



Original Research Article

L-leucine stimulates glutamate dehydrogenase activity and glutamate synthesis by regulating mTORC1/SIRT4 pathway in pig liver

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ABSTRACT

The liver is the most essential organ for the metabolism of ammonia, in where most of ammonia is removed by urea and glutamine synthesis. Regulated by leucine, glutamate dehydrogenase (GDH) catalyzes the reversible inter-conversion of glutamate to ammonia. To determine the mechanism of leucine regulating GDH, pigs weighing 20 ± 1 kg were infused for 80 min with ammonium chloride or alanine in the presence or absence of leucine. Primary pig hepatocytes were incubated with or without leucine. In the *in vivo* experiments with either ammonium or alanine as the nitrogen source, addition of leucine significantly inhibited ureagenesis and promoted the production of glutamate and glutamine in the perfused pig liver ($P < 0.05$). Similarly, leucine stimulated GDH activity and inhibited sirtuin4 (*SIRT4*) gene expression ($P < 0.01$). Leucine could also activate mammalian target of rapamycin complex 1 (mTORC1) signaling ($P < 0.05$), as evidenced by the increased phosphorylation levels of ribosomal protein S6 kinase 1 (S6K1) and ribosomal protein S6 (S6). Interestingly, the leucine-induced mTORC1 pathway activation suitably correlated with increased GDH activity and decreased expression of *SIRT4*. Similar results were observed in primary cultured hepatocytes. Notably, leucine exerted no significant change in GDH activity in *SIRT4*-deficient hepatocytes ($P > 0.05$), while mTORC1 signaling was activated. Leucine exerted no significant changes in both GDH activity and *SIRT4* gene expression in rapamycin treated hepatocytes ($P > 0.05$). In conclusion, L-leucine increases GDH activity and stimulates glutamate synthesis from different nitrogen sources by regulating mTORC1/SIRT4 pathway in the liver of pigs.

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1. Introduction

In a previous study in pigs, about 50% of total ingested nitrogen was found to be excreted by urine (Jongbloed and Lenis, 1992). Nitrogen urinary excretion occurs mainly as urea, which is mainly converted from ammonia in the liver. Ammonia is produced in the catabolism of proteins, amino acids (AA) and other nitrogen compounds (Häussinger, 2007). There are 2 major pathways for

ammonia detoxification by the liver: urea and glutamine synthesis (Häussinger, 1990; Olde Damink et al., 2002). As a final product, urea cannot be metabolized by mammalian enzymes and is exclusively excreted by the kidney (Hallbrucker et al., 1994). Glutamine, however, is a functional nonessential amino acid (NEAA) that can be further utilized. Notably, glutamate dehydrogenase (GDH) plays a central role in hepatic ammonia and AA metabolism, because the GDH activity is the highest in mammals liver and it can catalyze the reversible inter-conversion of glutamate to α -ketoglutarate and ammonia (Broshnan and Broshnan, 2009).

Leucine is one of branched-chain, essential amino acid (BCAA; EAA) that cannot be synthesized *de novo* by mammals (Hutson, 2006). Recently, studies have indicated that BCAA has a low metabolic rate in pig liver and it is now well recognized that leucine not only has a role as a substrate for protein synthesis, but also acts as a signal that regulates nutrient metabolism (Li et al., 2015; Anthony et al., 2001). Previous studies have shown that leucine can activate the mammalian target of rapamycin complex-1

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(mTORC1) pathway and GDH activity but the molecular mechanism of GDH regulated by leucine remains unclear (Stipanuk, 2007; Yang et al., 2010; Yoshizawa et al., 2013). The sirtuins (SIRT) are a family of nicotinamide adenine dinucleotide (NAD⁺) dependent deacetylase proteins with broad cellular functions including regulation of energy production and nutrition metabolism (Zhong and Mostoslavsky, 2011; Rajendran et al., 2011). Three sirtuins, SIRT3–5, localize to mitochondria, were all associated with the metabolism of ammonia. Sirtuin3 directly deacetylates ornithine transcarbamylase (OTC) and stimulates its activity (Hallows et al., 2011). Sirtuin5 plays a pivotal role in ammonia detoxification and disposal by activating carbamoyl phosphate synthetase 1 (CPS1) (Nakagawa et al., 2009). Interestingly, Leprieur et al. (2013) demonstrated that the mammalian Sir2 homolog SIRT4 could regulate GDH activity. Csibi et al. (2013) demonstrated that the mTORC1 pathway could stimulate glutamine metabolism and cell proliferation by repressing SIRT4. Therefore, we hypothesized that leucine might increase GDH activity and stimulate glutamate synthesis by regulating mTORC1/SIRT4 pathway in pigs liver.

The aim of this study was to investigate the mechanism by which leucine regulates the GDH activity via mTORC1/SIRT4 pathway in pigs liver. For that purpose, ¹⁵N-ammonium chloride and ¹⁵N-alanine were utilized as metabolic tracers in order to identify sources of nitrogen for hepatic urea, glutamate and glutamine synthesis in a pig liver perfusion system. The pig liver perfusion system is a practical technique to study hepatic nutrient metabolism in the organ level. These experiments were done in the presence and absence of physiologic concentrations of portal venous leucine with ammonium chloride and alanine as different nitrogen sources. Primary pig hepatocytes were transfected with small interfering RNA (siRNA) for SIRT4 knockdown or treated with rapamycin followed by treatment with leucine.

2. Materials and methods

2.1. Animals

The study protocol was approved by the Animal Care and Use Committee of College of Animal Sciences and Technology, Huazhong Agricultural University, and was carried out in accordance with the National Research Council's Guide for the Care and Use of Laboratory Animals. Three male piglets (Duroc × Landrace × Yorkshire) were killed at 3 days of age for isolation of hepatocytes. Fifteen castrated male pigs (Duroc × Landrace × Yorkshire, 20 ± 1 kg body weight) were individually housed in 1.5 m × 0.75 m metabolic cages located in a 20 to 21 °C air-conditioned room.

