

Fossil-calibrated phylogeny of Onagraceae: patterns of floral and chromosome evolution

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Abstract

blah blah

1. Introduction

blah blah

2. Methods

Supermatrix assembly. I downloaded all available DNA sequences from Gen-
Bank release 200 PLN division and performed an exhaustive all-by-all BLASTn
5 (Camacho et al., 2009) comparison for sequences in Onagraceae and Lythraceae.
Using a BLASTn e-value of $1.0e^{-10}$ threshold and a sequence length percent sim-
ilarity cutoff of 0.5, I constructed clusters of putative homologs using a single-
linkage hierarchical clustering algorithm. Subspecies names were removed from
10 all sequences, and all but one sequence of each species was pruned from each
cluster. Clusters that were not phylogenetically informative (< 4 taxa) were
discarded, and each cluster was aligned using MUSCLE (Edgar, 2004). The
alignments were concatenated by species, and any species that was not present
in at least two clusters was removed from the supermatrix. Code written to as-
15 semble the supermatrix is available as the Python module SUMAC (Freyman,

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Group	Age (Mya)	Prior Distribution	Mean	SD	Offset	Reference
<i>Circaea</i> (Onagraceae)	12	lognormal	0.0	2.0	12	(Grímsson et al., 2012)
<i>Epilobium</i> (Onagraceae)	12	lognormal	0.0	2.0	12	(Grímsson et al., 2012)
S. Pacific <i>Fuschia</i> (Onagraceae)	23	lognormal	0.0	1.0	23	(Lee et al., 2013)
<i>Ludwigia</i> (Onagraceae)	Paleocene	normal	60.0	3.0	-	(Zhi-Chen et al., 2004)
Lythraceae	82	lognormal	0.0	2.0	82	(Graham, 2013)

Table 1: Fossils used as priors in the Bayesian divergence time analysis.

2014, <https://github.com/wf8/sumac>), and can be used to assemble supermatrices for other taxonomic groups recognized in GenBank.

Phylogenetic analyses. Maximum likelihood analyses were performed with RAxML-
 HPC (Stamatakis, 2014) on the CIPRES Scientific Gateway (Miller et al., 2010)
 20 using the rapid bootstrap heuristic and the GTRCAT nucleotide substitution
 model. I used the ML tree to select 15 taxa phylogenetically widely distributed
 in Lythraceae to act as outgroup for the divergence time analysis; all other mem-
 bers of Lythraceae were subsequently removed from the supermatrix. Bayesian
 estimates of divergence times were inferred using BEAST v1.8 (Drummond and
 25 Rambaut, 2007; Suchard and Rambaut, 2009) on CIPRES and calibrated with
 five fossils identified with morphological synapomorphies (Table 1). The *Lud-*
wigia fossil pollen was dated broadly to the Paleocene (Grímsson et al., 2012),
 so I set the prior to a normal distribution with a wide standard deviation to
 cover the entire time period. For all other calibration points I used a lognormal
 30 prior distribution with the offset (the minimum age of the node) corresponding
 to the fossil age. The BEAST analysis utilized the GTR+ Γ nucleotide substitu-
 tion model with a relaxed molecular clock (uncorrelated lognormal model) and
 a Yule process tree prior. The Markov Chain Monte Carlo (MCMC) was run for
 100 million generations, sampling every 10 thousand generations. Tracer v1.6
 35 (Rambaut et al., 2013) was used to assess the MCMC output for parameter
 convergence and ensure that the effective sample size for all parameters was
 above 200. The first 1000 trees were discarded as burn-in, and the remaining
 9000 trees were summarized as a maximum clade credibility (MCC) tree with
 mean divergence times.

40 *Character state reconstruction.* I scored six characters, including chromosome
number, floral merosity, petal color, and self-compatibility/incompatibility. Char-
acter data was assembled from the comprehensive Wagner et al. (2007) Ona-
graceae monograph. Ancestral character state reconstructions of petal number
and petal color were performed using Mesquite v2.75 (Maddison and Maddison,
45 2011) over the Bayesian MCC tree. Characters were treated as unordered cate-
gorical data, and optimized using maximum likelihood with the Markov k-state
1 parameter (Mk1) model (Lewis, 2001).

3. Results

Supermatrix assembly. blah blah

50 *Divergence time estimates.* Here is a table with the mean divergence time esti-
mates including %95 confidence intervals....

