

Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file <FirstLast>_A03_DataExploration.Rmd (replacing <FirstLast> with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Canvas.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Load necessary packages (tidyverse, lubridate, here), check your current working directory and upload two datasets: the ECOTOX neonicotinoid dataset (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON dataset for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these datasets “Neonics” and “Litter”, respectively. Be sure to include the sub-command to read strings in as factors.

```
#load necessary packages
library(tidyverse)
library(lubridate)
library(here)
#check current working directory
getwd()

## [1] "/home/guest/R/EDE_Fall2025"
```

```

#output is "/home/guest/R/EDE_Fall2025"
here()

## [1] "/home/guest/R/EDE_Fall2025"

#import data
Neonics <- read.csv(
  file = here("./Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv"),
  stringsAsFactors = TRUE
)
NeonicsRAW <- Neonics

Litter <- read.csv(
  file = here("./Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv"),
  stringsAsFactors = TRUE
)
LitterRAW <- Litter

```

Learn about your system

2. The neonicotinoid dataset was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The dataset that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Since these neonicotinoids are insecticides that means that they are used to kill insects from damaging agricultural crops. To understand which neonicotinoids impact which insects and how, it would be useful to understand the ecotoxicology mechanisms involved.

3. The Niwot Ridge litter and woody debris dataset was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Studying the litter and woody debris helps you understand the health and functioning of these ecosystems. They can give insights to processes like nutrient cycling, carbon storage, and habitat creation.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON_Litterfall_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. In spatial sampling design, sampling is executed at terrestrial sites that contain woody vegetation greater than 2m tall. 2. In temporal sampling design, ground traps are sampled once per year. 3. Material definitions: "litterfall" includes material dropped from the canopy, butt-end diameter < 2 cm and length < 50 cm; "fine woody debris" is similar diameter (< 2 cm) but length > 50 cm.

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the dataset?

```
dim(Neonics)
```

```
## [1] 4623 30
```

#dimensions are 4623 rows by 30 columns

6. Using the **summary** function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest? [Tip: The **sort()** command is useful for listing the values in order of magnitude...]

```
NEON_summary <- summary(as.factor(Neonics$Effect))
NEON_summary_sorted <- sort(NEON_summary)
NEON_summary_sorted
```

	Hormone(s)	Histology	Physiology	Cell(s)
##	1	5	7	9
##	Biochemistry	Accumulation	Intoxication	Immunological
##	11	12	12	16
##	Morphology	Growth	Enzyme(s)	Genetics
##	22	38	62	82
##	Avoidance	Development	Reproduction	Feeding behavior
##	102	136	197	255
##	Behavior	Mortality	Population	
##	360	1493	1803	

Answer: Most common effects are population, mortality, and behavior, population and mortality, the highest two, can show how effective the pesticides are at killing the pests, while looking at behavior can maybe lead to bugs being inactive to not damage crops or different behaviors like that

7. Using the **summary** function, determine the six most commonly studied species in the dataset (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed.[TIP: Explore the help on the **summary()** function, in particular the **maxsum** argument...]

```
NEON_commonname <- summary(as.factor(Neonics$Species.Common.Name))
NEON_commonname_sorted <- sort(NEON_commonname)
NEON_commonname_sorted
```

	Ant Family	Apple Maggot
##	9	9
##	Glasshouse Potato Wasp	Lacewing
##	10	10
##	Southern House Mosquito	Two Spotted Lady Beetle
##	10	10
##	Spotless Ladybird Beetle	Braconid Parasitoid

