Image Classification for Identification of Cancerous Lesion

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## **1 Abstract**

When determining whether or not a skin lesion is cancerous or non cancerous, doctors and dermatologists rely on an initial visual examination of the area before making other decisions on how to proceed[[1]](#footnote-0). If some of the key variables are concerning, doctors usually perform a biopsy for a more detailed examination as a last step[[2]](#footnote-1). Dermatologists assess several critical variables when evaluating skin lesions. These include asymmetry, where one half of the lesion differs from the other; irregular or poorly defined borders; color variation; size, particularly when the diameter exceeds 6 millimeters; and the degree of protrusion above the skin surface[[3]](#footnote-2). We extracted features to proxy the variables using HSV, Gabor, Linear Binary Patterns, measuring distance from centroid, reflection score, and features returned from neural network models like ResNet50. For classification we used several methods where random forest and logistic regression returned the best accuracy scores of 75.8% and 75.7% respectively.

## **2 Introduction**

The accurate identification of lesions as cancerous or noncancerous in modern healthcare will influence treatment efficacy and help plan necessary treatment for patients. Early and precise diagnosis help plan timely treatment, particularly critical in conditions like melanoma where early detection improves prognosis, whereas misdiagnoses, including false negatives, can delay necessary interventions.

This project focuses on classifying skin lesions between cancerous and noncancerous classifications by extracting features using techniques such as HSV, Gabor, Linear Binary Patterns, measuring distance from centroid, reflection score and ResNet-50 convolutional neural network which are modeled through Random Forest and Logistic Regression. Authoritative bodies such as the World Health Organization, American Cancer Society, National Cancer Institute, and International Agency for Research on Cancer play pivotal roles in standardizing diagnostic criteria and using the HAM10000 image dataset, we intend to contribute to potential new ways to classify lesions to benefit patients and diagnostic healthcare professionals.

## **3 Data**

Our project focuses on classifying skin lesions from the [HAM10000 dataset](https://dataverse.harvard.edu/dataset.xhtml?persistentId=doi:10.7910/DVN/DBW86T)[[4]](#footnote-3), which consists of 10,015 dermatoscopic images of seven different types of pigmented lesions. Each image has dimensions of 450x600 pixels per color channel and a corresponding segmentation mask for the lesion. Over 50% of the lesions are confirmed through histopathology, while the ground truth for the remaining cases is established through follow-up examinations, expert consensus, or in-vivo confocal microscopy.

| Cancerous | Non-Cancerous |
| --- | --- |
| **AKIEC** : Actinic Keratoses & Intraepithelial Carcinoma | **BKL** :Benign Keratosis-like Lesions |
| **BCC** : Basal Cell Carcinoma | **DF** : Dermatofibroma |
| **MEL** : Melanoma | **NV** : Melanocytic Nevi |
|  | **VASC** : Vascular Lesions |

*Figure 1. Example images and segmentations from each skin lesion classification.*

## **2** **Feature Extraction**

The HAM10000 dataset comes with metadata that contains patients’ age at time of the image being taken, as well as their sex and localisation, which is where the lesion is on their body. Additional features then require extraction from the images themselves.

### **2.1** **Preprocessing**

The following section outlines the feature extraction process and details the features incorporated into our model.

#### **2.1.1 Hair Removal**

Upon reviewing the images, we noticed that a number of them had hair covering the lesion or ruler markers to measure the lesion. When initially attempting to extract features, such as edges of the lesions, the feature extraction algorithms would pick up characteristics of the hair or ruler markers. Since we wanted to focus on information about the lesion itself, we used an algorithm called Dullrazor[[5]](#footnote-4) to remove the hair.

The algorithm converts the image to grayscale, uses BlackHat to enhance dark objects of interest, blur the output image, and then apply binary thresholding to obtain a mask of the hair. This mask is then used to remove the hair pixels, which is then replaced with image inpainting.

|  |  |  |  |  |  |
| --- | --- | --- | --- | --- | --- |
| Original Image | Grayscale | BlackHat | Gaussian Blur | Binary Threshold | Final Image |

*Figure 2. Example of the Dullrazor algorithm on one image.*

### **2.1.2 Property Identification**

Due to a wide variation in lesions, despite the lesion itself generally being somewhat in the main focus of the image, in order to apply the metric for features like eccentricity, each property in the masked image is calculated for their respective size and the largest property is determined to be the main lesion to calculate metrics from.

### **2.2 Symmetry**

Lesion asymmetry can be an indicator of the lesion's characteristics and potential malignancy. Typically, benign lesions, like nevi, exhibit symmetry, whereas malignant lesions, like melanomas, often present asymmetry[[6]](#footnote-5).

The following steps were taken to calculate symmetry scores on a 0-1 scale for each lesion:

1. Find contours of lesion and extract from the largest lesion
2. Compute bounding box around largest contour and extract lesion region
3. Calculate symmetry scores across the x-axis, y-axis, main diagonal, and anti diagonal by reflecting lesion and compute percentage overlap on a 0-1 scale
4. Average symmetry scores to compute combined symmetry score on a 0-1 scale

The x-axis and y-axis symmetry scores are calculated by splitting the lesion region into top/bottom and right/left halves and flipping each pixel at (i,j) on one half to compare it with the respective pixel on the other half. To calculate the main diagonal symmetry score, for each pixel at (i,j) in the lesion region, it is reflected across the main diagonal to (j,i). To calculate the anti diagonal symmetry score, for each pixel at (i,j) in the lesion region, it is reflected across the anti diagonal to (h−j−1,w−i−1), where h and w are the height and width of the region.

