

Advanced Measurement Theory Course Notebook

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Introduction to the Course

Welcome! This is a notebook for ERMA 8350 Advanced Measurement Theory. The class will be using the textbook *Modern Psychometrics with R* (Mair, 2018), which will be the primary source for the course. I will use this notebook to make available additional information to help you learn the material. It may include some examples from the textbook, with some elaborations, additional readings, and some more details about implementing the methods in R. These web-based notes will make it easy for you to use code, by allowing you to copy and paste code found within. Some of you will have experience with R and others not. So I will try to also point you to additional resources that may be helpful. For example, in this preface I will provide links to resources to help you setup R and RStudio. RStudio is a platform to make using R more productive. I will use it extensively in this course.

Software

There are at least two way you can access the software needed for this course. You can use the virtual labs on campus. I know at least the education virtual labs have R and RStudio installed. IF you go this route you can watch the following video. Note you will need Duo setup for this to work.

Using Vlab to acces R/RStudio

A better option if you have a laptop, you can install both programs on your computer. They are both absolutely free and available on all major operating systems, so you will not have to worry about transferring information across computers, limited connection speeds, or other hassles inherent with the VLab route.

The following links take you to videos instructing you how to install them.

Installing R and RStudio

Organizing Projects in RStudio

Resources for Learning R

While such experience is certainly helpful, I do not assume you have prior knowledge of using R. I will demonstrate the use of R and provide (particularly in this notebook) the R code needed to use the methods we will learn. However, even if you have prior experience with R, you should plan to spend time learning to program in R. Some people find this intimidating initially, but most of you will grow to find R programming rewarding, and even fun by the end of the course. But, there will be frustration for sure.

Here are some good places to start learning R:

CRAN

R Packages

R is, among other paradigms, a functional programming language, which means it heavily utilizes functions. R's functions are stored in packages. While base R has a long list of very useful functions, to fully realize the power of R you will have to use additional packages. So, learning how to **install** packages (downloading from the web to your computer) and **loading** packages (making the package's functions accessible to your current R session) are important skills to master.

How To Use These Notes

Before going further, it may be helpful to watch the following video about how to use the code in this notebook with RStudio:

How to Use RStudio with this Notebook

Chapter 1

Classic Test Theory

1.1 Overview

Three important concepts:

1. We can build a model of instruments similar to the way we model conceptual relationships (measurement model vs. structural model).
2. The concept of reliability
3. Measurement models can account for different sources of error.

1.2 Classical True Score Model

The true score model is:

$$X = T + E$$

where X is the **observed score**, T is the **true score**, which is unknown, and E is the **error**

To demonstrate this let's assume we have the following data (R script is at end of this chapter),

```
source("code/simulate_CTTdata.R")
CTTdata
```

	id	time	x1	x2	x3	x4	x5	x6	x7	x8	x9	x10	Tau
1	1	1	3	6	5	3	5	5	4	5	3	3	4
2	1	2	6	3	5	3	4	2	4	4	3	5	4
3	1	3	4	4	2	4	4	3	5	3	4	5	4
4	2	1	3	6	8	6	5	4	5	5	5	5	5
5	2	2	6	4	6	6	4	6	6	5	4	4	5
6	2	3	4	6	6	5	5	5	1	3	6	4	5
7	3	1	6	5	6	6	6	6	9	6	6	5	6

8	3	2	6	6	6	7	5	6	6	6	6	7	6
9	3	3	6	5	8	6	6	6	7	7	5	7	6
10	4	1	4	3	5	4	2	3	3	5	5	2	4
11	4	2	4	5	5	4	5	5	3	5	3	4	4
12	4	3	2	4	4	4	6	4	3	4	5	4	4
13	5	1	5	6	5	4	5	5	5	6	5	6	5
14	5	2	6	6	4	6	4	5	4	5	5	5	5
15	5	3	6	4	5	4	5	5	4	4	5	5	5
16	6	1	6	6	7	8	6	6	7	6	6	4	6
17	6	2	4	5	7	5	5	7	4	5	6	7	6
18	6	3	5	6	6	6	4	5	4	5	7	6	6

where `id` is a variable indicating individual test-takers, `time` indicated which of 3 times each individual was assessed, `x1` - `x10` are the scores on 10 items that comprise the test, and `Tau` is the true value of the individuals ability. I use `Tau` here instead of `T`, because `T` is a protected symbol in R which is short-hand for `TRUE`. Note that we would not know `Tau` in most situations, but because this is simulated data we will pretend we do.

We can create a composite score for the ten items for each individual on each occasion by averaging columns 3 through 12.

```
CTTdata$X <- rowMeans(CTTdata[,3:12])
```

And we can also create `E`, the error with:

```
CTTdata$E <- CTTdata$X - CTTdata$Tau
```

Again, in practice we would not be able to directly compute `E` because we would not know `Tau`, but we will use it to build an understanding of what error is.

Now we have:

```
CTTdata
```

	id	time	x1	x2	x3	x4	x5	x6	x7	x8	x9	x10	Tau	X	E
1	1	1	3	6	5	3	5	5	4	5	3	3	4	4.2	0.2
2	1	2	6	3	5	3	4	2	4	4	3	5	4	3.9	-0.1
3	1	3	4	4	2	4	4	3	5	3	4	5	4	3.8	-0.2
4	2	1	3	6	8	6	5	4	5	5	5	5	5	5.2	0.2
5	2	2	6	4	6	6	4	6	6	5	4	4	5	5.1	0.1
6	2	3	4	6	6	5	5	5	1	3	6	4	5	4.5	-0.5
7	3	1	6	5	6	6	6	6	9	6	6	5	6	6.1	0.1
8	3	2	6	6	6	7	5	6	6	6	6	7	6	6.1	0.1
9	3	3	6	5	8	6	6	6	7	7	5	7	6	6.3	0.3
10	4	1	4	3	5	4	2	3	3	5	5	2	4	3.6	-0.4
11	4	2	4	5	5	4	5	5	3	5	3	4	4	4.3	0.3
12	4	3	2	4	4	4	6	4	3	4	5	4	4	4.0	0.0
13	5	1	5	6	5	4	5	5	5	6	5	6	5	5.2	0.2
14	5	2	6	6	4	6	4	5	4	5	5	5	5	5.0	0.0


```

15 5    3 6 4 5 4 5 5 4 4 5    5 5 4.7 -0.3
16 6    1 6 6 7 8 6 6 7 6 6    4 6 6.2  0.2
17 6    2 4 5 7 5 5 7 4 5 6    7 6 5.5 -0.5
18 6    3 5 6 6 6 4 5 4 5 7    6 6 5.4 -0.6

