

# **Comparative Genomics and Visualisation**

## **BS32010**



**The James  
Hutton  
Institute**

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# Table of Contents



# What Is Comparative Genomics?

**The combination of genomic data, and comparative and evolutionary biology, to address questions of genome structure, evolution, and function.**



# Evolution is the central concept



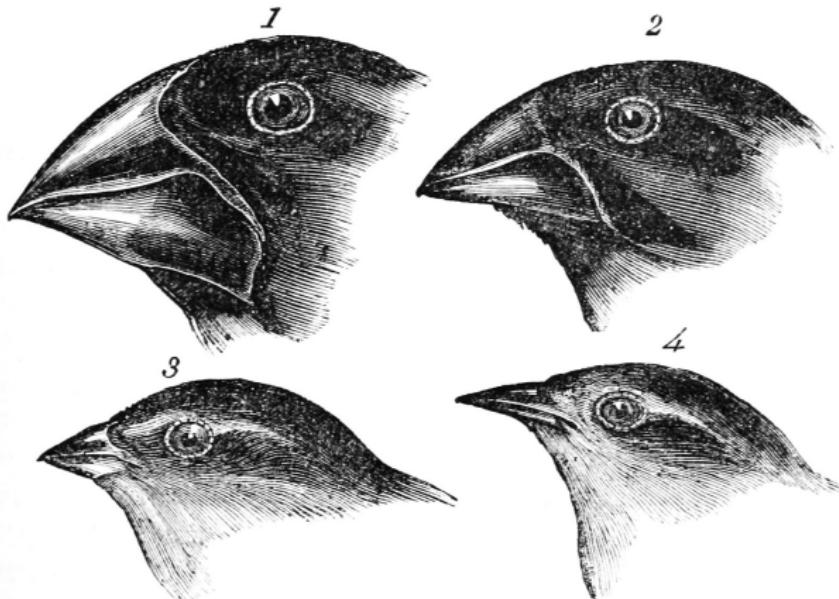
**“NOTHING IN BIOLOGY MAKES SENSE EXCEPT  
IN THE LIGHT OF EVOLUTION.”**

THEODOSIUS DOBZHANSKY



# Comparison of physical features

How do we determine that features share a common ancestor?



1. *Geospiza magnirostris*.  
3. *Geospiza parvula*.

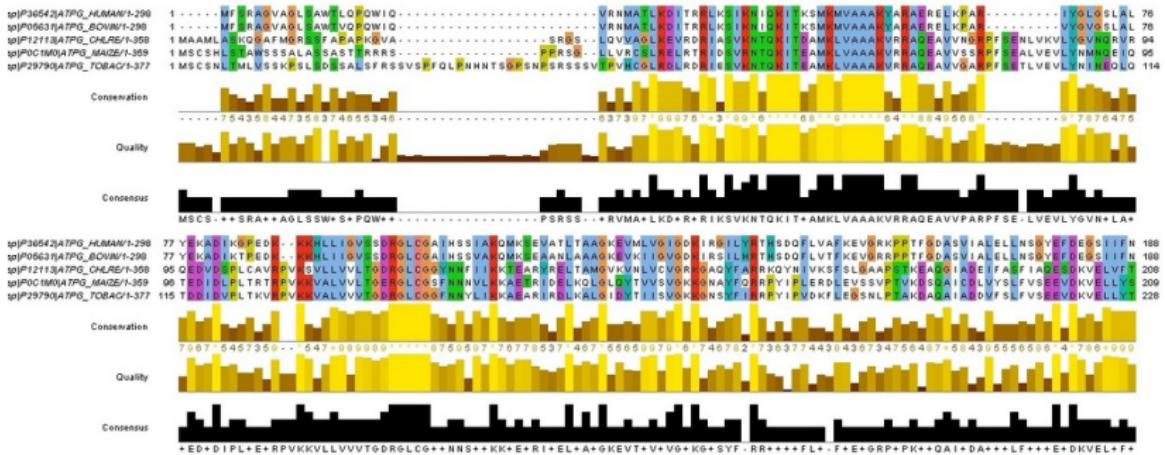
2. *Geospiza fortis*.  
4. *Certhidea olivacea*.



## Comparison of sequence features



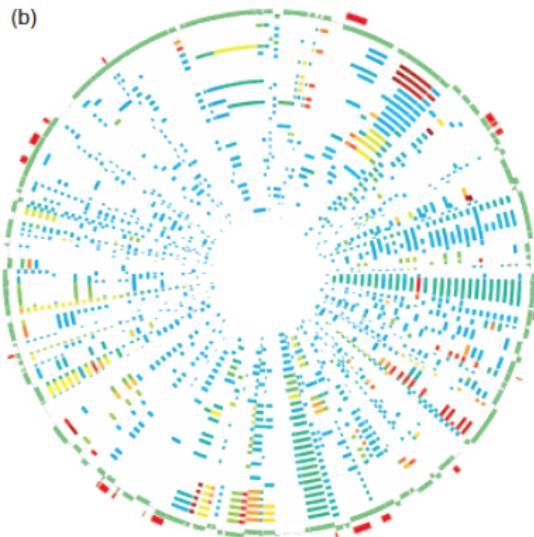
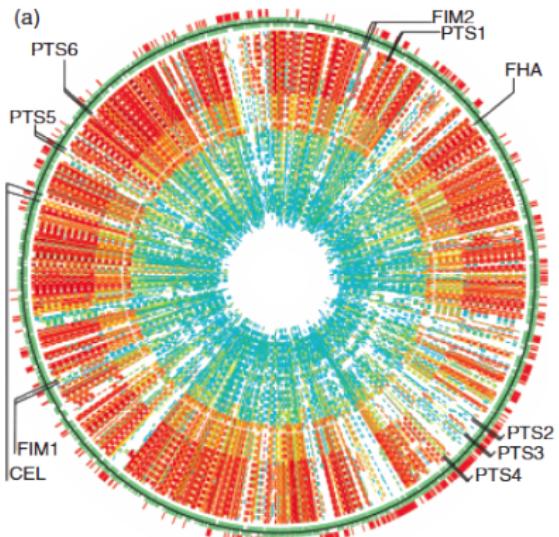
How do we determine that features share a common ancestor?  
Multiple sequence alignment of ATP synthase





# Comparison of genome features

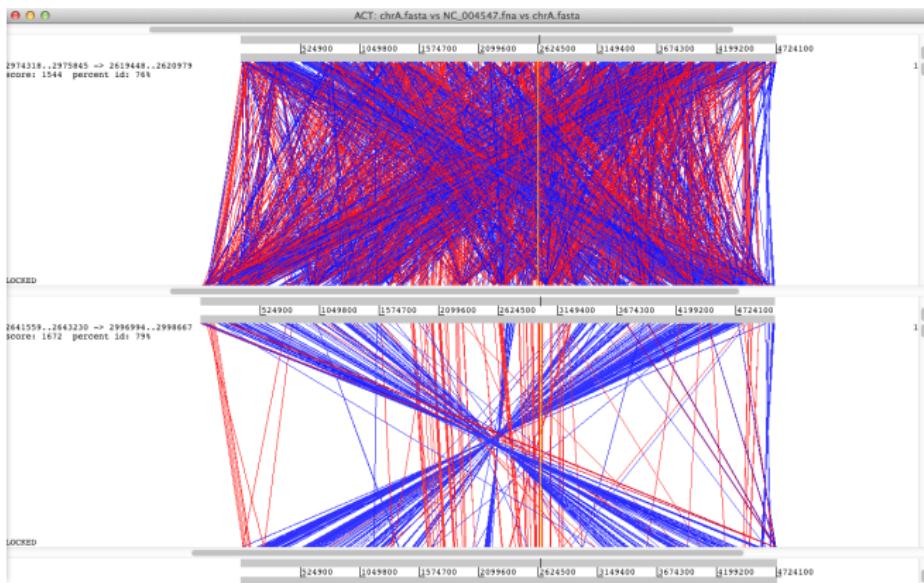
How do we determine that features share a common ancestor?  
Similarity of individual features (feature sequence)





# Comparison of genome features

How do we determine that features share a common ancestor?  
Similarity of individual features (ordering and arrangement)



# Why comparative genomics?

- Genome features are heritable characteristics
- Related organisms share ancestral genomes
- Related organisms inherit common genome features
- Genome similarity  $\propto$  relatedness? (phylogenomics)

I think



Then between A + B. *causes*  
less relation. C + B. the  
first generation, B + D  
rather greater distinction  
Then genome would be  
formed. - binary relation

# Why comparative genomics?

- Genomes carry functional elements under selection pressure
- Deleterious functional elements are lost through selection
- Organisms with similar phenotype carry similar functional elements
- Genome similarity  $\propto$  phenotype?  
(functional genomics)

I think



Then between A + B. *causes*  
less of relation. C + B. the  
first generation, B + D  
rather greater distinction  
Then genome would be  
formed. - binary relation



# Why comparative genomics?

- Functionally-optimised elements are conserved
- (Functional elements can be transferred non-heritably)
- Genome feature similarity  $\Rightarrow$  common function? (genome annotation)
- Transfer functional information from model systems (*E. coli*, *A. thaliana*, *D. melanogaster*) to non-model systems

I think



then between A & B. same  
box of relation. C & B. the  
first generation, B & D  
rather greater distinction  
then genome will be  
formed. - binary relation



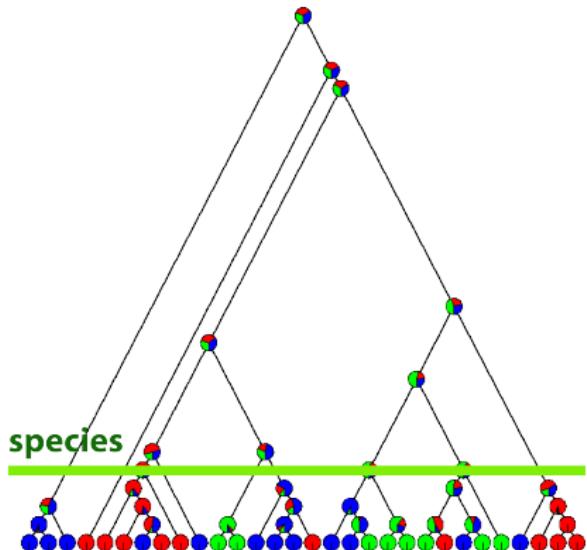
# Table of Contents



# Types of comparison

## Within species

- e.g. between isolates/individuals (or between tissues...)
- Which genome features may account for unique characteristics of organisms or cell-types (e.g. tumours)?
- what epigenetic changes occur in an individual?

