# Training and testing a K-Top-Scoring-Pair (KTSP) classifier with switchBox.

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## 1 Introduction

The switchBox package allows to train and validate a K-Top-Scoring-Pair (KTSP) classifier, as used by Marchionni et al in [1]. KTSP is an extension of the TSP classifier described by Geman and colleagues [2, 3, 4]. The TSP algorithm is a simple binary classifier based on the ordering of two measurements.

#### ADD about TSP and KTSP

The switchBox package contains several utilities enabling to:

- 1. Filter the features to be used to develop the classifier (*i.e.*, differentially expressed genes);
- 2. Compute the scores for all available feature pairs to identify the top performing TSP;
- 3. Compute the scores for selected feature pairs to identify the top performing TSP;
- 4. Identify the number of K TSP to be used in the final classifier;
- 5. Compute individual TSP votes for one class or the other and aggregate the votes based on various methods;
- 6. Classify new samples based on the top KTSP based on various methods;

# 2 Installing the package

Download and install the package switchBox from Bioconductor.

```
> source("http://bioconductor.org/biocLite.R")
> biocLite("switchBox")
```

Load the library.

```
> require(switchBox)
```

## 3 Data structure

## 3.1 Training set

Load the example training data contained in the switchBox package.

```
> ### Load the example data for the TRAINING set
> data(matTraining)
```

The object matTraining is a numeric matrix containing gene expression data for the 78 breast cancer patients and the 70 genes used to implement the MammaPrint assay [5]. This data was obtained from from the MammaPrintData package, as described in [1]. Samples are stored by column and genes by row. Gene annotation is stored as rownames (matTraining).

```
> class(matTraining)
[1] "matrix"
> dim(matTraining)
[1] 70 78

> str(matTraining)

num [1:70, 1:78] -0.0564 0.0347 -0.0451 -0.1556 0.1394 ...
- attr(*, "dimnames")=List of 2
    ..$ : chr [1:70] "AA555029_RC_Hs.370457" "AF257175_Hs.15250" "AK000745_Hs.377155" "AKAP2_Hs.516834"
    ..$ : chr [1:78] "Training1.Bad" "Training2.Bad" "Training3.Good" "Training4.Good" ...
```

Prognostic information can be extracted from the column names of the expression matrix, as follows:

```
> ### The FIRST level is for "Bad" group
> trainingGroup <- factor(gsub(".+\\.", "", colnames(matTraining)))
> ### Show group variable for the TRAINING set
> table(trainingGroup)

trainingGroup
Bad Good
34 44
```

## 3.2 Testing set

Load the example testing data contained in the switchBox package.

```
> ### Load the example data for the TEST set
> data(matTesting)
```

The object matTesting is a numeric matrix containing gene expression data for the 307 breast cancer patients and the 70 genes used to validate the MammaPrint assay [6]. This data was obtained from from the MammaPrintData package, as described in [1]. Also in this case samples are stored by column and genes by row. Gene annotation is stored as rownames (matTraining).

```
> class(matTesting)
[1] "matrix"
> dim(matTesting)
[1] 70 307
> str(matTesting)
num [1:70, 1:307] 0.0035 -0.0599 -0.0678 0.1139 -0.094 ...
- attr(*, "dimnames") = List of 2
    ..$ : chr [1:70] "AA555029_RC_Hs.370457" "AF257175_Hs.15250" "AK000745_Hs.377155" "AKAP2_Hs.516834"
    ..$ : chr [1:307] "Test1.Good" "Test2.Good" "Test3.Good" "Test4.Good" ...
```

Prognostic information can be extracted from the column names of the expression matrix, as follows:

```
> ### Create the group variable from the column names
> testGroup <- factor(gsub(".+\\.", "", colnames(matTesting)))
> ### Show group variable for the TEST set
> table(testGroup)

testGroup
Bad Good
47 260
```

# 4 Training KTSP algorithm

#### 4.1 Unrestricted KTSP classifiers

We can train the KTSP algoritm using all possible feature pairs – unrestricted KTSP classifier – with or without statistical feature filtering, using the SWAP.KTSP.Train.