```

Look over the last three columns and make sure you understand their relation. For example, in the first row, note that X is .2 points above Tau , which is exactly the value of E we computed ($X_1 - T_1 = E_1 = 4.2 - 4 = .2$). The 1 subscript in the previous expression indicated row 1 (i.e. $i = 1$).

```
CTTdata$X_t <- round(ave(CTTdata$X, CTTdata$id, FUN = mean),1)
```

```
CTTdata
```

	id	time	x1	x2	x3	x4	x5	x6	x7	x8	x9	x10	Tau	X	E	X_t
1	1	1	3	6	5	3	5	5	4	5	3	3	4	4.2	0.2	4.0
2	1	2	6	3	5	3	4	2	4	4	3	5	4	3.9	-0.1	4.0
3	1	3	4	4	2	4	4	3	5	3	4	5	4	3.8	-0.2	4.0
4	2	1	3	6	8	6	5	4	5	5	5	5	5	5.2	0.2	4.9
5	2	2	6	4	6	6	4	6	6	5	4	4	5	5.1	0.1	4.9
6	2	3	4	6	6	5	5	5	1	3	6	4	5	4.5	-0.5	4.9
7	3	1	6	5	6	6	6	6	9	6	6	5	6	6.1	0.1	6.2
8	3	2	6	6	6	7	5	6	6	6	6	7	6	6.1	0.1	6.2
9	3	3	6	5	8	6	6	6	7	7	5	7	6	6.3	0.3	6.2
10	4	1	4	3	5	4	2	3	3	5	5	2	4	3.6	-0.4	4.0
11	4	2	4	5	5	4	5	5	3	5	3	4	4	4.3	0.3	4.0
12	4	3	2	4	4	4	6	4	3	4	5	4	4	4.0	0.0	4.0
13	5	1	5	6	5	4	5	5	5	6	5	6	5	5.2	0.2	5.0
14	5	2	6	6	4	6	4	5	4	5	5	5	5	5.0	0.0	5.0
15	5	3	6	4	5	4	5	5	4	4	5	5	5	4.7	-0.3	5.0
16	6	1	6	6	7	8	6	6	7	6	6	4	6	6.2	0.2	5.7
17	6	2	4	5	7	5	5	7	4	5	6	7	6	5.5	-0.5	5.7
18	6	3	5	6	6	6	4	5	4	5	7	6	6	5.4	-0.6	5.7

1.3 Reliability

$$\text{reliability} = \frac{\sigma_T^2}{\sigma_X^2} = \frac{\sigma_T^2}{\sigma_T^2 + \sigma_E^2} = \rho_{XT}^2$$

The reliability is the proportion of variance of T in X , which is also the squared correlation between X and T .

```

Tau <- CTTdata$Tau
X <- CTTdata$X
E <- CTTdata$X - CTTdata$Tau

```

```
var(Tau)/var(X)
```

```
[1] 0.9170806
```

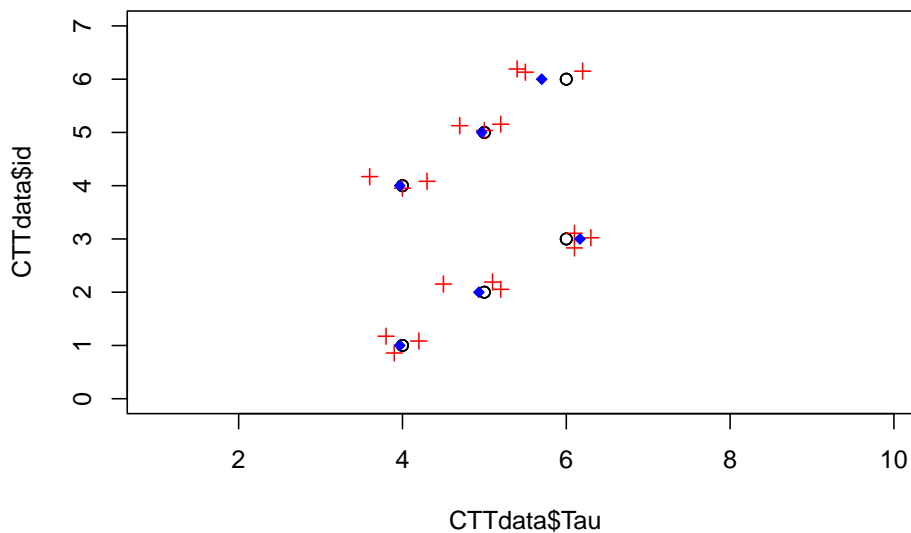
```
var(Tau)/(var(Tau) + var(E))
```

```
[1] 0.8898776
```

```
cor(Tau, X)^2
```

```
[1] 0.886766
```

```
plot(x = CTTdata$Tau, y = CTTdata$id, xlim = c(1,10),
     ylim = c(0,7))
points(x = CTTdata$X, y = jitter(CTTdata$id), pch = 3, col = "red")
points(x = ave(x = CTTdata$X, factor(CTTdata$id), FUN = mean), y = CTTdata$id,
     col = "blue", pch = 18)
```



```
# points(x = CTTdata$X_t, pch = 2, factor(CTTdata$id))
```

1.3.1 Cronbach's α

In the notes for this chapter, I demonstrate aspects of classical test theory, reliability and generalizability theory using data from a study exploring the motivation of R package authors (Mair et al., 2015). This tutorial is based on Chapter 1 of Mair (2018), which can be consulted for a more in depth exposition of the underlying theory. Here I focus on demonstrating some of those concepts in R, as well as describing how to get certain results in R.

First, I load the packages used in this tutorial:

```
# Packages used:
library(psych)
library(MPsychoR)
```

Next, I load the full data set from the MPsychoR package (Mair, 2020), then as in the chapter, I subset the data to only include hybrid motivation items, followed by removing rows with missing values.

```
data("Rmotivation")

# Create data frame with only Hybrid Motivation items.
HybMot <- subset(Rmotivation,
                 select = grep("hyb", names(Rmotivation)))
# Remove rows with any missing data.
HybMot <- na.omit(HybMot)
```

This leads to a data set with 777 authors and 19 items.

```
# How many authors(rows) and items(columns)?
dim(HybMot)
```

```
[1] 777  19
```

```
# Note they are all dichotomous items.
head(HybMot)
```

	hyb1	hyb2	hyb3	hyb4	hyb5	hyb6	hyb7	hyb8	hyb9	hyb10	hyb11	hyb12	hyb13	hyb14
1	1	0	1	0	1	1	0	0	0	1	1	1	1	0
3	0	0	1	0	1	0	0	0	0	1	0	1	1	0
4	1	1	1	1	1	0	1	0	0	1	0	1	1	0
5	1	0	0	1	1	0	0	0	0	1	1	1	1	0
8	1	1	1	1	1	1	1	1	1	1	0	1	1	1
9	1	0	0	1	1	0	0	0	0	1	0	0	1	0

	hyb15	hyb16	hyb17	hyb18	hyb19
1	1	1	1	1	1
3	1	0	0	1	0
4	0	1	1	1	1
5	1	0	1	1	1
8	1	1	1	1	1
9	0	1	1	1	1

```
# Variance/Covariance Matrix
vcmat <- cov(HybMot)
scroll_box(kable(vcmat, digits = 2), width = "100%")
```

