

# Silicon fertilizer application promotes phytolith accumulation in rice plants

Xing Sun<sup>1, 2</sup>, Qin Liu<sup>1\*</sup>, Tongtong Tang<sup>1</sup>, Xiang Chen<sup>1</sup>, Xia Luo<sup>2</sup>

<sup>1</sup>Institute of Soil Science (CAS), China, <sup>2</sup>School of Biological Science and Food Engineering, Chuzhou University, China

Submitted to Journal: Frontiers in Plant Science

Specialty Section: Functional Plant Ecology

ISSN: 1664-462X

Article type:

Original Research Article

Received on: 12 Dec 2018

Accepted on: 21 Mar 2019

Provisional PDF published on:

21 Mar 2019

Frontiers website link: www.frontiersin.org

#### Citation

Sun X, Liu Q, Tang T, Chen X and Luo X(2019) Silicon fertilizer application promotes phytolith accumulation in rice plants. *Front. Plant Sci.* 10:425. doi:10.3389/fpls.2019.00425

#### Copyright statement:

© 2019 Sun, Liu, Tang, Chen and Luo. This is an open-access article distributed under the terms of the <u>Creative Commons Attribution License (CC BY)</u>. The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

# Silicon fertilizer application promotes phytolith accumulation in rice

2	plants
3	Xing Sun <sup>1,2</sup> , Qin Liu <sup>1</sup> *, and Tongtong Tang <sup>1</sup> , Xiang Chen <sup>1</sup> , Xia Luo <sup>2</sup>
4	1, Institute of Soil Science, Chinese Academy of Sciences, Nanjing,PR China,210008
5	2, School of Biological Science and Food Engineering, Chu Zhou University, Chuzhou, 239000,
6	China
7	
8	*Correspondence:
9	Qin Liu
10	qliu@issas.ac.cn
11	



#### Abstract

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

31

32

In this study, a pot experiment was designed to elucidate the effect of varying dosages of silicon(Si) fertilizer application in Si-deficient and enriched paddy soils on rice phytolith and the carbon (i.e., C) bio-sequestration within phytoliths (PhytOC). The maximum silicon fertilizer dosage treatment(XG3) in the Si-deficit paddy soil resulted in an increase in the rice phytolith content by 100.77% in the stem, 29.46% in the sheath and 36.84% in the leaf compared with a lack of silicon fertilizer treatment(CK). However, the maximum silicon fertilizer dosage treatment(WG3) in the silicon-enriched soil increased the rice phytolith content by only 32.83% in the stem, 27.01% in the sheath and 32.06% in the leaf. Overall, Si fertilizer application significantly (p<0.05) increased the content of the rice phytoliths in the stem, leaf and sheath in both the Si-deficient and enriched paddy soils, and the statistical results showed a positive correlation between the amount of Si fertilizer applied and the rice phytolith content with correlation coefficients of 0.998 (p<0.01) in the Si-deficient soil and 0.952 (p<0.05) in the Si-enriched soil. In addition, the existence of phytoliths in the stem, leaf and sheath of rice and its content in the Si-enriched soil were markedly higher than that in the Si-deficient soil. Therefore, silicon fertilizer application helped to improve the phytolith content of the rice plant.

Key Words: Si fertilizer, phytolith accumulation, Si-deficient paddy soils, PhytOC, Rice organs

# Introduction

Phytoliths derive from bio-mineralization in plants and usually take the shape of the plant cell or cell spatium where Si is deposited. Phytoliths content of plants ranges from less than 50 g kg<sup>-1</sup> to as high as 150 g kg<sup>-1</sup> (Epstein, 1994; Ji et al, 2017; Parr et al, 2010; Song et al, 2013; Song et al,

2017), mainly due to phylogenetic differences in Si requirements of most dicotyledons and some Gramineae (Hodson et al, 2005), as well as the amount of available silica in the environment (Guo et al, 2015; Seyfferth et al, 2013; Si et al, 2018; Wen et al, 2018).

Rice is a staple crop, with a global planting area of approximately 1.64× 10<sup>8</sup> ha as of 2014(Prajapati et al, 2016). When rice is harvested, the rice straw and husks are removed from the paddy field and used for other purposes, including animal feeding and firewood, or simply incinerated (Savant et al, 1996). Thus, most of the Si taken up by rice is removed from a field when the rice straw is removed, and the loss of SiO<sub>2</sub> is from 75 kg hm<sup>-2</sup> to 130 kg hm<sup>-2</sup> every production season(Zhang et al, 2014). Such large losses of Si make it difficult to maintain the balance of Si in soils from natural weathering alone. Currently, most paddy soils in China are Si-deficient. For example, 73% of paddy soils in Zhejiang Province, and approximately 60% in Henan Province are Si-deficient.(Cai, 2015). Some research has shown that Si fertilizer application can significantly increase the biomass of rice(Wu et al, 2014; Zhang et al, 2014).