### 4.1.1 Default statistical filtering

Training an unrestricted KTSP predictor using a statistical feature filtering is the default and it is achieved by using the default parameters, as follows:

```
> ### The arguments to the "SWAP.KTSP.Train" function
> args(SWAP.KTSP.Train)
function (inputMat, phenoGroup, krange = c(3, 5, 7:10), FilterFunc = SWAP.Filter.Wilcoxon,
   RestrictedPairs, ...)
> ### Train a classifier using default filtering function based on the Wilcoxon test
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup, krange=c(3:15))</pre>
Applying filtering function to 'inputMat'...
Computing scores for 70 features.
This will require enough memory for 2415 pairs.
Selecting K...
7 TSP will be used to build the final classifier.
> ### Show the classifier
> classifier
[1] "7TSPs"
$TSPs
[, 1] [, 2] [, 2] [1,] "GNAZ_Hs.555870" "Cont
                                  "Contig32185_RC_Hs.159422"
[1,] "GNAZ_Hs.555870"
[2,] "Contig46223_RC_Hs.22917" "OXCT_Hs.278277"
[3,1] "RFC4 Hs.518475" "L2DTL_Hs.445885"
[4,] "Contig40831_RC_Hs.161160" "CFFM4_Hs.250822"
[5,] "FLJ11354_Hs.523468" "LOC57110_Hs.36761"
[6,] "Contig55725_RC_Hs.470654" "IGFBP5_Hs.184339"
[7,] "UCH37_Hs.145469" "SERF1A_Hs.32567"
[1] 0.6029423 0.5467924 0.5347600 0.5280755 0.5267389 0.5200542 0.5133699
$labels
[1] "Bad" "Good"
> ### Extract the TSP from the classifier
> classifier$TSPs
```

```
[,1] [,2]
[1,] "GNAZ_Hs.555870" "Contig32185_RC_Hs.159422"
[2,] "Contig46223_RC_Hs.22917" "OXCT_Hs.278277"
[3,] "RFC4_Hs.518475" "L2DTL_Hs.445885"
[4,] "Contig40831_RC_Hs.161160" "CFFM4_Hs.250822"
[5,] "FLJ11354_Hs.523468" "LOC57110_Hs.36761"
[6,] "Contig55725_RC_Hs.470654" "IGFBP5_Hs.184339"
[7,] "UCH37_Hs.145469" "SERF1A_Hs.32567"
```

Below is shown the way the default feature filtering works. The SWAP.Filter.Wilcoxon function takes the phenotype factor, the predictor data, the number of feature to be returned, and a logical value to decide whether to include equal number of featured positively and negatively associated with the phenotype to be predicted.

```
> ### The arguments to the "SWAP.KTSP.Train" function
> args(SWAP.Filter.Wilcoxon)
function (phenoGroup, inputMat, featureNo = 100, UpDown = TRUE)
NULL
> ### Retrieve the top best 4 genes using default Wilcoxon filtering
> ### Note that there are ties
> SWAP.Filter.Wilcoxon(trainingGroup, matTraining, featureNo=4)
[1] "KIAA0175_Hs.184339" "IGFBP5_Hs.184339" "RFC4_Hs.518475"
[4] "FLJ11354_Hs.523468" "GNAZ_Hs.555870"
```

Train a classifier using the SWAP.Filter.Wilcoxon filtering function.

```
> ### Train a classifier from the top 4 best genes
> ### according to Wilcoxon filtering function
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup,</pre>
                              FilterFunc=SWAP.Filter.Wilcoxon, featureNo=4)
Applying filtering function to 'inputMat'...
Computing scores for 5 features.
This will require enough memory for 10 pairs.
Selecting K...
The required range of k is not available!
The minimum number of available TSP (2) will be used instead.
> ### Show the classifier
> classifier
$name
[1] "2TSPs"
                          [,2]
[1,] "FLJ11354_Hs.523468" "IGFBP5_Hs.184339"
[2,] "RFC4_Hs.518475" "KIAA0175_Hs.184339"
[1] 0.5173798 0.4826204
$labels
[1] "Bad" "Good"
```

