

Unix II – Scripting, web clients, databases and formats

Biol4230 Thurs, Jan 25, 2017

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Goals of today's lecture:

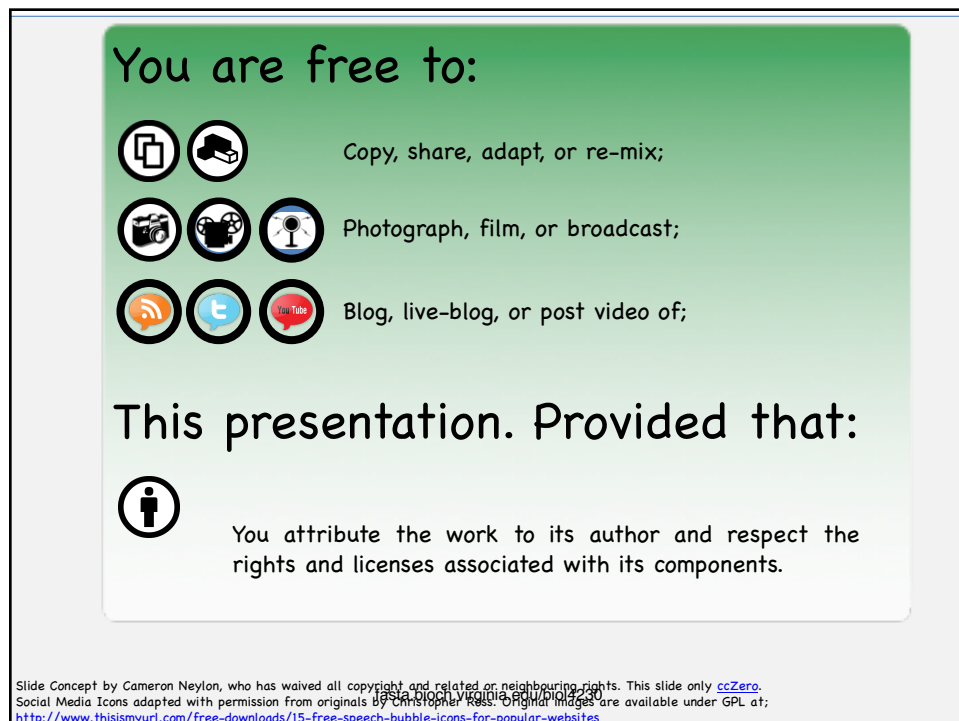
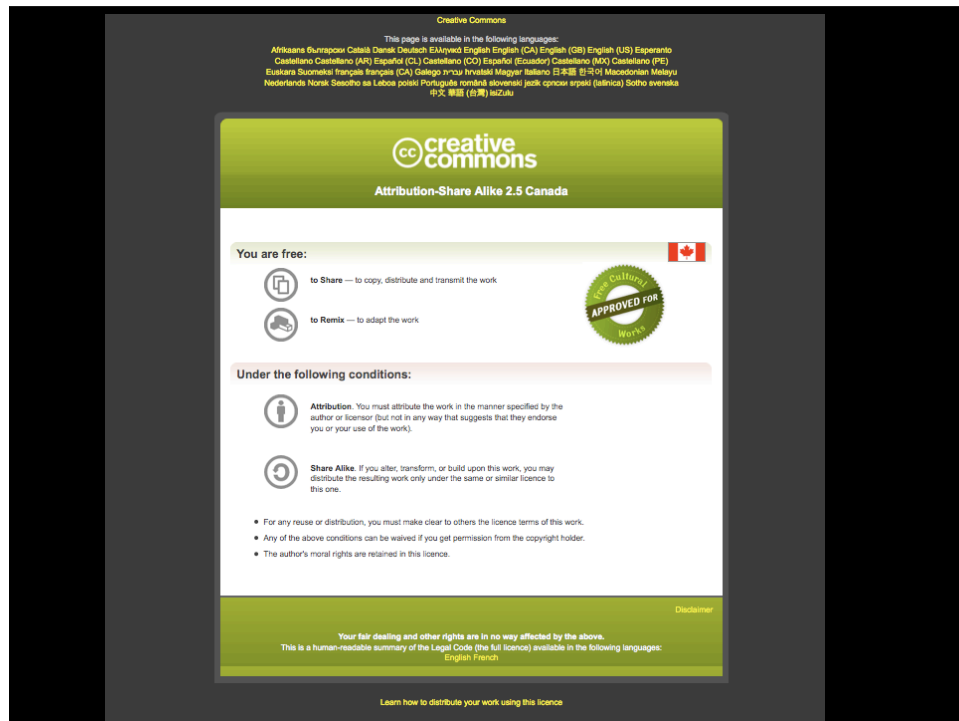
- Creating simple bash scripts
- Survey of Bioinformatics databases (Ouellette)
 - Primary vs reference
 - Annotations and cross-references
 - Survey of file formats
- Scripts as web browsers