	hyb1	hyb2	hyb3	hyb4	hyb5	hyb6	hyb7	hyb8	hyb9	hyb10	hyb11	hyb12
hyb1	0.18	0.06	0.04	0.03	0.03	0.05	0.01	0.05	0.04	0.04	0.03	0.03
hyb2	0.06	0.25	0.06	0.05	0.03	0.05	-0.01	0.04	0.05	0.02	0.04	0.04
hyb3	0.04	0.06	0.23	0.13	0.03	0.05	0.00	0.03	0.05	0.04	0.05	0.06
hyb4	0.03	0.05	0.13	0.21	0.03	0.04	0.01	0.03	0.04	0.03	0.05	0.05
hyb5	0.03	0.03	0.03	0.03	0.11	0.02	0.00	0.01	0.01	0.03	0.03	0.03
hyb6	0.05	0.05	0.05	0.04	0.02	0.24	0.01	0.11	0.15	0.05	0.06	0.06
hyb7	0.01	-0.01	0.00	0.01	0.00	0.01	0.22	0.04	0.01	0.03	0.00	0.02
hyb8	0.05	0.04	0.03	0.03	0.01	0.11	0.04	0.25	0.10	0.06	0.05	0.06
hyb9	0.04	0.05	0.05	0.04	0.01	0.15	0.01	0.10	0.20	0.04	0.04	0.05
hyb10	0.04	0.02	0.04	0.03	0.03	0.05	0.03	0.06	0.04	0.15	0.03	0.06
hyb11	0.03	0.04	0.05	0.05	0.03	0.06	0.00	0.05	0.04	0.03	0.23	0.10
hyb12	0.03	0.04	0.06	0.05	0.03	0.06	0.02	0.06	0.05	0.06	0.10	0.23
hyb13	0.03	0.03	0.02	0.02	0.03	0.02	0.00	0.02	0.01	0.03	0.03	0.04
hyb14	0.03	0.03	0.02	0.02	0.02	0.07	0.00	0.04	0.05	0.02	0.04	0.03
hyb15	0.04	0.03	0.06	0.04	0.04	0.06	0.01	0.06	0.04	0.04	0.10	0.11
hyb16	0.05	0.03	0.02	0.02	0.02	0.05	0.02	0.05	0.04	0.04	0.03	0.04
hyb17	0.04	0.01	0.03	0.03	0.02	0.05	0.02	0.05	0.03	0.04	0.03	0.03
hyb18	0.03	0.00	0.02	0.02	0.01	0.02	0.01	0.03	0.01	0.03	0.01	0.02
hyb19	0.06	0.03	0.04	0.04	0.03	0.07	0.02	0.06	0.05	0.05	0.02	0.04

```
k <- ncol(HybMot)
sigma2_Xi <- tr(vcmat) # trace of matrix or sum(diag(vmat))
sigma2_X <- sum(vcmat)
```

1.3.2 Other Reliability Coefficients

1.4 Generalizability Theory

Generalizability theory, or G-theory for short, is an extension of CTT, which decomposes the one error term in CTT into multiple sources of error called *facets*. These could include sources such as items, raters, or measurement occasions. These were each given a subscript on page 2 of the text.

Before looking at these different sources of error, let's calculate Cronbach's α in a different way, that will allow this decomposition going forward.

We will first need to reshape the data from wide to long format. A great tutorial on reshaping data with the `reshape2` package can be found here:

<https://seananderson.ca/2013/10/19/reshape/>

Basically, we need to transform the data so that instead of each item being in a separate column are reshaped so there is one column with the cell values, and one column that identifies which item the score is from.

```
library("reshape2")
# Add person variable
```

```
Hyb1 <- data.frame(HybMot, person = 1:nrow(HybMot))
Hyblong <- melt(Hyb1, id.vars = c("person"), variable.name = "item")
Hyblong$person <- as.factor(Hyblong$person)
```

1.4.1 Reliability and Generalizability

```
summary(aov(value ~ person + item, data = Hyblong))
```

```
              Df Sum Sq Mean Sq F value Pr(>F)
person        776  663.0    0.85   5.549 <2e-16 ***
item           18  573.8   31.88 207.048 <2e-16 ***
Residuals    13968 2150.5    0.15
---
```

```
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

From this output we can calculate Cronbach's α with the following values:

```
MSp <- 0.85
MSe <- 0.15
```

```
alpha <- (MSp - MSe)/MSp
print(alpha, digits = 2)
```

```
[1] 0.82
```

```
ICC(HybMot)
```

```
Call: ICC(x = HybMot)
```

```
Intraclass correlation coefficients
```

	type	ICC	F	df1	df2	p	lower bound
Single_raters_absolute	ICC1	0.15	4.4	776	13986	7.2e-279	0.14
Single_random_raters	ICC2	0.16	5.5	776	13968	0.0e+00	0.14
Single_fixed_raters	ICC3	0.19	5.5	776	13968	0.0e+00	0.17
Average_raters_absolute	ICC1k	0.77	4.4	776	13986	7.2e-279	0.75
Average_random_raters	ICC2k	0.78	5.5	776	13968	0.0e+00	0.75
Average_fixed_raters	ICC3k	0.82	5.5	776	13968	0.0e+00	0.80
	upper bound						
Single_raters_absolute		0.17					
Single_random_raters		0.18					
Single_fixed_raters		0.21					
Average_raters_absolute		0.79					
Average_random_raters		0.81					
Average_fixed_raters		0.84					

```
Number of subjects = 777      Number of Judges = 19
See the help file for a discussion of the other 4 McGraw and Wong estimates,
```

```

icchyb <- ICC(HybMot)
sqrt((0.85-0.15)/19)

[1] 0.191943
sqrt((31.88-0.15)/777)

[1] 0.2020806
library("lme4")

Loading required package: Matrix
VarCorr(lmer(value ~ (1|person) + (1|item), data = Hyblong))

      Groups      Name      Std.Dev.
person  (Intercept) 0.19200
item    (Intercept) 0.20206
Residual                                0.39238

library("gtheory")
gfit <- gstudy(data = Hyblong, formula = value ~ (1|person) + (1|item))
dfit <- dstudy(gfit, colname.objects = "person", colname.scores = "value",
              data = Hyblong)
round(dfit$generalizability, 3)

[1] 0.82

```

1.4.2 Multiple Sources of Error

Generalizability theory acknowledges that multiple sources of error can impact scores simultaneously, and allow for estimating the effects of each (Raykov and Marcoulides, 2011). These various sources of error, or facets (e.g., items, raters, measurement occasions). All measurements of behavior are conceptualized as being sampled from a *universe* of admissible observations (Raykov and Marcoulides, 2011). If the observed score is expected to vary across a facet (e.g. vary across occasions, or vary depending on the items included, or the rater scoring), then that facet is a defining characteristic of the universe. The idea of reliability is replaced with the idea of generalizability, which, instead of asking how accurately observed scores can reflect the true score (CTT), generalizability theory asks how accurately observed scores allow us to generalize about behavior of an individual in a particular universe.

Below is the code from the Mair (2018) text.

```

library(gtheory)

data("Lakes")
phydat <- subset(Lakes, subtest == "physical")

```

```

phydat$item <- droplevels(phydat$item)
head(phydat)

  personID raterID item score subtest
12611      1       7  phy1     5 physical
12612      1       1  phy1     5 physical
12613      1       3  phy1     5 physical
12614      1       8  phy1     4 physical
12615      1       5  phy1     6 physical
12616      2       3  phy1     5 physical

formula <- score ~ (1|personID) + (1|raterID) + (1|item) +
  (1|personID:raterID) + (1|personID:item) + (1|raterID:item)
gfit <- gstudy(formula = formula, data = phydat)
gfit

$components
      source      var percent  n
1 personID:raterID 0.127298630   10.7 1
2  personID:item 0.151229568   12.7 1
3    personID 0.458766113   38.5 1
4  raterID:item 0.008802081    0.7 1
5    raterID 0.139048763   11.7 1
6      item 0.033388424    2.8 1
7   Residual 0.272703419   22.9 1

attr(,"class")
[1] "gstudy" "list"

dfit <- dstudy(gfit, colname.objects = "personID", colname.scores = "score",
  data = phydat)
dfit$components

      source      var percent  n
1 personID:raterID 0.0254597259    4.3 5
2  personID:item 0.0504098560    8.5 3
3    personID 0.4587661131   77.4 1
4  raterID:item 0.0005868054    0.1 15
5    raterID 0.0278097526    4.7 5
6      item 0.0111294746    1.9 3
7   Residual 0.0181802279    3.1 15

dfit$var.error.abs

[1] 0.1335758

dfit$sem.abs

```

```
[1] 0.3654803
dfit$var.error.rel

[1] 0.09404981
dfit$sem.rel

[1] 0.3066754
dfit$dependability

[1] 0.7744954
dfit$generalizability

[1] 0.8298714
```

1.5 Additional Readings

For more information of G theory, see Raykov and Marcoulides (2011). For an example using the R package *lavaan* with G theory, see Jorgensen (2021).

1.6 R Scripts

1.6.1 Simulating CTT data

