
368
      DE DEL$corrected z \leftarrow sqrt(0.743 - (2.404*log(DE DEL$adj.P.Val))) - 0.862
369
      #calculate the one directional 95% confidence interval
370
      DE DEL$error <- abs(DE DEL$logFC /DE DEL$corrected z) * 1.96
371
372
373
374
      DE DEL <- DE DEL[!is.na(DE DEL$AveExpr),]
375
      DE DEL$CountWT <- NA
376
      DE DEL$CountExp <- NA
377
378
      for (i in 1:nrow(DE DEL)) {
379
        CountDF <- brSummaryWide[brSummaryWide$Accession == DE DEL$Accession[i],]</pre>
380
        DE DEL$CountWT[i] <- sum(!is.na(CountDF[,grepl("WT", names(CountDF))][1,]))</pre>
381
        DE DEL$CountExp[i] <- sum(!is.na(CountDF[,grepl("c1839", names(CountDF))][1,]))</pre>
382
383
384
      DE DEL$logFC[DE DEL$CountWT == 0] <- Inf
385
      DE DEL$logFC[DE DEL$CountExp == 0] <- -Inf
386
387
388
      adjSigCutoff <- 0.05
389
      NonAdjSigCutoff <- 0.05
390
      LFCcutoff <- 1
391
392
393
394
     adj <- T
395
396
      if (adj) {
397
        proteins sig <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff]))</pre>
```

```
398
        proteins sig2 <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
399
                                                             abs(DE DEL$logFC) > LFCcutoff]))
400
        proteins label pos <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
401
                                                                   DE DEL$logFC > LFCcutoff]))
402
        proteins label neg <- na.omit(unique(DE DEL$Accession[DE DEL$adj.P.Val < adjSigCutoff &
403
                                                                  DE DEL$logFC < -LFCcutoff]))</pre>
404
      } else {
405
       proteins sig <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff]))</pre>
        proteins sig2 <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
406
407
                                                             abs(DE DEL$logFC) > LFCcutoff]))
408
        proteins label pos <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
409
                                                                   DE DEL$logFC > LFCcutoff]))
410
        proteins label neg <- na.omit(unique(DE DEL$Accession[DE DEL$P.Value < adjSigCutoff &
411
                                                                   DE DEL$logFC < -LFCcutoff]))</pre>
412
413
414
415
416
417
418
      #add in gene and protein name info from saved dictionaries
419
      DE DEL$gene <- sapply(DE DEL$Accession, function(Accession) geneDict[[Accession]])
420
421
422
423
      plotBufferFactor <- 1.5
424
      maxY <- max(DE DEL$logFC[is.finite(DE DEL$logFC)]) * plotBufferFactor</pre>
425
      minY <- min(DE DEL$logFC[is.finite(DE DEL$logFC)]) * plotBufferFactor</pre>
426
427
428
      ggplot() +
429
        theme bw (base size = 25) +
430
        theme (panel.grid = element blank(),
431
              legend.position = "none") +
432
        labs(y = "Log2 Fold Change", x = "Normalized Average Expression (Log)",
             title = expression("WT vs " * Delta * italic(emp1) * "/comp")) +
433
434
        geom point (data = DE DEL[!DE DEL$Accession %in% proteins sig &
435
                                    is.finite(DE DEL$logFC),],
436
                    aes(x = AveExpr, y = logFC), color = "grey50",
437
                    alpha = 0.8, size = 0.8) +
438
        geom hline(yintercept = 0, linetype = 1) + #add line at LFC = 0
439
        geom hline(yintercept = c(-LFCcutoff, LFCcutoff), linetype = 2) +
440
        geom point(data = DE DEL[DE DEL$Accession %in% proteins sig,],
                    aes(x = AveExpr, y = logFC, color = logFC > \overline{0}), size = 0.8) +
441
442
        scale_color_manual(values = colorsUPDOWN) +
443
        scale fill manual(values = colorsUPDOWN) +
444
        geom segment (data = DE DEL[DE DEL$Accession %in% proteins sig2,],
445
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = 1) +
                      #add 95% CI lines
446
        geom segment (data = DE DEL[DE DEL$Accession %in% proteins sig2,],
447
                      aes (x = AveExpr, y = logFC - error, yend = logFC + error,
448
                          color = logFC > 0), lwd = 0.5) +
449
