Supplementary Material

Main document: A compositional model to assess expression changes from single-cell RNA-seq data

Authors: Ma, Korthauer, Kendziorski, and Newton

Version: February 20, 2019

This supplement is organized to match the sectioning of the main document. In summary,

- 1. Introduction
 - R package
- 2. Modeling
 - Data Structure, Sampling Model, and Parameters

Proof of Theorem 2

Method Structure and Clustering

 ${\tt EBSeq}$

modalClust

Randomized *K*-means

Selecting *K*

• Double Dirichlet Mixture

Proof of Properties 1-6 and Theorem 3

- 3. Numerical Experiments
 - Synthetic data, splatter
 - Empirical study, conquer
 - Null cases
 - Local fdr and
- 4. Other asymptotic properties
 - proof of theorem 4

1. Introduction.

1.1. R package. Reference can be found at github site ...

on scDDboost, web page, etc

2. Modeling.

2.1. Data Structure, Sampling Model, and Parameters. Proof of Theorem 2.

PROOF. Recall $\theta = (\phi, \psi, \mu, \sigma)$. Through the sampling procedure of our model (Figure 3), assuming we known number of cells within each conditions (n_1, n_2) . We have z^1, z^2 are multinomial draw given ϕ and ψ , thus the generation of y, z only depends on (ϕ, ψ) Also given $z, X_{g,c}$ is sampled through NB (μ_{g,z_c}, σ_g) , only depends on (μ, σ) . Thus $P(X, y, z | \theta) = P(y, z | \phi, \psi) P(X | z, \mu, \sigma)$, and we independently give priors for (μ, σ) and (ϕ, ψ) By the Baye's rule,

$$\begin{split} P(\theta|X,y,z) &\propto P(X,y,z|\theta)P(\theta) \\ P(X,y,z|\theta)P(\theta) &= P(y,z|\phi,\psi)P(X|z,\mu,\sigma)P(\mu,\sigma|z)P(\phi,\psi) \\ P(\phi,\psi|y,z) &\propto P(y,z|\phi,\psi)P(\phi,\psi) \\ P(\mu,\sigma|X,z) &\propto P(X|z,\mu,\sigma)P(\mu,\sigma|z) \\ \text{Thus } P(\theta|X,y,z) &\propto P(\phi,\psi|y,z)P(\mu,\sigma|X,z) \end{split}$$

From this we know

- 1. Given X, y and z, $(\phi, \psi) \perp (\mu, \sigma)$
- 2. Given condition and subtypes label $y, z, (\phi, \psi)$ is independent with X
- 3. Given *X* and *z*, (μ, σ) is independent with *y*

Thus we have
$$P\left(A_{\pi} \cap M_{g,\pi} | X, y, z\right) = P\left(A_{\pi} | y, z\right) P\left(M_{g,\pi} | X, z\right)$$
.

- 2.2. Method Structure and Clustering.
- 2.2.1. *EBSeq.* Suppose we have K subtypes, let $X_g^I = X_{g,1}^I, ..., X_{g,S_1}^I$ denote transcripts at gene g from subtype I, I = 1, ...K. In the EBSeq model it assumed that counts within subtype I are distributed as Negative Binomial: $X_{g,s}^I | r_{g,s}, q_g^I \sim NB(r_{g,s}, q_g^I)$ Where

$$P(X_{g,s}^{I}|r_{g,s},q_{g}^{I}) = {X_{g,s} + r_{g,s} - 1 \choose X_{g,s}} (1 - q_{g}^{I})^{X_{g,s}^{I}} (q_{g}^{I})^{r_{g,s}}$$

and $\mu_{g,s}^I = r_{g,s}(1 - q_g^I)/q_g^I$; For the ease of later deriving the prior predictive function of EB framework we use q rather than μ to parameterize the NB.

The EBSeq model assumed a prior distribution on $q_g^I:q_g^I|\alpha,\beta^{I_g}\sim Beta(\alpha,\beta^{I_g})$. The hyperparameter α is shared by all the isoforms and β^{I_g} is I_g specific. Now we are using EBSeq for expression inference of genes rather than isoforms, we made a modification to the original prior from EBSeq. We still make α to be shared by all the genes and β^g becomes gene specific parameter.

The modeling of $r_{g,s}$ is the same as what EBSeq does. Specifically, we further assume that $r_{g,s} = r_{g,0} * l_s$ where $r_{g,0}$ is an isoform specific parameter common across subtypes and $r_{g,s}$ depends on it through the sample-specific normalization factor l_s . Here we expected normalized data, so $l_s = 1$ for all samples, that is $r_{g,s}$ degenerate to σ_g , which does not depend on sample. We use σ_g instead of $r_{g,s}$ in the following.

What we are interested at those *K* groups comparison is the expression pattern,

$$M_{g,\pi} = \{\theta \in \Theta : \mu_{g,k} = \mu_{g,k'} \iff k,k' \in b, b \in \pi\}.$$

For example K = 3, there are 5 expression pattern, $P_1, P_2, ..., P_5$, Comparison between μ is equivalent as comparison between q.

P1:
$$q_g^1 = q_g^2 = q_g^3$$

P2: $q_g^1 = q_g^2 \neq q_g^3$
P3: $q_g^1 \neq q_g^2 = q_g^3$
P4: $q_g^1 = q_g^3 \neq q_g^2$
P5: $q_g^1 \neq q_g^2 \neq q_g^3$ and $q_g^1 \neq q_g^3$

Under the assumption that two groups I and J share the same q_g we can pool the counts from the two groups by viewing them come from same distribution i.e. $X_g^{I,J}|\sigma_g, q_g \sim NB(\sigma_g, q_g), q_g|\alpha, \beta^g \sim Beta(\alpha, \beta^g)$ and obtained the prior predictive function $f_0^g(X_g^{I,J}) = \int_0^1 P(X_g^{I,J}|r_g, q_g) * P(q_g|\alpha, \beta^g) dq_g = \left[\prod\limits_{s=1}^S {X_{g,s} + \sigma_g - 1 \choose X_{g,s}} \right] \frac{Beta(\alpha + \sum_{s=1}^S \sigma_g, \beta^g + \sum_{s=1}^S X_{g,s})}{Beta(\alpha, \beta^g)}$. Consequently, we have prior predictive function for P1, ..., P5 as

$$\begin{split} h_1^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) &= f_0^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) \\ h_2^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) &= f_0^{\mathcal{S}}(X_{\mathcal{S}}^{1,2}) f_0^{\mathcal{S}}(X_{\mathcal{S}}^3) \\ h_3^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) &= f_0^{\mathcal{S}}(X_{\mathcal{S}}^1) f_0^{\mathcal{S}}(X_{\mathcal{S}}^{2,3}) \\ h_4^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) &= f_0^{\mathcal{S}}(X_{\mathcal{S}}^{1,3}) f_0^{\mathcal{S}}(X_{\mathcal{S}}^2) \\ h_5^{\mathcal{S}}(X_{\mathcal{S}}^{1,2,3}) &= f_0^{\mathcal{S}}(X_{\mathcal{S}}^1) f_0^{\mathcal{S}}(X_{\mathcal{S}}^2) f_0^{\mathcal{S}}(X_{\mathcal{S}}^3) \end{split}$$

Then the marginal distribution of counts $X_g^{1,2,3}$ is $\sum_{k=1}^5 p_k h_k^g(X_g^{1,2,3})$, where the marginal $p_k = P(M_{g,\pi}|z)$ (shared by all genome) satisfying $\sum_{k=1}^5 p_k = 1$ and are estimated by EM algorithm. Thus, the posterior probability of an expression pattern k is obtained by:

$$\frac{p_k h_k(X_g^{1,2,3})}{\sum_{k=1}^5 p_k h_k^g(X_g^{1,2,3})}$$

In the optimization steps for determining the hyper parameters (α, β^g, p) , the computation and memory increase exponentially with the number of subtypes K. We use one-step EM as an approximation for the solution, that is α and β^g are updated through gradient ascent. p is updated by the explicit form of the maximizer of the log likelihood.