**2.2.2 Eccentricity**

Cancerous lesions are known to be less circular[[7]](#footnote-6) and to quantify this, eccentricity is a measure used in image processing to quantify how much a shape deviates from being circular. Eccentricity (e) quantitatively assesses the degree to which a shape diverges from a perfect circle, expressed as a value ranging from 0 to 1. A value of e approaching 0 indicates a shape closely matching a circle, whereas values nearing 1 suggest a significant deviation from circularity.

| ISIC\_0031476 | ISIC\_0026199 | ISIC\_0031471 | ISIC\_0028623 |
| --- | --- | --- | --- |
|  |  |  |  |
| Eccentricity: 0.087699 | Eccentricity: 0.250231 | Eccentricity: 0.750013 | Eccentricity: 0.964014 |

*Figure 3. Examples of eccentricity*

The eccentricity (e) of an ellipse is calculated using the formula:

e = sqrt(1 - b2 / a2)

where b is associated with the y-factor for the ellipse and a is related to the x-factor of the ellipse

To determine the significance of eccentricity, the below breakdown of the distribution by each classification can be seen by the distribution. AKIEC, BCC, and MEL are cancerous, whereas BKL, DF, NV, and VASC are not.

| Cancerous Lesions | Non-Cancerous Lesions |
| --- | --- |
|  |  |
|  |  |
|  |  |
|  |  |

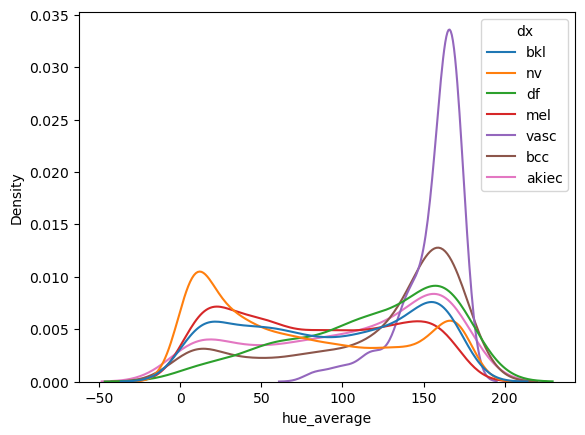
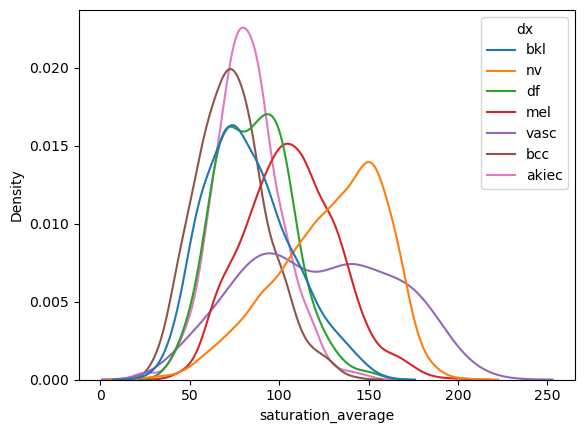
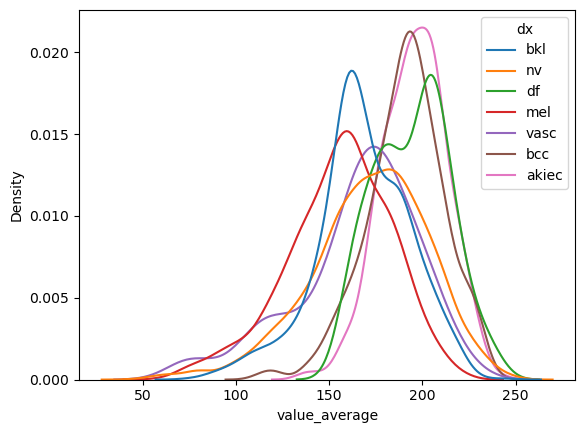
*Figure 4. Distribution of eccentricity across classifications*

**2.2.3 Color**

Due to color variation being an indicator of skin lesion malignancy and type[[8]](#footnote-7), we decided to characterize the color variances within the lesion itself and also compared to the surrounding skin.

***Histogram of Color***

The initial approach taken was to analyze the histogram of color along the axis of hue, saturation, and value to distinguish between skin lesion types. Hue represents the color itself across the red, blue, and green spectrum; saturation reflects the color's intensity or dullness; and value quantifies the color's lightness or darkness.

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*Figure 5. Distribution of HSV averages across groups normalized independently*