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To learn more:

- Scripting the bash shell (Google "bash introduction", focus on variables, flow control)
 - tldp.org/LDP/abs/html/ (concise intro)
 - Learning the Bash Shell, 3rd edition (Ch 4 and 5)
proquest.safaribooksonline.com/book/operating-systems-and-server-administration/unix/0596009658
 - Practical Computing, Ch. 4, 5, 6
 - Practical Computing, App. 3
practicalcomputing.org/files/PCfB_Appendices.pdf
- Bioinformatics databases:
 - Pevsner (2004) "Bioinformatics and Functional Genomics 2nd ed" Wiley-Blackwell, Ch. 1 (on reserve)
- Web clients – `curl`, `wget` (`man curl`, `man wget`)

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Unix II – scripting, web clients, databases

- Scripting – putting commands in a file
 - bash commands:


```
for file in *.fasta ; do ... ; done
```
 - Essential for reproducibility – your electronic lab notebook
 - Automation of repetitive tasks (run blast search using 20 files)
- Web clients – curl/wget – allow scripting of web access
 - Download a list of protein sequences using accessions
 - Homework – (a) do a blast search with tabular output; (b) extract accessions of hits; (c) download those sequences; (d) search with them

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(bash) shell scripts

- files ending with `.sh` suffix
- shebang: `#!/bin/bash` or `#!/bin/sh`
- useful to capture (potentially long) history of UNIX commands into a reproducible analysis
 - you will always need to repeat your analysis
 - you will never remember all the necessary steps
- with some modification, your script can be made generic, and reusable for other data

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shell scripts contain commands

```
franklin: 1 $ echo $PATH    # a simple command
/home/wrp/bin:/usr/local/bin:/bin:/usr/bin:./seqprg/bin

franklin: 2 $ echo_path.sh
                # echo_path.sh contains "echo $PATH"
bash: ./echo_path.sh: Permission denied
                # cannot execute because -rw-r--r--

franklin: 3 $ sh echo_path.sh # can execute with 'sh'
/home/wrp/bin:/usr/local/bin:/bin:/usr/bin:./seqprg/bin

franklin: 4 $ chmod +x echo_path.sh # make executable

franklin: 5 $ echo_path.sh    # now it works
/home/wrp/bin:/usr/local/bin:/bin:/usr/bin:./seqprg/bin
```

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(bash) shell variables

- Your unix session has two kinds of variables, `env` (environment) variables, and `SHELL` variables, refer to them with `$NAME` (`env`) / `$name` (`shell`)
 - Individual variables can be seen with 'echo'


```
echo $PATH
```
 - All environment variables are listed with 'env'
- You can make your own variables for a command as well:


```
files=$(ls *.aa)
echo $files
```

 - shell variables never have a '\$' on the left of the '=', and ALWAYS have a '\$' on the right side.
 - no spaces around the '='


```
new_files=$files
```
- `$SHELL` variables are transient; to make them permanent, use:


```
export PATH=$PATH:/seqprg/bin
```

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(bash) shell flow control

- `for name in [...] ; do [...] ; done`
– do something for each item in a list
- `if [...] ; then [...] ;`
 `elif [...] ; then [...] ;`
 `else [...]`
 `fi`
– specify behavior depending on conditions
- `';'` are only necessary when putting multiple commands on one line.
 `for ... ; do ... ; done`

