

Gibbs Sampling, More Bayesian Topics

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Computational Methods in Statistics and Data Science (Stats 406)

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- Inference is performed using **integration on posterior** (or other properties of π):

$$E(g(\theta) | x)$$

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 - **Rejection rate** trades off exploration for time to get a sufficient sample size
 - We frequently have to **tune parameters** to get samplers

Non-Conjugate Prior

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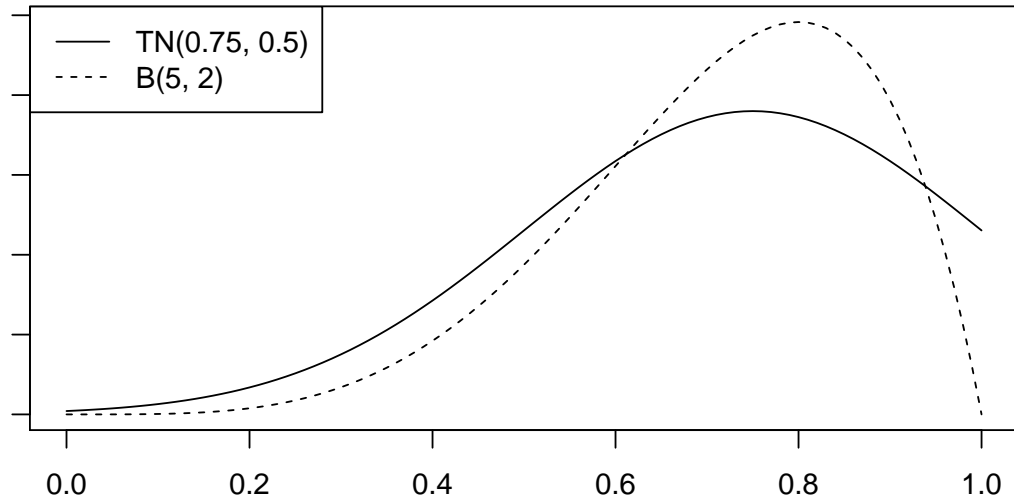
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What about other priors? Suppose instead described our beliefs using a **truncated Normal**?:

$$p(\theta) \propto \phi((\theta - \mu)/\sigma)$$

where ϕ is PDF of $N(0, 1)$. (Notice use of **proportionality**.)

Truncated Normal Prior



Posterior after seeing $X = 24$

What is the posterior distribution (proportional to)? Dropping **all terms not involving θ** ,

$$\pi(\theta \mid x) \propto \theta^x (1 - \theta)^x \exp \left\{ -\frac{(\theta - 0.75)^2}{0.5} \right\}$$

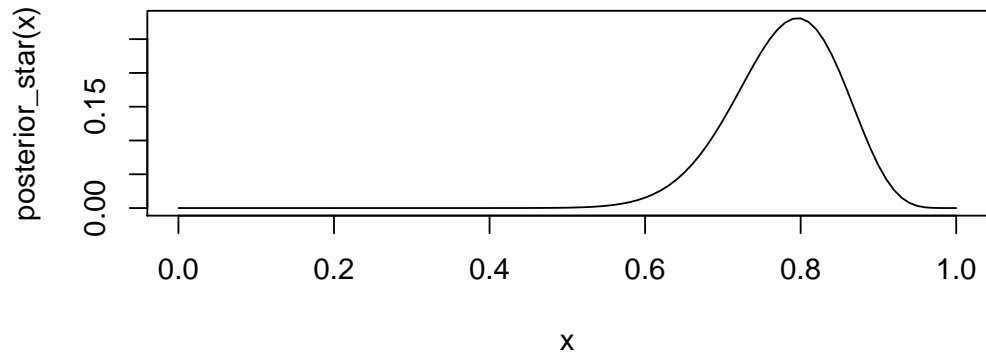
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```
> posterior_star <- function(theta) {  
+   dbinom(24, 30, prob = theta) * dnorm(theta, 0.75, 0.25)  
+ }
```

Unnormalized Posterior



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$$\theta(t)^* \sim U[a(\theta(t-1)), b(\theta(t-1))], a(x) = \max(0, x - \delta), b(x) = \min(x + \delta, 1)$$

so θ^* has density $1/(b(\theta(t-1)) - a(\theta(t-1)))$.

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Recall, we will need to compute:

$$\frac{\pi^*(\theta^* | x)}{\pi^*(\theta(t-1) | x)} \frac{g(\theta(t-1) | \theta^*)}{g(\theta^* | \theta(t-1))}$$

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$$\begin{aligned} \frac{\pi^*(\theta^* | x)}{\pi^*(\theta(t-1) | x)} \frac{g(\theta(t-1) | \theta^*)}{g(\theta^* | \theta(t-1))} &= \frac{\pi^*(\theta^* | x)}{\pi^*(\theta(t-1) | x)} \frac{\left[\frac{1}{b(\theta^*) - a(\theta^*)} \right]}{\left[\frac{1}{b(\theta(t-1)) - a(\theta(t-1))} \right]} \\ &= \frac{\pi^*(\theta^* | x)}{\pi^*(\theta(t-1) | x)} \frac{b(\theta(t-1)) - a(\theta(t-1))}{b(\theta^*) - a(\theta^*)} \end{aligned}$$

```
> findDeltas <- function(point, delta) {  
+   c(max(0, point - delta), # must be positive  
+     min(1, point + delta)) # must be less than 1  
+ }  
  
> drawCandidate <- function(previous, delta) {  
+   ab <- findDeltas(previous, delta)  
+   runif(1, ab[1], ab[2])  
+ }
```

```

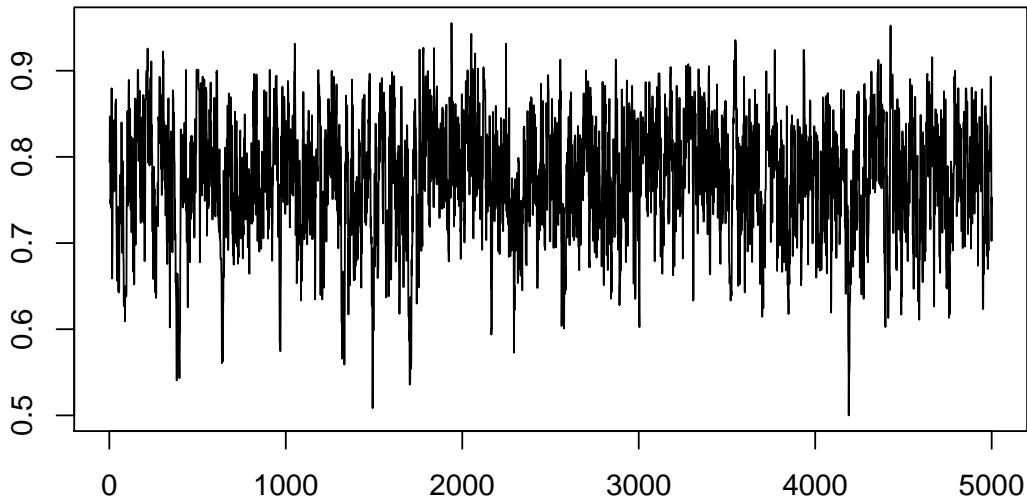
> nextDraw <- function(previous, delta = 0.1) {
+   candidate <- drawCandidate(previous, delta)
+
+   post_ratio <- posterior_star(candidate) / posterior_star(previous)
+   cand_ratio <- diff(findDeltas(previous, delta)) /
+                 diff(findDeltas(candidate, delta))
+
+   u <- runif(1)
+   if (u <= post_ratio * cand_ratio) {
+     return(candidate)
+   } else {
+     return(previous)
+   }
+ }

```

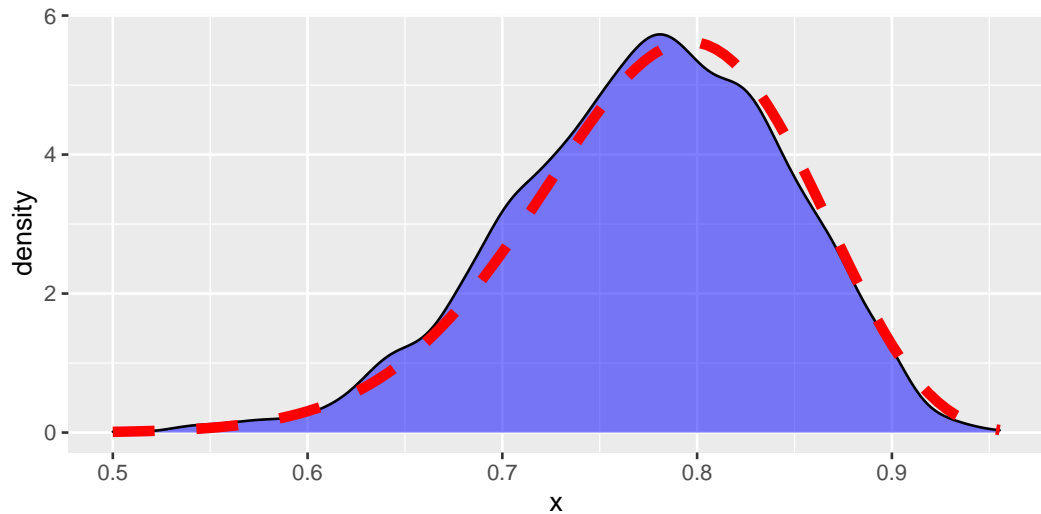
```
> B <- 10000 ; chain <- numeric(B) ; chain[1] <- 0.75
> for(i in 2:B) {
+   chain[i] <- nextDraw(chain[i - 1], 0.1)
+ }
> half_chain <- chain[5001:B] # drop burn in
> mean(half_chain > 0.75) # probability of successful teaching