#### Train a classifier using all possible features:

```
> ### To use all features "FilterFunc" must be set to NULL
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup, FilterFunc=NULL)
No feaure filtering procedure will be used...
Computing scores for 70 features.
This will require enough memory for 2415 pairs.
Selecting K...
7 TSP will be used to build the final classifier.
> ### Show the classifier
> classifier
$name
[1] "7TSPs"
$TSPs
[,2]
[1,] "GNAZ_Hs.555870" "Cont
                                "Contig32185_RC_Hs.159422"
[2,] "Contig46223_RC_Hs.22917" "OXCT_Hs.278277" [3,] "RFC4_Hs.518475" "L2DTL_Hs.445885"
[4,] "Contig40831_RC_Hs.161160" "CFFM4_Hs.250822"
[5,] "FLJ11354_Hs.523468" "LOC57110_Hs.36761"
[6,] "Contig55725_RC_Hs.470654" "IGFBP5_Hs.184339"
[7,] "UCH37_Hs.145469" "SERF1A_Hs.32567"
[1] 0.6029423 0.5467924 0.5347600 0.5280755 0.5267389 0.5200542 0.5133699
$labels
[1] "Bad" "Good"
```

#### 4.1.2 Altenative filtering methods

Training can also be achieved using alternative filtering methods. These methods can be specified by passing a different filtering function to SWAP.KTSP.Train. These functions should use th phenoGroup, inputData arguments, as well as any other necessary argument (passed using . . .), as shown below.

For instance, we can define an alternative filtering function selecting 10 random features.

```
> ### An alternative filtering function selecting 20 random features
> random10 <- function(situation, data) { sample(rownames(data), 10) }
> random10(trainingGroup, matTraining)

[1] "AA555029_RC_Hs.370457" "TMEFF1_Hs.336224" "DKFZP564D0462_Hs.318894"
[4] "ESM1_Hs.129944" "Contig55377_RC_Hs.463089" "PRC1_Hs.366401"
[7] "Contig28552_RC_Hs.508141" "MMP9_Hs.297413" "CENPA_Hs.1594"
[10] "AF257175_Hs.15250"
```

Below is a more realistic example of an alternative filtering function. In this case we use the Rt.test function to select the features with an absolute t-statistics larger than a specified quantile.

```
> ### An alternative filtering function based on a t-test
> topRttest <- function(situation, data, quant = 0.75) {
      out <- apply(data, 1, function(x, ...) t.test(x ~ situation)$statistic)
      names(out[ abs(out) > quantile(abs(out), quant) ])
}
> ### Show the top 5% features using the newly defined filtering function
> topRttest(trainingGroup, matTraining, quant=0.95)

[1] "Contig32185_RC_Hs.159422" "FLJ11354_Hs.523468" "IGFBP5_Hs.184339"
[4] "KIAA0175_Hs.184339"
```

Train a classifier using the alternative filtering function based on the t-test and also define the max number of TSP using krange.

```
> ### Train with t-test and krange
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup,
                              FilterFunc = topRttest, quant = 0.9, krange=c(15:30) )
Applying filtering function to 'inputMat'...
Computing scores for 7 features.
This will require enough memory for 21 pairs.
Selecting K ...
The required range of k is not available!
The minimum number of available TSP (3) will be used instead.
> ### Show the classifier
> classifier
[1] "3TSPs"
$TSPs
[,1] [,2] [,2] [1,] "GNAZ_Hs.555870" "Contig32185_RC_Hs.159422"
[2,] "FLJ11354_Hs.523468" "IGFBP5_Hs.184339"
[3,] "SERF1A_Hs.32567" "MMP9_Hs.297413"
[1] 0.6029413 0.5173798 0.1631016
$labels
[1] "Bad" "Good"
```

## 4.2 Training a Restricted KTSP algorithm

The swithcBox3 allows to training a KTSP classifier using a pre-specified set of restricted feature pairs. This can be useful to implement KTSP classifiers mechanistically restricted. To achieve this the specific TSP to be used must be defined using the RestrictedPairs. As an example we can define a set of specific pairs to be used for classifier development by randolmly selecting some of the rownames from the inputMat matrix. In a real example these pairs would be provided by the user. The restricted pairs must be encoded using valid names.