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Producing new filenames

```
$ for f in *.aa; # file glob (*)
> do
> n=$(basename $f .aa) # $(command) makes output
                        into a string
> nn=${f%.*} # basename() requires a suffix string
> new=$n.new # ${n} if no '.' or '/'
> new2="this${n}that"
> echo $f $new $new2
> done
gstml_human.aa gstml_human.new thisgstml_humanthat
sequence.aa sequence.new thissequencethat
```

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Extracting parts of lines: cut

```
# do a blastp search:
$ blastp -outfmt 6 -query atp6_human.aa -d /slib2/bl_dbs/pir1 > atp6.bl_out

# look at first three lines
$ head -n 3 atp6.bl_out
sp|P00846|ATP6_HUMAN      P00846  100.000 226    0    0    1    226    1    226    2.00e-157    434
sp|P00846|ATP6_HUMAN      P00847   77.876 226    50    0    1    226    1    226    3.58e-124    349
sp|P00846|ATP6_HUMAN      P00848   75.664 226    55    0    1    226    1    226    5.66e-112    318

#ssid          ssid perc alen mism gaps  qstart qend sstart sendevalue  bits

# extract only the ssid column (field)
$ cut -f 2 atp6.bl_out | head -n 3
P00846
P00847
P00848

# change field delimiter with -d " ", -d "|", etc.
```

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COMPUTATIONAL & COMPARATIVE GENOMICS: Understanding and Using Biological Databases

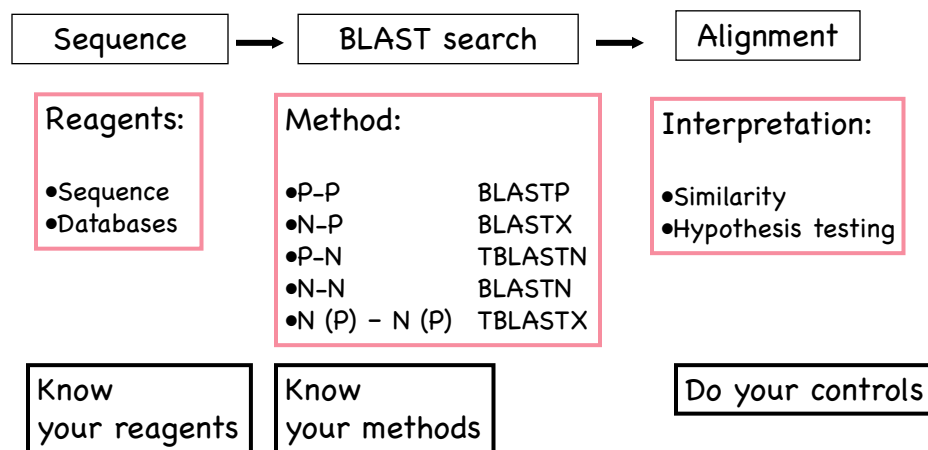
November 30th, 2012

- B.F. Francis Ouellette** francis@oicr.on.ca
- Associate Director, Informatics and Biocomputing, Ontario Institute for Cancer Research, Toronto, ON
 - Associate Professor, Department of Cell and Systems Biology, University of Toronto, Toronto, ON.

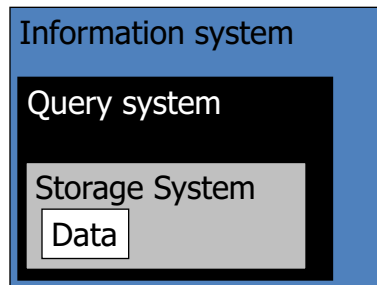
Bioinformatics reagent: **Databases**

- Organized array of information
- Place where you put things in, and (if all is well) you should be able to get them out again.
- Resource for other databases and tools.
- Simplify the information space by specialization.
- Bonus: Allows you to make discoveries.
- Important question to ask:
what is the data model?

