

Death comes on two wings: a review of dipteran natural enemies of arachnids

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REVIEW

Death comes on two wings: a review of dipteran natural enemies of arachnids

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Abstract. Though the best known natural enemies of arachnids are Hymenoptera, Diptera also form an important group of arachnid enemies, attacking 31 spider families in all three suborders of Araneae, as well as members of the Acari, Amblypygi and Scorpiones. Some species of Bombyliidae, Chloropidae, Drosophilidae, Ephydriidae, Phoridae and Sarcophagidae are known to attack eggs of several families of arachnids, acting as predators, parasitoids and/or parasites of egg sacs. Alternatively, members of Acroceridae and Tachinidae are internal parasitoids, attacking juvenile and/or adult spiders. One species of Sarcophagidae is reported as a predator of individual Liphistiidae (Mesothelae) spiders. We summarize the available information on all lineages of Diptera known to attack arachnids, including predators, parasites, kleptoparasites and parasitoids. A table including host records pertaining to the aforementioned dipteran families is presented. Particular emphasis is given to Acroceridae, the only lineage of Diptera known to develop exclusively on arachnids, and one of the most significant groups of natural enemies of spiders.

Keywords: Araneae, parasitoid, spider egg sacs, scorpions, amblypygids

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1. INTRODUCTION

Some of the best known natural enemies of arachnids belong to the order Hymenoptera, with species either feeding on the individual arachnid, or on its eggs. Several groups of Hymenoptera develop on arachnid hosts, including wasps in the families Diapriidae, Eulophidae, Eupelmidae, Eurytomidae, Ichneumonidae, Platygastriidae sensu lato, Pompilidae, Pteromalidae and Sphecidae (Fitton et al. 1987; Noyes 2016). Hymenopteran parasitoids and predators exhibit a breadth of life strategies, including species that feed in cocooned spider egg masses, endoparasitoids that develop individually within eggs, external koinobiont parasitoids of mobile spiders, and idiobionts that paralyze one or more spiders as prey (Austin 1985; Gauld & Dubois 2006).

Species in the order Diptera are also significant natural enemies of arachnids. As opposed to the large taxonomic

diversity of hymenopteran arachnid enemies, dipteran parasites, parasitoids and predators of arachnids are restricted to fewer families and vary considerably in their mode of action. All species of Acroceridae and a single species of Tachinidae are internal parasitoids of juvenile or adult spiders, while a few members of the Chloropidae, Drosophilidae, Ephydriidae, Phoridae and Sarcophagidae are known to attack egg sacs of several arachnids, acting as parasitoids and/or predators of arachnid eggs (Clausen 1940; Vincent 1985; Schlinger 1987; Marshall 2012). One species of Sarcophagidae is a predator of Mesothelae spiders, attacking both adult and immature prey (Schwendinger & Pape 2000).

Although a clear cut distinction between predator and parasite is not always possible in nature, the three extremes of the spectrum (i.e., true parasite vs. true predator vs. true parasitoid) can be easily distinguished. Predators are organ-

isms that consume one or multiple prey during their lifetime, thus establishing a relationship that positively affects the predator and negatively affects the prey. Parasites, in general, live on/in the body of the organism they feed upon, and in contrast to predators, only take resources from one host, which they tend not to kill. Parasitoids, like parasites, only attack one host, but they enter or attach to their host, feed upon it, and ultimately kill it. Thus, the essential difference between parasites and parasitoids is that parasitoids always kill their hosts, while parasites tend to not kill their hosts. Additionally, the main difference between predators and parasitoids relates to the number of individuals attacked; predators usually kill multiple prey during their lifetime, while parasitoids only kill one host (Price 1980).

Parasitoids are found in five orders of holometabolous insects: Hymenoptera, Diptera, Coleoptera, Lepidoptera, and Neuroptera. Hymenopteran parasitoids account for nearly 78% of the estimated number of species, and consequently have served as models for nearly all recent research on insect parasitoids (Feener & Brown 1997). However, parasitoids in the Hymenoptera represent a single evolutionary lineage in contrast to the hundreds of parasitoid lineages in Diptera, Coleoptera, and other orders (Eggleton & Belshaw 1992). There are over 16,000 species of described dipteran parasitoids, which are distributed in 21 families and represent about 20% of the known parasitoid diversity (Eggleton & Belshaw 1992). Dipteran parasitoids are hypothesized to be derived from over 100 independent lineages, thus offering an unparalleled system for understanding the origin, evolution and diversification of parasitoid life history.

In this review, we summarize the available information on all lineages of Diptera that are known to attack arachnids, including predators, parasites, kleptoparasites and parasitoids. A table summarizing the egg parasitoids and/or predators, as well as internal parasitoids of adults and immature arachnid hosts is presented. Predators and kleptoparasites of individual arachnids were not included in the table because they represent opportunistic rather than adaptive interactions. All species and genus names given in the literature were checked and updated using current dipteran and arachnid taxonomy (Pape & Thompson (2013) and World Spider Catalog (2016) respectively).

2. DIPTERAN NATURAL ENEMIES OF ARACHNIDS

2.1 Predators.—Species of Asilidae are among the best known dipteran predators, but it has been indicated that arachnids do not comprise a significant portion of robber fly diet, making up less than 2% of all their consumed prey (Dennis et al. 2012). Robber flies are found worldwide and may occasionally take spiders as prey, especially species in the asilid genera *Daspletis* Loew, 1858, *Efferia* Coquillett, 1893, *Euscelidia* Westwood, 1850, *Holopogon* Loew, 1847, *Laphystia* Loew, 1847, *Psilonyx* Aldrich, 1923 and *Stichopogon* Loew, 1847 (Dennis et al. 2012). Robber flies are recorded to prey on spiders belonging to the suborder Araneomorphae, mainly in the families Agelenidae, Araneidae, Clubionidae, Lycosidae, Salticidae and Theridiidae, among others (Dennis et al. 2012). This suborder comprises species that are generally readily visible, including orbweavers, crab spiders and jumping spiders. In contrast, spiders in the other two suborders of Araneae (Mygalomorphae and Mesothelae) generally have reclusive habits (such as

trapdoor and funnel-web spiders) or are highly mobile. There are also smaller dipterans that play a role as predators of small arachnids. For instance, larvae of species in the gall midge genus *Feltiella* Rubsaamen, 1910 (Cecidomyiidae) are specialized predators of all spider mite life stages (Acarina: Tetranychidae), as are many members of the gall midge tribe Lestodiplosini (Gagné 1995). One species of Ceratopogonidae, *Forcipomyia araneivora* Clastrier & Legrand, 1991, also plays a role as an enemy of spiders by feeding directly on their haemolymph (Clastrier & Legrand 1991). Other common dipteran predators (e.g., Muscidae and several families in Empidoidea) may take arachnids as prey, but no records of these were found.

2.2 Kleptoparasites.—Kleptoparasitism is a form of competition that involves the stealing of a portion of already acquired food items, and is one of the most common types of exploitation between animals. Kleptoparasites steal food that is either already in a predator's possession, or which the predator has already spent energy pursuing and capture by the predator is imminent. In this interaction, the kleptoparasite is benefited and the host may potentially be negatively affected by the loss of food resources (Brockmann & Barnard 1979; Barnard 1984). Some fly species in the families Cecidomyiidae, Ceratopogonidae, Chloropidae, Dolichopodidae, Lonchaeidae, Milichiidae and Phoridae are known kleptoparasites of spiders. These organisms typically exploit cadavers captured by spiders, feeding on the semi-digested prey. In general, spiders most commonly attacked by kleptoparasites are orbweavers (Araneidae and Nephilidae), crab spiders (Thomisidae) and jumping spiders (Salticidae). The most common spider kleptoparasites are species in the genera *Didactylomyia* Felt, 1911 (Cecidomyiidae), *Microphor* Macquart, 1827 (Dolichopodidae), *Desmometopa* Loew, 1866, *Neophyllomyza* Melander, 1913, *Paramyia* Williston, 1897, and *Phyllomyza* Fallen, 1810 (Milichiidae) and *Megaselia* Rondani, 1856 (Phoridae) (Robinson & Robinson 1977; Sivinski & Stowe 1980; Weinmann & Disney 1997; Sivinski et al. 1999; Brake 2000; Brake & von Tschirnhaus 2010; van Helsdingen 2011; Marshall et al. 2015).

Due to their generalistic feeding nature (taking a range of taxa as prey/host), predators and kleptoparasites of individual arachnids were not included in the summary table.

2.3 Egg parasitoids and predators.—A large variety of Diptera attack the eggs of arachnids, either as parasitoids or predators; it is often difficult to distinguish between these alternatives due to the limited number of natural history and rearing observations. Therefore, we have decided not to distinguish between these modes, in order to avoid making incorrect assumptions about poorly known dipteran-spider interactions. Each dipteran family is discussed in more detail below.

