# **Daily Records**

# **Caccone PostDoc**

# Gus Dunn

# February, 2015

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2 2015-02-02 (Monday)	
2.1 Updating maps: current trap locations	
2.1.1 spartan dev: GPS stuff	
• testing (test_utils_maps_gps.py):	

[x] GPSCoordTree.\_grow\_branch[x] GPSCoordTree.\_get\_subtree

- [x] GPSCoordTree.mean

### 2.2 Creating Uganda Data Repo

• local location:

/home/gus/Dropbox/uganda\_data/data\_repos/field\_data

• github address: https://github.com/CacconeLabYale/field\_data.git

# 3 2015-02-03 (Tuesday)

# 3.1 Updating maps: current trap locations

- established comprehensive lists of village-ID-map and trap GPS locations for Uganda:
  - village-ID-map:
     field data/locations/names/uganda village id map.csv
  - trap GPS coords:
     field\_data/locations/gps/traps/uganda\_traps\_gps.csv

# 4 2015-02-04 (Wednesday)

### 4.1 General ToDo

- [x] email to confirm HR got my letter
- [x] meet with Gisella and Andrea [1130]
  - [X] write up notes from meeting: gisella andrea 2015-02-04.pdf
- [x] Talk to Ben E about the MAD idea.
- [x] create git repo for this paper
- [] begin development of the MAD idea
- [X] install LDna and R-studio
- [X] Located space to move the EPH *G. pallidipes* samples here to ESC with Rob

#### 4.2 ddRAD stuff

### 4.2.1 LD: detect 'outlier' SNP-pairs

#### • I propose this method:

- 1. for each distance group: collect  $r^2$  from  $\pm \sim 5$  bp distance window
  - a. across genome
  - b. across scaffold
- calculate modified z-score (based on median absolute deviation rather than standard deviation: MAD is more robust than SD for HTS-type data)
- 3. flag any SNP-pair with  $z \ge 3.5$
- 4. possibly randomize data and calculate FDR to evaluate performance.
  - a. perhaps vary the window from step 1 to use FDR to chose window that minimizes FDR.

### • Ben E's thoughts:

- basically: this is probably a waste of time and energy
  - \* other more sophisticated methods have already been applied to this data with not much significance detected
  - \* why do we expect this work to yield better/more results?

### • Gisella's thoughts:

- still should do it be we will need it when we have more data

#### 4.2.2 Install LDna

- github page: github.com/petrikemppainen/LDna
- paper reference: http://onlinelibrary.wiley.com/doi/10.1111/1755-0998. 12369/abstract
- installed devtools with RStudio gui: [successful]
- installed LDna with devtools: [successful]

```
devtools::install_github("petrikemppainen/LDna")
```

documentation: LDna/html/00Index.html

#### 4.2.3 LDna notes

• operates on:

Lower diagonal matrix of pairwise LD values,  $r^2$  is strongly recommended

• the code below should generate what I want (I think):

```
\label{lem:plink} $$ --vcf tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf $$ --allow-extra-chr $$ --r2 bin $$ --out plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bin $$ --out plink_out/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_f2_53.recode.put/tsetseFINAL_140ct2014_
```

#### 4.2.4 PLINK run for LDna

• ran the command below:

```
wd238 at compute-23-2 in ~GENOMES/glossina_fuscipes/annotations/SNPs (py278)
$ plink --vcf tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf \
> --allow-extra-chr \
> --r2 bin \
> --out plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bi
```

• waiting for it to finish: [failed]

#### 4.2.5 Louise Scratch Request Email

netid: wd238 group: caccone anticipated usage:

- ~ 100G
- < 100 files purpose of usage:
- running plink all\_v\_all linkage disequilibrium calculations on ~40K
   SNPs
- current attempt (documented below) gave a write failure which I think may be bc of some rather large tmp files generated during the process?
- Does bumping up against our space quota have hard/immediate consequences like that?