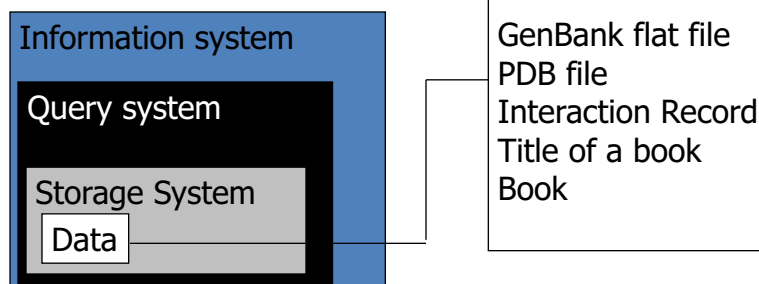
Bioinformatics experiments:



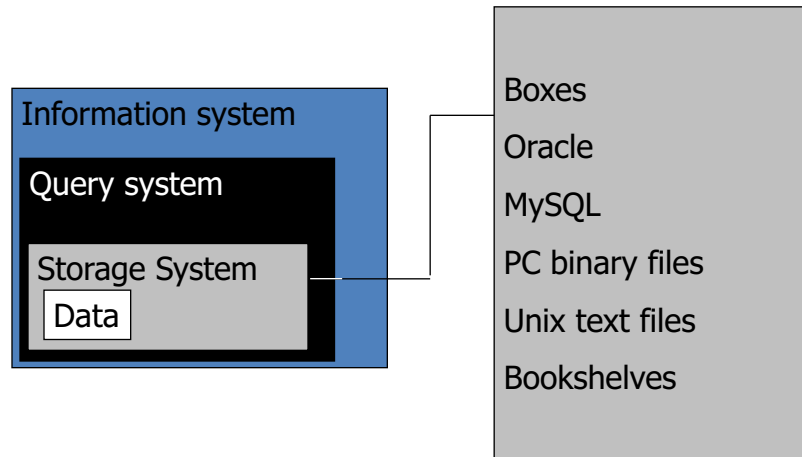
Databases



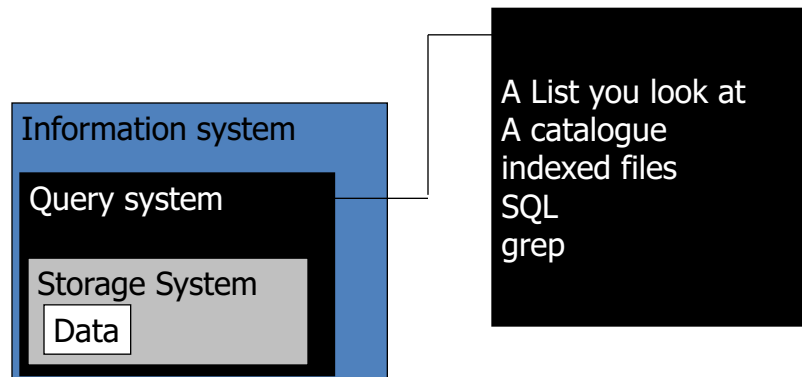
Databases



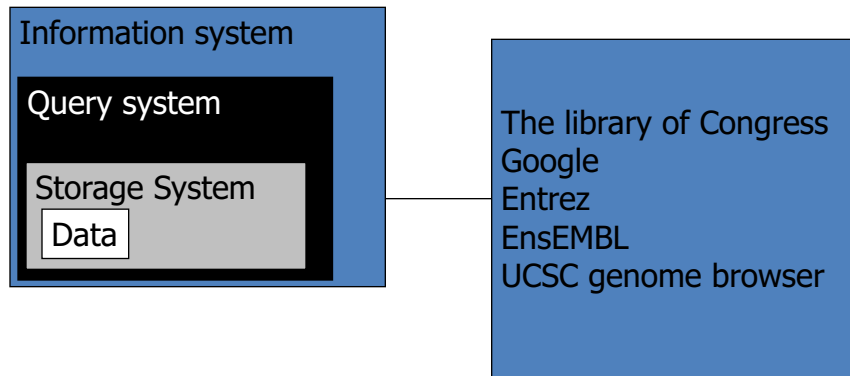
Databases



Databases



Databases



[www.ncbi.nlm.nih.gov/gquery/?term=all\[filter\]](http://www.ncbi.nlm.nih.gov/gquery/?term=all[filter])