2.3.1 Family Bombyliidae: Bombyliidae is a large group of Diptera commonly known as bee flies. Despite the great number of species (over 5,000 valid species), the habits of most immature stages (>80%) are still poorly understood (Yeates & Greathead 1997). Known species are mostly ecto- or endoparasitoids on the larvae and/or pupae of other insects, mainly in the Lepidoptera, Hymenoptera, Coleoptera, Diptera and Neuroptera. The only known exception is *Petrorossia feti* Zaitsev & Charykuliev, 1981, which develops as a parasitoid or predator on egg sacs of oecobiid spiders (Zaitsev & Charykuliev 1981; Yeates & Greathead 1997). It is unknown

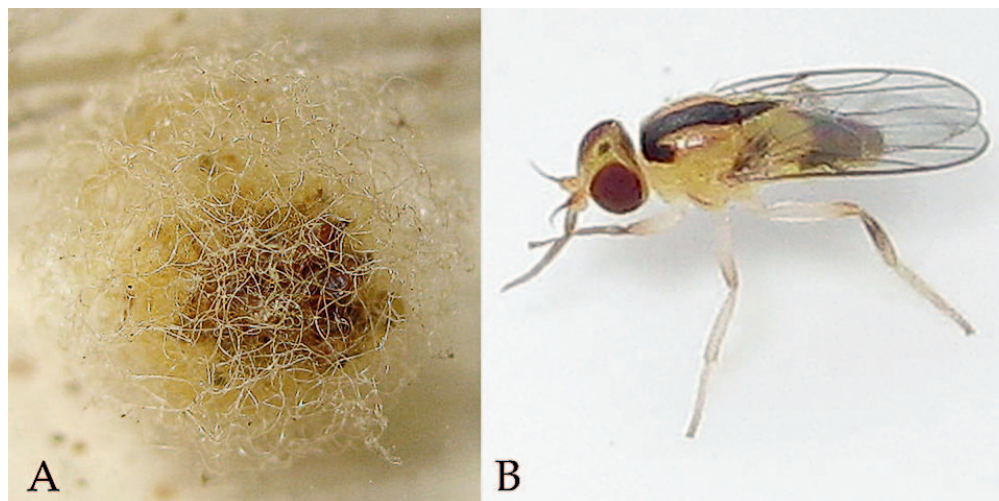


Figure 1.—Chloropidae development on spider egg sac. A. Egg mass of *Mimetus puritanus* Chamberlin, 1923 containing larvae of *Pseudogaurax anchora* (Loew, 1866). B. Adult *P. anchora*. Photos: John R. Maxwell.

whether or not this species has a specific host association with these spiders (Zaitzev & Charykuliev 1981).

2.3.2 Family Chloropidae: Several species of Chloropidae are known to be either parasitoids or predators of arachnid eggs. Genera of chloropids that have been reported as arachnid egg predators include *Conioscinella* Duda, 1929, *Gaurax* Loew, 1863, *Oscinella* Becker, 1909, *Oscinisoma* Lioy, 1864, *Pseudogaurax* Malloch, 1915, *Siphonella* Macquart, 1835, and *Tricimba* Lioy, 1864 (Barnes et al. 1992). Eason et al. (1967), Rollard (1984) and Barnes et al. (1992) provide a review of host records for several groups of Diptera and Hymenoptera, including chloropids. Several species of *Gaurax* and *Pseudogaurax* are suggested to be parasitoids of arachnid eggs (including amblypygids (Viquez & de Armas 2009; Chapin & Hebet 2016)) and have been raised from a variety of host eggs (see details in Table I) (Figs. 1 & 2). However, very little is known about the life histories of most of these species, and there is an indication that the larvae do not complete their development on a single egg, consuming a few or several eggs and, as such, must be considered as predators (Barnes et al. 1992). Some species are apparently opportunists, utilizing a variety of host species or even hosts of different orders, including lepidopteran cocoons and mantid oothecae. *Pseudogaurax signatus* (Loew, 1876) has been suggested as a biocontrol agent of black widow spiders, *Latrodectus* Walckenaer, 1805, since many specimens have been reared from egg sacs of this spider (Barnes et al. 1992; Vetter et al. 2012), with infestation rates reported to range from 6% (Vetter et al. 2012) to 40%, and estimated spider mortality >90% (Barnes et al. 1992). Female *P. signatus* flies lay the eggs on the surface of the host egg sac. Once hatched, larvae enter the egg sacs and consume multiple eggs, developing inside the sac.

2.3.3 Family Drosophilidae: Most species of Drosophilidae feed on decaying fruit and fungal material, as well as on fresh sap and nectar from flowers. Species in the subgenus *Titanochaeta* Knab, 1914 (genus *Scaptomyza* Hardy, 1850), however, differ from all other drosophilids in being parasitoids (or predators) of spider eggs. Unlike the remaining species of *Scaptomyza*, species in the subgenus *Titanochaeta* exhibit a

slender, sharply pointed, stylet-like ovipositor, which is likely an adaptation to a lifestyle as a spider egg sac parasitoid or predator (O'Grady et al. 2003). The group is endemic to Hawaii and comprises 11 species known to develop on spider egg sacs (Wirth 1952; Eason et al. 1967; O'Grady et al. 2003). Little biological information is available on the host usage of species of *Titanochaeta*, but available rearing data indicate a preference for spiders in the family Thomisidae (Hardy 1965; Lapoint et al. 2013). Further study is needed in order to investigate the number of eggs consumed by each individual, thus confirming whether species of *Titanochaeta* are predators or parasitoids of spider eggs.

2.3.4 Family Ephydriidae: The vast majority of species of Ephydriidae feed primarily on autotrophic microorganisms such as algae (Foote 1984). One exception is *Trimerina madizans* (Fallen, 1813), which has been indicated as an egg parasitoid of marsh-inhabiting spiders (Wirth et al. 1987). The fly has been reported to attack egg sacs of the linyphiid *Hypselistes florens* (O. Pickard-Cambridge, 1875) (Wirth et al. 1987). However, it has been shown that each fly larva consumes on average six eggs of *H. florens* during its development (Foote 1984); in this case, the species is better regarded as a predator rather than a parasitoid.

2.3.5 Family Phoridae: Although most species of Phoridae associated with arachnids are merely saprophagous, some are known to be associated with living rather than dead arachnids. Most of these are predators of arachnid eggs. However, specimens of *Apocephalus borealis* Brues, 1924 have been reared from black widow egg sacs, *Latrodectus mactans* (Fabricius, 1775), (Araneae: Theridiidae), and this has been suggested to be a case of true parasitism, although further research is needed for proper confirmation (Disney 1994). Several species in *Megaselia* Rondani, 1856 and *Phalacrotophora* Enderlein, 1912 (Fig. 3) are known to prey on eggs of spiders in the families Araneidae (*Argiope* Audouin, 1826, *Gasteracantha* Sundevall, 1833, and *Larinioides* Caporiacco, 1934), Linyphiidae (*Pityohyphantes* Simon, 1929), Salticidae (*Phidippus* C. L. Koch, 1846), Tetragnathidae (*Meta* C. L. Koch, 1836) and Theridiidae (*Enoplognatha* Pavesi, 1880,



Figure 2.—Amblypigid female carrying pupae of *Pseudogaurax* (Diptera: Chloropidae) that were egg parasitoids of her egg case as larvae. Photographed at the Soltis Center, San Juan de Peñas Blancas, San Ramón, Costa Rica, by Kristie Reddick and Jessica Honaker.

Latrodectus and *Robertus* O. Pickard-Cambridge, 1879) (Disney & Evans 1980; Disney 1982, 1994, 1999).

2.3.6 Family Sarcophagidae: Sarcophagidae is a large family of Diptera with species exhibiting a breadth of habits and life histories. Most species are generalist scavengers and insect predators, while some species are kleptoparasites of solitary bees and wasps (Pape et al. 2012). Several species in the genus *Sarcophaga* Meigen, 1826 are spider egg parasitoids or predators (e.g., Eason et al. 1967; Auten 1925; Prakash & Pandian 1977; Austin 1985; Souza Lopes 1985; Cantrell 1986; Hieber & Uetz 1990). However, as in the dipteran families mentioned above, more research is needed in order to confirm whether these species are parasitoids or predators. Schwendinger & Pape (2000) reported an unusual case of a predatory species of *Metopia* Meigen, 1803, a genus that mostly comprises kleptoparasites in nests of solitary aculeate Hymenoptera (Pape 1986). Larvae of *Metopia sinensis* Pape, 1986 apparently kill the spider host and complete most of their larval life on the carcass, behaving more like predators rather than parasites (following Price 1980).

2.3.7 Family Tachinidae: As discussed below, tachinids are well known as parasitoids of a range of insects. However, there is only a single record of a tachinid attacking spider eggs, a specimen of *Tachina* Meigen, 1803 which had been reared from egg sacs of the araneid *Larinioides cornutus* (Clerck, 1757) (Bertkau 1880).

2.4 Endoparasitoids.—The majority of dipteran endoparasitoids of arachnids are species of the family Acroceridae, which have exclusively been reared from a variety of spider families. There are also a few examples from other Diptera families (Phoridae, Sarcophagidae and Tachinidae). The specific details of the behaviors in each case are given below.

