## Bioinformatic analysis of Magnaporthe oryzae polyadenylation sites from next generation sequencing data

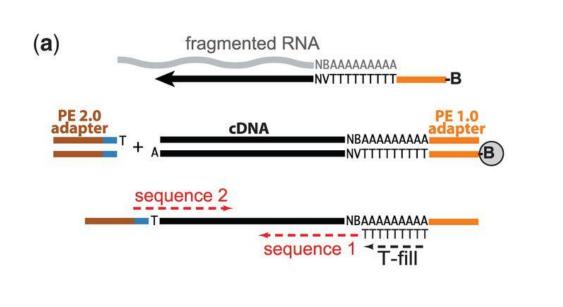
#### Background

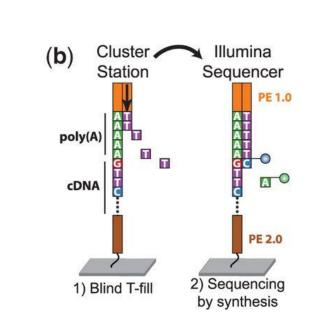
Several proteins have been shown to regulate alternative polyadenylation (APA), including Cleavage Factor I (CFIm) in metazoan. The ascomycetous fungus Magnaporthe oryzae, also known as rice blast, is a plant-pathogenic fungus that causes a serious disease affecting rice. Rbp35 is the functional M. oryzae equivalent of Human CFIm68. Δrbp35 knock-out mutant is viable indicating that Rbp35 is not an essential components of the polyadenylation machinery in the rice blast fungus. However,  $\Delta rbp35$  mutants shows developmental and virulence defects.

#### Results

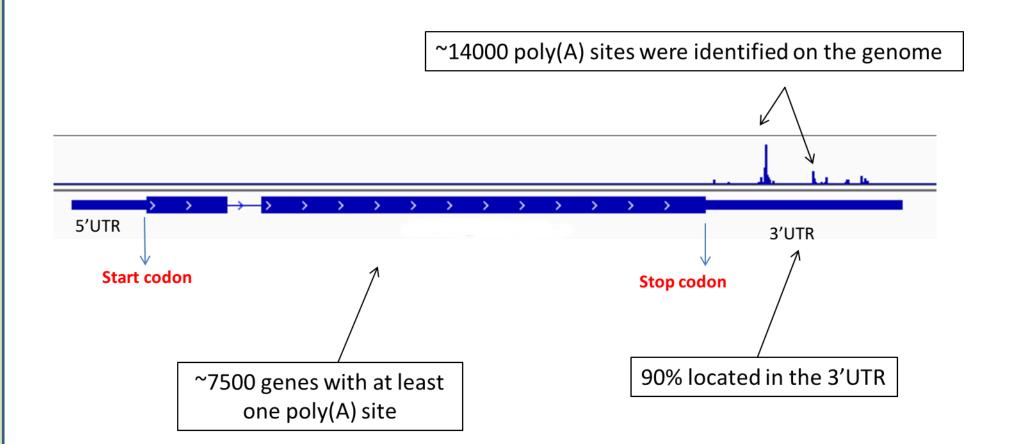
- Using a novel sequencing protocol, we mapped the polyadenylation sites of M. oryzae in four different growing conditions and identified more than 14000 high-confidence polyadenylation sites, accounting for more than 7,000 protein coding genes
- 30% of *M. oryzae* genes are alternatively polyadenylated, and grouped in specific functional groups.
- The nucleotide context and protein-binding regions differ from budding yeast.
- Polyadenylation sites possess a specific predicted RNA secondary structure, also depending on the elements defining the polyadenylation site.
- Under carbon starvation, polyadenylation site selection is altered in more than 400 genes, producing longer 3'UTR isoforms.
- 25% of the alternatively-polyadenylated transcripts found in the wild type were affected in the Δrbp35 mutant, which indicated that alternative site selection was Rbp35-dependent. Lack of Rbp35 in Δrbp35 affects poly(A) site selection by promoting proximal cuts, resulting in a global shortening of 3'UTRs.
- A UGUAH motif is enriched in Rbp35-dependent poly(A) sites, suggesting that these are the ribonucleotides recognized by Rbp35.
- The  $\Delta rbp35$  mutant seems to have lost the ability to adapt to nitrogen starvation.