In plants, monosilicic acid is taken up from the soil by a specific transporter(Ma et al, 2006; Song et al, 2014a) and deposited throughout the cellular structures, thereby forming amorphous Si particles known as 'phytoliths'(Pearsall, 1989; Piperno, 1988). There is a significant correlation between the Si content and the phytolith content of crop materials, including the leaves, stems and sheaths, the Si concentration of the plant phytoliths is approximately 90%. (Song et al, 2014a)

Phytoliths can occlude small amounts of many elements, such as C, N, S, and so on (Anala and Nambisan, 2015; Kameník et al, 2013; Li et al, 2014). The C-occluded content of phytoliths ranges from less than 1 g kg<sup>-1</sup> to as high as 100 g kg<sup>-1</sup>. (Clarke, 2003). This PhytOC can be stored in

the soil for thousands of years(Parr and Sullivan, 2005). Thus, it plays a vital role in global carbon pools(Song et al, 2014a). The Si cycle is tightly coupled to the C cycle, and this interaction is relevant for research on climate change(Chadwick et al, 1994). The formation of phytoliths in rice plants depends not only on the crops(Guo et al, 2015; Li et al, 2013a), but also on the plant cultivars(Henriet et al, 2008; Hodson et al, 2005; Yang et al, 2015) and soil Si availability(Henriet et al, 2008; Klotzbücher et al, 2018), and so on.

The application of silicon fertilizer on soils with different available silicon content needs further study on the accumulation of Phytolith in rice. Thus, in this work, we designed a pot experiment to elucidate the effect of varying dosages of Si fertilizer application on the rice phytolith and PhytOC contents of plants grown in Si-deficient and enriched paddy soils.

# 1.Methods

# 1.1 Experimental Soils and Rice cultivar

The Si-deficient paddy soil (red paddy soil) was obtained from Yangliu Town, Xuanchen City, Anhui Province, P.R. China. The Si-enriched paddy soil (Wushan soils) was obtained from the Changshu Agroecological Experimental Station, Chinese Academy of Sciences. The base is located in Xinzhuang County, South Changshu, Suzhou, Jiangsu Province, P.R. China. The physicochemical properties of two soils were showed in Table 1.

The rice cultivar (*Oryza* sativa) Nanjing 46 was achieved from the Changshu Agroecological Experimental Station, Chinese Academy of Sciences.

# 1.2 Pot Experiment

Two soils (Si-deficient and enriched paddy soils) were selected from Xuanchen City and the Changshu Agroecological Experimental Station, Chinese Academy of Sciences, respectively. Four

available Si dosages were designed in the pot experiments: 1. CK (Si fertilizer not applied);2. low slag Si fertilizer I (SiO<sub>2</sub>150 kg ha<sup>-1</sup>);3. high slag silicon fertilizer II (300 kg ha<sup>-1</sup>), and 4. high slag silicon fertilizer III (600 kg ha<sup>-1</sup>). Thus, this experiment comprised 8 treatments repeated 3 times. Two soils were placed in the pot bowl for a total volume of 0. 0175 m<sup>3</sup>; each pot contained N 46%, P<sub>2</sub>O<sub>5</sub>13.5% and K<sub>2</sub>O 60%, Si fertilizer as base fertilizer was applied, 3 rice plants was planted in every pot. Pots were placed in greenhouse of the Changshu Agroecological Experimental Station, Chinese Academy of Sciences in June 2014, and the whole rice growth period was maintained using conventional management.

# 1.3 Sample preparation

After the rice cultivar harvest, each rice plant was separated into five different organs: sheath, leaf, root, stem and grain. All rice samples were rinsed twice in distilled water, placed in an ultrasonic bath for 20 min and subsequently dried in oven at 70°C for 24 h. After hulling, the rice organs samples were stored for phytolith extraction and PhytOC determination.

#### 1.4 Phytolith extraction from rice organs and PhytOC analysis

The phytolith extraction was used for a revised wet digestion measurement previously described by Sun et al(Sun et al, 2016; Zuo and Lü, 2011). Phytolith extraction samples assemblages were installed on glass slides in Balsam Canada mounting medium. The slides were viewed at 400 × magnification using a microscope (Jiangnan XP-213, China) fitted with a polarizing filter and a 5.0 MP color CCD camera to ensure that no organic material residue as showed by Parr et al. (Parr et al., 2010) (Fig.1) The PhytOC was measured using an Elemental Analyzer 3000(GmbH Company, Germany).

#### 1.5 Statistical Analyses

The mean values of all parameters were calculated from the determination of three replicates, and the standard errors of the means were determined. A one-way ANOVA was used to measure the significance of the results between different varieties, and Tukey's multiple range tests (p<0.05) were subsequently performed. All of the statistical analyses were performed using SPSS v.17 for Windows.

#### 2 Results

# 2.1 Phytolith and C contents of the phytoliths in the rice organs

With the increase in the application of the Si fertilizer dosages, the content of the phytoliths in the rice organs could be increased in the Si-deficient red paddy soil (Table 2). For example, the content of the phytoliths in the XG3(26.10 g kg<sup>-1</sup>) and XG2(18.50 g kg<sup>-1</sup>) stems was significantly (p<0.05) higher than that of the control(13.00 g kg<sup>-1</sup>), and the rate increased by100.7% and 42.3%, respectively. In addition, the content of the phytoliths of XG1 in the stem was not significantly (p>0.05) different than that of the control. However, the content of the phytoliths in the rice sheath and leaf could be significantly (p<0.05) increased by the application of all the Si fertilizer dosages. The content of the phytoliths in the XG3 treatment rice grains could only be increased by a high dose of Si fertilizer application. However, the content of the phytoliths in all the root treatments was not significantly (p>0.05) different from that of the control.