2.2. Surgery and postoperative management

After a 5-day adaptation period, 12 pigs were surgically catheterized through the portal and hepatic veins. Catheters (MMVCVB1, Tiandihexie Technology Company, Beijing, China) were made of modified polyurethane, measuring 30 cm in length with 2.41 mm outer diameter. The procedure was conducted under isoflurane anesthesia and strict aseptic conditions. The catheters were flushed aseptically daily with 200 IU of heparinized normal saline to maintain their patency, protected with gauze pads and secured with a suitable dressing. The pigs received an intramuscular injection of 1.6 × 10⁷ IU penicillin and 1.0 × 10⁷ IU streptomycin twice daily during the first 3 days following the operation. The pigs were

progressively returned to their previous levels of dietary intake for 5 d before the experiments started.

2.3. Treatments and infusions

After post-surgery recovery (5 days) 12 healthy pigs with well-kept catheters that were food deprived overnight were randomly divided into 2 equal groups ($n = 6$) to be treated with ammonia or alanine. Each group was then subdivided into 2 subgroups of 3 animals each to be infused with ¹⁵N-ammonium chloride (NLM-467-PK, 99% enriched in ¹⁵N; Cambridge Isotope Laboratories) with or without leucine (L8912, Sigma-Aldrich) and those infused with ¹⁵N-alanine (NLM-454-PK, 98% enriched in ¹⁵N; Cambridge Isotope Laboratories), also with or without leucine. All pigs were perfused for 10 min with sterile saline, and then infused through the hepatic portal vein with the corresponding substrate (with or without leucine) for 80 min. The ammonium chloride and alanine infusions were done 1.5 h after feeding at a rate according to their physiological concentrations following a previously reported method. Blood samples were taken from the hepatic vein at 0, 20, 40, 60 and 80 min after perfusion. After 80 min the pigs were killed and tissue samples were collected from their livers. Moreover, another 3 pigs without treatments/perfusion were killed at the same time and their liver samples were also collected to normalize the expression corresponding signaling molecules. The plasma and tissue samples were quick-frozen in liquid nitrogen and maintained at −70 °C until analyzed for urea, individual AA concentrations and related gene expressions. Other freshly collected liver samples were homogenized and centrifuged at 10,000 × g for 10 min at 4 °C. Supernatants were diluted in sample buffer, frozen in liquid nitrogen, and stored at −70 °C until protein analysis (Suryawan et al., 2008).

2.4. Isolation and culture of hepatocytes

Hepatocytes were isolated from normal livers by collagenase perfusion and mechanical disruption as described previously (Donato et al., 1999). Cell viability was assessed by Trypan blue exclusion and was always greater than 85%. Cells were incubated in a medium containing 10% fetal bovine serum (10,099-141, Gibco) and an antibiotic solution (15,140-122, Gibco) at 37 °C in a 5% CO₂ humidified atmosphere. After isolation and culture of hepatocytes for 48 h, primary cultured hepatocytes were transfected with SIRT4 siRNA (Production No. sc-63024, Santa Cruz), or scrambled control siRNA to silence SIRT4 protein. The hepatocytes were transfected with Lipofectamine 2000 (11,668,019, Invitrogen) using 150 pmol/well of SIRT4 siRNA and scrambled SCR as a control (Santa Cruz Biotechnology) according to the manufacturer's instructions (Ho et al., 2013). The hepatocytes were then transfected with corresponding siRNAs and incubated in the presence or absence (control) of 5 mmol/L leucine for 4 h. Correspondingly, hepatocytes were treated with or without 20 ng/mL rapamycin (37,094, Sigma-Aldrich) for 24 h followed by treatment with 5 mmol/L leucine (L8912, Sigma-Aldrich) for the last 4 h.

2.5. Analyses of metabolites and enzymatic activities

Plasma urea was measured using an automatic biochemical analyzer (Siemens). ¹⁵N-labeled AA and aspartate from plasma and

tissue samples were detected by LC-MS as described by Nissim et al. (2012). The enzymatic activity of GDH was measured using a Sigma-Aldrich kit (MAK099), according to the manufacturer's instructions.

2.6. RNA isolation and quantitative real-time PCR

Total RNA was extracted from the livers and hepatocytes using the TRIzol reagent (Invitrogen Corporation, Carlsbad, CA, US) according to the manufacturer's specifications. Reverse transcription-PCR and real-time quantitative PCR analysis were detected as described by Huang et al. (2011). Briefly, reverse transcription of total RNA was performed using an avian myeloblastosis virus RT with a first-strand complementary DNA synthesis kit for reverse transcription-PCR. Aliquots of the reverse transcription reactions were then submitted in duplicate to online quantitative PCR with the LightCycler 480 Real-Time PCR system (Roche Applied Science) with SYBR green using the FastStart DNA-Master SYBR Green I kit (Roche Applied Science). Primer specific for *SIRT4* (Accession No. EU357900, forward 5'-TCTCTCCTGGTGGGGTCATCC-3', reverse 5'-GATGCCAAGTTACCCGACCGTGTG-3') was based on previous studies (Ren et al., 2013). β -actin was used as a housekeeping gene to normalize target gene expression levels (Gessner et al., 2016). Notably, the expression of β -actin was unchanged between the control and treatment groups (CT value: ammonia vs. leucine + ammonia, $P = 0.975$; alanine vs. leucine + ammonia, $P = 0.922$). Specific primers were synthesized commercially (Shanghai Sangon Biological Engineering Technology and Services Company, Limited, Shanghai, China). Primer sequences, genes accession number and optimal PCR annealing temperatures (t_a) are listed in Table 1.