Literature

Books	546,057	books and reports
MeSH	268,267	ontology used for PubMed indexing
NLM Catalog	1,557,061	books, journals and more in the NLM Collections
PubMed	26,893,130	scientific & medical abstracts/citations
PubMed Central	4,232,030	full-text journal articles

Health

ClinVar	267,768	human variations of clinical significance
dbGaP	225,719	genotype/phenotype interaction studies
GTR	48,724	genetic testing registry
MedGen	301,782	medical genetics literature and links
OMIM	25,098	online mendelian inheritance in man
PubMed Health	63,102	clinical effectiveness, disease and drug reports

Genomes

Assembly	107,981	genome assembly information
BioProject	211,589	biological projects providing data to NCBI
BioSample	5,685,167	descriptions of biological source materials
Clone	38,262,163	genomic and cDNA clones
dbVar	6,436,080	genome structural variation studies
Genome	22,828	genome sequencing projects by organism
GSS	39,772,962	genome survey sequences
Nucleotide	225,976,870	DNA and RNA sequences
Probe	32,405,227	sequence-based probes and primers
SNP	825,832,256	short genetic variations
SRA	3,625,864	high-throughput DNA and RNA sequence read archive
Taxonomy	1,658,042	taxonomic classification and nomenclature catalog

Genes

EST	76,324,767	expressed sequence tag sequences
Gene	26,489,867	collected information about gene loci
GEO DataSets	2,161,756	functional genomics studies
GEO Profiles	128,414,055	gene expression and molecular abundance profiles
HomoloGene	141,268	homologous gene sets for selected organisms
PopSet	265,235	sequence sets from phylogenetic and population studies
UniGene	6,473,284	clusters of expressed transcripts

Proteins

Conserved Domains	52,411	conserved protein domains
Protein	358,019,768	protein sequences
Protein Clusters	820,546	sequence similarity-based protein clusters
Structure	125,495	experimentally-determined biomolecular structures

Chemicals

BioSystems	944,494	molecular pathways with links to genes, proteins and chemicals
PubChem BioAssay	1,252,713	bioactivity screening studies
PubChem Compound	93,305,710	chemical information with structures, information and links
PubChem Substance	227,858,788	deposited substance and chemical information

<http://www.ncbi.nlm.nih.gov/>
All [filter] Jan, 2017

Formats

- DNA sequence (GenBank Flat Files)
- Protein Sequences
- Other formats to know about
 - FASTA
 - GFF3
 - XML

GenBank Flat File (GBFF)

[illegible]

Header

- Title
- Taxonomy
- Citation

Features (AA seq)

DNA Sequence

FASTA

NCBI

```
>P03069.1 RecName: Full=General control protein GCN4; ...
MSEYQPSLFALNPMGFSPLDGSKSTNENVSASTSTAKPMVGQLIFDKFIKTEEDPI
IKQDTPSNLDFDFALPQTATAPDAKTVLPPELDDAVVESFFSSSTDSTPMFEYEN
LEDNSKEWTSFLFDNDIPVTTDDVSLADKAIESTEEVSLVPSNLEVSTTSFLPTPVL
EDAKLTQTRKVKKPNVSVVKKSHHVGKDDESRDLHLGVVAYNRKQRSIPLSPIVPES
SDPAALKRARNTAARRSRARKLQRMKQLEDKVEELLSKNYHLENEVARLKKLVGE
R
```

uniprot.org

```
>sp|P03069|GCN4_YEAST General control protein GCN4 ... GN=GCN4 PE=1 SV=1
MSEYQPSLFALNPMGFSPLDGSKSTNENVSASTSTAKPMVGQLIFDKFIKTEEDPIIKQD
TPSNLDFDFALPQTATAPDAKTVLPPELDDAVVESFFSSSTDSTPMFEYENLEDNSKEW
TSFLFDNDIPVTTDDVSLADKAIESTEEVSLVPSNLEVSTTSFLPTPVLEDAKLTQTRVK
KPNVSVVKKSHHVGKDDESRDLHLGVVAYNRKQRSIPLSPIVPESDPAALKRARNTAAR
RSRARKLQRMKQLEDKVEELLSKNYHLENEVARLKKLVGER
```

