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Spatial Transcriptomic Data

2022-01-09



CONTENTS

- 01 **Introduction of STD**
- 02 **ClusterMap**
- 03 **Summary and weekly plan**

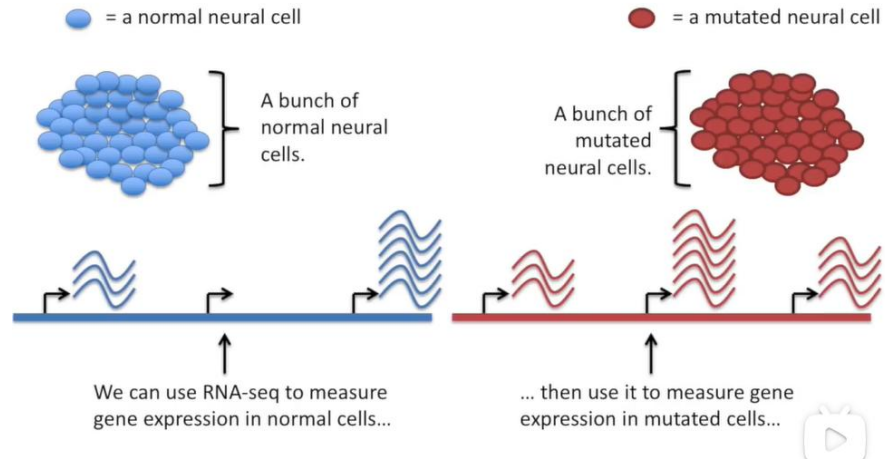




01

Introduction of STD

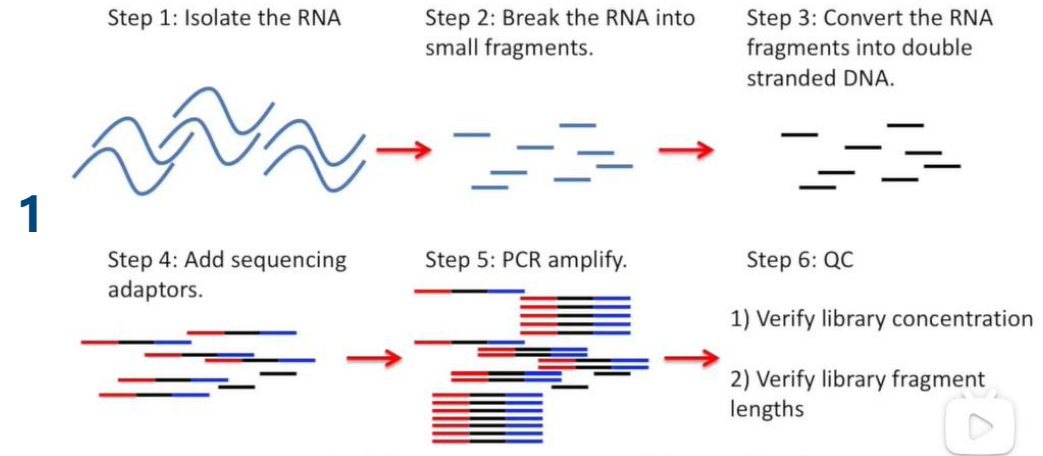
RNA-seq



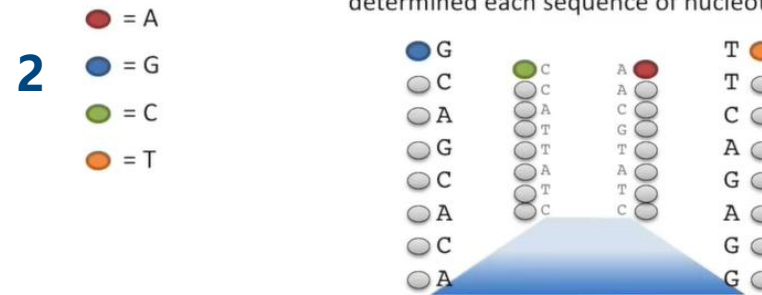
3 main steps for RNA-seq:

1. prepare a sequencing library
2. sequence
3. data analysis

Preparing an RNA-seq library



And the process repeats until the machine has determined each sequence of nucleotides.



