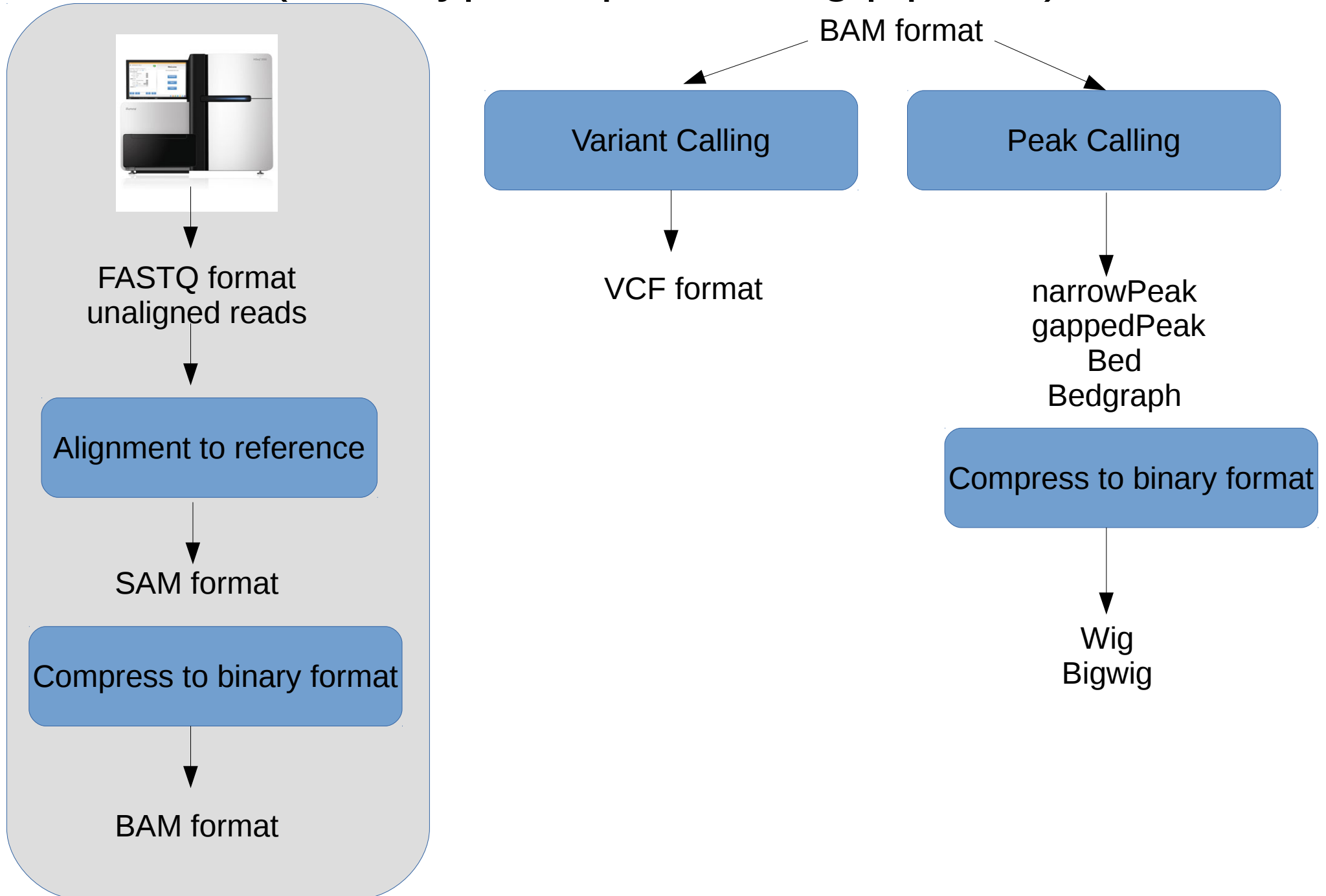


Data Formats Cheat Sheet

(for a “typical” processing pipeline)





FASTQ format
unaligned reads

Alignment to reference

SAM format

Compress to binary format

BAM format

BAM format

Variant Calling

VCF format

Peak Calling

NarrowPeak
Bed
Bedgraph

Compress to binary format

Wig
Bigwig

FASTA and FASTQ Files

FASTQ FORMAT:

[illegible]

- Read name
 - Preceded by @
 - Unique identifier for each read in the dataset
- Base sequence (A,T,C,G,N)
- + sign
- Phred quality score
 - ASCII encoding