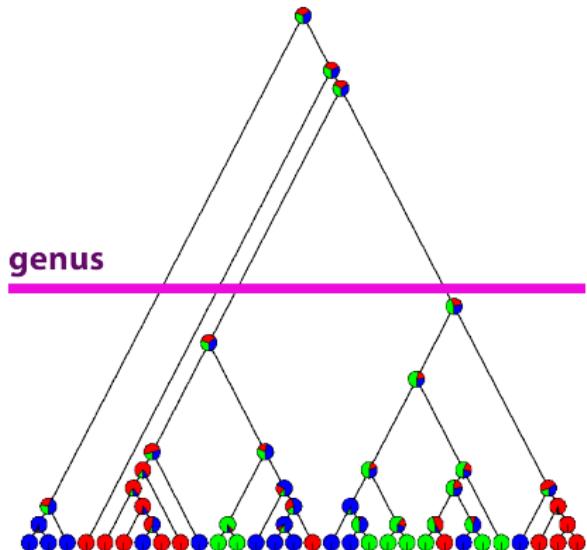




# Types of comparison

## Within genera/between species

- comparison between groups of individuals
- what genome features show evidence of selective pressure?
- which features/changes are associated with species phenotype/adaptation?



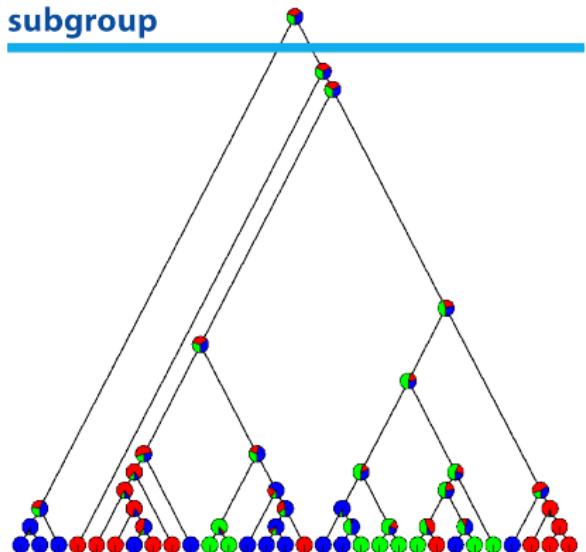


## Types of comparison



## Between subgroups

- e.g. comparisons across many diverse individuals
  - what are the *core set* of genome features that define a subgroup or genus?
  - what functions are present/absent between groups?





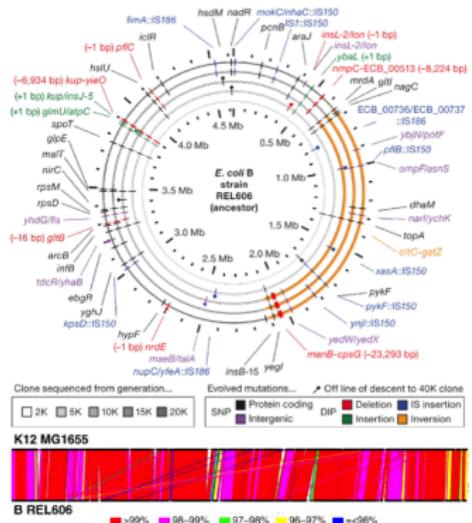
# **E. coli LTEE** *a b c*

<sup>a</sup> Jeong et al. (2009) *J. Mol. Biol.* doi:10.1016/j.jmb.2009.09.052

<sup>b</sup> Barrick et al. (2009) *Nature* doi:10.1038/nature08480

<sup>c</sup>Wiser et al. (2013) Science doi:10.1126/science.1243357

- Run by the Lenski lab, Michigan State University since 1988 (<http://myxo.css.msu.edu/ecoli/>)
  - 12 flasks, citrate usage selection
  - >50,000 generations of *E coli*!
    - Cultures propagated every day
    - Every 500 generations (75 days), mixed-population samples stored
    - Mean fitness estimated at 500 generation intervals





# Comparative genomics in the news <sup>a b c</sup>

<sup>a</sup> Bosi et al. (2015) Proc. Natl. Acad. Sci. USA 10.1073/pnas.1523199113

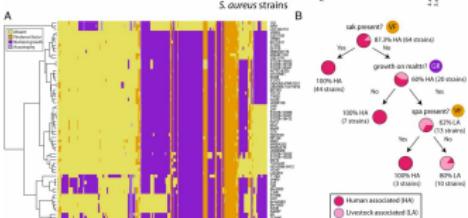
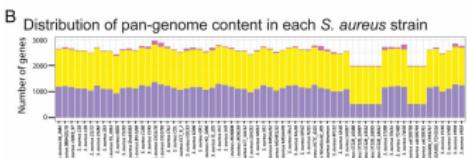
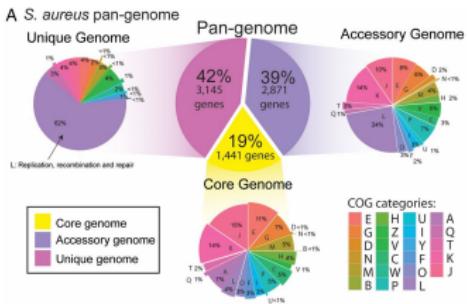
<sup>b</sup> World Health Organisation 27/2/2017

<sup>c</sup> Financial Times 27/2/2017

- Antibiotic resistance led to 40% rise in prescription of 'last-hope' colistin
- 12 bacterial priority pathogens identified by WHO
- Comparison of hundreds of *S. aureus* genomes produces models for metabolic capacity and insights into epidemiology



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# Levels of comparison

## Bulk Properties

- chromosome/plasmid counts and sizes, nucleotide content, etc.

## Whole Genome Sequence

- sequence similarity
- organisation of genomic regions (synteny), etc.

## Genome Features/Functional Components

- numbers and types of features (genes, ncRNA, regulatory elements, etc.)
- organisation of features (synteny, operons, regulons, etc.)
- complements of features
- selection pressure, etc.



# Table of Contents



## Bulk property comparisons

You don't have to sequence genomes to  
compare them  
(but it helps)



# Genome comparisons predate NGS

- Sequence data wasn't always cheap and abundant
- Practical, experimental genome comparisons were needed





## Bulk property comparisons

**Calculate values for individual genomes,  
then compare them.**

- Number of chromosomes
- Ploidy
- Chromosome size
- Nucleotide (A,C,G,T) frequency



# Nucleotide frequency/genome size

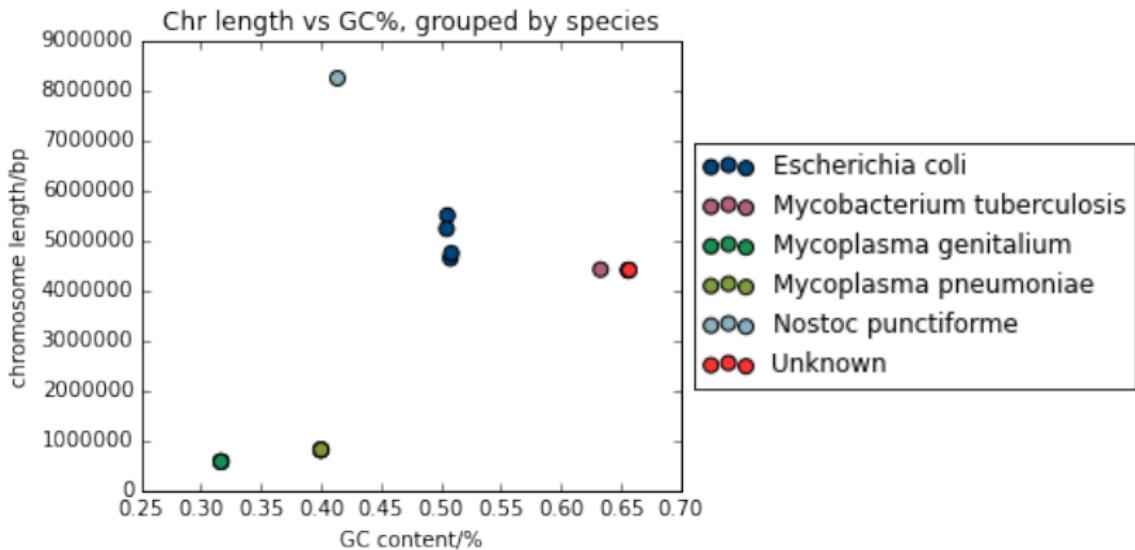
Very easy to calculate from complete or draft genome sequence

```
In [1]: from Bio import SeqIO
In [2]: s = SeqIO.read("data/NC_000912.fna", "fasta")
In [3]: a, c, g, t = s.seq.count("A"), s.seq.count("C"), s.seq.count("G"), s.seq.count("T")
In [4]: float(g + c)/len(s)
Out[4]: 0.40008010837904245
In [5]: float(g - c)/(g+c)
Out[5]: 0.002397259225467894
```

GC content, chromosome size can be characteristic of an organism.



# Genome Size and GC%





# Blobology <sup>a b</sup>

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<sup>a</sup>Kumar & Blaxter (2011) *Symbiosis* doi:10.1007/s13199-012-0154-6

<sup>b</sup><http://nematodes.org/bioinformatics/blobology/>



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Sequence data can be contaminated by other organisms

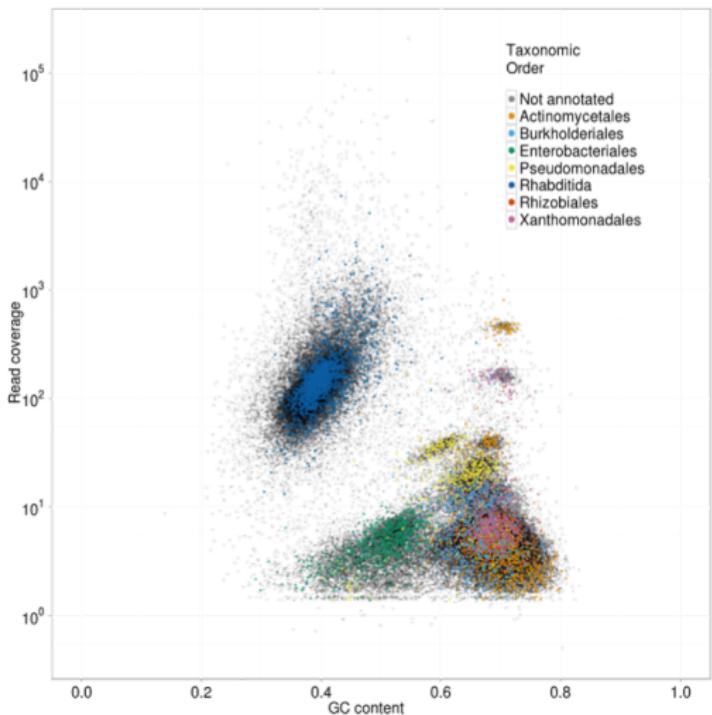
- Host and symbiont DNA have different %GC
- Host and symbiont DNA differ in coverage
- Assemble genome
- Map reads
- Plot coverage against %GC