Train a classifier using the set of restricted feature pairs and the default filtering:

```
> ### Train
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup,</pre>
                                    RestrictedPairs = somePairs, krange=3:16)
Applying filtering function to 'inputMat'...
Restricting the analysis to the provided candidate TSPs
Computing scores for 18 features.
This will require enough memory for 315 pairs.
Selecting K...
11 TSP will be used to build the final classifier.
> ### Show the classifier
> classifier
Sname
[1] "11TSPs"
$TSPs
       [,1]
                                       [,2]
 [1,] "DKFZP564D0462_Hs.318894" "RFC4_Hs.518475"
 [2,] "MP1_Hs.26010" "KIAA1442_Hs.471955" [3,] "SERF1A_Hs.32567" "PK428_Hs.516834" [4,] "FGF18_Hs.87191" "Contig46223_RC_Hs.22917" [5,] "AK000745_Hs.377155" "IGFBP5_Hs.511093"
```

```
[6,] "FLJ22477_Hs.149004" "FLT1_Hs.507621"
[7,] "KIAA0175_Hs.184339" "EXT1_Hs.492618"
[8,] "TMEFF1_Hs.336224" "Contig55377_RC_Hs.463089"
[9,] "ALDH4_Hs.133062" "Contig55725_RC_Hs.470654"
[10,] "ESM1_Hs.129944" "FLJ11190_Hs.516834"
[11,] "AA555029_RC_Hs.370457" "COL4A2_Hs.508716"

$score
[1] 0.4532088 0.3943852 0.3836903 0.3810164 0.3128345 0.2834227 0.2500002 0.2259360
[9] 0.2032088 0.1938504 0.1911766

$labels
[1] "Bad" "Good"
```

Train a classifier using a set of restricted feature pairs, defining the maximum number of TSP using krange and also filtering the features by T-test.

```
> ### Train
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup,</pre>
                                    RestrictedPairs = somePairs,
                                    FilterFunc = topRttest, quant = 0.3,
                                   krange=c(3:10) )
Applying filtering function to 'inputMat'...
Restricting the analysis to the provided candidate TSPs
Computing scores for 10 features.
This will require enough memory for 125 pairs.
Selecting K...
9 TSP will be used to build the final classifier.
> ### Show the classifier
> classifier
$name
[1] "9TSPs"
$TSPs
 [,1] [,2]
[1,] "SERF1A_Hs.32567" "PK428_Hs.516834"
[2,] "FGF18_Hs.87191" "Contig46223_RC_Hs.22917"
 [3,] "AK000745_Hs.377155" "IGFBP5_Hs.511093"
 [4,] "KIAA0175_Hs.184339" "EXT1_Hs.492618"
 [5,] "TMEFF1_Hs.336224" "Contig55377_RC_Hs.463089"
 [6,] "ESM1_Hs.129944" "FLJ11190_Hs.516834"
 [7,] "AL137718_Hs.508141" "PECI_Hs.15250"
 [7,] "ALI3//10_10_10

[8,] "ECT2_Hs.518299" "ORC6L_Hs.49/00

"CFFM4_Hs.250822"
 [9,] "OXCT_Hs.278277"
 \begin{smallmatrix} 11 \end{smallmatrix} \ \ 0.38369019 \ \ 0.38101624 \ \ 0.31283440 \ \ 0.25000012 \ \ 0.22593590 \ \ 0.19385035 \ \ 0.17780769 
[8] 0.13636374 0.08021399
$labels
[1] "Bad" "Good"
```

# 5 Calculate and aggregates the TSP votes

The SWAP.KTSP.Statistics function can be used to compute and aggregate the TSP votes using alternative functions to combine the votes. The default method is the count of the signed TSP votes. We can also pass a different function to combine the KTSPs. This function takes an argument x-a logical vector corresponding to the TSP votes – of length equal to the number of columns (e.g., the number of cancer patients under analysis) and aggregates the votes of all K TSP of the classifier.

Here we will use the default parameters (the count of the signed TSP votes)

```
> ### Train a classifier
> classifier <- SWAP.KTSP.Train(matTraining, trainingGroup,
                              FilterFunc = NULL, krange=8)
No feaure filtering procedure will be used...
Computing scores for 70 features.
This will require enough memory for 2415 pairs.
Selecting K...
8 TSP will be used to build the final classifier.
> ### Compute the statistics using the default parameters:
> ### counting the signed TSP votes
> ktspStatDefault <- SWAP.KTSP.Statistics(inputMat = matTraining,
     classifier = classifier)
> ### Show the components in the output
> names(ktspStatDefault)
[1] "statistics" "comparisons"
> ### Show some of the votes
> head(ktspStatDefault$comparisons[ , 1:2])
              GNAZ_Hs.555870>Contig32185_RC_Hs.159422
Training1.Bad
                                                 FALSE
Training2.Bad
                                                 FALSE
Training3.Good
Training4.Good
                                                  TRIIF
Training5.Bad
                                                 FALSE
Training6.Bad
               Contig46223_RC_Hs.22917>OXCT_Hs.278277
Training1.Bad
Training2.Bad
                                                FALSE
Training3.Good
                                                 TRUE
Training4.Good
                                                 TRUE
Training5.Bad
                                                 TRUE
Training6.Bad
                                                FALSE
> ### Show default statistics
> head(ktspStatDefault$statistics)
```

#### Here we will use the sum to aggregate the TSP votes

#### Here, for instance, we will apply a hard treshold equal to 2

We can also make a heatmap showing the individual TSPs votes (see Figure 1 below).

