

Assignment 3: Data Exploration

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Fall 2025

OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Canvas.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Load necessary packages (tidyverse, lubridate, here), check your current working directory and upload two datasets: the ECOTOX neonicotinoid dataset (`ECOTOX_Neonicotinoids_Insects_raw.csv`) and the Niwot Ridge NEON dataset for litter and woody debris (`NEON_NIWO_Litter_massdata_2018-08_raw.csv`). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
#load necessary packages
library(tidyverse)
library(lubridate)
library(here)
#check current working directory
getwd()
```

```
## [1] "/home/guest/R/EDE_Fall2025"
```

```
#output is "/home/guest/R/EDE_Fall2025"
here()
```

```
## [1] "/home/guest/R/EDE_Fall2025"
```

```
#import data
Neonics <- read.csv(
  file = here("./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv"),
  stringsAsFactors = TRUE
)
NeonicsRAW <- Neonics

Litter <- read.csv(
  file = here("./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv"),
  stringsAsFactors = TRUE
)
LitterRAW <- Litter
```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Since these neonicotinoids are insecticides that means that they are used to kill insects from damaging agricultural crops. To understand which neonicotinoids impact which insects and how, it would be useful to understand the ecotoxicology mechanisms involved.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Studying the litter and woody debris helps you understand the health and functioning of these ecosystems. They can give insights to processes like nutrient cycling, carbon storage, and habitat creation.

4. How is litter and woody debris sampled as part of the NEON network? Read the `NEON_Litterfall_UserGuide.pdf` document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. In spatial sampling design, sampling is executed at terrestrial sites that contain woody vegetation greater than 2m tall. 2. In temporal sampling design, ground traps are sampled once per year. 3. Material definitions: "litterfall" includes material dropped from the canopy, butt-end diameter < 2 cm and length < 50 cm; "fine woody debris" is similar diameter (< 2 cm) but length > 50 cm.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics)
```

```
## [1] 4623 30
```

```
#dimensions are 4623 rows by 30 columns
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest? [Tip: The `sort()` command is useful for listing the values in order of magnitude...]

```
NEON_summary <- summary(as.factor(Neonics$Effect))
NEON_summary_sorted <- sort(NEON_summary)
NEON_summary_sorted
```

```
##      Hormone(s)      Histology      Physiology      Cell(s)
##           1           5           7           9
##      Biochemistry      Accumulation      Intoxication      Immunological
##           11          12          12          16
##      Morphology      Growth      Enzyme(s)      Genetics
##           22          38          62          82
##      Avoidance      Development      Reproduction      Feeding behavior
##          102          136          197          255
##      Behavior      Mortality      Population
##          360          1493          1803
```

Answer: Most common effects are population, mortality, and behavior, population and mortality, the highest two, can show how effective the pesticides are at killing the pests, while looking at behavior can maybe lead to bugs being inactive to not damage crops or different behaviors like that

7. Using the `summary` function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: Explore the help on the `summary()` function, in particular the `maxsum` argument...]

```
NEON_commonname <- summary(as.factor(Neonics$Species.Common.Name))
NEON_commonname_sorted <- sort(NEON_commonname)
NEON_commonname_sorted
```