2.2.2. modalClust. Product Partition Model

Let $X = (X_1, X_2, ..., X_n)$ be n one dimension observed data, given a partition for the data $\pi = \{S_1, ..., S_q\}$, where S_i are disjoint subsets of $\{1, 2, ..., n\}$ and $\bigcup_{i=1}^q S_i = \{1, 2, ..., n\}$. The likelihood for X satisfying such partition is

$$p(X|\pi) = \prod_{i=1}^{q} f(X_{S_i})$$

where X_{S_i} is the vector of observations corresponding to the items of component S_i , The component likelihood $f(X_S)$ is defined for any non-empty component S and can take any form. The partition π is the only parameter we are interested at. Any other parameters that may have been involved in the model have been integrated over their prior.

The prior distribution for a partition π is also taken as a product form. We use the MAP partition(maximize the posterior $p(\pi|X) \propto p(X|\pi)p(\pi)$) as the estimated clustering.

Dahl demonstrated by some choice of f and prior of π , we can reduce the time complexity of finding the MAP partition from factorial(n) to $O(n^2)$ (Dahl, 2009), And the crucial condition for f is that if X_{S_1} and X_{S_2} are overlapped in the sense that $\min\{X_{S_2}\} < \max\{X_{S_1}\} < \max\{X_{S_2}\}$ or $\min\{X_{S_1}\} < \max\{X_{S_2}\} < \max\{X_{S_1}\}$, let $X_{S_1^*}$ and $X_{S_2^*}$ be the sets of swapping one pair of those overlapped terms and keep the other unchanged. Then $f(X_{S_1})f(X_{S_2}) \leq f(X_{S_1^*})f(X_{S_2^*})$. Under such condition, we know that possible MAP candidates must be those partition π s that for any two blocks $b_1, b_2 \in \pi$, either $\max_{i \in b_1}(X_i) \leq \min_{j \in b_2}(X_j)$ or $\min_{i \in b_1}(X_i) \geq \max_{j \in b_2}(X_j)$.

In Poisson-Gamma Model we assuming:

$$X_{i}|\pi, \lambda \sim Poisson(X_{i}|\lambda_{1}\mathbf{I}\{i \in S_{1}\} + ... + \lambda_{q}\mathbf{I}\{i \in S_{q}\})$$

$$\pi \sim p(\pi)$$

$$\lambda_{i} \sim Gamma(\alpha_{0}, \beta_{0})$$

where $p(\pi) \propto \prod_{i=1}^{q} \eta_0 \Gamma(|S_i|)$. Integrate out λ , $f(X_S)$ is obtained as:

$$f(X_S) = \frac{\beta^{\alpha}}{(|S| + \beta)^{\sum\limits_{i \in S} X_i + \alpha}} \frac{\Gamma(\sum\limits_{i \in S} X_i + \alpha)}{\Gamma(\alpha)} \frac{1}{\prod\limits_{i \in S} X_i}$$

To apply modal-cluster on Poisson-Gamma model, we need to show $f(X_S)$ still satisfying the condition mentioned before.

PROOF. if X_{S_1} and X_{S_2} are overlapped, without loss of generality, we assume $\min\{X_{S_2}\} < \max\{X_{S_1}\} < \max\{X_{S_2}\}$, and we swap $\max\{X_{S_1}\}$ with $\min\{X_{S_2}\}$ and keep the rest unchanged or we could also swap $\max\{X_{S_1}\}$ with $\max\{X_{S_2}\}$. We denote the new set forming by swap of $\max\{X_{S_1}\}$ with $\min\{X_{S_2}\}$ as S_1^* and S_2^* and swap of $\max\{X_{S_1}\}$ with $\max\{X_{S_2}\}$ as S_1^{**} , S_2^{**} accordingly.

Then we need to show at least one of the following happens

(1)
$$f(X_{S_1^*})f(X_{S_2^*}) \ge f(X_{S_1})f(X_{S_2})$$

(2)
$$f(X_{S_1^{**}})f(X_{S_2^{**}}) \ge f(X_{S_1})f(X_{S_2})$$

Let $a = \max\{X_{S_1}\}$, $b = \min\{X_{S_2}\}$ and $c = \max\{X_{S_2}\}$. $h_1 = \sum_{i \in S_1} X_i - a$ and $h_2 = \sum_{i \in S_2} X_i - b$, n_1 and n_2 are the number of elements in S_1 and S_2 . Then

$$f(X_{S_{1}^{*}})f(X_{S_{2}^{*}}) \geq f(X_{S_{1}})f(X_{S_{2}}) \iff \frac{\Gamma(h_{1}+a+\alpha)}{(n_{1}+\beta)^{h_{1}+a+\alpha}} \frac{\Gamma(h_{2}+b+\alpha)}{(n_{2}+\beta)^{h_{2}+b+\alpha}} \leq \frac{\Gamma(h_{2}+a+\alpha)}{(n_{2}+\beta)^{h_{2}+a+\alpha}} \frac{\Gamma(h_{1}+b+\alpha)}{(n_{2}+\beta)^{h_{1}+b+\alpha}} \iff \frac{\Gamma(h_{1}+a+\alpha)}{\Gamma(h_{1}+b+\alpha)} \frac{\Gamma(h_{2}+b+\alpha)}{\Gamma(h_{2}+a+\alpha)} \leq (\frac{n_{1}+\beta}{n_{2}+\beta})^{a-b}$$

Left hand side of above formula is LHS₁ = $\frac{(h_1+b+\alpha)...(h_1+a-1+\alpha)}{(h_2+b+\alpha)...(h_2+a-1+\alpha)}$ by the property of Gamma function and X_i are integer.

Similarly,

$$f(X_{S_1^{**}})f(X_{S_2^{**}}) \ge f(X_{S_1})f(X_{S_2}) \iff \frac{\Gamma(h_2 + c + \alpha)}{\Gamma(h_2 + a + \alpha)} \frac{\Gamma(h_1 + a + \alpha)}{\Gamma(h_1 + c + \alpha)} \le (\frac{n_2 + \beta}{n_1 + \beta})^{c - a}$$

Left hand side of above formula is LHS₂ = $\frac{(h_2+a+\alpha)...(h_2+c-1+\alpha)}{(h_1+a+\alpha)...(h_1+c-1+\alpha)}$

If
$$h_1 \le h_2$$
, then LHS₁ $\le (\frac{h_1 + a - 1 + \alpha}{h_2 + a - 1 + \alpha})^{a - b}$ and LHS₂ $\le (\frac{h_2 + c - 1 + \alpha}{h_1 + c - 1 + \alpha})^{a - b}$

So if
$$\frac{h_1+a-1+\alpha}{h_2+a-1+\alpha} \le \frac{n_1+\beta}{n_2+\beta}$$
 then (12) holds, if $\frac{h_2+c-1+\alpha}{h_1+c-1+\alpha} \le \frac{n_1+\beta}{n_2+\beta}$ then (13) holds

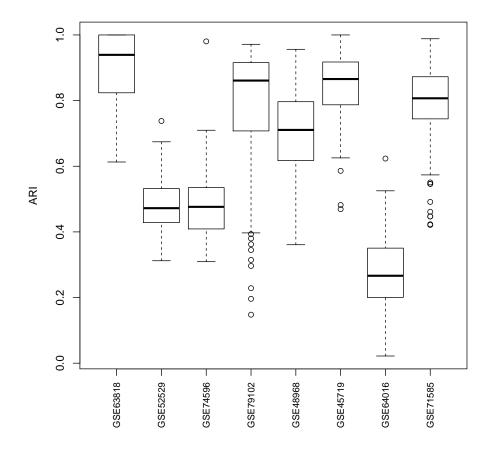
We multiply those two inequalities, we found that $\frac{h_1+a-1+\alpha}{h_2+a-1+\alpha}*\frac{h_2+c-1+\alpha}{h_1+c-1+\alpha}=\frac{h_1+a-1+\alpha}{h_1+c-1+\alpha}*\frac{h_2+c-1+\alpha}{h_2+a-1+\alpha}\leq 1$ as c>a and $h_1\leq h_2$ But $\frac{n_1+\beta}{n_2+\beta}*\frac{n_1+\beta}{n_2+\beta}=1$. At least one equality holds, consequently at least one of (12) and (13) holds.