# Blobology <sup>a b</sup>

<sup>a</sup> Kumar & Blaxter (2011) *Symbiosis* doi:10.1007/s13199-012-0154-6

<sup>b</sup> <http://nematodes.org/bioinformatics/blobology/>





# Nucleotide $k$ -mers

Sequence data is necessary to determine  $k$ -mers/frequencies  
*Not possible by experiment*

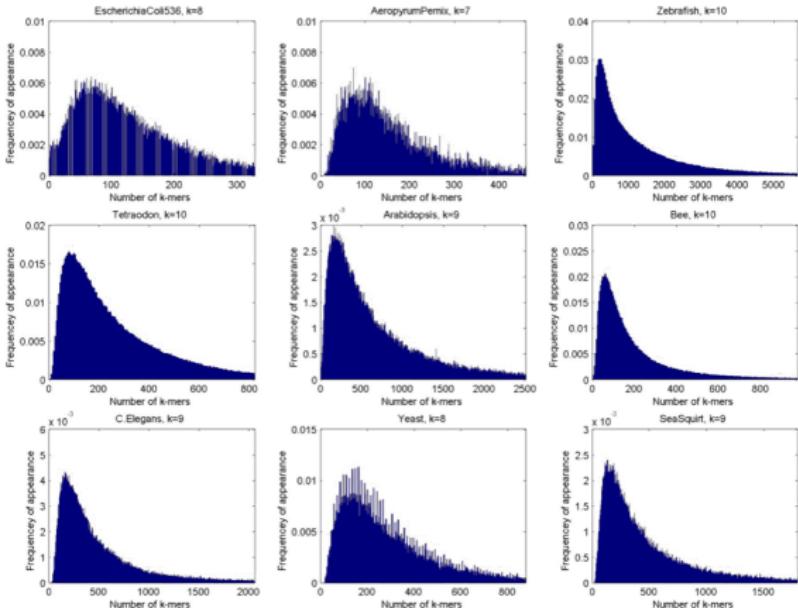
- Nucleotides,  $k = 1$ ,  $4 \times 1$ -mers  
A, C, G, T
- Dinucleotides,  $k = 2$ ,  $16 \times 2$ -mers  
AA, AC, AG, AT, CA, CC, CG, CT, GA, GC, GG, GT, TA, TC, TG, TT
- Triucleotides,  $k = 3$ ,  $64 \times 3$ -mers
- $k$ -nucleotides,  $4^k \times k$ -mers



# *k*-mer spectra <sup>a</sup>

<sup>a</sup>Chor et al. (2009) *Genome Biol.* doi:10.1186/gb-2009-10-10-r108

*k*-mer spectrum: frequency distribution of observed *k*-mer counts.  
Most species have a unimodal *k*-mer spectrum ( $k \approx 9$ )



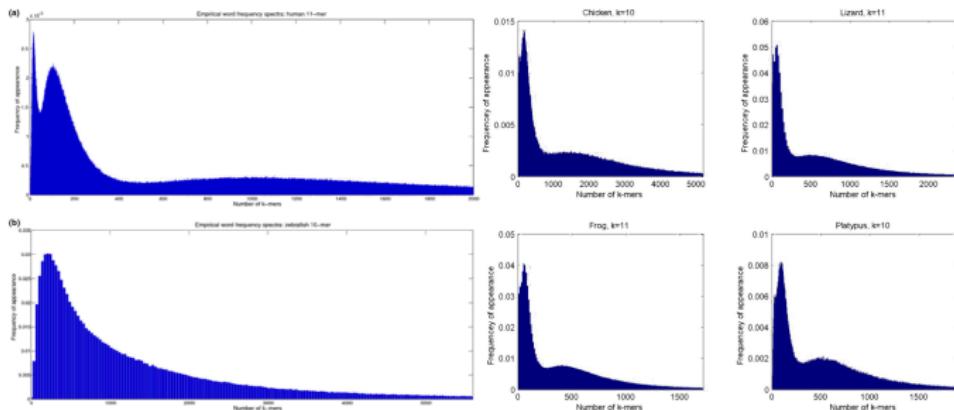


# *k*-mer spectra <sup>a</sup>

<sup>a</sup>Chor et al. (2009) *Genome Biol.* doi:10.1186/gb-2009-10-10-r108

All mammals tested (and some other species) have *multimodal k-mer spectra*

Genomic regions also differ in this property





# Table of Contents



## Whole genome comparisons

**Comparisons of one whole or draft genome  
with another  
(...or many others)**



# Whole genome comparisons

Minimum requirement: **two genomes**

- Reference Genome
- Comparator Genome

The experiment produces a comparative result *that is dependent on the choice of genomes.*



# Whole genome comparisons

Experimental methods mostly involve direct or indirect DNA hybridisation

- DNA-DNA hybridisation (DDH)
- Comparative Genomic Hybridisation (CGH)
- Array Comparative Genomic Hybridisation (aCGH)



# Whole genome comparisons

Analogously, *in silico* methods mostly involve sequence alignment

- Average Nucleotide Identity (ANI)
- Pairwise genome alignment
- Multiple genome alignment

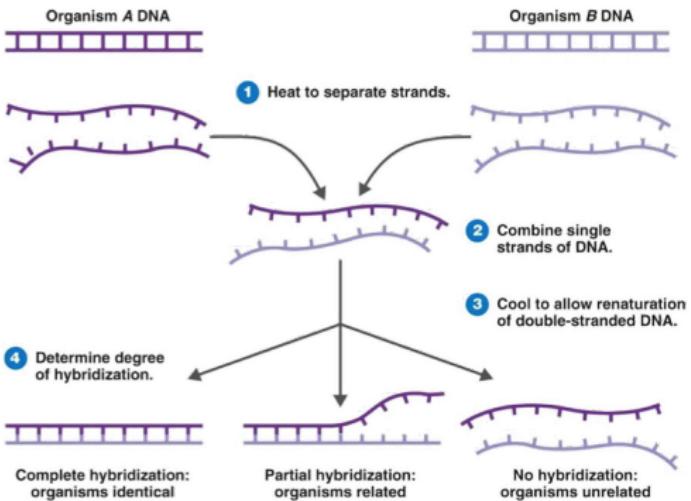


# DNA-DNA hybridisation (DDH) <sup>a</sup>

<sup>a</sup> Morelló-Mora & Amann (2011) *FEMS Microbiol. Rev.* doi:10.1016/S0168-6445(00)00040-1

Several similar methods based on the same principle

- Denature gDNA mixture for organisms *A, B*
- Allow gDNA to anneal; hybrids result



Reassociation of gDNA  $\approx$  sequence similarity

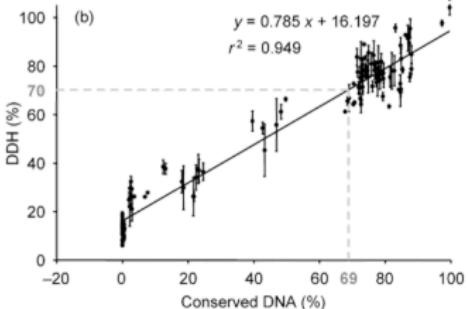
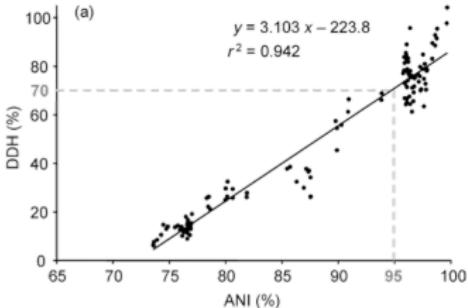


# Average Nucleotide Identity (ANI) <sup>a</sup>

<sup>a</sup> Goris et al. (2007) *Int. J. System. Evol. Biol.* doi:10.1099/ijss.0.64483-0

Introduced as an *in silico* substitute for DDH in 2007:

- 70% identity (DDH) = "gold standard" prokaryotic species boundary
- 70% identity (DDH)  $\approx$  95% identity (ANI)

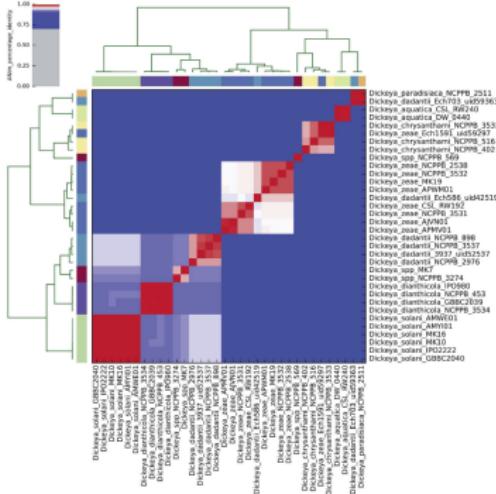


## ANI in practice <sup>a b</sup>

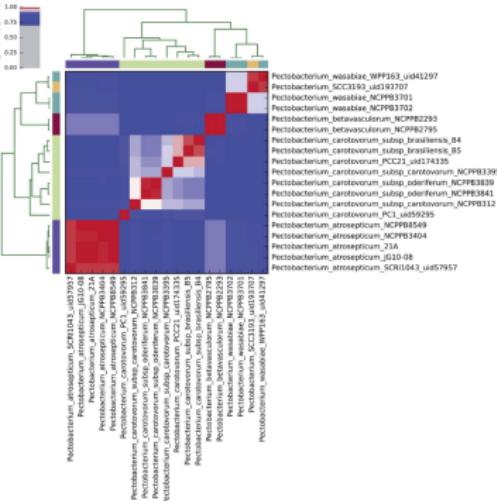
<sup>a</sup>van der Wolf et al. (2014) *Int. J. Syst. Evol. Micr.* **64**:768-774 doi:10.1099/ijss.0.052944-0

<sup>b</sup>Pritchard et al. (2016) *Anal. Methods* **8**:12-24 doi:10.1039/C5AY02550H