```
##      Ant Family      Apple Maggot
##           9           9
##      Glasshouse Potato Wasp      Lacewing
##          10          10
##      Southern House Mosquito      Two Spotted Lady Beetle
##          10          10
##      Spotless Ladybird Beetle      Braconid Parasitoid
```

##	11	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Armoured Scale Family	Diamondback Moth
##	13	13
##	Eulophid Wasp	Monarch Butterfly
##	13	13
##	Predatory Bug	Yellow Fever Mosquito
##	13	13
##	Corn Earworm	Green Peach Aphid
##	14	14
##	House Fly	Ox Beetle
##	14	14
##	Red Scale Parasite	Spined Soldier Bug
##	14	14
##	Western Flower Thrips	Hemlock Woolly Adelgid Lady Beetle
##	15	16
##	Hemlock Woolly Adelgid	Mite
##	16	16
##	Onion Thrip	Araneoid Spider Order
##	16	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17
##	Oystershell Scale Parasitoid	Black-spotted Lady Beetle
##	17	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Codling Moth	Flatheaded Appletree Borer
##	19	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Argentine Ant	Beetle
##	21	21
##	Mason Bee	Mosquito
##	22	22
##	Citrus Leafminer	Ladybird Beetle
##	23	23
##	Spider/Mite Class	Tobacco Flea Beetle
##	24	24
##	Chalcid Wasp	Convergent Lady Beetle

##	25	25
##	Stingless Bee	Ground Beetle Family
##	25	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ladybird Beetle Family
##	29	30
##	Parasitoid	Braconid Wasp
##	30	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Sweetpotato Whitefly	Aphid Family
##	37	38
##	Cabbage Looper	Buff-tailed Bumblebee
##	38	39
##	True Bug Order	Sevenspotted Lady Beetle
##	45	46
##	Beetle Order	Snout Beetle Family, Weevil
##	47	47
##	Erythrina Gall Wasp	Parasitoid Wasp
##	49	51
##	Colorado Potato Beetle	Parastic Wasp
##	57	58
##	Asian Citrus Psyllid	Minute Pirate Bug
##	60	62
##	European Dark Bee	Wireworm
##	66	69
##	Euonymus Scale	Asian Lady Beetle
##	75	76
##	Japanese Beetle	Italian Honeybee
##	94	113
##	Bumble Bee	Carniolan Honey Bee
##	140	152
##	Buff Tailed Bumblebee	Parasitic Wasp
##	183	285
##	Honey Bee	(Other)
##	667	670

Answer: From highest to lowest 1.Other 2.Honey Bee 3.Parasitic Wasp 4.Buff Tailed Bumblebee 5.Carniolan Honey Bee 6.Bumble Bee Most of these species are types of bees. Bees are pollinators, and thus an important part of the ecosystem.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the dataset, and why is it not numeric? [Tip: Viewing the dataframe may be helpful...]