Similar proof for the case $h_1 > h_2$.

2.2.3. Randomized K-means. To find the value of a_0 , a_1 and d_0 , we have the marginal likelihood of $d_{i,j}$.

$$P(d_{i,j}|a_0,a_1,d_0) = \frac{\Gamma(a_0 + a_1)}{\Gamma(a_0)\Gamma(a_1)} \frac{d_0^{a_0} d_{i,j}^{a_1 - 1} a_1^{a_1}}{(d_0 + a_1 * d_{i,j})^{a_0 + a_1}}$$

We estimate d_0 by treating $d_{i,j} \approx \Delta_{i,j}$ and based on the mean-variance ratio $(\frac{\mathbb{E}(1/\Delta_{i,j})}{\text{Var}(1/\Delta_{i,j})} = d_0)$, d_0 can be approximately estimated by moments of $1/d_{i,j}$, then we obtain a_0, a_1 from MLE of marginal density of $d_{i,j}$. The MLE estimators are obtained through "nlminb" function in r, one issue is that the default value for tolerance rate of stopping is 1e-10, which yields large value of $a_1 + a_0$ and resulting in non-randomness of our weighting matrix. We set tolerance rate as 1e-3. And obtained moderate deviation from D (supplementary Figure 1)



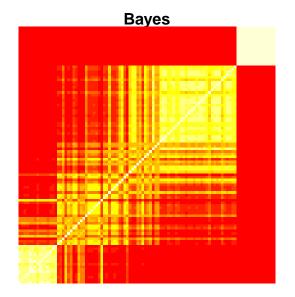
Supplementary Figure 1: Adjusted rand indexes to the clustering based on the original distance matrix without dividing weights. We investigate the randomness of clustering given by our weights through 8 datasets. All have stopping threshold for nlminb optimizing function in r with relative tolerance as 0.001

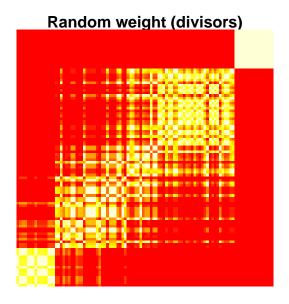
We plot the ARI(adjusted random index) between the randomly generated clustering to clustering under the original distance across eight datasets. Though the mean varies, the interquartile range is wide enough presenting a reasonable variation of our random weighting scheme.

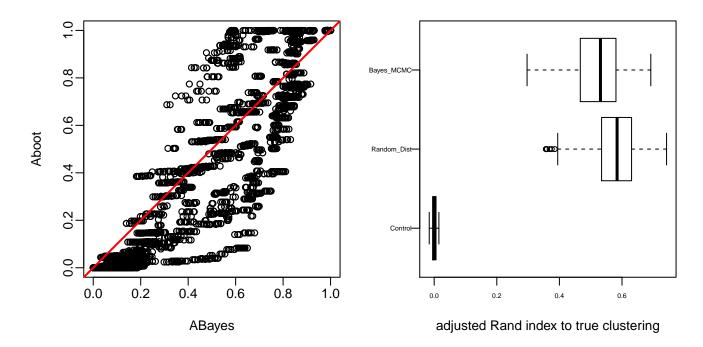
We also check validity of random weighting on simulated dataset. We random generate one-dimensional data *X* from a mixture of 5 normal distributions with different means and same variance. We compare clustering results between random weighting and bayesian clustering with Dirichlet process prior in terms of posterior probabilities that two elements belong to the same class given the whole data. We also compare accuracy of the two procedures by looking at the ARI comparing to true class label (supplementary Figure 2). We found that random weighting scheme tends to give better results than classical bayesian clustering.

2.2.4. Selecting K. In order to determine the number of clusters and inspired by validitydefined in Ray and Turi (2000), We consider a modified validity= $\frac{\text{intra}}{\text{inter}}$. where $\text{intra} = \frac{1}{N}\sum_{i=1}^{K}\sum_{x\in C_i}||x-z_i||^2$, $\text{inter} = mean(||z_i-z_j||^2)$, i,j=1,2,...K and z_i is the center(medoids) of cluster i. intra is the average of distance of a point to its corresponding cluster center, which measures the compactness of clusters. We made a small change here, in original paper inter was defined as minimum distance between

medoids, we use average instead for the purpose of getting a smoother quantity. **inter** is the average distance of two cluster centers, which measures the separation between clusters. We want to have a small intra-cluster distance and a big inter-cluster distance, consequently we want to minimize







Supplementary Figure 2: comparison between random weighting scheme and bayesian clustering procedure in terms of posterior probabilities that two elements belong to the same class given the whole data and adjusted rand index comparing to the underlying true class label

the validity. From empirical study, we constantly observe a monotone decreasing relation between number of clusters and validity. However this quantity stabilize when K is sufficiently large. The stopping rule for searching K is when validity $K < \epsilon$ is satisfied. We set the default value of ϵ to be 1. As we found DD analysis results to be most consistent with other scRNA method.

2.3. *Double Dirichlet Mixture.* On the double Dirichlet masses, using notation as in Section 2.3 we have density functions:

$$p_{\pi}(\phi, \psi) = q_{\pi}(\Phi_{\pi}, \Psi_{\pi}) \prod_{b \in \pi} [p(\tilde{\phi}_b)p(\tilde{\psi}_b)]$$

with

$$q_{\pi}(\Phi_{\pi}, \Psi_{\pi}) = \frac{\Gamma(\sum_{b \in \pi} \beta_b)}{\prod_{b \in \pi} \Gamma(\beta_b)} \left[\prod_{b \in \pi} \Phi_b^{\beta_b - 1} \right] 1 \left[\Phi_{\pi} = \Psi_{\pi} \right]$$

and

$$p(\tilde{\phi}_b) = \frac{\Gamma(\sum_{k \in b} \alpha_k)}{\prod_{k \in b} \Gamma(\alpha_k)} \prod_{k \in b} \tilde{\phi}_k^{\alpha_k - 1}, \qquad p(\tilde{\psi}_b) = \frac{\Gamma(\sum_{k \in b} \alpha_k)}{\prod_{k \in b} \Gamma(\alpha_k)} \prod_{k \in b} \tilde{\psi}_k^{\alpha_k - 1}.$$

Those computing units will serve as key components for proofing property $1 \sim 6$ in section 2.3 Proof of property 1

Proof. When ϕ and ψ only satisfy the coarsest constrants: $\sum_{i=1}^K \phi_i = \sum_{i=1}^K \psi_i = 1$. ϕ and ψ are independently Dirichlet distributed. When ϕ and ψ satisfy finer constraints, $P(\phi|\psi) \neq P(\phi)$ as there is some subsets b such that $\sum_{i \in b} \phi = \sum_{i \in b} \psi$. So ϕ and ψ are dependent

Proof of property 2

PROOF. $E_{\pi}(\phi_k) = E_{\tilde{\phi}_b}(\tilde{\phi}_k)E_{\Phi}(\Phi_b)$ where b is the block containing subtype index k. As $\tilde{\phi}_b \sim \text{Dirichlet}_{N(b)}[\alpha_b^1]$ and $\Phi_{\pi} \sim \text{Dirichlet}_{N(\pi)}[\beta_{\pi}]$ We have $E_{\tilde{\phi}_b}(\tilde{\phi}_k) = \frac{\alpha_k^1}{\sum_{k' \in b} \alpha_{k'}^1}$ and $E_{\Phi}(\Phi_b) = \frac{\beta_b}{\sum_{b' \in \pi} \beta_{b'}}$. Similarly we could proof the case for $E_{\pi}(\psi_k)$

Proof of property 3

PROOF. t^1/t_{π}^1 is independent with t^2/t_{π}^2 conditioning on t_{π}^1 and t_{π}^2 by the Neutrality property of dirichlet distribution