Phred quality scores Q are defined as a property which is logarithmically related to the base-calling error probabilities P .¹

$$Q = -10 \log_{10} P$$

or

$$P = 10^{\frac{-Q}{10}}$$

FASTA and FASTQ Files

FASTA FORMAT:

```
>J00118:203:HFL3VBBXX:4:1101:23399:1349 1:N:0:TAAGGCGA+TCTACTCT
GNTCTAGGGTGTAGCCTGAGAATAGGGGAAATCAGTGAATGCTGTCTCTTATACACATCTCCGAGCCCACGAGACT
>J00118:203:HFL3VBBXX:4:1101:24292:1349 1:N:0:TAAGGCGA+TCTACTCT
GNCTGAGAGGGCCCCTGTTAGGGGTCATGGGCTGGGTTTACTATATGATAGGCATGTGATTGGTGGGTCATTATG
```

- Read name
 - Preceded by >
 - Unique identifier for each read in the dataset
- Base sequence (A,T,C,G,N)



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SAM and BAM format

- Alignment algorithms such as BWA and Bowtie2 will accept input data in FASTA format and will generate aligned output in SAM or BAM format.
- BAM format stores data in a compressed, indexed, binary format.
- SAM is the human readable version of BAM – **never store data in SAM format! It is much larger than BAM and contains the same information.**
- Use the **samtools** program to work with bam files – sort, index, look for variants, and so much more
 - **<http://www.htslib.org/doc/samtools.html>**

Samtools is your friend!

(essential commands for viewing/working with (s/b)am files)

```
samtools view -bt ref_list.txt -o aln.bam aln.sam.gz
```

```
samtools sort -T /tmp/aln.sorted -o aln.sorted.bam aln.bam
```

```
samtools index aln.sorted.bam
```

```
samtools stats aln.sorted.bam
```

```
samtools bedcov aln.sorted.bam
```

```
samtools depth aln.sorted.bam
```

```
samtools view aln.sorted.bam chr2:20,100,000-20,200,000
```

So what's actually in a bam (sam) file?

“Row” for a single read:

```
J00118:203:HFL3VBBXX:4:2205:13626:34495 Read name  
99          a bitwise set of information describing the alignment, FLAG  
chr10       Chromosome where the read aligned  
60573      Starting position of alignment  
1           Phred-scaled quality score  
76M  
=  
60621  
124  
GTTATTAGATGATTCAAATATGAAGTGCTGTTATGCCAAACAATGAATCTTGTGTTATACA Sequence  
AafffJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJ Quality  
AS:i:-6  
XS:i:-6  
XN:i:0  
XM:i:1  
XO:i:0  
XG:i:0  
NM:i:1  
MD:Z:24A51  
YS:i:0  
YT:Z:CP
```




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VCF (Variant Call Format)

```
##fileformat=VCFv4.1
##fileDate=20130207
##source=GenerateReportDataAndVCFv2.2.0.0
##reference=HumanNCBI37_UCSC
##phasing=none
##INFO=<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO=<ID=TI,Number=.,Type=String,Description="Transcript ID">
##INFO=<ID=GI,Number=.,Type=String,Description="Gene ID">
##INFO=<ID=EXON,Number=0,Type=Flag,Description="Exon Region">
##INFO=<ID=FC,Number=.,Type=String,Description="Functional Consequence">
##FILTER=<ID=q20,Description="Quality below 20">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT VACO_576_MAXGT VACO_576_POLY
chr1 10291 . C T 3 q20 DP=4 GT:GQ 0/0:3 0/1:31
chr1 10327 . T C 46 PASS DP=9 GT:GQ 1/0:45 1/0:51
chr1 10552 . G A 5 q20 DP=2 GT:GQ 1/0:5 1/0:32
chr1 14907 rs6682375;rs79585140 A G 1 q20 DP=2;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 0/0:6 0/1:26
chr1 14930 rs6682385;rs75454623 A G 4 q20 DP=2;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 0/1:4 0/1:31
chr1 15190 rs71230572 G A 5 q20 DP=1;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/0:2 1/0:3
chr1 15211 rs11586607;rs78601809 T G 10 q20 DP=1;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/0:3 1/0:3
chr1 15817 rs2691316;rs78436736 G T 1 q20 DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON GT:GQ 0/0:9 0/1:24
chr1 15820 rs2691315;rs75570658 G T 10 q20 DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON GT:GQ 0/1:10 0/1:41
chr1 16014 rs75082847;rs80035579 C T 5 q20 DP=2;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/1:2 1/1:4
chr1 16068 rs79696773 T C 11 q20 DP=2;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/1:3 1/1:4
chr1 16103 rs76959363;rs78376469 T G 67 PASS DP=3;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/1:7 1/1:7
chr1 17222 rs2981830;rs62530147;rs80270096 A G 32 PASS DP=6;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 0/1:32 0/1:56
chr1 17407 . G A 1 q20 DP=3;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 0/0:6 1/0:27
chr1 17538 rs71260068 C A 52 PASS DP=11;TI=NR_024540;GI=WASH7P;FC=Silent GT:GQ 1/0:52 1/0:85
chr1 17626 rs11555814;rs77744836 G A 2 q20 DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON GT:GQ 0/0:5 1/0:29
```