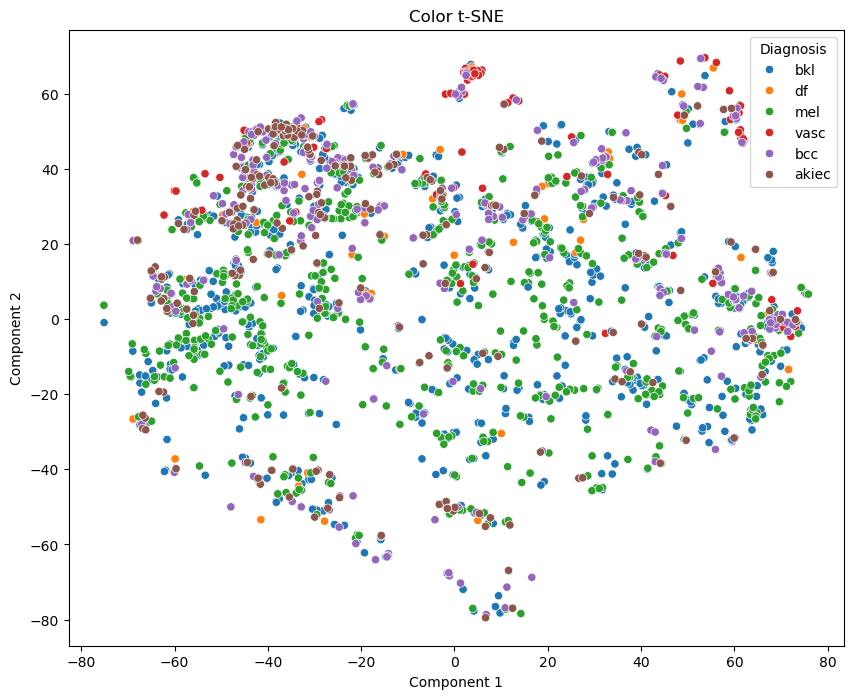
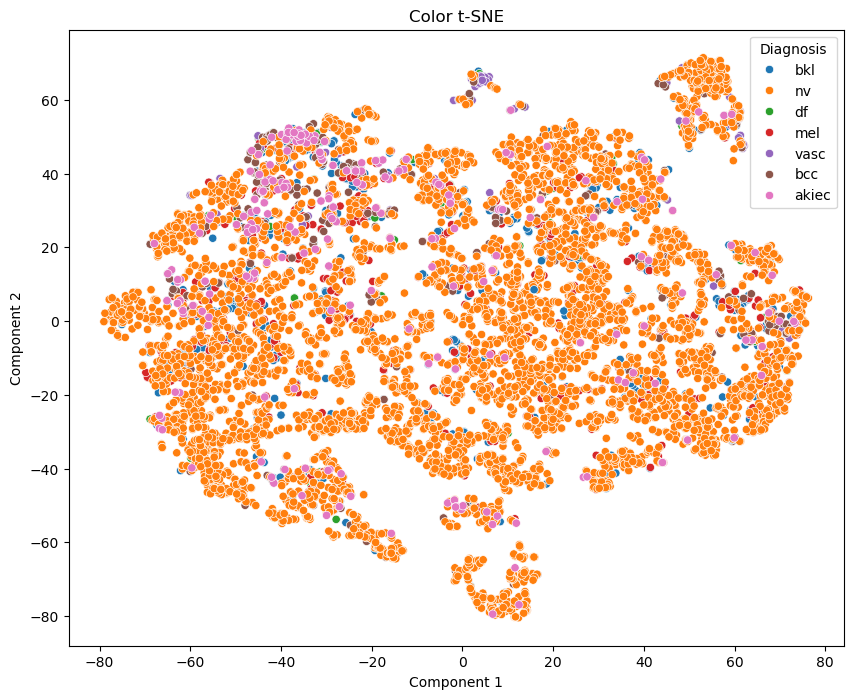
While most of the distributions appear to be similar, there are a few outliers that distinguish one lesion type from another. For example, in hue, VASC lesions have a peak distinct from other groups and the saturation averages for NV differs from the other groups.

***HSV of Lesion v. Surrounding Skin***

Another approach involved calculating the mean and median hue within each lesion and comparing these metrics with those of the surrounding skin. For each image and its corresponding mask, we computed the average and median hue values both within and outside the segmented lesion area. This provides insights into the color differences between the lesion and the surrounding skin.

Another approach involved dividing each lesion into quadrants and computing the average and median hue for each quadrant. The goal is to identify the darkest and lightest hue pixels within the lesion to help identify if there was any difference in color within each lesion. This would help identify color-based patterns or characteristics that might be relevant for understanding different types of skin lesions.

Some of the data manipulations that we accounted for was removing the color black from the min/max hue calculation, otherwise for most lesions any detection of the color black would lead to that skewing the data.



*Figure 6. t-SNE of color*

Using t-SNE, which is a method of dimensionality reduction to visualize how well a feature is distinguished between classes, it appears that NV covers cases in most classes. This is reasonable because NV is a noncancerous skin lesion that is commonly known as a mole. Having a mole can sometimes warrant further follow-up to diagnose if it is any type of cancerous lesion. However, after removing NV, there appears to be some differences between classes, especially with VASC lesions and AKEIC lesions.

**2.2.4 Texture**

We make use of classical texture-capturing methods to supply our model with discrimination power beyond that available in color distributions. Texture features are invariant to skin pigmentation and therefore, we hope, contribute a degree of robustness to the model.

The first texture feature we use is a Gabor filter. Gabor filters are a family of parametrized linear filters whose impulse response is defined by a sinusoidal plane wave multiplied by a Gaussian. This constitutes a 2-dimensional band-pass filter, able to identify the significance of frequencies in some predefined bandwidth. Since that bandwidth, along with the direction of measurement are limited in a single filter, a collection of these is usually used, called a filter bank. However, each filter in the bank would lead to another set of features with cardinality matching the number of pixels in the image. To control for the overall number of features in our model, we use a single Gabor filter, whose hyperparameters were optimized visually for discrimination between the classes. By post-processing the output of this convolution (more below), we manage to extract useful information from this single filter.