With the increase in the application of the Si fertilizer doses, the content of the phytolith in the rice organs could be increased in the Si-enriched Wushan paddy soil (Table 2). For example, the content of the phytoliths of the WG3 (100.60 g kg<sup>-1</sup>) and WG2 (93.13 g kg<sup>-1</sup>) leaves was significantly (p<0.05) higher than that of the control(76.18 g kg<sup>-1</sup>), and the increase in the rate

was 32.06% and 22.25%, respectively. In addition, the content of the phytoliths of WG1 in the stem was not significantly (p>0.05) different from that of the control. However, the content of the phytoliths in the other rice organs could be significantly (p<0.05) increased by the application of high Si fertilizer dosages.

Thus, different Si fertilizer doses might increase the content of the phytoliths in the rice organs in either Si-deficient red paddy soil or Si-enriched Wushan paddy soil. The C content in the phytoliths in the organs was not affected by the increase in the Si fertilizer dose. However, the content of the C in the phytoliths was different in all the organs. Generally the content of the C of the leaf phytoliths was higher than that of the other organs(Table 3).

# 2.2 Phytolithcontent and the estimated PhytOC fluxes in whole rice plants

Compared with the control treatment, the content of phytoliths in the whole rice plant was significantly (p<0.05) increased by the use of a high Si fertilizer dose in the two types of soils(Table 4). The C content of the phytoliths and the PhytoC content of the dry organ weights were not significantly (p>0.05) different in the rice plant. In Si-deficient red paddy soil, the estimated PhytoC fluxes were calculated by the content and proportion of the phytoliths and the C content of the phytoliths in each part of the rice plant. The results showed that the application of Si fertilizer could significantly (p<0.05) increase the content of the estimated PhytoC fluxes in the whole plant with the increase in the Si fertilizer dosage. The estimated PhytoC fluxes of the XG2 (11.36 kg-CO<sub>2</sub> ha<sup>-1</sup>yr<sup>-1</sup>) and XG3(12.93 kg-CO<sub>2</sub> ha<sup>-1</sup>yr<sup>-1</sup>) treatments were 43.04% and 49.70%, respectively, and were significantly (p<0.05) higher than those of the control treatment(8.41 kg-CO<sub>2</sub> ha<sup>-1</sup>yr<sup>-1</sup>). In the Si-enriched soil, the phytolith content of all the Si fertilizer treatments in the rice plants was higher than that of the control treatment, but it was not significantly (p>0.05)

different in all the Si fertilizer treatments compared with the control treatment. The estimated PhytOC fluxes of WG1 were 1.3% lower than those of the control.

# 2.3The correlation coefficients between the six variables of the red paddy soil

As shown in Table 5, the coefficient of variation in the different factors in the Si-deficient red paddy soils was high, illustrating considerable variation among these different Si fertilizer dosages. The results demonstrated that there was a significant correlation (R=0.998, p<0.01) between the phytolith content and the silicon fertilizer dose. The C contents of the phytoliths were not correlated(R=-0.177, p>0.05) with the phytolith content in the rice plants treated with different fertilizer doses. The correlation coefficient was 0.986, indicating a significant relationship(p<0.05) between the phytolith content and the estimated PhytOC fluxes. The biomass of the rice was significantly related to the phytolith content (R=0.972, p<0.05)and the estimated PhytOC fluxes(R=0.994, p<0.01).

# 2.4 The Correlation coefficients between the six variables of the Wushan soil

As shown in Table 6, the coefficient of variation in the different factors in the Si-deficient red paddy soils was high, illustrating considerable variation among the different Si fertilizer doses. The results demonstrated that there was a significant correlation (R=0.952, p<0.05) between the phytolith content and the silicon fertilizer dose. The C contents of the phytoliths were not correlated(R=-0.035, p>0.05) with the phytolith content in the rice plants of different fertilizer treatments. The correlation coefficient was 0.598 and was significantly correlated (p>0.05) between the phytolith content and the estimated PhytOC fluxes. The biomass of the rice was significantly correlated with the phytolith content (R=-0.890, p>0.05)and the estimated PhytOC fluxes(R=0.076, p>0.05).

#### 3 Discussion

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

182

183

184

185

Rice accumulates Si(Seyfferth et al, 2013), and the Si concentration is approximately 10%-15% in the rice plant (Marschner,1995) with approximately 90% of the Si present in the phytolith (Wang, 1998). There was a significant correlation between the Si content and the phytolith content of the crop materials, such as the phytolith contents of the rice leaves, stems, and sheaths (Song et al, 2014a). The shape of the phytoliths in the different rice organs varied (e.g., double-peaked, bulliform and parallel dumbbell phytoliths)(Prajapati et al, 2016). Prajapati et al. reported that the phytolith content in the different rice organs (stem, sheath, leaf and grain) ranged from 0.14 g kg<sup>-1</sup> to 26.4 g kg<sup>-1</sup>(Prajapati et al, 2016). Similar results and trends were reported by other researchers (Guo et al, 2015; Li et al, 2013c). Our results showed that whether the paddy soil was Si-deficient or Si-enriched, the utilization of Si fertilizer could significantly (p<0.05) improve the phytolith content of the rice organs(Tables 2,3), such as the stem, sheath, leaf, grain and root. According to formation mechanism of phytolith, the soil available Si is taken up by rice plant the roots, usually take the shape of the plant cell or cell spatium where Si is deposited(Ma, 2003; Neumann, 2003; Piperno, 1988; Song et al, 2016). Thus, the use of Si fertilizer increased the content of effective Si in the soil(Cai, 2015; Liu et al, 2006; Ma et al, 2004) and increased the absorption capacity of Si in the rice(Guo et al, 2015; Huan et al, 2018; Li et al, 2013c; Seyfferth et al, 2013; Zuo et al, 2016), therefore increasing the phytolith content of the rice plant (Table 4). A substantial amount of research reported that the factors of the PhytOC content were as follows: different varieties(Li et al, 2013b; Parr et al, 2010; Parr et al, 2009; Parr and Sullivan, 2011; Song et al, 2017; Sun et al, 2017), pest and disease resistances(Ma et al, 2002), nitrogen utilization (Zhao et al, 2016), basalt powder(Guo et al, 2015), soil-effective Si content(Klotzbücher