##		11		12
##	Common Thrip		Eastern Subterranean Termite	
##		12		12
##	Jassid		Mite Order	
##		12		12
##	Pea Aphid		Pond Wolf Spider	
##		12		12
##	Armoured Scale Family		Diamondback Moth	
##		13		13
##	Eulophid Wasp		Monarch Butterfly	
##		13		13
##	Predatory Bug		Yellow Fever Mosquito	
##		13		13
##	Corn Earworm		Green Peach Aphid	
##		14		14
##	House Fly		Ox Beetle	
##		14		14
##	Red Scale Parasite		Spined Soldier Bug	
##		14		14
##	Western Flower Thrips	Hemlock Woolly Adelgid	Lady Beetle	
##		15		16
##	Hemlock Wooly Adelgid		Mite	
##		16		16
##	Onion Thrip		Araneoid Spider Order	
##		16		17
##	Bee Order		Egg Parasitoid	
##		17		17
##	Insect Class		Moth And Butterfly Order	
##		17		17
##	Oystershell Scale Parasitoid		Black-spotted Lady Beetle	
##		17		18
##	Calico Scale		Fairyfly Parasitoid	
##		18		18
##	Lady Beetle		Minute Parasitic Wasps	
##		18		18
##	Mirid Bug		Mulberry Pyralid	
##		18		18
##	Silkworm		Vedalia Beetle	
##		18		18
##	Codling Moth		Flatheaded Appletree Borer	
##		19		20
##	Horned Oak Gall Wasp		Leaf Beetle Family	
##		20		20
##	Potato Leafhopper		Tooth-necked Fungus Beetle	
##		20		20
##	Argentine Ant		Beetle	
##		21		21
##	Mason Bee		Mosquito	
##		22		22
##	Citrus Leafminer		Ladybird Beetle	
##		23		23
##	Spider/Mite Class		Tobacco Flea Beetle	
##		24		24
##	Chalcid Wasp		Convergent Lady Beetle	

##		25		25
##	Stingless Bee		Ground Beetle Family	
##		25		27
##	Rove Beetle Family		Tobacco Aphid	
##		27		27
##	Scarab Beetle		Spring Tiphia	
##		29		29
##	Thrip Order		Ladybird Beetle Family	
##		29		30
##	Parasitoid		Braconid Wasp	
##		30		33
##	Cotton Aphid		Predatory Mite	
##		33		33
##	Sweetpotato Whitefly		Aphid Family	
##		37		38
##	Cabbage Looper		Buff-tailed Bumblebee	
##		38		39
##	True Bug Order		Sevenspotted Lady Beetle	
##		45		46
##	Beetle Order		Snout Beetle Family, Weevil	
##		47		47
##	Erythrina Gall Wasp		Parasitoid Wasp	
##		49		51
##	Colorado Potato Beetle		Parastic Wasp	
##		57		58
##	Asian Citrus Psyllid		Minute Pirate Bug	
##		60		62
##	European Dark Bee		Wireworm	
##		66		69
##	Euonymus Scale		Asian Lady Beetle	
##		75		76
##	Japanese Beetle		Italian Honeybee	
##		94		113
##	Bumble Bee		Carniolan Honey Bee	
##		140		152
##	Buff Tailed Bumblebee		Parasitic Wasp	
##		183		285
##	Honey Bee		(Other)	
##		667		670

Answer: From highest to lowest 1.Other 2.Honey Bee 3.Parasitic Wasp 4.Buff Tailed Bumblebee 5.Carniolan Honey Bee 6.Bumble Bee Most of these species are types of bees. Bees are pollinators, and thus an important part of the ecosystem.

- Concentrations are always a numeric value. What is the class of Conc.1..Author. column in the dataset, and why is it not numeric? [Tip: Viewing the dataframe may be helpful...]

```
class(Neonics$Conc.1..Author.)
```