        geom point (data = DE DEL[DE DEL$Accession %in% proteins sig2 &
450
                                    is.finite(DE DEL$logFC),],
451
                    aes(x = AveExpr, y = logFC, fill = logFC > 0), size = 3,
452
                    shape = 21, stroke = 1.5) +
        geom point(data = DE_DEL[DE_DEL$logFC > 0 & !is.finite(DE_DEL$logFC),],
453
454
                    shape = 2\overline{3}, aes(x = AveExpr, size = CountExp, fill = logFC > 0), y = maxY,
                    stroke = 1, alpha = 0.6) +
455
        geom point(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC),],</pre>
456
                    shape = 23, aes(x = AveExpr, size = CountWT, fill = logFC > 0), y = minY,
                    stroke = 1, alpha = 0.6) +
457
        coord cartesian(ylim = c(minY, maxY)) +
458
        scale size (range = c(1,3)) +
459
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteins label pos,],
460
                          aes(x = AveExpr, y = logFC, label = gene), nudge y = \frac{1}{1}, nudge x = \frac{-1.2}{1}
461
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteins label neg,],
462
                          aes(x = AveExpr, y = logFC, label = gene), nudge y = -1, nudge x = -1
```

```
1.2) +
        geom label repel(data = DE DEL[DE DEL$logFC > 0 & !is.finite(DE DEL$logFC) & DE DEL$
463
        CountExp > 1, ],
464
                          aes(x = AveExpr, label = gene), y = maxY, nudge y = -1, nudge x = -1)
465
        geom label repel(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC) & DE DEL$
        CountWT > 1, ],
466
                          aes(x = AveExpr, label = gene), y = minY, nudge y = \frac{1}{1}, nudge x = \frac{1}{1})
467
468
469
470
      ggsave("WTvsComp MAplot.png", width = 15, height = 12)
471
      write.table(DE DEL, "WTvsCOMPprotein.tsv", sep = "\t", row.names = F)
472
473
474
475
      proteinList <- acetylation$Accession2</pre>
476
477
478
      ggplot() +
479
        theme bw (base size = 25) +
480
        theme (panel.grid = element blank(),
481
              legend.position = "none") +
482
        labs(y = "Log2 Fold Change", x = "Normalized Average Expression (Log)",
              title = expression("WT vs " * Delta * italic(emp1) * "/comp")) +
483
484
        geom point(data = DE DEL,
485
                    aes(x = AveExpr, y = logFC), color = "grey50",
486
                    alpha = 0.8, size = 0.8) +
487
        geom hline(yintercept = 0, linetype = 1) + #add line at LFC = 0
488
        geom hline(yintercept = c(-LFCcutoff, LFCcutoff), linetype = 2) +
489
        geom segment(data = DE DEL[DE DEL$Accession %in% proteinList,],
490
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = \frac{1}{2} +
                      #add 95% CI lines
        geom segment(data = DE DEL[DE DEL$Accession %in% proteinList,],
491
492
                      aes(x = AveExpr, y = logFC - error, yend = logFC + error), lwd = 0.5,
493
                      color = green) +
        geom point (data = DE DEL[DE DEL$Accession %in% proteinList &
494
495
                                     is.finite(DE DEL$logFC),],
496
                    aes(x = AveExpr, y = logFC), size = 3,
497
                    fill = green,
498
                    shape = 21, stroke = 1.5) +
499
        geom point(data = DE DEL[DE DEL$logFC > 0 & !is.finite(DE DEL$logFC),],
500
                    shape = 23, aes(x = AveExpr), y = maxY, stroke = 1, alpha = 0.6) +
501
        geom point(data = DE DEL[DE DEL$logFC < 0 & !is.finite(DE DEL$logFC),],</pre>
502
                    shape = 23, aes(x = AveExpr), y = minY, stroke = 1, alpha = 0.6) +
503
        coord cartesian(ylim = c(minY, maxY)) +
504
        scale size(range = c(1,3)) +
        geom label repel(data = DE DEL[DE DEL$Accession %in% proteinList,],
505
506
                          aes(x = AveExpr, y = logFC, label = gene), nudge_y = \frac{3}{2}, nudge_x = \frac{-1.2}{2}
507
                          max.overlaps = 40)
508
509
510
      ggsave("WTvsCOMP MAplot acetylationChangers.png", width = 15, height = 12)
511
512
513
514
      Qprot <- addAssay(Qprot,</pre>
515
                         logTransform((Qprot[["raw proteins"]]),
516
                                       base = 10),