et al, 2018; Song et al, 2014b) and net production on the ground(Blecker et al, 2006). It has been shown that Si is an important element for rice growth and the deficiency of plant-available Si may exert an adverse effect on the rice yield through biotic stresses, disease and pests, and so on(Ma et al, 2004; Ma, 2004). Our results also showed that the content of phytolith in rice plants were different in Si-deficient and Si-enriched paddy soil. The content of Phytolith in rice plants with Si-enriched paddy soils was higher than that in rice plants with Si-deficient paddy soil (Table 2, 4). Moreover, whether in silicon-deficient soil or in silicon-deficient soil, The phytolith content of rice plants was positive correlation(p<0.05) with the Si fertilizer dose in two types paddy soil((Table 5, 6). Previous studied have demonstrated that the content of the Si (phytoliths) in crops may be promoted through the Si fertilizers application (Alvarez and Datnoff, 2001; Liang et al, 2010; Mecfel et al, 2010). Further, in the Si-deficient paddy soil, the estimated PhytOC fluxes was significantly related to the Si fertilizers (R=0.973,p<0.05), and the phytolith content (R=0.986, p<0.05) and the biomass of the rice (R=0.994, p<0.01) (Table 5). However, in the Si-enriched paddy soil, the estimated PhytOC fluxes was not correlation(P>0.05) with these factors. Zhang et al showed (Zhang et al, 2014)that the yield of rice was increased 14.5% by the use of 225 kg ha<sup>-1</sup> Si fertilizer; when the application of Si fertilizer was increased to 375 kg ha<sup>-1</sup>, the yield of the rice was only increased by 10.1%. Similarly, Wu et al also recommended the use of 225 kg ha<sup>-1</sup> Si fertilizer as the most economical measure(Wu et al, 2014). We also obtained the same results. The application of Si fertilizer to the Si-enriched paddy soil did not increase the biomass of the rice but reduced it. In particular, when the amount of the Si fertilizer reached 600 kg ha<sup>-1</sup>, the rice biomass decreased significantly by 29.10% compared with the control treatment (Table 4). Therefore, excessive Si fertilizer not only has no benefit to the accumulation of estimated PhytOC

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

205

206

fluxes in rice plant, but also reduces the yield of rice. However, for Si-deficient soils, the application of Si fertilizer can not only increase rice yield, but also increase phytoliths content of rice plants and the estimated PhytOC fluxes(Table 4). Thus, different Si fertilizer doses were one of measures to improve the phytolith content and the biomass of the rice plant. Thus, how to promote the phytolith content and C content of phytoliths will require further in-depth study.

208

209

210

211

212

213

214

215

216

217

218

219

220

221

222

223

224

225

226

227

228

229

The global rice cultivation area was approximately 1.64×10<sup>8</sup> ha in 2014(Prajapati et al, 2016); When rice is harvested, the rice straw and husks are removed from the paddy field and used for other purposes, including animal feeding and firewood, or simply incinerated (Savant et al, 1996). Thus, most of the Si taken up by rice is removed from a field when the rice straw is removed, and the loss of SiO<sub>2</sub> is from 75 kg ha<sup>-1</sup> to 130 kg ha<sup>-1</sup> every production season(Zhang et al, 2014). Such large losses of Si make it difficult to maintain the balance of Si in soils from natural weathering alone. Appropriate dosages of Si fertilizer could solve the problem of Si deficiency in soil, and increase the biomass of rice and the content of phytolith in rice plants, and indeed result in the occlusion of increased CO<sub>2</sub> in the rice plants (Liang et al, 2010; Mecfel et al, 2010). The estimated PhytOC fluxes increased from 0.49 Kg-e-CO<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup>to 4.52 Kg-e-CO<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup> (Table 4). More than  $8.04\times10^4$  Mg-e-CO<sub>2</sub> to  $7.41\times10^5$  Mg-e-CO<sub>2</sub> would have been occluded within the phytolith of the rice plants per year globally. Considering that the highest estimated PhytOC flux of the rice plants, 2.12×10<sup>6</sup> Mg-e-CO<sub>2</sub>, would have been occluded within the phytolith of rice plants every year. Although the annual CO<sub>2</sub> bio-sequestration within the rice phytoliths of the unit area is likely to be lower than that of other plants, such as bamboo leaf litter(1.56×10<sup>7</sup> Mg-e-CO<sub>2</sub>yr<sup>-1</sup>)(Parr, 2006), plants  $(4.39\times10^7 \text{ Mg-e-CO}_2\text{yr}^{-1})$  (Guo et al, 2015), grasslands  $(4.14\times10^7 \text{ mg})$  $Mg-e-CO_2yr^{-1})$  (Song et al, 2012), millet (2.37×10<sup>6</sup>  $Mg-e-CO_2yr^{-1})$  (Pan et al, 2017) and sugarcane

leaf(0.72×10<sup>7</sup> Mg-e-CO<sub>2</sub>yr<sup>-1</sup>) (Parr et al, 2009).In this study, we showed that Si fertilizer application could promote the phytolith content and biomass of rice plants and further improve the estimated PhytOC flux of rice plants. Thus, the measure provided a theoretical basis for the bio-carbon sequestration of the rice plant and laid a foundation for PhytOC fixation in paddy soil by the return of straw.