## *Dickeya* species structure



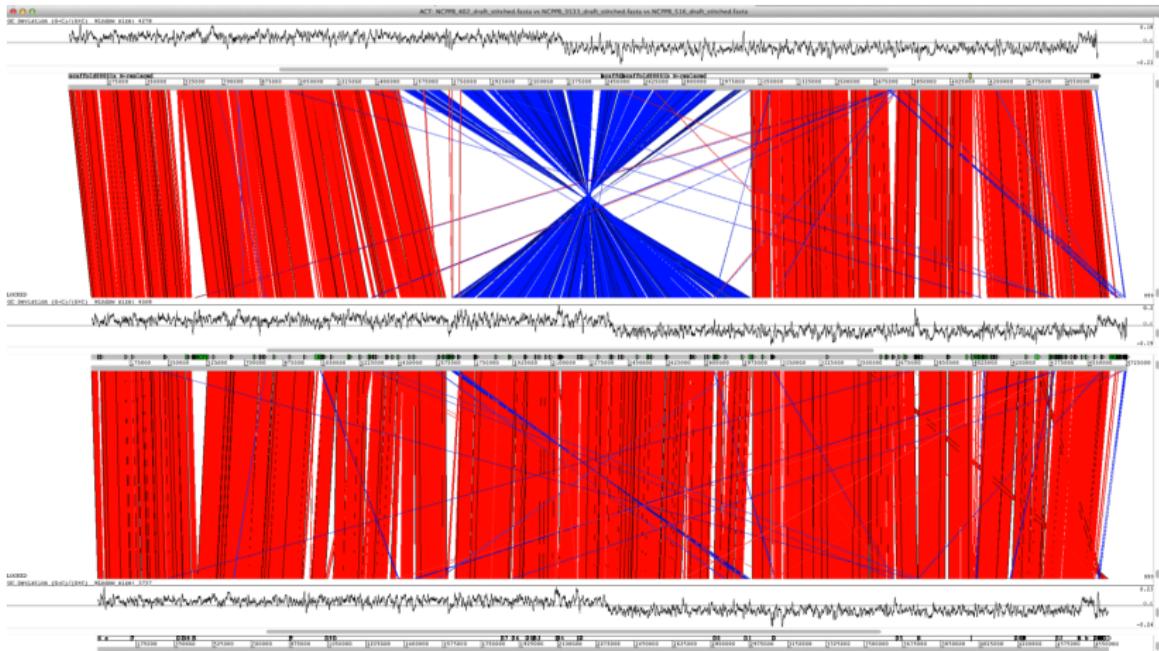
## *Pectobacterium* species structure:





# Pairwise genome alignments

Pairwise comparisons require alignment of similar regions.

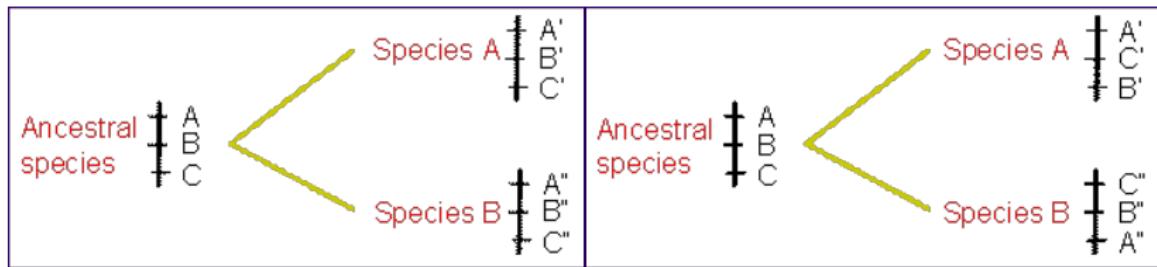




# Synteny and Collinearity

Genome rearrangements may occur post-divergence

Sequence similarity, and order of similar regions, may be conserved



- *collinear* conserved elements lie in the same linear sequence
- *syntenous* (or *syntenic*) elements:
  - (orig.) lie on the same chromosome
  - (mod.) are collinear

Evolutionary constraint (e.g. indicated by synteny) may indicate a functional constraint (and help determine *orthology*)



# Alignment algorithms/programs

I assume you're familiar with BLAST...

BLASTN and naïve alignment algorithms are not appropriate for whole-genome alignment:

- Needleman-Wunsch: optimal global alignment
- Smith-Waterman: optimal local alignment

Cannot handle rearrangement

Computationally expensive



# Alignment algorithms/programs

Many whole-genome alignment algorithms proposed  
Handle genome-scale evolutionary processes, scalable

- LASTZ (<http://www.bx.psu.edu/~rsharris/lastz/>)
- **BLAT** (<http://genome.ucsc.edu/goldenPath>)
- Mugsy (<http://mugsy.sourceforge.net/>)
- **megaBLAST** (<http://www.ncbi.nlm.nih.gov/blast/>)
- **MUMmer** (<http://mummer.sourceforge.net/>)
- LAGAN ([http://lagan.stanford.edu/lagan\\_web/index.shtml](http://lagan.stanford.edu/lagan_web/index.shtml))
- WABA, etc?



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<sup>a</sup>Zhang *et al.* (2000) *J. Comp. Biol.* **7**(1-2): 203-214

<sup>b</sup>Korf *et al.* (2003) *BLAST* O'Reilly & Associates, Sebastopol, CA

Optimised for:

- speed and genome-level searching
- queries on large sequence sets: "query-packing"
- long alignments of very similar sequences (dc-megablast for divergent sequences)

Uses Zhang et al. greedy algorithm, **not BLAST algorithm**

BLASTN+ defaults to megaBLAST algorithm  
(see <http://www.ncbi.nlm.nih.gov/blast/Why.shtml>)

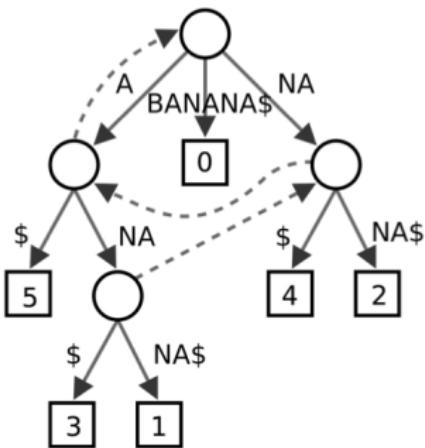


Conceptually completely different to BLAST/BLAT/megaBLAST  
Uses *suffix trees* for pattern matching

- Finds maximal exact matches
- Memory use depends only on reference sequence size

Suffix Tree:

- Constructed and searched in  $O(n)$  time
- Useful algorithms are nontrivial
- BANANA\$





# Pairwise genome alignments

Which genomes should you align (or not bother with)?

For reasonable analysis, genomes should:

- derive from a sufficiently **recent** common ancestor, so that homologous regions can be identified
- derive from a sufficiently **distant** common ancestor, so that biologically meaningful changes are likely to be found



# Vibrio mimicus <sup>a</sup>

<sup>a</sup> Hasan et al. (2010) Proc. Natl. Acad. Sci. USA 107:21134-21139 doi:10.1073/pnas.1013825107

Chromosome C-II carries genes associated with environmental adaptation; C-I carries virulence genes.  
C-II has undergone extensive rearrangement; C-I has not.

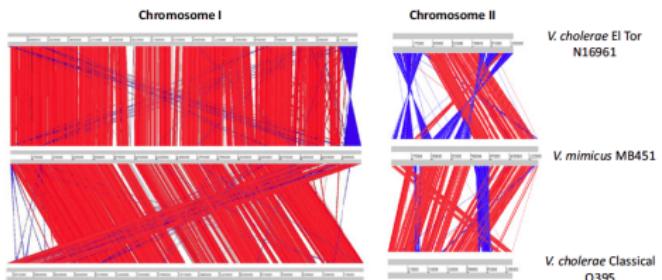


Fig. 2. Linear pairwise comparison of the *Vibrio mimicus* genome by Artemis Comparison Toll. Regions with similarity are highlighted by connecting red or blue lines between the genomes; red lines indicate homologous blocks of sequence, and blue lines indicate inversions. Gaps indicate unique DNA. The gray bars represent forward and reverse strands.

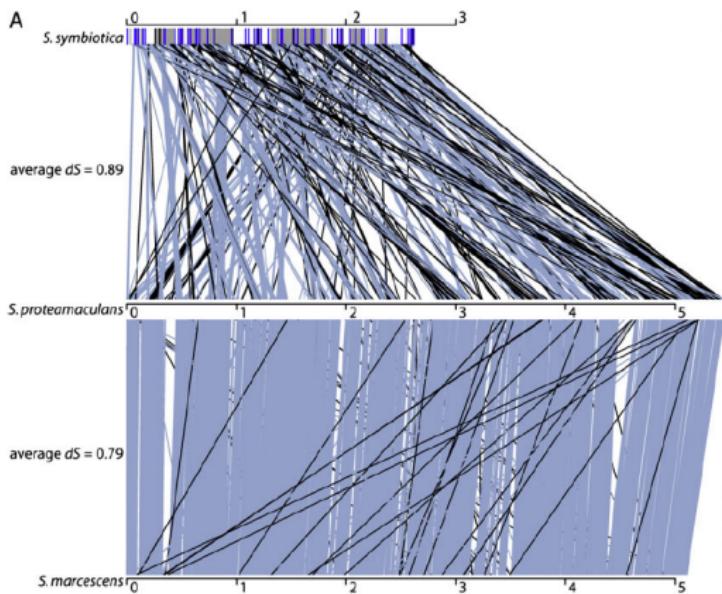
Suggests modularity of genome organisation, as a mechanism for adaptation (HGT, two-speed genome).



# Serratia symbiotica <sup>a</sup>

<sup>a</sup> Burke and Moran (2011) *Genome Biol. Evol.* 3:195-208 doi:10.1093/gbe/evr002

*S. symbiotica* is a recently evolved symbiont of aphids  
Massive genomic decay is an adaptation to the new environment.





# Multiple genome alignments

Multiple genome alignments are “harder” than pairwise

- Computationally difficult to produce
- Lead to NP-complete optimisation problems!