Databases

- Primary (archival)
 - GenBank/EMBL/DDBJ
 - UniProt
 - PDB
 - Medline (PubMed)
 - Intact
- Secondary (curated)
 - RefSeq
 - Taxon
 - UniProt
 - OMIM
 - SGD
 - Biosamples/BioProjects

<https://nar.oxfordjournals.org/content/45/D1.toc> January 2017

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Database issue
Volume 44 Issue D1 04 January 2016

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☒ **Nucleic acid sequence, structure, and regulation**

☐ Daniel J. Rigden, Xosé M. Fernández-Suárez, and Michael Y. Galperin
The 2016 database issue of *Nucleic Acids Research* and an updated molecular biology database collection
Nucleic Acids Res. (04 January 2016) 44 (D1): D1-D6 doi:10.1093/nar/gkv1356
» Abstract » Full Text (HTML) » Full Text (PDF) » Database Summaries » Permissions

☐ NCBI Resource Coordinators
Database resources of the National Center for Biotechnology Information
Nucleic Acids Res. (04 January 2016) 44 (D1): D7-D19 doi:10.1093/nar/gkv1290
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☐ Charles E. Cook, Mary Todd Bergman, Robert D. Finn, Guy Cochrane, Ewan Birney, and Rolf Apweiler
The European Bioinformatics Institute in 2016: Data growth and integration
Nucleic Acids Res. (04 January 2016) 44 (D1): D20-D26 doi:10.1093/nar/gkv1352
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Alert me to new issues

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Sequence Databases

- Primary DNA (archive) (avoid)
 - DDBJ/ENA/GenBank
- Primary protein (curated/automation)
 - UniProtKB
- Curated Databases (lots of human labour)
 - RefSeq (Genomic, mRNA and protein)
 - UniProtKB/SwissProt and neXtprot

Identifiers

- You need identifiers which are stable through time
- Need identifiers which will always refer to specific sequences
- Need these identifiers to track history of **sequence** updates
- Also need feature and annotation identifiers (need to track important things)
 - Genes
 - Transcripts
 - Proteins
 - (((Phenotype)))

LOCUS, Accession, NID and protein_id

LOCUS: Unique string of 10 letters and numbers in the database. Not maintained amongst databases, and is therefore a poor sequence identifier.

ACCESSION: A unique identifier to that record, citable entity; does not change when record is updated. A good record identifier, ideal for citation in publication.

VERSION: ID system where the accession and version play the same function as the accession and gi number.

protein_id: Identifier which has the same structure and function as the nucleotide Accession.version numbers, but slightly different format.

LOCUS, Accession, (gi) and PID

LOCUS	HSU40282	1789 bp	mRNA	PRI	21-MAY-1998
DEFINITION	Homo sapiens integrin-linked kinase (ILK) mRNA, complete cds.				
ACCESSION	U40282				
VERSION	U40282.1				

LOCUS: HSU40282
 ACCESSION: U40282
 VERSION: U40282.1
 protein_id: AAC16892.1

LOCUS
 ACCESSION
 Protein_id

CDS	157..1515
	/gene="ILK"
	/note="protein serine/threonine kinase"
	/codon_start=1
	/product="integrin-linked kinase"
	/protein_id="AAC16892.1"

In closing ...

