# DSA 8020 R Session 10: Random and Mixed Effects Models and Computer Experiments

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## Contents

Random Effects Example	1
Read the data into R	. 1
Fitting a fixed effects model	. 2
Fitting a random effects model	. 3
RCBD: Fixed vs. Random Block	. 3
Load R libraries	. 3
Read the data	. 4
Fixed block	. 4
Random block	. 4
Computer Experiments	. 5
Design: Latin hypercube	5
Analysis: Gaussian Process	7

## Random Effects Example

Suppose that an agronomist is studying a large number of varieties of soybeans for yield. The agronomist randomly selects three varieties, and then randomly assigns each of those varieties to 10 of 30 available plots.

Model:  $y_{ij} = \mu + \alpha_i + \epsilon_{ij}$ ,  $\alpha_i s \overset{i.i.d.}{\sim} N(0, \sigma_{\alpha}^2)$ ,  $\epsilon_{ij} s \overset{i.i.d.}{\sim} N(0, \sigma^2)$ .  $\alpha_i s$  and  $\epsilon_{ij} s$  are independent to each other

#### Read the data into R

```
v1 <- c(6.6, 6.4, 5.9, 6.6, 6.2, 6.7, 6.3, 6.5, 6.5, 6.8)

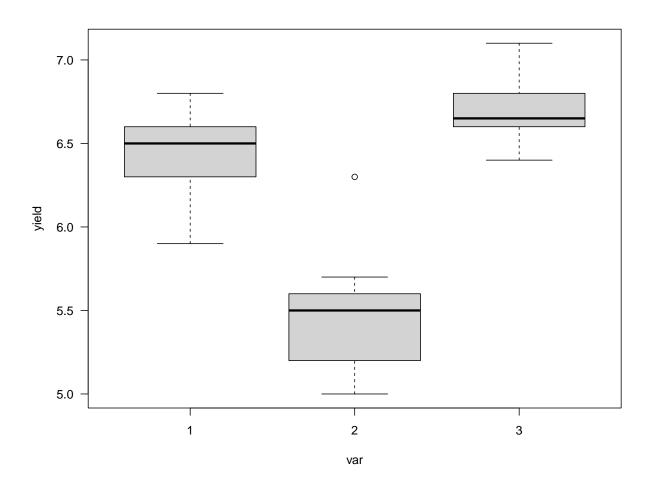
v2 <- c(5.6, 5.2, 5.3, 5.1, 5.7, 5.6, 5.6, 6.3, 5.0, 5.4)

v3 <- c(6.9, 7.1, 6.4, 6.7, 6.5, 6.6, 6.6, 6.6, 6.8, 6.8)

yield <- c(v1, v2, v3)

var <- factor(c(rep(1, 10), rep(2, 10), rep(3, 10)))

plot(yield ~ var, las = 1)
```



## Fitting a fixed effects model

6.45

-0.97

##

```
fixef <- lm(yield ~ var)</pre>
anova(fixef)
## Analysis of Variance Table
##
## Response: yield
##
             Df Sum Sq Mean Sq F value
                                          Pr(>F)
              2 8.306 4.1530 49.593 9.114e-10 ***
## var
## Residuals 27 2.261 0.0837
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' ' 1
coefficients(fixef)
## (Intercept)
                      var2
                                  var3
```

0.25

#### Fitting a random effects model

```
library(lme4)
## Loading required package: Matrix
randef <- lmer(yield ~ 1 + (1|var), REML = TRUE)</pre>
summary(randef)
## Linear mixed model fit by REML ['lmerMod']
## Formula: yield ~ 1 + (1 | var)
## REML criterion at convergence: 21.6
## Scaled residuals:
##
       Min
               1Q Median
                                ЗQ
                                       Max
## -1.8839 -0.6181 0.1118 0.4962 2.7828
##
## Random effects:
## Groups Name
                         Variance Std.Dev.
            (Intercept) 0.40693 0.6379
## var
                         0.08374 0.2894
## Residual
## Number of obs: 30, groups: var, 3
##
## Fixed effects:
              Estimate Std. Error t value
## (Intercept) 6.2100
                            0.3721 16.69
Let's construct CIs for \sigma_{\alpha}^2, \sigma^2, and \mu
## Compute the confidence intervals (CIs) using profile likelihood
CIs <- confint(randef, oldNames = FALSE)</pre>
## Computing profile confidence intervals ...
CIs
                          2.5 %
                                   97.5 %
## sd_(Intercept)|var 0.2637525 1.5512218
## sigma
                  0.2265053 0.3877781
## (Intercept)
                      5.3618584 7.0581407
RCBD: Fixed vs. Random Block
```