error log:

```
wd238 at compute-23-2 in ~GENOMES/glossina_fuscipes/annotations/SNPs (py278)
$ plink --vcf tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf \
> --allow-extra-chr \
> --r2 bin \
> --out plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_
PLINK v1.90b2o 64-bit (25 Nov 2014)
                                          https://www.cog-genomics.org/plink2
(C) 2005-2014 Shaun Purcell, Christopher Chang GNU General Public License v3
Logging to plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/
516842 MB RAM detected; reserving 258421 MB for main workspace.
--vcf: 73k variants complete.
plink out/tsetseFINAL 140ct2014 f2 53.recode.renamed scaffolds.maf0 05.vcf/ld/r2 bin-temp
plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bin-temp
plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bin-temp
73297 variants loaded from .bim file.
53 people (0 males, 0 females, 53 ambiguous) loaded from .fam.
Ambiguous sex IDs written to
plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bin.nose
Using up to 63 threads (change this with --threads).
Before main variant filters, 53 founders and 0 nonfounders present.
Calculating allele frequencies... done.
Total genotyping rate is 0.965098.
73297 variants and 53 people pass filters and QC.
Note: No phenotypes present.
--r2 square bin to
plink_out/tsetseFINAL_140ct2014_f2_53.recode.renamed_scaffolds.maf0_05.vcf/ld/r2_bin.ld.h
... done.
Error: File write failure.
```

#### 4.2.6 Github repo for this paper

• github page:

https://github.com/CacconeLabYale/gloria soria ddRAD 2015.git

# 5 2015-02-05 (Thursday)

### 5.1 Mariangela blacktie install

- turns out i did NOT send Mariangela install instructions for the development branch
- wrote quick install script for her to use and sent it

#### 5.2 MAD idea

- 1. for each group of SNPs x bp apart: collect  $r^2$  from  $\pm \sim 5$  bp distance window around x:
  - a. across genome
  - b. across scaffold
- 2. calculate modified z-score (based on *median absolute deviation* rather than standard deviation: **MAD** is more robust than **SD** for **HTS-type** data)
- 3. flag any SNP-pair with  $z \ge 3.5$
- 4. possibly randomize data and calculate FDR to evaluate performance.
  - a. perhaps vary the window-size from step 1 to use FDR to chose window-size that minimizes FDR.

### 5.2.1 Development

• ipython notebook: ddrad58/2015-02-05 MAD idea.ipynb

### 5.3 G. pallidipes

- Rob brought most to ESC this morning
- doesn't expect to need my truck for the rest

# 6 2015-02-06 (Friday)

### 6.1 MAD idea

#### 6.1.1 Development

- LOTS of progress at ipython notebook: ddrad58/2015-02-05 MAD idea.ipynb
- See notes about plotting median and MAD with bootstrapped CIs near the bottom of above (commit dd7fe5da5733406edeaab6ce3c25b523b94552f2)

# 7 2015-02-09 (Monday)

#### Goals:

- [x] Zimmer Workshop
- [x] Start Professional Development notebook
- [x] Find out how to process health reimbursement
  - [ ] Get them ready for mailing
    - \* [x] form
    - \* [] receipts
  - [X] Assemble list of information I need from Sarah and send it to her
- [] Progress on MAD idea
- [ ] Generate strategy for the week
- [ ] Sketch out abstract for Keystone? meeting
- [] find out if there is data available on tsetse control by area in Uganda
  - chemicals sold
  - etc

#### 7.1 Health reimbursement

- http://yalehealth.yale.edu/claims
- Supplemental Claim form: http://yalehealth.yale.edu/sites/default/files/supplemental\_claims\_form.pdf
- pharmacy claim form: http://yalehealth.yale.edu/sites/default/files/pharmacy\_claim\_form\_restat\_catamaran.pdf

#### 7.1.1 Instructions for pharmacy process

#### • from website above

Include copies of prescription receipts showing the following information:

- Pharmacy Name, Address & Phone Number
- Patient Name
- Prescription Number
- Prescription Fill Date
- Drug Name, Strength and NDC Code
- Drug Quantity & Days supply
- Drug Cost
- Amount Paid

Please mail the Prescription Drug Claim Form and receipts to:

Restat

Patient Reimbursement 11900 W. Lake Park Drive Milwaukee, WI 53224

Claims are honored for one year from the date of service. If you haven't received a response to a claim within 60 days of filing, contact the Claims Department. You may call sooner to inquire if the claim has been received and is in process.