Proof of property 4

PROOF. For j=1,2, let T_b^j be the vector of t_k^j such that $k \in b$. Recall $t_b^j = \sum_{k \in b} t_k^j$. Without loss of generality, we consider the case condition j=1. We can decompose the density to each blocks by the property of multinomial distribution

$$p_{\pi}(t^{1}|t_{\pi}^{1},y) = \prod_{b \in \pi} (p(T_{b}^{1}|t_{b}^{1},y))$$

and prior predictive function can be obtained via integral out $\tilde{\phi}_b$ given the prior Dirichlet $[\alpha_b^1]$ and $p(T_b^1|\tilde{\phi}_b)$ is multinomial $(\tilde{\phi}_b)$ distributed.

$$\begin{split} p(T_b^1|t_b^1,y) &= \int_{\tilde{\phi}_b} p(T_b^1|\tilde{\phi}_b) p(\tilde{\phi}_b) d\tilde{\phi}_b \\ &= \left\{ \left[\frac{\Gamma(t_b^j+1)}{\prod_{k\in b} \Gamma(t_k^j+1)} \right] \left[\frac{\Gamma(\sum_{k\in b} \alpha_k^j)}{\prod_{k\in b} \Gamma(\alpha_k^j)} \right] \left[\frac{\prod_{k\in b} \Gamma(\alpha_k^j+t_k^j)}{\Gamma(t_b^j+\sum_{k\in b} \alpha_k^j)} \right] \right\} \end{split}$$

Proof. t_{π}^1 and t_{π}^2 given the condition label y are independent identical distributed. $t_{\pi}^1|\Phi\sim \text{multinomial}(\Phi)$

$$p_{\pi}(t_{\pi}^{1}, t_{\pi}^{2}|y) = \int_{\Phi} p(t_{\pi}^{1}|\Phi)p(t_{\pi}^{2}|\Phi)p(\Phi)d\Phi$$

$$= \left[\frac{\Gamma(n_{1}+1)\Gamma(n_{2}+1)}{\prod_{b\in\pi}\Gamma(t_{b}^{1}+1)\Gamma(t_{b}^{2}+1)}\right] \left[\frac{\Gamma(\sum_{b\in\pi}\beta_{b})}{\prod_{b\in\pi}\Gamma(\beta_{b})}\right] \left[\frac{\prod_{b\in\pi}\Gamma(\beta_{b}+t_{b}^{1}+t_{b}^{2})}{\Gamma(n_{1}+n_{2}+\sum_{b\in\pi}\beta_{b})}\right].$$

As prior of Φ is Dirchlet[β] and $n_j = \sum_{b \in \pi} t_b^j$ for j = 1, 2

Lemma 1. If π_2 is not refinement of π_1 then $A_{\pi_1} \cap A_{\pi_2}$ is a lower dimensional subset of A_{π_2}

in order to proof property 6 we gave a lemma of dimensionality of the intersection of two A_{π} s Proof of lemma 1

PROOF. Let V denote the orthogonal space of $\phi - \psi$, when $(\phi, \psi) \in A_{\pi_1} \cap A_{\pi_2}$, and $\dim(A_{\pi_1} \cap A_{\pi_2}) = \dim(\phi - \psi) + \dim(\psi) = 2K - \dim(V) - 1$. Also let $\pi_1 = \{b_1^1, ..., b_s^1\}$, $\pi_2 = \{b_1^2, ..., b_t^2\}$. The corresponding vectors are $v_1^1, ..., v_s^1$ and $v_1^2, ..., v_t^2$. We claim there must be a $b_i^1 \in \pi$ whose corresponding v_i^1 is linear independent with $v_1^2, ..., v_t^2$. If not, for every v_i^1 there exists $\alpha_1^i, ..., \alpha_t^i$ such that

$$v_i^1 = \sum_{i=1}^t \alpha_j^i v_j^2 \tag{*}$$

If $b_j^2 \cap b_i^1 \neq \emptyset$, then multiply v_j^2 on both sides of (*), we obtain $v_i^1 * v_j^2 = \alpha_j^i (v_j^2)^2$, as v_j^2 are orthogonal vectors, and $v_i^1 * v_j^2 > 0$ implies $\alpha_j^i > 0$. Consider $x = f(b_j^2 \setminus b_i^1)$, we have $x * v_i^1 = 0$ and we multiply x on both sides of (*) to obtain $\alpha_j^i v_j^2 * x = 0$, thus x must be zero vector and $b_j^2 \setminus b_i^1 = \emptyset$, which implies $b_j^2 \subset b_i^1$. That is to say when $b_j^2 \cap b_i^1 \neq \emptyset$, b_j^2 must be subset of b_i^1 . So b_i^1 is union of some blocks in π_2 . Which implies π_2 is refinement of π_1 , contradiction.

Consequently there exists $b \in \pi_1$ with v(b) linear independent with v(b'), $b' \in \pi_2$. dim(V) is at least $N(\pi_2) + 1$, dim $(A_{\pi_1} \cap A_{\pi_2}) < \dim(A_{\pi_2})$

Proof of property 6

Proof. by lemma 1, it is easy to verify.

Proof of theorem 3

PROOF. $p_{\pi}(t^1|t_{\pi}^1,y)p_{\pi}(t^2|t_{\pi}^2,y)p_{\pi}(t_{\pi}^1,t_{\pi}^2|y)$ The DDM: $p(\phi,\psi)=\sum_{\pi\in\Pi}p_{\pi}(\phi,\psi)$. we know $p(\phi,\psi|y,z)\propto p(\phi,\psi,y,z)=\sum_{\pi\in\Pi}p(y,z|\phi,\psi)p_{\pi}(\phi,\psi)\omega_{\pi}$ And $p(y,z|\phi,\psi)p_{\pi}(\phi,\psi)=p(y,z|\tilde{\phi},\tilde{\psi},\Phi_{\pi})p(\tilde{\phi})p(\tilde{\psi})p(\Phi_{\pi})$ consider the support of $p_{\pi}(\phi,\psi)$.

Right hand side of the above equation is

$$U_{\pi} = A_1 * A_2 * A_3 * \prod_{k=1}^{K} (\tilde{\phi}_k)^{t_k^1 + \alpha_k^1} (\tilde{\psi}_k)^{t_k^2 + \alpha_k^2} \prod_{b \in \pi} (\Phi_b)^{t_b^1 + t_b^2 + \beta_b}$$

Where A_1 is the product of normalizing terms from multinomial distribution of z^1 and z^2 , $A_1 = \frac{\Gamma(n_1+1)\Gamma(n_2+1)}{\prod_{i=1}^2\prod_{k=1}^K\Gamma(t_k^i+1)}$

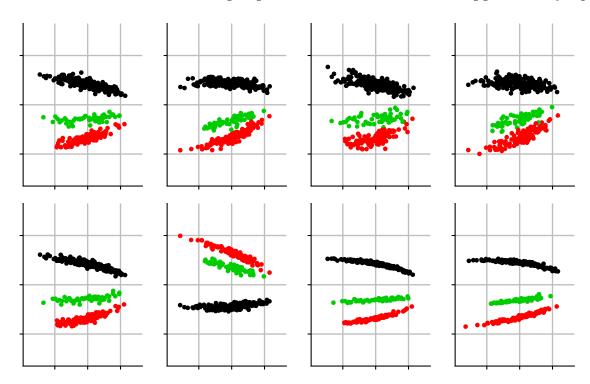
 A_2 is the product of normalizing terms from Dirichlet distribution of $\tilde{\phi}$ and $\tilde{\psi}$, $A_2 = \frac{\Gamma(\sum_{k=1}^K \alpha_k^1 + 1)\Gamma(\sum_{k=1}^K \alpha_k^2 + 1)}{\prod_{i=1}^2 \prod_{k=1}^2 \Gamma(\alpha_i^i + 1)}$