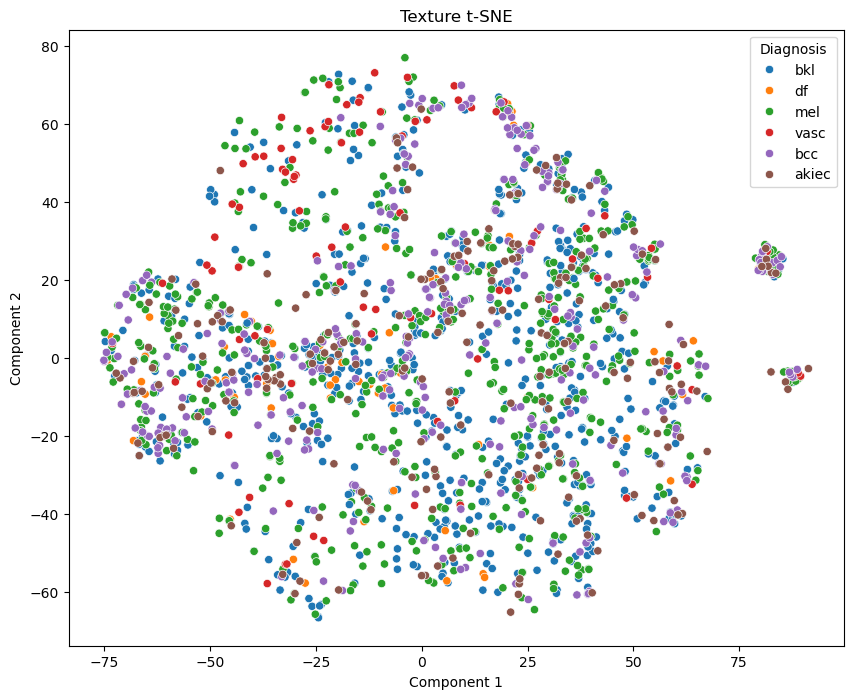
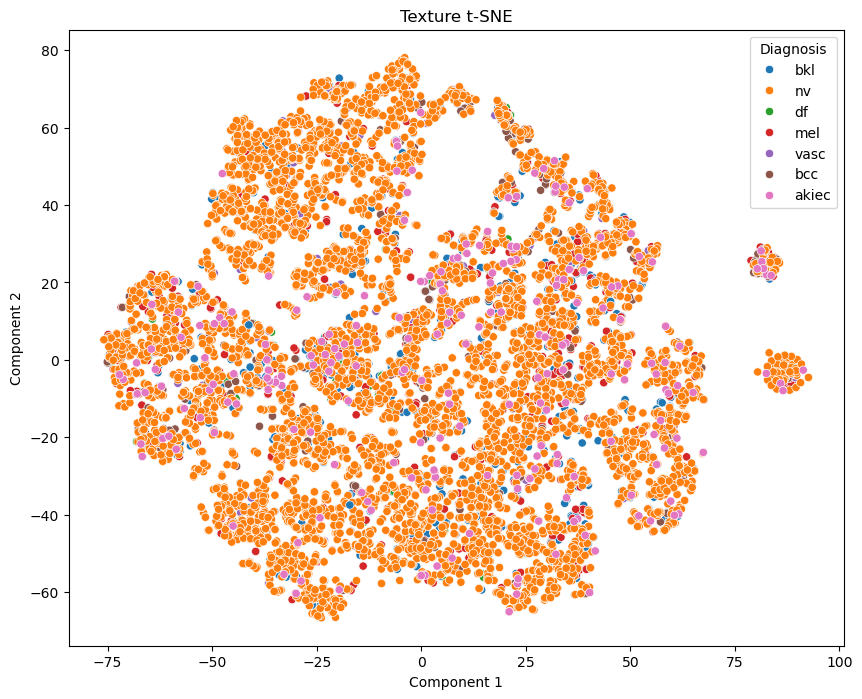
The second texture feature is the linear binary pattern. This is an encoding of the region surrounding a pixel obtained by thresholding each neighboring pixel against the pixel in question, and mapping the relationship between the two to 0 or 1. We then collect the resulting values into a binary string and use the numerical value of that binary string as the texture representation of the pixel. This encodes local contrast observed in the vicinity of each pixel. In much the same way as the Gabor filter, hyperparameters were selected visually for discrimination power between images.

The features described above, as with most commonly used texture methods, yield an output whose dimension matches that of the image. To control for the number of features, and to make these features invariant to position and rotation of the lesion, we aggregate the values produced in the following way. First, we leverage the segmentation masks to identify the center and radius of each lesion’s largest bounded circle. We then identify the lesion’s smallest bounding circle with the same center. Next, we average the two radii, and use the resulting circle as a set of polar-coordinate axes around the lesion. Next, we bin the radius into 8 equally spaced radii, and compute aggregating statistics over each concentric ring. Finally, aggregate over the radial direction.

The aggregations used are listed below:

* Standard deviation of ring means
* Kurtosis of ring means
* Skew of ring means
* Interquartile range of entire circle’s values
* Radial mean slope: OLS coefficient of each ring’s mean regressed on the ring number (size-invariant measure of distance from center)
* Radial standard deviation slope: same as above, but aggregate each ring by the standard deviation of its values.
* Ring number of ring with maximum internal variation (Std)
* Ring number of ring with maximum internal mean
* Lesion region mean relative to outer (non lesion skin) mean

The 18 (9+9) features are then standardized, and then combined with the meta-features and fed into a simple logistic regression classifier for evaluation and features selection. The meta-features alone achieve an accuracy score of 25.6% on the validation set (30% of the dataset). When adding the LBP features, we get 34.9%. The meta and Gabor features combination yield 34.3%, and the combination of meta, Gabor, and LBP features yields 39.2%. A backward stepwise selection algorithm is then applied to remove a total of five features and achieve an accuracy score of 40.4% on the validation set. While they certainly do not discriminate well on their own, these features capture information from the images, at a far cheaper computational cost and risk of overfitting than we would experience by using all pixel values from multiple filters.