```
class(Neonics$Conc.1..Author.)
```

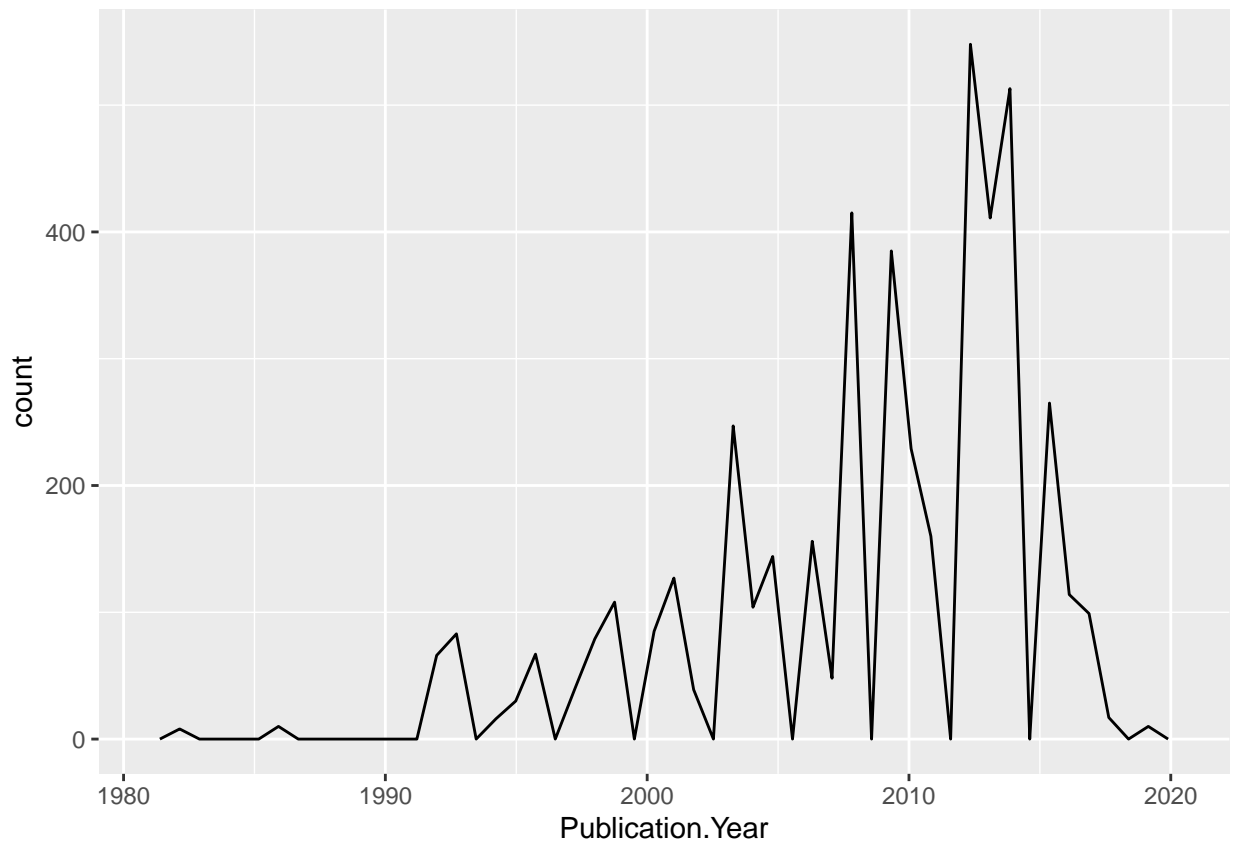
```
## [1] "factor"
```

Answer: This column is a factor, this is because while the column does include numbers, it also includes non number characters like letters, ~, /

Explore your data graphically (Neonics)

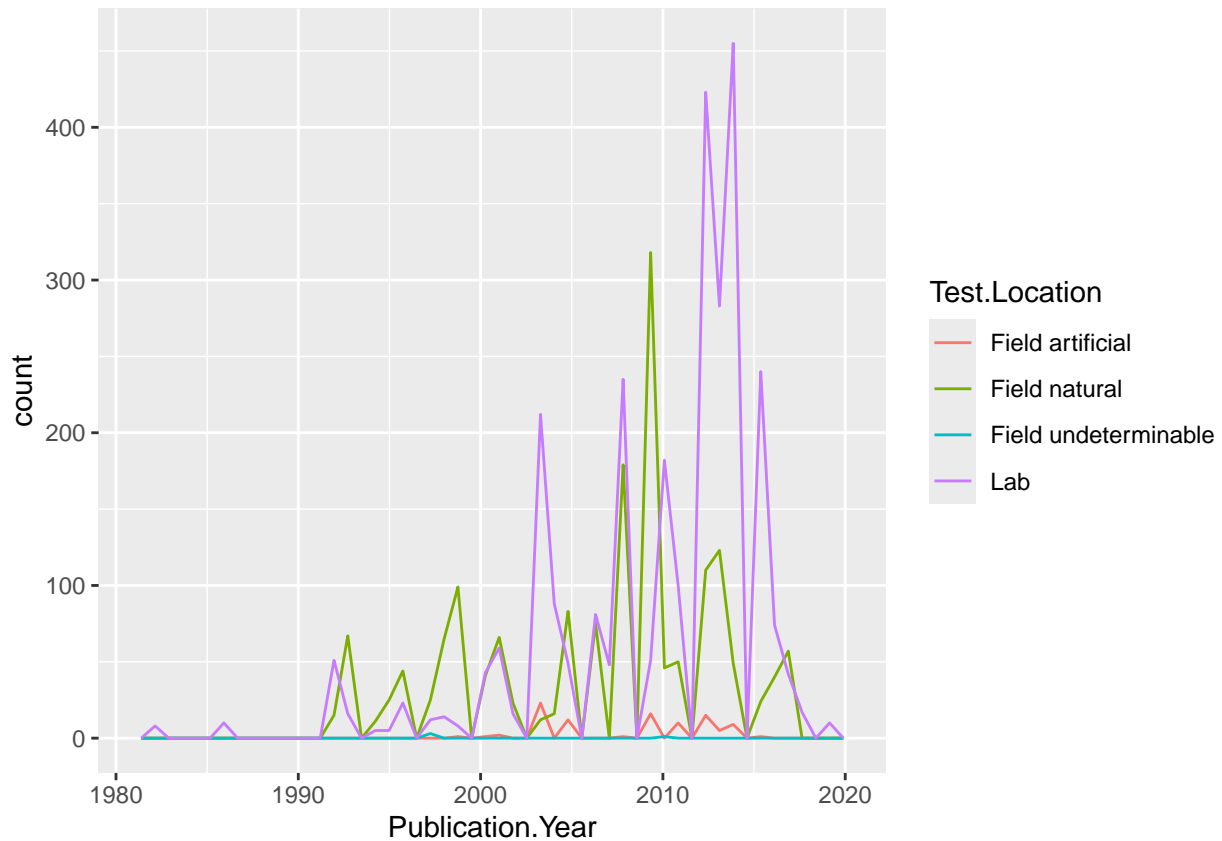
9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year), bins = 50 )
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
Pub_test_fig <- ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 50 )  
Pub_test_fig
```



```
# Plotting the frequency polygon
```