 A_3 is the normalizing term from Dirichle distribution of Φ_{π} , $A_3 = \frac{\Gamma(\sum_{b \in \pi} \beta_b + 1)}{\prod_{b \in \pi} \Gamma(\beta_b + 1)}$. To convert U_{π} to have form similar of p_{π} , We need $U_{\pi} \propto f_1 f_2 f_3$ proportional to product of three Dirichlet densities. We know $f_1 \sim \text{Dirichlet}[\alpha^1 + t^1]$, $f_2 \sim \text{Dirichlet}[\alpha^2 + t^2]$ and $f_3 \sim \text{Dirichlet}[\beta + t^1 + t^2]$. Considering the normalizing factors for f_1 , f_2 and f_3 , and multiplying them with A_1 , A_2 and A_3 . We have the $U_{\pi} = C_{\pi} * f_1 f_2 f_3$. The final normalizing term is $p_{\pi}(t^1 | t_{\pi}^1, y) p_{\pi}(t^2 | t_{\pi}^2, y) p_{\pi}(t_{\pi}^1, t_{\pi}^2 | y)$. Then we have $(\phi, \psi) | y, z \sim \text{DDM}\left[\omega^{\text{post}} = (\omega_{\pi}^{\text{post}}), \alpha^1 + t^1, \alpha^2 + t^2\right]$ and $\omega_{\pi}^{\text{post}} \propto p_{\pi}(t^1 | t_{\pi}^1, y) p_{\pi}(t^2 | t_{\pi}^2, y) p_{\pi}(t_{\pi}^1, t_{\pi}^2 | y) \omega_{\pi}$.

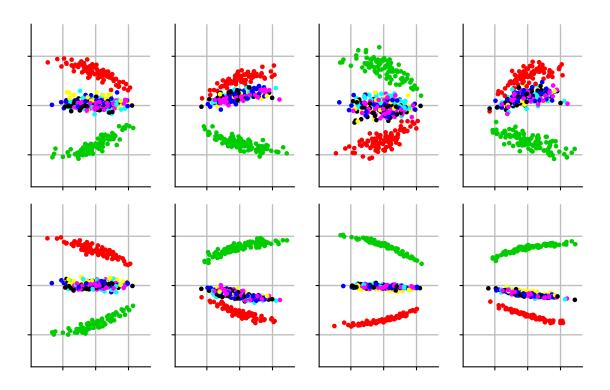
Notice in DDM, we constrained $\beta = \alpha^1 + \alpha^2$.

3. Numerical Experiments.

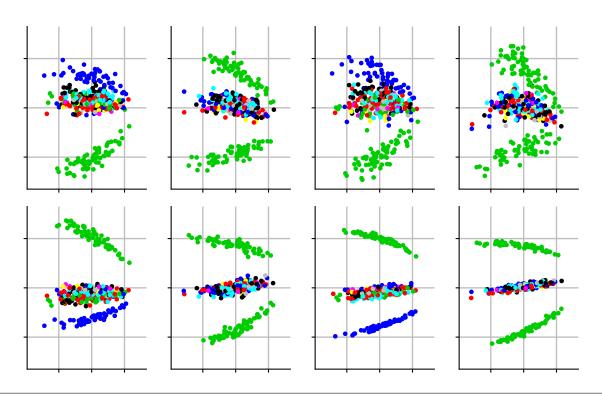
3.1. Synthetic Data. We first look at the pca plots of the simulated data (supplementary figure 2,3,4)



Supplementary Figure 3: first two principal components of transcripts under different parameters for simulated data. Different parameters eters resulted in different degree of separation of subtypes. We have 4 different settings for hyper-parameters of simulation, each setting has 2 replicates K = 3



Supplementary Figure 4: K = 7



Supplementary Figure 5: K = 12

We also have roc curve for the simulated data, each sub-figure is averaged over two replicates under the same parameters setting. scDDboost tends to outperform other methods (supplementary figure 5)



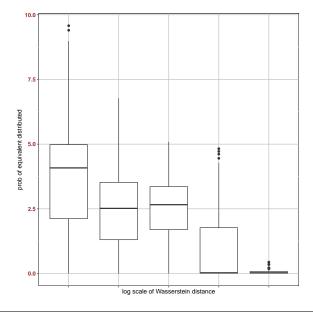
Supplementary Figure 6: Roc curve of the 12 simulation settings, under each setting, TPR and FPR are averaged over two replicates, generally we found scDDboost perform better than other methods

3.2. *Empirical Study.* **Data sets** details for the datasets used in the empirical studies of the main paper and the estimated number of subtypes *K* (supplementary table 1)

| Data set | Conditions | Number of | Organism | Ref | K |
|-----------------------|--|----------------|----------|----------------------------|---|
| | | cells/conditio | | | |
| GSE94383 | 0 min unstim vs 75min stim | 186,145 | human | (Lane et al., 2017) | 9 |
| GSE48968- GPL13112 | BMDC (2h LPS stimulation) vs 6h LPS | 96,96 | mouse | (Shalek et al., 2014) | 4 |
| GSE52529 | T0 vs T72 | 69,74 | human | (Trapnell et al., 2014) | 7 |
| GSE74596 | NKT1 vs NTK2 | 46,68 | mouse | (Engel et al., 2016) | 7 |
| EMTAB2805 | G1 vs G2M | 95,96 | mouse | (Buettner et al., 2015) | 6 |
| GSE71585- GPL13112 | Gad2tdTpositive vs Cux2tdTnegative | 80,140 | mouse | (Tasic et al., 2016) | 4 |
| GSE64016 | G1 vs G2 | 91,76 | human | (Leng et al., 2015) | 6 |
| GSE79102 | patient1 vs patient2 | 51, 89 | human | Kiselev et al. (2017) | 4 |
| GSE45719 | 16-cell stage blastomere vs mid blastocyst cell | 50, 60 | mouse | (Deng et al., 2014) | 4 |
| GSE63818 | Primordial Germ Cells, develop- mental stage: 7 week gestation vs Somatic Cells, developmental stage: 7 week gestation | 40,26 | mouse | (Guo et al., 2015) | 6 |
| GSE75748 | DEC vs EC | 64, 64 | human | (Chu et al., 2016) | 5 |
| GSE84465 | neoplastic cells vs non- neoplastic cells | 546, 664 | human | (Darmanis et al., 2017) | 7 |

Supplementary Table 1

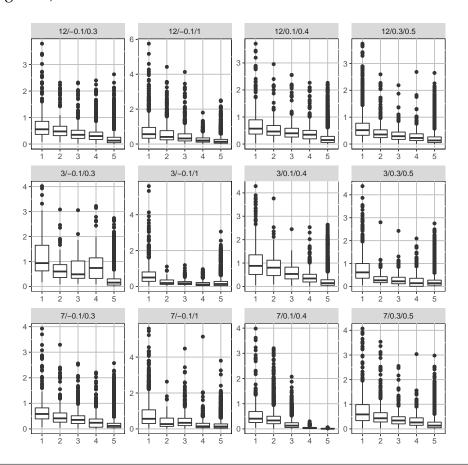
datasets used for comparisons of DD analysis under different methods



Supplementary Figure 7: $P(ED_g|X,y)$ given by scDDboost versus empirical Wasserstein distance. Genes associated with boxes from left to right having $P(ED_g|X,y)$ range from 0 - 0.2, 0.2 - 0.4, 0.4 - 0.6, 0.6 - 0.8, 0.8 - 1, data used: FUCCI

We observed consistent results with scDDboost estimation and wasserstein distance between the

transcripts . Lower probabilities of equivalent distributed are associated with bigger distances. (supplementary Figure 7)



Supplementary Figure 8: $P(ED_g|X,y)$ given by scDDboost versus empirical Wasserstein distance. Genes associated with boxes from left to right having $P(ED_g|X,y)$ range from 0 - 0.2, 0.2 - 0.4, 0.4 - 0.6, 0.6 - 0.8, 0.8 - 1, similar plots as supplementary Fig 7, now is for the simulation cases

3.3. Null cases. datasets used for generating the Null cases (supplementary table 2)

| Data set | Conditions | Number of | Organism |
|---------------|---------------------------|-----------------|----------|
| | | cells/condition | |
| GSE63818null | 7 week gestation | 20,20 | mouse |
| GSE75748null | DEC | 32, 32 | human |
| GSE94383null | T0 | 93, 93 | human |
| GSE48968- | BMDC (2h LPS stimulation) | 48,48 | mouse |
| GPL13112null | | | |
| GSE74596null | NKT1 | 23,23 | mouse |
| EMTAB2805null | G1 | 48,48 | mouse |
| GSE71585- | Gad2tdTpositive | 40,40 | mouse |
| GPL13112null | | | |
| GSE64016null | G1 | 46,45 | human |
| GSE79102null | patient1 | 26, 25 | human |