Character evolution. blah blah

4. Conclusion

blah blah

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Clade	Mean Age (Mya)	%95 HPD Min	%95 HPD Max
Onagraceae / Lythraceae	109	88	131
<i>Ludwigia</i>	97	76	118
<i>Hauiya</i>	49	35	64
<i>Circaea</i> / <i>Fuchshia</i>	37	28	47
<i>Lopezia</i>	71	55	68
<i>Gongylocarpus</i>	60	45	77
<i>Epilobium</i>	49	38	60
<i>Chamerion</i>	47	36	57
<i>Xylonagra</i>	43	33	52
<i>Clarkia</i>	40	32	48
<i>Terapteron</i>	19	10	29
<i>Camissoniopsis</i> / <i>Neoholmgrenia</i>	14	5	23
<i>Eremothera</i> / <i>Camissonia</i>	24	16	33
<i>Taraxia</i>	30	22	38
<i>Chylismiella</i> / <i>Gayophytum</i>	20	10	30
<i>Eulobus</i>	26	19	34
<i>Chylismia</i> / <i>Oenothera</i>	25	18	31

Table 2: Bayesian divergence time estimates of major clades.

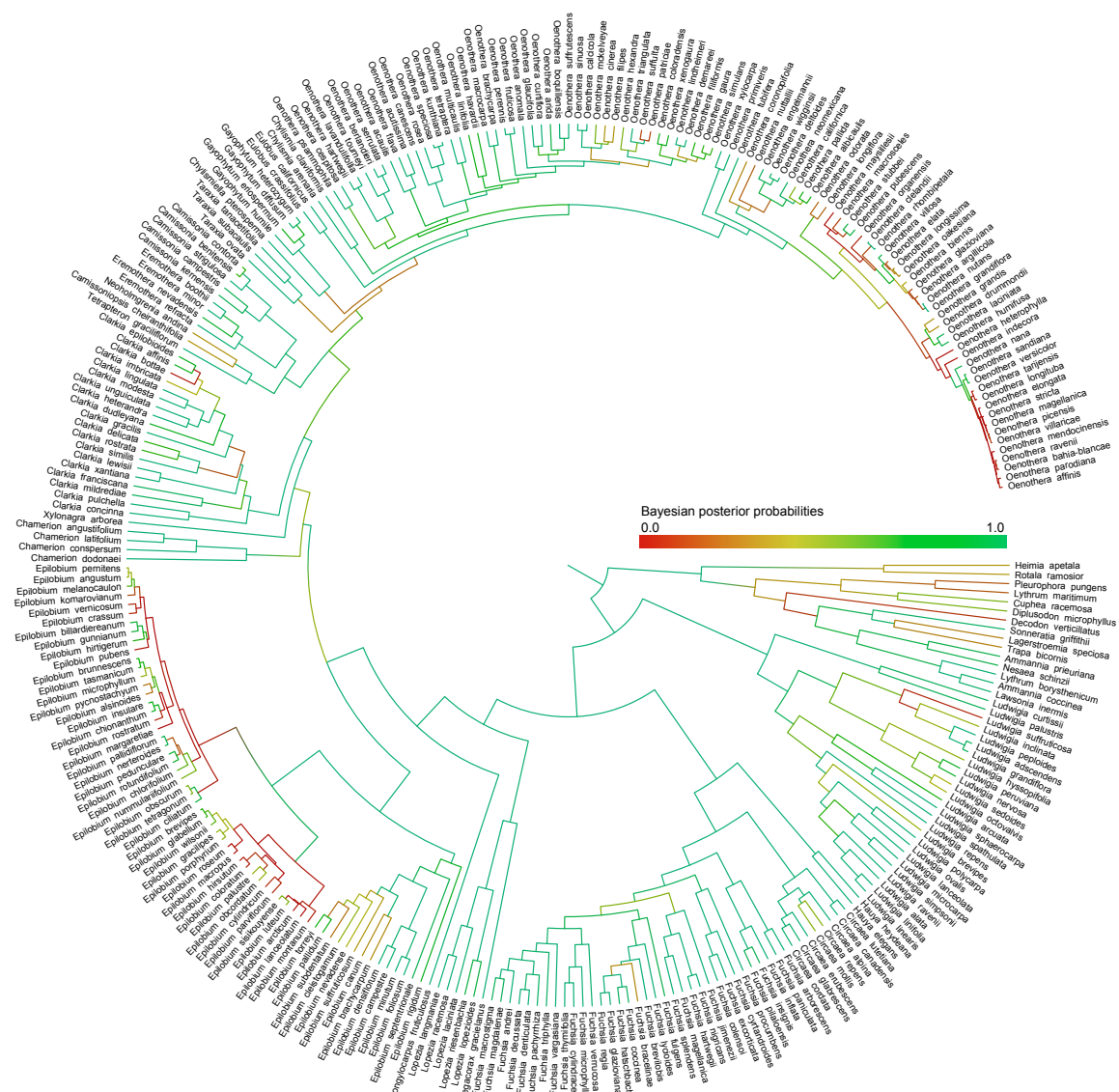


Figure 1: Bayesian maximum clade credibility phylogeny of 280 Onagraceae taxa and 15 Lythraceae taxa. Estimated posterior probabilities close to 1.0 are shown in green. All genera described in Wagner et al. (2007) are monophyletic with posterior probabilities of > 0.95 except for sister genera *Neoholmgrenia* and *Camissoniopsis* (posterior = 0.31).