2.7. Immunoblotting

Protein extracts were prepared from liver tissues and whole-cell lysates. Equal amounts of protein samples were separated by electrophoresis in polyacrylamide gels and then transferred to a polyvinylidene difluoride (PVDF) membrane that was incubated with the appropriate primary antibodies, washed, and exposed to an appropriate secondary antibody. Western blot analysis was performed using antibodies against *SIRT4* (ab139128, Abcam), *GDH* (ab34786, Abcam), ribosomal protein S6 kinase 1 (*S6K1*) (total 9202 and phosphorylated 9205, Cell Signaling Technology), ribosomal protein S6 (*S6*) (total and phosphorylated 2211 or 2215, Cell Signaling Technology), α -tubulin (2144, Cell Signaling Technology), *CPS1* (ab3682, Abcam) to detect protein expression.

2.8. Cellular viability and MTT assay

The activity of lactate dehydrogenase (LDH) was used as a measure of cell viability. Lactate dehydrogenase leakage into the culture media was respectively measured after hepatocytes isolation and 24 h siRNA transfection using an LDH assay kit (K726-500, Biovision) as following the manufacturer's instructions. The MTT was assayed as described by Soria et al. (2013).

2.9. Statistics

Analysis was performed using an one-way analysis of variance (ANOVA) or a *T*-test analysis in SPSS (17.0), and significant differences were indicated by single asterisk (*) when $P < 0.05$ and double asterisk (**) when $P < 0.01$.

3. Results

3.1. L-leucine stimulates glutamate synthesis and inhibits ureagenesis from ammonium chloride

The effect of leucine on the metabolism of ammonia in perfused pig liver was determined using ammonium chloride as a substrate, which caused an increase in hepatic urea output throughout the perfusion experiment (Fig. 1A). When leucine was used concurrently with ammonium chloride, the urea output was significantly reduced after 40 min and the hepatic vein ^{15}N -labeled glutamate and glutamine concentrations were significantly increased at 40- and 60-min, respectively ($P < 0.05$, Fig. 1A–C). Consistent with this, the amounts of ^{15}N -labeled glutamate and glutamine in liver tissue samples were significantly greater when leucine was added during perfusion with ammonium chloride ($P < 0.01$, Fig. 1D). Additionally, leucine caused a relatively low but significant consumption of aspartate when the nitrogen source was ammonium chloride ($P < 0.05$, Fig. 1E).

3.2. L-leucine inhibits ureagenesis and stimulates glutamate synthesis from alanine

The effect of leucine on the dynamic changes of hepatic vein urea, glutamate and glutamine from alanine in perfused pig liver was included in this study. On the whole, the infusion of alanine had an increase in the output of urea and of ^{15}N -labeled glutamate and glutamine. In the hepatic vein the addition of leucine significantly decreased the concentration of urea and increased the concentrations of ^{15}N -labeled glutamate and glutamine after 40 min ($P < 0.05$, Fig. 2A to C). After 80 min of perfusion the hepatic

Table 1
Oligonucleotide PCR primers.

Gene	Orientation	Primer source	Primer sequences (5' to 3')	Accession No.	Product size, bp	t_a , °C
<i>GDH</i>	Forward	Sus	ACATTGGGCTCCCTCTTTGG	NM_001244501	124	59.96
	Reverse		CCCCTGGGAATGAAGCTGT			
<i>CPS1</i>	Forward	Sus	AAGGCAAAGGAGATTGGGTCT	XM_005672159	147	59.89
	Reverse		TACTGATGGGTATTCTGCAGCC			
<i>GLUL</i>	Forward	Sus	GGGCGAGAAAGTCCAAGCTAT	NM_213909	187	60.26
	Reverse		CATGGCAGCAGGACAAGAT			
β -actin	Forward	Sus	GACATCCGCAAGGACCTCTA	XM_003124280	205	—
	Reverse		ACATCTGCTGGAAGTGGAC			

t_a = annealing temperatures; *GDH* = glutamate dehydrogenase; *CPS1* = carbamoyl phosphate synthetase 1; *GLUL* = glutamate-ammonia ligase; Sus = *S. acrofa* (Pig).

