Data Integration in the Life Science

Report - Integration of Big Biological Data

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Introduction

This report serves as a presentation for our project of Data Integration course. The idea is to concretely integrate several biological datasets while following a process enhancing the reproducibility using the general methods like scripts and data set setting.

Data integration involves combining data residing in different sources and providing users with a unified view of these data. This process becomes significant in a variety of situations, which include both commercial and scientific domains especially in bioinformatics.

Bioinformatics uses the methods of mathematics, informatics, statistics, and computer science to study biology. Now, the main research directions of bioinformatics are: sequence alignment, gene recognition, gene recombination, protein structure prediction, gene expression, protein response prediction, and the establishment of evolutionary models.

This project follows following steps: Firstly, we download the datasets from UniProtKB and NCBI. The dataset of UniProtKB can be downloaded with different formats like plain text, XML and RDF. We used XML format for this project. The datasets downloaded from NCBI are already well annotated with column names. Then, we should design a database model for store the data. For this project, we used the relational database management system MySQL. The step followed is to populate the data into the model. The database model we designed will decide how we populate the data. Finally, we will do some experiments and analyses based on the data we populated, especially relationship of the data from two different sources (UniPort and NCBI).

For the following part, we will first get a glimpse of the data, especially the data from UniProtKB which is more difficult to handle. Then, we will make a detailed description on the work we did. And in the last part, we will make a conclusion and talk about the difficulties we encountered during this project.

Datasets

UniProtKB

In this project, we used the congenital disease dataset in the form of XML downloaded from UniProtKB. To get this dataset, we first go to http://www.uniprot.org, then search *congenital disease AND reviewed:yes*, we will get the query result like this:

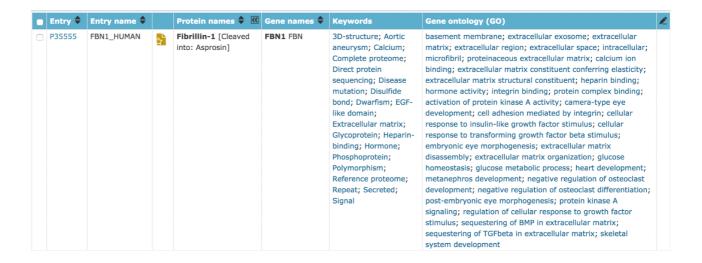


Figure 1 - Query result of e congenital disease

As it is said in the subject, for this project, we are only interested in these fields:

- ID, AC, DE, GN, KW
- DR but only the lines starting with GO (to get the gene ontology annotations)

In the above of the table, we can modify the column configuration to get only the columns that we are interested. For instance, we selected to show the columns: Entry, Entry name, Protein names, Gene names, Keywords and Gene ontology (GO).

In fact, these are columns are exactly correspond to the fields we mentioned above. The mapping is:

ID	Entry name
AC	Entry
DE	Protein names
GN	Gene names
KW	Keywords

DR with GO Gene ontology (GO)

It should be noted that, in the figure 1, the column only shows the gene ontology term without the GO id like GO:0005604.

We can more details on every entry and the characteristics of the columns by clicking on the Entry.

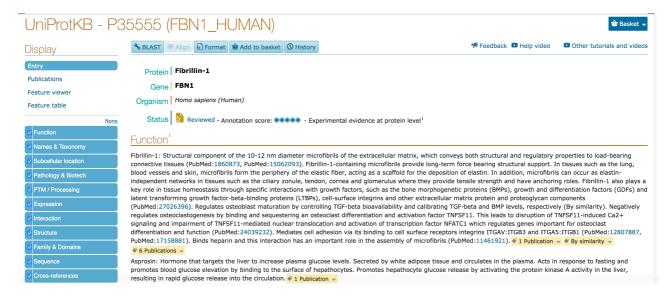


Figure 2 - Entry details

The figure 2 above shows the details of an entry. For instance, we can see the protein name, the gene name and many other details.



Figure 3 - Protein details

Besides, we can find the protein name details here. The figure 3 shows the protein details of the entry **P35555** we illustrated above. From this table information, we know that **Fibrillin-1** is the recommended name of the protein which linked to the disease entry.

Moreover, we can click on the Protein names (with a letter 's' as the superscript) to find more details about this column.