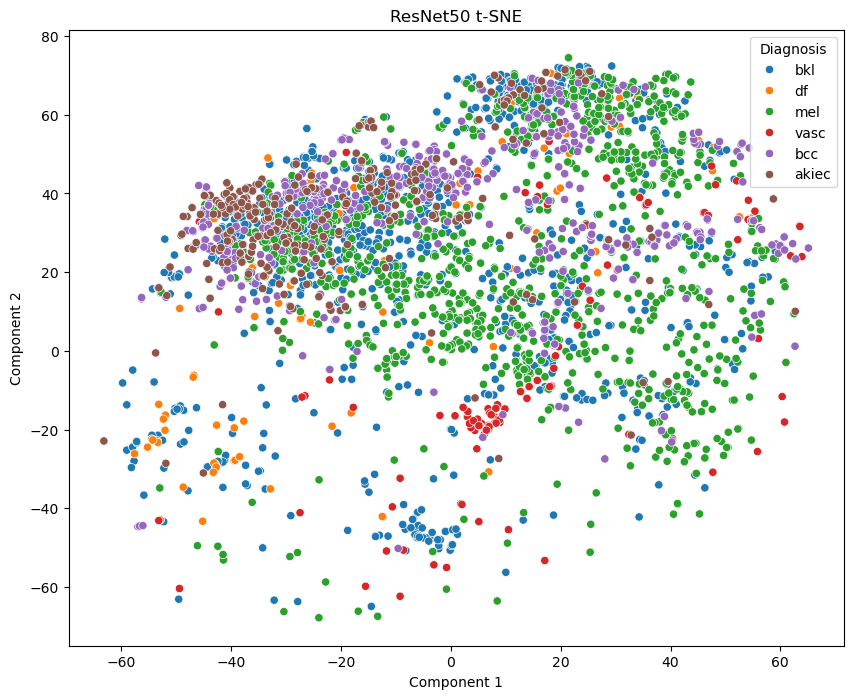
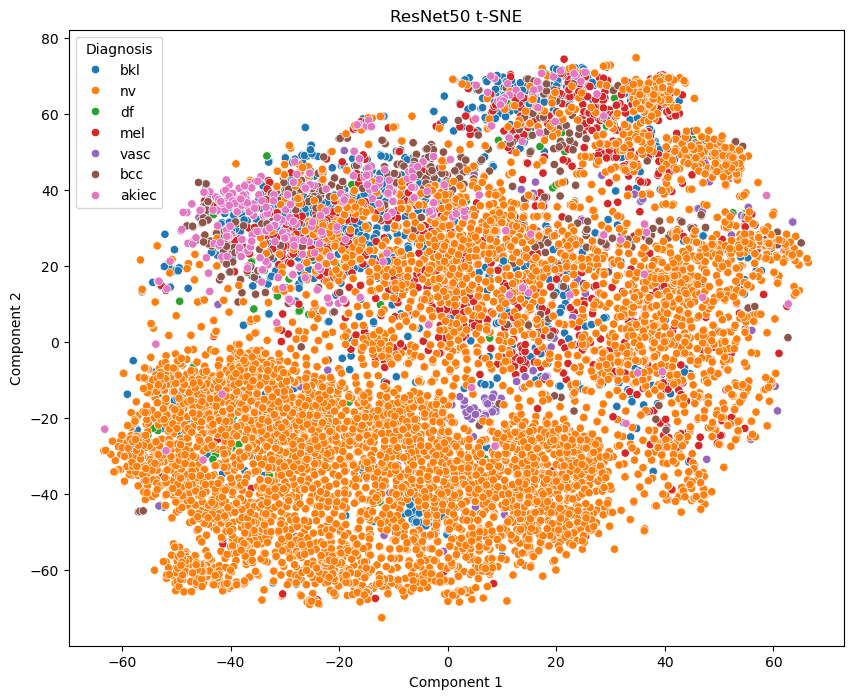
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*Figure 7. t-SNE of Texture*

Using t-SNE to visualize if there is any separation of classes from the images, we noticed that all of the information appears to be clustered together. Even after removing NV, which is the dominant class, it appears that the feature doesn’t provide great distinction between the different classes.

**2.3 ResNet-50 with Principal Component Analysis**

In an effort to retrieve features that are not captured by our own feature extractions, we leveraged a pre-trained neural network, ResNet-50 (Residual Network-50), to output a feature vector. This network is a deep convolutional neural network (CNN) with 50 layers trained on millions of images and classes from the ImageNet database. To obtain a feature vector to be placed within our models, we input the original images that were in RGB, resized them to 244 by 244, and completed Principal Component Analysis (PCA) to reduce the dimensionality of the output.



*Figure 8. t-SNE of ResNet50*

Using t-SNE to visualize if there is any separation of classes from the images, we noticed that all of the information appears to be clustered together. But after removing NV, there appears to be some clusters that are separated from other classes, such as AKEIC lesions. One potential explanation for this is that ResNet-50 is trained to identify different objects rather than the same object with different characteristics. Therefore, these features may be less useful for our final model than we anticipated.

**2.4 Pretrained ResNet on ImageNet Output as Features**

In an effort to improve classification accuracy, we combined the output of an image classification residual neural network trained on the very large “ImageNet” dataset. We passed the network’s output through a max pooling layer, applied dropout with a rate of 0.5, passed the result through a 64-unit hidden layer with relu activation, then concatenated the result with our manually constructed features, and fed the result through another hidden layer, this time with 32 hidden units. Finally, the output was fed to a softmax layer for classification.

We observed no improvement after including these features. This is most likely due to the unique visual content of the images in our dataset, while public classification datasets focus on object recognition, our closeup medical images of skin lesions share little common visual features with those datasets. We believe complex features constructed using neural networks on this dataset or others like it only may yield better results, but leave that for further research since such efforts are outside the scope of this project.

## **3** **Classification**

In this section, we discuss two classification models we trained and analyzed: Random Forest and Logistic Regression. As discussed in Section 2, we used the following features: (1) metadata provided in the dataset, such as age, sex, and lesion location, (2) symmetry, (3) eccentricity, (4) mean HSV values, (5) texture statistical features, and (6) embeddings from the ResNet50 neural network model. There were a total of 792 features used for model training.