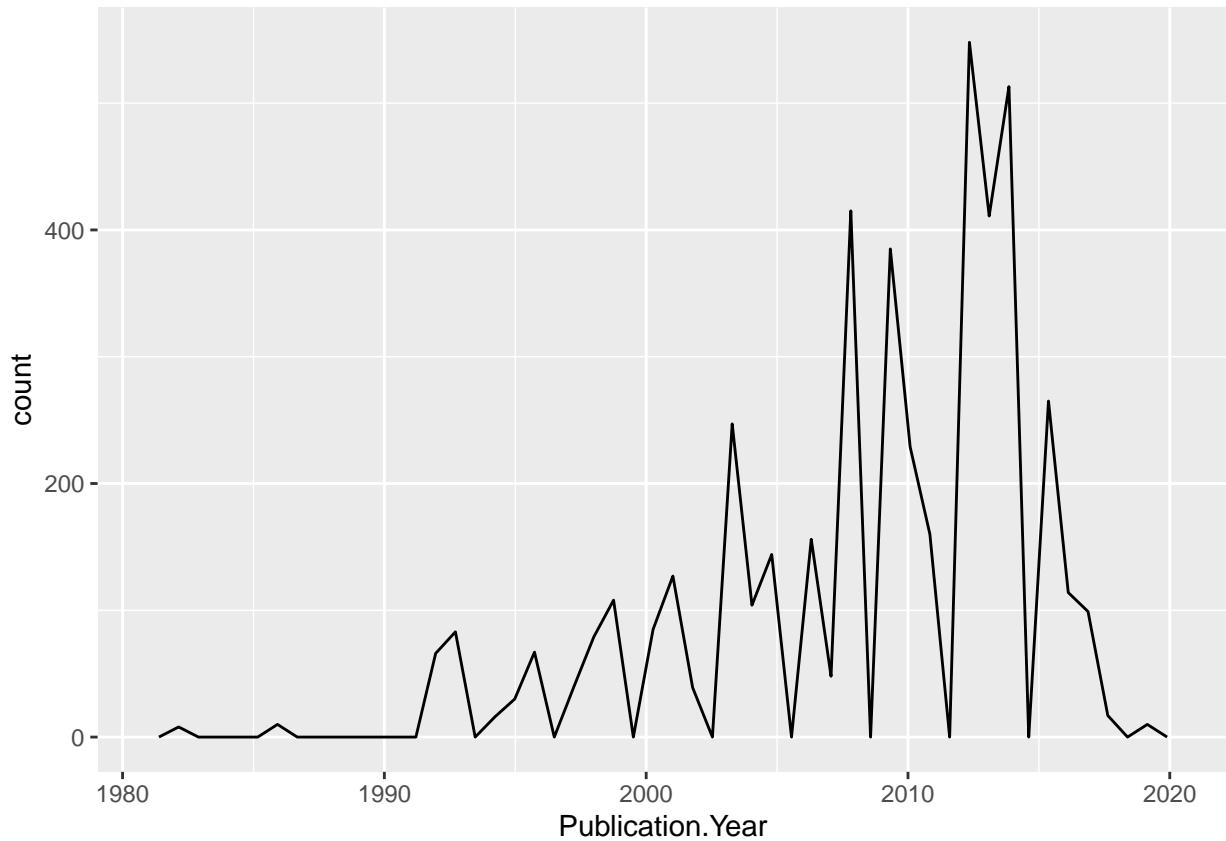
```
## [1] "factor"
```

Answer: This column is a factor, this is because while the column does include numbers, it also includes non number characters like letters, ~, /

Explore your data graphically (Neonics)

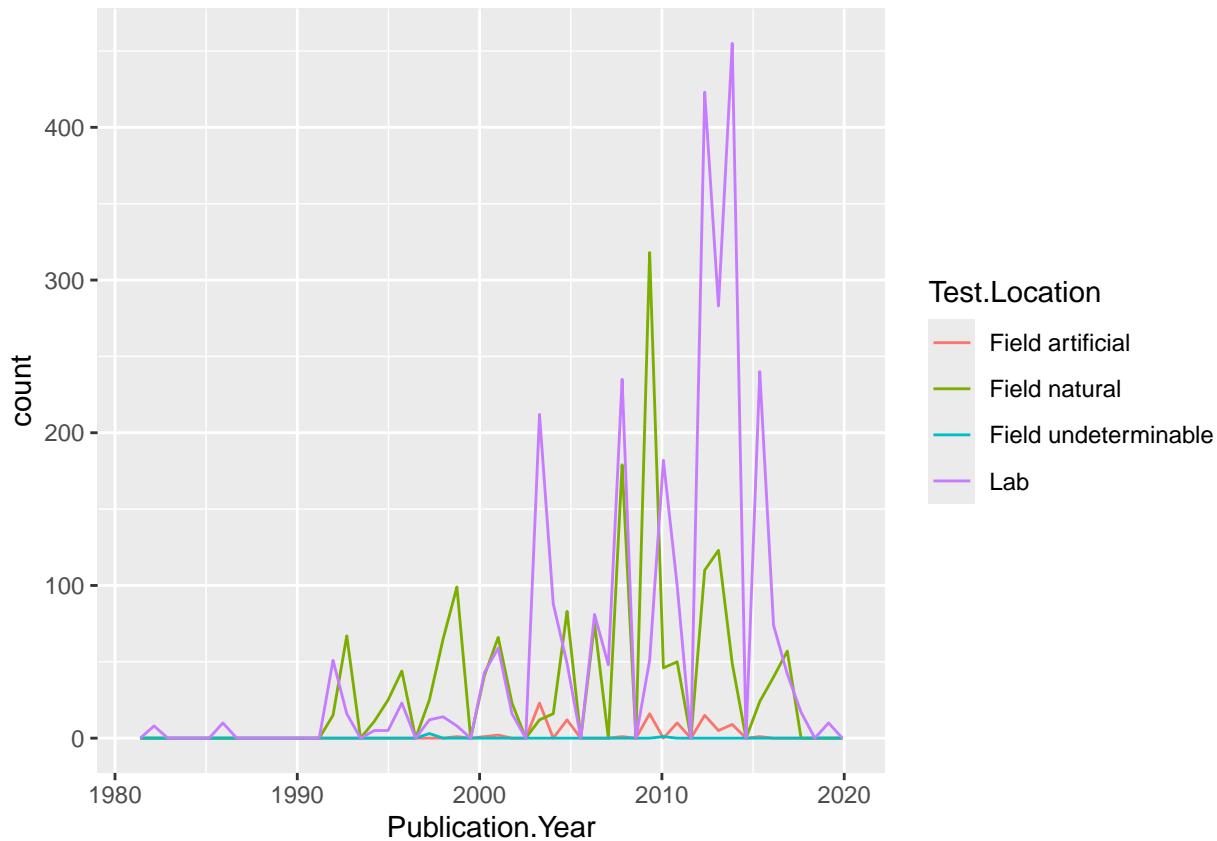
9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year), bins = 50 )
```



10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
Pub_test_fig <- ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 50 )  
Pub_test_fig
```



```
# Plotting the frequency polygon
```