# 8 2015-02-10 (Tuesday)

### Goals:

- [ ] Get pharm claim ready for mailing
  - -[x] form
  - [] receipts
- [] Progress on MAD idea
- [ ] Generate strategy for the week
- [ ] Sketch out abstract for Keystone? meeting
- [ ] find out if there is data available on tsetse control by area in Uganda
  - chemicals sold

- etc
- [ ] figure out how to download zimmer files

### 8.1 Health reimbursement

• printed form

# 8.2 Met with Postdoc applicant (Christina)

• had lunch

# 9 2015-02-12 (Thursday)

# 9.1 Health reimbursement

• Need Catherine's member ID

### 9.2 MAD idea

### 9.2.1 Development

- yesterday:
  - bootstrap confidence intervals are functional
  - modified z-score is functional
  - used ggplot to provide nice figure showing rough progression of z-scored  $r^2$  through distance between snps

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# 10 2015-02-13 (Friday)

# 10.1 G. pallidipes Sample catalog

## 10.1.1 Summary table

- data types:
  - location
  - symbols when present (*I assume you mean location symbol?*)
  - number of individuals
  - date range
  - is tissue?
  - is extraction?
  - analysis status
- will be done in python for increased flexibility by [Gus]
- notebook file: 2015-02-12 sample catalog summary.ipynb
- Showed output to Gisella and she signed off on it after asking whether I could accommodate GEO COORDS when we get them.
- STATUS: [completed]

#### 10.1.2 Primers etc

- RobH reports that he and KirstinD found many primers etc that were either designed for *G. pallidipes* or shown to work with it in the past.
- testing on the primers will begin next week.

# 10.1.3 Leg extractions

- Rob did Xymogen extractions on 5 legs
- NanoDrop indicates absorption at 260 but peaks look weird
  - probably bc the kit leaves EVERYTHING still in solution
  - [ ] RobH will check with KirstinD about her extraction traces on G.
     f. fuscipes legs

# 10.2 MAD idea

## 10.2.1 Development

- [completed]: functions to
  - update df with distance\_bin and mad\_z
  - plot mad z by bins
- [to do]:
  - implement printing/saving snp-pairs that pass the z-filter

# 11 2015-02-14 (Saturday)

### 11.1 MAD idea

### 11.1.1 Development

• implement printing/saving snp-pairs that pass the z-filter

# 12 2015-02-16 (Monday)

### 12.1 G. f. fuscipes: infection summaries

- ipython to get pivot table for infected flies
  - file: 2015-02-16 g f fuscipes pandas import.ipynb
    - \* file of dumped pandas table of collection records for 2014 in hdf5 format:
- add PCR detected fly statuses to main DB

### 12.2 *G. pallidipes*: MicroSat extraction pilot

- RobH spoke with KirstinD about strange NanoDrop traces:
  - KirstinD: hers looked the same, just used 260/280 values as presented
  - likely explanation is that the extraction kit is EXTREMELY dirty by design so the spec peaks are shifted around
- RobH is beginning PCRs with ITS primers (same that KirstinD is using on the *G. f. fuscipes*) today.
- RobH is researching location names on the SerapA tubes (n ~ 6) bc
   GisellaC is not convinced the sheet SerapA included makes since.
  - RobH will google first
  - GusD will get GIS admin layers to search if google fails
- v0.2.1.2-1.tar.gz

# 13 2015-02-17 (Tuesday)

# 13.1 meeting

- escarpment Nguruman:
  - GisellaC try to get samples from extremes and in the middle
- [ ] GusD send most recent version of protocol to BrianW

# 14 2015-02-18 (Wednesday)

### 14.1 *G. pallidipes* status update meeting

- GusD
- RobH
- KirstinD
- extractions not working for a while with KirstinD
- trouble shooting

KirstinD moving forward with extractions now

# 15 2015-02-21 (Saturday)

### **GOALS:**

- [worked on] G. f. fuscipes infection summaries/maps for GisellaC meeting
- [no work] script for MariangelaB
- [small work]  $r^2$  per bin model