#### Conclusion

The use of silicon fertilizer could significantly increase the phytolith content of rice plants in Si-deficient red paddy soil or Si-enriched Wushan soil. The phytolith content of rice plants was positive correlation with the silicon fertilizer dose in two types paddy soil. The estimated PhytoC fluxes in Si-deficient red paddy soil was a positive correlation with the phytolith content, the biomass of the rice and the silicon fertilizer dose, respectively. In this study, we estimated that the PhytoC fluxes increased from0.49 Kg-e-CO<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup> to 4.52Kg-e-CO<sub>2</sub> ha<sup>-1</sup> yr<sup>-1</sup>. More than  $8.04 \times 10^4$  Mg-e-CO<sub>2</sub> to  $7.41 \times 10^5$  Mg-e-CO<sub>2</sub> would have been occluded within the phytoliths of the rice plants per year globally. Therefore, Si fertilizer application might provide a new approach to increase the atmospheric CO<sub>2</sub> occluded within the phytoliths, offering a potential method.

# **Acknowledgements**

This work was partially supported by the National Natural Science Foundation of China (No.41271208; No.31400464;), the Anhui Province University Natural Science Research Foundation (No.KJ2017A423; No.KJ2018A0430),and the Excellent Researcher Program of the Education Department of Anhui Province(No.gxyq2018097), and Laboratory Opening Subject of the School of Biology and Food Engineering of Chuzhou University (No.SWSP201816KF) ..

#### References

- Alvarez, J. and Datnoff, L. E. (2001). The economic potential of silicon for integrated management and sustainable rice production *Crop Prod* 20, 43-48.

  Anala, R. and Nambisan, P. (2015). Study of morphology and chemical composition of phytoliths on the surface of paddy straw. *Paddy & Water Environment* 13, 521-527.
- Blecker, S. W., Mcculley, R. L., Chadwick, O. A. and Kelly, E. F. (2006). Biologic cycling of silica across a grassland bioclimosequence. *Global Biogeochemical Cycles* 20, 4253-4274.
- Cai, D. (2015). Silicon fertilizer: a new resource-saving and environment-friendly fertilizer. *China* Agri-Production News, 23-23.
- Chadwick, K. M., Johnson, R. G., Dhaul, A. K., Steinmeyer, R. D. and Halm, R. L. (1994). Conversion of
   direct process high-boiling component to chlorosilane monomers in the presence of chlorine.
   Clarke, J. (2003). The occurrence and significance of biogenic opal in the regolith. *Earth Science*
- 263 Reviews 60, 175-194.
- Epstein, E. (1994). The anomaly of silicon in plant biology. *Proceedings of the National Academy of*Sciences of the United States of America 91, 11-17.
- Guo, F., Song, Z., Sullivan, L., Wang, H., Liu, X., Wang, X., Li, Z. and Zhao, Y. (2015). phytolith carbon
   sequestration in rice ecosystems through basalt powder amendment. *Science Bulletin* 60,
   591-597.
- Henriet, C., Bodarwé, L., Dorel, M., Draye, X. and Delvaux, B. (2008). Leaf silicon content in banana

  ( Musa spp.) reveals the weathering stage of volcanic ash soils in Guadeloupe. *Plant and Soil* 313,

  71-82.
- Hodson, M., White, P., Mead, A. and Broadley, M. (2005). Phylogenetic variation in the silicon composition of plants. *Annals of Botany* 96, 1027-1046.

- Huan, X., Lu, H., Zhang, J. and Wang, C. (2018). Phytolith assemblage analysis for the identification of
- 275 rice paddy. Scientific Reports 8
- 276 Ji, Z., Yang, X., Song, Z., Liu, H., Liu, X., Qiu, S., Li, J., Guo, F., Wu, Y. and Zhang, X. (2017). Silicon
- 277 distribution in meadow steppe and typical steppe of northern China and its implications for
- phytolith carbon sequestration. *Grass & Forage Science*, DOI: 10.1111/gfs.12316.
- 279 Kameník, J., Mizera, J. and Řanda, Z. (2013). Chemical composition of plant silica phytoliths.
- 280 Environmental Chemistry Letters 11, 189-195.
- 281 Klotzbücher, T., Klotzbücher, A., Kaiser, K., Merbach, I. and Mikutta, R. (2018). Impact of agricultural
- practices on plant-available silicon. *Geoderma* 331, 15-17.
- Li, B., Song, Z., Li, Z., Wang, H., Gui, R. and Song, R. (2014). Phylogenetic variation of phytolith carbon
- sequestration in bamboos. Sci Rep 4, 4710.
- 285 Li, Z., Song, Z. and Li, B. (2013a). The production and accumulation of phytolith-occluded carbon in
- Baiyangdian reed wetland of China. *Applied Geochemistry* 37, 117-124.
- 287 Li, Z., Song, Z., Li.Beilei and Cai, Y. (2013b). Phytolith production in wetland plants of the Hangzhou Xixi
- Wetlands ecosystem. *Journal of Zhejiang A & F University* 30, 470-476.
- 289 Li, Z., Song, Z., Parr, J. F. and Wang, H. (2013c). Occluded C in rice phytoliths: implications to
- biogeochemical carbon sequestration. *Plant and Soil* 370, 615-623.
- Liang, Y., Haixia, H., Yong-Guan, Z., Jie, Z., Chunmei, C. and Volker, R. M. (2010). Importance of plant
- species and external silicon concentration to active silicon uptake and transport. New Phytologist
- 293 172, 63-72.
- Liu, M. d., Zhang, Y. I., Meng, X. f., Li, J., Yang, D., Wang, Y.-j. and Yu, N. (2006). Eva lua tion of Silicon
- 295 Supply ing Capac ity of Paddy Soil by Silicon Libera tion Dynamics Equa tion