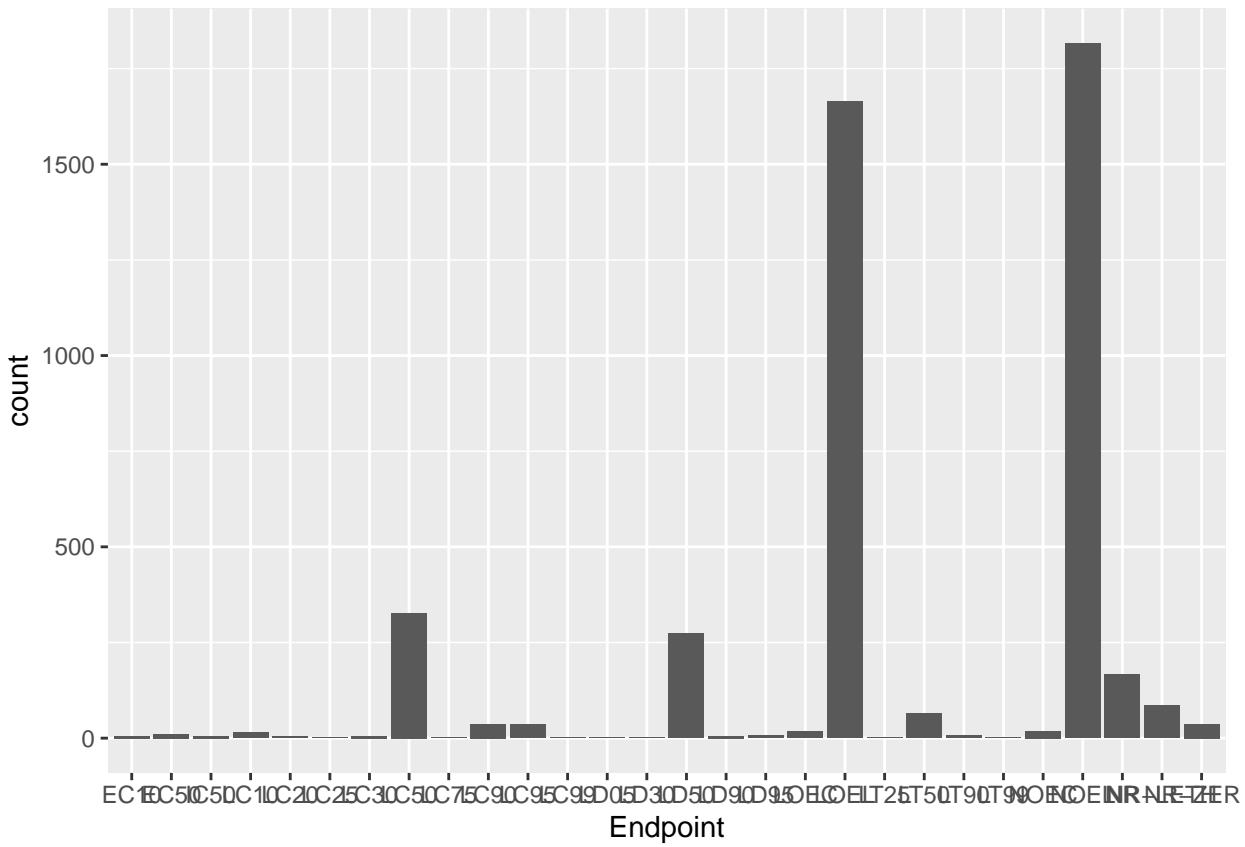
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: The most common test location is the lab, but they do differ over time with sometimes the field natural being the greatest, like just before 2010.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[TIP: Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics) +
  geom_bar(aes(x = Endpoint))
```



Answer: NOEL and LOEL are the two most common end points. LOEL is the Lowest Observed Effect Level - the lowest concentration where there is a statistically adverse effect. NOEL is the no observed effect level - the highest concentration where there is no statistically observed effect.

Explore your data (Litter)

12. Determine the class of `collectDate`. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate)

## [1] "factor"

#answer is factor, not date
Litter$collectDate <- as.Date(Litter$collectDate, format = "%Y-%m-%d")
class(Litter$collectDate)
```