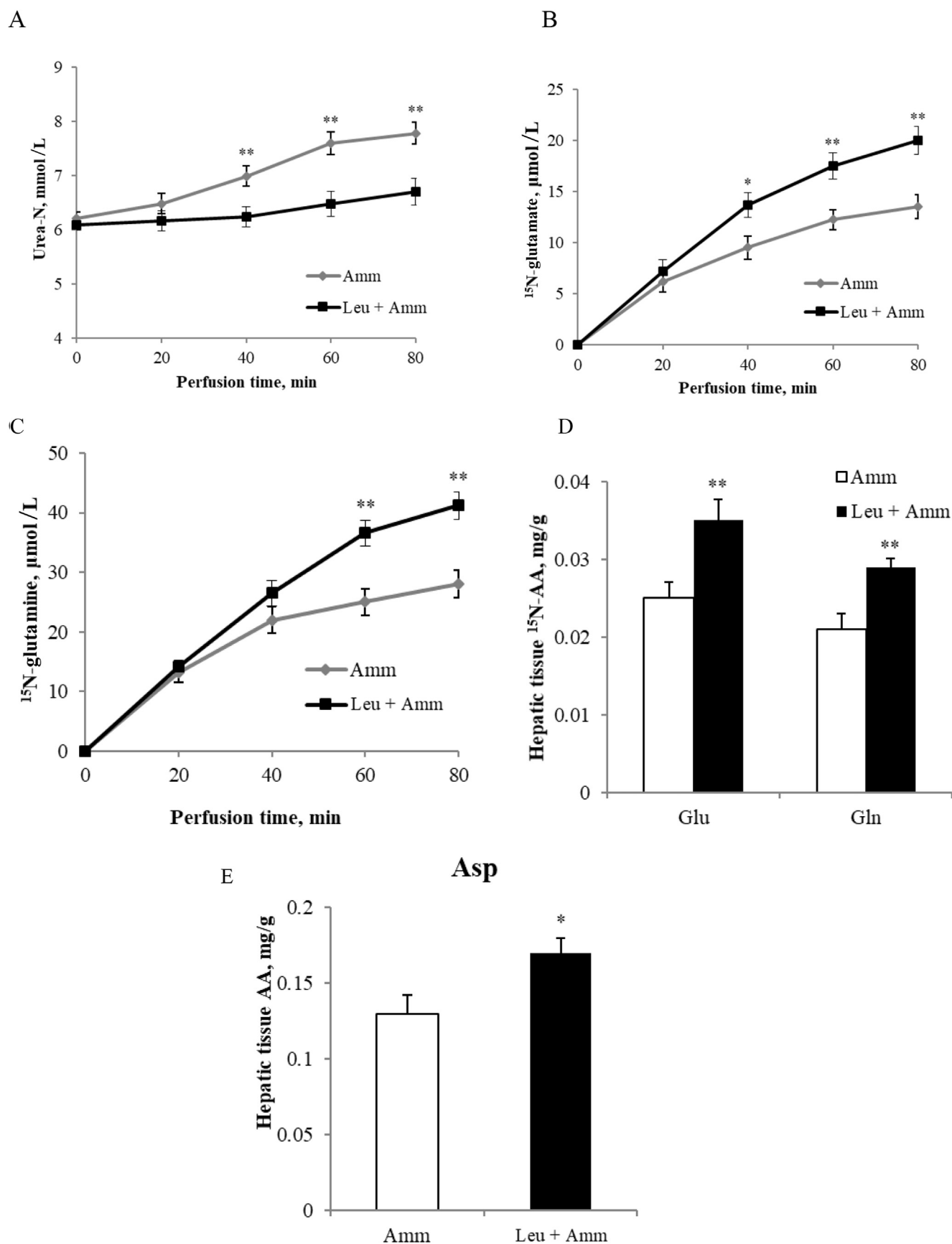


Fig. 1. L-leucine inhibits ureagenesis and stimulates glutamate synthesis from ammonium chloride (NH_4Cl). (A) hepatic vein plasma urea-N levels (mmol/L); (B) (C) livers perfused with ^{15}N -labeled ammonium chloride for 80 min, plasma ^{15}N -labeled glutamate and glutamine ($\mu\text{mol/L}$) were measured every 20 min; (D) (E) ^{15}N -labeled glutamate and glutamine ($\mu\text{mol/L}$) and the total aspartate (mg/g) in tissue samples after 80 min. Amm represents ammonium chloride. Data are expressed as the mean values and standard deviations ($n = 3$). * represents $P < 0.05$, ** represents $P < 0.01$.

metabolites showed that leucine significantly stimulated the synthesis of ^{15}N -labeled glutamate and glutamine from alanine ($P < 0.01$, Fig. 2D). In addition, there was no significant difference in aspartate between the alanine and leucine plus alanine treatments during the perfusion experiment ($P > 0.05$, Fig. 2E).

3.3. L-leucine inhibits ureagenesis and stimulates glutamate synthesis via GDH

With ammonium chloride or alanine as a substrate, the addition of leucine inhibited the output of hepatic urea and strongly stimulated glutamate synthesis in perfused livers. Notably, the GDH activity and mRNA level were significantly higher in the presence of leucine in both the ammonium chloride and alanine groups ($P < 0.01$, Fig. 3A, B, D). Leucine induced a significantly lower *SIRT4* gene expression both in mRNA and protein level ($P < 0.01$, Fig. 3A to C). Notably, addition of leucine increased the *S6K1* and *S6* phosphorylation levels, suggesting that leucine activated mTORC1 signaling ($P < 0.05$, Fig. 3C). Additionally, leucine decreased the *CPS1* mRNA level in the ammonium chloride group, but increased the glutamate-ammonia ligase (*GLUL*) mRNA level in the alanine group ($P < 0.05$, Fig. 3A and B). There were no significant changes in GDH, *CPS1* and GS protein levels of these 2 groups with or without leucine.

3.4. L-leucine increases GDH activity by regulating mTORC1/SIRT4 pathway

With either ammonium chloride or alanine as nitrogen sources in perfused livers, the addition of leucine have activated mTORC1 pathway, repressed the *SIRT4* expression and strongly increased GDH activity. Whether mTORC1/SIRT4 exerts an important function in the regulation of GDH by leucine was further studied in primary pig hepatocytes.

To induce a decrease in *SIRT4* expression, primary hepatocytes were transfected with siRNAs containing sequences specific for *SIRT4*. After transfection, *SIRT4* protein expression decreased by approximately 85% (Fig. 4A). The viability of *SIRT4* knockdown hepatocytes assessed either by lactate dehydrogenase leakage or by the activity of the mitochondrial complex II/succinate dehydrogenase (MTT assay) was unaffected. The effect of *SIRT4* knockdown on GDH activity in response to leucine was demonstrated. Leucine also decreased *SIRT4* mRNA level in control hepatocytes (Fig. 4B). Notably, leucine could markedly increase mTORC1 signaling, as evidenced by sustained phosphorylation of *S6K1* and its target *S6* both in control and lacking *SIRT4* hepatocytes ($P < 0.05$, Fig. 4C). However, the leucine-induced mTORC1 pathway activation nicely correlated with an increase in GDH activity only in control hepatocytes ($P < 0.01$, Fig. 4C and D). It is important to note that no significant change in GDH activity was observed in leucine-treated *SIRT4*-deficient hepatocytes ($P > 0.05$, Fig. 4D).