```
#-----
# Title: simulate_CTTdata
# Author: William Murrah
# Description: Simulate data to demonstrate CTT and reliability
# Created: Monday, 09 August 2021
# R version: R version 4.1.0 (2021-05-18)
# Project(working) directory: /Users/wmm0017/Projects/Courses/AdvancedMeasurementTheory
#-----

simx <- function(truescore, sigmax = 1) {
  x <- rnorm(18, truescore, sigmax)
  return(round(x))
}
id <- rep(1:6, each = 3)
Tau <- rep(rep(4:6, each = 3), 2)
set.seed(20210805)
CTTdata <- data.frame(
  id = id,
  time = rep(1:3, 6),
  x1 = simx(Tau),
```



```
x2 = simx(Tau),  
x3 = simx(Tau),  
x4 = simx(Tau),  
x5 = simx(Tau),  
x6 = simx(Tau),  
x7 = simx(Tau),  
x8 = simx(Tau),  
x9 = simx(Tau),  
x10 = simx(Tau),  
Tau = Tau  
)  
rm(id, Tau, simx)
```


Chapter 2

Factor Analysis

2.1 Correlation Coefficient

Pearson product-moment correlation:

$$r_{xy} = \frac{\sum_{n=1}^n (x_k - \bar{x})(y_i - \bar{y})}{\sqrt{\sum_{n=1}^n (x_i - \bar{x})^2} \sqrt{\sum_{n=1}^n (y_i - \bar{y})^2}} = \frac{s_{xy}}{s_x s_y}.$$

The equation looks very daunting, until you see that it is just the covariance of x and y divided by the product of their standard deviations.

```
library("MPsychorR")
data("YouthDep")
item1 <- YouthDep[, 1]
levels(item1) <- c("0", "1", "1")
item2 <- YouthDep[, 14]
levels(item2) <- c("0", "1", "1")
table(item1, item2)
```

```
      item2
item1    0    1
0  1353  656
1   115  166
```

```
## ----- correlation coefficients
```

```
library("psych")
tetcor <- tetrachoric(cbind(item1, item2))
```

```
tetcor
```

```
Call: tetrachoric(x = cbind(item1, item2))
tetrachoric correlation
```

```

      item1 item2
item1 1.00
item2 0.35  1.00

with tau of
item1 item2
  1.16  0.36
item1 <- YouthDep[, 1]
item2 <- YouthDep[, 14]
polcor <- polychoric(cbind(item1, item2))
polcor

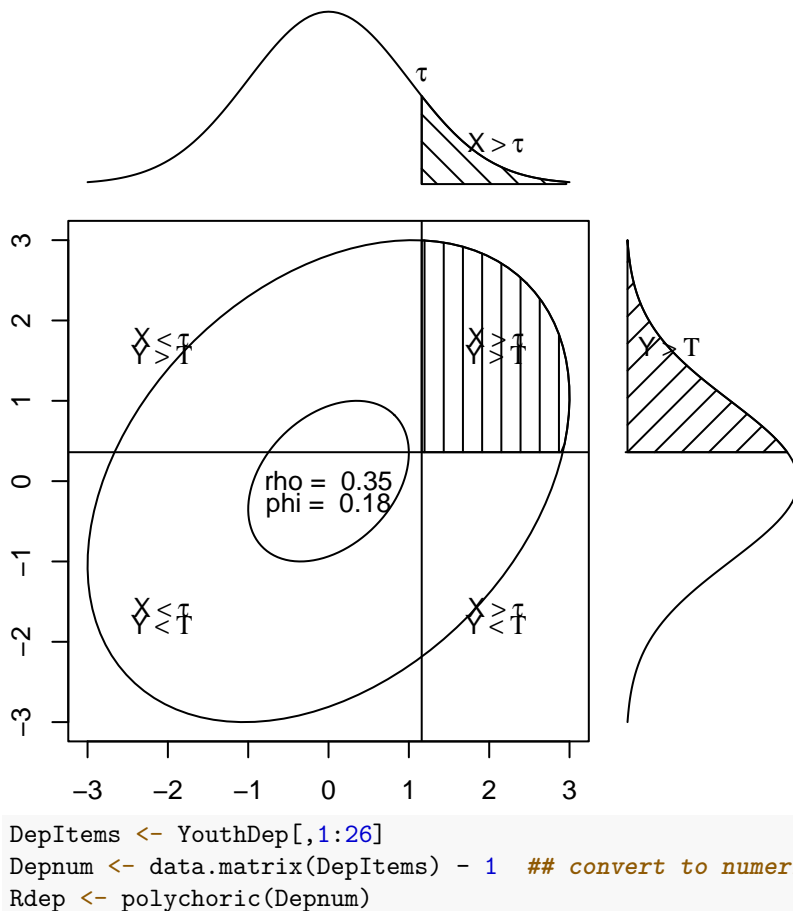
```

```

Call: polychoric(x = cbind(item1, item2))
Polychoric correlations
      item1 item2
item1 1.00
item2 0.33  1.00

with tau of
      1  2
item1 1.16 2.3
item2 0.36 1.2
draw.tetra(r = .35, t1 = 1.16, t2 = .36)

```



2.2 Think about these situations

What do you do when you have a large number of variables you are considering as predictors of a dependent variable?

- Often, subsets of these variables are measuring the same, or very similar things.
- We might like to reduce the variables to a smaller number of predictors.

What if you are developing a measurement scale and have a large number of items you think measure the same construct

- You might want to see how strongly the items are related to the construct.

2.3 Solutions

1. Principal Components Analysis

- transforming the original variables into a new set of linear combinations (principal components).

2. Factor Analysis

- setting up a mathematical model to estimate the number of factors

2.4 Principal Components Analysis

- Concerned with explaining variance-covariance structure of a set of variables.
- PCA attempts to explain as much of the total variance among the observed variables as possible with a smaller number of components.
- Because the variables are standardized prior to analysis, the total amount of variance available is the number of variables.
- The goal is **data reduction** for subsequent analysis.
- Variables *cause* components.
- Components are not representative of any underlying theory.

2.5 Factor Analysis

- The goal is understanding underlying constructs.
- Uses a modified correlation matrix (reduced matrix)
- factors *cause* the variables.
- Factors represent theoretical constructs.
- Focuses on the common variance of the variables, and purges the unique variance.

2.6 Components

The principal components partition the total variance (the sum of the variances of the original variables) by finding the linear combination of the variables that account for the maximum amount of variance:

$$PC1 = a_{11}x_1 + a_{12}x_2 \dots a_{1p}x_p,$$

This is repeated as many times as there are variables.

2.7 PC Extraction

draw pretty pictures on the board

2.8 Eigenvalues

Eigenvalues represent the variance in the variables explained by the success components.

2.9 Determining the Number of Factors

1. Kaiser criterion: Retain only factors with eigenvalues > 1 . (generally accurate)
2. Scree plot: plot eigenvalues and drop factors after leveling off.
3. Parallel analysis: compare observed eigenvalues to parallel set of data from randomly generated data. Retain factors in original if eigenvalue is greater than random eigenvalue.
4. Factor meaningfulness is also very important to consider.

2.10 Example data