### More than 14000 poly(A) sites were identified using a novel sequencing protocol

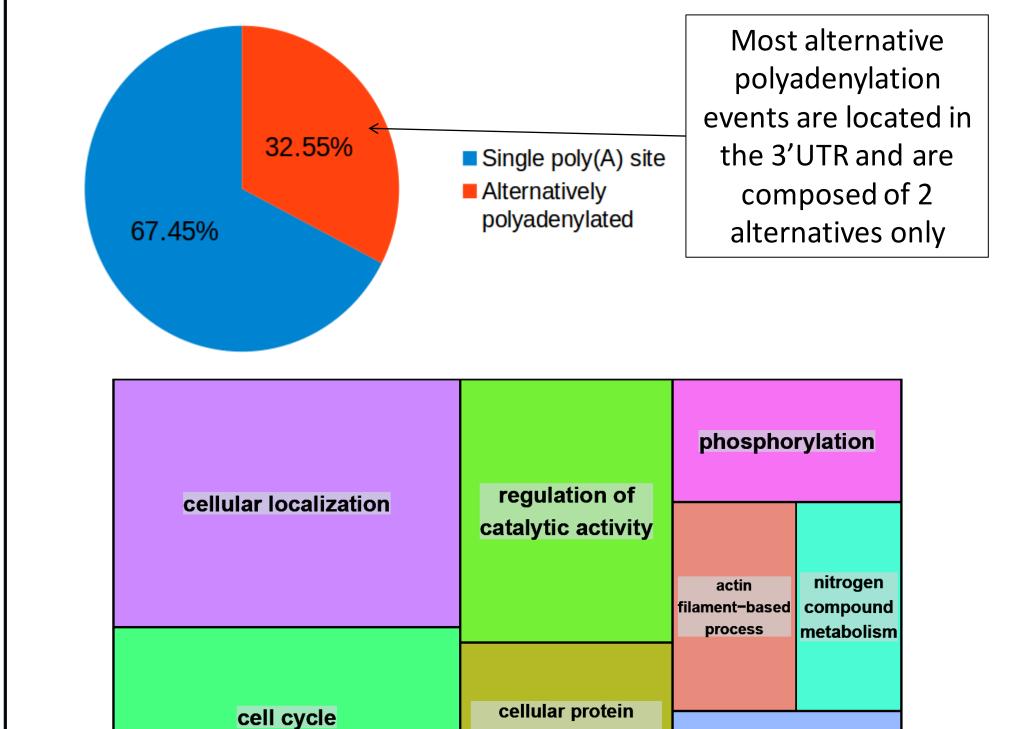




Wilkening, S. et al. An efficient method for genome-wide polyadenylation site mapping and RNA quantification. Nucleic acids research 1-8 (2013).doi:10.1093/nar/gks1249