2.4.1 Family Acroceridae: Species of Acroceridae are commonly called spider flies due to their tight relationship with spiders as internal parasitoids. They are sometimes also referred to as small-headed flies in reference to the disproportionately small heads of some species. Spider flies represent a morphologically heterogeneous assemblage of lineages, currently classified into three subfamilies (Acrocerinae, Panopinae and Philopotinae), 55 genera and approximately 530

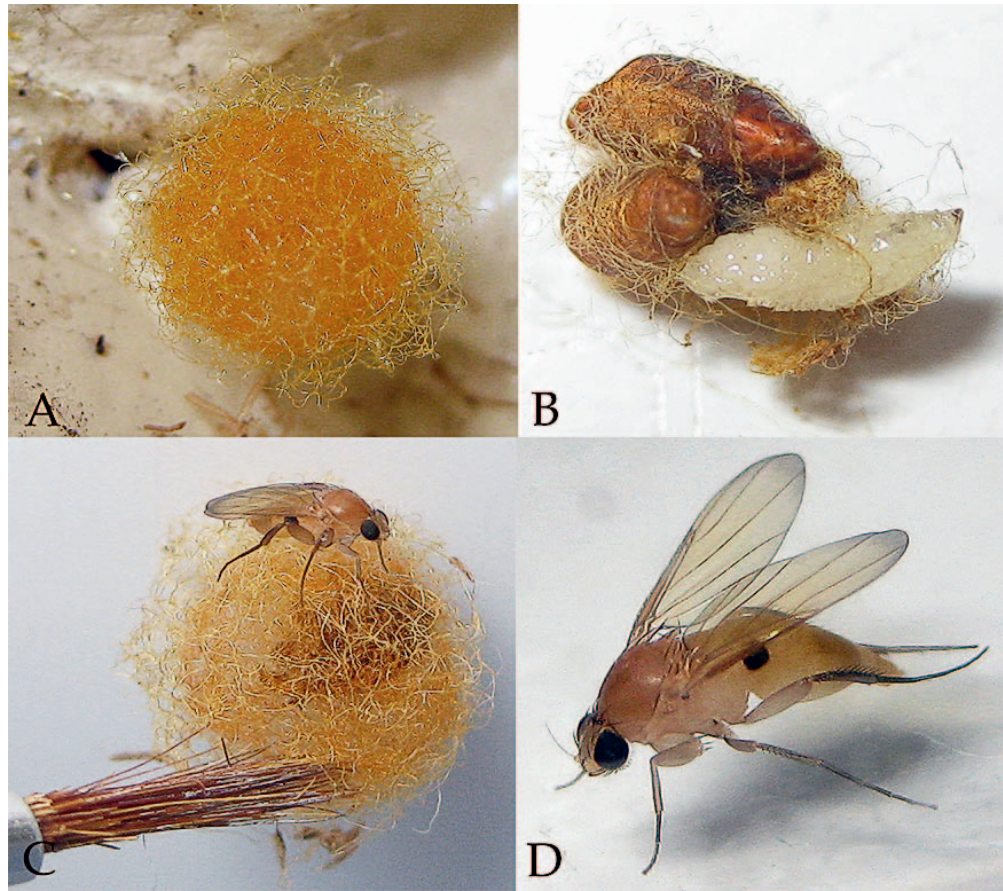


Figure 3.—Phoridae development on spider egg sac. A. Egg mass of *Mimetus puritanus* Chamberlin, 1923. B. Pupae and larva of *Phalacrotophora epeirae* (Brues, 1902). C. Spider egg mass, pupae and adult of *P. epeirae*. D. Adult *P. epeirae*, oblique view. Photos: John R. Maxwell.

species (Schlinger et al. 2013). Adults are morphologically very distinctive, usually with inflated or hunchbacked bodies, and occasionally with metallic coloration. Several species have long, modified mouthparts for nectar feeding and are considered important pollinators (e.g., Goldblatt et al. 1997; Potgieter et al. 1999; Pujol-Luz 2004; Carvalho & Machado 2006; Borkent & Schlinger 2008).

Spider fly larvae attack spiders in the Mygalomorphae (Fig. 4) and Araneomorphae. The only exception to the exclusive endoparasitic mode in Acroceridae is found in the Chilean genus *Carvalhoa* Koçak & Kemal, 2013 (= *Sphaerops* Philippi, 1865), which is reported to remain ectoparasitic on its host spider for at least three weeks (Schlinger 1987). It is still unknown whether the entire lifecycle is spent as an ectoparasite, but Schlinger (1987) reports this ectoparasitic behavior in multiple rearings of later instars of the species.

Records of acrocerids parasitizing Acari are extremely rare (Sferra 1986; Kerr & Winterton 2008), and no spider fly planidium has been reared from living mites to confirm, or identify, the acrocerid species in question. It is likely that the presence of acrocerid first instar larvae in mites is merely accidental parasitism, and the larvae probably do not develop further due to the small size of the mites relative to known acrocerid adults. Acrocerids are known to attack juvenile spiders, but even the smallest known spider hosts are several

times larger than mites found in association with acrocerid planidia (Sferra 1986; Kerr & Winterton 2008). In the cases of mite parasitism, it is likely that the planidium is simply indiscriminately adhering to arachnids in an attempt to find a suitable host. In this case, the association between Acroceridae and Acari would not represent a true host-parasitoid relationship.

Generally, adult spider fly females scatter eggs during flight or oviposit large numbers of microtype eggs on twigs, branches or foliage (Schlinger 1987). In most species, oviposition seems to be entirely independent of the presence of a host, though naturally the flies occur in a habitat favorable to spider populations. Females of *Eulonchus* Gerstaecker, 1856 are reported to be attracted to burrows of trapdoor spiders in the genus *Antrodiaetus* Ausserer, 1871. Coyle (1971) reports a case in which, as trapdoor spider burrows were being excavated, a large number of spider flies quickly approached, hovering close to the ground and landing near closed burrow entrances, apparently attracted by some chemical released during the excavation process.

The first-instar acrocerid is a free-living planidium, which actively seeks out a spider host by crawling, looping or jumping, with the aid of well-developed setae, spines and a caudal suction disk (Schlinger 1981). Planidia in the genus *Acrocera* Meigen, 1803, however, differ from all other spider

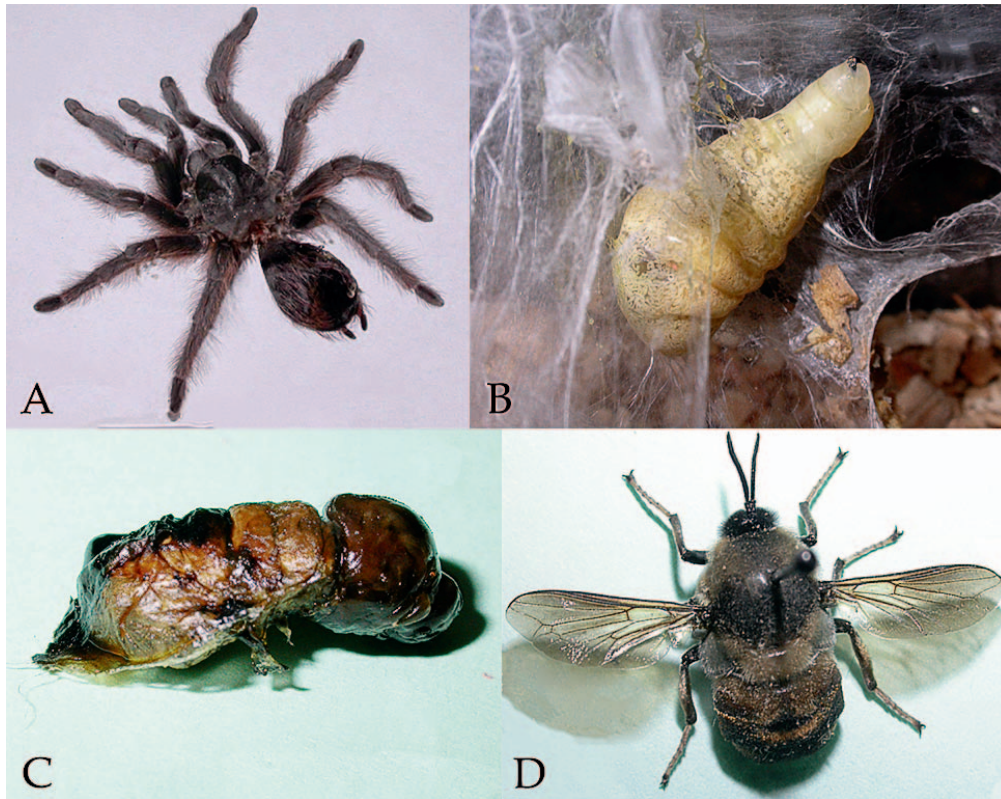


Figure 4.—Acroceridae development in the spider host. A. Spider host, *Aphonopelma duplex* (Chamberlin, 1925). B. Fourth instar larva of *Ocnaea* sp. C. Pupa of *Ocnaea* sp. D. Adult *Ocnaea* sp. Photos: Dr. A. Alagon.

flies in having a poorly sclerotized body and an abdomen with numerous annulations obscuring the regular body segmentation, as well as the absence of setal pile characteristic of acrocerid planidia (Schlinger 1981). *Acrocera* also exhibits a unique mode of entering its host, in which the first-instar larva injects itself into the spider. Planidia of *Acrocera orbicula* (Fabricius, 1787) have been observed entering a wolf spider through this sophisticated mode, in which first-instars firmly attach themselves to the host's cuticle by the mouthparts, presumably feeding externally for about a week (Overgaard Nielsen et al. 1999; Toft et al. 2012). Rather than entering the spider, the first-instar subsequently molts and the small, flexible, and glabrous second-instar is injected directly into the spider via the wound, leaving the exuvia of the ectoparasitic first-instar covering the site of infection. This mode of host invasion may reduce physical damage to the host in the initial phase of endoparasitism, thus enhancing parasitoid survival (Overgaard Nielsen et al. 1999). More research is needed in order to investigate whether this mechanism for infection is also present in other Acroceridae lineages.