```
lower <- "
1.00
0.70 1.00
0.65 0.66 1.00
0.62 0.63 0.60 1.00
"
cormat <- getCov(lower, names = c("d1", "d2", "d3", "d4"))
cormat
```

```
      d1  d2  d3  d4
d1 1.00 0.70 0.65 0.62
d2 0.70 1.00 0.66 0.63
d3 0.65 0.66 1.00 0.60
d4 0.62 0.63 0.60 1.00
```

2.11 Kaiser

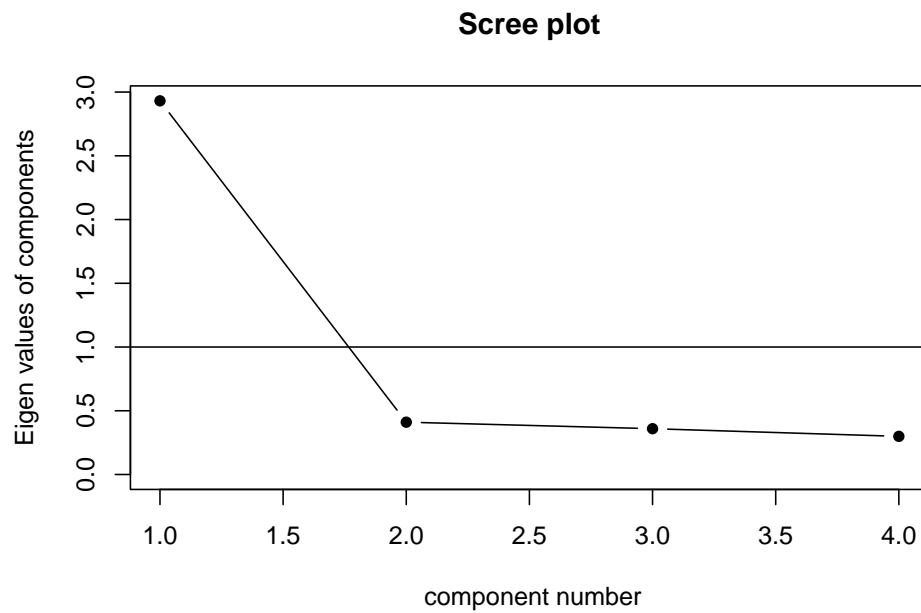
Retain factors with eigenvalues greater than 1

```
eigen(cormat)$values
```

```
[1] 2.9311792 0.4103921 0.3592372 0.2991916
```

```
scree(cormat, factors = FALSE)
```

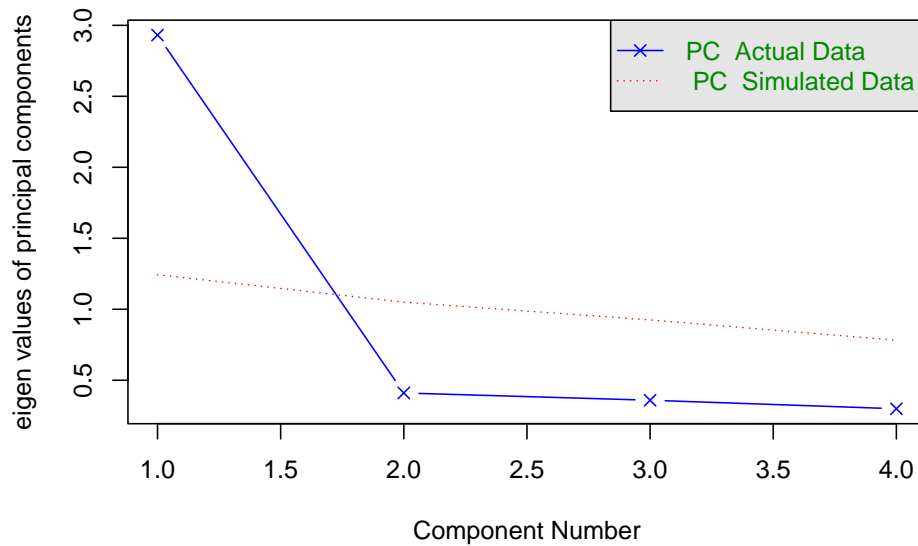
2.12 Scree Plot



```
fa.parallel(cormat, fa = "pc")
```

2.13 Horn's Parallel Analysis

Parallel Analysis Scree Plots

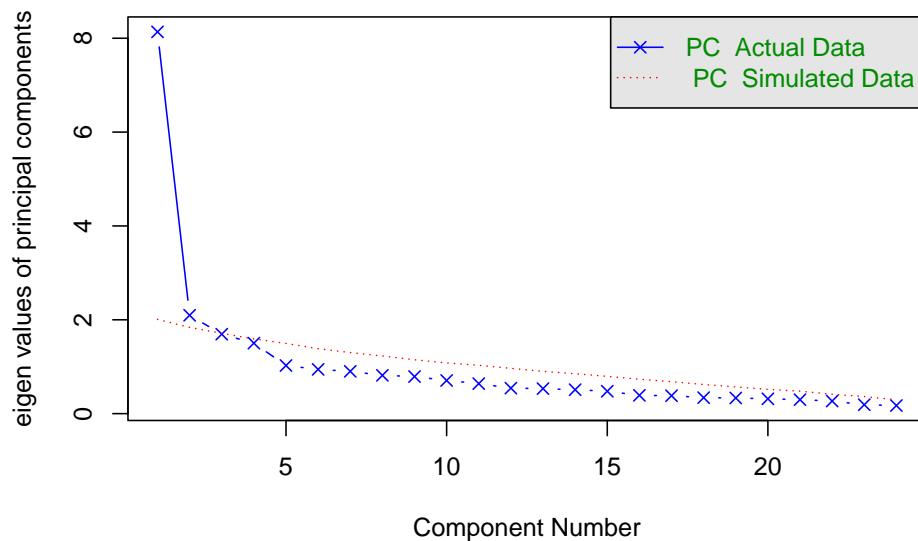


Parallel analysis suggests that the number of factors = NA and the number of components = 1

2.14 Another example