[1] 0.6544
```

Chain (5000 to 10000)



Density



Multivariate Sampling

Multidimensional Posterior

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- β_0, β_1 and σ^2 from the model:

$$Y = \beta_0 + \beta_1 x + \epsilon, \quad \epsilon \sim N(0, \sigma^2)$$

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- Mixture distributions with $\sigma^2 > 0, \theta \in \{0, 1\}$:

$$\theta N(0, \sigma^2) + (1 - \theta) \text{Laplace}(0, \sigma^2)$$

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Example: Generating zero-mean bivariate Normals with variance-covariance matrix:

$$\begin{pmatrix} 1 & 0.8 \\ 0.8 & 1 \end{pmatrix}$$

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As a candidate, we'll use a **random walk** with $\epsilon = N(0, 0.25I_2)$.

Setup

```
> library(mvtnorm) # includes the multivariate normal density  
> B <- 5000  
> chain <- matrix(0, nrow = 2, ncol = B)  
> S1 <- matrix(c(1, 0.8, 0.8, 1), ncol = 2)  
> S2 <- matrix(c(0.25, 0, 0, 0.25), ncol = 2)
```

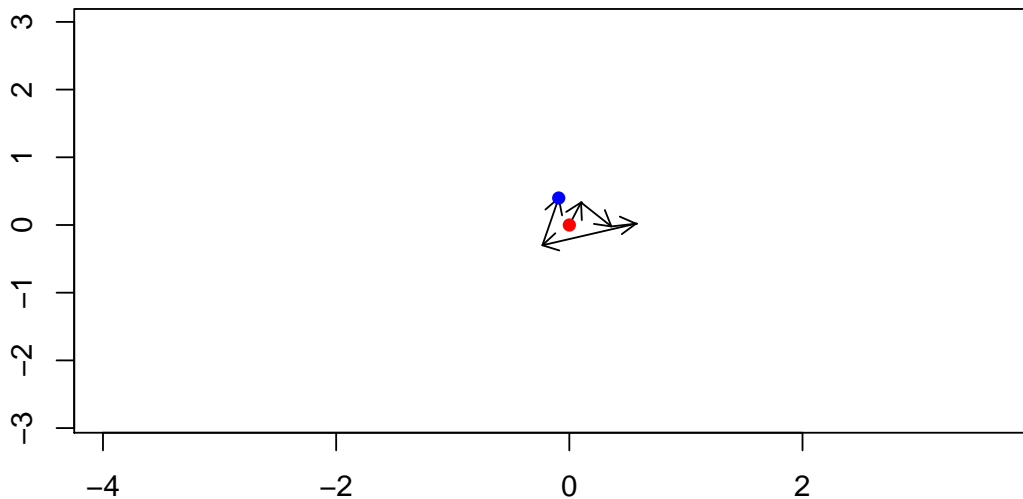
Computing Ratio

```
> ratio <- function(candidate, previous) {  
+  
+   ## evaluate  $\pi(\theta^*) / \pi(\theta(t - 1))$   
+   a <- dmvnorm(candidate, mean = c(0, 0), sigma = S1) /  
+       dmvnorm(previous, mean = c(0, 0), sigma = S1)  
+  
+   ## evaluate the candidate density  
+   b <- dmvnorm(previous, mean = candidate, sigma = S2) /  
+       dmvnorm(candidate, mean = previous, sigma = S2)  
+  
+   return(a * b)  
+ }
```

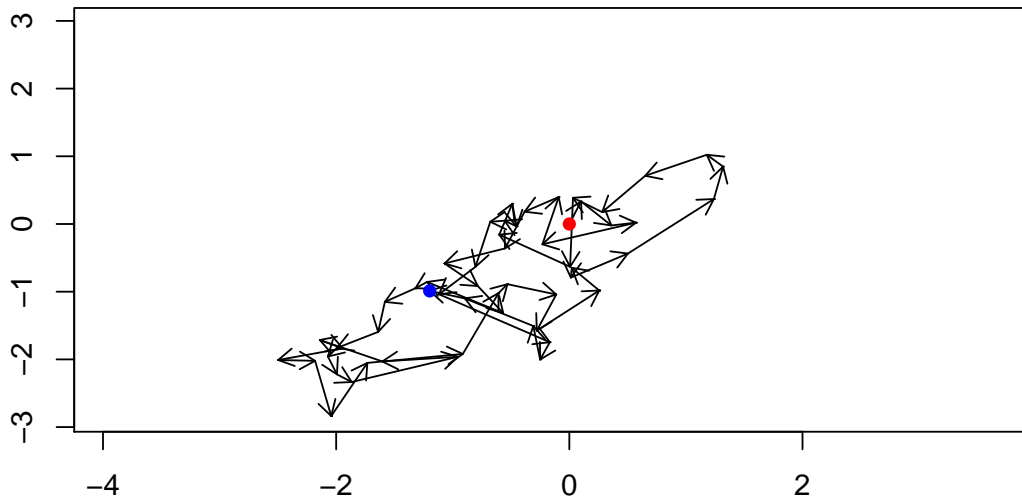
Making the Chain