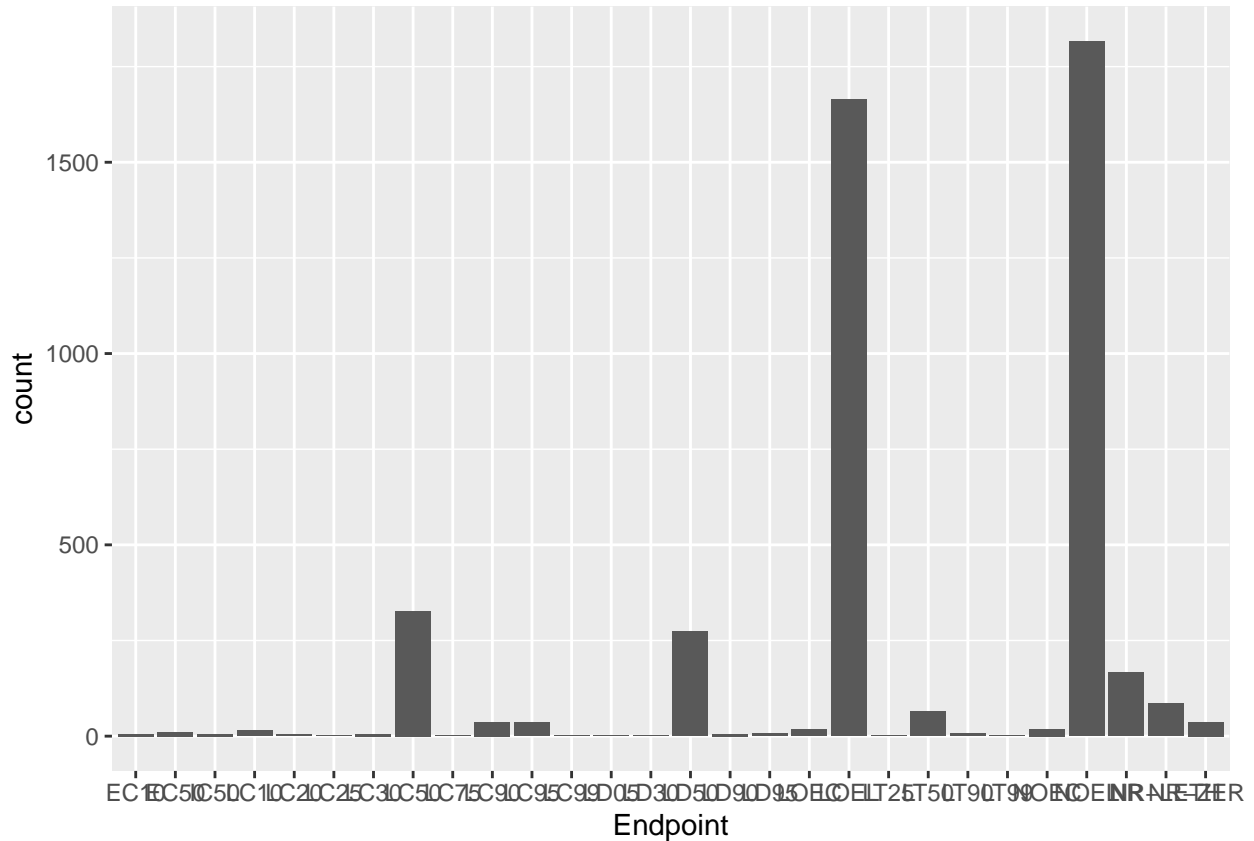
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test location is the lab, but they do differ over time with sometimes the field natural being the greatest, like just before 2010.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics) +  
  geom_bar(aes(x = Endpoint))
```



Answer: NOEL and LOEL are the two most common end points. LOEL is the Lowest Observed Effect Level - the lowest concentration where there is a statistically adverse effect. NOEL is the no observed effect level - the highest concentration where there is no statistically observed effect.

Explore your data (Litter)

12. Determine the class of `collectDate`. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)
```

```
## [1] "factor"
```