3

```
@NS500177:196:HFTTTAFXX:1:11101:10916:1458 2:N:0:CGCGGCTG
ACACGACGATGAGGTGACAGTCACGGAGGATAAGATCAATGCCCTCATTAAGCAGCCGGTGTA
+
AAAAEEEEEEEEEEEEEE//AEEEEEEEEEEEEEEEE/EE/<<EE/AEEEEEE//EEEEEEEEEEA<
```

index; sequencing; ' + ' ; quality score

RNA-seq



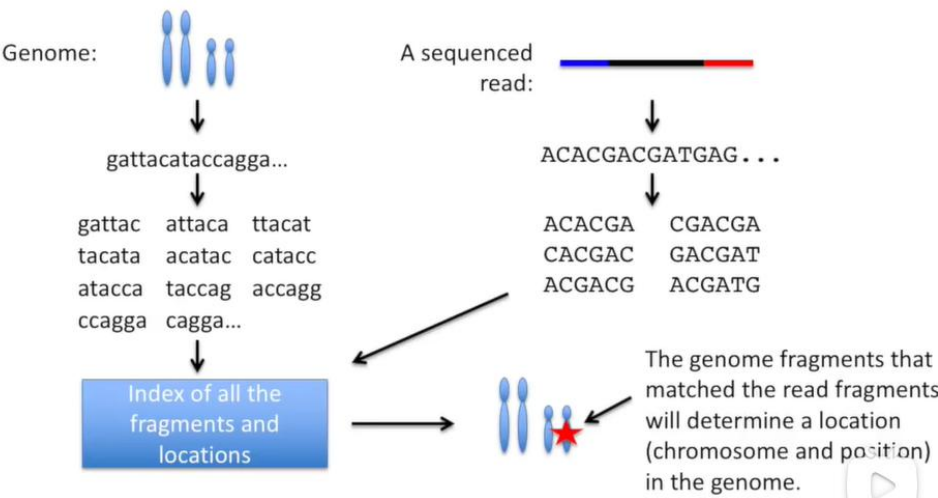
Preprocessing

Filter out garbage reads

Garbage reads are:

1. Reads with low quality base calls
2. Reads that are clearly artifacts of chemistry

Align the high quality reads to a genome

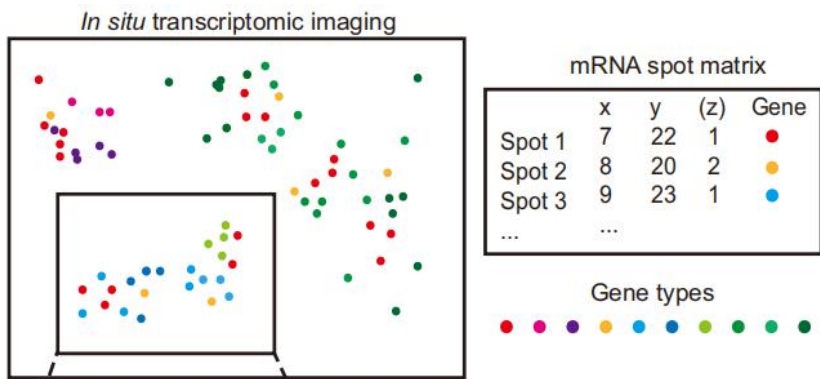


Count the number of reads per gene

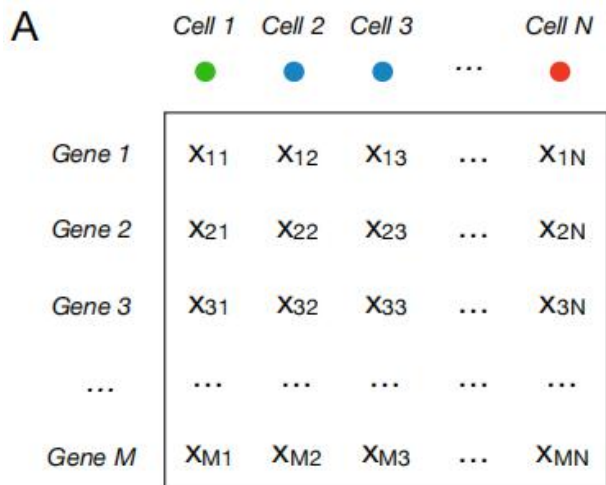
Gene	Sample1	Sample2	Sample3...
A1BG	30	5	13...
A1BG-AS1	24	10	18...
A1CF	0	0	0...
A2M	5	9	7...
A2M-AS1	3563	5771	4123...
A2ML1	13	8	7...
...