Most host records for Acroceridae involve rearing of mature larvae from parasitized spiders, and observations of infestation of the host by the planidium stage are rare. The planidium presumably enters the host directly through the cuticle of the cephalothorax, opisthosoma or leg joints (Schlinger 1981, 1987; Nartshuk 1997), but these apparently have not been verified by direct observations. Acrocerid larvae eventually locate themselves in the opisthosoma and attach their posterior spiracle to the spider's book lung for

respiration (Schlinger 1981, 1987). Schlinger (1987) proposed that upon entering the host, the larva may enter a state of diapause that can last for several months (Acrocerinae) up to 10 years (Panopinae), and, upon cessation of diapause, the actively feeding larva completes its life cycle relatively quickly (days to weeks), undergoing up to four instars. McQueen (1983) observed wolf spiders in the genus *Geolycosa* Montgomery, 1904 parasitized by *Pterodontia* Gray, 1832 spider flies and stated that infected spiders carry the larvae for approximately a year until they are entirely consumed by the parasitoid. However, further evidence is needed to support the hypothesis of diapause within the host and to verify the actual number of larval instars in Acroceridae, because the typical number of instars in lower brachyceran flies is three.

Many parasites and parasitoids have evolved remarkable strategies to manipulate the behavior of their hosts in order to promote their own survival and reproduction. These behavioral manipulations may include alterations in phototaxis, locomotion, foraging and defensive behaviors (Moore 2002). Most spider hosts of acrocerids, however, do not exhibit any obvious external indication of parasitism, although there are reports of enlargement of the spider's opisthosoma due to the presence of spider fly larvae (e.g., Lamore 1960; Barraclough & Croucamp 1997). It has been repeatedly reported that the host displays some abnormal, agitated behavior during the final stages of parasitism, in which the spiders walk aimlessly and incessantly scratch the lateral portions of the opisthosoma with the legs, presumably where the parasitoid is located (Cady et al. 1993; Barneche et al. 2013). In some species,

especially in the Mygalomorphae, the emergence of the parasite seems to correspond with the premolting behavior of the spider (e.g., Montgomery 1903; Cady et al. 1993; Barneche et al. 2013). However, a causative relationship between the production of the premolting web and parasitoid emergence can only be speculative at this time, but the presence of hook-like processes on the head and/or abdomen of the spider fly larva, which are supposedly used as attachment to the web, suggests that this is not coincidental. Lamore (1960) reported an example in which a larva of *Ogcodes dispar* (Macquart, 1855) parasitizing a basilica spider, *Mecynogea lemniscata* (Walckenaer, 1841), hung itself on the spider web while also hanging on to the spider on one end for a while, then released its hold on the exoskeleton in order to pupate. The last instar acrocerid larva typically only kills the host shortly prior to emergence, when it consumes the entire contents of the host body, leaving an empty, unbroken exoskeleton (Schlinger 1987; Nartshuk 1997).

While super-parasitism is common in early instars, usually only a single acrocerid adult ultimately emerges from its host (Cady et al. 1993; Overgaard Nielsen et al. 1999). Multiple emergences of larvae from a single host are more likely found in Panopinae, which attack large Mygalomorphae spiders capable of sustaining multiple parasitoids (Schlinger 1987; Cady et al. 1993).

Host records for acrocerids parasitizing spiders are known for at least 60 species, recorded from about 25 spider families (Schlinger 1987). Schlinger (1987, table 24) presented an extensive list of spider taxa parasitized by acrocerids, while Winterton et al. (2007) mapped host use onto a DNA-based phylogeny of the family. These studies clearly demonstrate that Panopinae are host-specific to Mygalomorphae spiders (Fig. 4), while Acrocerinae and Philopotinae are host-specific to the Araneomorphae. Only the genera *Acrocera* and *Carvalhoa* have been reared from Haplogynae spiders, while Philopotinae and all remaining Acrocerinae have only been recorded to attack Entelegynae. The cosmopolitan genera *Acrocera*, *Ogcodes* Latreille, 1797 and *Pterodontia* have been reared from numerous hosts in multiple spider families, but most geographically restricted and species-poor genera tend to be more host-specific, generally attacking only a single spider family (Schlinger 1987). Strict host specificity between particular acrocerid and spider species seems to be rare, but some trends are evident where host preference generally follows spider guilds instead of lineages (Schlinger 1987; Cady et al. 1993). Spiders most susceptible to parasitism by Acroceridae belong to guilds of cursorial or fossorial species (e.g., Mygalomorphae: Antrodiaetidae, Ctenizidae, Migidae, Theraphosidae; Araneomorphae: Anyphaenidae, Clubionidae, Lycosidae, Salticidae, Thomisidae), or those that occupy sac, tangle or funnel-like web retreats that are close to the ground, have webs with many connections to vegetation, or visit surrounding vegetation or substrate frequently (e.g., Agelenidae, Amaurobiidae, some Araneidae, Dipluridae, Segestriidae) (Schlinger 1987; Cady et al. 1993; Overgaard Nielsen et al. 1999). There are very few records of acrocerids parasitizing true web-dwelling spiders and exceptions typically involve comb-footed spiders (e.g., Theridiidae), where the spider may still be proximal to the substrate (e.g., Lamore 1960).

2.4.2 Family Sarcophagidae: As discussed above, the majority of members of Sarcophagidae are generalist scavengers and insect predators (Pape et al. 2012). However, there is one example of a sarcophagid acting as an endoparasitoid of a scorpion. *Sarcophaga dux* (Thomson, 1869), a species generally considered to be of forensic importance, was found to attack the Chinese scorpion *Mesobuthus martensii* (Karsch, 1879) (Scorpiones: Buthidae). Multiple larvae (3–6) attacked more than 50 individuals of this scorpion species, with more than 100 flies emerging as adults by the end of the study (Shi et al. 2015).

2.4.3 Family Tachinidae: Species of Tachinidae are internal parasitoids on a wide range of arthropod hosts. The most commonly used hosts are phytophagous insects, primarily Lepidoptera, Coleoptera, Hymenoptera, Heteroptera, and Orthoptera. Several genera of tachinids attack non-insect arthropods, including centipedes and scorpions (Stireman et al. 2006). There is a single record of a spider serving as a tachinid host (Vincent 1985). Two tachinid larvae in the genus *Lypha* Robineau-Desvoidy, 1830 emerged from the abdomen of immature individuals of *Antrodiaetus riversi* (O. Pickard-Cambridge, 1883) in the laboratory. The two larvae pupated outside of the host but neither adult emerged. Other known hosts of *Lypha* species are Lepidoptera in the families Gelechiidae and Tortricidae, and it is likely that the two spiders may have been accidental hosts that did not manage to survive and become adults. Vincent (1985) reported rearing over 340 spiders, which were examined and inspected for signs of parasitism, and only two contained larval tachinids.

The tachinid *Spilochaetosoma californicum* Smith, 1917 has been found to be an endoparasitoid of two Nearctic scorpions, *Anuroctonus phaiodactylus* (Wood, 1863) and *Paravaejovis spinigerus* (Wood, 1863) (Scorpiones: Vaejovidae). Multiple larvae of *S. californicum* emerged from wild caught individuals of both species and it is likely that this tachinid is a general parasitoid of burrowing scorpions (Williams et al. 1994).

3. CONCLUSION

In this review we summarized the available information on all lineages of Diptera known to attack arachnids, including predators, parasites, kleptoparasites and parasitoids. A summary table (Table 1) containing over 200 host records for eight families of Diptera attacking 36 arachnid families is included. Even though hymenopterans are among the best known natural enemies of arachnids, species of Diptera clearly also comprise a large component of arachnid enemies, attacking families in four orders of arachnids and all three suborders of Araneae.

