

Rainbow: Automated Air-Liquid Interface Cell Culture Analysis Using Deep Optical Flow

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Summary

Rainbow is a free, open-source and cross-platform Python-based tool for Air-Liquid Interface (ALI) cell culture image sequence analysis. Rainbow accepts input image sequences in standard image formats, such as TIFF, or microscopy file formats, such as ND2, and then computes the optical flow, that is: the apparent motion of individual pixels in an image ([Beauchemin & Barron, 1995](#); [Turaga et al., 2010](#); [Zhai et al., 2021](#)), across multiple frames using a deep learning based pretrained optical flow model ([Jiang et al., 2021](#)). Rainbow then uses the pixel-level optical flow information and applies circular data analysis to calculate the average magnitude $[0, \infty]$ (m) and direction $[0, 360]$ (°) of motion between adjacent frames to quantify cell motility. Additionally, the variance of the magnitude $[0, \infty]$ (m) and direction $[0, 360]$ (°) of motion between adjacent frames is calculated to quantitatively capture the degree of heterogeneity in cell motility.

For each experiment, a CSV file with minimum, maximum, mean, standard deviation and variance values of the magnitude and direction of cell movement between adjacent frames in an image sequence is produced. Multiple CSV files from different experiments can be combined into one file that can be analysed for differences in cell motility across experiments. Rainbow also includes a high-resolution and easily readable unified hue/saturation-based visualisation scheme for the instantaneous vector field of motion between adjacent frames of an image sequence to qualitatively show cell motion. Rainbow can be used through a graphical user interface or command line interface and can generate a HTML report containing output images, videos, publication ready figures, and CSV files detailing cell dynamics (refer to Examples folder on GitHub) in one command. Importantly, our software is not limited to ALI culture image analysis, and developers can extend the software's existing pipeline to other use cases. For example, the optical flow model can be readily substituted with different models, as we utilised the Factory Method creational software design pattern. The data analyses and report generated can be adjusted through interactive Jupyter Notebooks, allowing for a flexible and versatile system. Some of Rainbow's visualizations are shown in [Figure 1](#).

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39 Statement of need

40 Assessment of airway epithelial cell (AEC) function in disease utilise in vitro models such as
41 the ALI cell culture (Chen & Schoen, 2019; Looi et al., 2018; Martinovich et al., 2017). The
42 integration of image analysis and ALI cultures has provided novel insights into cell dynam-
43 ics, such as the recently identified unjammed-to-jammed transition of AEC characterised by
44 changes in cell motility (Mitchel et al., 2020; Park et al., 2015). In chronic respiratory diseases
45 like asthma, increased cell motility and AEC unjamming have been linked to airway remod-
46 elling and disease development (Mitchel et al., 2020; Park et al., 2015). However, the image
47 analyses performed in these studies are limited. For example, handcrafted methods from the
48 MATLAB Computer Vision Toolbox that compute optical flow have been used to extract cell
49 motion information from ALI culture images (Mitchel et al., 2020). This approach requires
50 licenced software and handcrafted optical flow estimation methods have been outperformed
51 in terms of accuracy by deep learning methods (Savian et al., 2020). Furthermore, commonly
52 used cell motion metrics, such as average cell speed, do not capture all unique aspects of
53 cell motion, such as the heterogeneity of cell migration patterns across time. Cell motion is
54 commonly visualised using vector fields, which are useful but bound by an inverse relationship
55 between resolution and readability (Henkes et al., 2020; Nnetu et al., 2012; O'Sullivan et al.,
56 2020).

57 To increase understanding of lung disease mechanisms and development of new treatment
58 options for patients, there is a need for open-source solutions for ALI culture image analyses
59 that can be broadly implemented across cell biology laboratories. Rainbow is the first single
60 easy-to-use tool that performs all the above analyses automatically for efficient utilisation by
61 non-programmers. Rainbow produces automatic cell motion quantifications, figures, and a
62 report that is easily transferrable into publications. We anticipate that Rainbow will provide
63 cell motion characterization for each experiment and allow for easy comparisons among mul-
64 tiple experiments to uncover cellular migration mechanisms previously undetermined in health
65 and disease.