```
> for (i in 2:B) {  
+   ## generate 2, independent Normals at the previous point  
+   candidate <- rnorm(2, mean = chain[, i - 1],  
+                       sd = sqrt(0.25))  
+  
+   r <- ratio(candidate, chain[, i - 1])  
+   if (runif(1) <= r) {  
+     chain[, i] <- candidate  
+   } else {  
+     chain[, i] <- chain[, i - 1]  
+   }  
+ }
```

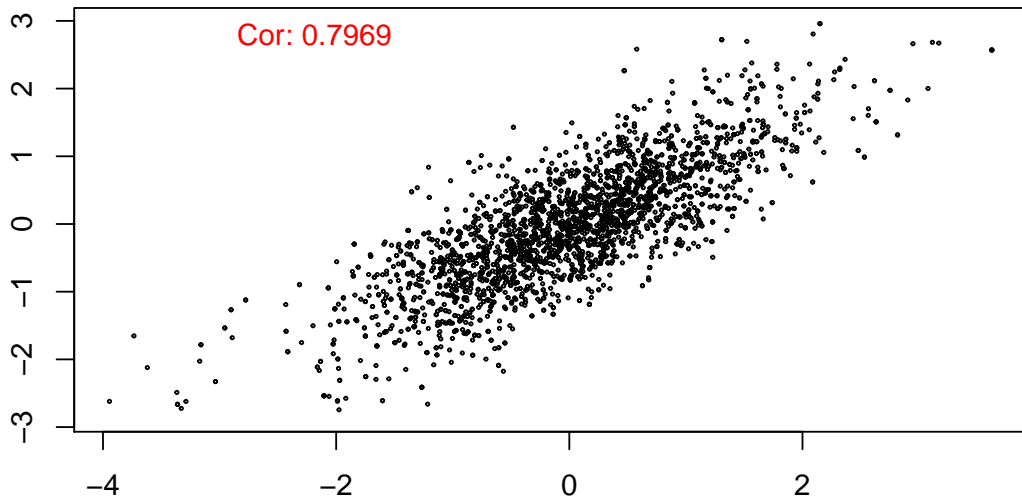
Chain: 1:10



Chain: 1:100



Chain: 2000:5000



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Gibbs sampling

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Define the **full conditional posterior** for parameter θ_i :

$$\pi(\theta_i \mid \theta_1, \theta_2, \dots, \theta_{i-1}, \theta_{i+1}, \dots, \theta_d, x) = \pi_i(\theta_i)$$

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Gibbs sampler: Step through the θ_i drawing from π_i conditional on the **rest of the parameters**.

Special case of Metropolis-Hastings that

- draws θ_i^* directly from π_i (which is always accepted)
- sets $\theta_j(t) = \theta_j(t-1)$ for all $j \neq i$.

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Gibbs for Multivariate Normal

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Two dimensional case:

$$\theta_1 | \theta_2 = t \sim N \left(\mu_1 + \frac{\text{Cov}(\theta_1, \theta_2)}{\text{Var}(\theta_2)}(t - \mu_2), \text{Var}(\theta_1) - \frac{\text{Cov}(\theta_1, \theta_2)^2}{\text{Var}(\theta_2)} \right)$$

$$\theta_2 | \theta_1 = t \sim N \left(\mu_2 + \frac{\text{Cov}(\theta_2, \theta_1)}{\text{Var}(\theta_1)}(t - \mu_1), \text{Var}(\theta_2) - \frac{\text{Cov}(\theta_2, \theta_1)^2}{\text{Var}(\theta_1)} \right)$$

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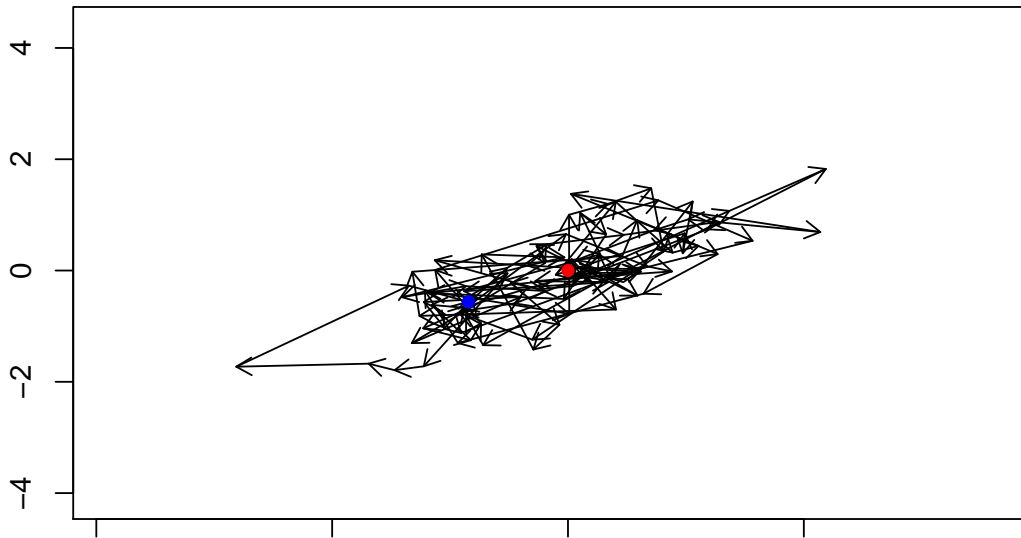
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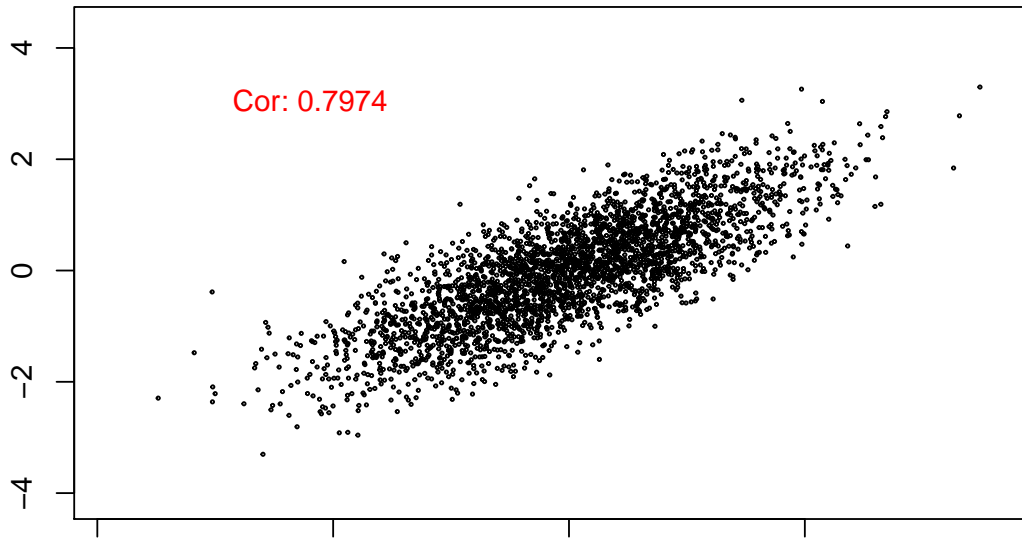
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In our example, $\mu_1 = \mu_2 = 0$, $\text{Var}(\theta_1) = \text{Var}(\theta_2) = 1$, and $\text{Cov}(\theta_1, \theta_2) = 0.8$.

