## Microbial Ecology and Biogeography

OF THE

## Southern Ocean

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September 22, 2012

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## **Abstract**

#### Introduction

#### Some test text to preview layout etc.

The Roseobacter clade is an abundant and ecologically significant group of marine bacteria, found at high (> 15%) abundance in most marine surface environments (Ano (2005) and references therein). Unlike some other major proteobacterial groups which are strongly associated with a particular ecological niche (e.g. the SAR11 clade), roseobacters have diverse metabolic abilities, with members capable (for example) of aerobic anoxygenic phototrophy (Biebl, 2005; ?), degradation of dimethylsulfoniopropionate (DMSP) by at least two pathways (Moran *et al.*, 2007; Miller and Belas, 2004), carbon monoxide oxidation (King, 2003) and heterotrophic utilisation of a broad range of substrates (reviewed in (Brinkhoff *et al.*, 2008)). Roseobacters are found in the planktonic fraction as well as in commensal association with phytoplankton and metazoans (reviewed in Ano (2005)).

#### Microbial ecology of the Southern Ocean

Oceanography of the Southern Ocean

Water masses and fronts

Effect of climate change

Role of the Polar Front in biogeography

Project questions and hypotheses

# The Polar Front as a major biogeographic boundary in the Southern Ocean

Sections of this chapter have been previously published in (TODO: cite PF manuscript)

#### **Summary**

#### Introduction

#### Methods

#### Sampling and metagenomic sequencing

Sampling<sup>1</sup> was conducted on board the RSV *Aurora Australis* during cruise V3 CEAMARC/CASO (Collaborative East Antarctic Marine Census / Climate of Southern Ocean) from 13 December 2007 – 26 January 2008. This cruise occupied the SR3 latitudinal transect from Hobart, Australia (44° S) to the Mertz Glacier, Antarctica (67° S) within a longitudinal range of 140–150° E. Nineteen samples (16 surface, 3 deep) were obtained along almost the entire latitudinal range (Figure 1).

At each station,  $\sim 500$  L of seawater was pumped from  $\sim 2$  m below the sea surface into drums stored at ambient temperature on deck. In the case of deep samples,  $\sim 10$ –50 L of seawater was collected opportunistically from Niskin bottles attached to a CTD (Conductivity, Temperature and Depth) instument operated by an unrelated oceanographic project. Seawater samples were prefiltered through a 20  $\mu$ m plankton net, then filtrate was captured on sequential 3.0  $\mu$ m, 0.8  $\mu$ m and 0.1  $\mu$ m polyethersulfone membrane filters (Supor membrane disc filter; Pall Life Sciences), and immediately stored at -20 °C (Rusch *et al.*, 2007; Ng *et al.*, 2010).

<sup>&</sup>lt;sup>1</sup>Sampling was performed by Jeffrey M. Hoffman and Jeffrey B. Mcquaid

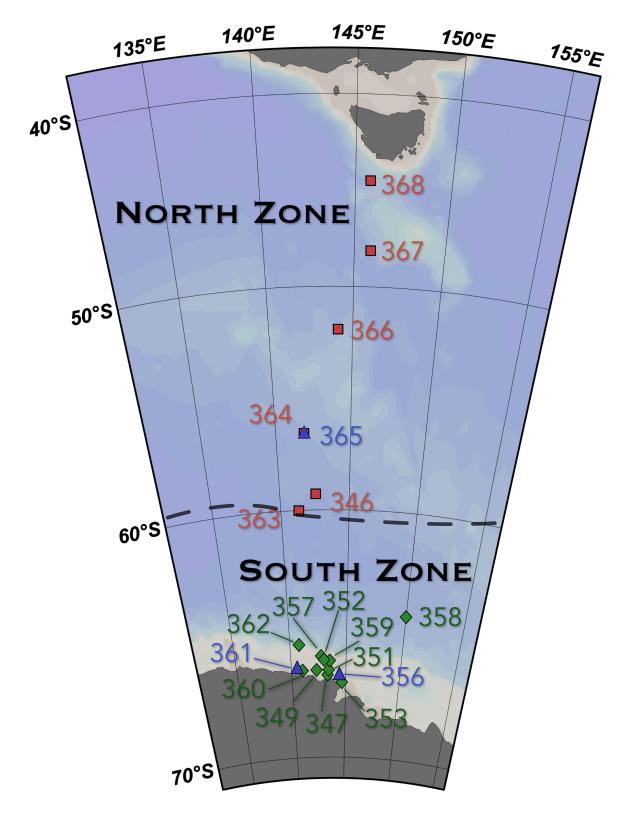


Figure 1: Sites of seawater samples used in this study. Red squares indicate surface samples from the North Zone; green diamonds samples from the South Zone; and blue triangles indicate deep samples. The dashed line gives the approximate location of the Polar Front.

Phylogenetic analysis of metagenomic data

Functional analysis of metagenomic data

#### **Results**

Metagenomic sequencing

Phylogenetic analysis of metagenomic data

Functional analysis of metagenomic data

#### Discussion

#### **Conclusions**

## Meso-scale biogeographic drivers of planktonic diversity

## **Conclusions**

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