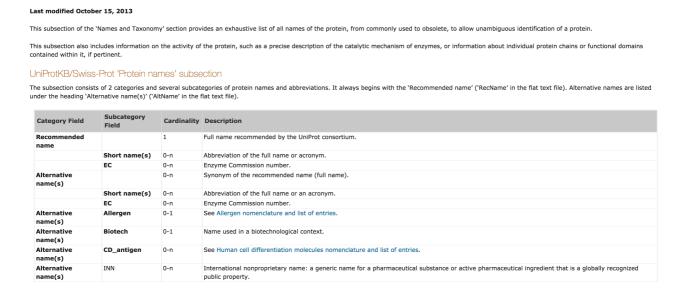


Figure 4 - Protein name details

The figure above shows the details of the column Protein names we talked about in the Figure 1. We can see the cardinality of every category field. For instance, the cardinality of the field Recommended name is 1. This means that in the Protein names section, it can only have one recommended name. This can be illustrated more concretely in the XML file as below.

```
3  <entry dataset="Swiss-Prot" created="1994-06-01" modified="2016-11-30" version="204">
4  <accession>P35555</accession>
5  <accession>B2RUU0</accession>
6  <accession>D2JYH6</accession>
7  <accession>015972</accession>
8  <accession>075N87</accession>
9  <name>FBN1_HUMAN</name>
10  <protein>
11  <recommendedName>
12  <fullName>Fibrillin-1</fullName>
13  </recommendedName>
14  <component>
15  <recommendedName>
16  <fullName evidence="107">Asprosin</fullName>
17  </recommendedName>
18  </component>
19  </protein>
```

Figure 5 - The protein element of XML data

Here, it's also the first entry P35555, we can see that inside the protein element, there is only one recommended name element named recommendedName. It should be noted

Protein names

that the second recommendedName inside the protein element clause belongs to component element. This correspond to the component of the protein, which we don't concern about here.

UCBI

As we said before, the two dataset, namely *Homo_sapiens.gene_info* and *gene2GO* are well annotated. Here is a simple exploration using Python script and Pandas library.

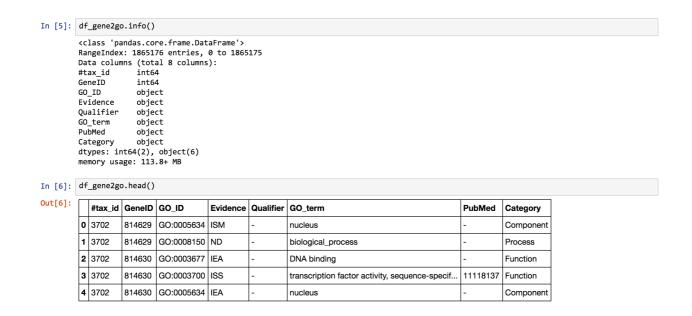


Figure 6 - gene2go dataset

```
In [7]: df_geneInfo = pd.read_table("data/Homo_sapiens.gene_info")
In [8]: df geneInfo.info()
         <class 'pandas.core.frame.DataFrame';</pre>
         RangeIndex: 59652 entries, 0 to 59651
         Data columns (total 15 columns):
                                                     59652 non-null int64
         GeneID
                                                     59652 non-null int64
         Symbol
                                                     59652 non-null object
                                                     59652 non-null object
         Synonyms
                                                     59652 non-null object
         dbXrefs
                                                    59652 non-null object
         chromosome
                                                     59652 non-null object
         map_location
                                                     59652 non-null object
         description
                                                    59652 non-null object
                                                    59652 non-null object
         type of gene
         Symbol_from_nomenclature_authority
                                                     59652 non-null object
         {\tt Full\_name\_from\_nomenclature\_authority}
                                                    59652 non-null object
                                                    59652 non-null object
         Nomenclature status
         Other_designations
        Modification_date
dtypes: int64(3), object(12)
                                                    59652 non-null int64
         memory usage: 6.8+ ME
```

Figure 7 - Home_sapiens_gene_info dataset (Jupyter notebook)

As we can see, the *gene2go* dataset downloaded from UCBI has eight columns and the *Home_sapiens_gene_info* dataset has total 15 columns. We can easily import these data directly into a relational database using the embedded data structure of Pandas like dataFrame.

Description of Work

Database Model

To populate the data, we need firstly design the database model. The figure below illustrates the UML model of the database.

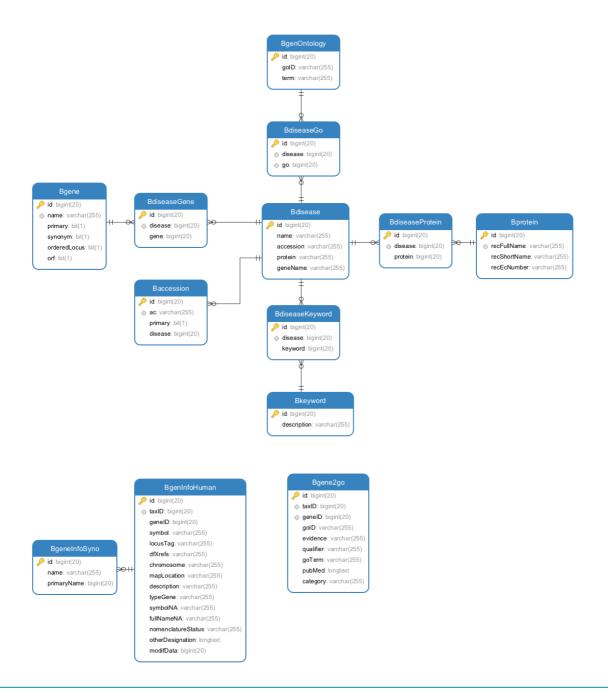


Figure 8 - Database model

The name of every table in the database starts with a capital B. Here, for instance, *Bdisease* represents one entry (one row) in the Figure 1. It has 5 fields: id, name, accession, protein and geneName. The filed *id* is the row number, the field *name* is the Entry name illustrated in the Figure 1, the field *accession* is the recommended (citable) ac number of this entry. The field *protein* is the recommended name of the protein and the filed *geneName* is the primary gene name (corresponding to the gene/name element with a primary attribut in the XML data).