Header information

↑ position
↑ chromosome
↑ variant id (if known)
↑ Reference allele
↑ Alternate allele
↑ Quality score
↑ Quality threshold used to filter variants
↑ Depth – how many reads aligned to the position
↑ Functional consequence
↑ Transcript ID/Gene ID

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```
##fileformat=VCFv4.1
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##source=GenerateReportDataAndVCFv2.2.0.0
##reference=HumanNCBI37_UCSC
##phasing=none
##INFO=<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO=<ID=TI,Number=.,Type=String,Description="Transcript ID">
##INFO=<ID=GI,Number=.,Type=String,Description="Gene ID">
##INFO=<ID=EXON,Number=0,Type=Flag,Description="Exon Region">
##INFO=<ID=FC,Number=.,Type=String,Description="Functional Consequence">
##FILTER=<ID=q20,Description="Quality below 20">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
```

Header information

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	VACO_576_MAXGT	VACO_576_POLY
chr1	10291	.	C	T	3	q20	DP=4	GT:GQ	0/0:3	0/1:31
chr1	10327	.	T	C	46	PASS	DP=9	GT:GQ	1/0:45	1/0:51
chr1	10552	.	G	A	5	q20	DP=2	GT:GQ	1/0:5	1/0:32
chr1	14907	rs6682375;rs79585140	A	G	1	q20	DP=2;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	0/0:6	0/1:26
chr1	14930	rs6682385;rs75454623	A	G	4	q20	DP=2;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	0/1:4	0/1:31
chr1	15190	rs71230572	G	A	5	q20	DP=1;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/0:2	1/0:3
chr1	15211	rs11586607;rs78601809	T	G	10	q20	DP=1;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/0:3	1/0:3
chr1	15817	rs2691316;rs78436736	G	T	1	q20	DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON	GT:GQ	0/0:9	0/1:24
chr1	15820	rs2691315;rs75570658	G	T	10	q20	DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON	GT:GQ	0/1:10	0/1:41
chr1	16014	rs75082847;rs80035579	C	T	5	q20	DP=2;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/1:2	1/1:4
chr1	16068	rs79696773	T	C	11	q20	DP=2;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/1:3	1/1:4
chr1	16103	rs76959363;rs78376469	T	G	67	PASS	DP=3;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/1:7	1/1:7
chr1	17222	rs2981830;rs62530147;rs80270096	A	G	32	PASS	DP=6;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	0/1:32	0/1:56
chr1	17407	.	G	A	1	q20	DP=3;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	0/0:6	1/0:27
chr1	17538	rs71260068	C	A	52	PASS	DP=11;TI=NR_024540;GI=WASH7P;FC=Silent	GT:GQ	1/0:52	1/0:85
chr1	17626	rs11555814;rs77744836	G	A	2	q20	DP=4;TI=NR_024540;GI=WASH7P;FC=Silent;EXON	GT:GQ	0/0:5	1/0:29

Genotype

0/0 → 2 reference alleles

0/1 → 1 reference allele, 1 alternate allele

1/1 → 2 alternate alleles

MAXGT – genotype compared to reference sequence (i.e. hg19)

POLY – genotype for a polymorphic site



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BED files (and variations)

Bed files denote genome regions/ positions of interest

chr1	9998	10674	
chr1	17378	17588	
chr1	235560	235770	
chr1	237617	237892	
chr1	521429	521692	
chr1	545957	546238	
chr1	564497	564727	
chr1	569808	570050	
chr1	713705	714679	
chr1	740197	740406	
chr1	753380	753554	
chr1	755417	756332	

Chromosome
Start position
End position

Bedgraph files are typically used to denote genome coverage in an added 4th column

chr1	40186	40187	0.00106931
chr1	40237	40238	0.0011158
chr1	40404	40405	0.00181824
chr1	40406	40407	0.00157195
chr1	40407	40408	0.00169206
chr1	40408	40409	0.00145328
chr1	40409	40410	0.00191194
chr1	40437	40438	0.00174463
chr1	40444	40445	0.00324619
chr1	40445	40446	0.00434136



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Browser Track Formats: BigWig and Hammock

- Allow for visualization of large datasets in a browser such as UCSC or Washu
- BigWigs are binary files
- Hammock tracks are human-readable
- Washu browser demo