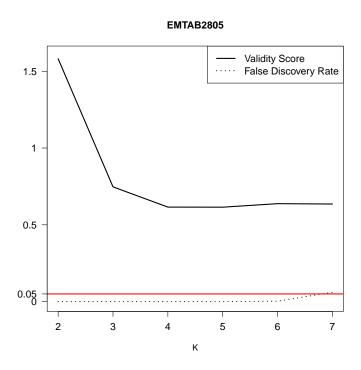
SUPPLEMENTARY TABLE 2

datasets used for null cases, as cells are coming from same biological condition, there should not be any differential distributed genes, any positive call is false positive

4. Random weighting.

4.1. Stability of posterior under random weighting. Number of subtypes *K* is a crucial factor controlling the accuracy of our modeling. Too small *K* may end up in an underfit such that cells within same subtype can still be very different, mean expression change among subtypes is incapable to capture the distribution change for some genes and consequently reducing the power of scDDboost. Too big *K* may end up in an overfit such that two subtypes can be very similar, given we have fixed number of samples (cells), allowing more clusters will introduce may patterns (both for mean expression change and proportion change) to infer. Also notice the limitation of DDM model (see section 4), overestimating *K* in scDDboost may losing FDR control (supplementary Figure 9).

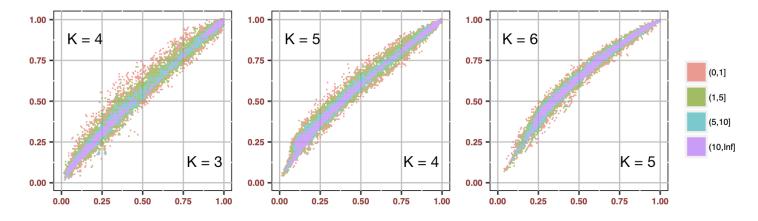
scDDboost may lose FDR control if we keep pushing number of subtypes K bigger.



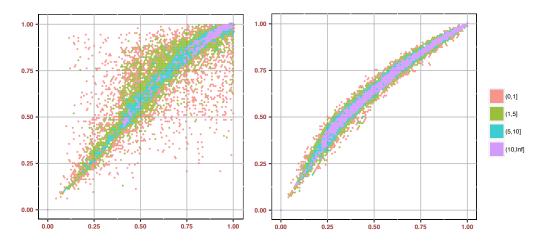
Supplementary Figure 9: under NULL case, using dataset EMTAB2805, when using too big *K* we may lose FDR control (black dashed line shows proportion of false positive identified by scDDboost under 0.05 threshold, while validity score did not vary too much after *K* is greater than 2

From our empirical experience, it would be sufficient to capture the heterogeneity underlying cells with number of clusters not greater than 9. And we generally obtain stable in validity score and PDD simultaneously (supplementary Table 1, supplementary Figure 10)

We demonstrate the change of posterior probabilities of differential distribution given different *K* at data GSE75748. If allowing one more subtype would result in a lot increases in posterior probabilities of DD, which suggests that the number of subtypes is underestimated since we found more distribution differences between conditions given one more mixture component. If posterior inference is stable after increasing the number of subtypes, then we consider previous number of subtypes to be optimal.



Supplementary Figure 10: PDD change under different number of subtypes *K*, dataset used DEC-EC, our rule for selecting *K* tends also to make PDD stabilize



Supplementary Figure 11: DEC-EC, PDD under K = 5 vs. K = 6, left panel is without the randomized distance and right panel is with randomized distance. We increase robustness of our methods through random weighting

5. Posterior consistency. As the density of DDM is computed by product or ratio over bunches of gamma function and gamma function is not easy to direct work on it and derive limiting theorem. To proof theorem 4, we need a crucial lemma which gave us an approximation to the gamma function, namely

Lemma 2. For $x \ge 1$, $\frac{x^{x-c}}{e^{x-1}} \le \Gamma(x) \le \frac{x^{x-1/2}}{e^{x-1}}$, where c = 0.577215... is the Euler-Mascheroni constant.

PROOF. By (Li and ping Chen, 2007), we have $\frac{x^{x-c}}{e^{x-1}} \le \Gamma(x) \le \frac{x^{x-1/2}}{e^{x-1}}$ for x > 1 and now we added the case when x = 1, $\Gamma(x) = 1$ so that both sides will include the equality case.

Lemma 3. For positive integer n, $\sqrt{2\pi}n^{n+1/2}e^{-n} \le \Gamma(n+1) \le en^{n+1/2}e^{-n}$

We have another two lemmas and theorem 1 and 2 are just proporsition of the lemma

Lemma 4. If $(\phi, \psi) \in A_{\pi_1} \cap A_{\pi_2}$, follow the conditions in theorem 1 then

$$\frac{\omega_{\pi_1}^{post}}{\omega_{\pi_2}^{post}} \xrightarrow[n \to \infty]{a.s.} 0 \quad \text{if } N(\pi_1) < N(\pi_2)$$

PROOF. Recall $\omega_{\pi}^{\text{post}} \propto p_{\pi}(t^{1}|t_{\pi}^{1},y) \ p_{\pi}(t^{2}|t_{\pi}^{2},y) \ p_{\pi}(t_{\pi}^{1},t_{\pi}^{2}|y) \ \omega_{\pi}$. and RHS = $g(\pi,\alpha,\beta,n_{1},n_{2}) f(\pi,t^{1},t^{2},\alpha,\beta)$ and $\frac{\omega_{\pi_{1}}^{\text{post}}}{\omega_{\pi_{2}}^{\text{post}}} = \frac{g(\pi_{1},\alpha,\beta,n_{1},n_{2})}{g(\pi_{2},\alpha,\beta,n_{1},n_{2})} \frac{f(\pi_{1},t^{1},t^{2},\alpha,\beta)}{f(\pi_{2},t^{1},t^{2},\alpha,\beta)}$ where

$$g(\pi, t^{1}, t^{2}, \alpha, \beta) = \left[\prod_{j=1}^{2} \prod_{b \in \pi} \frac{\Gamma(\Sigma_{k \in b} \alpha_{k}^{j})}{\prod_{k \in b} \Gamma(\alpha_{k}^{j})}\right] \frac{\Gamma(n_{1}+1)\Gamma(n_{2}+1)}{\prod_{b \in \pi} \Gamma(\beta_{b})} \frac{\Gamma(\Sigma_{b \in \pi} \beta_{b})}{\Gamma(n_{1}+n_{2}+\Sigma_{b \in \pi} \beta_{b})}$$
$$f(\pi, t^{1}, t^{2}, \alpha, \beta) = \left[\prod_{j=1}^{2} \prod_{b \in \pi} \frac{1}{\prod_{k \in b} \Gamma(t_{k}^{j}+1)} \frac{\prod_{k \in b} \Gamma(\alpha_{k}^{j}+t_{k}^{j})}{\Gamma(t_{b}^{j}+\Sigma_{k \in b} \alpha_{k}^{j})}\right] \prod_{b \in \pi} \Gamma(\beta_{b}+t_{b}^{1}+t_{b}^{2})$$