# 15.1 *G. f. fuscipes*: infection summaries

### 15.1.1 Converting dates to YYYY-MM-DD

- 2014\_spring\_summer\_from\_rob.xlsx
  - added new function to TsetseCheckout:
     TsetseCheckout/data/utils.py:convert brit dates to yyyy mm dd(string)
  - added new cell magic to ipython to send variable to clipboard: clip\_magic.py
  - used new function and the cell magic to copy, change, then paste back into spreadsheet.
- 2014 fall for pandas.xlsx
  - dates already fine

### 15.1.2 Adding Village names to the spring/summer excel file

- [COMPLETED]: 2015-02-22
- created python hack to use the summary sheet info to generate the Village rows

```
YalePostDoc/project stuff/g f fucipes uganda/collection data/traps to villages.py
```

summary sheets:2014 full surveyreport 20140820/summary survey data.xlsx

#### 15.1.3 ALERT: errors detected in fly name code combinations

- during this process i detected instances where the fly number code combinations (example: OLW-14 038) were **NOT** correct!
- the following IDs illustrate this:
  - 0L0-14 033 is Olobo
  - 0L0-14 034 is Olobo
  - OLW-14 035 is Olwi
  - OLW-14 036 is Olwi
  - OLW-14 037 is Olobo
  - OLW-14 038 is Olobo
- additionally, the Dissection Data-Kole sheet has **ALL** fly IDs starting KO regardless of the source village.
- RECOMEND NOT DEPENDING ON FLY ID FOR VILLAGE SOURCE!

# 16 2015-02-22 (Sunday)

### **GOALS:**

- [worked on] G. f. fuscipes infection summaries/maps for GisellaC meeting
- [none] script for MariangelaB
- ullet [none]  $r^2$  per bin model

### 16.1 *G. f. fuscipes*: infection summaries

### 16.1.1 HDF5 import and data cleaning

- standardized the spreadsheet column titles by hand to allow import and correct dataframe referencing
- file: 2015-02-16 g f fuscipes pandas import.ipynb
- recode\_villages(df, map\_func=map\_func):
  - renaming villages to letter codes

- [degenerate names discovered] and accommodated in uganda\_village\_id\_map.csv by mapping the letter code to more than one long form:
  - \* AKAYODEBE vs AKAYO-DEBE
- corrected misspellings of
  - \* "Orubakulemi" from "Orubakulem"
  - \* "JIAKO" from "JAIKO"
- recode\_positives(df):
  - recode prob, midgut, sal.gland as 0 or 1.
  - [NOTE] this will change to a trivalent state (class Tristate) soon
- recode\_tenerals(df)
  - implemented but needs conversion to Tristate
- recode\_dead(df)
  - implemented but needs conversion to Tristate
- add\_infection\_state\_col(df)
  - implemented but failing to actually alter the dataframe
- spartan.utils.misc.Tristate
  - implements three state logic that mostly supports normal boolean arithmetic (just ignoring the None state)

# 17 2015-02-23 (Monday)

#### **GOALS:**

- [] G. f. fuscipes infection summaries/maps for GisellaC meeting
- [] script for MariangelaB
- []  $r^2$  per bin model

### 17.1 *G. f. fuscipes*: infection summaries

### 17.1.1 HDF5 import and data cleaning

• spartan.utils.misc.Tristate

- I found an existing "Tribool" class on github and forked it: https://github.com/xguse/python tribool
- it did not support boolean arithmetic but was much more sophisticated in all other ways.
- I added support for boolean addition but will also add \*, -, and / before writing the tests and submitting a pull request to upstream.
- I am now using Tribool instead of Tristate
- running into serious hashable issues df.midgut.unique() throws \_\_nonezero\_\_'s ValueError.
  - possible solutions:
    - \* override \_\_new\_\_ might allow me to mimic the "always the same mem address" behavior of True etc?
    - \* Look into implementation of Factories in Python
    - \* perhaps a hint in behavior/class code for np.NaN?
    - \* [best bet] use enum class
- looking for more fertile ground to cover while I think