Solutions: **heuristics**

- Progressive (build a tree, combine pairwise alignments)
- Iterative (realign initial sequences as new genomes added)
- Positional homology
- *Glocal* alignments



# Multiple genome alignment

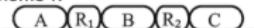
Many tools use either positional homology or glocal alignment

Several tools available:

- **Mugsy:** (<http://mugsy.sourceforge.net/>)
- **MLAGAN:**  
([http://lagan.stanford.edu/lagan\\_web/index.shtml](http://lagan.stanford.edu/lagan_web/index.shtml))
- **TBA/MultiZ:**  
([http://www.bx.psu.edu/miller\\_lab/](http://www.bx.psu.edu/miller_lab/))
- **Mauve:**  
(<http://gel.ahabs.wisc.edu/mauve/>)

Given a set of genomes:

Genome 1:



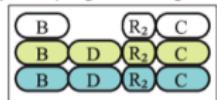
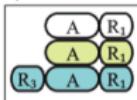
Genome 2:



Genome 3:



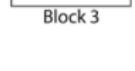
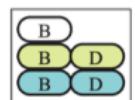
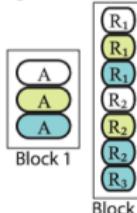
Ideal *positional homology* multiple genome alignment:



Block 1

Block 2

Ideal *glocal* multiple genome alignment:



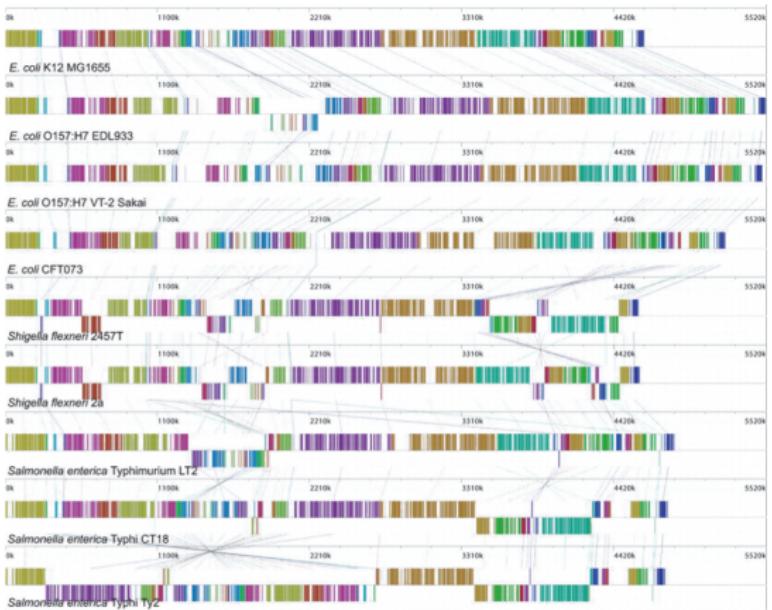
Block 2

Block 3

Block 4



## MAUVE alignment of nine enterobacterial genomes Evidence for rearrangement of homologous backbone sequence



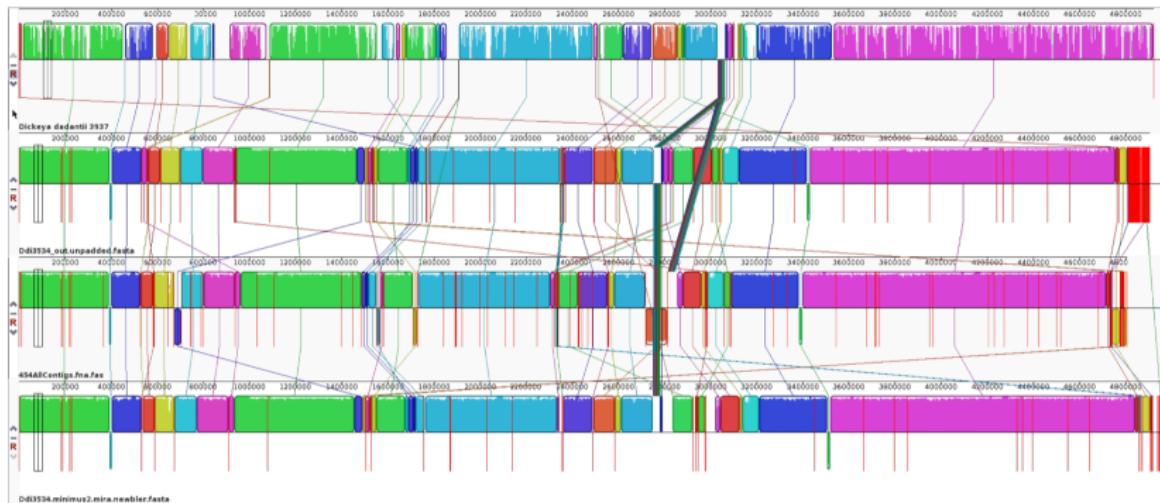


# Draft genome alignment

High-throughput genome assemblies are often fragmented (contigs)

Contigs can be ordered (*scaffolded*):

- **without alignment**, by long or paired-end reads
- **by alignment**, to complete *reference* genomes or to other draft incomplete genomes





# Chromosome painting<sup>a</sup>

<sup>a</sup>Yahara et al. (2013) *Mol. Biol. Evol.* 30:1454–1464 doi:10.1093/molbev/mst055

“Chromosome painting” infers recombination-derived ‘chunks’  
Genome’s haplotype constructed in terms of recombination events  
from a ‘donor’ to a ‘recipient’ genome



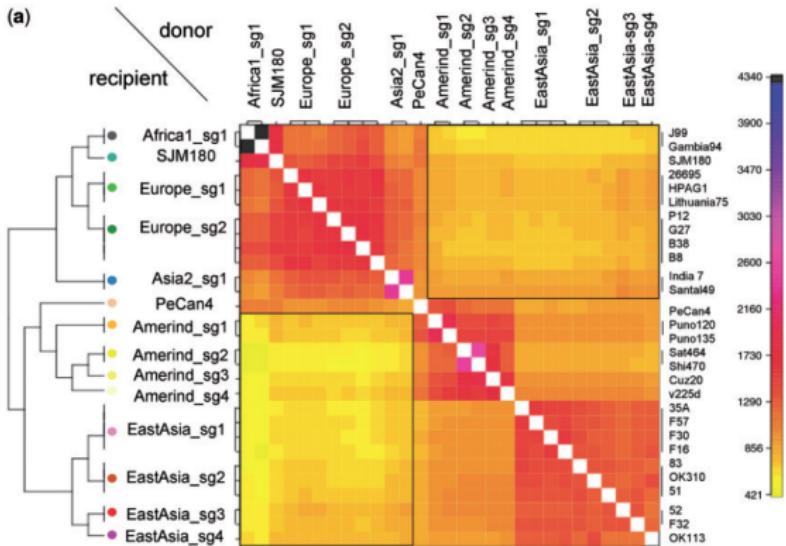
FIG. 1. Chromosome painting *in silico*. Each lane indicates the chromosome of a strain shown on the right. The strains are classified by fineSTRUCTURE into subgroups labeled by colors (table 1 and fig. 2) on the left. A color along the chromosome indicates the subgroup that donated a chunk of SNPs through homologous recombination. All genomic positions are transformed to those of a reference strain (26695).



# Chromosome painting<sup>a</sup>

<sup>a</sup>Yahara et al. (2013) Mol. Biol. Evol. 30:1454-1464 doi:10.1093/molbev/mst055

Recombination events summarised in a *coancestry matrix*.  
*H. pylori*: most within geographical bounds, but asymmetrical donation from Amerind/East Asian to European isolates.





# Whole Genome Comparisons

## Physical and computational genome comparisons

- Similar biological questions
- ∴ similar concepts

## Modern biology: lots of sequence data

- Conservation ≈ evolutionary constraint
- Many choices of algorithms/software
- Many choices of visualisation tools/software



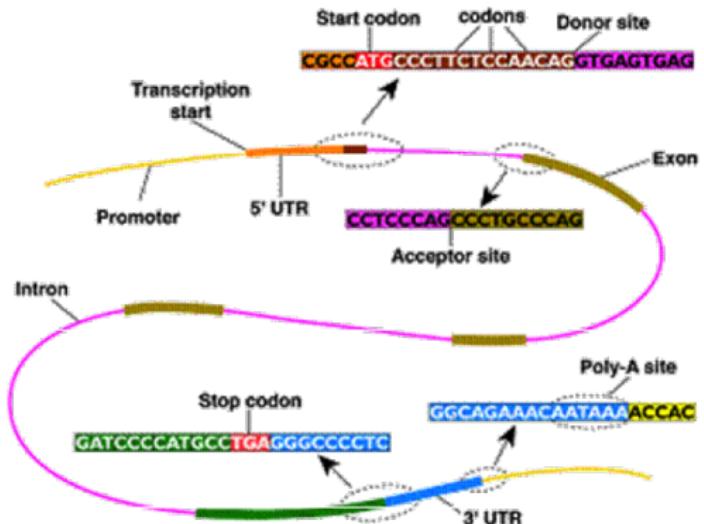
# Table of Contents



# Gene features

Significant substructure, especially in eukaryotes

- translation start
- introns
- exons
- translation stop
- translation terminator

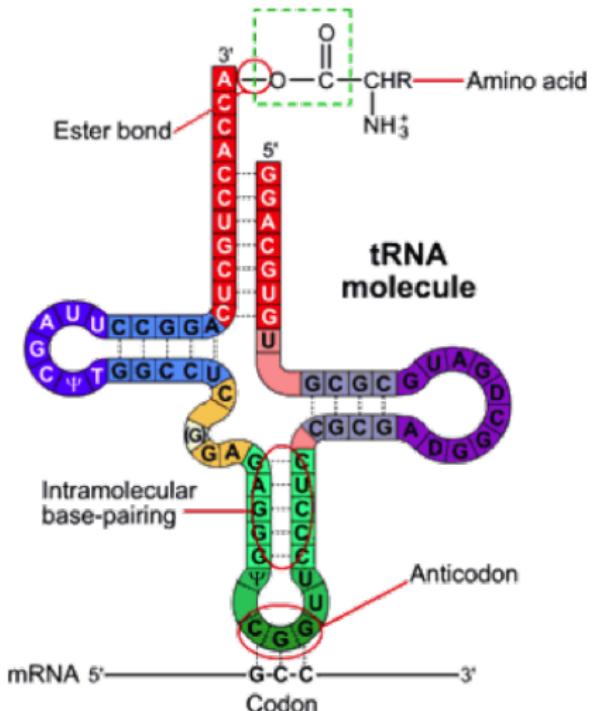




# RNA features

RNA/ncRNA: characterised by complex secondary structure

- tRNA - transfer RNA
- rRNA - ribosomal RNA
- CRISPRs - prokaryotic defence, and genome editing
- many other functional classes, including enhancers

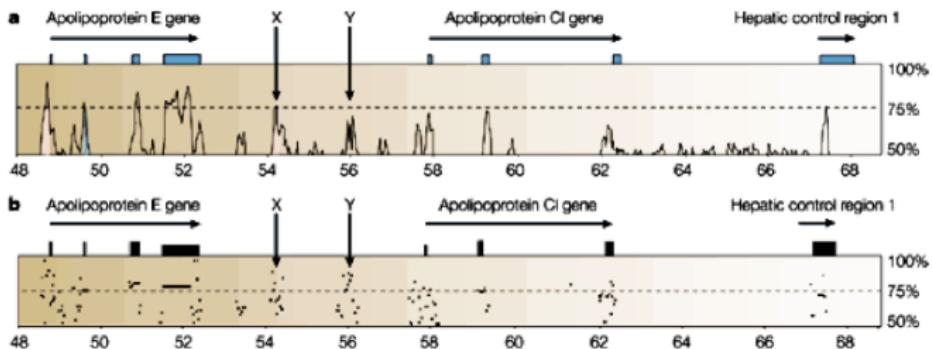




# Regulatory features <sup>a</sup>

<sup>a</sup>Pennacchio & Rubin (2001) *Nature Rev. Genet.* doi:10.1038/35052548

- transcription start sites (TSS)
- RNA polymerase (RNAP) binding sites
- transcription factor binding sites (TFBS)
- core, proximal and distal promoter regions





# Gene finding <sup>a b c</sup>

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<sup>a</sup> Liang et al. (2009) *Genome Res.* doi:10.1101/gr.088997.108

<sup>b</sup> Brent (2007) *Nat. Biotech.* doi:10.1038/nbt0807-883

<sup>c</sup> Korf (2004) *BMC Bioinf.* doi:10.1186/1471-2105-5-59

At genome scales, we need to automate functional prediction

Empirical (evidence-based) methods:

- Inference from known protein/cDNA/mRNA/EST sequence
- Interference from mapped RNA reads (e.g. RNAseq)

*Ab initio* methods:

- Prediction on the basis of gene features (TSS, CpG islands, Shine-Dalgarno sequence, stop codons, nucleotide composition, etc.)