```
fa.parallel(Harman74.cor$cov, fa = "pc")
```

Parallel Analysis Scree Plots



Parallel analysis suggests that the number of factors = NA and the number of components

2.15 Rotation

- Principal components are derived to maximize the variance accounted for (data reduction).
- Rotation is done to make the factors more interpretable (i.e. meaningful).
- Two major classes of rotation:
 - Orthogonal - new factors are still uncorrelated, as were the initial factors.
 - Oblique - new factors are allowed to be correlated.

Essentially reallocates the loadings. The first factor may not be the one accounting for the most variance.

2.16 Orthogonal Rotation

1. **Quartimax** - idea is to clean up the *variables*. Rotation done so each variable loads mainly on one factor. Problematic if there is a general factor on which most or all variables load on (think IQ).
2. **Varimax** - to clean up *factors*. So each factor has high correlation with a smaller number of variables, low correlation with the other variables. Generally makes interpretation easier.

2.17 Oblique Rotation

- Often correlated factors are more reasonable.
- Therefore, oblique rotation is often preferred.
- But interpretation is more complicated.

2.18 Factor Matrices

1. Factor pattern matrix:
 - includes *pattern coefficients* analogous to standardized partial regression coefficients.
 - Indicated the unique importance of a factor to a variable, holding other factors constant.
2. Factor structure matrix:
 - includes *structure coefficients* which are simple correlations of the variables with the factors.

2.19 Which matrix should we interpret?

- When orthogonal rotation is used interpret *structural coefficients* (but they are the same as pattern coefficients).
- When oblique rotation is used pattern coefficients are preferred because they account for the correlation between the factors and they are parameters of the correlated factor model (which we will discuss next class).

2.20 Which variables should be used to interpret each factor?

- The idea is to use only those variables that have a strong association with the factor.
- Typical thresholds are $|.30|$ or $|.40|$.
- Content knowledge is critical.

2.21 Examples

Let's look at some examples

2.22 Steps in Factor Analysis

1. Choose extraction method
 - So far we've focused on PCA
2. Determine the number of components/factors
 - Kaiser method: eigenvalues > 1
 - Scree plot: All components before leveling off
 - Horn's parallel analysis: components/factors greater than simulated values from random numbers
3. Rotate Factors
 - Orthogonal
 - Oblique
4. Interpret Components/Factors

2.23 Tom Swift's Electric Factor Analysis Factory

"Little Jiffy" method of factor analysis

1. Extraction method : PCA
2. Number of factors: eigenvalues > 1
3. Rotation: orthogonal(varimax)

Table 2.1: Functional Definitions of Tom Swift's Original 11 Variables

Dimension	Derivation
Thickness	x
Width	y
Length	z
Volume	xyz
Density	d
Weight	xyzd
Surface area	$2(xy + xz + yz)$
Cross-section	yz
Edge length	$4(x + y + z)$
Diagonal length	(x^2)
Cost/lb	c

4. Interpretation

2.24 Metal Boxes

```

'data.frame':  63 obs. of  11 variables:
 $ thick   : num  1.362 2.385 3.101 0.934 0.845 ...
 $ width   : num  1.71 2.83 4.32 3.2 3.84 ...
 $ length  : num  2.93 5.01 5.99 4.15 4.09 ...
 $ volume  : num  6.02 30.2 72.01 11.78 16.1 ...
 $ density : int  10 7 16 22 11 16 11 21 6 13 ...
 $ weight  : num  60 210 1152 264 176 ...
 $ surface : num  22 62.1 108.2 38 48 ...
 $ crosssec: num  5.87 15.06 23.53 12.02 16.13 ...
 $ edge    : num  23.9 39.9 51.5 31.9 36.1 ...
 $ diagonal: num  196 1444 3721 676 1089 ...
 $ cost    : num  4.48 2.37 9.77 22.21 15.86 ...

```

Dimension	Derivation
Thickness	x
Width	y
Length	z
Volume	xyz
Density	d
Weight	$xyzd$
Total surface area	$2(xy + xz + yz)$
Cross-sectional area	yz
Total edge length	$4(x + y + z)$
Internal diagonal length	$(x^2 + y^2 + z^2)^{1/2}$
Cost per pound	c

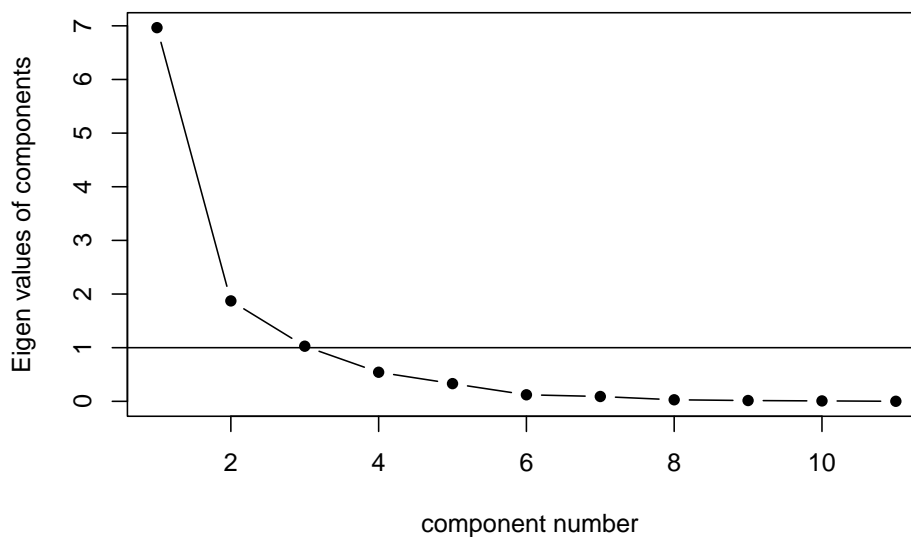
Table 2.2: Correlations between dimensions

	thick	width	length	volume	density	weight	surface	crossec	edge	diagonal	cost
thick	1.00	0.49	0.24	0.84	-0.13	0.59	0.74	0.46	0.61	0.51	-0.02
width	0.49	1.00	0.61	0.77	-0.15	0.55	0.87	0.92	0.88	0.78	0.03
length	0.24	0.61	1.00	0.58	-0.02	0.45	0.72	0.83	0.84	0.86	-0.02
volume	0.84	0.77	0.58	1.00	-0.22	0.65	0.97	0.81	0.87	0.85	-0.11
density	-0.13	-0.15	-0.02	-0.22	1.00	0.44	-0.20	-0.15	-0.15	-0.18	0.62
weight	0.59	0.55	0.45	0.65	0.44	1.00	0.65	0.56	0.61	0.57	0.24
surface	0.74	0.87	0.72	0.97	-0.20	0.65	1.00	0.92	0.97	0.91	-0.07
crossec	0.46	0.92	0.83	0.81	-0.15	0.56	0.92	1.00	0.96	0.93	-0.03
edge	0.61	0.88	0.84	0.87	-0.15	0.61	0.97	0.96	1.00	0.92	-0.04
diagonal	0.51	0.78	0.86	0.85	-0.18	0.57	0.91	0.93	0.92	1.00	-0.12
cost	-0.02	0.03	-0.02	-0.11	0.62	0.24	-0.07	-0.03	-0.04	-0.12	1.00

2.25 Correlations

2.26 Eigenvalues > 1

Scree plot



2.27 Orthogonal Rotation

Loadings:

	RC1	RC3	RC2	RC5	RC4
thick		0.968			
width				0.734	
length	0.986				
volume		0.754			
density			0.864		
weight					
surface	0.703				
crosssec	0.829				
edge	0.819				
diagonal	0.875				
cost					0.955

	RC1	RC3	RC2	RC5	RC4
SS loadings	4.425	2.662	1.318	1.225	1.106
Proportion Var	0.402	0.242	0.120	0.111	0.101
Cumulative Var	0.402	0.644	0.764	0.876	0.976

2.28 Orthogonal Rotation with Loadings $> .70$

Loadings:

	RC1	RC3	RC2

```

thick      0.947
width      0.801
length     0.936
volume     0.744
density    0.930
weight
surface    0.792
crosssec   0.942
edge       0.892
diagonal   0.905
cost       0.841

          RC1  RC3  RC2
SS loadings 5.298 2.699 1.868
Proportion Var 0.482 0.245 0.170
Cumulative Var 0.482 0.727 0.897

```

2.29 R Code for Chapter 2