Hepatocytes were then treated with or without rapamycin followed by treatment with leucine. Rapamycin completely blocked the leucine-induced mTORC1 activation, as indicated by the unchanged phosphorylation levels of *S6K1* and *S6* (Fig. 4E). In addition, the administration of rapamycin also blocked the stimulatory effect of leucine on GDH activity and *SIRT4* gene expression (Fig. 4E and F).

4. Discussion

As the major site for ammonia metabolism, the liver is worthy of special emphasis and meticulous investigation. In contrast to

conversion of ammonia into urea, ammonia is also responsible for glutamine and other AA synthesis in liver (Peh et al., 2010). So the hepatic metabolic regulation of ammonia is of important significance in reduction of endogenous nitrogen excretion. Notably, GDH is highly expressed in mammalian liver and of central importance in the metabolism of nitrogen (Spanaki and Plaitakis, 2012). In the deamination direction, the enzyme is thought to provide ammonia and α -ketoglutarate, while in the reverse direction it catalyzes the synthesis of glutamate (McGivan et al., 1973). In addition, GDH may produce glutamate for glutamine synthesis. The major finding in this study relates to the molecular mechanism by which leucine regulates GDH in pigs liver. We determined that leucine increases GDH activity and stimulates glutamate synthesis by regulating the mTORC1/SIRT4 pathway.

Liver controls the urea cycle as well as the catabolic enzymes for most of the AA. However, one of the distinguishing features of BCAA catabolism is the relatively small fraction of the capacity that resides in the liver (Brosnan and Brosnan, 2006). In a previous paper we showed that leucine was not much degraded by pig liver (Li et al., 2015). It seems likely that leucine exerts its effect directly and not as a result of its metabolic degradation.

In the present study, the hepatic portal vein perfusion technique was introduced to study the effect of leucine on GDH activity and ammonia metabolism. The results presented that the addition of leucine inhibited the formation of urea and increased the formation of glutamate and glutamine using ammonium chloride or alanine as nitrogen sources in perfused liver. This is in agreement with previous findings in which leucine administration increased the production of glutamate and glutamine and inhibited the synthesis of urea in isolated rat liver cells (Mourão et al., 1975).

Ammonia and alanine are the important nitrogen sources in the liver. Ammonia and alanine are also the vital nitrogen sources for the study of ammonia metabolism (Soria et al., 2013). The net portal outflow of ammonia accounts for 18% of total amino acid nitrogen intake (Stoll et al., 1998). Alanine is recognized as an important hepatic nitrogen donor that is involved in gluconeogenesis and in the glucose–alanine cycle between liver and muscle (Felig et al., 1970). Interestingly in our study, leucine caused a relatively low consumption of aspartate when ammonium chloride was used as nitrogen source, but when alanine was used instead there was no significant difference in aspartate consumption. Taken together, leucine could cause an inhibition of urea synthesis and a corresponding low consumption in aspartate. This was because the formation of urea required two nitrogen atoms, one from ammonia and another from aspartate (Brosnan et al., 2001). Alanine is very effective in providing nitrogen for urea synthesis via converting to cytosolic aspartate by the combined action of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) (Brosnan et al., 2001).

In this study leucine strongly increased the activity of hepatic GDH, which is located exclusively in the mitochondrial matrix and its activity is to some extent dependent on leucine stimulation (McGivan et al., 1973). From this it is apparent that leucine potently activates GDH, whereas valine, isoleucine, and methionine activate it weakly (Yang et al., 2010). It is evident that with both nitrogen sources leucine-increased GDH activity played an essential role in the inhibition of urea formation by acting in the reverse direction catalyzing production of glutamate. Moreover, the different effects of leucine on the *CPS* and *GLUL* mRNA levels in both study groups might be caused by different substrates. Taken together, these studies clearly reveal that leucine-mediated GDH regulation is the key step controlling AA and ammonia metabolism in liver. In view