```
> pi_i <- function(t) {  
+   rnorm(1, mean = 0.8 * t, sd = sqrt(1 - 0.8^2))  
+ }  
> chain_gibbs <- matrix(0, ncol = B, nrow = 2)  
> for (i in 2:B) {  
+   chain_gibbs[1, i] <- pi_i(chain_gibbs[2, i - 1])  
+   chain_gibbs[2, i] <- pi_i(chain_gibbs[1, i])  
+ }
```



Gibbs: 2000:5000



- Must be able to draw from the full conditional posterior directly.
- Often use conjugate prior distributions to make this possible.
- Gibbs sampling is frequently the choice for **hierarchical models** in which pieces are built on top of each other.

Hierarchical Model: Normal mean

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The Inverse Gamma distribution has density:

$$f(\sigma^2) = \frac{b^a}{(\sigma^2)^{a+1}} \frac{\exp(-b/\sigma^2)}{\Gamma(a)}$$

Full conditional posteriors: useful tricks

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The full conditional posterior for any parameter is **proportional to the joint posterior**. Writing θ_{-i} for all other parameters,

$$\pi(\theta_i \mid \theta_{-i}, x, \dots) = \frac{\pi(\theta_i, \theta_{-i} \mid x, \dots)}{\pi(\theta_{-i} \mid x, \dots)} \propto \pi(\theta_i, \theta_{-i} \mid x, \dots)$$

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Remember that addition in exponents is multiplicative:

$$g(a \mid b) = \exp(a + b) = \exp(a) \exp(b) \propto \exp(a)$$

Joint Posterior Distribution

$$\pi(\mu, \sigma^2 \mid \theta, \tau^2, a, b, \mathbf{x}) \propto f(\mathbf{x} \mid \mu, \sigma^2) p(\mu, \sigma^2)$$

Joint Posterior Distribution

$$\begin{aligned}\pi(\mu, \sigma^2 \mid \theta, \tau^2, a, b, \mathbf{x}) &\propto f(\mathbf{x} \mid \mu, \sigma^2) p(\mu, \sigma^2) \\ &\propto \exp \left\{ -\frac{\sum_{i=1}^n (x_i - \mu)^2}{2\sigma^2} \right\} \frac{1}{(\sigma^2)^{a+1}} \exp \{ -b/\sigma^2 \} \exp \left\{ -\frac{(\mu - \theta)^2}{2\tau^2} \right\}\end{aligned}$$

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Full Conditional for μ

$$\pi(\mu \mid \sigma^2, \theta, \tau^2, a, b, x) \propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \frac{1}{(\sigma^2)^{a+1}} \exp \{ -b/\sigma^2 \}$$

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$$\begin{aligned}\pi(\mu \mid \sigma^2, \theta, \tau^2, a, b, x) &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \frac{1}{(\sigma^2)^{a+1}} \exp \{ -b/\sigma^2 \} \\ &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x_i - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\}\end{aligned}$$

Full Conditional for μ

$$\begin{aligned}\pi(\mu \mid \sigma^2, \theta, \tau^2, a, b, x) &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x_i - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \frac{1}{(\sigma^2)^{a+1}} \exp \{ -b/\sigma^2 \} \\ &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x_i - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \\ &= \exp \left\{ -\frac{\tau^2 \left(\left[\sum_{i=1}^n x_i^2 \right] - 2n\bar{x}\mu + n\mu^2 \right) + \sigma^2 \mu^2 - 2\sigma^2 \mu \theta + \sigma^2 \theta^2}{2\sigma^2 \tau^2} \right\}\end{aligned}$$

Full Conditional for μ

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Full Conditional for μ

$$\begin{aligned}\pi(\mu \mid \sigma^2, \theta, \tau^2, a, b, x) &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x_i - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \frac{1}{(\sigma^2)^{a+1}} \exp \{ -b/\sigma^2 \} \\ &\propto \exp \left\{ -\frac{\tau^2 \sum_{i=1}^n (x_i - \mu)^2 + \sigma^2 (\mu - \theta)^2}{2\sigma^2 \tau^2} \right\} \\ &= \exp \left\{ -\frac{\tau^2 \left(\left[\sum_{i=1}^n x_i^2 \right] - 2n\bar{x}\mu + n\mu^2 \right) + \sigma^2 \mu^2 - 2\sigma^2 \mu \theta + \sigma^2 \theta^2}{2\sigma^2 \tau^2} \right\} \\ &\propto \exp \left\{ -\frac{(\tau^2 + n\sigma^2)\mu^2 - 2(\sigma^2 \mu \theta - \tau^2 n\bar{x})\mu}{2\sigma^2 \tau^2} \right\} \\ &\propto \exp \left\{ -\frac{(\mu - (\sigma^2 \mu \theta - \tau^2 n\bar{x})/(\tau^2 + n\sigma^2))^2}{2\sigma^2 \tau^2 / (\tau^2 + n\sigma^2)} \right\}\end{aligned}$$

After working through $\pi(\mu, \sigma^2 | x)$, we find:

$$\mu | \sigma^2, x \sim N \left(\frac{\sigma^2 \theta + n \tau^2 \bar{x}}{\sigma^2 + n \tau^2}, \frac{\sigma^2 \tau^2}{\sigma^2 + n \tau^2} \right)$$

$$\sigma^2 | \mu, x \sim \text{Inv. Gamma} \left(\frac{n}{2} + a, \frac{1}{2} \sum_{i=1}^n (x_i - \mu)^2 + b \right)$$

Prior for μ

We'll work on the **systolic blood pressure** data again.

Prior for μ

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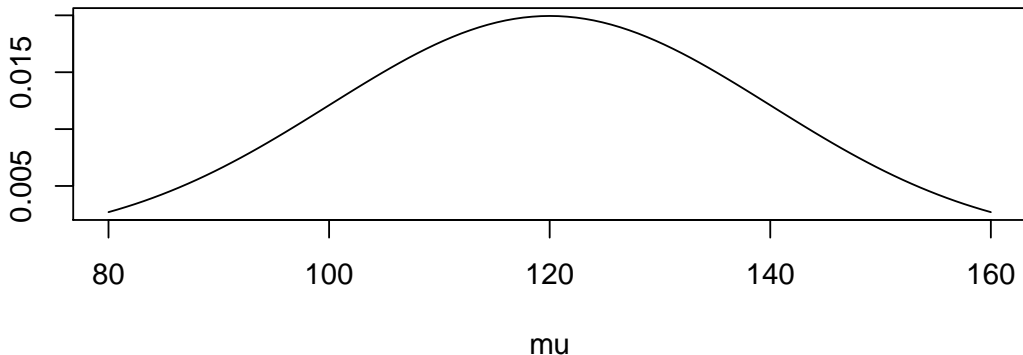
If we start from the idea that a **healthy systolic BP** is about **120**, so $\theta = 120$.

Prior for μ

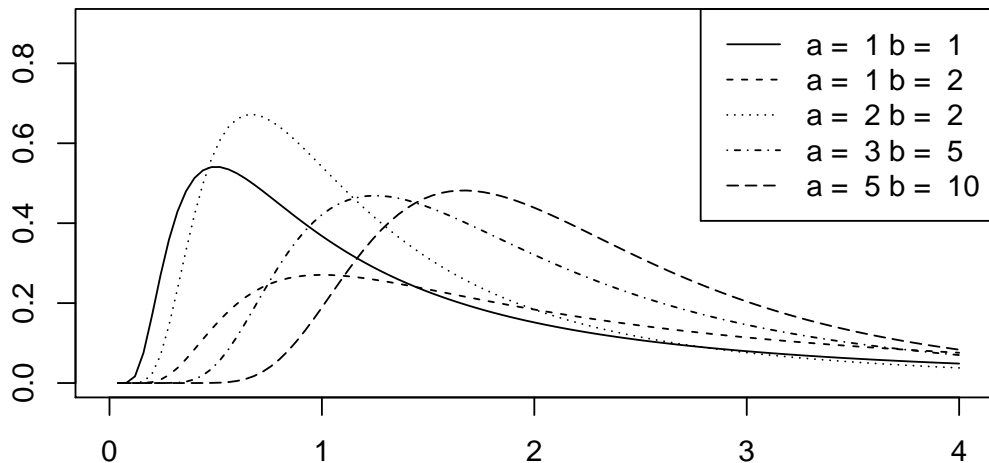
We'll work on the **systolic blood pressure** data again.

If we start from the idea that a **healthy systolic BP** is about **120**, so $\theta = 120$.

We might not be very certain about that, so let's provide a **wide variance**: $\tau = 20$.



Inverse Gamma



Prior for σ^2 , setup

For my **subjective beliefs**, I think $a = 2$, $b = 5$ best captures my position.

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```
> param_a <- 3  
> param_b <- 5  
> param_theta = 120  
> param_tau2 = 20^2  
> n <- dim(combined)[1] # the BP data
```

Gibbs for BP: μ step

$$\mu | \sigma^2, x \sim N \left(\frac{\sigma^2 \theta + n \tau^2 \bar{x}}{\sigma^2 + n \tau^2}, \frac{\sigma^2 \tau^2}{\sigma^2 + n \tau^2} \right)$$

```
> xsum <- sum(combined$sys_mean)
> gibbs_mu <- function(s2) {
+   denom <- s2 + n * param_tau2
+   rnorm(1, mean = (s2 * param_theta + param_tau2 * xsum) / denom,
+         sd = sqrt(s2 * param_tau2 / denom))
+ }
```

Gibbs for BP: σ^2 step

$$\sigma^2 \mid \mu, x \sim \text{Inv. Gamma} \left(\frac{n}{2} + a, \frac{1}{2} \sum_{i=1}^n (x - \mu)^2 + b \right)$$

```
> shape_1 <- n/2 + param_a # doesn't depend on mu
> gibbs_s2 <- function(mu) {
+   shape_2 <- (0.5) * sum((combined$sys_mean - mu)^2) + param_b
+   g <- rgamma(1, shape_1, shape_2)
+   1 / g # recall: inverse gamma
+ }
```

Creating the chain

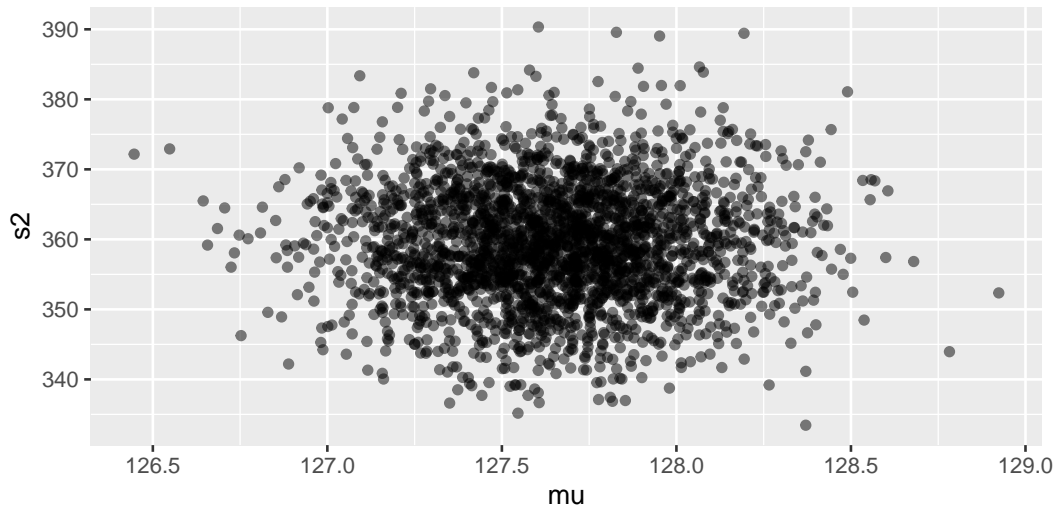
```
> gibbs_mu_sigma2 <- matrix(0, nrow = 2, ncol = B)
> ## draw from the prior to set up the chain
> gibbs_mu_sigma2[1, 1] <- rnorm(1, param_theta, sqrt(param_tau2))
> gibbs_mu_sigma2[2, 1] <- 1 / rgamma(1, param_a, param_b)
```

