Explaining gene expression variation using neural network derived image features in histopathology

William Jones

Wellcome Trust Sanger Institute Hinxton UK, CB10 1SA wj2@sanger.ac.uk

Abstract

How does gene expression influence visual characteristics of human tissues? To what extent can the variation in expression be predicted using features extracted from these images alone? We use data from the Genotype Tissue Expression (GTEx) project which contains high resolution histopathology images annotated with bulk RNA expression and genotype data. We define image features by taking the penultimate layer activation from InceptioNet-v3, after retraining it to differentiate between 10 different tissue types. Across all tissues, we find that a substantial amount of variation of gene expression can be explained using a combination of neural network derived image features and known technical covariates. After taking into account technical variation, we find over 66% (in Liver)of the variation of certain transcripts can be explained by individual image features. This work indicates that expression biomarkers could soon be well estimated using biomedical images.

1 Introduction

Biomedical images are routinely used by doctors in pathology to diagnose disease. For example, Lung biopsies are taken from patients when grading severities of Lung cancers [1]. Specifically for lung cancer, a great deal of work has focussed on the identification of specific tissue characteristics which are indicative of metastasis. Indeed, this has been the topic of many supervised machine learning competition and approaches. We can now accurately identify metastatic regions from high resolution histopathology lung images, with the state of the art achieved by Convolutional Neural Networks (CNNs) [2].

What other types of information and biomarkers can be estimated from these high resolution images? To what extent are the visual characteristics of biomedical images influenced by our DNA, and gene expression levels within our tissues? Pathologists are trained to identify visual characteristics in histological slides that are indicative of disease, and it is known that disease onset triggers changes in bulk RNA expression data in tissues where the disease presents. [3] Furthermore, in many cases the onset of these has a genetic component [4]. Therefore, it is reasonable to conclude that high resolution images contain information pertaining to the bulk RNA expression profile taken from the sample, and perhaps even the donor genotype.

Until recently the absence such datasets has prohibited work in this direction. With the addition of high resolution histopathology images annotated with gene expression data, and genotypes, as part of the Genotype Tissue Expression Project (GTEx Project) [5], it is now possible to investigate the interplay of histopathology images, gene expression and genetics. At the time of analysis, the repository (v6), consisted of 449 genotyped individuals, along with bulk RNA expression from 44 tissue types [6]. A median of 15 tissues were donated per individual, and a median of 155 samples

of each tissue are available in total. High resolution histopathology images were available for 34 of these tissue types.

2 Methods

We generated image features using the 1024 penultimate layer activations of an InceptioNet CNN retrained to differentiate between square patches originating from 10 different tissue types. These activations are the output of the layer just before the classification layer. We mean aggregate these activations across all patches situated within the tissue boundary for each image sample (Figure 1). This results in a length 1024 vector representation that we use for downstream association analyses.

To investigate the amount of shared variation between the the expression data and these image features, we performed Pearson Correlation tests between the principal components describing the 99.9% of the variation in the image features and expression respectively. To investigate individual transcript-feature relationships for a given tissue, we select the top 2000 varying transcripts with mean expression greater than 1 (log RPKMs). We calculate the Pearson Correlation for each of these pairs or transcript and feature.

3 Results

We find that across 10 different tissue types, a large amount of variation in expression can be explained in large part with a combination of known technical factors and image features. (Figure 3). We find a greater amount of variance is explained after the InceptioNet model is retrained to differentiation between tissue type, but also find that raw InceptioNet feature explain a large amount of residual variance.

RNA degradation number (SMRIN) was the factor that explained the most (11.8%) variation in the image features, wheres Ischemic time explained the most the variance (12.7%) in the expression data (Figure 4). For Lung tissue, technical factors explained 51.1% of the variation in expression and 40.6% of the variation in the image features generated at a patch size of 256.

After regressing out the effect of technical covariates from both the image features and expression, we find that for some transcripts as much as 66% of the variation can be explained by individual image features.

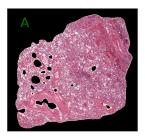
For Lung tissue, we find that feature 501 explains a significant amount of variation for gene transcripts that are enriched for the gene ontology term: Defective ABCA3 causes pulmonary surfactant metabolism dysfunction type 3 (SMDP3) (Table 1). This condition is known to be visually identifiable by pathologists via histopathology [7]. Also in Lung tissue, a specific image feature that was highly predictive of Ischemic time (the duration of time between donor death and tissue harvesting) appeared to capture the global colour (Figure 2b) across the entire tissue area (Figure 2a).

We investigated the genetic basis of variation for individual image features. However we found that no associations passed Benjamini-Hochberg correction at a 5% FDR rate.