For notation simplicity, we use the abbreviation $g(\pi)$, $f(\pi)$ to substitute $g(\pi,\alpha,\beta,n_1,n_2)$, $f(\pi,t^1,t^2,\alpha,\beta)$. We take \log on $\frac{\omega_{\pi_1}^{post}}{\omega_{\pi_2}^{post}}$, denote it as LR. $LR = \ln g(\pi_1) - \ln g(\pi_2) + \ln f(\pi_1) - \ln f(\pi_2)$. Denote $C(\pi_1,\pi_2,\alpha,\beta) = \ln g(\pi_1) - \ln g(\pi_2)$, $C(\pi_1,\pi_2,\alpha,\beta)$ does not change with sample size n_1,n_2 and is a constant determined by partition π_1,π_2 and hyper parameters α,β . For further convenience of notation let $h(x) = \ln \Gamma(x)$ and $\gamma_b^j = \Sigma_{k \in b} \alpha_k^j$. Denote $R(\pi_1,\pi_2,t^1,t^2,\alpha,\beta) = \ln f(\pi_1) - \ln f(\pi_2)$. And removing the common part of $f(\pi_1)$ and $f(\pi_2)$, we have

$$R(\pi_1, \pi_2, t^1, t^2, \alpha, \beta) = d(\pi_1, t^1, t^2, \alpha, \beta) - d(\pi_2, t^1, t^2, \alpha, \beta)$$

where

$$d(\pi, t^{1}, t^{2}, \alpha, \beta) = \sum_{b \in \pi} h(\beta_{b} + t_{b}^{1} + t_{b}^{2}) - \sum_{j=1}^{2} \sum_{b \in \pi} h(t_{b} + \gamma_{b}^{j})$$

Recall $\beta_b = \gamma_b^1 + \gamma_b^2$ and from lemma 2, $(x-c)\ln(x) - x + 1 \le h(x) \le (x-1/2)\ln(x) - x + 1$ we have

(3)
$$d(\pi, t^1, t^2, \alpha, \beta) \ge \sum_{b \in \pi} (\beta_b + t_b^1 + t_b^2 - c) \ln(\beta_b + t_b^1 + t_b^2) - \sum_{j=1}^2 \sum_{b \in \pi} (t_b^j + \gamma_b^j - 1/2) \ln(t_b^j + \gamma_b^j) + N(\pi)$$

(4)
$$d(\pi, t^1, t^2, \alpha, \beta) \leq \sum_{b \in \pi} (\beta_b + t_b^1 + t_b^2 - 1/2) \ln(\beta_b + t_b^1 + t_b^2) - \sum_{j=1}^2 \sum_{b \in \pi} (t_b^j + \gamma_b^j - c) \ln(t_b^j + \gamma_b^j) + N(\pi)$$

$$\begin{aligned} \text{RHS of (4)} &= \Sigma_b \big[(t_b^1 + \gamma_b^1) \ln(1 + \frac{t_b^2 + \gamma_b^2}{t_b^1 + \gamma_b^1}) + (t_b^2 + \gamma_b^2) \ln(1 + \frac{t_b^1 + \gamma_b^1}{t_b^2 + \gamma_b^2}) \\ &+ (1 - c) \ln(\beta_b + t_b^1 + t_b^2) - 1/2 \big(\ln(1 + \frac{t_b^2 + \gamma_b^2}{t_b^1 + \gamma_b^1}) + \ln(1 + \frac{t_b^1 + \gamma_b^1}{t_b^2 + \gamma_b^2}) \big) \big] + N(\pi) \end{aligned}$$

By Taylor expansion at x = 1, $\ln(x+1) = \ln 2 + 1/2(x-1) - 1/8(x-1)^2 + g(\xi)(x-1)^3$, where $g(\xi)$ is the reminder term of form $\frac{1}{3(1+\xi)^3}$ for $0 < \xi < x$ For a fixed n_1, n_2 , we have

RHS of (4) =
$$(n_1 + n_2)\ln 2 - \sum_{b \in \pi} (1/8(X_b^1 + X_b^2) + g(\xi_b)(Y_b^1 + Y_b^2)) + T(\pi) + N(\pi)$$

where $X_b^1 = \frac{(t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)^2}{t_b^1 + \gamma_b^1}$, $X_b^2 = \frac{(t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)^2}{t_b^2 + \gamma_b^2}$, $Y_b^1 = \frac{(t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)^3}{(t_b^1 + \gamma_b^1)^2}$, $Y_b^2 = \frac{(t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)^3}{(t_b^2 + \gamma_b^2)^2}$ and $T(\pi) = \sum_{b \in \pi} \left[(1 - c) \ln(\beta_b + t_b^1 + t_b^2) - 1/2 (\ln(1 + \frac{t_b^2 + \gamma_b^2}{t_b^1 + \gamma_b^1}) + \ln(1 + \frac{t_b^1 + \gamma_b^1}{t_b^2 + \gamma_b^2})) \right]$ Similarly

RHS of (5) =
$$(n_1 + n_2)\ln 2 - \sum_{b \in \pi} (1/8(X_b^1 + X_b^2) + g(\xi_b)(Y_b^1 + Y_b^2)) + U(\pi) + N(\pi)$$

$$U(\pi) = \sum_{b \in \pi} \left[(2c - 1/2) \ln(\beta_b + t_b^1 + t_b^2) - c(\ln(1 + \frac{t_b^2 + \gamma_b^2}{t_b^1 + \gamma_b^1}) + \ln(1 + \frac{t_b^1 + \gamma_b^1}{t_b^2 + \gamma_b^2})) \right]$$

Using above inequalities, we have

$$R(\pi_1, \pi_2, t^1, t^2, \alpha, \beta) \leq U(\pi_1) - T(\pi_2) - 1/8(\Sigma_{b \in \pi_1}(X_b^1 + X_b^2) - \Sigma_{b \in \pi_2}(X_b^1 + X_b^2)) + \Sigma_{b \in \pi_1} g(\xi_b)(Y_h^1 + Y_h^2) - \Sigma_{b \in \pi_2} g(\xi_b)(Y_h^1 + Y_h^2)$$

 $Y_b^j = \frac{((t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)/\sqrt{n})^3/\sqrt{n}}{((t_b^i + \gamma_b^i)/n)^2}, \text{ by LLN the denominator goes to a constant and by CLT in the numerator } (t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)/\sqrt{n} \rightarrow (t_b^1 - t_b^2)/\sqrt{n} \rightarrow \sqrt{n}[(t_b^1/n - \Phi_b) - (t_b^2/n - \Psi_b)], \text{ which goes to a normal distributed random variables when } \Phi_b = \Psi_b. \text{ So } Y_b^j \text{ is } o_p(1). \text{ Similarly, } X_b^j = \frac{((t_b^1 - t_b^2 + \gamma_b^1 - \gamma_b^2)/\sqrt{n})^2}{t_b^j + \gamma_b^j/n} \text{ is asymptotic gamma}(\chi\text{-square}) \text{ distributed. } g(\xi_b) \text{ has bounded variance, } U(\pi_1) - T(\pi_2) = -\ln(n) \text{ if } N(\pi_2) < N(\pi_1) \text{ as } \ln(\beta_b + t_b^1 + t_b^2) - \ln(\beta_{b'} + t_{b'}^1 + t_{b'}^2) = \ln(\frac{\beta_b + t_b^1 + t_b^2}{n}) - \ln(\frac{\beta_{b'} + t_b^1 + t_b^2}{n}) \rightarrow O(1) \quad a.s. \text{ so we complete the proof}$

Lemma 5. If $(\phi, \psi) \in A_{\pi_1} \cap A_{\pi_2}$, follow the conditions in theorem 1 and further we have α^j , j = 1, 2 be vectors of integers then

$$\frac{\omega_{\pi_1}^{post}}{\omega_{\pi_2}^{post}} \xrightarrow[n \to \infty]{d} v \quad \textit{if } N(\pi_1) = N(\pi_2)$$

PROOF. follow almost same procedure in lemma 4, but instead of using inequalities in lemma 2, we use lemma 3. And we still have

$$d(\pi, t^{1}, t^{2}, \alpha, \beta) = \sum_{b \in \pi} h(\beta_{b} + t_{b}^{1} + t_{b}^{2}) - \sum_{j=1}^{2} \sum_{b \in \pi} h(t_{b} + \gamma_{b}^{j})$$

and by lemma 3

(5)

$$d(\pi, t^1, t^2, \alpha, \beta) \ge \sum_{b \in \pi} (\beta_b + t_b^1 + t_b^2 - 1/2) \ln(\beta_b + t_b^1 + t_b^2) - \sum_{j=1}^2 \sum_{b \in \pi} (t_b^j + \gamma_b^j - 1/2) \ln(t_b^j + \gamma_b^j) + \ln(\sqrt{2\pi}) - 1$$