Creating the chain

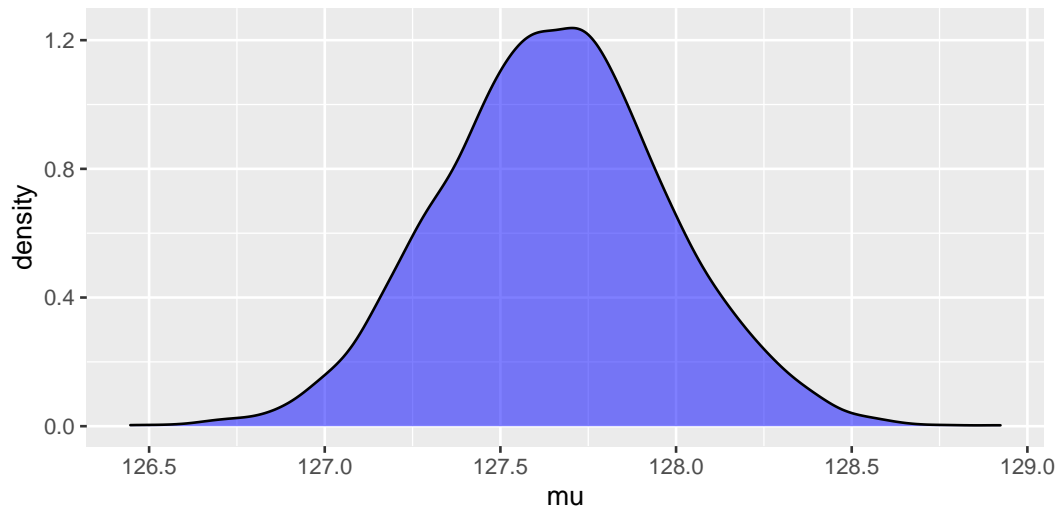
```
> gibbs_mu_sigma2 <- matrix(0, nrow = 2, ncol = B)
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> gibbs_mu_sigma2[2, 1] <- 1 / rgamma(1, param_a, param_b)

> for (i in 2:B) {
+   chain[1, i] <- gibbs_mu(chain[2, i - 1])
+   chain[2, i] <- gibbs_s2(chain[1, i])
+ }
```

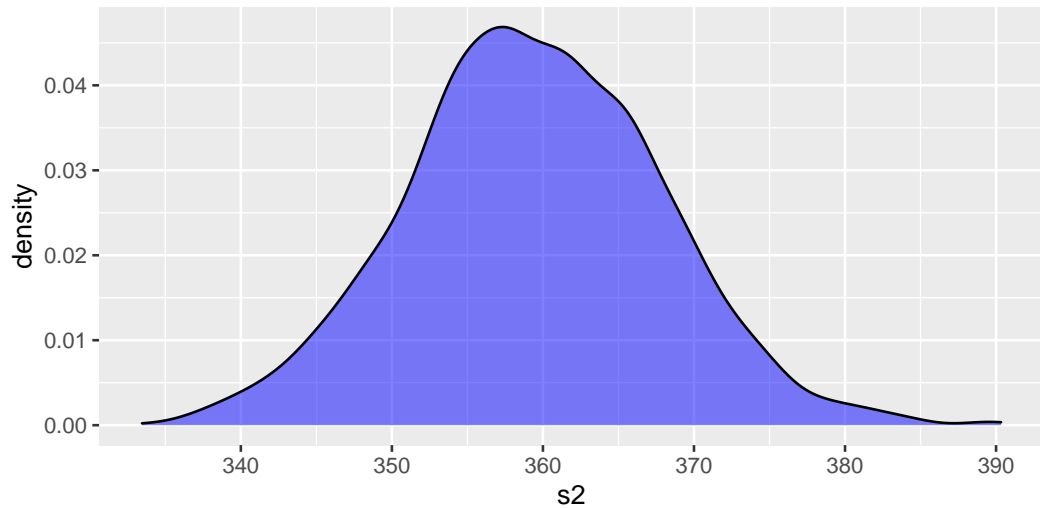
Joint distribution (discarding first 1/2 chain)



Marginal: μ



Marginal: σ^2



How much did we decrease our uncertainty?

For μ :

```
> 1 - var(chain[1, -(1:500)]) / 20^2 # recall, tau^2 = 20^2  
[1] 0.9997
```

We've greatly increased our certainty on the mean.

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For μ :

```
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[1] 0.9997
```

We've greatly increased our certainty on the mean.

For σ^2 :

```
> prior_var <- param_b^2 / ((param_a - 1)^2 * (param_a - 2))  
> 1 - var(chain[2, -(1:500)]) / prior_var  
[1] -10.05
```

But now we are much less certain about the variance (perhaps we were over confident before!).

Inference

```
> # discard the first 500 draws  
> stationary <- chain[, -(1:500)]
```

Inference

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> stationary <- chain[, -(1:500)]
```

Bayes estimators (with regard to squared loss/MSE) for the parameters:

```
> # mu  
> mean(stationary[1,])
```

```
[1] 127.7
```

```
> # sigma^2  
> mean(stationary[2,])
```

```
[1] 359.2
```

More inference

95% credible intervals for the parameters (intervals that contain 95% of the mass for marginal posterior distributions:

```
> # mu  
> quantile(stationary[1, ], c(0.025, 0.975))
```

```
 2.5% 97.5%  
127.0 128.3
```

```
> # sigma^2  
> quantile(stationary[2, ], c(0.025, 0.975))
```

```
 2.5% 97.5%  
343.0 375.3
```

Gibbs Sampling Summary

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 1. Draw θ_1 conditional on $\theta_2, \theta_3, \dots, \theta_k$
 2. Draw θ_2 conditional on $\theta_1, \theta_3, \dots, \theta_k$
 3. ...
 4. Draw θ_k conditional on $\theta_1, \theta_2, \dots, \theta_{k-1}$
- Most of the work is getting the FPCs.

Bayesian Computation with Stan

Stan: A Bayesian Programming Language



- A programming language for specifying Bayesian models
- A library R, Python, and other languages
- A collection of advanced Bayesian algorithms

Example: BP Model

$$X_i \sim N(\mu, \sigma^2)$$

$$\mu \sim N(120, 400)$$

$$\sigma^2 \sim \text{Inv. Gamma}(2, 5)$$

Data and Parameters

```
data {  
  int<lower=0> n;  
  real y[n];  
}
```

```
parameters {  
  real mu;  
  real<lower=0> sigmasq;  
}
```

Model

```
model {  
  mu ~ normal(120, 20);  
  sigmasq ~ inv_gamma(3, 5);  
  for (i in 1:n)  
    y[i] ~ normal(mu, sqrt(sigmasq));  
}
```

Running the `bp.stan`

The data, parameters, and model sections are put into `bp.stan`.

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```
> library(rstan)
> bp_stan <- stan("bp.stan",
+               data = list(n = dim(combined)[1],
+               y = combined$sys_mean))
```

Running the bp.stan

The data, parameters, and model sections are put into `bp.stan`.

```
> library(rstan)
> bp_stan <- stan("bp.stan",
+               data = list(n = dim(combined)[1],
+               y = combined$sys_mean))
```

After compiling `bp.stan` into its own executable, we see

```
SAMPLING FOR MODEL 'bp' NOW (CHAIN 1).
```

```
Chain 1:
```

```
Chain 1: Iteration:    1 / 2000 [  0%]   (Warmup)
```

```
Chain 1: Iteration:   200 / 2000 [ 10%]   (Warmup)
```

```
...
```

Results

```
> print(bp_stan)
```

Inference for Stan model: bp.