Inference from genome comparisons/sequence conservation



# Regulatory element finding <sup>a b c</sup>

<sup>a</sup>Zhang *et al.* (2011) *BMC Bioinf.* doi:10.1186/1471-2105-12-238

<sup>b</sup>Kilic *et al.* (2013) *Nucl. Acids Re.* doi:10.1093/nar/gkt1123

<sup>c</sup>Vavouris & Elgar (2005) *Curr. Op. Genet. Deve.* doi:10.1016/j.gde.2005.05.002

## Empirical (evidence-based) methods:

- Inference from protein-DNA binding experiments
- Interference from co-expression

## *Ab initio* methods:

- Identification of regulatory motifs (profile/other methods; TATA,  $\sigma$ -factor binding sites, etc.)
- Statistical overrepresentation of motifs
- Identification from sequence properties

## Inference from genome comparisons/sequence conservation



# Equivalent genome features



When comparing two features (e.g. genes) between two or more genomes, there must be some basis for making the comparison.  
They have to be *equivalent* in some way, such as:

- common evolutionary origin
- functional similarity
- a family-based relationship

It's common to define equivalence of genome features in terms of evolutionary relationship.



# Why look at equivalent features?



## The real power of genomics is comparative genomics!

- Makes catalogues of genome components comparable between organisms
- Differences, e.g. presence/absence of equivalents may support hypotheses for functional or phenotypic difference
- Can identify characteristic signals for diagnosis/epidemiology
- Can build parts lists and wiring diagrams for systems and synthetic biology



# Who let the -logues out?

Genome features can have complex evolutionary relationships

We have precise terms to describe these relationships





# The -logues drop <sup>a</sup>

<sup>a</sup>Fitch et al. (1970) *Syst. Zool.* doi:10.2307/2412448



How do we understand the relationships between features in more than one genome?

- Functional similarity: **analogy**
- Evolutionary common origin: **homology, orthology, etc.**
- Evolutionary/functional/family relationship: **paralogy**

## DISTINGUISHING HOMOLOGOUS FROM ANALOGOUS PROTEINS

WALTER M. FITCH

### *Abstract*

*Fitch, W. M. (Dept. Physiological Chem., U. Wisconsin, Madison 53706) 1970. Distinguishing homologous from analogous proteins. Syst. Zool., 19:99–113.—This work provides a means by which it is possible to determine whether two groups of related proteins have a common ancestor or are of independent origin. A set of 16 random*

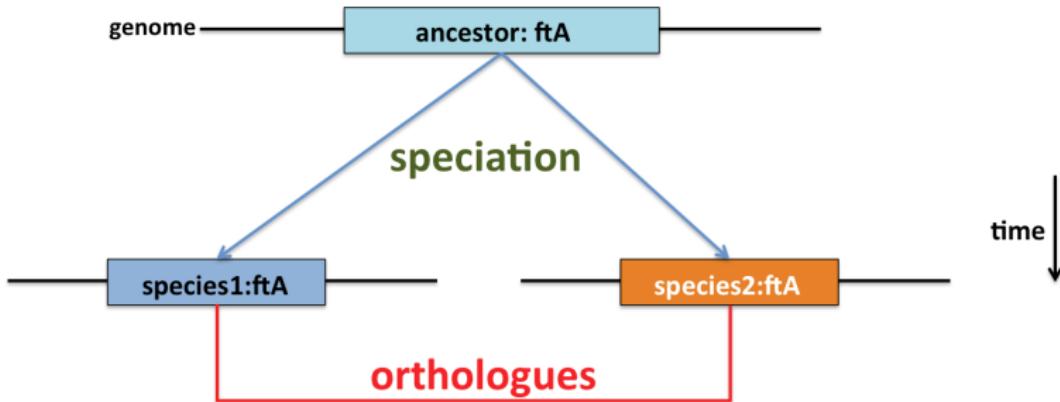


# Who let the -logues out?





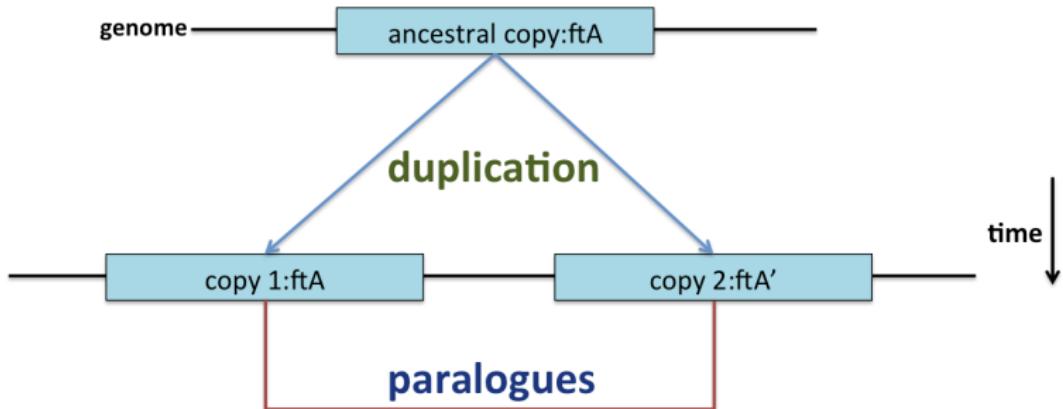
# Who let the -logues out?



- **Orthologues:** homologues that diverged through speciation



# Who let the -logues out?



**Paralogues:** homologues that diverged through duplication within the same genome



# Orthology <sup>a</sup>

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<sup>a</sup>Storm & Sonnhammer (2002) *Bioinformatics* doi:10.1093/bioinformatics/18.1.92

- Frequently abused/misused as a term
- “Orthology” is an evolutionary relationship, bent into service as a functional descriptor
- Orthology is strictly defined *only for two species or clades!* (cf. OrthoMCL)
- Orthology is not transitive:  
( $A$  is an orthologue of  $C$ , and  $B$  is an orthologue of  $C$ , does **not** imply that  $A$  is an orthologue of  $B$ )

**All classifications of orthology/paralogy are inferences!**



# The Ortholog Conjecture <sup>a</sup> <sup>b</sup>

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<sup>a</sup> Nehrt et al. (2011) *PLoS Comp. Biol.* doi:10.1371/journal.pcbi.1002073

<sup>b</sup> Chen et al. (2012) *PLoS Comp. Biol.* doi:10.1371/journal.pcbi.1002784



**Without duplication, a gene product is unlikely to change its basic function, because this would lead to loss of the original function, and this would be harmful.**



# Why focus on orthologues? <sup>a b c</sup>

<sup>a</sup>Chen and Zhang (2012) *PLoS Comp. Biol.* doi:10.1371/journal.pcbi.1002784

<sup>b</sup>Dessimoz (2011) *Brief. Bioinf.* doi:10.1093/bib/bbr057

<sup>c</sup>Altenhoff and Dessimoz (2009) *PLoS Comp. Biol.* 5:e1000262 doi:10.1371/journal.pcbi.1000262

Formalisation of the idea of *corresponding genes* in different organisms.

Orthologues serve two purposes:

- **Evolutionary equivalence**
- **Functional equivalence** ("The Ortholog Conjecture")

Applications in comparative genomics, functional genomics and phylogenetics.

Over 30 databases attempt to describe orthologous relationships ([http://questfororthologs.org/orthology\\_databases](http://questfororthologs.org/orthology_databases))



# Finding orthologues <sup>a b c</sup>

<sup>a</sup> Kristensen *et al.* (2011) *Brief. Bioinf.* **12**:379-391 doi:10.1093/bib/bbr030

<sup>b</sup> Trachana *et al.* (2011) *Bioessays* **33**:769-780 doi:10.1002/bies.201100062

<sup>c</sup> Salichos and Rokas (2011) *PLoS One* **6**:e18755 doi:10.1371/journal.pone.0018755.g006

## Multiple methods and databases

- **Pairwise genome**

- RBBH (aka BBH, RBH),  
RSD, InParanoid, RoundUp

- **Multi-genome**

- *Graph-based*: COG, eggNOG,  
OrthoDB, OrthoMCL, OMA,  
MultiParanoid
- *Tree-based*: TreeFam,  
Ensembl Compara,  
PhylomeDB, LOFT

### List of orthology databases

*If you know of any other database, please edit this page directly or contact us.*

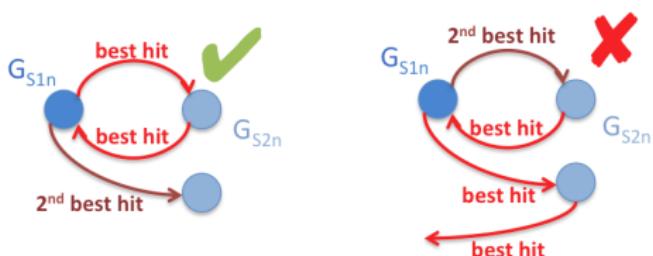
1. COG/WEGeKOGs
2. COGe-COCO-CL
3. COGe-LOFT
4. eggNOG
5. EGO
6. Ensembl Compara
7. Gene-Oriented Ortholog Database
8. GreenPhyDB
9. HCDP
10. Hemisite
11. HOGENOM
12. HOVERGEN
13. HOMOLENS
14. HOPS
15. INVHOGEN
16. JnParanoid
17. KEGG Orthology
18. MetaphOrn
19. MiGD
20. MiGD
21. OMA
22. OrthoDB (OrthoDB on Wikipedia)
23. OrthoLogID
24. ORTHOLUGE
25. OrthoMepster
26. OrthoMCL
27. Panther
28. PhOG
29. PHOG
30. PhylomeDB
31. PLAZA
32. P-POD
33. ProMMap
34. Proteinortho
35. RoundUp
36. TreeFam
37. YOBY