<https://doi.org/10.1038/s41467-021-26044-x>



Gene type & Coordinate



How to get 2D(3D) coordinate:

1. spatial barcoding
2. in situ hybridization
3. in situ sequencing

Method	Type	Resolution	Genes	Reference
Visium	Spatial barcoding	55μm	Whole transcriptome	(16)
Slide-seq	Spatial barcoding	10μm	Whole transcriptome	(17, 18)
HDST	Spatial barcoding	2μm	Whole transcriptome	(19)
DBiT-Seq	Spatial barcoding	10μm	Whole transcriptome	(20)
Seq-scope	Spatial barcoding	0.6μm	Whole transcriptome	(21)
Stereo-seq	Spatial barcoding	500nm	Whole transcriptome	(22)
seqFISH	<i>in situ</i> hybridization	single-molecule	>10,000	(23, 24)
merFISH	<i>in situ</i> hybridization	single-molecule	100 – 1,000	(25, 26)
STARmap	<i>in situ</i> sequencing	single-cell	160 – 1020	(27)
FISSEQ	<i>in situ</i> sequencing	subcellular	~ 8000	(28)

<http://arxiv.org/abs/2110.07787>



STD: Data analysis



data filter

1. genes and cells may be filtered based on threshold specific to the dataset.

2. gene expression per cell may be normalized to have the same total library size so that expression levels are comparable across cells.

analysis and visualization in expression domain

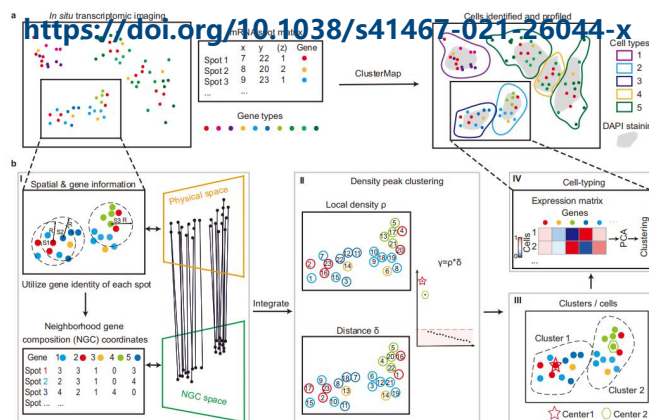
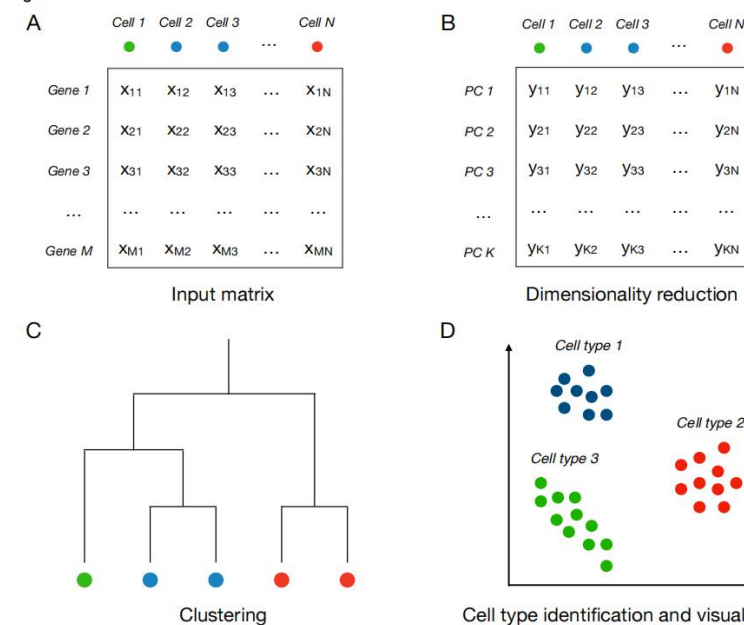
A first step in the spatial transcriptomic analysis is to identify the cell type (for datasets of single-cell resolution) or cell mixture (for datasets of multicellular resolution) of each spatial unit or spot. Cell type identification usually starts with dimensionality reduction technique to reduce time and space complexity for downstream analysis. The reduced representations are used to cluster cells based on the assumption that cell of the same type falls into the same cluster.