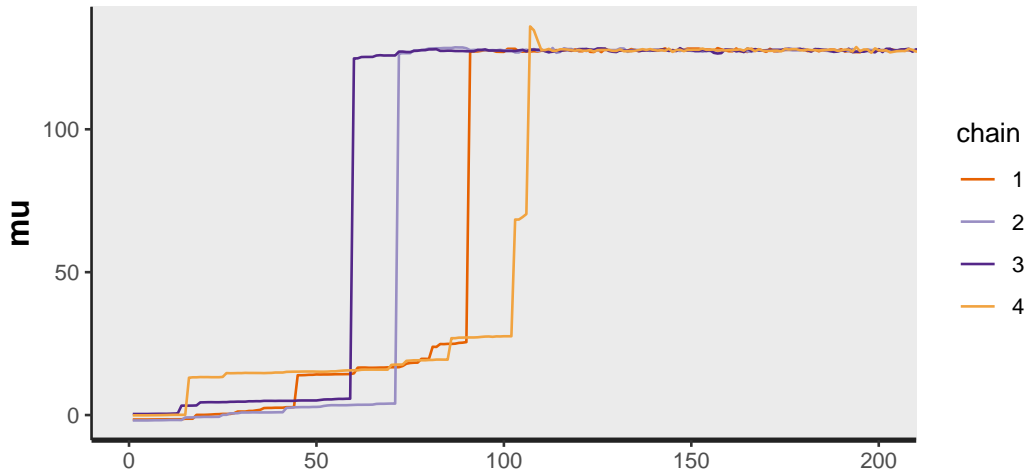
4 chains, each with iter=2000; warmup=1000; thin=1;

post-warmup draws per chain=1000, total post-warmup draws=4000.

	mean	se_mean	sd	2.5%	25%	50%
mu	127.6	0.01	0.31	127.0	127.4	127.6
sigmasq	358.6	0.15	8.37	342.4	353.0	358.4
lp__	-12243.4	0.02	0.96	-12246.0	-12243.8	-12243.1
	75%	97.5%	n_eff	Rhat		
mu	127.8	128.2	3644	1		
sigmasq	364.1	375.4	3086	1		
lp__	-12242.7	-12242.5	2055	1		

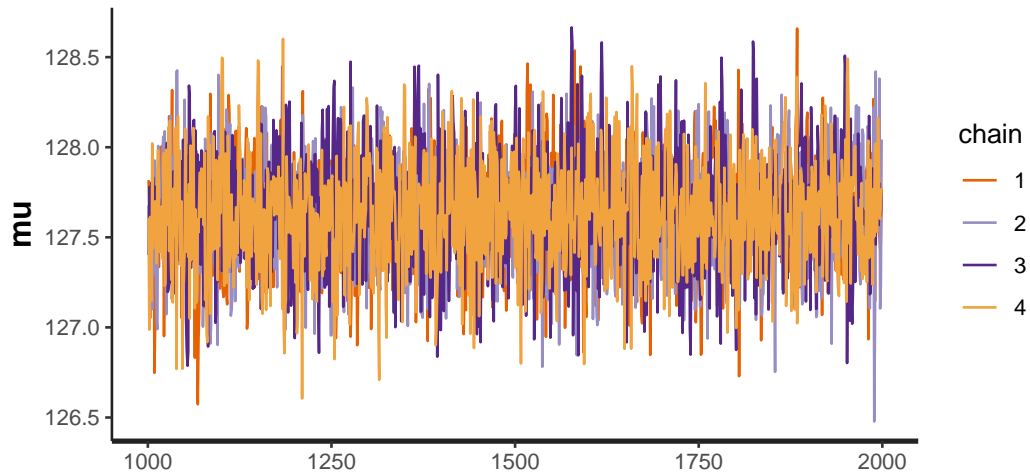
Plotting chains, warmup period

```
> traceplot(bp_stan, pars = "mu", inc_warmup = TRUE, window = c(0, 200))
```



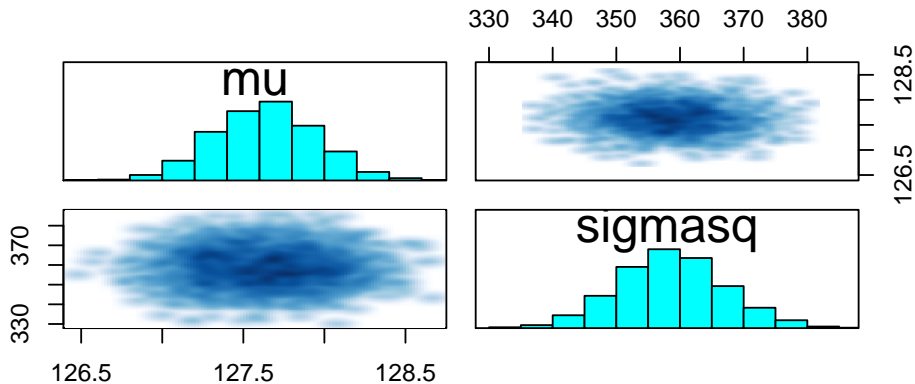
Plotting chains, convergence

```
> traceplot(bp_stan, "mu")
```



Joint Posterior

```
> pairs(bp_stan, pars = c("mu", "sigmasq"))
```



A more involved model

We previously investigated the **effect of aspirin** on BP, comparing subjects who took to those who did not.

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We previously investigated the **effect of aspirin** on BP, comparing subjects who took to those who did not.

$$\mu_1 \sim N(120, 20)$$

$$\delta \sim N(0, 10)$$

$$\sigma^2 \sim \text{Inv. Gamma}(3, 5)$$

$$X_i \sim N(\mu_1 + I(T_i = 0)\delta, \sigma^2)$$

Data and Parameters

```
data {  
  int<lower=1> n;  
  int<lower=1> m;  
  real takers[n];  
  real nontakers[m];  
}  
  
parameters {  
  real mu_1;  
  real delta;  
  real<lower=0> sigmasq;  
}
```

Transformed Parameters and Model

```
transformed parameters {  
  real mu_2 = mu_1 + delta;  
}  
  
model {  
  mu_1      ~ normal(120, 20);  
  delta     ~ normal(0, 10);  
  sigmasq   ~ inv_gamma(3, 5);  
  takers    ~ normal(mu_1, sqrt(sigmasq));  
  nontakers ~ normal(mu_2, sqrt(sigmasq));  
}
```

Running the model

```
> aspirin_effect <- stan("aspirin_effect.stan",  
+   data = with(combined,  
+               list(  
+                 n = sum(taking_aspirin),  
+                 m = sum(!taking_aspirin),  
+                 takers = sys_mean[taking_aspirin],  
+                 nontakers = sys_mean[!taking_aspirin])))
```


Results

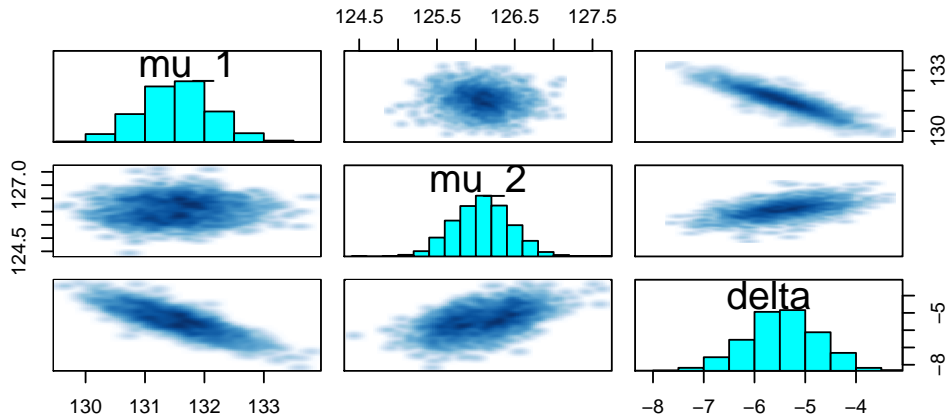
Inference for Stan model: aspirin_effect.