## Load R libraries

```
library(lsmeans)
library(lmerTest)
```

#### Read the data

```
### Create the data set
x <- c(52, 47, 44, 51, 42, 60, 55, 49, 52, 43, 56, 48, 45, 44, 38)
trt <- rep(c("A", "B", "C"), each = 5)
blk <- rep(1:5, 3)
dat <- data.frame(x = x, trt = trt, blk = as.factor(blk))</pre>
```

#### Fixed block

#### Random block

```
randef <- lmer(x ~ trt + (1|blk), REML = TRUE, data = dat)</pre>
summary(randef)
## Linear mixed model fit by REML. t-tests use Satterthwaite's method [
## lmerModLmerTest]
## Formula: x ~ trt + (1 | blk)
##
     Data: dat
##
## REML criterion at convergence: 71.1
## Scaled residuals:
      Min 1Q Median
##
                             3Q
                                     Max
## -1.1417 -0.6147 -0.1494 0.5772 1.3390
##
## Random effects:
## Groups Name
                       Variance Std.Dev.
## blk
            (Intercept) 28.35
                              5.324
## Residual
                        5.85
                                2.419
## Number of obs: 15, groups: blk, 5
##
## Fixed effects:
##
              Estimate Std. Error
                                  df t value Pr(>|t|)
## (Intercept) 47.200 2.615 5.054 18.047 8.76e-06 ***
```

```
## trtB
                 4.600
                           1.530 8.000
                                         3.007
                                                  0.0169 *
## trtC
                -1.000
                           1.530 8.000 -0.654
                                                  0.5316
## ---
## Signif. codes: 0 '*** 0.001 '** 0.01 '* 0.05 '.' 0.1 ' 1
## Correlation of Fixed Effects:
       (Intr) trtB
## trtB -0.292
## trtC -0.292 0.500
lsmeans(randef, list(pairwise ~ trt), adjust = "none")
## $'lsmeans of trt'
## trt lsmean SE
                     df lower.CL upper.CL
## A
         47.2 2.62 5.05
                           40.5
                                    53.9
## B
         51.8 2.62 5.05
                           45.1
                                    58.5
## C
         46.2 2.62 5.05
                           39.5
                                    52.9
##
## Degrees-of-freedom method: kenward-roger
## Confidence level used: 0.95
##
## $'pairwise differences of trt'
         estimate SE df t.ratio p.value
             -4.6 1.53 8 -3.007 0.0169
## A - B
## A - C
             1.0 1.53 8
                           0.654 0.5316
## B - C
              5.6 1.53 8
                           3.661 0.0064
## Degrees-of-freedom method: kenward-roger
```