As we know, every disease (i.e. every entry) has more than one accession numbers, has more than one protein names, gene names, keywords and gene ontology, vice versa. Consequently, we have to put an intermediate between the table *Bdisease* and each of the other four. This is the reason why there exist tables like BdiseaseProtein which contains mapping between the two tables to simplify the N-N relationship.

For the other two datasets, we integrate them directly keeping the columns structures. Here, it should be noted that the synonyms of protein in the dataset UCBI are not well formed. Every gene has several synonyms, separating by a vertical line. So we separate them using scripts and store them in the table *BgeneInfoSyno*.

Populate the data

To populate the three datasets, we use Python Script. Firstly, to make sure the reproducibility. The version of techniques and dependencies that we used in this project are listed here:

Python	2.7.11
Pandas	0.19
mysql-python	1.2.5
MySQL	5.7.12
Jupyter notebook	4.2.3

We used the package manage software Anaconda to manage these libraries and software. The Anaconda is running on a Macbook computer and is the latest version of 64bits.

Firstly, we should populate the XML data. Here we have used a very import method: **XPath**. It is an XML-based tree structure which provides the ability to find nodes in a data structure tree. We can easily find the interesting element we want using this tool.

Python already has XML package installed. To use the XPath, we should import ElementTree module. We use the parse() method to parse the XML file and then get the root element. Then, we can use the findall() method to find the interested element in the XML tree structure.

After finding the data we want, we should then populate it into the relational database. To do this, we use the mysql-python package listed before. It's a popular package in the Python Eco-system to handle the MySQL connection problem.

We have implemented a module named MySQLConnector, which is a Python class for connecting the MySQL server and accelerating the development of project. In this class, there are a set of general methods such as **select()**, **insert()**, **update()** and **delete()**. These methods take the name of table, the condition, the selected columns as parameters to form the SQL command and return a list of results. This module can be used to handle most of the iteration problems with MySQL databases.

```
def __open(self):
    try:
        cnx = MySQLdb.connect(self.__host, self.__user, self.__password, self.__database, port=3306)
        self.__connection = cnx
        self.__session = cnx.cursor()
    except MySQLdb.Error as e:
        print "Error %d: %s" % (e.args[0], e.args[1])
```

Figure 9 - General method **open()**

```
def select(self, table, where=None, *args, **kwargs):
    result = None
query = 'SELECT'
    keys = args
    values = tuple(kwargs.values())
    1 = len(keys) - 1
    for i, key in enumerate(keys):
        query += "`" + key +
if i < 1:</pre>
             query += ","
    query += 'FROM %s' % table
    if where:
        query += " WHERE %s" % where
    self.__open()
    self.__session.execute(query, values)
    number_rows = self.__session.rowcount
number_columns = len(self.__session.description)
    if number_rows >= 1 and number_columns > 1:
        result = [item for item in self.__session.fetchall()]
        result = [item[0] for item in self.__session.fetchall()]
    self.__close_con()
    return result
```

Figure 10 - Method select()

The two figures above illustrated the ideas of this module. For instance, the select method takes a table name as the first parameter, a condition, a set of selected column named (*args) and a set of conditions (**kwargs).

The whole process is a streaming method, which means we iterate the XML file from the beginning to the end only one time and we should get all we want. Concretely, the entrance of the element we want is **entry** element. An entry corresponds to one row of Figure 1, but contains all the information about this entry. For instance, to get the accession number of an entry, we use the findall() method to looking for the direct descend child the entry element whose element name is <accession>. Then we get the text data and populate to the database.

The population of the other two datasets is more simple, we populate directly into two table, except for the synonyms of protein, which we use another table to store.

Analysis/Queries

After populating the data, we should write two queries/scripts to find the genes having the same official first name UCBI and UniProtKB but with different synonyms names. Also, we want to find the genes having the same first name but with different gene ontology terms.

For the first one, we just need to find out the genes with the same official first names in the two sources. Then, we compare their corresponding synonym name one by one to find check out if they have different synonym (secondary) names. The same for the comparison of gene ontology, but we should use the gene2go dataset to get the gene ontology information.

Difficulties

The parser program can correctly parse the dataset. But there is still a problem we haven't overcome. The MySQL server can be suddenly closed during the execution of a command. We have to wait until the connection is re-established. This problem may due to the frequent close actions during the whole parsing processus.

Conclusion

This project gives us a new vision of data integration. We have experienced each step of a data integration processus including downing the source, pre-precessing, populating the data and data analysis based on the data model.

The bioinformatics data is well formed and important. We can use these data to apply to many different interesting areas like medicine, health-care industry.