# Reciprocal Best BLAST Hits <sup>a</sup>

<sup>a</sup>On Reciprocal Best BLAST Hits 19/7/2012

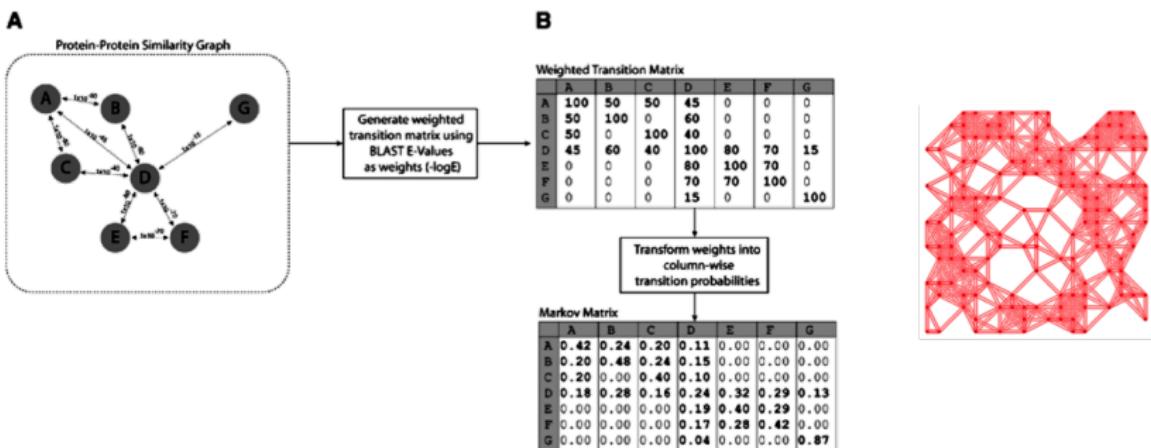
- $S_1, S_2$  are the gene sequence sets from two organisms
- Use sequence search tool (BLAST/FASTA):
  - Query= $S_1$ , Subject= $S_2$
  - Query= $S_2$ , Subject= $S_1$

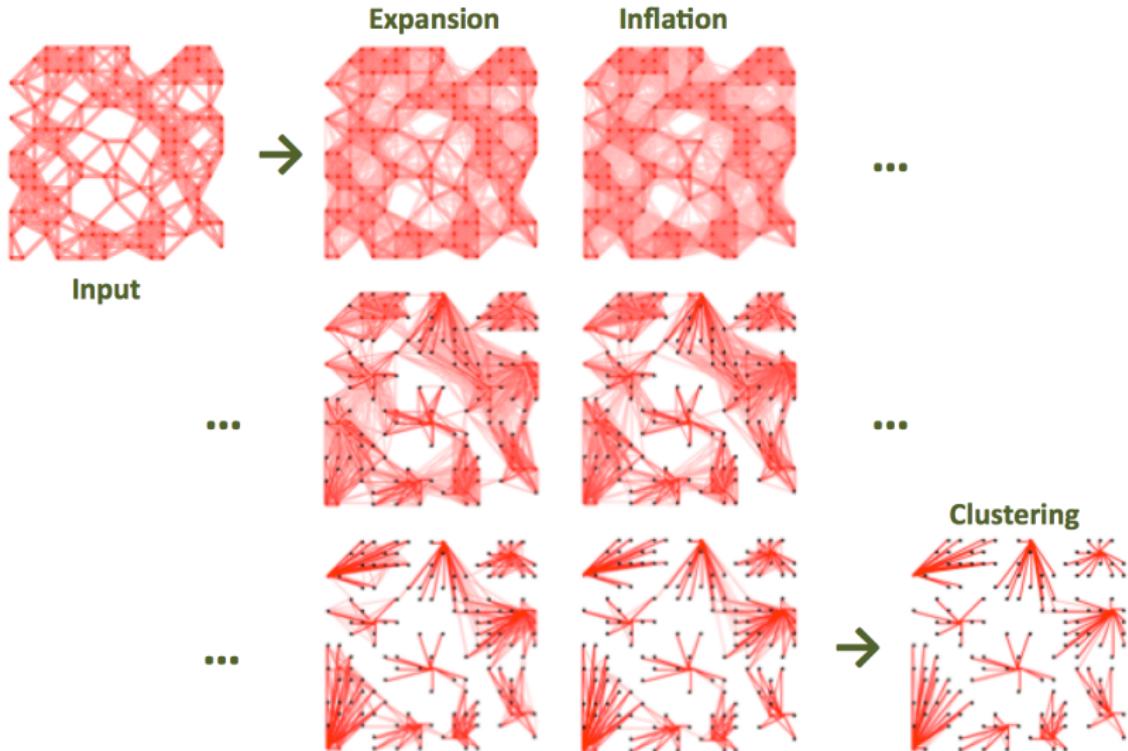


- Optionally filter hits (e.g. on %identity and %coverage)
- Find all pairs of sequences  $\{G_{S1n}, G_{S2n}\}$  in  $S_1, S_2$  where  $G_{S1n}$  is the best BLAST match to  $G_{S2n}$  and  $G_{S2n}$  is the best BLAST match to  $G_{S1n}$ .



- MCL constructs a network (*graph*) from all-against-all BLAST results
- Matrix operations (*expansion, inflation*) are applied
- Expansion, inflation iterated until the network converges







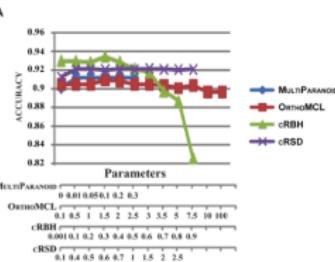
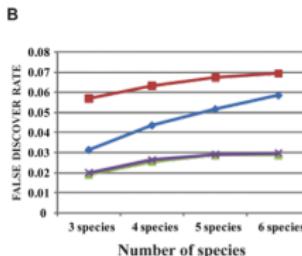
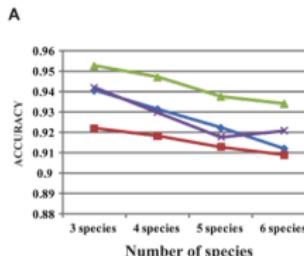
# Which prediction methods work best? <sup>a</sup>

<sup>a</sup>Salichos and Rokas (2011) *PLoS One* 6:e18755 doi:10.1371/journal.pone.0018755.g006

Four methods tested against 2,723 curated orthologues from six *Saccharomycetes*

- RBBH (and cRBH); RSD (and cRSD); MultiParanoid; OrthoMCL
- Rated by statistical performance metrics: sensitivity, specificity, accuracy, FDR

**cRBH most accurate and specific, with lowest FDR.**





# Which prediction methods work best? <sup>a</sup> <sup>b</sup>

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<sup>a</sup>Wolf and Koonin (2012) *Genome Biol. Evol.* **4**:1286-1294 doi:10.1093/gbe/evs100

<sup>b</sup>Altenhoff and Dessimoz (2009) *PLoS Comp. Biol.* **5**:e1000262 doi:10.1371/journal.pcbi.1000262

- Performance varies by choice of method, and interpretation of “orthology”
- Biggest influence is genome annotation quality
- Relative performance varies with choice of benchmark
- **(clustering) RBH outperforms more complex algorithms under many circumstances**



# How orthologues help

Defining core groups of genes as “orthologues” allows analysis of groups of genes by:

- synteny/collocation
- gene neighbourhood changes (e.g. *genome expansion*)
- pan genome (core/accessory genomes)

and of individual genes within those groups, by:

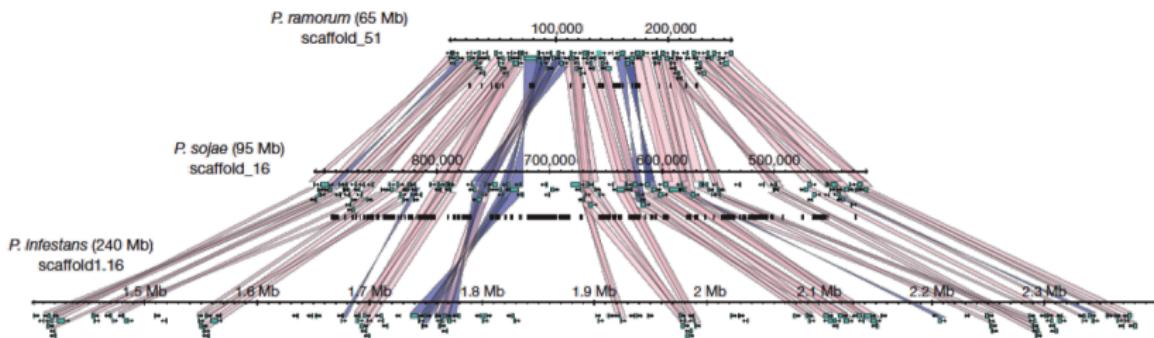
- multiple alignment
- domain detection
- identification of functional sites
- inference of directional selection (stabilising/positive selection)



# Genome expansion <sup>a</sup>

<sup>a</sup>Haas et al. (2009) *Nature* doi:10.1038/nature08358

- Mobile/repeat elements reproduce and expand during evolution
- Generates a “sequence laboratory” for variation and experiment
- e.g. *Phytophthora infestans* effector protein expansion and arms race



**Figure 1 |** Repeat-driven genome expansion in *Phytophthora infestans*. Conserved gene order across three homologous *Phytophthora* scaffolds. Genome expansion is evident in regions of conserved gene order, a

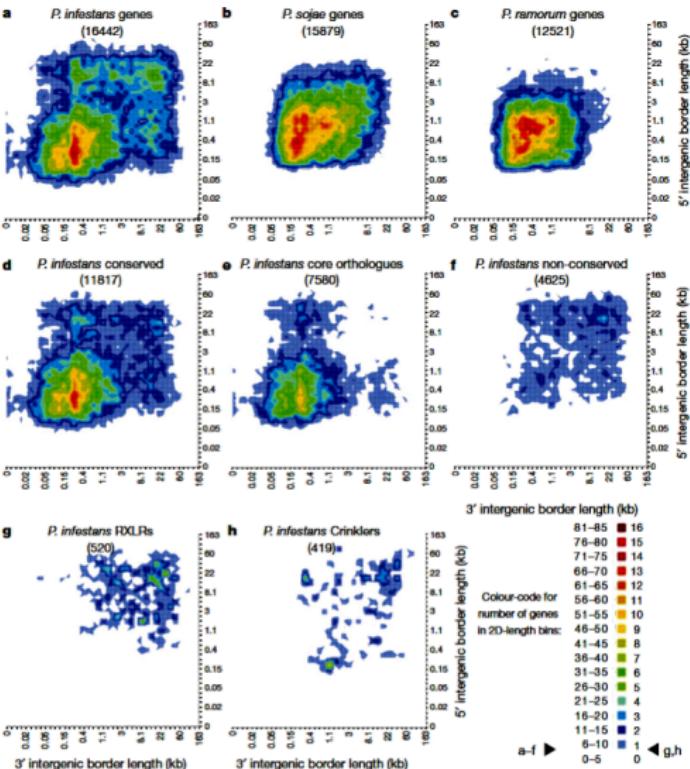
consequence of repeat expansion in intergenic regions. Genes are shown as turquoise boxes, repeats as black boxes. Collinear orthologous gene pairs are connected by pink (direct) or blue (inverted) bands.