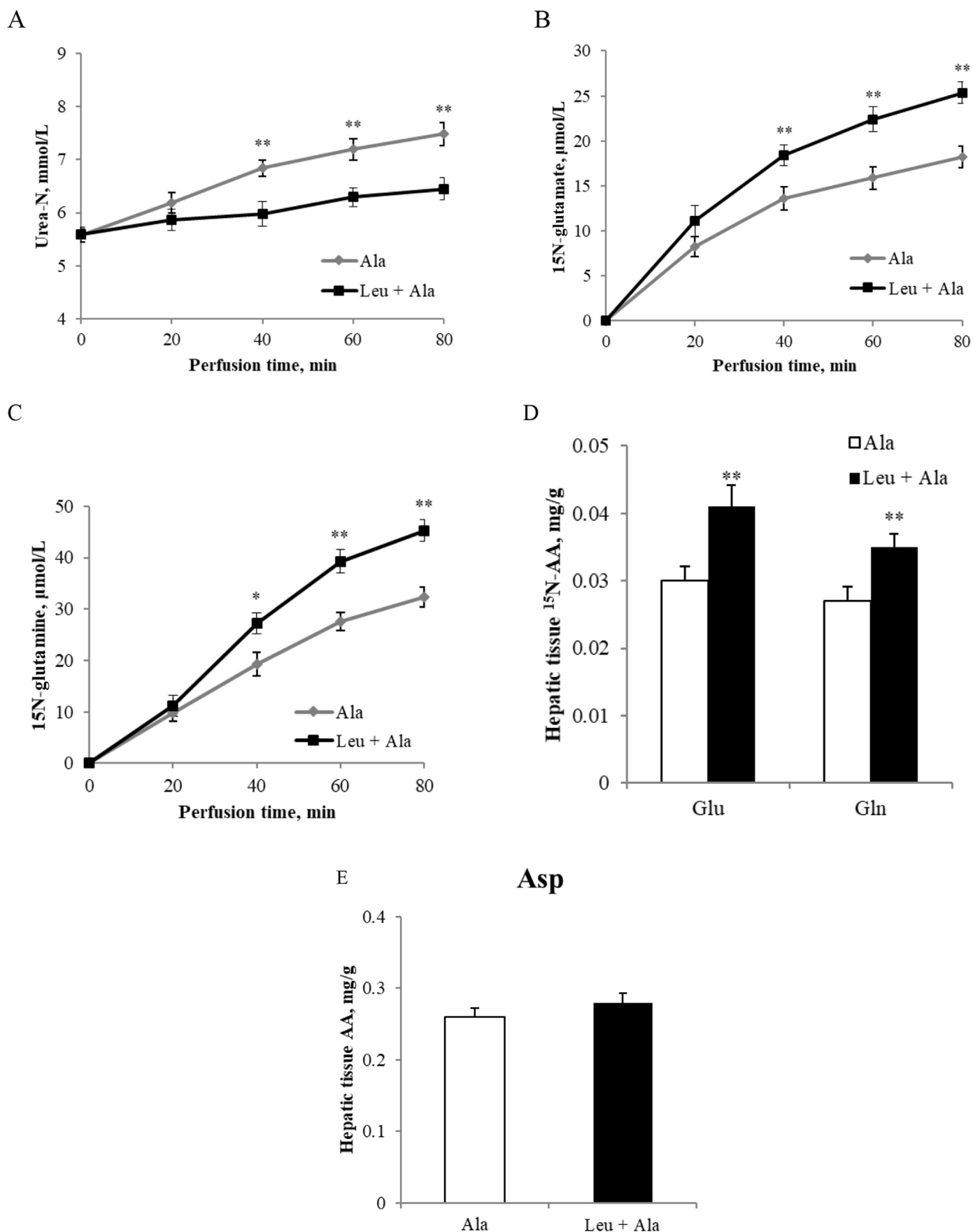


Fig. 2. L-leucine inhibits ureagenesis and stimulates glutamate synthesis from alanine. (A) plasma urea-N (mmol/L) at 0, 20, 40, 60 and 80 min; (B) (C) livers perfused with ^{15}N -labeled alanine for 80 min, plasma ^{15}N -labeled glutamate and glutamine ($\mu\text{mol/L}$) every 20 min; (D) (E) ^{15}N -labeled glutamate and glutamine ($\mu\text{mol/L}$) and total aspartate (mg/g) in pig liver tissue samples. Data are expressed as the mean values and standard deviations ($n = 3$). * represents $P < 0.05$, ** represents $P < 0.01$.