#### Alternative polyadenylation seems to be involved in specific biological processes

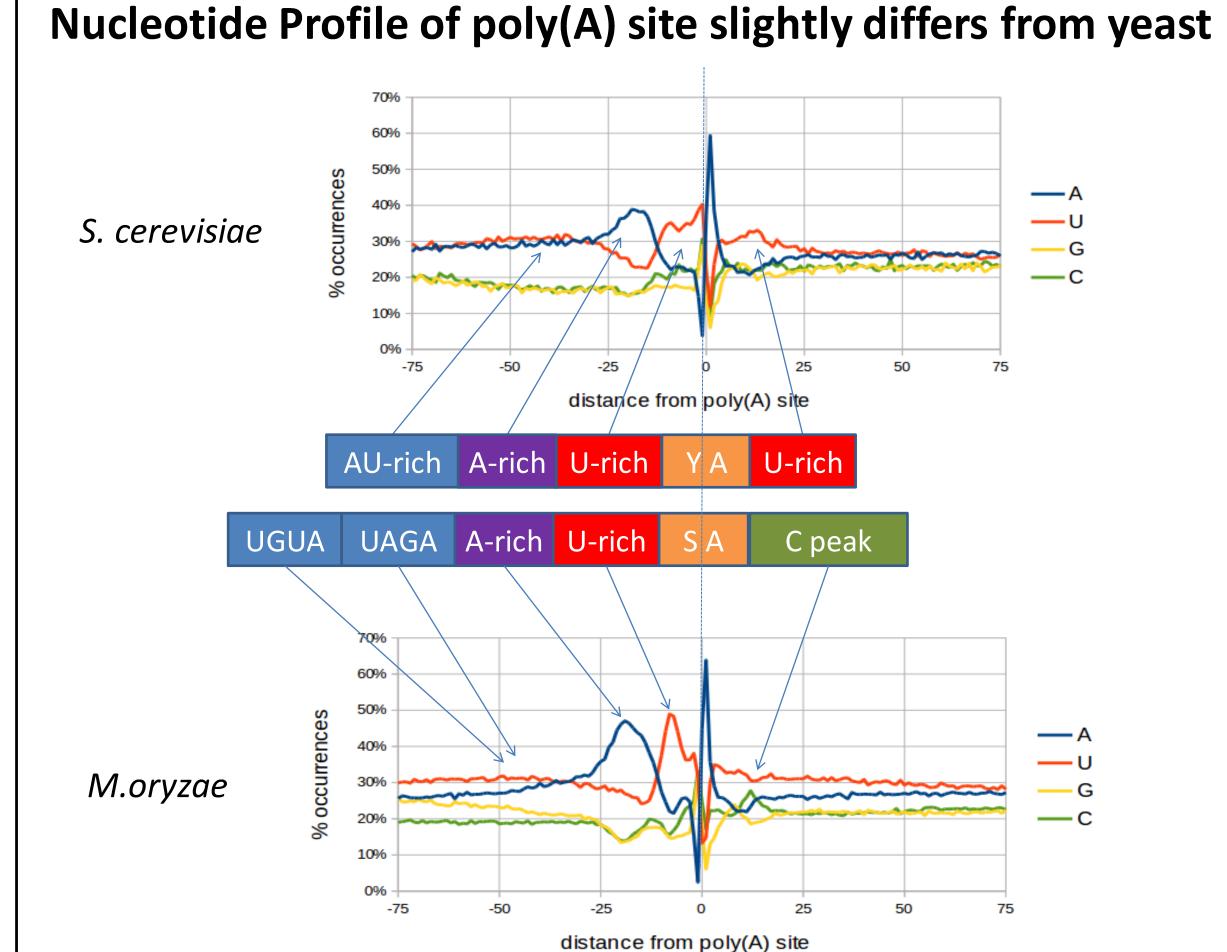


Supek F, Bošnjak M, Škunca N, Šmuc T. "REVIGO summarizes and visualizes long lists of Gene Ontology terms" PLoS ONE 2011. doi:10.1371/journal.pone.0021800

