# Pritchard2014\_Graph\_Generation

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### Introduction

This document describes the production of the Figure 1 graphs from the Pritchard and Birch (2014) opinion paper in *Mol. Plant. Pathol.* (doi:10.1111/mpp.12210). In that manuscript, a toy dynamic model of the plant immune system is presented (this is deposited in the BioModels database with accession MODEL1408280000: http://www.ebi.ac.uk/biomodels-main/). This document uses the output from five runs of the model using COPASI, representing 200s timecourses:

- no\_path.txt: The default model, with k1("Pathogen arrival")=0
- no\_pti\_no\_supp\_no\_eti.txt: Default model, with k1("PAMP recognition")=0, k1("Callose suppression")=0, and k1("Effector recognition")=0
- pti\_no\_supp\_no\_eti.txt: Default model, with k1("Callose suppression")=0, and k1("Effector recognition")=0
- pti\_supp\_no\_eti.txt: Default model, with k1("Effector recognition")=0
- pti\_supp\_eti.txt: Default model

# Load model output into dataframes

COPASI tab-separated output starts the header line with the # character, which is annoyingly also the comment line character in R, so we need to specify that no comment characters should be respected, when we load the dataframes. We initially store each run in its own dataframe, then bind the output into a single dataframe, with columns Time, variable, value, and runtype where Time is the model run timepoint, variable is either Callose or Pathogen (the two variables of interest), value is the dimensionless level of the variable, and runtype is a human-readable label for the run.

```
# We use ggplot2, and the handy reshape2 libraries
library(ggplot2)
library(reshape2)

# Load data
# These files are timecourse output from COPASI, but COPASI's header line
# starts with the default comment character, so we need to lose this.
# (possibly by manual editing)
```

```
nopathdata = read.table('no_path.txt', sep='\t',
                        header=T, comment.char="")
pathdata = read.table('no_pti_no_supp_no_eti.txt', sep='\t',
                      header=T, comment.char="")
ptidata = read.table('pti_no_supp_no_eti.txt', sep='\t',
                     header=T, comment.char="")
ptietsdata = read.table('pti_supp_no_eti.txt', sep='\t',
                        header=T, comment.char="")
ptietsetidata = read.table('pti_supp_eti.txt', sep='\t',
                           header=T, comment.char="")
# Reshape all five datasets and lose the columns we're not interested in
dflist = list(nopath=nopathdata, noresponse=pathdata,
              pti=ptidata, ptiets=ptietsdata,
              ptietseti=ptietsetidata)
dflist = lapply(dflist, function(df) {
  colnames(df)[1] = "Time"
  colnames(df)[7] = "Pathogen"
 df = melt(df, id=c("Time"))
 df = df[df$variable %in% c("Callose", "Pathogen"),]
})
# Bind all our data into a single dataframe
data = do.call(rbind, dflist)
# Convert row.names to run names
vals = 402 # How many rows per run?
data$runtype = c(rep("No Pathogen", vals), rep("No Response", vals),
                 rep("PTI", vals), rep("PTI+ETS", vals),
                 rep("PTI+ETS+ETI", vals))
```

### Plot steady-states

The code below renders a plot of steady-state values of Pathogen and Callose variables across the five run types. The output behaves as we might expect.

```
# We assume that the model reaches a steady-state (or near enough), by
# Time=200s, and just pull out those values
ssdata = data[data$Time==200,]
print(ssdata)
```

```
##
                  Time variable
                                   value
                                             runtype
## nopath.1005
                   200 Callose 0.000000 No Pathogen
## nopath.1206
                   200 Pathogen 0.000000 No Pathogen
## noresponse.1005 200 Callose 0.000000 No Response
## noresponse.1206 200 Pathogen 1.000000 No Response
## pti.1005
                   200 Callose 0.414207
                                                 PTI
## pti.1206
                   200 Pathogen 0.707112
                                                 PTI
## ptiets.1005
                   200 Callose 0.300568
                                             PTI+ETS
## ptiets.1206
                   200 Pathogen 0.768893
                                             PTI+ETS
## ptietseti.1005
                   200 Callose 0.280057 PTI+ETS+ETI
## ptietseti.1206
                   200 Pathogen 0.640030 PTI+ETS+ETI
```

When no pathogen is present, Pathogen and Callose (a proxy for overall PTI response) are both zero. If pathogen is introduced to a host that does not produce PTI or ETI (No Response), then Pathogen reaches its full arbitrary level of 1.

However, if the host exhibits PTI, and the pathogen does not produce effectors, the result is that Callose is produced (0.41), and the incidence of Pathogen falls (0.71). Against this background, if the pathogen is capable of producing effectors that result in callose supression, then steady-state Callose falls (0.30), and Pathogen rises (0.76).

Finally, for a model where the host exhibits both PTI and ETI, but the pathogen produces an effector that can suppress callose, both Callose and Pathogen fall to their lowest steady-state values (0.28 and 0.64, respectively).

### Plot time series

The code below renders a small multiple/facet plot for the four timecourses, where a pathogen is present. These are presented as plots of the values for Path (local microbial population) and Callose (abstracted callose deposition/PTI level), as a function of time. For these plots, we exclude the run with no pathogen present.

## Output

The two figures are plotted below.

```
library(gridExtra)
grid.arrange(p2, p1, ncol=1)
```