13. Using the `unique` function, list the different plotIDs sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

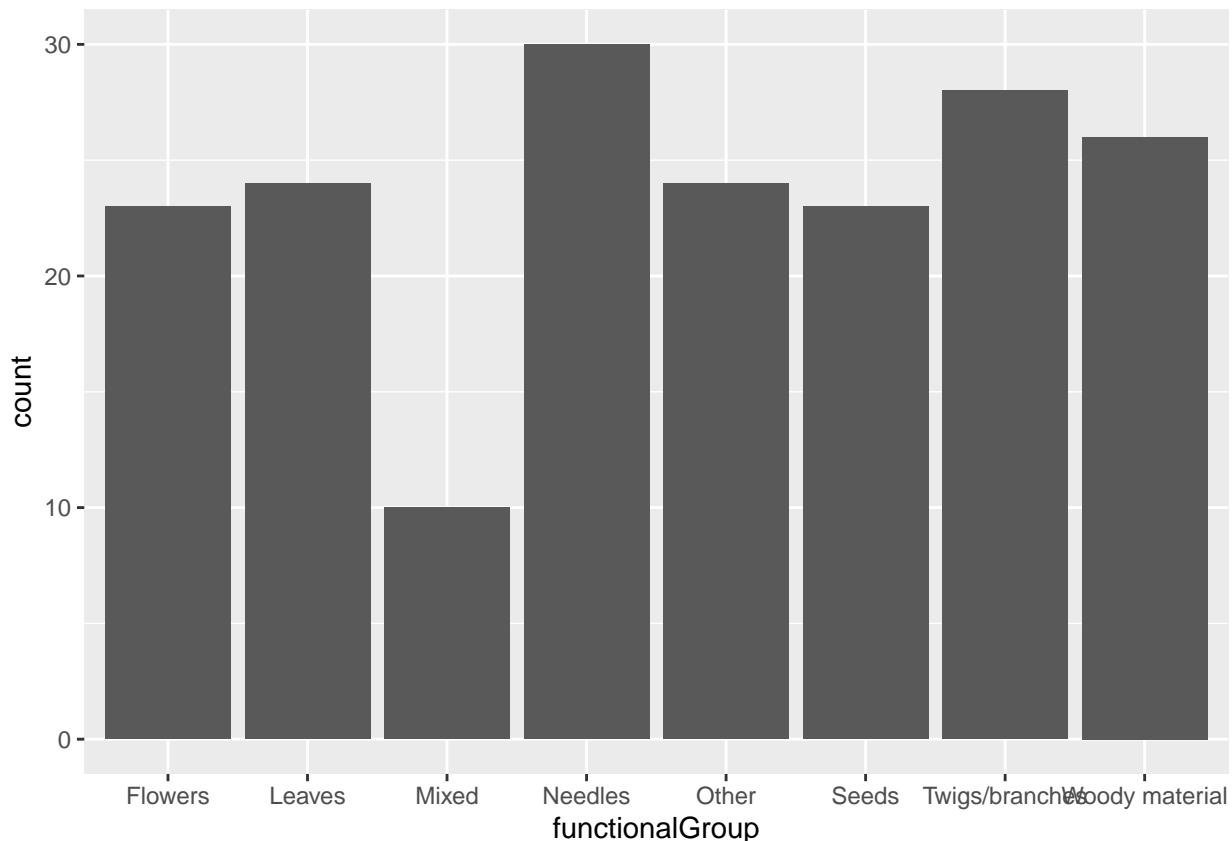
```
unique(Litter$plotID)
```

```
## [1] NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051
## [9] NIWO_058 NIWO_046 NIWO_062 NIWO_057
## 12 Levels: NIWO_040 NIWO_041 NIWO_046 NIWO_047 NIWO_051 NIWO_057 ... NIWO_067
```