- 296 I The Relationship between Parameter of Soil Silicon Liberation Dynamics Equation and Relative Yield.
- 297 Chinese Journal of Soil Science 37, 107-110.
- 298 Ma, F., Namiki, M., Sakiko, N., Saeko, K., Kazunori, T., Takashi, I. and Masahiro, Y. (2004).
- 299 Characterization of the Silicon Uptake System and Molecular Mapping of the Silicon Transporter
- 300 Gene in Rice. *Plant Physiology* 136, 3284-3289.
- 301 Ma, F. J. (2004). Role of Silicon in Enhancing the Resistance of Plants to Biotic and Abiotic Stresses. Soil
- 302 *Sci Plant Nutr* 50, 11 18.
- 303 Ma, J. F. (2003) Functions of Silicon in Higher Plants: Springer Berlin Heidelberg.
- Ma, J. F., Kazunori, T., Masahiko, I. and Guo Feng, W. (2002). A rice mutant defective in Si uptake. *Plant*
- 305 *Physiology* 130, 2111-2117.
- 306 Ma, J. F., Tamai, K., Yamaji, N., Mitani, N., Konishi, S., Katsuhara, M., Ishiguro, M., Murata, Y. and Yano,
- 307 M. (2006). A silicon transporter in rice. *Nature* 440, 688-691.
- Mecfel, J., Hinke, S., Goedel, W. A., Marx, G., Fehlhaber, R., Bäucker, E. and Wienhaus, O. (2010). Effect
- of silicon fertilizer on silicon accumulation in wheat. Journal of Plant Nutrition and Soil Science =
- Zeitschrift fuer Pflanzenernaehrung und Bodenkunde 170, 769-772.
- 311 Neumann, D. (2003) Silicon in Plants: Springer Berlin Heidelberg.
- 312 Pan, W., Song, Z., Liu, H., Zwieten, L. V., Li, Y., Yang, X., Han, Y., Liu, X., Zhang, X. and Xu, Z. (2017). The
- 313 accumulation of phytolith-occluded carbon in soils of different grasslands. Journal of Soils &
- 314 *Sediments* 17, 1-8.
- 315 Parr, J., Sullivan, L., Chen, B., Gongfu, Y. E. and Zheng, W. (2010). Carbon bio-sequestration within the
- phytoliths of economic bamboo species. *Global Change Biology* 16, 2661-2667.
- 317 Parr, J., Sullivan, L. and Quirk, R. (2009). Sugarcane phytoliths: encapsulation and sequestration of a

- 318 long-lived carbon fraction. Sugar Tech 11, 17-21.
- 319 Parr, J. F. (2006). Effect of fire on phytolith coloration. Geoarchaeology-an International Journal 21,
- 320 171-185.
- 321 Parr, J. F. and Sullivan, L. A. (2005). Soil carbon sequestration in phytoliths. Soil Biology & Biochemistry
- 322 37, 117-124.
- Parr, J. F. and Sullivan, L. A. (2011). Phytolith occluded carbon and silica variability in wheat cultivars.
- 324 Plant and Soil 342, 165-171.
- 325 Pearsall, D. M. (1989) Paleoethnobotany. A Handbook of Procedures. Academic Press., pp. 1-470.
- Piperno, D. R. (1988) *Phytolith Analysis: An Archaeological and Geological Perspective*: San Diego:
- 327 Academic Press.
- Prajapati, K., Rajendiran, V. Coumar, Dotaniya, Ajay, Kundu and Patra (2016). Carbon occlusion
- potential of rice phytoliths: Implications for global carbon cycle and climate change mitigation.
- 330 Applied Ecology and Environmental Research 14, 265-281.
- 331 Savant, N. K., Snyder, G. H. and Datnoff, L. E. (1996). Silicon Management and Sustainable Rice
- Production. *Advances in Agronomy* 58, 151-199.
- 333 Seyfferth, A. L., Kocar, B. D., Lee, J. A. and Fendorf, S. (2013). Seasonal dynamics of dissolved silicon in
- a rice cropping system after straw incorporation. Geochimica Et Cosmochimica Acta 123,
- 335 120-133.
- 336 Si, Y., Wang, L., Zhou, Q. and Huang, X. (2018). Effects of lanthanum and silicon stress on
- 337 bio-sequestration of lanthanum in phytoliths in rice seedlings. Environmental Science & Pollution
- 338 Research 25, 10752-10770.
- 339 Song, Z., Liu, H., Li, B. and Yang, X. (2013). The production of phytolith-occluded carbon in China's