# 6 Classifiy samples and compute the classifier performance

## 6.1 Classifiy training samples

The SWAP.KTSP.Classify function allows to classify one or more samples. Below is shown the **resubstitution** performance in the training set.

```
> ### Show the classifier
> classifier
[1] "8TSPs"
STSPS
[,1] [,2] [,2] [1,] "GNAZ_Hs.555870" "Contig32185_RC_Hs.159422"
[2,] "Contig46223_RC_Hs.22917" "OXCT_Hs.278277"
[3,] "RFC4_Hs.518475" "L2DTL_Hs.445885"
[4,] "Contig40831_RC_Hs.161160" "CFFM4_Hs.250822"
[5,] "FLJ11354_Hs.523468" "LOC57110_Hs.36761"
[6,] "Contig55725_RC_Hs.470654" "IGFBP5_Hs.184339" [7,] "UCH37_Hs.145469" "SERF1A_Hs.32567" [8,] "GSTM3_Hs.2006" "KIAA0175_Hs.18433
                                 "KIAA0175_Hs.184339"
[1] 0.6029423 0.5467924 0.5347600 0.5280755 0.5267389 0.5200542 0.5133699 0.5080221
$labels
[1] "Bad" "Good"
> ### Apply the classifier to the TRAINING set
> trainingPrediction <- SWAP.KTSP.Classify(matTraining, classifier)
> str(trainingPrediction)
Factor w/ 2 levels "Bad", "Good": 1 1 2 2 1 1 2 2 1 1 ...
 - attr(*, "names") = chr [1:78] "Training1.Bad" "Training2.Bad" "Training3.Good" "Training4.Good" ...
> ### Resubstitution performance in the TRAINING set
> table(trainingPrediction, trainingGroup)
                 trainingGroup
trainingPrediction Bad Good
            Bad 30 6
Good 4 38
```

We can apply the classifier using a specific decision to combine the K TSP as specified with the <code>DecideFunc</code> argument of <code>SWAP/KTSP.Classify</code>. This argument is a function working on a logical vector  $\mathbf x$  containing the votes of each TSP. We can for instance count all votes for class one and then classify a patient in one class or the other based on a specific threshold.

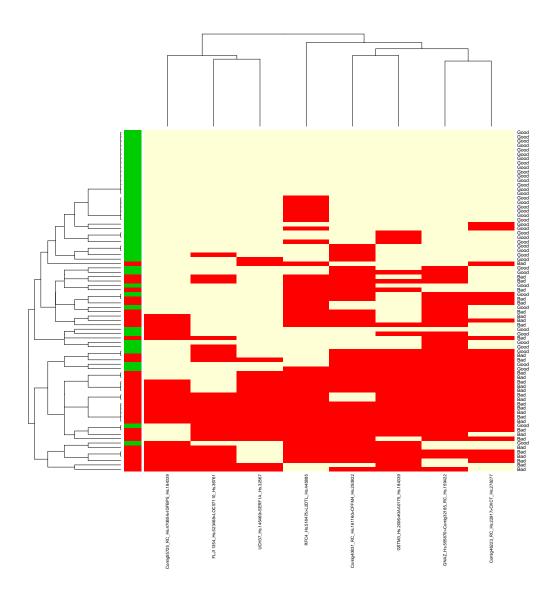


Figure 1: Heatmap showing the individual TSP votes.

## 6.2 Classifiy validation samples

We can apply the trained classifier to one new sample of the test set:

We can apply the trained classifier to a new set of samples, using the defaul decision rule based on the "majority wins" principle:

We can apply the trained classifier to a new set of samples, using an alternative decision rule based as specified by DecideFunc. For instance we can apply the classifier to the test counting all votes for class one and then classify a patient in one class or the other based on a specific threshold.

## 7 Compute the signed TSP scores

The switchBox allows also to compute the individual scores for each TSP of interest. This can be achieved by using the SWAP.CalculateSignedScore function as shown below.

