Curriculum Vitae

W. Zane Billings

Contact Information

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Education

Western Carolina University

Mathematics and Biology, B.S., 4.0 GPA

2016-2020

• Concentrations: Applied mathematics, molecular biology

• Minor: Chemistry

Honors and Awards:

• Chancellor's List, every semester

- WCU Distinguished Achievement Scholarship (2016 2020)
- Several other scholarship awards from Biology, Mathematics, and the Honors College.
- Math and Computer Science Department Ambassador, Fall 2018 onward
- COMAP Mathematical Contest in Modeling 2019: Meritorious Winnter (Problem A)

Employment

Coan Lab at Western Carolina University

Cullowhee, NC

Cullowhee, NC

Fall 2018-Present

Lab manager, researcher

- Supervised by Dr. Heather Coan at WCU.
- Researched the autophagy pathway, focused on stress-induced autophagy in human cells.
- Utilized several techniques from mammalian cell culture and fluorescence microscopy.
- Ensured the lab environment, including the cell culture facility, was tidy and complied with BSL-2 and OSHA saftey standards.
- Prepped cell cultures and solutions for other lab workers and assisted with use of lab technology such as the fluorescence microscope and culture room.
- Worked with the Advanced Techniques in Microscopy Laboratory, mainly assisting with data analysis protocols.

Mathematics and Computer Science Department, WCU

Cullowhee NC

Mathematics lab assistant

Spring 2019 – present

- Worked as a lab assistant for the hands-on, programming based course MATH 340: Introduction to Scientific Computing. This course is designed as a first introduction to programming and scientific computing, and is primarily taken by math, computer science, and engineering students.
- Assisted several students with LATEX, MATLAB, R, and Mathematica coding, as well as Excel spreadsheet development.
- Developed educational activities to strengthen coding skills for in-class use.

Population Biology of Infectious Diseases REU

Athens, GA

Summer 2019

Researcher; REU participant, Handel Research Group

- Participated in the 2019 Summer NSF-funded REU program at the University of Georgia, Athens.
- Wrote R code to perform a variety of tasks, including cleaning and joining data, summarizing and visualizing data, and constructing models.
- Analyzed cross-sectional data from a study performed at UGA's Student Health Clinic, in order to analyze the clinical revelance of qPCR data to influenza prognosis and care.
- Communicated results of data analysis through markdown, intermediate progress presentations, and a final poster presentation.
- Worked with Dr. Handel to prepare walkthroughs and solutions to several learning modules for the DSAIDE R

Math-Bio REU at the University of North Carolina, Greensboro

Greensboro, NC

Researcher; REU participant, Bobay Research Group

Summer 2018

- Utilized methods from genomics to analyze large DNA sequence datasets.
- Visualized data for the project using R.
- Contributed to mathematical models of convergent mutations between DNA strands.
- Assisted in the design of Python simulations to verify findings.

Writing and Learning Commons at WCU

Cullowhee, NC

Course Tutor

Spring 2017–Spring 2019

- Tutored peers in material from introductory biology, organic chemistry and genetics. As of Spring 2019, I am tutoring Immunology, Advanced Techniques in Microscopy, and Evolutionary Biology.
- Identified strategies for working with students who had different learning techniques.
- Communicated with professors regarding material students did and did not know, and questions which came up commonly during tutoring.

New River Engineering, PLLC

Mount Holly, NC

July 2014– August 2016

I proofread emails, project proposals, and technical reports to provide grammatical and stylistic corrections.

Research Experience

Cell Biology

Western Carolina University

Mentor: Heather Coan

Spring 2016–Present

I conducted independent research on autophagic flux. My research utilized several tools from molecular biology and microscopy in order to elucidate stress-induced autophagic flux in human cells. I maintained bacterial cultures and extracted plasmids, maintained human cell cultures, transfected plasmids into human cells, and stressed cells before visualization using widefield fluoresence microscopy. After visualization, I was involved in image processing and designed analyses for the data.

Epidemiology and Biostatistics

University of Georgia, Athens

Mentor: Andreas Handel

Summer 2019

As part of the Population Biology of Infectious Diseases REU, funded by the NSF and NIH, I worked with the Handel research group. My research included the development of software packages in the R language for epidemiology education, and analysis of flu data collected at the UGA student health center.

Computational Biology

University of North Carolina, Greensboro

Mentor: Louis-Marie Bobay

Summer 2018

As part of the Math-Biology REU site, I worked on a team modeling the rate of convergent mutation between bacterial genomes. In order to predict rates of homologous recombination, we constructed a probabilistic model of recombination and designed and ran several simulations to verify.

Organic Chemistry

Western Carolina University

Mentor: William Kwochka

Fall 2017–Spring 2018

I conducted independent research in organic chemistry. The lab group's work focused on characterizing the nature of the "dative bond" between Lewis acid-base complexes. I synthesized several derivatives of an organic boroncontaining compound and complexed several of these bonds. Additionally, I characterized these compounds using spectroscopic methods.

Behavioral Ecology

Western Carolina University

Mentor: Jeremy Hyman

Summer 2016

As part of a two-week program (SURP) prior to my first semester at WCU, I assisted Dr. Hyman and a senior undergraduate student with her thesis work on the behavior of song sparrows. I mainly participated in the field component of the research, assisting with bird song playback experiments as well as catching a bird.

Skills

Lab techniques: Aseptic technique, mammalian Programming: Proficient with R (including Base Program responsible conduct in research certification.

cell culture and transfection, fluorescence and Tidyverse), LATEX, and Markdown, intermediate microscopy, PCR, ELISA, Western blot, CITI proficiency with Python, familiar with Git and CSS.

Relevant Coursework

Biology: Immunology, Advanced Techniques in Mathematics: Microscopy, Protein Systems and Bioinformatics, Algebra, Statistical Methods (I-Intro & II-Cell Biology, Evolutionary Biology, Origin of Time Series and Linear Regression), Scientific Species

Differential Equations, Linear Computing, Linear Optimization, Introductory Probability Theory, Mathematical Modeling