**3.1 Results**

For both classification models, our dataset was split into 3 groups: (1) training set, representing 70% (5229 images) of the dataset, (2) validation set, representing 15% (1120 images) of the dataset, and (3) test set, representing 15% (1121 images) of the dataset. The training set for the Random Forest model was also upsampled using synthetic minority oversampling technique (SMOTE) to account for the class imbalance, totalling 26,474 samples. Training the Logistic Regression model using SMOTE was attempted but we struggled with convergence.

After initial model training, we use grid search to tune hyperparameters and use accuracy to evaluate performance. Figure 9 shows the hyperparameter(s) we tune and the best value(s). We then re-train our models using these hyperparameters and evaluate the performance on validation and test set. Figure 10 shows the classification accuracy of our final models.

| **Model** | **Parameter** | **Search Values** | **Best Value** | **Best Accuracy** | **Search Time** |
| --- | --- | --- | --- | --- | --- |
| Random Forest | n\_estimators | 100, 200, 300 | 300 | 76.7% | 8mins 42.3s |
| max\_depth | 3, 5, 10, 12 | 12 |
| class\_weight | None, ‘balanced’, ‘balanced\_subsample’ | ‘balanced’ |
| Logistic Regression | max\_iter | 100, 500, 1000, 2000 | 2000 | 75.7% | 2mins |
| class\_weight | None, ‘balanced’ | ‘balanced’ |

*Figure 9. Summary of hyperparameters*

| **Model** | **Accuracy** | |
| --- | --- | --- |
| Validation Set | Test Set |
| Random Forest | 77.6% | 76.7% |
| Logistic Regression | 72.5% | 75.7% |

*Figure 10. Summary of results of final models*

**3.2 Model Performance**

Both models performed well with the Random Forest model performing better on both the validation set and test set. With hyperparameter tuning, we saw uplifts in accuracy for both classifiers. The inclusion of embeddings from the ResNet50 neural network model did not improve model performance. The features extracted from ResNet50 might not be entirely relevant to the lesion classification task. This could be because ResNet50 is powerful for general image classification, but the features it learns might not align well with the nuances required for distinguishing between different types of lesions. The confusion matrices in Figures 11 and 12 provides a detailed overview of the performance by lesion type and highlights misclassifications.

## **4** **Generalizability**

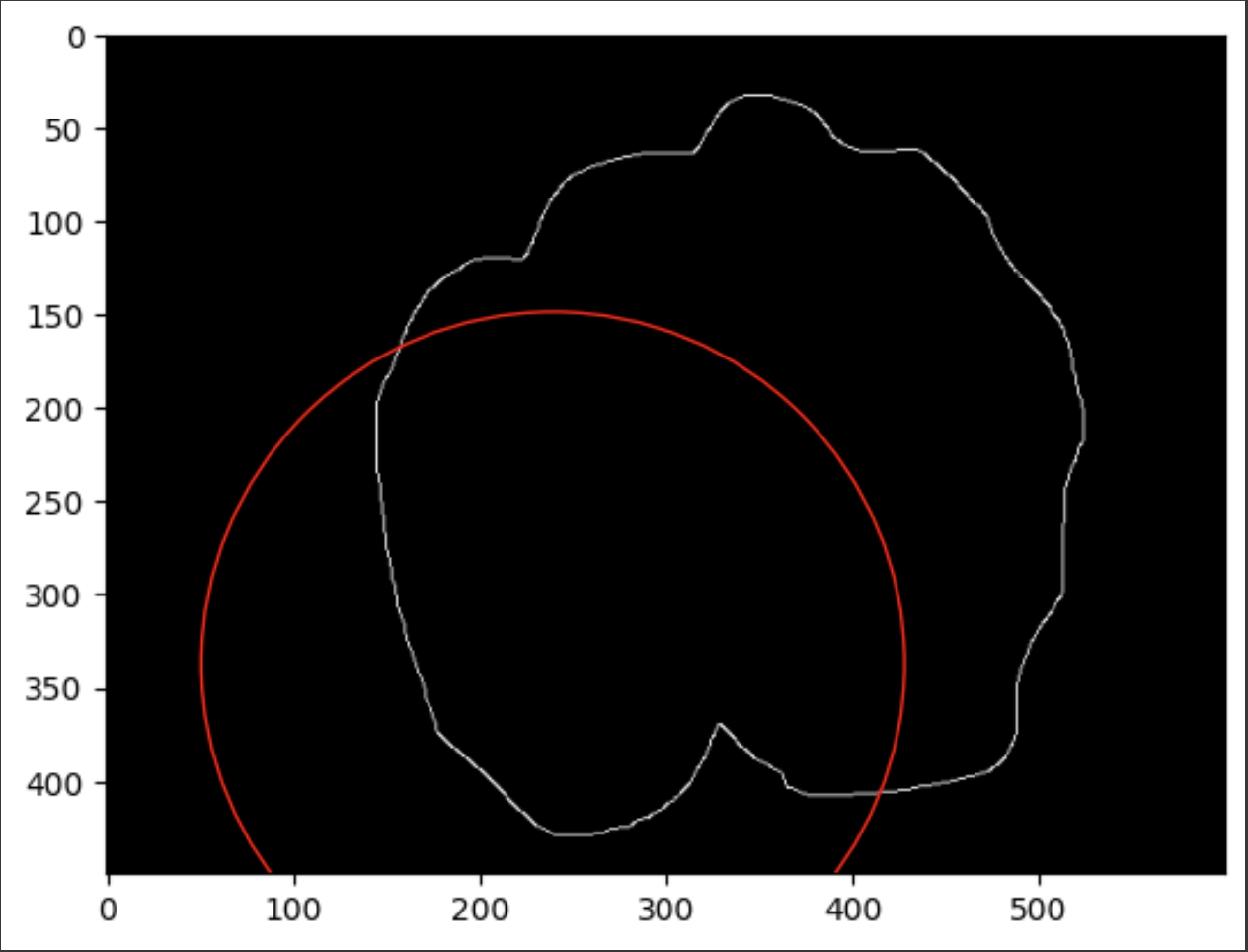
Our logistic regression model generalizes well to the test dataset, in fact it performs better on it than on the validation set. This outperformance is probably coincidental because the validation set was not used to optimize any hyperparameters in this case, the difference illustrates the variance of our model’s performance on different datasets more than anything else.