Compute the scores using all features for all possible pairs:

```
> ### Compute the scores using all features for all possible pairs
> scores <- SWAP.CalculateSignedScore(matTraining, trainingGroup, FilterFunc=NULL, )
No feaure filtering procedure will be used...
Computing scores for 70 features.
This will require enough memory for 2415 pairs.
> ### Show scores
> class(scores)
[1] "list"
> dim(scores$score)
[1] 70 70
```

Extract the TSP scores of interest – the absolute value correspond to the scores returned by SWAP.KTSP.Train.

```
> ### Get the scores
> scoresOfInterest <- diag(scores$score[ classifier$TSPs[,1] , classifier$TSPs[,2] ])
> ### Their absolute value should corresponf to the scores returned by SWAP.KTSP.Train
> all(classifier$score == abs(scoresOfInterest))
```

```
[1] TRUE
```

The SWAP.CalculateSignedScore function accept the same argumets used by SWAP.KTSP.Train. It can compute the scores with or without a filtering function and using or not the restricted pairs, as specified by FilterFunc and RestrictedPairs respectively.

```
> ### Compute the scores with default filtering function
> scores <- SWAP.CalculateSignedScore(matTraining, trainingGroup, featureNo=20)
Applying filtering function to 'inputMat'...
Computing scores for 21 features.
This will require enough memory for 210 pairs.
> ### Show scores
> dim(scores$score)
[1] 21 21
> ### Compute the scores without the default filtering function
> ### and using restricted pairs
> scores <- SWAP.CalculateSignedScore(matTraining, trainingGroup,
                                     FilterFunc = NULL, RestrictedPairs = somePairs )
No feaure filtering procedure will be used...
Restricting the analysis to the provided candidate TSPs
Computing scores for 18 features.
This will require enough memory for 315 pairs.
> ### Show scores
> class(scores$score)
[1] "numeric"
> length(scores$score)
[11 18
```

In Figure 2 is shown the histograms for all possible TSP scores.

```
> hist(scores$score, col="salmon", main="TSP scores")
```

# 8 Use of deprecated functions

The two functions KTSP. Train and KTSP. Classify are deprecated and are included in the package only for backward compatibility. They have been substituted by respectively SWAP.KTSP.Train and SWAP.KTSP.Classify. These

> hist(scores\$score, col="salmon", main="TSP scores")

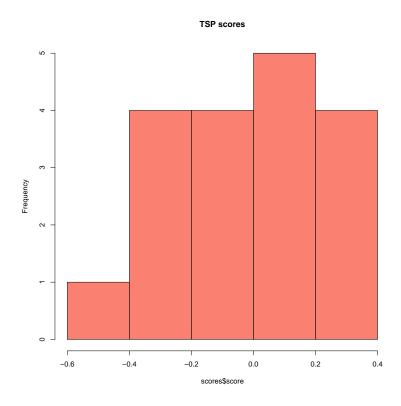


Figure 2: Histograms of all TSP socres.

functions were used to train and validate the 8-TSP classifier described by Marchionni et al [1] and are maintained for reproducibility purposes. Example on the way they are used follows.

Preparation of phenotype information (a numeric vector with values equal to 0 or 1) for training the KTSP classifier:

```
> ### Load gene expression data for the training set
> data(matTraining)
> ### Create a phenotypic group variable for the 78 samples
> trainingGroup <- factor(gsub(".+\\.", "", colnames(matTraining)))
> levels(trainingGroup)

[1] "Bad" "Good"

> ### Turn into a numeric vector with values equal to 0 and 1
> trainingGroup <- as.numeric(trainingGroup) - 1
> ### Show group variable for the TRAINING set
> table(trainingGroup)

trainingGroup
0 1
34 44
```

