I519: Homework 1

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Overview

Set working directory

```
rm(list=ls())
getwd()

## [1] "/Users/WRShoemaker/github/PopGen/I519/HW1"

setwd("~/github/PopGen/I519/HW1/")
```

Import packages

```
library("quantmod")
## Loading required package: xts
## Loading required package: zoo
##
## Attaching package: 'zoo'
##
## The following objects are masked from 'package:base':
##
       as.Date, as.Date.numeric
##
##
## Loading required package: TTR
## Version 0.4-0 included new data defaults. See ?getSymbols.
library("ggplot2")
library("reshape2")
library("wesanderson")
library(data.table)
##
## Attaching package: 'data.table'
## The following object is masked from 'package:xts':
##
##
       last
```

Import the data and add headers

```
draft <- read.table("./genesize-draft.txt",header=F)
complete <- read.table("./genesize-complete.txt",header=T)

colnames(draft) <- c("Strain", "Genes")
colnames(complete) <- c("Strain", "Genes")</pre>
```

Mean gene number

```
mean(draft[,2])
## [1] 2678.527
mean(complete[,2])
```

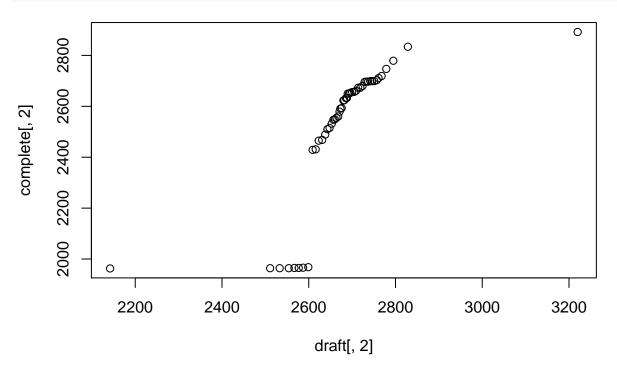
[1] 2521.816

So the draft genomes have a mean gene number of $\sim 2,679$ and the complete genomes have a mean gene number of ~ 2522 . Let's look if there's a significant difference.

Examine the quantile-quantile plots for the samples

First we make a Q-Q plot that examines both samples.

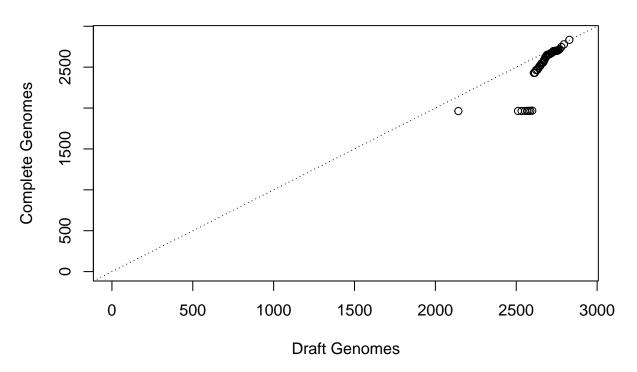
```
qqplot(draft[,2], complete[,2])
```



```
xlim = range(1800, draft)
ylim = range(1800, complete)

qqplot(draft[,2], complete[,2],
xlim = ylim, ylim = ylim,
xlab = "Draft Genomes",
ylab = "Complete Genomes",
main = "Gene Number")
abline(a=0, b=1, lty="dotted")
```

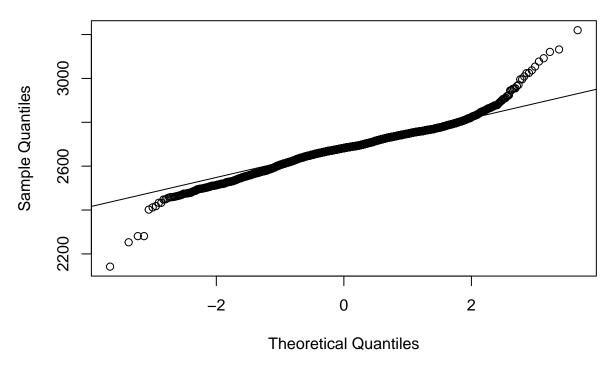
Gene Number



Then we make a Q-Q plot for each sample that compares the theoretical quantiles with sample quantiles.

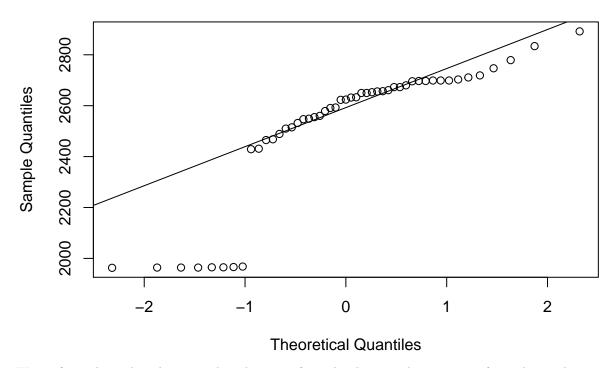
```
qqnorm(draft[,2],
main = "Gene number in draft genomes")
qqline(draft[,2])
```

Gene number in draft genomes



```
qqnorm(complete[,2],
main = "Gene number in complete genomes")
qqline(complete[,2])
```

Gene number in complete genomes



We see from above that there is a clear deviation from the theoretical expectation for each sample, suggesting

that it would not be a good idea to assume normality. We can then proceed with a Wilcoxon rank-sum test.

```
wilcox.test(draft[,2], complete[,2], alternative = c("two.sided"), paired = FALSE, conf.level = 0.95)
##
##
   Wilcoxon rank sum test with continuity correction
##
## data: draft[, 2] and complete[, 2]
## W = 140410, p-value = 6.078e-07
## alternative hypothesis: true location shift is not equal to 0
There is a significant difference in the number of genes between draft and complete genomes.
draft$Type <- rep("Draft",nrow(draft))</pre>
complete$Type <- rep("Complete",nrow(complete))</pre>
mergeData <- rbind(draft, complete)</pre>
mergeData$logGenes <- log(mergeData[,2])</pre>
class(mergeData)
## [1] "data.frame"
head(mergeData)
##
                                         Strain Genes Type logGenes
## 1 Staphylococcus_aureus_06BA18369_uid170654 2804 Draft 7.938802
## 2 Staphylococcus_aureus_07_03450_uid226303 2476 Draft 7.814400
## 3 Staphylococcus_aureus_07_03451_uid226304 2484 Draft 7.817625
## 4 Staphylococcus_aureus_08_01059_uid226297 2566 Draft 7.850104
## 5 Staphylococcus_aureus_08_01062_uid226298 2566 Draft 7.850104
## 6 Staphylococcus aureus 08 01084 uid226305 2574 Draft 7.853216
palette <- wes_palette(5, name = "FantasticFox", type = "discrete")</pre>
ggplot(mergeData, aes(x=Type, y=Genes, fill = Type)) +
  geom_violin(trim=FALSE) +
  geom_boxplot(width=0.1) +
  scale_fill_manual(values=palette[-2]) +
  xlab("Genome Type") +
  scale_size_area("Genome Type") +
  ylab("Gene Number") +
  guides(fill=guide legend(title=NULL))
```