4 Discussion

Gene expression reflects the biological activity of a cell at a given point in time. Technical factors are already known to have a large impact on gene expression. For example, Ischemic time (SMTISCH), reflects the period time between the donor's death, and tissue extraction. During apoptosis, multiple biological pathways are activated which have a profound impact on the biological activity within a cell. In our data, this technical factor explains the largest amount of variance in expression out of all over technical factors (Figure 4). Across all tissue types, more variance is explained by image features generated from large sized patches (Figure 3). This is surprising because intuitively one might imagine that variation in gene expression would only affect visual characteristics in tissues at smaller scales. However, these results provide evidence against this intuition. It is also possible that features generated by patches at larger scales are more robust to the mean aggregation, and so are more representative of the entire tissue.

This study motivates further work that aims to close the information boundary between biomedical images and gene expression markers. Future work will aim to predict the variation of specific expression markers for a given disease for diagnosis. This work will require more comprehensive datasets, but with modern day neural networks able to successfully perform function approximation for a wide variety of complex tasks, this might soon be possible.



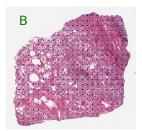
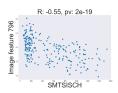


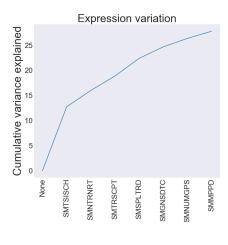
Figure 1: A We illustrate how the tissue is segmented into the tissue foreground and background by Gaussian blurring followed by Otsu thresholding. B We illustrate where patch centers are located within a tissue boundary. The dots represent patch centers that fit inside the tissue boundary.





(a) Image feature 796 appears to capture global tissue colour. Top (b) Image feature 796 strongly row displays top five samples for the feature, bottom row displays SISCH). (b) Image feature 796 strongly predicts Ischemic time (SMT-bottom five lung samples for the feature.

Figure 2



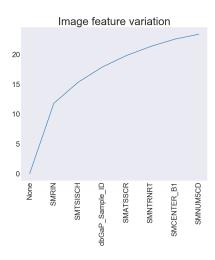


Figure 4: Only displaying the top 8 technical factors. In total, technical factors account for 40.6% of variation in the image features and 51.1% of variation in expression.

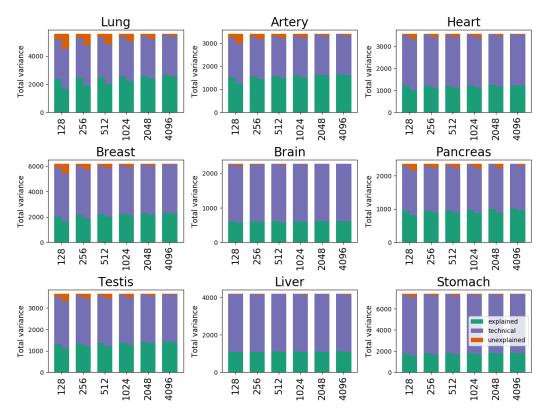


Figure 3: Proportion of the variation in expression explained by image features, explained by technical factors, unexplained, for 10 different tissue types using image features from 6 different patch-sizes. For each patch-size, the **left stacked bar** uses retrained InceptioNet features, and **right stacked bar** uses raw Inception features.

Table 1: Enriched Ontology terms for image feature 501

p-value 1.21e-15 6.87e-09	Ontology lamellar body Defective ABCA3 causes pulmonary sur- factant metabolism dysfunction type 3	Genes LAMP3,CTSH,SFTPA1,NAPSA,SFTPD SFTPA1,SFTPD,SFTPB,SFTPA2
	(SMDP3)	

References

- [1] Lamb D. Histological classification of lung cancer. Thorax. 1984;39(3):161-165.
- [2] Yun Liu & Krishna Gadepalli (2017) Detecting Cancer Metastases on Gigapixel Pathology Images http://arxiv.org/abs/1703.02442v2
- [3] Lewis, P. A., & Cookson, M. R. (2012). Gene expression in the Parkinson's disease brain. Brain Research Bulletin, 88(4), 302?312. http://doi.org/10.1016/j.brainresbull.2011.11.016
- [4] Mitchell, K. J. (2012). What is complex about complex disorders? Genome Biology, 13(1), 237. http://doi.org/10.1186/gb-2012-13-1-237
- [5] Lonsdale, J., Thomas, J., Salvatore, M., Phillips, R., Lo, E., Shad, S., et al. (2013). The Genotype-Tissue Expression (GTEx) project. Nature Genetics, 45(6), 580?585. http://doi.org/10.1038/ng.2653
- [6] GTEx Image Viewer https://brd.nci.nih.gov/brd/image-search/searchhome
- [7] Susan E. Wert Jeffrey A. Whitsett (2009) Genetic Disorders of Surfactant Dysfunction http://journals.sagepub.com/doi/10.2350/09-01-0586.1

[8] McCall, M. N., Illei, P. B., & Halushka, M. K. (2016). Complex Sources of Variation in Tissue Expression Data: Analysis of the GTEx Lung Transcriptome. The American Journal of Human Genetics, 99(3), 624ÅŞ635. http://doi.org/10.1016/j.ajhg.2016.07.007