(6)

$$d(\pi, t^1, t^2, \alpha, \beta) \leq \sum_{b \in \pi} (\beta_b + t_b^1 + t_b^2 - 1/2) \ln(\beta_b + t_b^1 + t_b^2) - \sum_{j=1}^2 \sum_{b \in \pi} (t_b^j + \gamma_b^j - 1/2) \ln(t_b^j + \gamma_b^j) + 1 - \ln(\sqrt{2\pi})$$

$$R(\pi_1, \pi_2, t^1, t^2, \alpha, \beta) \approx D(\pi_1) - D(\pi_2) - 1/8(\Sigma_{b \in \pi_1}(X_b^1 + X_b^2) - \Sigma_{b \in \pi_2}(X_b^1 + X_b^2)) - \Sigma_{b \in \pi_1}g(\xi_b)(Y_b^1 + Y_b^2) - \Sigma_{b \in \pi_2}g(\xi_b)(Y_b^1 + Y_b^2)$$

where
$$D(\pi) = \sum_{b \in \pi} \left[1/2 \ln(\beta_b + t_b^1 + t_b^2) - c(\ln(1 + \frac{t_b^2 + \gamma_b^2}{t_b^1 + \gamma_b^1}) + \ln(1 + \frac{t_b^1 + \gamma_b^1}{t_b^2 + \gamma_b^2})) \right]$$
 And $D(\pi_1) - D(\pi_2)$ is $O(1)$ if $N(\pi_1) = N(\pi_2)$ as $\ln(\beta_b + t_b^1 + t_b^2) - \ln(\beta_{b'} + t_{b'}^1 + t_{b'}^2) = \ln(\frac{\beta_b + t_b^1 + t_b^2}{n_1}) - \ln(\frac{\beta_{b'} + t_b^1 + t_b^2}{n_1}) \to 0$ a.s.

Proof of theorem 4

PROOF. Recall $\sum_{\pi \in \Pi} \omega_{\pi}^{\text{post}} = 1$ and $P(A_{\pi}|y,z) = \sum_{\tilde{\pi} \in \Pi} \omega_{\tilde{\pi}}^{\text{post}} 1[\tilde{\pi} \text{ refines } \pi]$. If $(\phi, \psi) \notin Q$, for all the A_{π} covers (ϕ, ψ) there is one finest π^* with the largest $N(\pi^*)$ and every other π that $(\phi, \psi) \in A_{\pi}$ is coarser than π^* . We get the results of theorem 4 by lemma 4.

1

References.

Buettner, F., Natarajan, K. N., Casale, F. P., Proserpio, V., Scialdone, A., Theis, F. J., Teichmann, S. A., Marioni, J. C. and Stegle, O. (2015). Computational analysis of cell-to-cell heterogeneity in single-cell RNA-sequencing data reveals hidden subpopulations of cells. *Nature Biotechnology* 33 155 EP -.

Chu, L.-F., Leng, N., Zhang, J., Hou, Z., Mamott, D., Vereide, D. T., Choi, J., Kendziorski, C., Stewart, R. and Thomson, J. A. (2016). Single-cell RNA-seq reveals novel regulators of human embryonic stem cell differentiation to definitive endoderm. *Genome Biology* 17 173.

Dahl, D. B. (2009). Modal clustering in a class of product partition models. Bayesian Anal. 4 243-264.

Darmanis, S., Sloan, S. A., Croote, D., Mignardi, M., Chernikova, S., Samghababi, P., Zhang, Y., Neff, N., Kowarsky, M., Caneda, C., Li, G., Chang, S. D., Connolly, I. D., Li, Y., Barres, B. A., Gephart, M. H. and Quake, S. R. (2017). Single-Cell RNA-Seq Analysis of Infiltrating Neoplastic Cells at the Migrating Front of Human Glioblastoma. *Cell reports* 21 1399–1410.

Deng, Q., Ramsköld, D., Reinius, B. and Sandberg, R. (2014). Single-Cell RNA-Seq Reveals Dynamic, Random Monoallelic Gene Expression in Mammalian Cells. *Science* **343** 193–196.

ENGEL, I., SEUMOIS, G., CHAVEZ, L., SAMANIEGO-CASTRUITA, D., WHITE, B., CHAWLA, A., MOCK, D., VIJAYANAND, P. and KRONENBERG, M. (2016). Innate-like functions of natural killer T cell subsets result from highly divergent gene programs. *Nature Immunology* **17** 728 EP --

Guo, F., Yan, L., Guo, H., Li, L., Hu, B., Zhao, Y., Yong, J., Hu, Y., Wang, X., Wei, Y., Wang, W., Li, R., Yan, J., Zhi, X., Zhang, Y., Jin, H., Zhang, W., Hou, Y., Zhu, P., Li, J., Zhang, L., Liu, S., Ren, Y., Zhu, X., Wen, L., Gao, Y. Q., Tang, F. and Qiao, J. (2015). The Transcriptome and DNA Methylome Landscapes of Human Primordial Germ Cells. *Cell* 161 1437–1452.

- KISELEV, V. Y., KIRSCHNER, K., SCHAUB, M. T., ANDREWS, T., YIU, A., CHANDRA, T., NATARAJAN, K. N., REIK, W., BARAHONA, M., GREEN, A. R. and HEMBERG, M. (2017). SC3: consensus clustering of single-cell RNA-seq data. *Nature Methods* 14 483 EP -.
- Lane, K., Van Valen, D., Defelice, M. M., Macklin, D. N., Kudo, T., Jaimovich, A., Carr, A., Meyer, T., Pe'er, D., Boutet, S. C. and Covert, M. W. (2017). Measuring Signaling and RNA-Seq in the Same Cell Links Gene Expression to Dynamic Patterns of NF-B Activation. *Cell Systems* 4 458–469.e5.
- LENG, N., CHU, L.-F., BARRY, C., LI, Y., CHOI, J., LI, X., JIANG, P., STEWART, R. M., THOMSON, J. A. and KENDZIORSKI, C. (2015). Oscope identifies oscillatory genes in unsynchronized single-cell RNA-seq experiments. *Nature Methods* 12 947 EP -.
- LI, X. and PING CHEN, C. (2007). Inequalities for the gamma function. In 2007), Art. 28. [ONLINE: http://jipam.vu.edu.au/ article.php?sid=842.
- RAY, S. and TURI, R. H. (2000). Determination of Number of Clusters in K-Means Clustering and Application in Colour Image Segmentation.
- SHALEK, A. K., SATIJA, R., SHUGA, J., TROMBETTA, J. J., GENNERT, D., LU, D., CHEN, P., GERTNER, R. S., GAUBLOMME, J. T., YOSEF, N., SCHWARTZ, S., FOWLER, B., WEAVER, S., WANG, J., WANG, X., DING, R., RAYCHOWDHURY, R., FRIEDMAN, N., HACOHEN, N., PARK, H., MAY, A. P. and REGEV, A. (2014). Single-cell RNA-seq reveals dynamic paracrine control of cellular variation. *Nature* 510 363 EP -.
- Tasic, B., Menon, V., Nguyen, T. N., Kim, T. K., Jarsky, T., Yao, Z., Levi, B., Gray, L. T., Sorensen, S. A., Dolbeare, T., Bertagnolli, D., Goldy, J., Shapovalova, N., Parry, S., Lee, C., Smith, K., Bernard, A., Madisen, L., Sunkin, S. M., Hawrylycz, M., Koch, C. and Zeng, H. (2016). Adult mouse cortical cell taxonomy revealed by single cell transcriptomics. *Nature Neuroscience* 19 335 EP -.
- Trapnell, C., Cacchiarelli, D., Grimsby, J., Pokharel, P., Li, S., Morse, M., Lennon, N. J., Livak, K. J., Mikkelsen, T. S. and Rinn, J. L. (2014). The dynamics and regulators of cell fate decisions are revealed by pseudotemporal ordering of single cells. *Nature biotechnology* **32** 381–386.