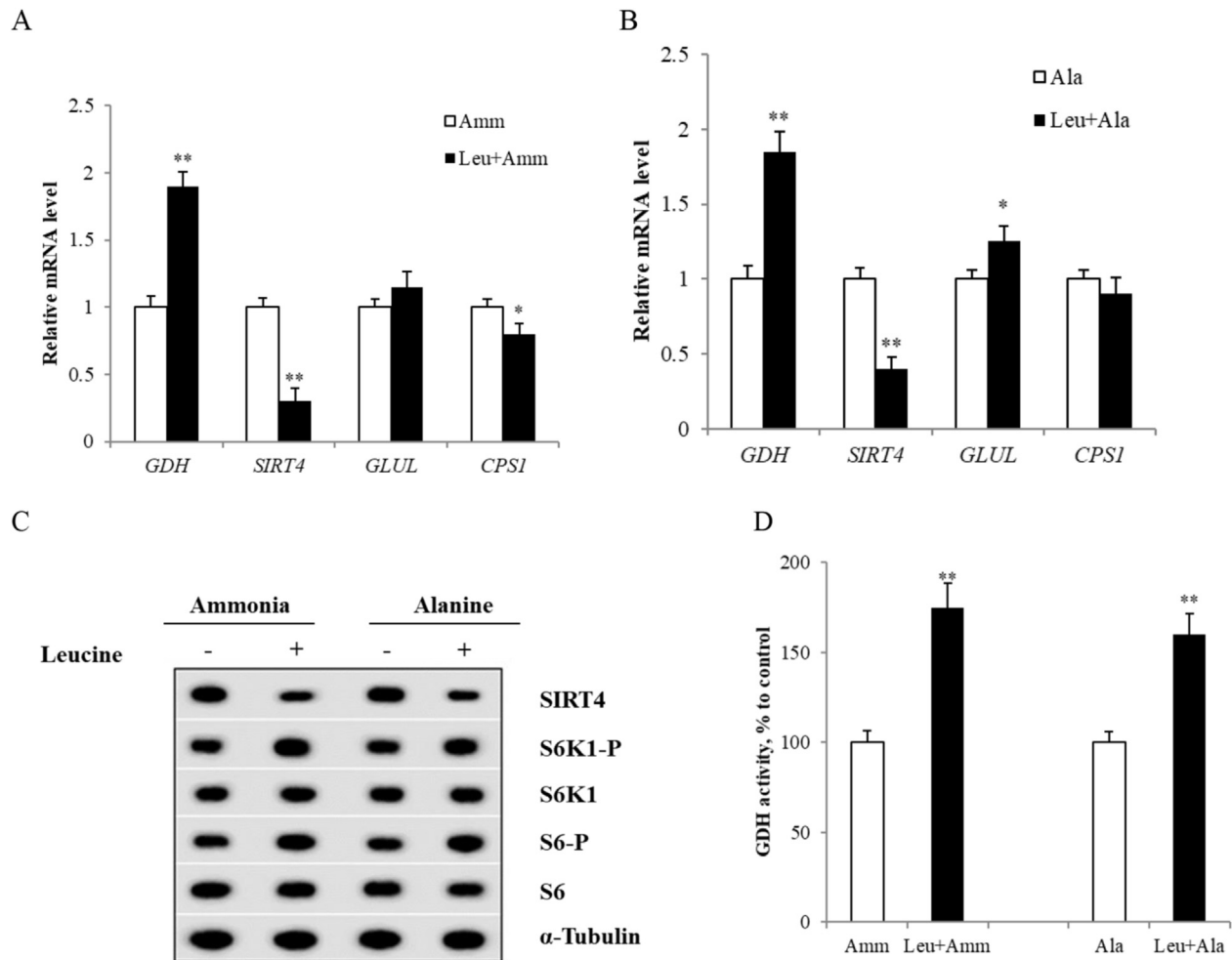


Fig. 3. L-leucine inhibits ureagenesis and stimulates glutamate synthesis via glutamate dehydrogenase (GDH). (A) (B) Glutamate dehydrogenase, sirtuin4 (*SIRT4*) and glutamate-ammonia ligase (*GLUL*) mRNA levels in tissue samples. The relative abundance of mRNA was calculated after normalization to β -actin; (C) Proteins prepared from liver tissues were analyzed by western blotting using antibodies to *SIRT4*, ribosomal protein S6 kinase 1-P (S6K1-P), ribosomal protein S6 kinase 1 (S6K1), ribosomal protein S6-P (S6-P), ribosomal protein S6 (S6) and normalized by α -Tubulin; (D) GDH activity in pig liver. Data are expressed as the mean values and standard deviations ($n = 3$). * represents $P < 0.05$, ** represents $P < 0.01$.

of its importance, the regulatory mechanism of leucine is clearly demonstrated by the results of this study.

Sirtuins are a family of NAD-dependent protein deacetylases. In mammals, there are seven sirtuins (SIRT1–7), 3 of which (SIRT3–5) are located in the mitochondrial matrix (Newman et al., 2012). During fasting SIRT3 and SIRT5 are reported to promote ureagenesis (Takashi and Leonard, 2009; Ogura et al., 2010; Hallows et al., 2011). Our previous study found PGC-1 α could promote ureagenesis in liver through SIRT3 and SIRT5 in response to glucagon (Li et al., 2016). Interestingly, it was also found that leucine inhibited *SIRT4* gene expression both in mRNA and protein levels in our study. This indicates *SIRT4* also plays key roles in metabolism. Haigis and Guarente (2006) demonstrated that *SIRT4* could act in the mitochondria to repress the activity of GDH through ADP ribosylation.

To learn whether leucine-increased GDH activity was mediated by *SIRT4*, the *SIRT4* protein was knockdown in primary pig hepatocytes. The use of cell cultures demonstrated that leucine

could promote GDH activity and repressed *SIRT4* gene expression in control hepatocytes, but no significant change in GDH activity was observed in *SIRT4*-deficient hepatocytes treated with leucine. So, it was reasonable to infer that leucine could stimulate GDH activity by repressing *SIRT4* gene expression in pig liver.

Another finding was that leucine could activate mTORC1 signaling both *in vivo* and *in vitro* experiments. Interestingly, the activation of the mTORC1 pathway nicely correlated with an increase in GDH activity and a decrease of *SIRT4* expression, suggesting that regulation of GDH by leucine might be directly associated to mTORC1 pathway. To further explore this question, hepatocytes were treated with rapamycin, an inhibitor of mTORC1. It was observed that rapamycin completely blocked the leucine-induced mTORC1 activation and the leucine-increased GDH activity. Therefore, the mTORC1 pathway might exert an important function in regulating GDH activity. Notably, the administration of leucine did not influence *SIRT4* expression in rapamycin-treated