Clustering

Agglomerative clustering : a class of methods that iteratively aggregate data points into clusters.

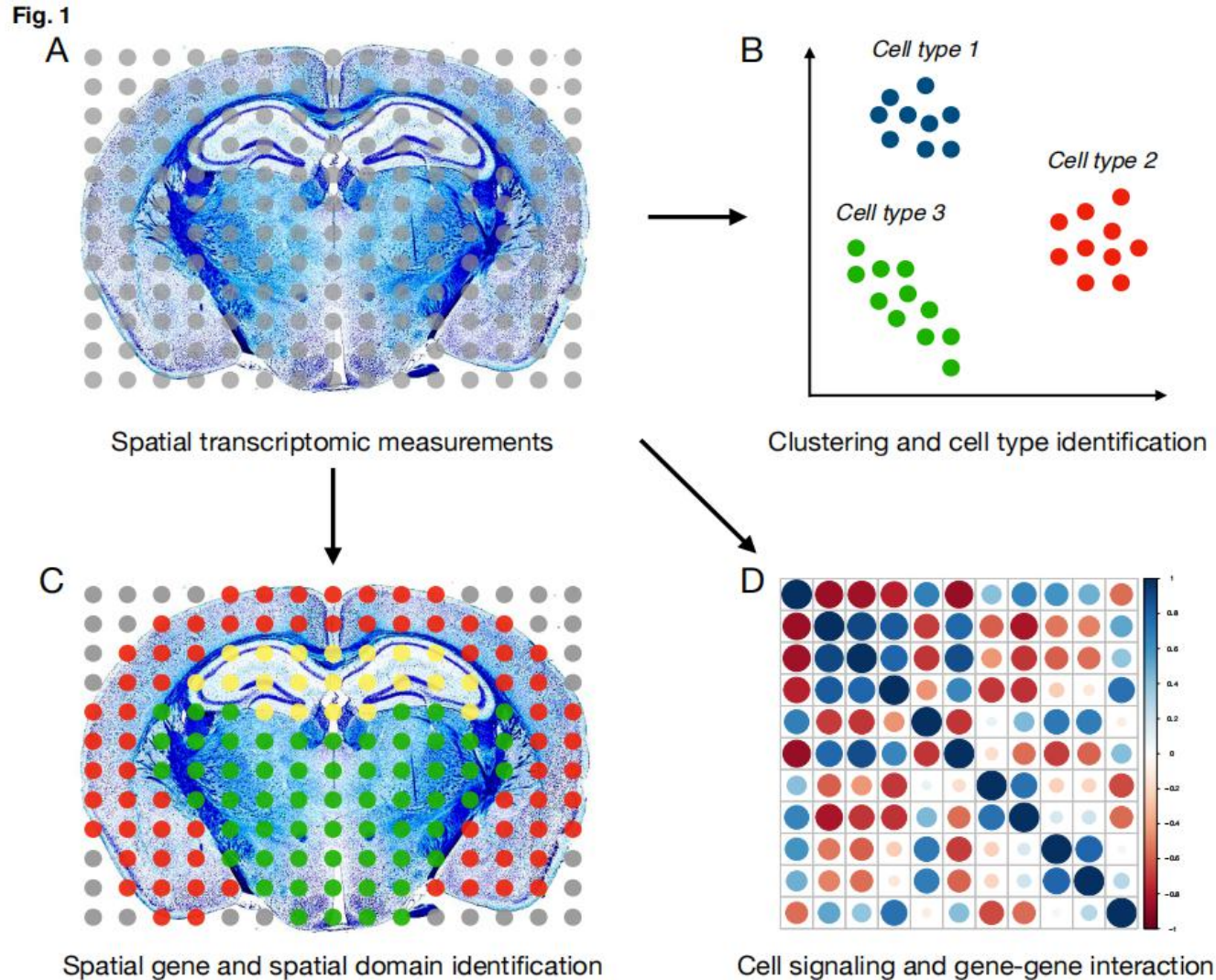
Identification of cell types

Fig. 2



<https://doi.org/10.1038/s41467-021-26044-x>

STD: Data analysis



Data analysis:

1. Clustering and cell type identification
2. Spatial gene and spatial domain identification
3. Cell signaling and gene-gene interaction

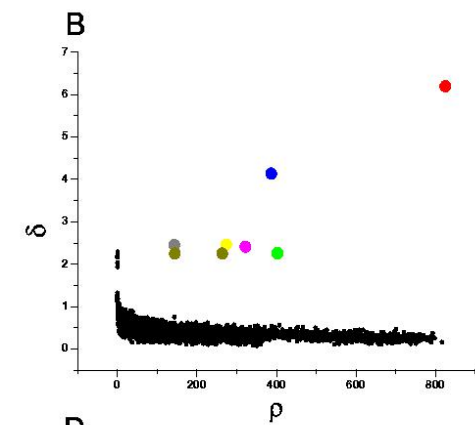