A single species of Sarcophagidae, one species of Tachinidae and all species of Acroceridae are internal parasitoids, attacking juvenile and/or adult arachnids. Spider flies (family Acroceridae) comprise the only lineage of Diptera known to develop exclusively on arachnid hosts, representing some of the most significant natural enemies of spiders. Multiple species of Chloropidae, Drosophilidae, Ephydriidae, Phoridae and Sarcophagidae are known to attack eggs of 11 families of arachnids, acting as predators and/or parasitoids of arachnid egg sacs (see Table 1). Some of these species are known to be true predators, while some are known to be true parasitoids. However, in most cases, the life history strategy is not clear,

Table 1.—Host records for the known natural enemies of arachnids in the order Diptera, organized by arachnid order and family. Due to their generalist feeding habits, predators and kleptoparasites of individual arachnids were not included. Host stage exploited by the dipteran is noted as egg, adult, immature or stage unknown (Adult/Immature). EIS = E.I. Schlinger Collection database record.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|----------------|--|-----------------------|-------------|--|-----------------------------|
| Amblypygi | Phrynidae | <i>Paraphrynus laevifrons</i> (Pocock, 1894) | Adult | Chloropidae | <i>Pseudogaurax</i> sp. | Viquez & De Armas 2009 |
| | | <i>Phrynus</i> <i>pseudoparvulus</i> Armas & Viquez, 2001 | Adult | Chloropidae | <i>Pseudogaurax</i> sp. | Viquez & De Armas 2009 |
| Araneae | Agelenidae | <i>Agelenopsis naevia</i> (Walckenaer, 1841) | Adult/ Immature | Acroceridae | <i>Turbopsebius</i> <i>sulphuripes</i> (Loew) | Melander 1902 |
| | | <i>Agelenopsis</i> <i>oregonensis</i> Chamberlin & Ivie, 1935 | Adult/ Immature | Acroceridae | <i>Acrocera bakeri</i> Coquillett | Schlinger 1987 |
| | | <i>Agelenopsis</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera melanderi</i> Cole | EIS database |
| | | <i>Agelenopsis</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes dispar</i> (Macquart) | Schlinger 1987 |
| | | <i>Agelenopsis</i> sp. | Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Guarisco 1990 |
| | | <i>Barronopsis</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera bimaculata</i> (Loew) | Schlinger 1987 |
| | | <i>Coras montanus</i> (Emerton, 1890) | Adult/ Immature | Acroceridae | <i>Acrocera bimaculata</i> (Loew) | Cady et al. 1993 |
| | | <i>Coras montanus</i> | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | Sabrosky 1948 |
| | | <i>Coras montanus</i> | Adult/ Immature | Acroceridae | <i>Turbopsebius</i> <i>sulphuripes</i> (Loew) | Cady et al. 1993 |
| | | <i>Eratigena sicana</i> (Brignoli, 1976) | Adult/ Immature | Acroceridae | <i>Ogcodes</i> sp. | Brignoli 1976 |
| | | <i>Hololena curta</i> (McCook, 1894) | Adult/ Immature | Acroceridae | <i>Acrocera subfasciata</i> Westwood | EIS database |
| | | <i>Hololena curta</i> | Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1960 |
| | | <i>Hololena curta</i> | Adult and immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1960 |
| | | <i>Hololena curta</i> | Immature | Acroceridae | <i>Turbopsebius diligens</i> (Osten Sacken) | Schlinger 1952 |
| | | <i>Hololena</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera melanderi</i> Cole | Cole 1969 |
| | | <i>Hololena</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera subfasciata</i> Westwood | Schlinger 1987 |
| | | <i>Rualena</i> sp. | Adult/ Immature | Acroceridae | <i>Turbopsebius diligens</i> (Osten Sacken) | Schlinger 1987 |
| | | <i>Textrix denticulata</i> (Olivier, 1769) | Adult/ Immature | Acroceridae | <i>Acrocera sanguinea</i> Meigen | Koch 1872 |
| Araneae | Amaurobiidae | <i>Amaurobius erberi</i> (Keyserling, 1893) | Immature | Acroceridae | <i>Acrocera orbicula</i> (Fabricius) | Kehlmaier & Almeida 2014 |
| | | <i>Amaurobius</i> sp. | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1960 |
| | | <i>Callobius bennetti</i> (Blackwall, 1846) | Adult/ Immature | Acroceridae | <i>Acrocera fasciata</i> Wiedemann | Emerton 1890 |
| | | Undetermined genus | Adult/ Immature | Acroceridae | <i>Megalybus pictus</i> Philippi | Schlinger 1987 |
| | | Undetermined genus | Adult/ Immature | Acroceridae | <i>Thyllis</i> sp. | Schlinger 2003 |
| Araneae | Amphinectidae | <i>Metaltella</i> sp. | Adult/ Immature | Acroceridae | <i>Holops cyaneus</i> Philippi | Schlinger 1987 |
| Araneae | Antrodiaetidae | <i>Aliatypus californicus</i> (Banks, 1896) | Adult/ Immature | Acroceridae | <i>Eulonchus tristis</i> Loew | Coyle & Icenogle 1994 |
| | | <i>Aliatypus erebus</i> Coyle, 1974 | Adult | Acroceridae | <i>Eulonchus tristis</i> Loew | Coyle & Icenogle 1994 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|----------------|---|--------------------|---------------|--|---------------------------------|
| Araneae | Antrodiaetidae | <i>Aliatypus</i> sp. | Adult/ Immature | Acroceridae | <i>Eulonchus tristis</i> Loew | Schlinger 1987 |
| | | <i>Antrodiaetus riversi</i> (O. Pickard- Cambridge, 1883) | Adult/ Immature | Acroceridae | <i>Eulonchus sapphirinus</i> Osten Sacken | EIS database |
| | | <i>Antrodiaetus riversi</i> | Adult/ Immature | Acroceridae | <i>Eulonchus</i> sp. | Vincent 1986 |
| | | <i>Antrodiaetus riversi</i> | Adult | Tachinidae | <i>Lypha</i> sp. | Vincent 1985 |
| | | <i>Antrodiaetus unicolor</i> (Hentz, 1842) | Adult/ Immature | Acroceridae | <i>Eulonchus marialiciae</i> Brimley | Coyle 1971; Adler et al 1997 |
| | | <i>Antrodiaetus</i> sp. | Adult/ Immature | Acroceridae | <i>Eulonchus</i> sp. | Schlinger 1987 |
| Araneae | Anyphaenidae | <i>Anyphaena californica</i> (Banks, 1904) | Adult/ Immature | Acroceridae | <i>Ogcodes</i> sp. | Cady et al. 1993 |
| | | <i>Wulfilia saltabundus</i> (Hentz, 1847) | Adult/ Immature | Acroceridae | <i>Ogcodes borealis</i> Cole | Sabrosky 1948 |
| | | <i>Wulfilia saltabundus</i> | Adult/ Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Sabrosky 1948 |
| Araneae | Araneidae | <i>Alpaida veniliae</i> (Keyserling, 1865) | Egg | Chloropidae | <i>Pseudogaurax</i> <i>cingulatus</i> Sabrosky | Sabrosky 1966 |
| | | <i>Araneus angulatus</i> Clerck, 1757 | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Davidson 1896 |
| | | <i>Araneus ejusmodi</i> (Boesenberg & Strand, 1906) | Egg | Chloropidae | <i>Gaurax chiyokae</i> (Kanmiya) | Kanmiya 1972, 1983 |
| | | <i>Araneus gemma</i> (McCook, 1888) | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Pierce 1942 |
| | | <i>Argiope aemula</i> (Walckenaer, 1841) | Egg | Phoridae | <i>Megaselia araneivora</i> Goto | Goto 1985 |
| | | <i>Argiope argentata</i> (Fabricius, 1775) | Egg | Sarcophagidae | <i>Sarcophaga davidsonii</i> Coquillett | Davidson 1894 |
| | | <i>Argiope aurantia</i> Lucas, 1833 | Egg | Phoridae | <i>Megaselia</i> sp. | Kaston & Jenks 1937 |
| | | <i>Argiope aurantia</i> | Egg | Chloropidae | <i>Pseudogaurax anchora</i> (Loew) | Kaston & Jenks 1937 |
| | | <i>Argiope aurantia</i> | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Coquillett 1898 |
| | | <i>Argiope aurantia</i> | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Lockley & Young 1993 |
| | | <i>Argiope aurantia</i> | Egg | Sarcophagidae | <i>Sarcophaga litsingeri</i> (Shinonaga & Barrion) | Davidson 1896 |
| | | <i>Argiope catenulata</i> (Doleschall, 1859) | Egg | Sarcophagidae | <i>Sarcophaga litsingeri</i> (Shinonaga & Barrion) | Shinonaga & Barrion 1980 |
| | | <i>Argiope pulchella</i> Thorell, 1881 | Egg | Sarcophagidae | <i>Sarcophaga banksi</i> Senior-White | Prakash & Pandian 1977 |
| | | <i>Argiope trifasciata</i> (Forsskål, 1775) | Egg | Sarcophagidae | <i>Tricharaea</i> (<i>Sarcophagula</i>) Thomson | de Armas & Garcia 1986 |
| | | <i>Argiope</i> sp. | Egg | Phoridae | <i>Megaselia</i> <i>argiopephaga</i> Disney | Disney 1982 |
| | | <i>Cyrtophora</i> <i>moluccensis</i> (Doleschall, 1857) | Egg | Sarcophagidae | <i>Sarcophaga</i> <i>arachnivora</i> (Lopes) | Cantrell 1986 |
| | | <i>Cyrtophora</i> <i>moluccensis</i> | Egg | Sarcophagidae | <i>Sarcophaga</i> <i>cyrtophorae</i> (Cantrell) | Cantrell 1986 |
| | | <i>Cyrtophora</i> <i>moluccensis</i> | Egg | Sarcophagidae | <i>Sarcophaga reposita</i> (Lopes) | Cantrell 1986 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|--------------|--|--------------------|---------------|---|---------------------------------|
| Araneae | Araneidae | <i>Gasteracantha cancriformis</i> (Linnaeus, 1785) | Egg | Phoridae | <i>Phalacrotophora epeirae</i> (Brues) | Muma & Stone 1971 |
| | | <i>Gasteracantha cancriformis</i> | Egg | Chloropidae | <i>Pseudogaurax lancifer</i> (Coquillett) | Hall 1937 |
| | | <i>Larinioides cornutus</i> (Clerck, 1757) | Egg | Chloropidae | <i>Conioscinella frontella</i> (Fallen) | Krijger 1910 |
| | | <i>Larinioides cornutus</i> | Egg | Chloropidae | <i>Oscinella halterata</i> (Lamb) | Auten 1925 |
