

Inference for quantitation parameters in q-PCR via branching processes with random effects

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Outline

1 Overview

- Overview of the model
- Overview of a qPCR experiment

2 Methodological details

- Sources of randomness
- Branching process details
- Estimating gene copies from qPCR
- Variance of the estimator

3 Analysis of experimental data

- Luteinizing hormone
- *S. vulgaris*

Goal

- Compare expression of some gene between treatment conditions

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What is qPCR?

- qPCR stands for quantitative polymerase chain reaction
- PCR is the chemical reaction by which DNA replicates
 - ▶ e.g. during cell division
- The technology is designed so that PCR replicates only a target gene
- "Quantitative" because we want to count the number of gene copies in a sample
 - ▶ Absolute quantitation: count the gene copies from the original sample
 - ▶ Relative quantitation: find the ratio of copies of one gene, compared to another
 - ▶ The paper covers both; we'll only consider absolute quantitation here

qPCR as a branching process

- PCR is controlled so that each replication cycle k is discrete
- Each particle either doubles or does not during each trial
 - ▶ Probability of replication is typically high ($0.9 < p$)
- This defines a supercritical branching process that leads to exponential growth
- During early cycles ($k < 15$, say), the count is obscured by noise
- Availability of reaction chemicals attenuates the reaction after ~ 30 cycles
- The cycles between 15 and 30 are called the exponential phase.

The experimental setup

A typical experimental setup is to have:

- Two treatment groups (T=treatment, C=control)
- Two genes under study
- Three replicates for each gene-treatment combination
- So an experiment typically involves twelve reactions

The experimental procedure

Do the following for each reaction:

```
for (cycle in 1:40):  
    count the gene copies  
    use PCR to produce a new generation
```


Sample experimental data

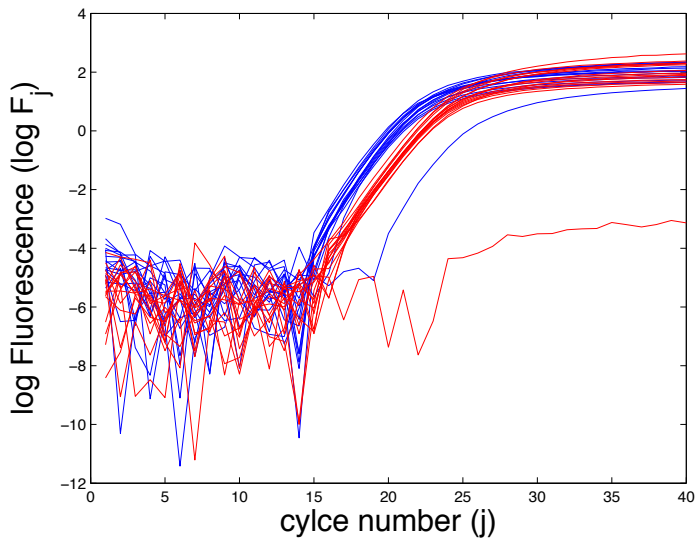


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Sources of randomness

- N_0 , the initial number of gene copies, is random
 - ▶ $E[N_0] = m_a$
 - ▶ $\text{var}(N_0) = \sigma_a^2$
- At each cycle of the reaction, each gene copy replicates randomly
 - ▶ $N_{n+1} = N_n + \text{Bin}(N_n, p)$

qPCR as a branching process

Note:

$$\begin{aligned} E[N_n] &= E[E(N_n|N_{n-1})] = E[(1+p)N_{n-1}] \\ &= \cdots = (1+p)^n \times E(N_0) \end{aligned}$$

- So $W_n = \frac{N_n}{(1+p)^n}$ is a positive martingale
- Thus, $W_n \rightarrow W$ almost surely for some W
- $E[W] = m_a$

qPCR as a branching process

Consider an idealized reaction experiment:

- If we knew p and could let the number of reaction cycles $n \rightarrow \infty$:
 - ▶ $W_i = \lim_{n \rightarrow \infty} \frac{N_{i,n}}{(1+p)^n}$
 - ▶ $W_1, W_2, \dots, W_r \stackrel{\text{iid}}{\sim} W$
 - ▶ So $\frac{1}{r} \sum_{i=1}^r W_i \xrightarrow{\text{a.s.}} E[W] = m_a$
- But p is unknown and we can only observe ≈ 15 reaction cycles, so we need some other estimator.

Estimating p

- Since p is unknown, we estimate it with \hat{p} via weighted least squares:

$$\begin{pmatrix} N_n \\ N_{n-1} \\ \vdots \\ N_1 \end{pmatrix} - \begin{pmatrix} N_{n-1} \\ N_{n-2} \\ \vdots \\ N_0 \end{pmatrix} = p \times \begin{pmatrix} N_{n-1} \\ N_{n-2} \\ \vdots \\ N_0 \end{pmatrix} + \epsilon$$

- Where $\epsilon_j \stackrel{\text{approx.}}{\sim} \text{Normal}(0, p(1-p)N_{j-1})$
- With weights $W_j = (N_{j-1})^{-1}$ the resulting estimator is:

$$\hat{p} = \frac{\sum_{i=1}^n (N_i - N_{i-1})}{\sum_{i=1}^n N_i}$$

Making the most of a finite sample

Reminder: our idealized estimator was $W(n) = \frac{N_n}{(1+p)^n}$

- W uses only the final observation (N_n)
- More efficient: use the sum $Y_n = \sum_{i=1}^n N_i$
- By the Toeplitz Lemma, $\frac{Y_n}{(1+p)^n} \xrightarrow{\text{a.s.}} \frac{1+p}{p} W \Rightarrow \frac{pY_n}{(1+p)^{n+1}} \xrightarrow{\text{a.s.}} W$
- Plug in \hat{p} and the limit still holds.