# Genome expansion <sup>a</sup>

<sup>a</sup>Haas et al. (2009) *Nature* doi:10.1038/nature08358

- Mobile elements (MEs) are large, and duplicate/carry genes with them
- Larger intergenic regions in MEs
- Effector proteins found preferentially in regions with large gaps
- Two-speed genome associated with adaptability





# The Pangenome

**The Core Genome Hypothesis:**  
**“The core genome is the primary cohesive unit defining a bacterial species”**

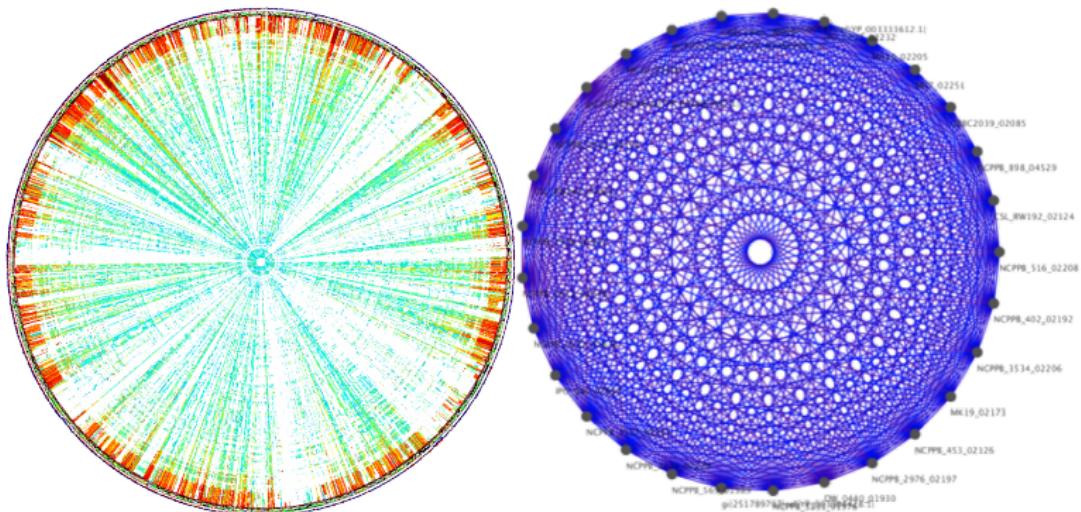


# Core genome <sup>a b</sup>

<sup>a</sup>Laing (2010) *BMC Bioinf.* doi:10.1186/1471-2105-11-461

<sup>b</sup>Lefébure *et al.* (2010) *Genome Biol. Evol.* doi:10.1093/gbe/evq048

Once equivalent genes have been identified, those present in all related isolates can be identified: **the core genome**.



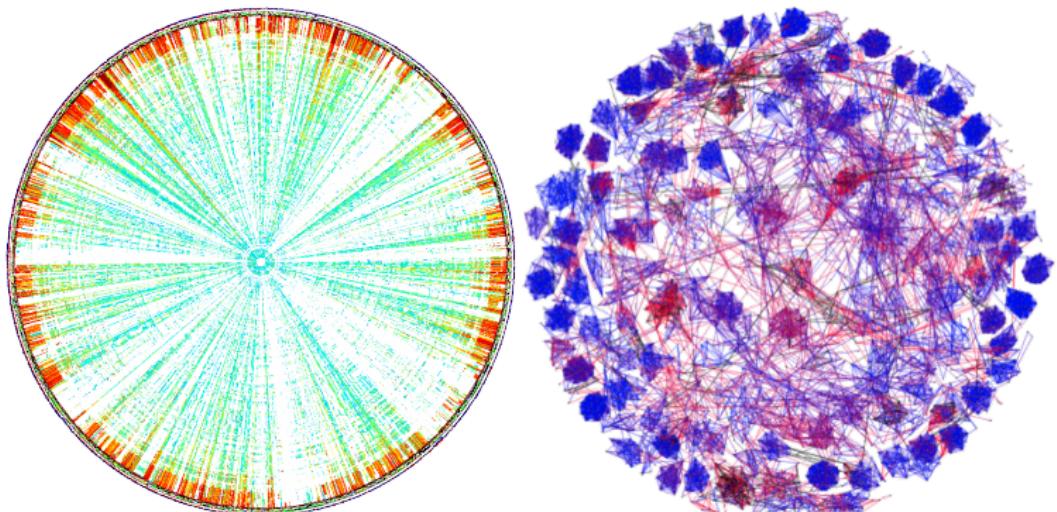


# Accessory genome <sup>a</sup> <sup>b</sup>

<sup>a</sup> Laing (2010) *BMC Bioinf.* doi:10.1186/1471-2105-11-461

<sup>b</sup> Lefébure et al. (2010) *Genome Biol. Evol.* doi:10.1093/gbe/evq048

The remaining genes are **the accessory genome**, and are expected to mediate function that distinguishes between isolates.



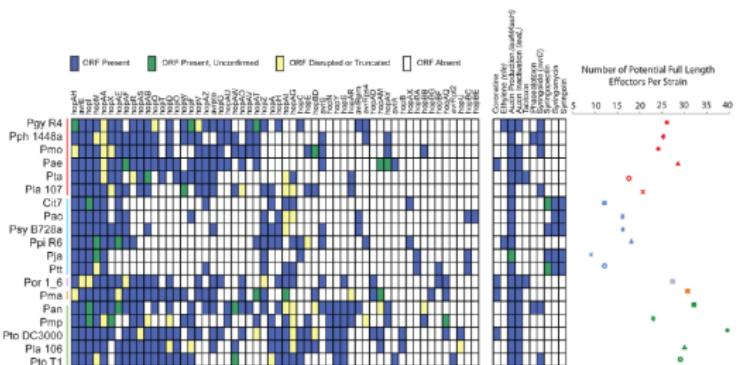


## Accessory genome <sup>a b</sup>

<sup>a</sup>Croll and McDonald (2012) *PLoS Path.* 8:e1002608 doi:10.1371/journal.ppat.1002608

<sup>b</sup> Baltrus et al. (2011) PLoS Pathog. 7:e1002132 doi:10.1371/journal.ppat.1002132.t002

Accessory genomes are a cradle for adaptive evolution  
This is particularly so for bacterial pathogens, such as  
*Pseudomonas* spp.



**Figure 3.** *S. pyogenes* isolates harbor extensive diversity in virulence gene repertoires. TTE, toxin, and plant hormone biosynthesis genes are listed across the top. *S. pyogenes* genomes, color-coded by phylogenetic group as in Figure 1. At the left, a blue box indicates presence of full-length ORFs or complete pathways within each genome. Green boxes indicate that genes or pathways are present by similarity searches, but the presence of full-length genes could not be verified by PCR, or the pathways are potentially incomplete. Yellow boxes indicate that genes are either significantly truncated or are disrupted by insertion sequence elements. White boxes indicate absence of genes or pathways from the strains based on homology searches. At the far right, the total number of potentially functional TTE proteins is shown for each genome and displayed according to the color-coded group symbols shown in Figure 1.



# Identifying the Pangenome <sup>a</sup>

<sup>a</sup>Page et al. (2015) *Bioinf.* **31**:3691-3693 doi:10.1093/bioinformatics/btv421

Roary can produce pangenomes for 1000s of prokaryotes on a desktop machine

- Pre-cluster with CD-HIT (reduce input size)
- All-against-all on reduced sequence set
- MCL clustering
- Merge clusters and use synteny to identify orthologues

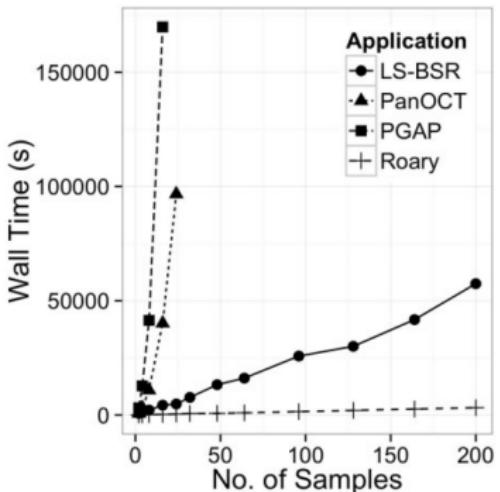


Fig. 1. Effect of dataset size on the wall time of multiple applications. Only analysis that completed within 2 days and 60 GB of RAM is shown



# What didn't I get to?

- Genome-Wide Association Studies (GWAS)
  - Try <http://genenetwork.org/> to play with some data
- Prediction of regulatory elements, e.g.
  - Kellis *et al.* (2003) *Nature* doi:10.1038/nature01644
  - King *et al.* (2007) *Genome Res.* doi:10.1101/gr.5592107
  - Chaivorapol *et al.* (2008) *BMC Bioinf.* doi:10.1186/1471-2105-9-455
  - CompMOBY <http://genome.ucsf.edu/compmoby>
- Detection of Horizontal/Lateral Gene Transfer (HGT/LGT), e.g.
  - Tsirigos & Rigoutsos (2005) *Nucl. Acids Res.* doi:10.1093/nar/gki187
- Phylogenomics, e.g.
  - Delsuc *et al.* (2005) *Nat. rev. Genet.* doi:10.1038/nrg1603
  - AMPHORA <https://phylogenomics.wordpress.com/software/amphora/>



## Messages to take away

- Comparative genomics is a powerful set of techniques for:
  - Understanding and identifying evolutionary processes and mechanisms
  - Reconstructing detailed evolutionary history
  - Identifying and understanding common genomic features
  - Providing hypotheses about gene function for experimental investigation



# Messages to take away

- Comparative genomics is comparisons
  - What is *similar* between two genomes?
  - What is *different* between two genomes?
- Comparative genomics *is* evolutionary genomics
  - Lots of scope for improvement in tools
- Tools that 'do the same thing' can give different output
  - BLAST vs MUMmer
  - RBBH vs MCL
  - The choice of application matters for correctness and interpretation



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