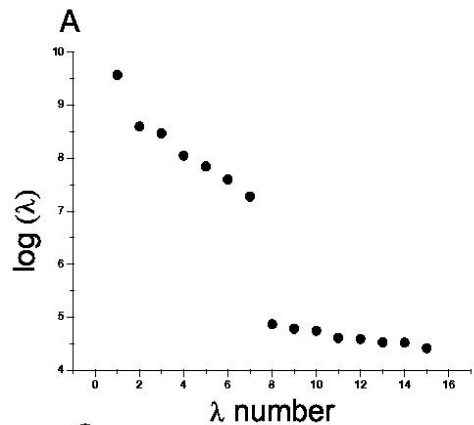


02

ClusterMap



Density peak clustering (DPC)



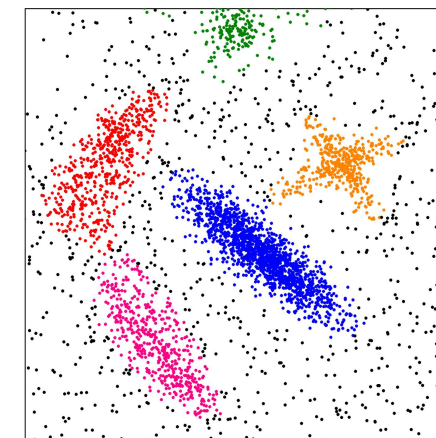
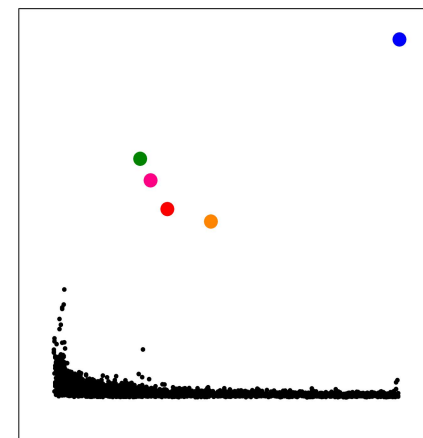
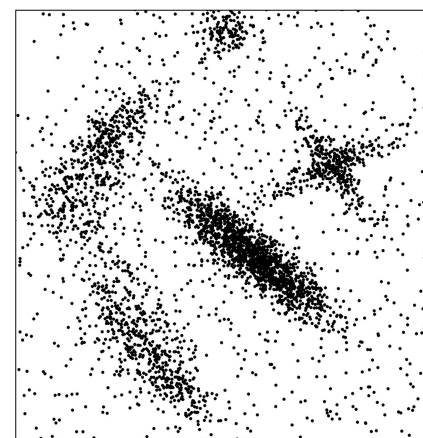
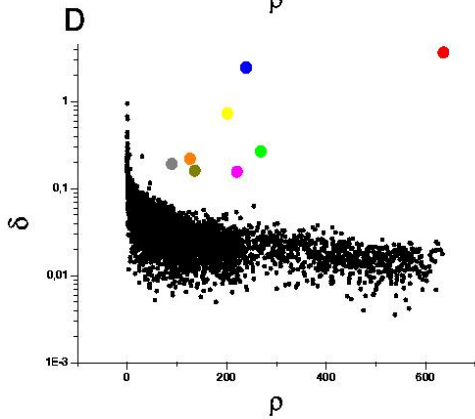
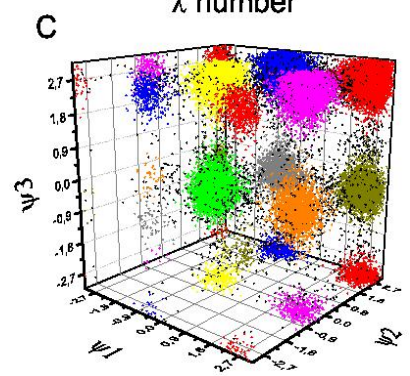
When applied to Molecular Dynamics trajectories, the results are coherent with a much more complex kinetic model, even when employing different similarity measures like RMSD or Dihedral distance.