- forests: implications to biogeochemical carbon sequestration. Global Change Biology 370,
- 341 615-623.
- Song, Z., Liu, H., Si, Y. and Yin, Y. (2012). The production of phytoliths in China's grasslands:
- implications to the biogeochemical sequestration of atmospheric CO2. Global Change Biology 18,
- 344 3647-3653.
- Song, Z., Liu, H., Strömberg, C., Yang, X. and Zhang, X. (2017). Phytolith carbon sequestration in global
- terrestrial biomes. *Science of the Total Environment* 603-604, 502-509.
- Song, Z., Müller, K. and Wang, H. (2014a). Biogeochemical silicon cycle and carbon sequestration in
- agricultural ecosystems. *Earth-Science Reviews* 139, 268-278.
- 349 Song, Z., Mcgrouther, K. and Wang, H. (2016). Occurrence, turnover and carbon sequestration
- potential of phytoliths in terrestrial ecosystems. *Earth-Science Reviews* 158, 19-30.
- 351 Song, Z., Wang, H., Strong, P. J. and Guo, F. (2014b). Phytolith carbon sequestration in China's
- 352 croplands. *European Journal of Agronomy* 53, 10-15.
- 353 Sun, X., Liu, Q., Zhao, G., Chen, X., Tang, T. and Xiang, Y. (2017). Comparison of phytolith-occluded
- carbon in 51 main cultivated rice (Oryzasativa) cultivars of China. Rsc Advances 7, 54726-54733.
- 355 Sun, X., Qin, L., Xiang, C. and Keya, Z. (2016). Evaluation of the occluded carbon within husk phytoliths
- of 35 rice cultivars. *Frontiers of Earth Science* 10, 683-690.
- Wang, Y. J. (1998). A study on the chemical compostion of phytolths. J Oceano Huanghai Bohai Seas 16,
- 358 33–38.
- Wen, C., Lu, H., Zuo, X. and Ge, Y. (2018). Advance of research on modern soil phytolith. Science China
- 360 *Earth Sciences* 61, 1-14.
- 361 Wu, H.-b., Shen, J.-n. and Hu, S.-s. (2014). Effect of Silicon on Plant Traits and Yield of Cold Region Rice.

362	North Rice(in chinese) 44, 12-15.
363	Yang, X., Song, Z., Liu, H., Bolan, N. S., Wang, H. and Li, Z. (2015). Plant silicon content in forests of
364	north China and its implications for phytolith carbon sequestration. Ecological Research 30,
365	347-355.
366	Zhang, L., Chen, X., Chang, B., Gu, X., Li, W., wang, S. and Wang, X. (2014). Application effect of silicon
367	fertilizer on rice. Heilongjiang Agricultural Sciences (in Chinese), 43-46.
368	Zhao, Y., Song, Z., Xu, X., Liu, H., Wu, X., Li, Z., Guo, F. and Pan, W. (2016). Nitrogen application
369	increases phytolith carbon sequestration in degraded grasslands of North China. Ecological
370	Research 31, 117-123.
371	Zuo, X. and Lü, H. (2011). Carbon sequestration within millet phytoliths from dry-farming of crops in
372	China. Science Bulletin 56, 3451-3456.
373	Zuo, X., Lu, H., Li, Z., Song, B., Xu, D., Zou, Y., Wang, C., Xiujia, H. and He, K. (2016). Phytolith and
374	diatom evidence for rice exploitation and environmental changes during the early mid-Holocene
375	in the Yangtze Delta. Quaternary Research 86, 304-315.
376	
377	
378	
379	
380	
381	
382	
383	
384	
385	
386	
387	
388	
389	
390	
391	

Figure captions
-----------------

**Figure 1.** Optical microscope images of phytoliths extracted from the rice samples using the wet ashing Method according to Zuo (2011) and Sun(2016), Magnification 400×, scale bar 30 μm.

Table 1 Basic chemical properties of the two soils

Experimental Soils	NH <sub>4</sub> OAc -extractable Si (mg kg <sup>-1</sup> )	рН	total N (g kg <sup>-1</sup> )	total P (g kg <sup>-1</sup> )	total K (g kg <sup>-1</sup> )	organic matter (mg kg <sup>-1</sup> )	Na <sub>2</sub> CO <sub>3</sub> - extractable P (mg kg <sup>-1</sup> )	NH₄OAc- extractable K (mg kg <sup>-1</sup> )
Si-deficient paddy soil (red paddy soil)	5.67	4.62	1.20	0.18	52.49	28.89	17.44	210.0
Si-enriched paddy soil (Wushan soils)	252.3	7.54	2.40	0.73	20.16	39.89	34.27	101.7



Table 2 Effect different of silicon fertilizers on rice organs content of phytoliths,(W: Wushan soil; X:

402		Red paddy soil;G: Leg silicon fertilizer)									
	Treatment s	stem(g kg <sup>-1</sup> )	sheath(g kg <sup>-1</sup> )	leaf(g kg <sup>-1</sup> )	grain(g kg <sup>-1</sup> )	root(g kg <sup>-1</sup> )					
	XCK	13.00±1.44c	36.01±1.45c	34.72±1.50c	11.59±2.41b	80.66±25.81a					
	XG1	16.84±1.07bc	42.73±2.74b	41.37±2.25b	13.86±1.61b	117.20±18.58a					
	XG2	18.50±0.91b	49.37±0.89a	48.72±2.09a	12.96±3.49b	92.87±36.86a					
	XG3	26.10±4.41a	46.62±2.38a	47.51±3.08a	19.37±1.38a	84.78±30.35a					
	WCK	37.22±4.19b	75.31±4.68bc	76.18±4.44b	16.60±3.29b	67.46±22.70b					
	WG1	38.57±4.63b	74.18±3.21c	76.14±8.10b	17.26±1.92b	56.09±1.76b					
	WG2	42.09±3.23b	84.00±5.19b	93.13±11.68a	21.69±1.83ab	96.66±14.10a					
	WG3	49.44±0.74a	95.72±5.59a	100.60±7.98a	24.72±4.96a	82.03±5.32ab					