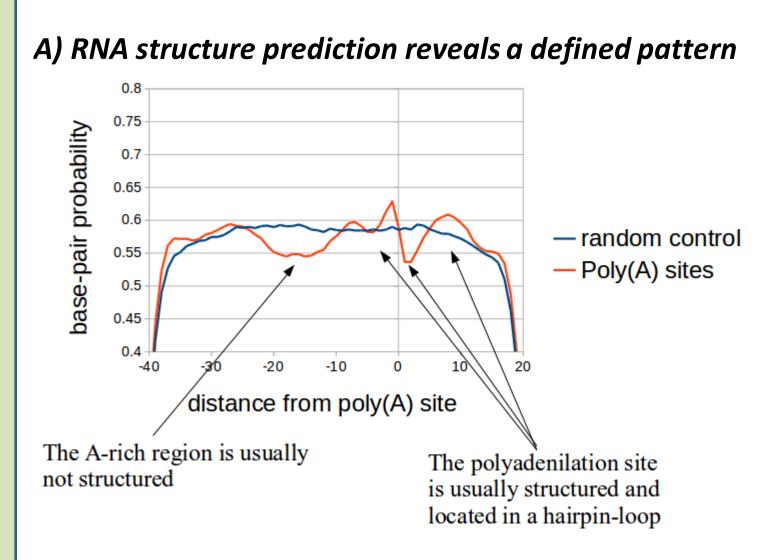
modification process

signaling

# **RBP35** ortholog found in animals, but not in yeast Millevoi, S. & Vagner, S. Nucleic Acids Res. 38, 2757–74 (2010)



#### The polyadenylation site region has a defined predicted secondary structure



C) Different polyadenylation signals have different

distance from poly(A) site

- random control

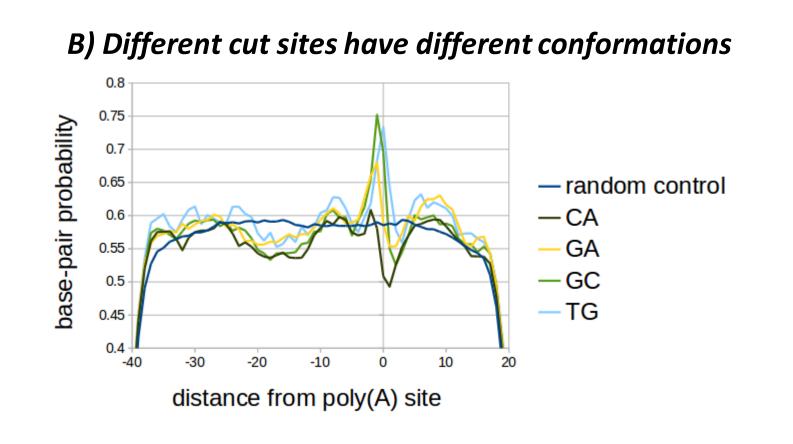
- AAGAAA

AATAAA

- AATAGA

AATTGA

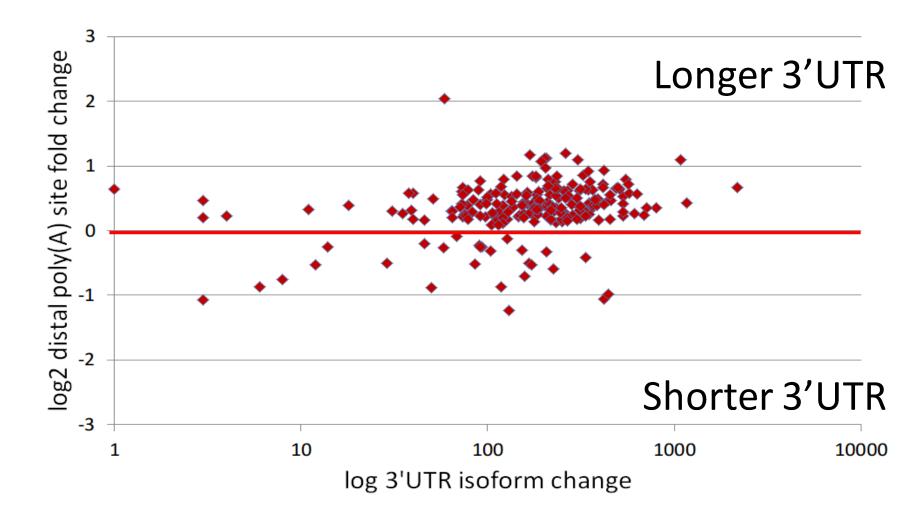
conformations



D) An example, the poly(A) site of 40S ribosomal

#### Carbon starvation medium and $\Delta rbp35$ gene deletion affect poly(A) site selection

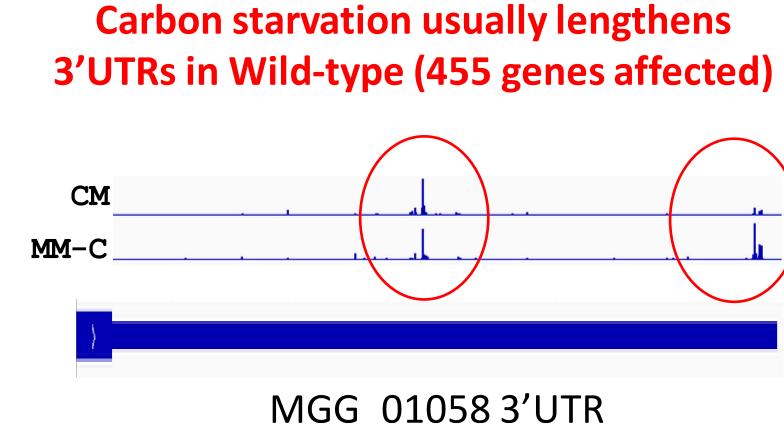
Carbon starvation presents a big number of genes differentially expressed, while