#answer is factor, not date

```
Litter$collectDate <- as.Date(Litter$collectDate, format = "%Y-%m-%d")
```

```
class(Litter$collectDate)
```

```
## [1] "Date"
```

13. Using the `unique` function, list the different plotIDs sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

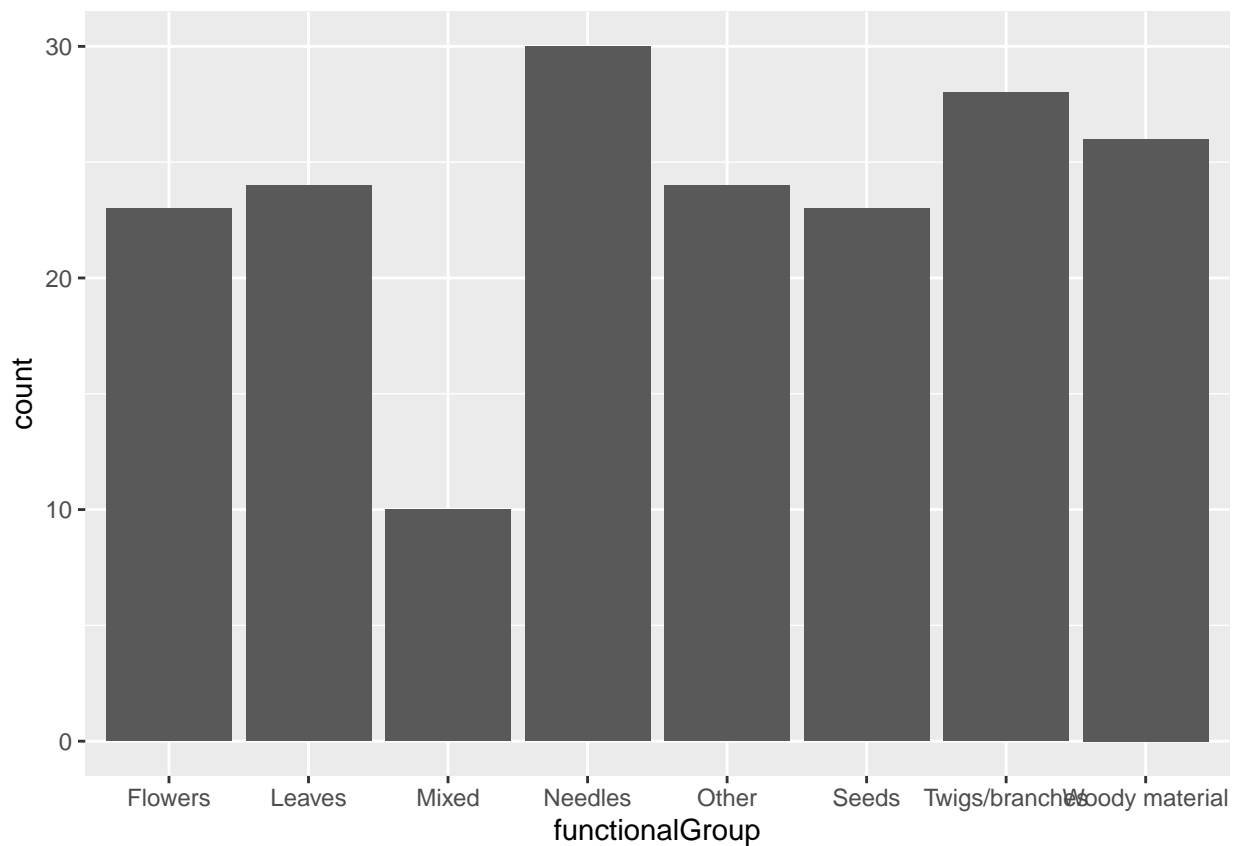

```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