517
                         name = "log proteins 10")
518
      Qprot <- addAssay(Qprot,</pre>
519
                         normalize(Qprot[["log proteins 10"]],
520
                                   method = "center.median"),
521
                         name = "norm proteins 10")
522
523
524
525
      #add in old column data to normalized data
```

```
526
      colData(Qprot[["norm proteins 10"]]) <- colData(Qprot)</pre>
527
528
529
      loglog <- Qprot[["norm proteins 10"]] %>% assay() %>% data.frame() %>% rownames to column
      ("Accession")
530
531
      loglogLong <- loglog %>% pivot longer(2:ncol(loglog))
532
      loglogLong$strain <- str extract(loglogLong$name, "^[^]+")</pre>
533
534
      loglogGrouped <- loglogLong %>% group by (Accession, strain) %>%
535
        summarise (Area = mean (value, na.rm = T),
536
                   count = sum(!is.na(value)))
537
538
539
      loglogWide <- loglogGrouped %>% pivot wider(id cols = Accession,
540
                                                     values from = Area,
541
                                                     names from = strain)
542
543
      ggplot(loglogWide) +
544
        geom point (aes (x = WT, y = c1839), color = "grey50",
545
                    size = 0.8, alpha = 0.8) +
546
        theme bw (base size = 25) +
547
        theme (panel.grid = element blank(),
               legend.position = "none") +
548
        geom abline(slope = \frac{1}{2}, intercept = \frac{1}{2}, linetype = \frac{1}{2}, size = \frac{1}{2}) +
549
550
        geom point (data = loglogWide[loglogWide$Accession %in% proteins sig2,],
551
                    aes(x = WT, y = c1839, fill = WT < c1839), size = 3,
552
                    shape = 21, stroke = 1.5) +
553
        scale fill manual(values = colorsUPDOWN) +
554
        labs(title = expression("WT vs " * Delta * italic(emp1) * "/comp"),
555
              x = "Normalized WT expression (log)",
556
              y = expression("Normalized " * Delta * italic(emp1) * "/comp expression (log)"))
557
558
      ggsave ("galaxyWTvsCOMPProteinDE.png", width = 10, height = 10)
559
560
561
      ggplot(loglogWide) +
562
        geom point (aes (x = WT, y = c1839), color = "grey50",
563
                    size = 0.8, alpha = 0.8) +
564
        theme bw (base size = 25) +
565
        theme (panel.grid = element blank(),
566
               legend.position = "none") +
567
        geom abline(slope = \frac{1}{2}, intercept = \frac{1}{2}, linetype = \frac{1}{2}, size = \frac{1}{2}) +
568
        geom point(data = loglogWide[loglogWide$Accession %in% proteinList,],
569
                    aes(x = WT, y = c1839), fill = green, size = 3,
570
                    shape = 21, stroke = 1.5) +
571
        scale fill manual(values = colorsUPDOWN) +
572
        labs(title = expression("WT vs " * Delta * italic(emp1) * "/comp"),
573
              x = "Normalized WT expression (log)",
574
              y = expression("Normalized " * Delta * italic(emp1) * "/comp expression (log)")) +
575
        geom label repel(data = loglogWide[loglogWide$Accession %in% proteinList,],
576
                          aes(x = WT, y = c1839, label = Accession), nudge y = \frac{1}{1}, nudge x = \frac{-1}{1},
577
                          max.overlaps = 40)
578
579
      ggsave("galaxyWTvsCOMPAcetylationChangers labs.png", width = 10, height = 10)
580
581
      ggplot(loglogWide) +
582
        geom point (aes (x = WT, y = c1839), color = "grey50",
583
                    size = 0.8, alpha = 0.8) +
584
        theme bw (base size = 25) +
585
        theme (panel.grid = element blank(),
586
               legend.position = "none") +
587
        geom abline(slope = 1, intercept = 0, linetype = 2, size = 1) +
588
        geom point(data = loglogWide[loglogWide$Accession %in% proteinList,],
589
                    aes(x = WT, y = c1839), fill = green, size = 3,
590
                    shape = 21, stroke = 1.5) +
591
        geom_point(data = loglogWide[loglogWide$Accession %in% proteins_sig2,],
592
                    aes(x = WT, y = c1839, fill = WT < c1839), size = 3,
593
                    shape = 21, stroke = 1.5) +
```