```

## ----- Chapter 2: Factor Analysis -----
library("MPSychoR")
data("YouthDep")
item1 <- YouthDep[, 1]
levels(item1) <- c("0", "1", "1")
item2 <- YouthDep[, 14]
levels(item2) <- c("0", "1", "1")
table(item1, item2)

## ----- correlation coefficients
library("psych")
tetcor <- tetrachoric(cbind(item1, item2))
tetcor
item1 <- YouthDep[, 1]
item2 <- YouthDep[, 14]
polcor <- polychoric(cbind(item1, item2))
polcor

DepItems <- YouthDep[,1:26]
Depnum <- data.matrix(DepItems) - 1 ## convert to numeric
Rdep <- polychoric(Depnum)

data("Rmotivation")
vind <- grep("ext|int", colnames(Rmotivation))
Rmotivation1 <- Rmotivation[, vind]
Rmot1 <- tetrachoric(Rmotivation1, smooth = FALSE)
tail(round(eigen(Rmot1$rho)$values, 3))
Rmot <- tetrachoric(Rmotivation1)
tail(round(eigen(Rmot$rho)$values, 3))

## ----- exploratory factor analysis
motFA <- fa(Rmot$rho, nfactors = 2, rotate = "none", fm = "ml")
print(motFA$loadings, cutoff = 0.2)
round(motFA$communality, 2)

motFA2 <- fa(Rmot$rho, nfactors = 2, rotate = "varimax", fm = "ml")
plot(motFA$loadings, asp = 1, xlim = c(-0.2, 0.9), ylim = c(-0.5, 0.9), type = "n", xlab = "Factor 1", ylab = "Factor 2")

```

```

text(motFA$loadings, labels = rownames(motFA$loadings), cex = 0.8, col = "gray")
abline(h = 0, v = 0, col = "lightgray", lty = 2)
text(motFA2$loadings, labels = rownames(motFA2$loadings), col = 1, cex = 0.8)
legend("bottomleft", legend = c("rotated", "unrotated"), col = c("black", "gray"), pch = 19)

Rmot2 <- tetrachoric(Rmotivation[,1:36])
motFA3 <- fa(Rmot2$rho, nfactors = 3, rotate = "oblimin", fm = "ml")
motFA3$loadings
round(motFA3$Phi, 3)

motFA2 <- fa(Rmotivation1, nfactors = 2, rotate = "varimax", cor = "tet", fm = "ml", scores = "regression",
            missing = TRUE, impute = "median")
dim(motFA2$scores)

Rdep <- polychoric(Depnum)$rho
evals <- eigen(Rdep)$values
scree(Rdep, factors = FALSE)
(evals/sum(evals)*100)[1:2]

set.seed(123)
resPA <- fa.parallel(Depnum, fa = "pc", cor = "poly", fm = "ml")
resvss <- vss(Rdep, fm = "ml", n.obs = nrow(Depnum), plot = FALSE)
resvss

fadep <- fa(Depnum, 1, cor = "poly", fm = "ml")
summary(fadep)

resnf <- nfactors(Depnum, n = 8, fm = "ml", cor = "poly")
resnf

## ----- Bayesian exploratory factor analysis
library("MPSychoR")
library("corrplot")
library("BayesFM")
data("Privacy")
Privstd <- scale(Privacy)
corrplot(cor(Privstd))

Nid <- 2 ## minimum number of variables per factor
pmax <- trunc(ncol(Privstd)/Nid) ## maximum number of factors
pmax

set.seed(123)
Rsim <- simul.R.prior(pmax, nu0 = pmax + c(1, 2, 5, 7, 10))
plot(Rsim)

Ksim <- simul.nfac.prior(nvar = ncol(Privstd), Nid = Nid, Kmax = pmax, kappa = c(.1, .2, .5, 1))
plot(Ksim)

set.seed(222)
fitbfa <- befa(Privstd, Nid = 2, Kmax = pmax, nu0 = 10, kappa = 0.2, kappa0 = 0.1, xi0 = 0.1,
            burnin = 5000, iter = 50000)
fitbfa <- post.column.switch(fitbfa) ## column reordering
fitbfa <- post.sign.switch(fitbfa) ## sign switching
sumbfa <- summary(fitbfa)

## ----- confirmatory factor analysis

```



```

library("MPSychoR")
library("lavaan")
data("Rmotivation")
vind <- grep("ext|int", colnames(Rmotivation)) ## ext/int items
Rmot <- na.omit(Rmotivation[, vind])
mot_model <- '
  extrinsic =~ ext1 + ext2 + ext3 + ext4 + ext5 + ext6 +
              ext7 + ext8 + ext9 + ext10 + ext11 + ext12
  intrinsic =~ int1 + int2 + int3 + int4 + int5'
fitMot <- lavaan::cfa(mot_model, data = Rmot, ordered = names(Rmot))

library("semPlot")
semPaths(fitMot, what = "est", edge.label.cex = 0.7, edge.color = 1, esize = 1, sizeMan = 4.5, asize = 2.5,
          intercepts = FALSE, rotation = 4, thresholdColor = "red", mar = c(1, 5, 1.5, 5), fade = FALSE, nCharNodes = 4)

inspect(fitMot, what = "est")$theta
inspect(fitMot, what = "est")$lambda
inspect(fitMot, what = "std")$lambda
inspect(fitMot, what = "est")$psi
inspect(fitMot, what = "std")$psi

parameterEstimates(fitMot, standardized = TRUE)
summary(fitMot, standardized = TRUE, fit.measures = TRUE)
parameterEstimates(fitMot)[5,]

mot_model2 <- '
  extrinsic =~ ext1 + ext2 + ext3 + ext4 + ext6 + ext7 +
              ext8 + ext9 + ext10 + ext11 + ext12
  intrinsic =~ int1 + int2 + int3 + int4 + int5'
fitMot2 <- lavaan::cfa(mot_model2, data = Rmot, ordered = names(Rmot)[-5])
vind <- c(1:4, 13:16, 32:35)
Rmot2 <- na.omit(Rmotivation[, vind])

mot_model3 <- '
  extrinsic =~ ext1 + ext2 + ext3 + ext4
  hybrid =~ hyb1 + hyb2 + hyb3 + hyb4
  intrinsic =~ int1 + int2 + int3 + int4
  motivation =~ extrinsic + hybrid + intrinsic'
fitMot3 <- lavaan::cfa(mot_model3, data = Rmot2, ordered = names(Rmot2))

semPaths(fitMot3, what = "std", edge.label.cex = 0.7, edge.color = 1, esize = 1, sizeMan = 5, asize = 2.5,
          intercepts = FALSE, rotation = 4, thresholdColor = "red", mar = c(1, 5, 1.5, 5), fade = FALSE, nCharNodes = 4)

summary(fitMot3, standardized = TRUE, fit.measures = TRUE)

vind <- c(1:4, 13:16, 32:35, 39:41)
Rmot3 <- na.omit(Rmotivation[, vind])
mot_model4 <- '
  extrinsic =~ ext1 + ext2 + ext3 + ext4
  hybrid =~ hyb1 + hyb2 + hyb3 + hyb4
  intrinsic =~ int1 + int2 + int3 + int4
  motivation =~ extrinsic + hybrid + intrinsic
  motivation ~ npkgs + phd'
fitMot4 <- lavaan::cfa(mot_model4, data = Rmot3, ordered = names(Rmot3[1:12]))

semPaths(fitMot4, what = "std", edge.label.cex = 0.7, edge.color = 1, esize = 1, sizeMan = 5, asize = 2.5,
          intercepts = FALSE, rotation = 4, thresholdColor = "red", mar = c(1, 5, 1.5, 5), fade = FALSE, nCharNodes = 4)

```