#### KTSP classifier training:

```
> ### Train a classifier using default filtering function based on the Wilcoxon test
> classifier <- KTSP.Train(matTraining, trainingGroup, n=8)</pre>
> ### Show the classifier
> classifier
$TSPs
    [,1] [,2]
[1,] 42 19
[2,] 24 58
[3,] 63 50
[4,] 22 13
[5,] 37 52
[6,] 27 46
[7,] 69 64
[8,] 43 48
[1] 0.6029417 0.5467919 0.5347597 0.5280752 0.5267384 0.5200538 0.5133694 0.5080218
$geneNames
"Contig32185_RC_Hs.159422"
[2,] "Contig46223_RC_Hs.22917" "OXCT_Hs.278277" [3,] "RFC4_Hs.518475" "L2DTL_Hs.445885"
[4,] "Contig40831_RC_Hs.161160" "CFFM4_Hs.250822"
[5,] "FLJ11354_Hs.523468" "LOC57110_Hs.36761"
[6,] "Contig55725_RC_Hs.470654" "IGFBP5_Hs.184339"
[7,] "UCH37_Hs.145469" "SERF1A_Hs.32567"
[8,] "GSTM3_Hs.2006"
                                 "KIAA0175_Hs.184339"
```

#### KTSP classifier performance:

Preparation of phenotype information (a numeric vector with values equal to 0 or 1) for testing the KTSP classifier on new data:

```
> ### Load the example data for the TEST set
> data(matTesting)
> ### Create the group variable from the column names
> testGroup <- factor(gsub(".+\\.", "", colnames(matTesting)))
> levels(testGroup)

[1] "Bad" "Good"

> ### Turn into a numeric vector with values equal to 0 and 1
> testGroup <- as.numeric(testGroup) - 1
> ### Show group variable for the TEST set
> table(testGroup)

testGroup
0 1
47 260
```

Testing on new data and getting KTSP classifier performance:

```
> ### Apply the classifier to one sample of the TEST set using
> ### sum of votes less than 2.5
> testPrediction <- KTSP.Classify(matTesting, classifier,
    combineFunc = function(x) sum(x) < 2.5
> ### Show prediction
> table(testPrediction)
testPrediction
 0 1
181 126
> ### Contingency table
> table(testPrediction, testGroup)
            testGroup
testPrediction 0 1
          0 43 138
            7
              4 122
```

# 9 System Information

#### Session information:

> toLatex(sessionInfo())

- R version 3.0.2 (2013-09-25), x86\_64-apple-darwin13.0.2
- Locale: C/en\_US.UTF-8/en\_US.UTF-8/C/en\_US.UTF-8/en\_US.UTF-8
- Base packages: base, datasets, grDevices, graphics, methods, stats, utils
- Other packages: switchBox 0.99.7
- Loaded via a namespace (and not attached): tools 3.0.2

## 10 Literature Cited

## References

- [1] Luigi Marchionni, Bahman Afsari, Donald Geman, and Jeffrey T Leek. A simple and reproducible breast cancer prognostic test. *BMC Genomics*, 14:336, 2013.
- [2] Donald Geman, Christian d'Avignon, Daniel Q Naiman, and Raimond L Winslow. Classifying gene expression profiles from pairwise mrna comparisons. *Stat Appl Genet Mol Biol*, 3:Article19, 2004.
- [3] Aik Choon Tan, Daniel Q Naiman, Lei Xu, Raimond L Winslow, and Donald Geman. Simple decision rules for classifying human cancers from gene expression profiles. *Bioinformatics*, 21(20):3896–904, Oct 2005.
- [4] Lei Xu, Aik Choon Tan, Daniel Q Naiman, Donald Geman, and Raimond L Winslow. Robust prostate cancer marker genes emerge from direct integration of inter-study microarray data. *Bioinformatics*, 21(20):3905–11, Oct 2005.

- [5] Annuska M Glas, Arno Floore, Leonie J M J Delahaye, Anke T Witteveen, Rob C F Pover, Niels Bakx, Jaana S T Lahti-Domenici, Tako J Bruinsma, Marc O Warmoes, René Bernards, Lodewyk F A Wessels, and Laura J Van't Veer. Converting a breast cancer microarray signature into a highthroughput diagnostic test. *BMC Genomics*, 7:278, 2006.
- [6] Marc Buyse, Sherene Loi, Laura van't Veer, Giuseppe Viale, Mauro Delorenzi, Annuska M Glas, Mahasti Saghatchian d'Assignies, Jonas Bergh, Rosette Lidereau, Paul Ellis, Adrian Harris, Jan Bogaerts, Patrick Therasse, Arno Floore, Mohamed Amakrane, Fanny Piette, Emiel Rutgers, Christos Sotiriou, Fatima Cardoso, Martine J Piccart, and TRANSBIG Consortium. Validation and clinical utility of a 70-gene prognostic signature for women with node-negative breast cancer. *J Natl Cancer Inst*, 98(17):1183–92, Sep 2006.