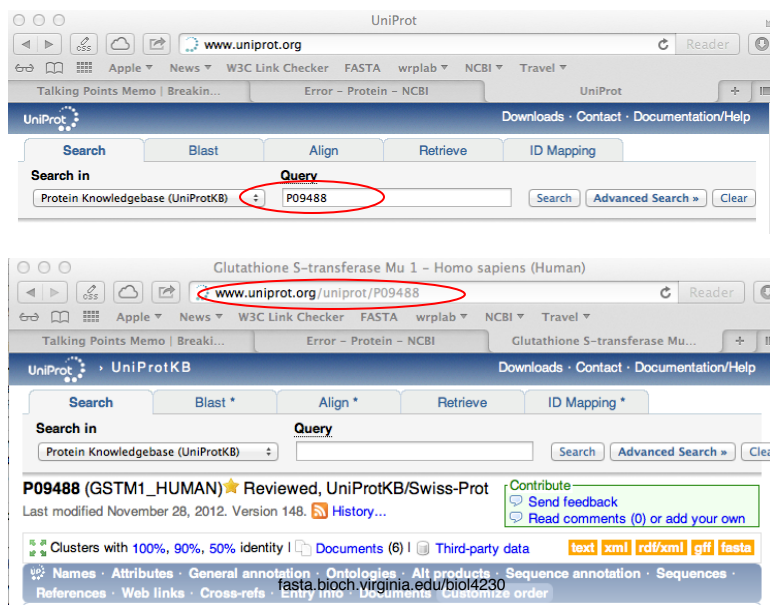
- Often only use FASTA files (e.g. for BLAST)
- Using any sequence where the coordinates are important, need an accession.version
- Keep in mind that GenBank is DNA centric and is a poor vehicle for protein and mRNA expression/interaction information: NCBI (and others) have other databases for these entities.
- All databases I mentioned today are fully “open” ...

Scripting from the WWW: `wget/curl`

- Most bioinformatics analyses require resources from the web, e.g. sequences, domain information, datasets, etc.
 - The NCBI and EBI resources are usually scriptable; e.g. write a script that takes a set of accessions from a file and get the sequences
 - Often all that is required is to recognize the URL of the information desired
<http://www.ncbi.nlm.nih.gov/protein/P09488>
 - Sometimes, you will need more information to get the desired format (e.g. FASTA)
<http://www.ncbi.nlm.nih.gov/protein/121735?report=fasta>
- `curl` and `wget` allow you to pull a web page into a file from the command line:
`curl http://uniprot.org/uniprot/P09488.fasta > p09488.fasta`
- Sometimes this is what you need; other times more work is required

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Finding a URL (www.uniprot.org)



Finding a URL to download (uniprot)

Glutathione S-transferase Mu 1 - Homo sapiens (Human)

www.uniprot.org/uniprot/P09488

UniProtKB

Search in: Protein Knowledgebase (UniProtKB) Query

P09488 (GSTM1_HUMAN) ★ Reviewed, UniProtKB/Swiss-Prot
Last modified November 28, 2012. Version 148. History...

Clusters with 100%, 90%, 50% identity | Documents (6) | Third-party data | text | xml | rdf/xml | **fasta**

Names · Attributes · General annotation · Ontologies · Alt products · Sequence annotation · Sequences · References · Web links · Cross-refs · Entry info · Documents · Customize order

www.uniprot.org/uniprot/P09488.fasta

www.uniprot.org/uniprot/P09488.fasta

```
>sp|P09488|GSTM1_HUMAN Glutathione S-transferase Mu 1 OS=Homo sapiens GN=GSTM1 PE=1 SV=3
MPMILGYWDIRGLAHAIRLLLEYTDSYEEKKTYMGDAPDYDRSOWLNEKFKLGLDFNL
PYLIDGAHKITQSNAILCYIARKHNLGCTEEKIRVDILENQTMDSNMQLGMICYNPEF
EKLKPKYLEELPEKILKYSEFLGKRPWFAGNKITFVDFLVYDVLDRHIFEPKCLDAPPN
LKDFISRFEGLKISAYMKSSRFLPRPVFSKMAVWGNK
```

```
curl http://www.uniprot.org/uniprot/P09488.fasta
fasta.bioch.virginia.edu/biol4230
```

Finding a URL to download (NCBI)

National Center for Biotechnology Information

www.ncbi.nlm.nih.gov

NCBI Resources How To

Protein **P09488** Search

RecName: Full=Glutathione S-transferase Mu 1; AltName: Full=GST HB sub - Protein - NCBI

www.ncbi.nlm.nih.gov/protein/P09488

Protein

Display Settings: GenPept

RecName: Full=Glutathione S-transferase Mu 1; AltName: Full=GST HB subunit 4; AltName: Full=GST class-mu 1; AltName: Full=GSTM1-1; AltName: Full=GSTM1a-1a; AltName: Full=GSTM1b-1b; AltName: Full=GTH4