## Computer Experiments

Design: Latin hypercube

```
# install.packages("lhs") # Latin Hypercube Sample #Package
library(lhs)
# Generate a good n x k LHD
LHD = maximinLHS(n = 30, k = 2, dup = 5)
# "dup" is an integer tuning parameter that determines the number of
# candidate points considered. Larger values should inprove results
# but require more computational resources.
# Display the LHD
LHD
```

```
## [,1] [,2]

## [1,] 0.153124122 0.74125924

## [2,] 0.700805556 0.87879513

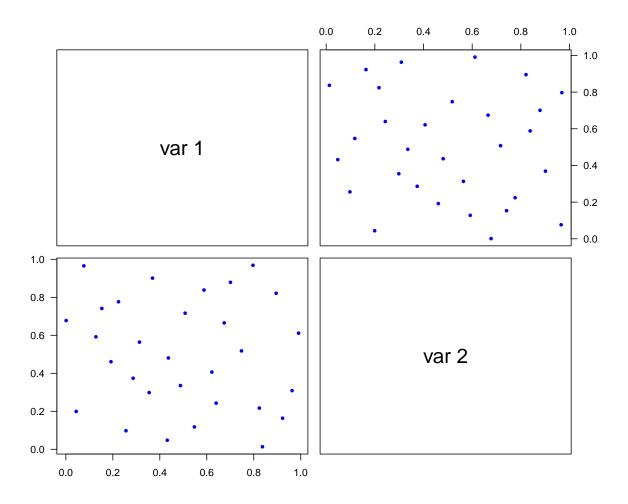
## [3,] 0.286366499 0.37400299

## [4,] 0.922708305 0.16367775

## [5,] 0.487996157 0.33548689

## [6,] 0.747629143 0.51789241
```

```
## [7,] 0.127992460 0.59206000
## [8,] 0.547269613 0.11773931
## [9,] 0.313338413 0.56407709
## [10,] 0.507805155 0.71654250
## [11,] 0.823827647 0.21695917
## [12,] 0.256026314 0.09764923
## [13,] 0.621629609 0.40638483
## [14,] 0.895286514 0.82115117
## [15,] 0.588290882 0.83823162
## [16,] 0.224194450 0.77652176
## [17,] 0.354814611 0.29852273
## [18,] 0.192092881 0.46089337
## [19,] 0.076504682 0.96544849
## [20,] 0.674212717 0.66531212
## [21,] 0.963176641 0.30887109
## [22,] 0.639809715 0.24298214
## [23,] 0.431783698 0.04765659
## [24,] 0.001182855 0.67762407
## [25,] 0.796737649 0.96853772
## [26,] 0.436685562 0.48074595
## [27,] 0.990953378 0.61141780
## [28,] 0.043934178 0.19928618
## [29,] 0.836982981 0.01321333
## [30,] 0.368944972 0.90109887
pairs(LHD, col = "blue", cex = 0.8, pch = 16, las = 1)
```



## Analysis: Gaussian Process

```
# Load the data
neuron <- read.table("http://deanvossdraguljic.ietsandbox.net/DeanVossDraguljic/R-data/neuron.txt", head(neuron, 10)</pre>
```

```
## 1 9NaFsc gKdrsc fr
## 1 0.38593729 0.2120652 33
## 2 0.04666927 0.4594742 0
## 3 1.0000000 0.4473344 46
## 4 0.95467637 0.3351407 44
## 5 0.53334929 0.7981310 41
## 6 0.59166751 0.6042714 41
## 7 0.18570301 0.3799469 31
## 8 0.49927784 0.2444170 36
## 9 0.74609113 0.3949591 42
## 10 0.07269414 1.0000000 0
```

```
# Fit a GP
library(mlegp)
GPFit <- mlegp(neuron[, 1:2], neuron[, 3])</pre>
## no reps detected - nugget will not be estimated
## ======= FITTING GP # 1 ==========
## running simplex # 1...
## ...done
## ...simplex #1 complete, loglike = -104.446501 (convergence)
## running simplex # 2...
## ...done
## ...simplex #2 complete, loglike = -104.446501 (convergence)
## running simplex # 3...
## ...done
## ...simplex #3 complete, loglike = -104.446502 (convergence)
## running simplex # 4...
## ...done
## ...simplex #4 complete, loglike = -104.446501 (convergence)
## running simplex # 5...
## ...done
## ...simplex #5 complete, loglike = -104.446501 (convergence)
## using L-BFGS method from simplex #1...
## iteration: 1,loglike = -104.446501
## ...L-BFGS method complete
## Maximum likelihood estimates found, log like = -104.446501
## creating gp object.....done
summary(GPFit)
## Total observations = 30
```