The random forest model achieved a higher validation score, but this did not generalize well to the test set. While we don’t see severe overfitting, the test score is further away from the deviation than in the logistic regression case, to the extent that the two test set scores are similar despite a significant improvement on the validation set. This is coming from a model that is more flexible in its ability to fit, and whose hyperparameters have been optimized on the validation set, suggestive of some overfitting, but the two are still close enough to one another to indicate some generalizability. It is however important to note, that when accounting for this worse generalizability by not optimizing anything on the test set, we conclude this model does not outperform the logistic regression model.

## **5 Efficiency vs. Accuracy**

The complex features included in our model yielded no better results than our simple logistic regression model, this rather simplifies the efficiency-accuracy tradeoff, we opt for the better and more efficient logistic regression model. Another model that performed similarly well was, as described in section 3, the random forest model. While this model is not much slower than the logistic regression model to train, it does not beat it, and so it is unnecessary to pay that additional cost.

In terms of the features computation, the texture features’ circles took about 10 hours to generate on 64 CPU cores. This is the most computationally intensive step in our feature pipeline, but we believe it can be improved with a better optimization algorithm. An attempt was made to improve efficiency by solving a linear regression on the circle’s parameters fitted to canny-identified edges from the segmentation masks, but the many of the resulting circles (as per example below) lacked precision. We opt for the grid-search approach and leave the algorithmic improvement to future works.

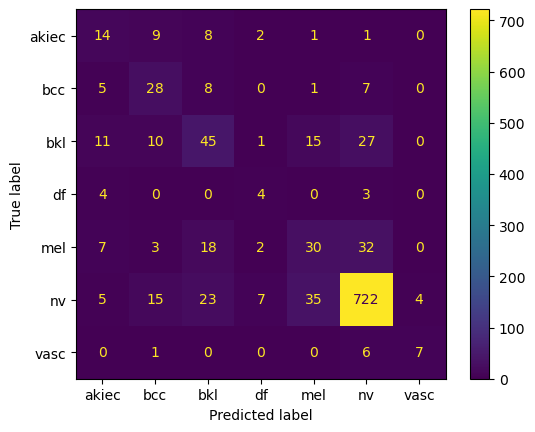


*Figure 12.*

| Model | Model Training Time | Prediction Time | | |
| --- | --- | --- | --- | --- |
| Training Data | Validation Data | Testing Data |
| Logistic Regression | 55s | 0.5s | 0.5s | 0.5s |
| Random Forest | 21.6s | 0.1s | 0.1s | 0.1s |

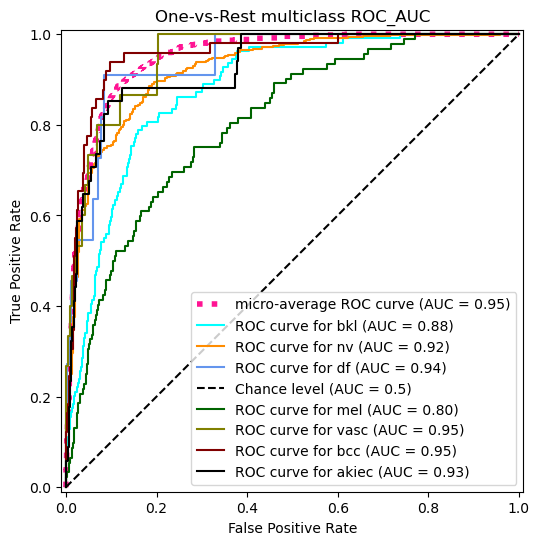
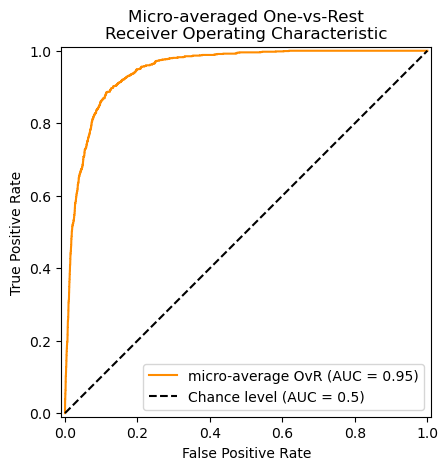
## **6** **Detailed Model Performance**

**Random Forest:** Random Forest is an ensemble learning method that builds multiple decision trees and merges them to improve the overall prediction accuracy and control over-fitting. It handles high-dimensional data well and can capture non-linear relationships, making it well-suited for the lesion image classification task where the visual patterns can be complex and varied. Our Random Forest model performed well in aggregate. This was driven by the model’s strong performance in classifying the “nv” class (F1 score of 0.88), which represented the most number of data points. However, the model had varying performance across the other classes.