| | | <i>Larinioides cornutus</i> | Egg | Sarcophagidae | <i>Sarcophaga sexpunctata</i> (Fabricius) | Auten 1925 |
| | | <i>Larinioides cornutus</i> | Egg | Tachinidae | <i>Tachina</i> sp. | Bertkau 1880 |
| | | <i>Larinioides scolopetarius</i> (Clerck, 1757) | Egg | Chloropidae | <i>Oscinella halterata</i> (Lamb) | Auten 1925 |
| | | <i>Larinioides scolopetarius</i> | Egg | Phoridae | <i>Phalacrotophora epeirae</i> (Brues) | Brues 1902, 1903; Auten 1925 |
| | | <i>Larinioides scolopetarius</i> | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | King 1916 |
| | | <i>Larinioides scolopetarius</i> | Egg | Sarcophagidae | <i>Sarcophaga sexpunctata</i> (Fabricius) | Auten 1925 |
| | | <i>Mecynogea lemniscata</i> (Walckenaer, 1841) | Adult | Acroceridae | <i>Ogcodes dispar</i> Macquart | Lamore 1960 |
| | | <i>Metepeira atascadero</i> Piel, 2001 | Egg | Sarcophagidae | <i>Sarcophaga lindae</i> (Lopes) | Hieber & Uetz 1990 |
| | | <i>Metepeira incrassata</i> O. Pickard-Cambridge, 1903 | Egg | Sarcophagidae | <i>Sarcophaga lindae</i> (Lopes) | Hieber & Uetz 1990 |
| | | <i>Neoscona nautica</i> (L. Koch, 1875) | Egg | Chloropidae | <i>Gaurax chiyokae</i> (Kanmiya) | Kanmiya 1972, 1983 |
| | | <i>Ordgarius magnificus</i> (Rainbow, 1897) | Egg | Sarcophagidae | <i>Sarcophaga arachnivora</i> (Lopes) | Souza Lopes 1985 |
| | | <i>Singa nitidula</i> C. L. Koch, 1844 | Egg | Chloropidae | <i>Conioscinella frontella</i> (Fallen) | Vachon 1952 |
| | | <i>Zygiella x-notata</i> (Clerck, 1757) | Adult/ Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | Holl et al. 1983 |
| | | Undetermined genus | Egg | Phoridae | <i>Megaselia longifurca</i> (Lundbeck) | Disney 1999 |
| Araneae | Clubionidae | <i>Clubiona leucaspis</i> (Simon, 1932) | Immature | Acroceridae | <i>Ogcodes reginae</i> Trojan | Kehlmaier & Almeida 2014 |
| | | <i>Clubiona putris</i> nom. dub. C. L. Koch, 1839 | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Emerton 1890 |
| | | <i>Clubiona putris</i> nom. dub. | Adult/ Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Menge 1866 |
| | | <i>Clubiona</i> sp. | Immature | Acroceridae | <i>Acrocera orbicula</i> (Fabricius) | Millot 1938 |
| Araneae | Ctenizidae | <i>Bothriocyrtum californicum</i> (O. Pickard-Cambridge, 1874) | Adult/ Immature | Acroceridae | <i>Ocnaea smithi</i> Sabrosky | Jenks 1938, 1940 |
| | | <i>Cyrtocarenum cunicularium</i> (Olivier, 1811) | Adult/ Immature | Acroceridae | <i>Astomella hispaniae</i> Lamarck | Brauer 1869 |
| Araneae | Desidae | <i>Matachia ramulicola</i> Dalmás, 1917 | Adult/ Immature | Acroceridae | <i>Ogcodes brunneus</i> (Hutton) | Dumbleton 1940 |
| Araneae | Dipluridae | <i>Linothele cousini</i> (Simon, 1889) | Adult/ Immature | Acroceridae | <i>Lasia ecuadorensis</i> Bequaert | Schlinger 1987 |
| Araneae | Euctenizidae | <i>Aptostichus stanfordianus</i> Smith, 1908 | Adult/ Immature | Acroceridae | <i>Eulonchus smaragdinus</i> Gerstaecker | Schlinger 1987 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|--------------|---|--------------------|---------------|---|-----------------------------|
| Araneae | Eutichuridae | <i>Cheiracanthium punctatorium</i> (Villers, 1789) | Egg | Sarcophagidae | <i>Sarcophaga sexpunctata</i> (Fabricius) | Krehenwinkel et al. 2016 |
| | | <i>Cheiracanthium</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes croucampi</i> Barraclough | Barraclough & Croucamp 1997 |
| Araneae | Gnaphosidae | <i>Cheiracanthium</i> sp. | Immature | Acroceridae | <i>Ogcodes</i> sp. | Schlinger 2003 |
| | | <i>Herpyllus</i> sp. | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1960 |
| | | <i>Zelotes</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera melanderi</i> Cole | Schlinger 1987 |
| | | <i>Zelotes</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes gibbosus</i> (Linnaeus) | Nielsen 1932 |
| Araneae | Linyphiidae | <i>Hypselistes florens</i> (O. Pickard-Cambridge, 1875) | Egg | Ephydriidae | <i>Trimerina madizans</i> (Fallen) | Wirth et al. 1987 |
| | | <i>Pityohyphantes costatus</i> (Hentz, 1850) | Egg | Phoridae | <i>Phalacrotophora epeirae</i> (Brues) | Manuel 1984 |
| Araneae | Liphistiidae | <i>Liphistius lahu</i> Schwendinger, 1998 | Adult/ Immature | Sarcophagidae | <i>Metopia sinensis</i> Pape | Schwendinger & Pape 2000 |
| | | <i>Liphistius</i> sp. | Adult | Sarcophagidae | <i>Metopia sinensis</i> Pape | Schwendinger & Pape 2000 |
| Araneae | Lycosidae | <i>Alopecosa accentuata</i> (Latreille, 1817) | Adult/ Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Eason et al. 1967 |
| | | <i>Alopecosa barbipes</i> (Sundevall, 1833) | Adult | Acroceridae | <i>Ogcodes gibbosus</i> (Linnaeus) | Locket 1930 |
| | | <i>Alopecosa barbipes</i> | Adult/ Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Locket 1930 |
| | | <i>Alopecosa kochi</i> (Keyserling, 1877) | Immature | Acroceridae | <i>Ogcodes melampus</i> (Loew) | Schlinger 1960 |
| | | <i>Geolycosa domifex</i> (Hancock, 1899) | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | McQueen 1978, 1983 |
| | | <i>Lycosa godeffroyi</i> L. Koch, 1865 | Immature | Acroceridae | <i>Ogcodes basalis</i> (Walker) | Humphreys 1976 |
| | | <i>Lycosa godeffroyi</i> | Immature | Acroceridae | <i>Pterodontia melli</i> Erichson | Humphreys 1976 |
| | | <i>Pardosa alacris</i> (C. L. Koch, 1833) | Adult | Acroceridae | <i>Ogcodes gibbosus</i> (Linnaeus) | Langer 2005 |
| | | <i>Pardosa distincta</i> (Blackwall, 1846) | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Sabrosky 1948 |
| | | <i>Pardosa lapidicina</i> Emerton, 1885 | Adult/ Immature | Acroceridae | <i>Acrocera fasciata</i> Wiedemann | Eason 1966 |
| | | <i>Pardosa lapidicina</i> | Adult/ Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Eason 1966 |
| | | <i>Pardosa littoralis</i> Banks, 1846 | Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Kaston 1937 |
| | | <i>Pardosa littoralis</i> | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Kaston 1937 |
| | | <i>Pardosa lugubris</i> (Walckenaer, 1802) | Adult/ Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | de Jong et al. 2000 |
| | | <i>Pardosa milvina</i> (Hentz, 1844) | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Eason et al. 1967 |
| | | <i>Pardosa milvina</i> | Immature | Acroceridae | Undetermined genus | Allard & Robertson 2003 |
| | | <i>Pardosa prativaga</i> (L. Koch, 1870) | Immature | Acroceridae | <i>Acrocera orbicula</i> (Fabricius) | Toft et al. 2012 |
| | | <i>Pardosa pullata</i> (Clerck, 1757) | Adult/ Immature | Acroceridae | <i>Ogcodes gibbosus</i> (Linnaeus) | Duffey 2000 |
| | | <i>Pardosa pullata</i> | Adult/ Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Schlinger 1987 |
| | | <i>Pardosa pullata</i> | Adult/ Immature | Acroceridae | <i>Ogcodes rufobdominalis</i> Cole | Eason et al. 1967 |
| | | <i>Pardosa saxatilis</i> (Hentz, 1844) | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Kaston 1937 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|---------------|--|--------------------|---------------|--|----------------------------------|
| Araneae | Lycosidae | <i>Pardosa sternalis</i> (Thorell, 1877) | Adult/ Immature | Acroceridae | <i>Acrocera convexa</i> Cole | Schlinger 1987 |
| | | <i>Pardosa sternalis</i> | Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1960 |
| | | <i>Pardosa sternalis</i> | Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1960 |
| | | <i>Pardosa sternalis</i> | Adult/ Immature | Acroceridae | <i>Pterodontia misella</i> Osten Sacken | Schlinger 1987 |
| | | <i>Pardosa tuoba</i> Chamberlin, 1919 | Adult/ Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1987 |
| | | <i>Pardosa utahensis</i> Chamberlin, 1919 | Adult/ Immature | Acroceridae | <i>Ogcodes</i> <i>rufoabdominalis</i> Cole | Capelle 1966 |
| | | <i>Pardosa vancouveri</i> Emerton, 1917 | Adult/ Immature | Acroceridae | <i>Acrocera convexa</i> Cole | Schlinger 1987 |
| | | <i>Pirata sedentarius</i> Montgomery, 1904 | Adult/ Immature | Acroceridae | <i>Ogcodes dispar</i> (Macquart) | Eason et al. 1967 |
| | | <i>Schizocosa crassipes</i> (Walckenaer, 1837) | Adult/ Immature | Acroceridae | <i>Acrocera fasciata</i> Wiedemann | Montgomery 1903, Johnson 1915 |
| | | <i>Schizocosa ocreata</i> (Hentz, 1844) | Adult/ Immature | Acroceridae | <i>Acrocera fasciata</i> Wiedemann | Montgomery 1903, Johnson 1915 |
| | | <i>Schizocosa rovneri</i> Uetz & Dondale, 1979 | Adult/ Immature | Acroceridae | <i>Ogcodes borealis</i> Cole | Cady et al. 1993 |
| | | <i>Schizocosa rovneri</i> | Adult/ Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Cady et al. 1993 |
| | | <i>Tigrosa helluo</i> (Walckenaer, 1837) | Adult/ Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Eason et al. 1967 |
| | | <i>Trochosa hispanica</i> Simon, 1870 | Adult/ Immature | Acroceridae | <i>Ogcodes lautereri</i> Chvala | Canzoneri & Hansen 1996 |