Answer: 12 levels - NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051 NIWO_058 NIWO_046 NIWO_062 NIWO_057 summary will tell you the count of each level, while unique just shows how many different types there are.

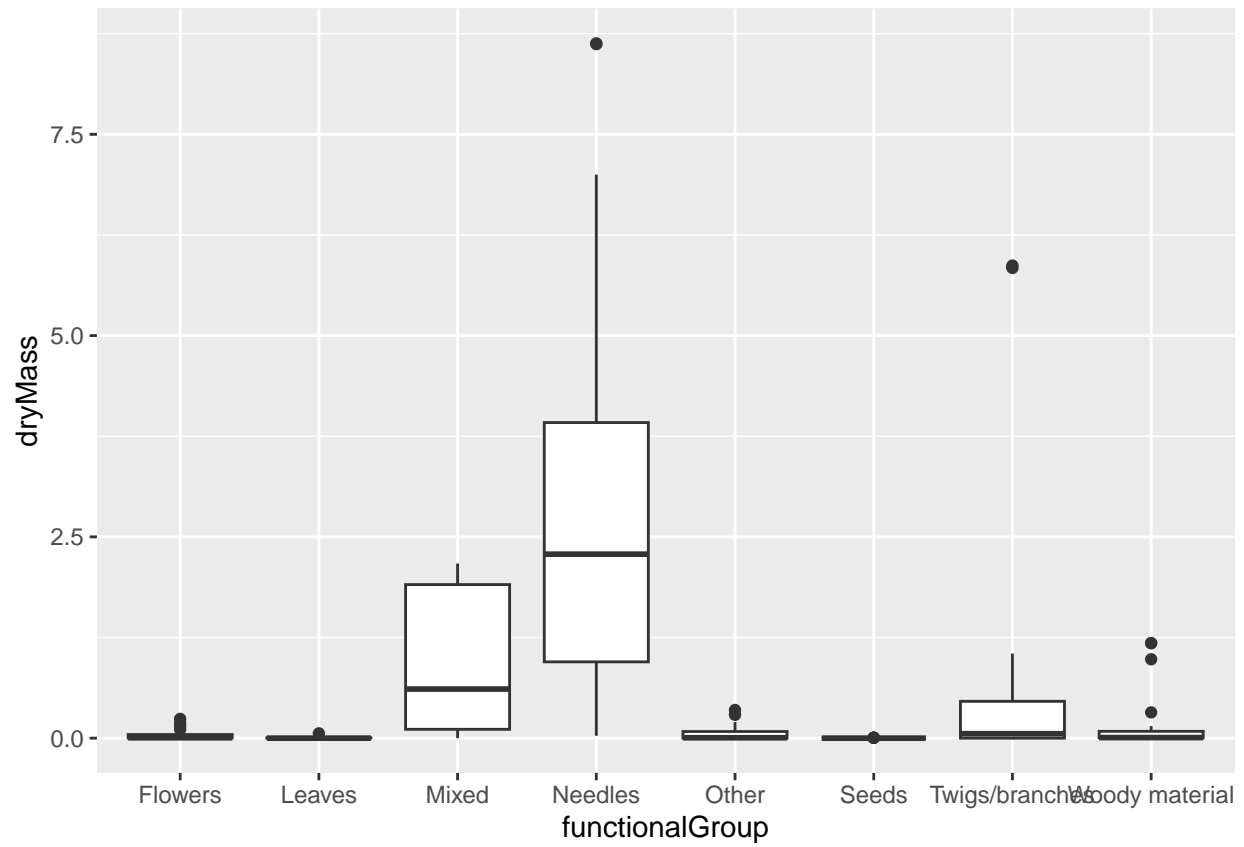
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter) +
  geom_bar(aes(x = functionalGroup))
```