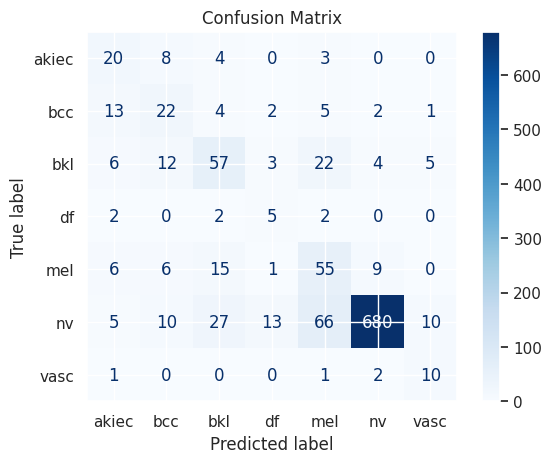


| **Class** | **Precision** | **Recall** | **F1** |
| --- | --- | --- | --- |
| akiec | 0.30 | 0.40 | 0.34 |
| bcc | 0.42 | 0.57 | 0.49 |
| bkl | 0.44 | 0.41 | 0.43 |
| df | 0.25 | 0.36 | 0.30 |
| mel | 0.36 | 0.33 | 0.34 |
| nv | 0.90 | 0.89 | 0.89 |
| vasc | 0.63 | 0.5 | 0.56 |

*Figure 11. Confusion matrix for Random Forest model*

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**Logistic Regression:** Logistic Regression is a simple and easy to interpret classifier. It predicts the probability of each class based on one or more input features. It is suitable for problems where relationships between features and the target class are linear. Despite the challenges with obtaining complex relationships using Logistic Regression, our model performs well in aggregate. This was driven by the model’s strong performance in classifying the “nv” class (F1 score of 0.88), which represented the most number of data points. However, similar to the Random Forest, the Logistic Regression model had varying performance across the other classes.



| **Class** | **Precision** | **Recall** | **F1** |
| --- | --- | --- | --- |
| akiec | 0.27 | 0.41 | 0.33 |
| bcc | 0.26 | 0.41 | 0.32 |
| bkl | 0.48 | 0.50 | 0.49 |
| df | 0.19 | 0.45 | 0.26 |
| mel | 0.36 | 0.59 | 0.44 |
| nv | 0.97 | 0.81 | 0.88 |
| vasc | 0.39 | 0.60 | 0.47 |

*Figure 12. Confusion matrix for Logistic Regression model*

## **7** **Discussion**

During the image pre-processing stage, we used the Dullrazor algorithm to remove hair and ruler marking within the image that impacted our ability to detect edges. One limitation of this method is that it can only detect dark hair. However, we did not notice an issue of edge detection with lighter color hair, therefore, this limitation minimally affects the feature extraction. Another limitation is that the algorithm sometimes removed blood vessels and other important color features within the images. This limitation may have affected the color extraction by decreasing the range of colors within the lesion, which may have impacted our ability to detect more granular differences between lesion types. Therefore, one future direction is to try a sharper hair removal method that only removes hair and ruler marking while leaving the other lesion structures in-tact. One potential next algorithm is SharpRazor[[9]](#footnote-8).

**Limitations**

A lesion may be deemed "suspicious" or concerning if it meets one or more of these criteria: asymmetry, border irregularity, color variation, or size. Additional criteria for assessing whether a lesion is cancerous but involve more advanced techniques, such as dermoscopy, which examines specific patterns and pigmentation.

A comprehensive evaluation of whether a skin lesion is cancerous typically involves visual inspection, dermoscopy, and biopsy. Based on this understanding, we focused our dataset on features observable through visual inspection, including symmetry, edges, and color. However, our source data lacks information on the lesion-to-skin ratio, as it consists only of close-up images of the lesion, and is limited to a two-dimensional view, preventing assessment of the lesion's protrusion above the skin. Consequently, we were constrained in developing a model that incorporates lesion size and skin protrusion.

TO DO (bullet points for features):

## **8** **Conclusion**

Identifying whether skin lesions are cancerous by using visual clues is extremely important for dermatologists. In our project, we utilized the HAM10000 dataset to explore various techniques for improving the classification of these lesions. We focused on specific features such as symmetry, eccentricity, color, and texture, and assessed the performance of Random Forest and Logistic Regression models.

One major challenge was the class imbalance in the dataset, with a larger number of a specific cancerous class compared to others. To address this, we applied class weighting to our models. This adjustment ensured that cancerous lesions, which were underrepresented, were given higher importance, preventing the models from being biased towards the noncancerous class. This approach was crucial for enhancing the models' ability to detect cancerous lesions accurately.

Despite some limitations in our feature extraction methods and the complexity of the images, our models yielded encouraging results. The Logistic Regression and or Random forest model, after hyperparameter tuning and applying class weights, showed the highest accuracy ((we can replace this sentence when we have some final numbers based on the features)). This highlighted the effectiveness of combining simpler models with well-chosen features.

In terms of next steps and potential improvement, some ideas could be enhancing the precision of hair removal algorithms and developing more advanced feature extraction techniques could lead to better model performance. Additionally, employing neural networks such as Sharprazor specifically designed for medical imaging might offer improved classification of lesion types.

In conclusion, our study demonstrates the potential of machine learning and computer vision in classifying skin lesions. Our goal is to assist healthcare professionals in making more accurate and timely diagnoses, ultimately benefiting patient outcomes.

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2. (*Tests For Melanoma Skin Cancer | Melanoma Diagnosis*, *2023*) [↑](#footnote-ref-1)
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4. *(Philipp Tschandl, 2018)* [↑](#footnote-ref-3)
5. *(Lee et al., 1997)* [↑](#footnote-ref-4)
6. *(Ali, 2020)* [↑](#footnote-ref-5)
7. *(Ali, 2020)* [↑](#footnote-ref-6)
8. *(Ali, 2020)* [↑](#footnote-ref-7)
9. *(Kasmi et al., 2023)* [↑](#footnote-ref-8)