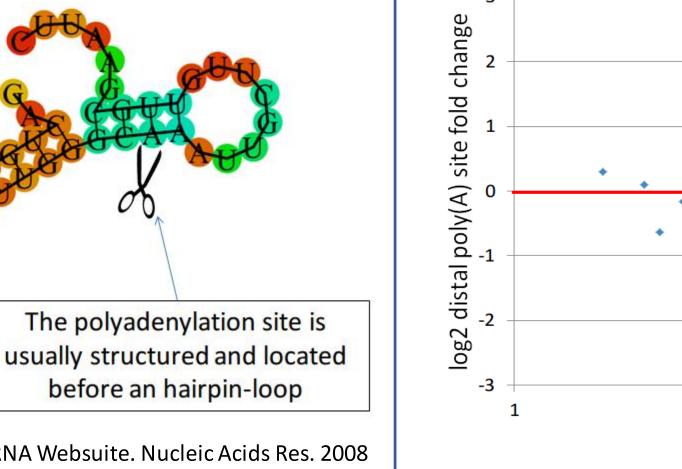


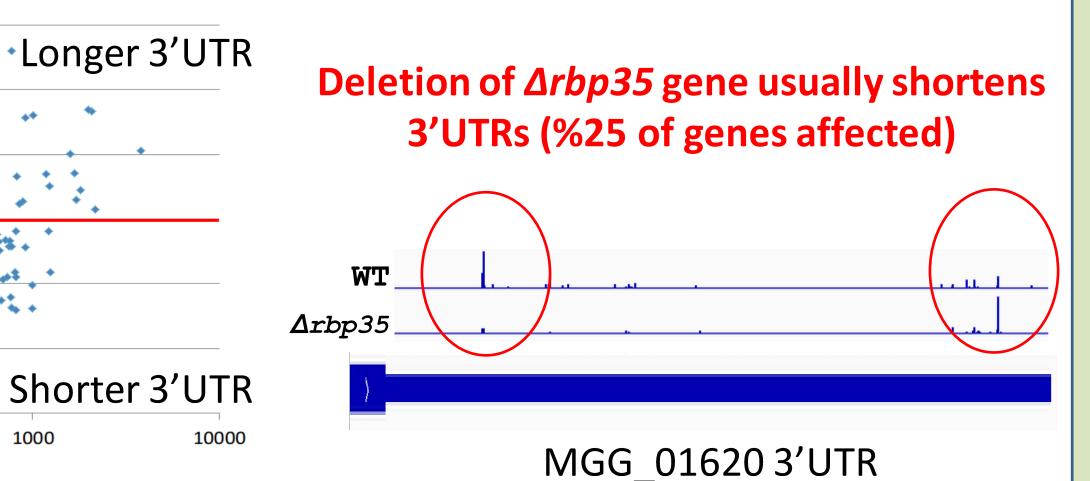
log 3'UTR isoform change

Wild-type

 $CM \rightarrow MM-C$ 