4 chains, each with iter=2000; warmup=1000; thin=1;

post-warmup draws per chain=1000, total post-warmup draws=4000.

	mean	se_mean	sd	2.5%	25%	50%
mu_1	131.53	0.02	0.59	130.37	131.1	131.54
delta	-5.45	0.02	0.70	-6.86	-5.9	-5.44
sigmasq	352.93	0.16	8.24	337.06	347.3	352.75
mu_2	126.08	0.01	0.37	125.39	125.8	126.08
lp__	-12213.71	0.03	1.21	-12216.65	-12214.3	-12213.41
	75%	97.5%	n_eff	Rhat		
mu_1	131.92	132.72	1531	1		
delta	-4.97	-4.13	1645	1		
sigmasq	358.32	369.39	2676	1		

Posterior Distribution



Using the posterior distribution

```
> samples <- as.array(aspirin_effect)
```

```
> dimnames(samples)
```

```
$iterations
```

```
NULL
```

```
$chains
```

```
[1] "chain:1" "chain:2" "chain:3" "chain:4"
```

```
$parameters
```

```
[1] "mu_1"      "delta"     "sigmasq"   "mu_2"      "lp_--"
```

```
> ## probability that the effect size is at least 5  
> mean(abs(samples[, "delta"]) >= 5)  
  
[1] 0.7362
```

Installing Stan

In theory, installing Stan should be as easy as:

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> install.packages("rstan")
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```
> install.packages("rstan")
```

In practice, I've had **mixed success** (yes on my linux machine, no on my OS X machine).

We won't use Stan in class, but feel free to use on your final project.

Convergence

Example: $N(0, 1)$

Recall this example from last class:

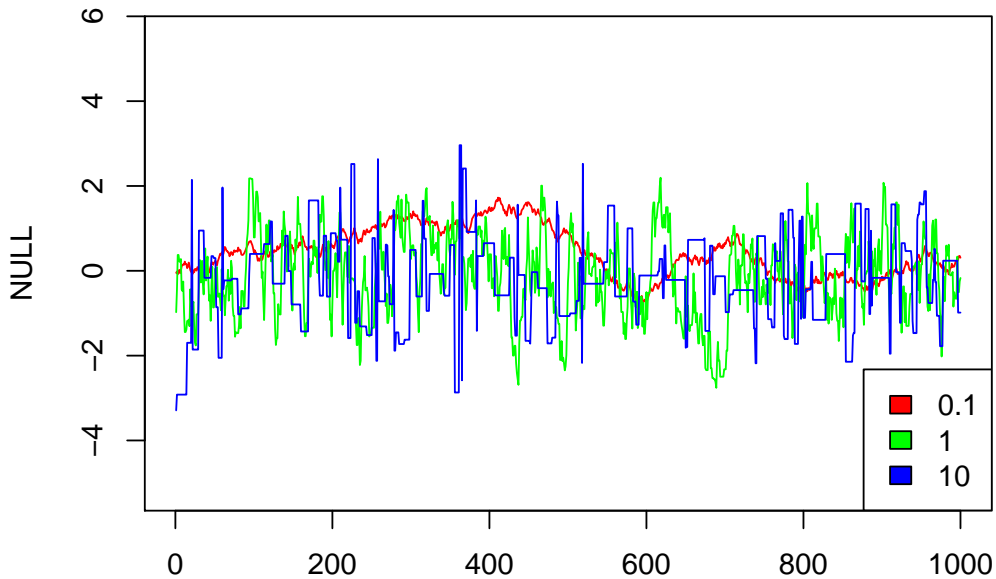
Generating draws from $N(0, 1)$ using Metropolis-Hastings with

$$\theta^* = \theta(t - 1) + U(-\delta, \delta)$$

as a proposal.

```
> unif_chain <- function(delta, B = 5000) {  
+   chain <- numeric(B); chain[1] <- runif(1, -delta, delta) ; rejects <-  
+   for (i in 2:B) {  
+     candidate <- chain[i - 1] + runif(1, -delta, delta)  
+     ratio <- dnorm(candidate) / dnorm(chain[i - 1])  
+     if (runif(1) <= ratio) {  
+       chain[i] <- candidate  
+     } else {  
+       chain[i] <- chain[i - 1]  
+       rejects <- rejects + 1  
+     }  
+   }  
+   list(reject_rate = rejects / B, chain = chain)  
+ }
```

```
> n01_chain_0.1 <- unif_chain(0.1)
> n01_chain_1 <- unif_chain(1)
> n01_chain_10 <- unif_chain(10)
```



Convergence

We can see that for $\delta = 0.1$, the **chain did not converge**.

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This conclusion was made easier **generating multiple chains**.

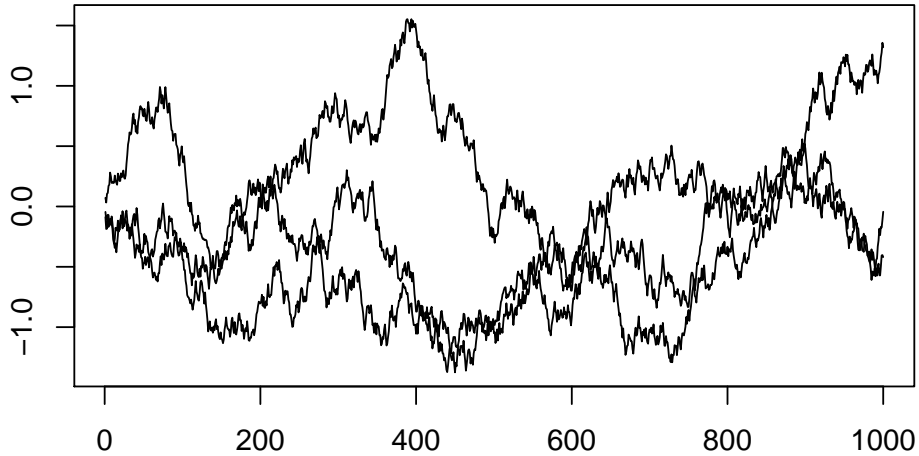
Convergence

We can see that for $\delta = 0.1$, the **chain did not converge**.

This conclusion was made easier **generating multiple chains**.

To check convergence for a single δ , we will need to generate several chains. Luckily, we can do this in **parallel**.

```
> library(parallel)
> RNGkind("L'Ecuyer-CMRG") # parallel safe PRNG
> chains_0.1 <- mclapply(rep(0.1, 3), unif_chain, mc.cores = 3, B = 1000)
```

Gelman-Rubin Method

Suppose we have k Markov Chains from 1 up to n :

$$X_1(0), X_1(2), \dots, X_1(n)$$

$$\vdots$$

$$X_k(0), X_k(1), \dots, X_k(n)$$

Gelman-Rubin Method

Suppose we have k Markov Chains from 1 up to n :

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$$\vdots$$

$$X_k(0), X_k(1), \dots, X_k(n)$$

Let ψ be a function of n arguments. We can compute:

$$\psi_{1n} = \psi_n(X_1(0), \dots, X_1(n))$$

$$\vdots$$

$$\psi_{kn} = \psi_n(X_k(0), \dots, X_k(n))$$

Gelman-Rubin Method

Suppose we have k Markov Chains from 1 up to n :

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Let ψ be a function of n arguments. We can compute:

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$$\vdots$$

$$\psi_{kn} = \psi_n(X_k(0), \dots, X_k(n))$$

Observation: If the chains are converging to the same distribution, then **all ψ_{in} should converge** as well.

Example: Mean

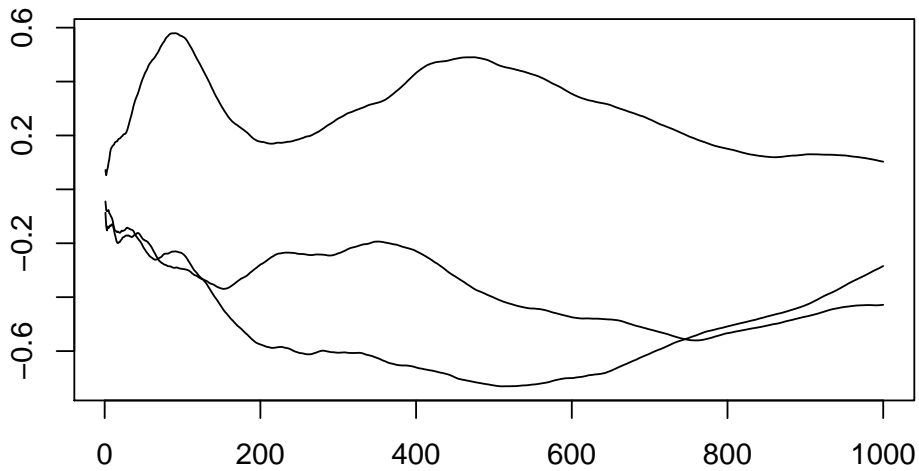
One common choice for ψ_n is the **mean of the chain up n** :

```
> psi <- function(chain) {  
+   n <- length(chain)  
+   cumsum(chain) / (1:n) # cumulative mean  
+ }
```

Example: Mean

One common choice for ψ_n is the **mean of the chain up n** :