k-NN by ρ

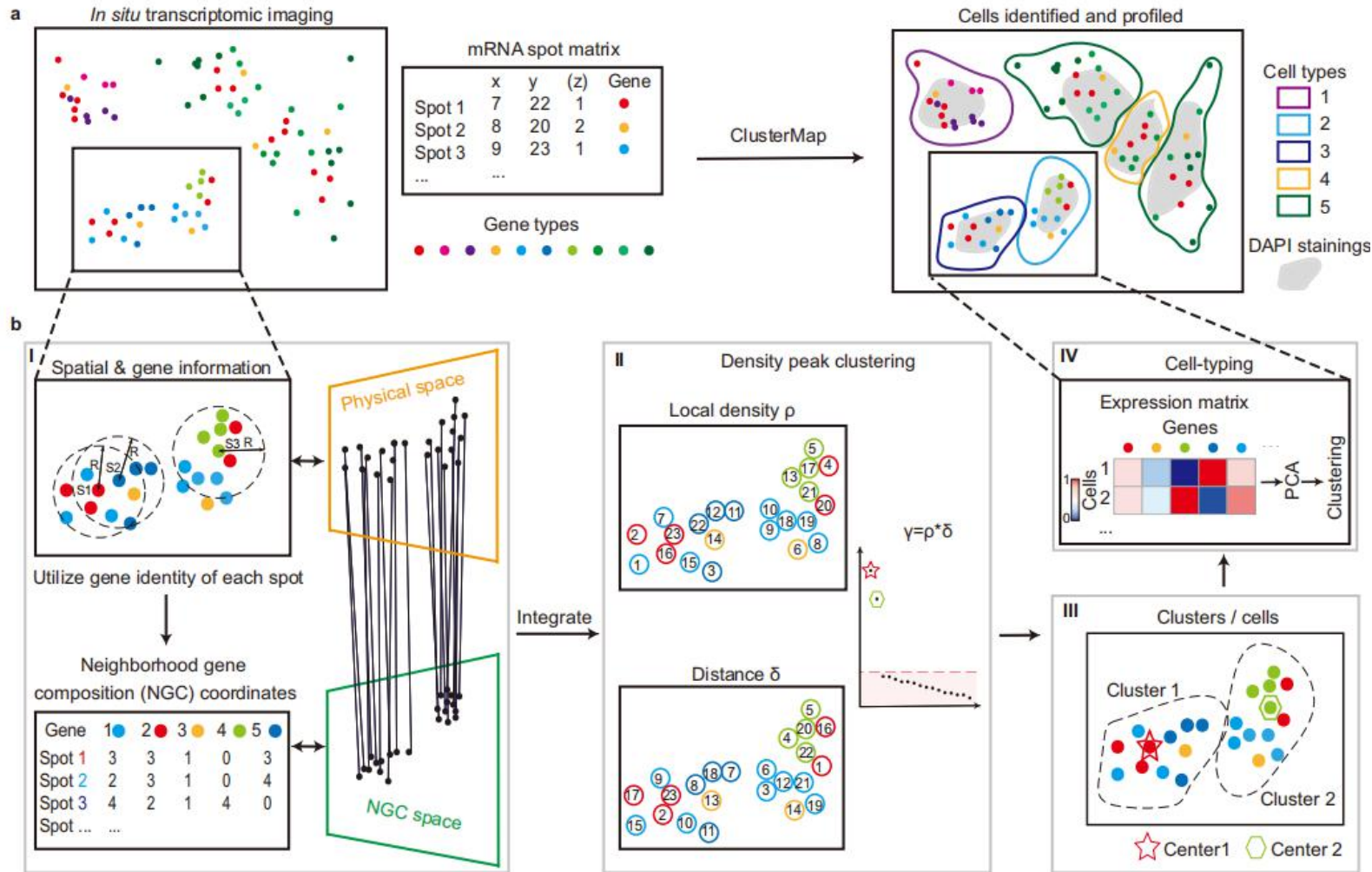
$$\rho_i = \sum \chi(d_{ij} - d_c)$$

$$\chi(x) = \begin{cases} 1 & \text{if } x < 0 \\ 0 & \text{if otherwise} \end{cases}$$

$$\delta_i = \min_{j: \rho_j > \rho_i} (d_{ij})$$



ClusterMap



Integration of the physical and NGC coordinate

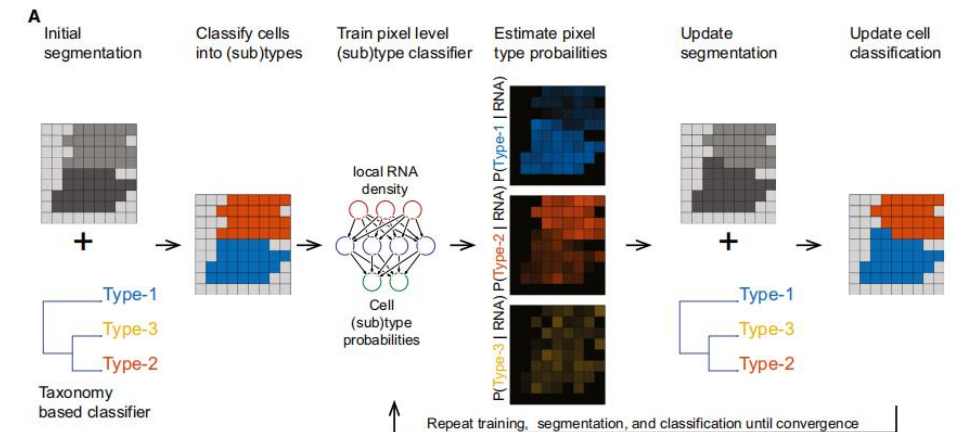
$$P(i) = \langle x_i, y_i, (z_i) \rangle \quad (6)$$

$$NGC(i) = \langle Num_{Gene\ 1}, Num_{Gene\ 2}, \dots, Num_{Gene\ t}, \dots, Num_{Gene\ T} \rangle \quad (7)$$

$$d_{ij} = \frac{Distance\{P(i), P(j)\}}{SpearmanCorr\{NGC(i), NGC(j)\}}$$

$$\rho_i = \sum_j I(d_{ij} - d_{max}) * e^{-(d_{ij}/R)^2} \quad (4)$$

$$\delta_i = \min(d_{ij}), j: \rho_j > \rho_i \quad (5)$$



<https://github.com/wanglab-broad/ClusterMap>

<https://doi.org/10.15252/msb.202010108>

<https://doi.org/10.1038/s41467-021-26044-x>