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66 **Figures**

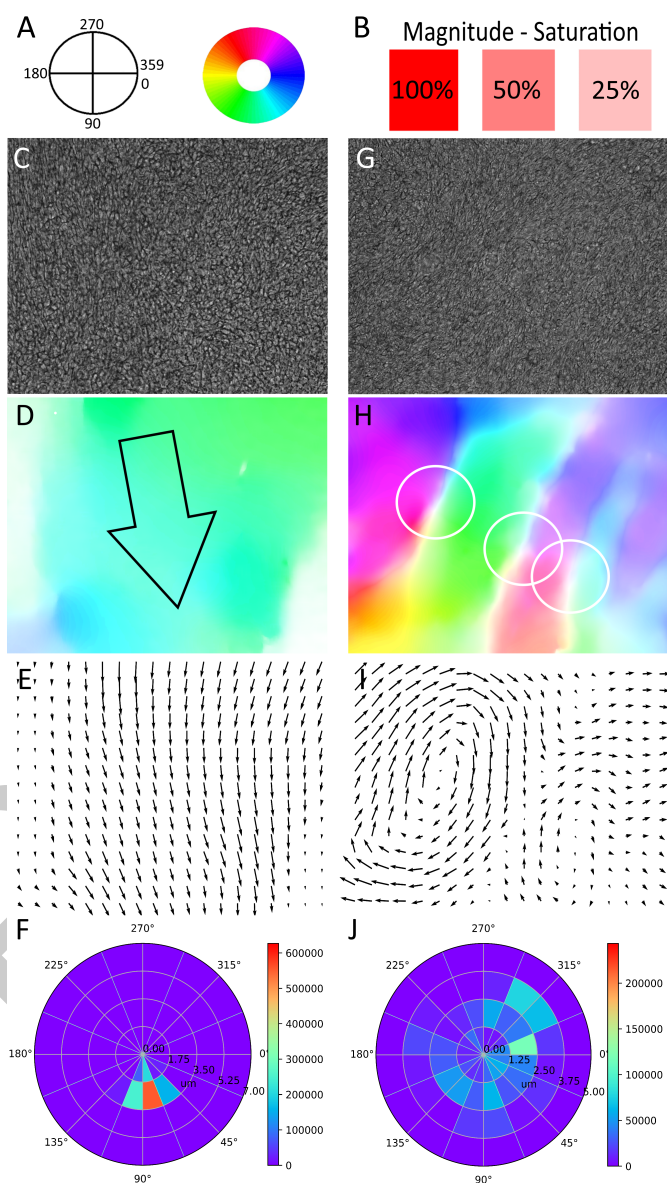


Figure 1: Rainbow optical flow visualisation. **A:** The direction of motion at any position within Rainbow generated optical flow images is measured clockwise from the initial horizontal position of a unit circle (left) and is shown using hue values (right). **B:** The magnitude of motion at any position within optical flow images is shown using saturation values. High saturation (100%) corresponds to high magnitude of motion and low saturation (25%) corresponds to low magnitude of motion. **C, G:** Still frames taken from two separate ALI culture image sequences. **D, H:** Unified visualisation of optical flow magnitude and direction between adjacent frames of two ALI culture image sequences using Rainbow. The arrow indicates the average direction of motion across the image sequence. The circles indicate three localised vortexes that the cells move around in a swirl-like motion as they change direction. **E, I:** Traditional visualisation of optical flow between adjacent frames of two ALI culture image sequences using quiver plots containing vector arrows at every 70 px. **F, J:** Polar plots visualising motion magnitude (concentric circles; μm) and direction (azimuthal angle; degrees) in the same frame of two ALI culture image sequences. Colour scale indicates the number of points migrating towards given direction. All left positioned subfigures from row 2 onwards correspond to the same ALI culture image sequence while right positioned subfigures correspond to a different image sequence. For complete insight, refer to Examples folder on GitHub containing videos.

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