```
> psi <- function(chain) {  
+   n <- length(chain)  
+   cumsum(chain) / (1:n) # cumulative mean  
+ }  
  
> psi_1 <- psi(chains_0.1[[1]]$chain)  
> psi_2 <- psi(chains_0.1[[2]]$chain)  
> psi_3 <- psi(chains_0.1[[3]]$chain)
```



Within and Between Variances

Define the following:

$$B = \frac{n}{k-1} \sum_{i=1}^l (\bar{\psi}_{i\cdot} - \bar{\psi}_{\cdot\cdot})^2$$
$$W = \frac{1}{nk} \sum_{i=1}^k \sum_{t=1}^n (\psi_{it} - \bar{\psi}_{i\cdot})^2$$

where the “dot” notation means taking the mean over that index.

Within and Between Variances

Define the following:

$$B = \frac{n}{k-1} \sum_{i=1}^l (\bar{\psi}_{i\cdot} - \bar{\psi}_{\cdot\cdot})^2$$
$$W = \frac{1}{nk} \sum_{i=1}^k \sum_{t=1}^n (\psi_{it} - \bar{\psi}_{i\cdot})^2$$

where the “dot” notation means taking the mean over that index.

B measures how much **each chain's average varies about the overall average**.

Within and Between Variances

Define the following:

$$B = \frac{n}{k-1} \sum_{i=1}^l (\bar{\psi}_{i\cdot} - \bar{\psi}_{\cdot\cdot})^2$$
$$W = \frac{1}{nk} \sum_{i=1}^k \sum_{t=1}^n (\psi_{it} - \bar{\psi}_{i\cdot})^2$$

where the “dot” notation means taking the mean over that index.

B measures how much **each chain's average varies about the overall average**.

W measures the **within chain variance**.

Variance of ψ

An unbiased estimate of the variance of ψ (with respect to the true posterior) is

$$V = \frac{n-1}{n}W + \frac{1}{n}B$$

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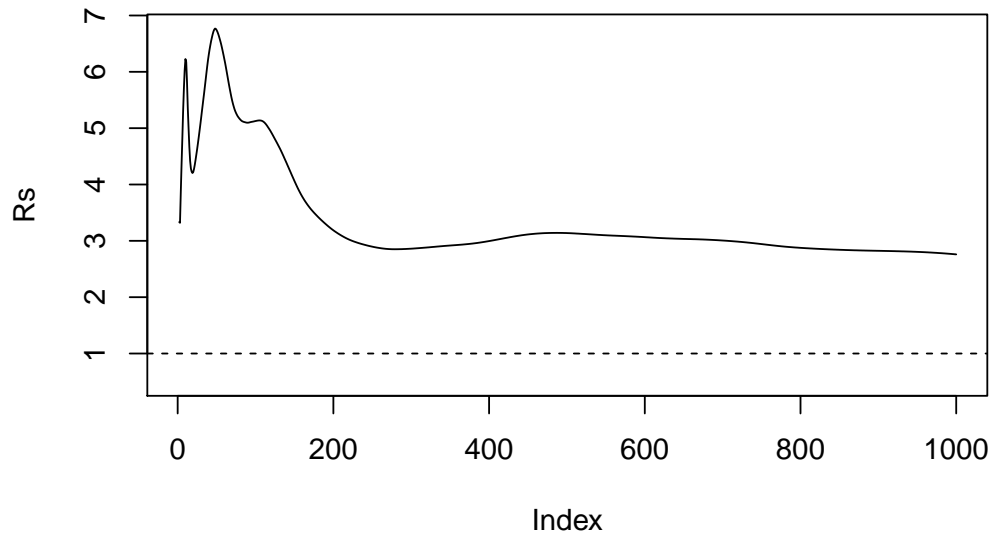
Conversely, W (on its own) **underestimates** the variance, but converges to the true variance.

This suggests:

$$\hat{R} = \sqrt{\frac{V}{W}} \approx 1 \iff \text{the chains have converged}$$

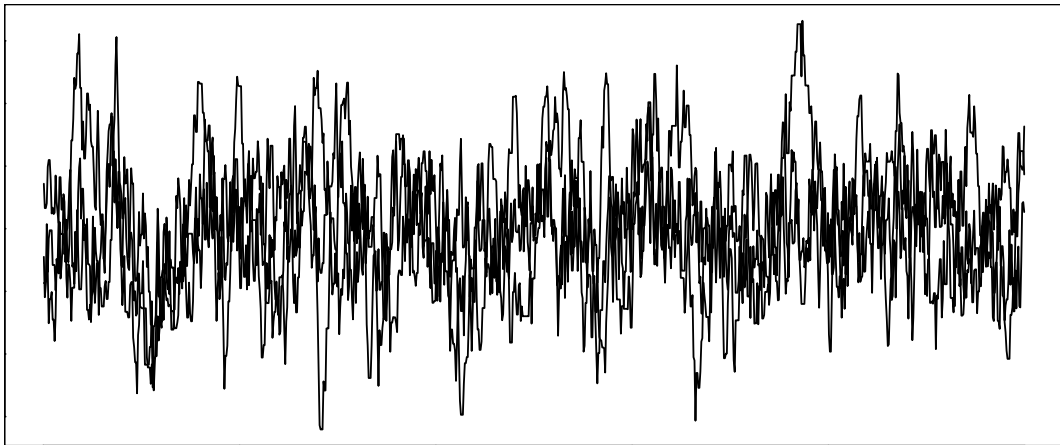
Computing \hat{R}

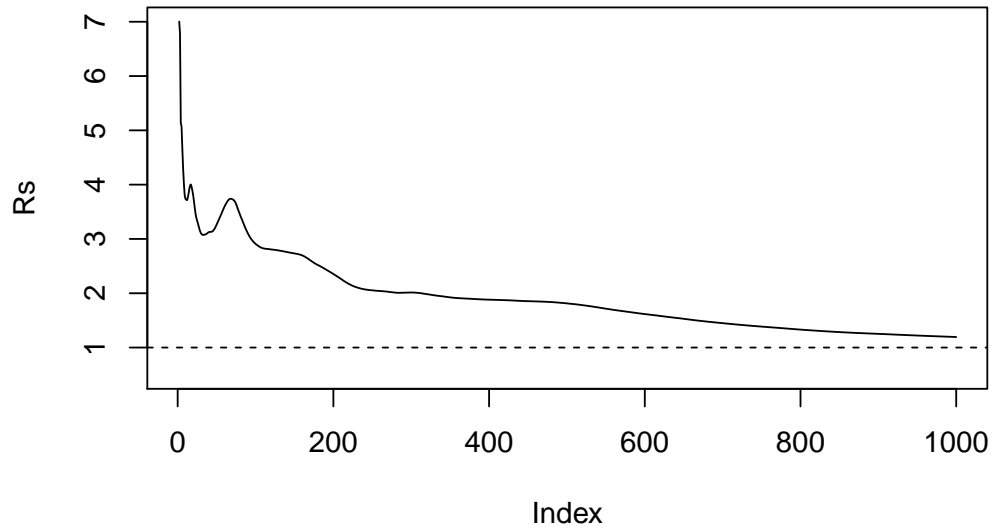
```
> Rs <- numeric(1000)
> for (t in 1:1000) {
+   p1 <- psi_1[1:t] ; p2 <- psi_2[1:t] ; p3 <- psi_3[1:t]
+   psiBars <- c(mean(p1), mean(p2), mean(p3))
+   psiBarBar <- mean(c(p1, p2, p3))
+
+   B <- (t / 2) * sum((psiBars - psiBarBar)^2)
+   W = (1 / (2 * t)) * (sum((p1 - psiBars[1])^2) +
+                        sum((p2 - psiBars[2])^2) +
+                        sum((p2 - psiBars[2])^2))
+   Rs[t] <- sqrt(((t - 1) / t * W + 1 / t * B) / W)
+ }
```



Repeat with better mixing

```
> chains_1 <- mclapply(rep(1, 3), unif_chain, mc.cores = 3, B = 1000)
```





Monitoring Convergence and Stan

Computing the \hat{R} values can be somewhat tedious. Luckily, **Stan will compute them for us:**

```
> summary(bp_stan)$summary[, c("mean", "sd", "n_eff", "Rhat")]
```

	mean	sd	n_eff	Rhat
mu	127.6	0.3149	3644	1.0006
sigmasq	358.6	8.3742	3086	0.9998
lp__	-12243.4	0.9593	2055	1.0019