```

parameterEstimates(fitMot4)[16:17,]

library("semTools")
data("Bergh")
GP_model <- 'GP =~ EP + HP + DP + SP'
minvfit1 <- measEq.syntax(GP_model, data = Bergh, group = "gender", return.fit = TRUE)
minvfit2 <- measEq.syntax(GP_model, data = Bergh, group = "gender",
  group.equal = c("loadings"), return.fit = TRUE)
minvfit3 <- measEq.syntax(GP_model, data = Bergh, group = "gender",
  group.equal = c("loadings", "intercepts"), return.fit = TRUE)
minvfit4 <- measEq.syntax(GP_model, data = Bergh, group = "gender",
  group.equal = c("loadings", "intercepts", "means"), return.fit = TRUE)
anova(minvfit1, minvfit2, minvfit3, minvfit4)

GP_model <- 'GP =~ c(v1,v1)*EP + c(v2,v2)*HP + c(v3,v3)*DP + SP'
fitBase <- lavaan::cfa(GP_model, data = Bergh, group = "gender", estimator = "MLR")

GP_model <- 'GP =~ EP + HP + DP + SP'
fitBase <- lavaan::cfa(GP_model, data = Bergh, group = "gender", group.equal = c("loadings"),
  group.partial = c("GP=~ SP"), estimator = "MLR")

fitBase1 <- lavaan::cfa(GP_model, data = Bergh, group = "gender", group.equal = c("loadings", "intercepts"),
  group.partial = c("GP=~SP", "DP~1", "HP~1", "SP~1"), estimator = "MLR")

GP_model2 <- 'GP =~ c(v1,v1)*EP + c(v2,v2)*HP + c(v3,v3)*DP + c(NA, 0)*SP'
fitI0 <- lavaan::cfa(GP_model2, data = Bergh, group = "gender", group.equal = c("intercepts"),
  group.partial = c("DP~1", "HP~1", "SP~1"), estimator = "MLR")

fitMarg <- lavaan::cfa(GP_model, data = Bergh, group = "gender", group.equal = c("loadings", "intercepts"),
  group.partial = c("DP~1", "HP~1", "SP~1"), estimator = "MLR")

anova(fitMarg, fitBase1)

library("MPSychoR")
library("lavaan")
data("SD0wave")
model_sdo1 <- '
  SD01996 =~ 1*I1.1996 + a2*I2.1996 + a3*I3.1996 + a4*I4.1996
  SD01998 =~ 1*I1.1998 + a2*I2.1998 + a3*I3.1998 + a4*I4.1998
  SD01996 ~~ SD01998

  ## intercepts
  I1.1996 ~ int1*1; I1.1998 ~ int1*1
  I2.1996 ~ int2*1; I2.1998 ~ int2*1
  I3.1996 ~ int3*1; I3.1998 ~ int3*1
  I4.1996 ~ int4*1; I4.1998 ~ int4*1

  ## residual covariances
  I1.1996 ~~ I1.1998
  I2.1996 ~~ I2.1998
  I3.1996 ~~ I3.1998
  I4.1996 ~~ I4.1998

  ## latent means: 1996 as baseline
  SD01996 ~ 0*1
  SD01998 ~ 1'
fitsdo1 <- cfa(model_sdo1, data = SD0wave, estimator = "MLR")

```

```

parameterEstimates(fitsdo1)[22:23,]

model_sdo2 <- '
  ## 1st CFA level, constant loadings across time
  SDOD1996 =~ 1*I1.1996 + d1*I2.1996
  SDOD1998 =~ 1*I1.1998 + d1*I2.1998
  SDOD1999 =~ 1*I1.1999 + d1*I2.1999
  SDOE1996 =~ 1*I3.1996 + a1*I4.1996
  SDOE1998 =~ 1*I3.1998 + a1*I4.1998
  SDOE1999 =~ 1*I3.1999 + a1*I4.1999

  ## 2nd CFA level, constant loadings across time
  SD01996 =~ 1*SDOD1996 + sd1*SDOE1996
  SD01998 =~ 1*SDOD1998 + sd1*SDOE1998
  SD01999 =~ 1*SDOD1999 + sd1*SDOE1999

  ## Constant 1st level intercepts
  I1.1996 ~ iI1*1; I1.1998 ~ iI1*1; I1.1999 ~ iI1*1
  I2.1996 ~ iI2*1; I2.1998 ~ iI2*1; I2.1999 ~ iI2*1
  I3.1996 ~ iI3*1; I3.1998 ~ iI3*1; I3.1999 ~ iI3*1
  I4.1996 ~ iI4*1; I4.1998 ~ iI4*1; I4.1999 ~ iI4*1

  ## residual covariances:
  I1.1999 ~~ I1.1998; I1.1996 ~~ I1.1998; I1.1999 ~~ I1.1996
  I2.1999 ~~ I2.1998; I2.1996 ~~ I2.1998; I2.1999 ~~ I2.1996
  I3.1999 ~~ I3.1998; I3.1996 ~~ I3.1998; I3.1999 ~~ I3.1996
  I4.1999 ~~ I4.1998; I4.1996 ~~ I4.1998; I4.1999 ~~ I4.1996

  ## latent means
  SD01996 ~ 0*1      ## 1996 baseline year
  SD01998 ~ 1        ## 1998 vs. 1996
  SD01999 ~ 1        ## 1999 vs. 1996
'

fitsdo2 <- cfa(model_sdo2, data = SD0wave, estimator = "MLR")

semPaths(fitsdo2, what = "est", edge.label.cex = 0.7, edge.color = 1, esize = 1, sizeMan = 6, asize = 2.5,
  intercepts = FALSE, rotation = 4, thresholdColor = "red", mar = c(1, 5, 1.5, 5), fade = FALSE)
parameterEstimates(fitsdo2)[43:45,]

data("FamilyIQ")
modelIQ <- '
  level: 1
    numeric =~ wordlist + cards + matrices
    perception =~ figures + animals + occupation
  level: 2
    general =~ wordlist + cards + matrices + figures + animals +
      occupation'
fitIQ <- cfa(modelIQ, data = FamilyIQ, cluster = "family", std.lv = TRUE)
fitIQ

## ----- bayesian confirmatory factor analysis
library("blavaan")
dpriors()[c("lambda", "itheta", "ipsi")]

library("MPSychoR")
data("Bergh")
GP_model <- 'GP =~ EP + HP + DP + SP'

```

```
set.seed(123)
fitBCFA <- bcfa(GP_model, data = Bergh, burnin = 2000, sample = 10000, n.chains = 2)

plot(fitBCFA, pars = 1:2, plot.type = "trace")
plot(fitBCFA, pars = 1:2, plot.type = "autocorr")
summary(fitBCFA)
```

Chapter 3

Path Analysis and Structural Equation Modeling

We describe our methods in this chapter.

Chapter 4

Item Response Theory

Some *significant* applications are demonstrated in this chapter.

4.1 Example one

4.2 Example two

Chapter 5

Principal Components Analysis

We have finished a nice book.

Chapter 6

Correspondence Analysis

Chapter 7

Gif Methods

Chapter 8

Multidimensional Scaling

Chapter 9

Graphing Multidimensional Data

Chapter 10

Networks

Chapter 11

Modeling Trajectories and Time Series

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