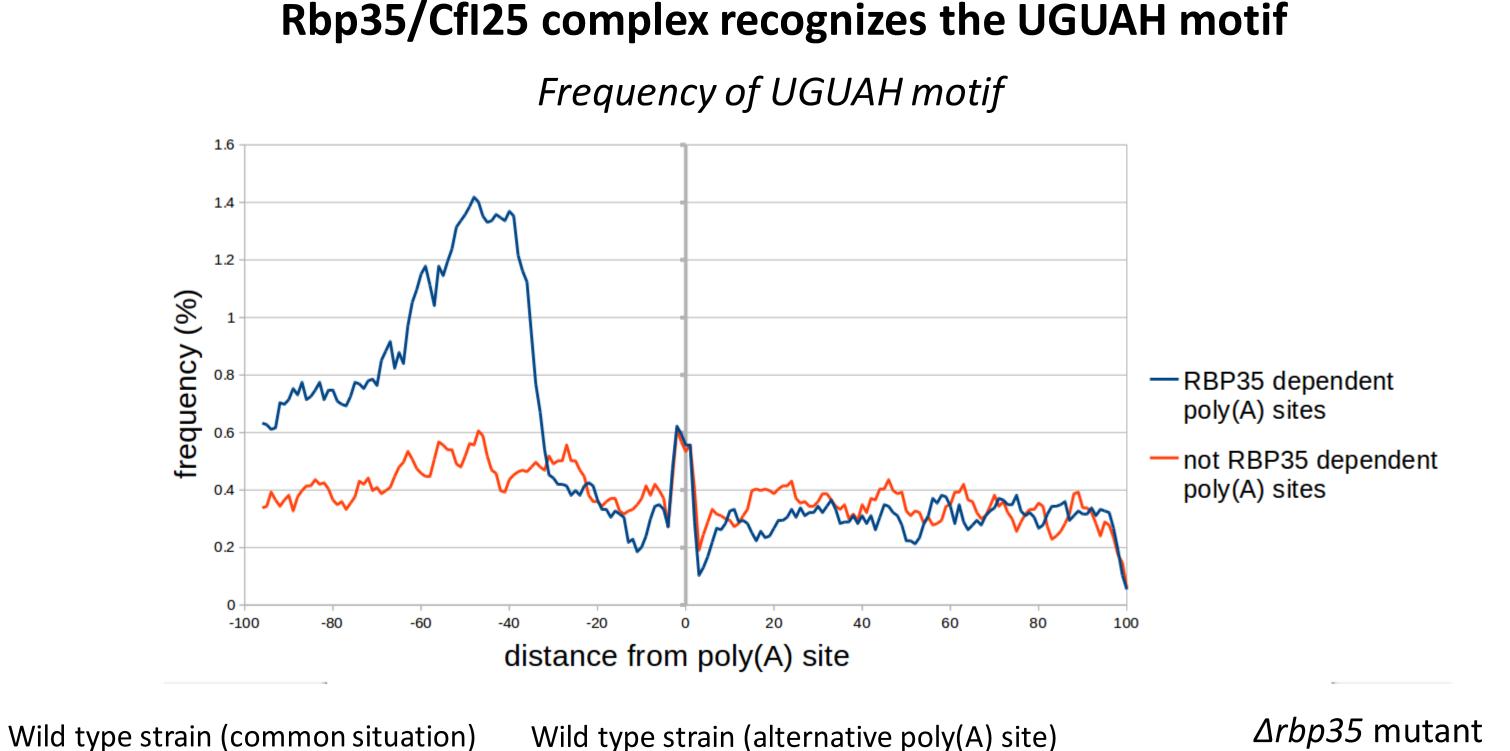


Gruber AR, Lorenz R, Bernhart SH, Neuböck R, Hofacker IL The Vienna RNA Websuite. Nucleic Acids Res. 2008