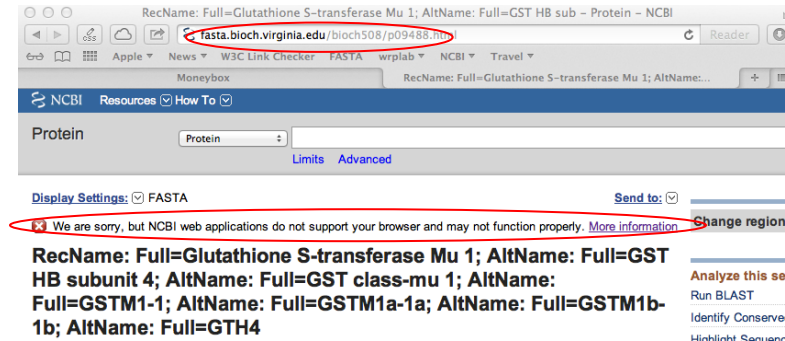
UniProtKB/Swiss-Prot: P09488.3

[FASTA](#) [Graphics](#)

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Finding a URL to download (NCBI)

```
curl http://www.ncbi.nlm.nih.gov/protein/P09488?report=fasta
```



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NCBI e-utilities

- The NCBI does not allow their web server to be used for large-scale, automated downloads (unlike Uniprot)
- NCBI provides e-utilities (esearch.cgi, efetch.cgi) for programmatic access to ALL NCBI databases (proteins, DNA, also PubMed)

www.ncbi.nlm.nih.gov/guide/howto/dwn-records/

www.ncbi.nlm.nih.gov/books/NBK25500/ (this document is currently out of date because it still users GI numbers)

In 2017, NCBI also uses accessions for downloads, so downloading a fasta file is easy:

```
curl
'https://eutils.ncbi.nlm.nih.gov/entrez/eutils/efetch.fcgi?db=protein&id=P09488&rettype=fasta&retmode=text'
```

Quotes are required to protect '&' and '?' from shell

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Homework (due Monday, Jan. 29, 12:00 noon)

1. Do a search of the SwissProt database using `blastp` using NP_001171499 (honeybee_gst.aa) saving the output in "tabular" format (-outfmt 6)
2. Repeat step 1, using the `ssearch36` program specifying the BLOSUM62 matrix (-s BP62). (you can produce tabular output using the -m 8 option):

```
ssearch36 -m 8 -s BP62 honeybee_gst.aa q > output
```
3. For both the `blastp` and `ssearch` results, make a *copy* of each results file and remove all the lines with $E() > 0.001$. Write a bash script to isolate the library (subject) accession information for each of the lines in the edited file, and save the accession in a new file
4. For each accession, split it into its component parts (hit 'man cut' to see how to change the delimiter).
 – Write a script to save the accessions (P12345.3) to a file, and isolate only the accessions without the version information.
5. Compare the list of SwissProt accessions with $E() < 0.001$ from BLASTP and SSEARCH. Which program finds more homologs? For the program that finds fewer homologs, what are the $E()$ -values of those hits in the list of hits from the other program?

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Homework (due Monday, Jan. 29)

7. Edit new *copies* of the original `blastp` and `ssearch` output files file to save the lines with $0.1 < E()$ -values < 2.0 (you can do this by hand, or with a script) The 'awk' program makes it very easy to parse tab-delimited files for lines that meet criteria and print the `sseqid`, e.g.

```
awk '($11 > 0.1 && $11 < 2.0){print $2}' tab.output
```

 In this example, the $E()$ -value is in column 11 (\$11), and the `sseqid` in column 2 (\$2)
7. For the accessions $0.1 < E() < 2.0$ from step 7, run the script from steps 4,5 to isolate the SwissProt accessions. Then use the protein accessions to get the sequences from UniProt.
8. Write a script to take the accessions from with $0.1 < E() < 2.0$ from the `blastp` search and re-search Swissprot for each of those accessions, saving the new search results in files named after the accession numbers.
9. Write a description of your work in the file "hwk2.notes", labeling the scripts that you wrote, and save the description, scripts, and results files in biol4230/hwk2.

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