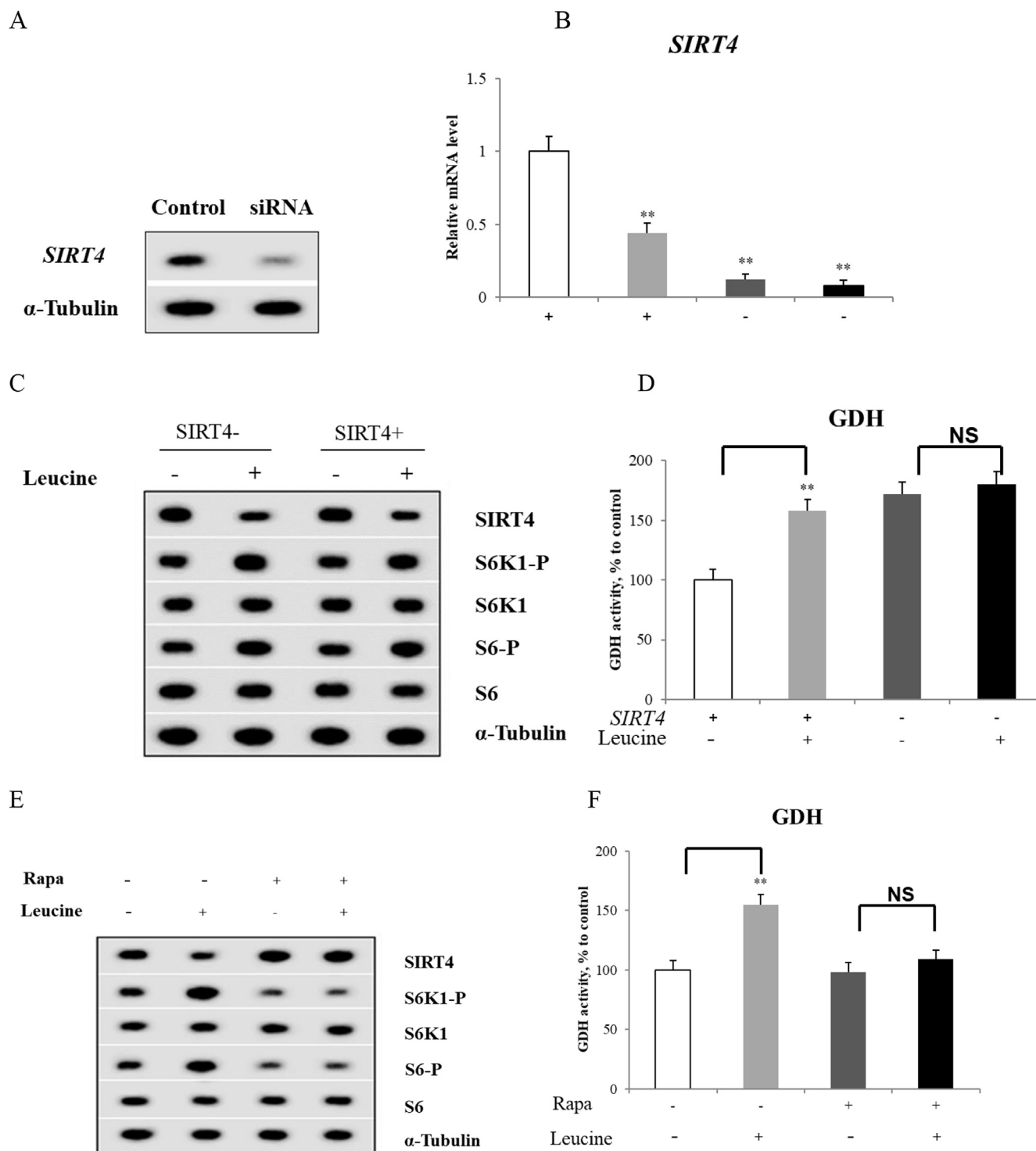


Fig. 4. L-leucine increases glutamate dehydrogenase (GDH) activity by regulating mammalian target of rapamycin complex 1 (mTORC1)/sirtuin4 (SIRT4) pathway. (A) Hepatocytes transfected with small interfering RNA (siRNA) for knockdown *SIRT4*. Protein extracts were analyzed by western blotting using anti- α -Tubulin or anti-SIRT4 antibodies; (B) *Sirtuin4* mRNA level in hepatocytes; (C) (E) Proteins prepared from hepatocytes were analyzed by western blotting using antibodies to SIRT4, ribosomal protein S6 kinase 1-P (S6K1-P), ribosomal protein S6 kinase 1 (S6K1), ribosomal protein S6-P (S6-P), ribosomal protein S6 (S6) and normalized by α -Tubulin; (D) (F) GDH activity in hepatocytes. Data are expressed as the mean values and standard deviations ($n = 3$). * represents $P < 0.05$, ** represents $P < 0.01$.

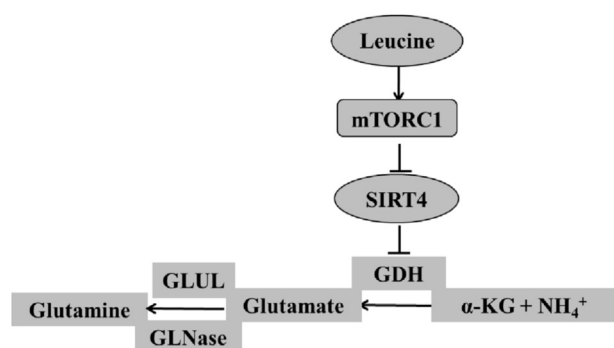


Fig. 5. Schematic of the regulation of leucine on glutamate dehydrogenase (GDH) activity and ammonia metabolism by the mammalian target of rapamycin complex 1 (mTORC1)/sirtuin4 (SIRT4) pathway. Leucine could activate mTORC1 signaling. Mammalian target of rapamycin complex 1 inhibits *SIRT4* gene expression, in turn; decrease the inhibition of GDH activity from SIRT4. Indeed, leucine increases GDH activity and stimulates glutamate synthesis from different nitrogen sources by regulating mTORC1/SIRT4 pathway. In addition, leucine promotes the reversible conversion from glutamate to glutamine. GLUL = glutamate-ammonia ligase; GLNase = Glutaminase; α -KG = α -ketoglutarate.

hepatocytes. Additionally, our findings also showed that the addition of leucine activates mTORC1 pathway, without changing the GDH activity in SIRT4-knockdown hepatocytes. The results demonstrated that SIRT4 was the downstream target in the mTORC1 pathway. This result was in agreement with that of Csibi et al. (2013) who reported that the mTORC1 pathway controls GDH activity by regulating SIRT4. Taken together, these results reveal that leucine might increase GDH activity via mTORC1/SIRT4 pathway (Fig. 5).

5. Conclusion

Overall, the results presented in this study show that addition of leucine inhibited urea output and stimulated glutamate synthesis in perfused pig livers receiving nitrogen from 2 different sources. Mechanistically, this result may be due to deactivation of the mTORC1 pathway by leucine, which in turn repressed *SIRT4* expression and strongly increased GDH activity and glutamate synthesis by regulating the mTORC1/SIRT4 pathway in pig liver.

Conflicts of interest

The authors have no financial or personal conflicts of interest to declare.

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