| | | <i>Trochosa terricola</i> Thorell, 1856 | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | King 1916 |
| Araneae | Migidae | <i>Moggridgea crudeni</i> Hewitt, 1913 | Adult/ Immature | Acroceridae | <i>Astomella capensis</i> Schlinger | Barracough 1984 |
| Araneae | Mimetidae | <i>Mimetes notius</i> Chamberlin, 1923 | Egg | Phoridae | <i>Phalacrotophora</i> <i>epeirae</i> (Brues) | Guarisco 2001 |
| Araneae | Miturgidae | <i>Griswoldia</i> sp. | Immature | Acroceridae | <i>Thyllis crassa</i> (Fabricius) | Schlinger 1987 |
| | | <i>Griswoldia</i> sp. | Adult/ Immature | Acroceridae | <i>Thyllis</i> sp. | Schlinger 2003 |
| Araneae | Nephilidae | <i>Nephila clavipes</i> (Linnaeus, 1767) | Egg | Chloropidae | <i>Pseudogaurax higginsii</i> Sabrosky | Barnes et al. 1992 |
| | | <i>Nephila clavipes</i> | Egg | Chloropidae | <i>Pseudogaurax</i> <i>mexoculatus</i> Sabrosky | Barnes et al. 1992 |
| | | <i>Nephila inaurata</i> (Vinson, 1863) | Egg | Chloropidae | <i>Pseudogaurax coyleae</i> Cogan | Cogan 1977 |
| | | <i>Nephila pilipes</i> (Fabricius, 1793) | Egg | Chloropidae | <i>Pseudogaurax seguyi</i> (Sabrosky) | Sabrosky 1990 |
| Araneae | Oecobiidae | <i>Uroctea limbata</i> (C. L. Koch, 1843) | Egg | Bombyliidae | <i>Petrorossia feti</i> Zaitsev & Charykuliev | Zaitsev & Charykuliev 1981 |
| Araneae | Oxyopidae | <i>Oxyopes lineatus</i> Latreille, 1806 | Adult/ Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | Schlinger 1987 |
| | | <i>Oxyopes salticus</i> Hentz, 1845 | Adult/ Immature | Acroceridae | <i>Ogcodes dispar</i> (Macquart) | Eason et al. 1967 |
| | | <i>Oxyopes salticus</i> | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Eason et al. 1967 |
| Araneae | Philodromidae | <i>Philodromus aureolus</i> (Clerck, 1757) | Egg | Sarcophagidae | <i>Sarcophaga</i> <i>sexpunctata</i> (Fabricius) | Auten 1925 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|---------------|--|--------------------|---------------|--|-------------------------------|
| Araneae | Philodromidae | <i>Philodromus cespitum</i> (Walckenaer, 1802) | Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | Kehlmaier et al. 2012 |
| | | <i>Philodromus cespitum</i> | Egg | Chloropidae | <i>Oscinella halterata</i> (Lamb) | Auten 1925 |
| | | <i>Philodromus cespitum</i> | Egg | Sarcophagidae | <i>Sarcophaga sexpunctata</i> (Fabricius) | Auten 1925 |
| | | <i>Philodromus</i> sp. | Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1960 |
| Araneae | Phyxelididae | <i>Ambohima sublima</i> Griswold, 1990 | Immature | Acroceridae | <i>Thyllis</i> sp. | Schlinger 2003 |
| Araneae | Plectreuridae | <i>Kibramoa</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera arizonensis</i> Cole | Schlinger 1987 |
| Araneae | Salticidae | <i>Aelurillus v-insignitus</i> (Clerck, 1757) | Adult | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Millot 1938 |
| | | <i>Aelurillus v-insignitus</i> (Clerck) | Adult/ Immature | Acroceridae | <i>Ogcodes varius</i> Latreille | Séguy 1926 |
| | | <i>Cobanus</i> sp. | Adult/ Immature | Acroceridae | <i>Terphis</i> sp. | Schlinger 1987 |
| | | <i>Cosmophasis bitaeniata</i> (Keyserling, 1882) | Adult/ Immature | Acroceridae | <i>Ogcodes basalis</i> (Walker) | Schlinger 1987 |
| | | <i>Cosmophasis bitaeniata</i> | Adult/ Immature | Acroceridae | <i>Ogcodes doddi</i> Wandolleck | Wandolleck 1906, Dodd 1906 |
| | | <i>Eris militaris</i> (Hentz, 1845) | Immature | Acroceridae | <i>Acrocera</i> sp. | Larivière & Borkent 2009 |
| | | <i>Evarcha jucunda</i> (Lucas, 1846) | Immature | Acroceridae | <i>Ogcodes reginae</i> Trojan | Kehlmaier & Almeida 2014 |
| | | <i>Habronattus hallani</i> (Richman, 1973) | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | <i>Heliophanus</i> sp. | Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Millot 1938 |
| | | <i>Heliophanus</i> sp. | Immature | Acroceridae | <i>Ogcodes zonatus</i> Erichson | Millot 1938 |
| | | Lyssomaninae gen. sp. | Adult/ Immature | Acroceridae | <i>Ogcodes guttatus</i> (Costa) | Schlinger 1987 |
| | | <i>Metaphidippus manni</i> (Peckham & Peckham, 1901) | Adult/ Immature | Acroceridae | <i>Pterodontia vix</i> Townsend | Schlinger 1987 |
| | | <i>Metaphidippus</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera bulla</i> Westwood | Schlinger 1987 |
| | | <i>Metaphidippus</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes boharti</i> Schlinger | Schlinger 1987 |
| | | <i>Pelegrina aeneola</i> (Curtis, 1892) | Adult/ Immature | Acroceridae | <i>Acrocera bulla</i> Westwood | Beckwith et al. 1987 |
| | | <i>Pelegrina aeneola</i> | Adult/ Immature | Acroceridae | <i>Ogcodes boharti</i> Schlinger | Beckwith et al. 1987 |
| | | <i>Pelegrina aeneola</i> | Adult/ Immature | Acroceridae | <i>Ogcodes borealis</i> Cole | Schlinger 1987 |
| | | <i>Pelegrina proterva</i> (Walckenaer, 1837) | Immature | Acroceridae | <i>Ogcodes eugonatus</i> Loew | Larivière & Borkent 2009 |
| | | <i>Pelegrina proterva</i> | Immature | Acroceridae | <i>Ogcodes melampus</i> Loew | Larivière & Borkent 2009 |
| | | <i>Phidippus ardens</i> Peckham & Peckham, 1901 | Adult/ Immature | Acroceridae | <i>Ogcodes boharti</i> Schlinger | Schlinger 1987 |
| | | <i>Phidippus audax</i> (Hentz, 1845) | Egg | Phoridae | <i>Phalacrotophora epeirae</i> (Brues) | Jones 1940 |
| | | <i>Phidippus comatus</i> Peckham & Peckham, 1901 | Adult/ Immature | Acroceridae | <i>Ogcodes boharti</i> Schlinger | Schlinger 1987 |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|------------|----------------|--|--------------------|---------------|--|-------------------------------------|
| Araneae | Salticidae | <i>Phidippus johnsoni</i> (Peckham & Peckham, 1883) | Adult/ Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1987 |
| | | <i>Phidippus johnsoni</i> | Adult/ Immature | Acroceridae | <i>Ogcodes boharti</i> Schlinger | Schlinger 1987 |
| | | <i>Phidippus johnsoni</i> | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | <i>Phidippus octopunctatus</i> (Peckham & Peckham, 1883) | Egg | Sarcophagidae | <i>Sarcophaga davidsonii</i> Coquillett | Coquillett 1892; Davidson 1896 |
| | | <i>Phidippus princeps</i> (Peckham & Peckham, 1883) | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | <i>Phidippus regius</i> C. L. Koch, 1846 | Egg | Phoridae | <i>Phalacrotophora epeirae</i> (Brues) | Manuel 1984 |
| | | <i>Phidippus rimator</i> nom. dub. (Walckenaer, 1837) | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | <i>Phidippus</i> sp. | Adult/ Immature | Acroceridae | <i>Acrocera</i> sp. | Schlinger 1987 |
| | | <i>Phlegra fasciata</i> (Hahn, 1826) | Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Millot 1938 |
| | | <i>Sassacus</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | <i>Sidusa</i> sp. | Adult/ Immature | Acroceridae | <i>Terphis</i> sp. | Schlinger 2003 |
| | | | | | | |
| | | | | | | |
| Araneae | Segestriidae | <i>Ariadna maxima</i> (Nicolet, 1849) | Adult/ Immature | Acroceridae | <i>Carvalhoa appendiculata</i> Philippi | Schlinger 1987 |
| Araneae | Tetragnathidae | <i>Meta menardi</i> (Latreille, 1804) | Egg | Phoridae | <i>Megaselia melanocephala</i> (Roser) | Decou-Burghelle 1961 |
| | | <i>Tetragnatha</i> sp. | Egg | Chloropidae | <i>Pseudogaurax silbergliedi</i> Sabrosky | Sabrosky 1990 |
| Araneae | Theraphosidae | <i>Tetragnatha</i> sp. | Egg | Chloropidae | <i>Siphonella</i> sp. | Kintner 1935 |
| | | <i>Acanthoscurria sternalis</i> Pocock, 1903 | Immature | Acroceridae | <i>Exetasis jujuyensis</i> Gillung | Barneche et al. 2013 |
| | | <i>Aphonopelma duplex</i> (Chamberlin, 1925) | Adult/ Immature | Acroceridae | <i>Ocnaea</i> sp. | Alagon & Odell 2004 |
| | | <i>Aphonopelma hentzi</i> (Girard, 1852) | Adult/ Immature | Acroceridae | <i>Lasia purpurata</i> Bequaert | Baerg 1958; Eason et al. 1967 |
| | | <i>Aphonopelma</i> sp. | Adult/ Immature | Acroceridae | <i>Ocnaea</i> sp. | Schlinger 1987 |
| | | <i>Chaetopelma</i> sp. | Adult/ Immature | Acroceridae | <i>Astomella gravis</i> Erichson | Schlinger 1987 |
| | | <i>Grammostola actaeon</i> (Pocock, 1903) | Adult | Acroceridae | <i>Exetasis</i> sp. | Vellard 1934 |
| | | <i>Lasiodora klugi</i> (C. L. Koch, 1841) | Immature | Acroceridae | <i>Exetasis eickstedtae</i> Schlinger | Eickstedt 1971; Schlinger 1972 |
| | | <i>Phrixotrichus scrofa</i> (Molina, 1788) | Immature | Acroceridae | <i>Arrhynchus maculatus</i> Schlinger | Schlinger 1968 |
| Araneae | Theridiidae | <i>Enoplognatha ovata</i> (Clerck, 1757) | Adult/ Immature | Acroceridae | <i>Ogcodes gibbosus</i> (Linnaeus) | Pichka 1977 |
| | | <i>Enoplognatha</i> sp. | Egg | Phoridae | <i>Megaselia tenebricola</i> Schmitz | Evans 1969 |
| | | <i>Enoplognatha</i> sp. or <i>Robertus</i> sp. | Egg | Phoridae | <i>Megaselia angusta</i> Wood | Disney & Evans 1980; Disney 1999 |
| | | <i>Enoplognatha</i> sp. or <i>Robertus</i> sp. | Egg | Phoridae | <i>Megaselia longifurca</i> (Lundbeck) | Disney & Evans 1980; Disney 1999 |
| | | | | | | |