*protein S15 (MGG\_10370)* 

The A-rich region is usually

not structured

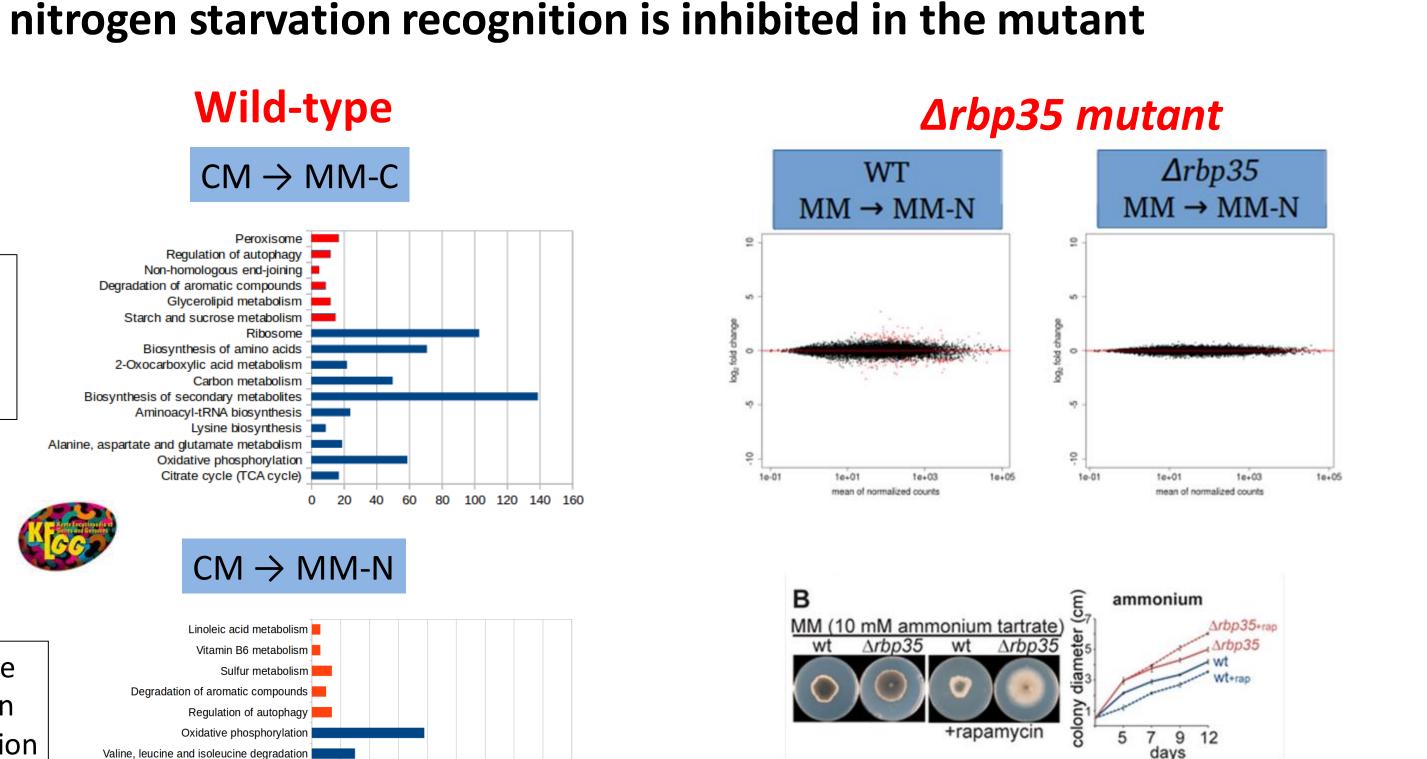


Poly(A) site

"occluded" by

Rbp35

Most metabolic Degradation of aromatic compounds pathways are down-regulated in carbon starvation Alanine, aspartate and glutamate metabolism  $CM \rightarrow MM-N$ **GATA** genes are up-regulated in nitrogen starvation



Both RNA-seq and plate growing display failed adaptation to nitrogen lacking medium



Poly(A) site



Poly(A) site

"revealed" in the

mutant