Answer: 12 levels - NIWO_061 NIWO_064 NIWO_067 NIWO_040 NIWO_041 NIWO_063 NIWO_047 NIWO_051 NIWO_058 NIWO_046 NIWO_062 NIWO_057 summary will tell you the count of each level, while unique just shows how many different types there are.

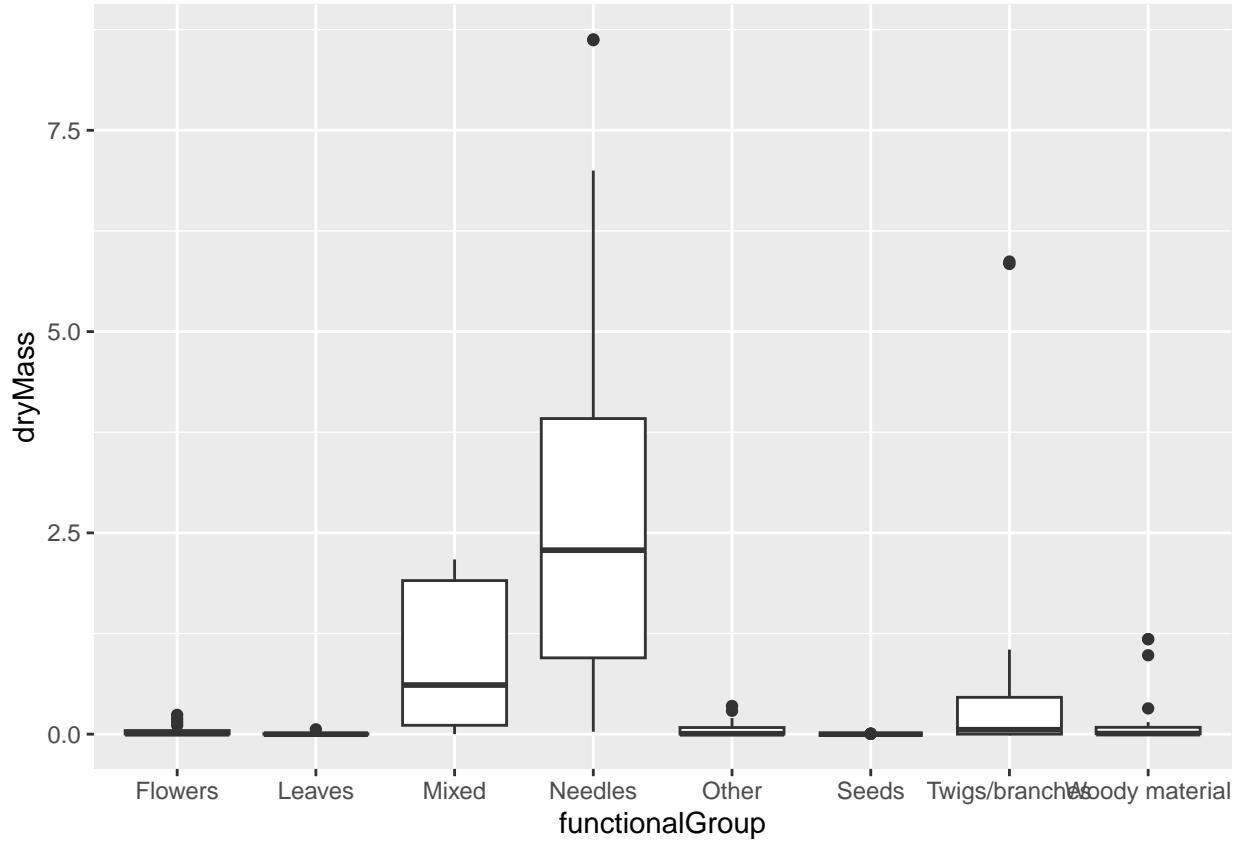
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter) +
  geom_bar(aes(x = functionalGroup))
```

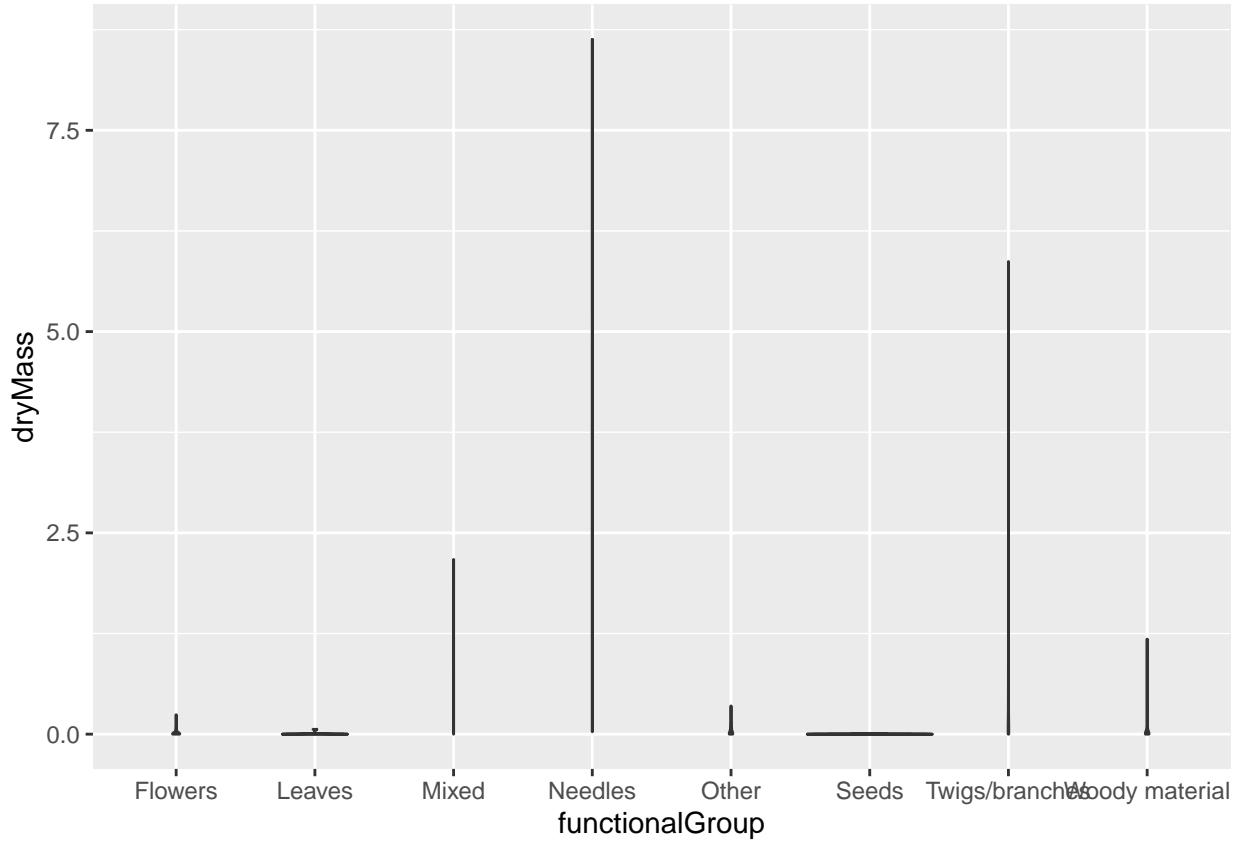


15. Using `geom_boxplot` and `geom_violin`, create a boxplot and a violin plot of dryMass by functionalGroup.

```
ggplot(Litter) +
  geom_boxplot(aes(x = functionalGroup, y = dryMass))
```



```
ggplot(Litter) +  
  geom_violin(aes(x = functionalGroup, y = dryMass),  
              draw_quantiles = c(0.25, 0.5, 0.75))
```



Why is the boxplot a more effective visualization option than the violin plot in this case?

Answer: In this case, the violin plots are not really showing violin shape or wide density (where the data is more concentrated), so the visual is lost and you are left with thin vertical lines which do not portray your data to viewers as nicely as the box plot does.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles has the highest, then mixed, then twigs/branches.