Strategy for quantitation

- Collect data on r independent reactions
- For reaction i ($i = 1, 2, \dots, r$), compute the statistic $M_i = \frac{\hat{p}_i Y_{n_i}}{(1+\hat{p}_i)^{n_i+1}}$
- Average M_1, M_2, \dots, M_r to get \bar{M}
- $\sqrt{r}(\bar{M} - m_a) \xrightarrow{d} \text{Normal}(0, \sigma_L^2)$
- Where $\sigma_L^2 = \sigma_a^2 + m_a E[\frac{1-p}{1+p}]$

Variance of the estimator

$$\begin{aligned}\sigma_L^2 &= \text{var}\left[\frac{N_j}{(1+p)^j}\right] = E(\text{var}\left[\frac{N_j}{(1+p)^j} | p\right]) + \text{var}(E\left[\frac{N_j}{(1+p)^j} | p\right]) \\ &= E(\text{var}\left[\frac{N_j}{(1+p)^j} | p\right]) + \text{var}(m_a) \\ &= E(\text{var}\left[\frac{N_j}{(1+p)^j} | p\right])\end{aligned}$$

Variance of the estimator

$$\begin{aligned}\text{var}\left[\frac{N_j}{(1+p)^j} \middle| p\right] &= \frac{1}{(1+p)^{2j}} \text{var}[N_j | p] \\&= \frac{1}{(1+p)^{2j}} (E(\text{var}[N_j | N_{j-1}, p] | p) + \text{var}(E[N_j | N_{j-1}, p] | p)) \\&= \frac{1}{(1+p)^{2j}} (E[N_{j-1} p (1-p) | p] + \text{var}((1+p)N_{j-1} | p)) \\&= \frac{1}{(1+p)^{2j}} (m_a (1+p)^{j-1} p (1-p) + (1+p)^2 \text{var}[N_{j-1} | p]) \\&= \frac{m_a p (1-p)}{(1+p)^{j+1}} + \frac{\text{var}[N_{j-1} | p]}{(1+p)^{2j-2}} \\&= \dots \\&= \frac{m_a p (1-p)}{(1+p)^{j+1}} + \frac{m_a p (1-p)}{(1+p)^j} + \dots + \frac{m_a p (1-p)}{(1+p)^2} \\&\quad + \frac{\text{var}[N_0 | p]}{(1+p)^{2j-2j}}\end{aligned}$$

Variance of the estimator

$$\begin{aligned}\text{var}\left[\frac{N_j}{(1+p)^j} \mid p\right] &= \frac{m_a p(1-p)}{(1+p)^2} \sum_{k=0}^{j-1} \frac{1}{(1+p)^k} + \sigma_a^2 \\ &= m_a \frac{1-p}{1+p} + \sigma_a^2\end{aligned}$$

So:

$$\begin{aligned}\text{var}\left[\frac{N_j}{(1+p)^j}\right] &= E(\text{var}\left[\frac{N_j}{(1+p)^j} \mid p\right]) \\ &= E\left(m_a \frac{1-p}{1+p} + \sigma_a^2\right) \\ &= m_a E\left(\frac{1-p}{1+p}\right) + \sigma_a^2 = \sigma_L^2\end{aligned}$$

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Experimental data - luteinizing hormone

- The goal with the experimental data was *relative* quantitation
 - ▶ Estimate ratio of gene expression between conditions C and T
- The sample was divided into two parts
- One part was diluted to one-third the original concentration
- Sixteen reactions were run under each condition (diluted, normal)

Experimental data - luteinizing hormone

Luteinizing Hormone

	BP	C_T method	Std. Curve	Adj C_T
\hat{R}	2.8221	3.3108	3.5558	3.9567
GCI	[1.8624, 3.7817]	.	.	.
tCI	[1.7719, 3.8722]	.	.	.
BCI	[1.6870, 3.6013]	[2.7935, 3.7477]	[2.8646, 4.1355]	[2.9024, 5.4157]

Experimental data - *Strongylus vulgaris*

- Again, the goal of the was relative quantitation
- One part diluted to one-tenth the original concentration
- Ten reactions run under each condition (diluted, normal)

Experimental data - *Strongylus vulgaris*

S. vulgaris

	BP	C_T method	Adj C_T
\hat{R}	10.1793	6.3622	303.1662
GCI	[2.8686, 17.4900]	.	.
tCI	[1.7414, 18.6172]	.	.
BCI	[1.0446, 15.7493]	[5.2733, 7.3176]	[29.7583, 892.6178]