Table 1.—Continued.

| Host order | Host family | Arachnid host | Host stage | Fly family | Fly parasitoid/ parasite/predator | Reference |
|----------------|--------------|---|--------------------|---------------|---|---|
| Araneae | Theridiidae | <i>Latrodectus geometricus</i> C.L. Koch, 1841 | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Vetter et al. 2012 |
| | | <i>Latrodectus hesperus</i> Chamberlin & Ivie, 1935 | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Vetter et al. 2012 |
| | | <i>Latrodectus mactans</i> (Fabricius, 1775) | Egg | Phoridae | <i>Apocephalus borealis</i> Brues | Disney 1994 |
| | | <i>Latrodectus mactans</i> | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Davidson 1896 |
| | | <i>Latrodectus mactans</i> | Egg | Chloropidae | <i>Pseudogaurax</i> sp. | Baerg 1959 |
| | | <i>Parasteatoda tepidariorum</i> (L. Koch, 1841) | Egg | Chloropidae | <i>Pseudogaurax signatus</i> (Loew) | Kaston & Jenks 1937 |
| | | <i>Robertus</i> sp. | Egg | Phoridae | <i>Megaselia tenebricola</i> Schmitz | Disney & Evans 1980 |
| | | <i>Steatoda palomara</i> Chamberlin & Ivie, 1935 | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1960 |
| Araneae | Thomisidae | <i>Diaea</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes nitens</i> (Hutton) | Schlinger 1987 |
| | | <i>Mecaphesa</i> sp. | Immature | Acroceridae | <i>Ogcodes eugonatus</i> Loew | Cokendolpher et al. 1979 |
| | | <i>Misumena vatia</i> (Clerck, 1757) | Adult/ Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | de Jong et al. 2000 |
| | | <i>Thomisus onustus</i> (Walckenaer, 1806) | Adult/ Immature | Acroceridae | <i>Ogcodes fumatus</i> (Erichson) | de Jong et al. 2000 |
| | | <i>Xysticus cunctator</i> Thorell, 1877 | Immature | Acroceridae | <i>Ogcodes adaptatus</i> Schlinger | Schlinger 1960 |
| | | <i>Xysticus cunctator</i> | Immature | Acroceridae | <i>Ogcodes melampus</i> (Loew) | Schlinger 1960 |
| | | <i>Xysticus luctuosus</i> (Blackwall, 1836) | Adult/ Immature | Acroceridae | <i>Ogcodes pallipes</i> Latreille | Trojan 1956 |
| | | <i>Xysticus montanensis</i> Keyserling 1887 | Immature | Acroceridae | <i>Ogcodes borealis</i> Cole | Schlinger 1960 |
| | | <i>Xysticus montanensis</i> | Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1960 |
| | | <i>Xysticus</i> sp. | Adult/ Immature | Acroceridae | <i>Ogcodes eugonatus</i> (Loew) | Schlinger 1987 |
| | | Undetermined genus | Egg | Drosophilidae | <i>Scaptomyza</i> (<i>Titanochaeta</i>) 11 spp. | Wirth 1952; Hardy 1965; Lapoint et al. 2013 |
| | | | | | | al. 2013 |
| Araneae | Trachelidae | <i>Trachela mexicana</i> Banks, 1898 | Adult/ Immature | Acroceridae | <i>Ogcodes pallidipennis</i> (Loew) | Schlinger 1987 |
| Araneae | Unknown | Undetermined genus | Egg | Phoridae | <i>Megaselia oviaraneae</i> Disney | Disney 1999 |
| Scorpiones | Buthidae | <i>Centruroides margaritatus</i> (Gervais, 1841) | Adult/ Immature | Sarcophagidae | <i>Sarcodexia sternodontis</i> Townsend | Townsend 1893 |
| | | <i>Mesobuthus martensii</i> (Karsch, 1879) | Adult/ Immature | Sarcophagidae | <i>Sarcophaga dux</i> Thomson | Shi et al. 2015 |
| Scorpiones | Vaejovidae | <i>Anuroctonus phaiodactylus</i> (Wood, 1863) | Adult | Tachinidae | <i>Spilochaetosoma californicum</i> Smith | Williams et al. 1994 |
| | | <i>Vaejovis spinigerus</i> (Wood, 1863) | Adult/ Immature | Tachinidae | <i>Spilochaetosoma californicum</i> Smith | Williams et al. 1994 |
| Trombidiformes | Anystidae | Undetermined genus | Adult/ Immature | Acroceridae | Undetermined genus | Kerr & Winterton 2008 |
| Trombidiformes | Erythraeidae | <i>Abrolophus</i> sp. | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | Sferra 1986 |
| Trombidiformes | Trombidiidae | <i>Podothrombium</i> sp. | Adult/ Immature | Acroceridae | <i>Pterodontia flavipes</i> Gray | Sferra 1986 |

and deeper investigation is needed in order to verify whether the species in question are predators, parasitoids, or both.

In general, there is still much to be learned about the interactions between Diptera and the arachnids they attack. Many more large rearing studies of spiders and other arachnids would be the best approach to fill this gap in knowledge, as the majority of reports of natural enemies are the result of individual incidental rearings. Unfortunately, there are still large numbers of arachnid species that we know almost nothing about in terms of their habits, habitats, mating behavior and natural enemies. This lack of basic natural history information seriously limits the ability of arachnologists and dipterists to address these issues, though this problem is not limited to these groups or questions (Tewksbury et al. 2014; Barrows et al 2016). It is imperative that we as biologists spend time observing our chosen study organisms in their natural environment, if we are to have any hope of both discovering and understanding the ecological web of organisms that covers our planet.

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