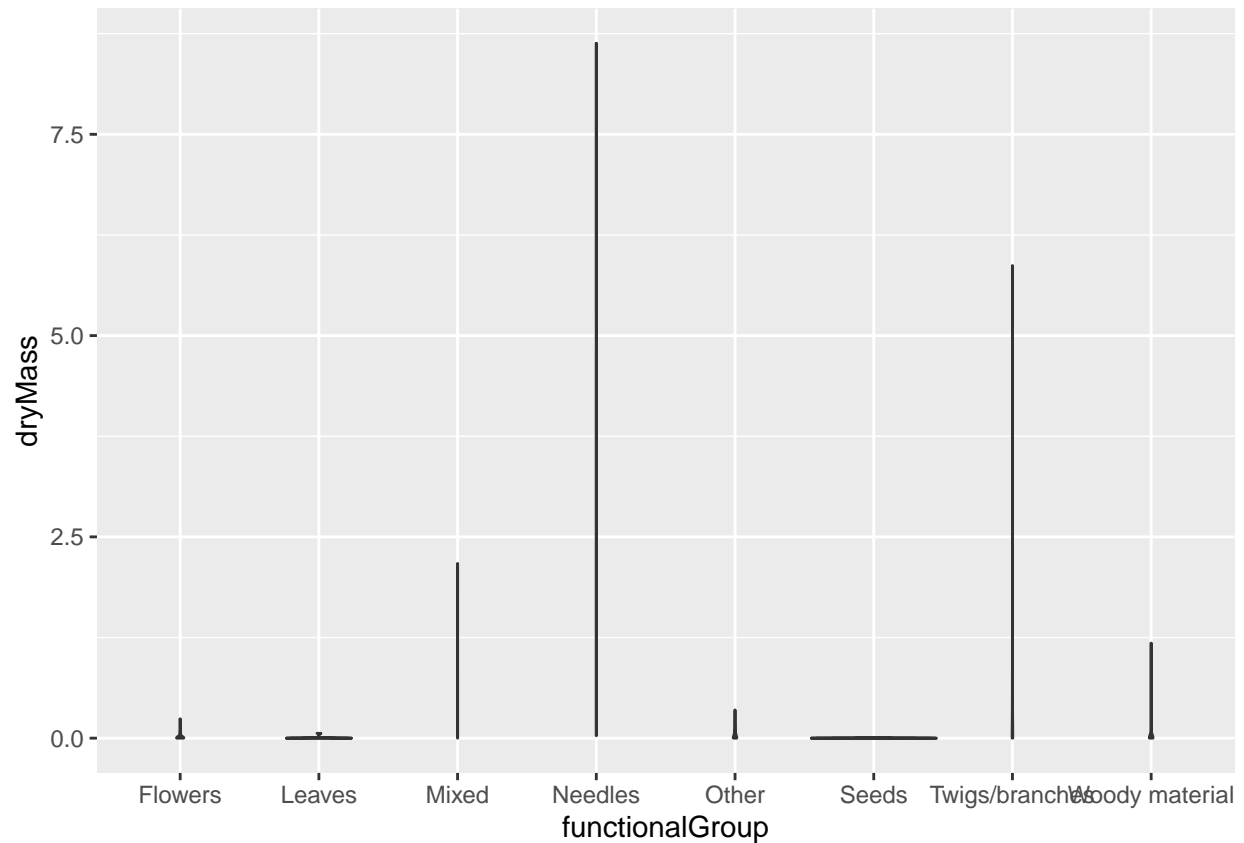


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter) +
  geom_boxplot(aes(x = functionalGroup, y = dryMass))
```



```
ggplot(Litter) +
  geom_violin(aes(x = functionalGroup, y = dryMass),
    draw_quantiles = c(0.25, 0.5, 0.75))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: In this case, the violin plots are not really showing violin shape or wide density (where the data is more concentrated), so the visual is lost and you are left with thin vertical lines which do not portray your data to viewers as nicely as the box plot does.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles has the highest, then mixed, then twigs/branches.