Different lowercase letters after the data means that the difference between different type of Si-fertilizers dosage treatments is significant (p<0.05)

Table 3 Effect different of silicon fertilizers on rice organs content of C content of phytoliths, (W: Wushan soil; X: Red paddy soil; G: Leg silicon fertilizer)

wushan soil, A. Reu paddy soil, G. Leg silicon Tertilizer)								
Treatment s	stem(g kg <sup>-1</sup> )	sheath(g kg <sup>-1</sup> )	leaf(g kg <sup>-1</sup> )	grain(g kg <sup>-1</sup> )	root(g kg <sup>-1</sup> )			
ХСК	8.60±1.34a	5.80±0.75a	8.02±1.81a	4.78±0.61a	1.67±0.01a			
XG1	4.85±1.20b	6.09±0.92a	9.21±3.84a	6.79±1.87a	1.88±0.31a			
XG2	5.28±1.20b	4.82±1.20a	9.47±1.75a	7.08±1.43a	1.68±0.24a			
XG3	4.64±0.38b	4.61±0.97a	7.23±1.20a	5.10±1.90a	2.19±0.34a			
WCK	2.28±0.36a	2.28±0.39ab	7.41±2.81a	6.53±2.91a	4.98±1.42a			
WG1	2.59±0.55a	2.16±0.11ab	3.22±0.82a	3.39±0.45a	4.79±2.22a			
WG2	4.13±2.0a	2.82±0.54a	5.58±3.37a	4.34±0.92a	4.68±0.59a			
WG3	2.23±0.67a	1.90±0.09b	4.31±2.57a	4.42±1.18a	4.03±1.12a			

Different lowercase letters after the data means that the difference between different type of Si-fertilizers dosage treatments is significant (p<0.05)

Table 4 Effect different of silicon fertilizers on rice plants content of phytoliths, C content of phytoliths, PhytOC content of dry organs weight, and the estimated PhytOC fluxes per ha in kg of CO<sub>2</sub> equivalents (kg~e~ CO<sub>2</sub>) for rice (W: Wushan soil; X: Red paddy soil;G: Leg silicon fertilizer)

Treatment s	Phytolith content(g kg <sup>-1</sup> )	C content of phytoliths(g kg <sup>-1</sup> )	PhytOC content of dry organs weight (g kg <sup>-1</sup> )	Estimated PhytOC fluxes (kg-CO <sub>2</sub> ha <sup>-1</sup> yr <sup>-1</sup> )	Biomass (t ha <sup>-1</sup> )
ХСК	24.23±0.39b	5.74±0.52a	0.13±0.01b	8.41±0.99b	16.38±2.34b
XG1	26.64±1.02a	6.61±1.48a	0.16±0.04a	10.38±2.43a	18.43±1.74a
XG2	28.32±2.06a	6.56±1.04a	0.17±0.03a	11.36±2.34a	18.90±1.67a
XG3	31.94±2.18a	5.14±1.05a	0.16±0.03a	12.93±0.54a	20.33±0.77a
WCK	34.96±0.88b	5.44±1.80a	0.17±0.05a	8.74±0.29a	17.94±0.46a
WG1	36.22±1.06b	3.21±0.15a	0.11±0.01a	7.92±0.95a	17.42±1.61a
WG2	48.06±4,34a	4.29±1.09a	0.21±0.08a	11.11±1.93a	17.78±4.38a
WG3	53.85±0.79a	4.39±0.68a	0.21±0.07a	9.23±0.09a	14.76±1.78b

Different lowercase letters after the data means that the difference between different type of

417 Si-fertilizers dosage treatments is significant (p<0.05)

419	Table 5 The Correlation coefficients between the six variables of the red paddy soil
713	Table 5 The correlation coefficients between the six variables of the rea paday son

Variables	Silicon fertilizer	Phytolith content	C content of phytoliths	PhytOC content of dry organs weight	Estimated PhytOC fluxes	Biomass
Silicon fertilizer	1					_
Phytolith content	0.998**	1				
C content of phytoliths	-0.238	-0.177	1			
PhytOC content of dry organs weight	0.620	0.665	0.612	1		
Estimated PhytOC fluxes	0.973*	0.986*	-0.008	0.795	1	
Biomass	0.953*	0.972*	0.041	0.799	0.994**	1

\* Correlation is significant at the 0.05 level (2-tailed).

\*\* Correlation is significant at the 0.01 level (2-tailed).

422



424	Table 6 The Correlation coefficients between the six variables of the Wushan soil
424	Table 6 The Correlation coefficients between the six variables of the wushan son

Variables	Silicon fertilizer	Phytolith content	C content of phytoliths	PhytOC content of dry organs weight	Estimated PhytOC fluxes	Biomass
Silicon fertilizer	1					
Phytolith content	0.952*	1				
C content of phytoliths	-0.209	-0.035	1			
PhytOC content of dry organs weight	0.599	0.796	0.526	1		
Estimated PhytOC fluxes	0.333	0.598	0.229	0.800	1	
Biomass	-0.890	-0.746	0.100	-0.393	0.076	1

\* Correlation is significant at the 0.05 level (2-tailed).

\*\* Correlation is significant at the 0.01 level (2-tailed).

427