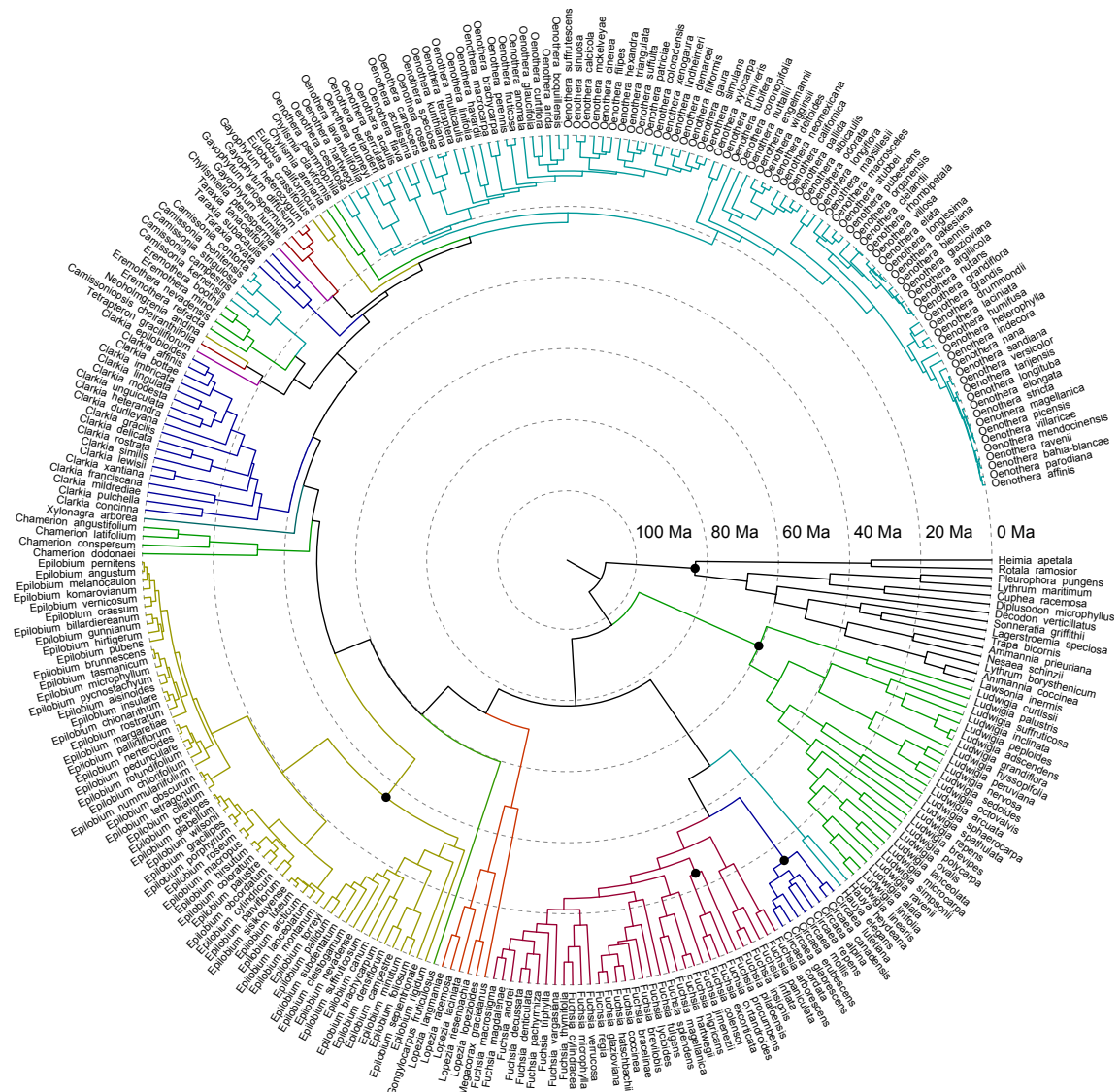


Figure 2: Bayesian chronogram of 280 Onagraceae taxa and 15 Lythraceae taxa. Approximate positions of fossil calibration points are shown as black circles. All genera described in Wagner et al. (2007) are colored, and their divergence time estimates and %95 HPD intervals can be seen in Table 2.

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