

**Supplementary Table 1: Effects of Gene-deficiency on Tfh cell formation**

	Gene/Protein	Effect of ko/mutation on Tfh cells	Comment/potential or actual mechanism	References
<b>Cytokines</b>	IL-6	<ul style="list-style-type: none"> <li>• mild/moderate decrease</li> <li>• no effect</li> </ul>	<ul style="list-style-type: none"> <li>• variable results probably reflects different immunisations and infections (eg acute vs chronic LCMV; influenza)</li> <li>• combined deficiency of IL-6 and IL-21 reduced Tfh numbers <i>in vivo</i>, consistent with their ability to induce Bcl-6 and c-Maf</li> </ul>	1-6
	IL-21, IL-21R	<ul style="list-style-type: none"> <li>• mild decrease</li> <li>• no effect</li> </ul>		1,3-5,7-11
	IL-27Ra	<ul style="list-style-type: none"> <li>• decreased</li> </ul>	<ul style="list-style-type: none"> <li>• induces c-Maf, ICOS, IL-21</li> <li>• promotes Tfh cell survival</li> </ul>	12,13
	IL-2	<ul style="list-style-type: none"> <li>• increased</li> </ul>	<ul style="list-style-type: none"> <li>• induces Blimp-1 which suppresses Tfh commitment</li> </ul>	14,15
	IL-10/IL-10R	<ul style="list-style-type: none"> <li>• increased</li> </ul>	<ul style="list-style-type: none"> <li>• IL-10 suppresses Tfh formation directly by acting on naïve CD4<sup>+</sup> T cells but also indirectly by suppressing production of cytokines (IL-6, IL-23) by DC that may promote Tfh formation. Molecular mechanism of inhibitory action of IL-10 is unknown.</li> </ul>	16-18
<b>Signalling</b>	SH2D1A (SAP)	<ul style="list-style-type: none"> <li>• decreased/non-functional</li> </ul>	<ul style="list-style-type: none"> <li>• requirement for SAP in Tfh cell can be overcome by providing excess Ag to circumvent the dependency on B cells as the primary APC</li> <li>• however SAP-deficient Tfh cells are still unable to “help” cognate B cells due to an inability to form stable T-B conjugates</li> <li>• mutations in SH2D1A results in impaired Ab production in humans due to a CD4<sup>+</sup> T cell intrinsic defect</li> </ul>	19-24
	STAT1	<ul style="list-style-type: none"> <li>• transient decrease</li> </ul>	<ul style="list-style-type: none"> <li>• functions downstream of IL-6 receptors; greater defect in Tfh formation when naïve CD4<sup>+</sup> T cells lack both STAT1 and STAT3</li> </ul>	6
	STAT3	<ul style="list-style-type: none"> <li>• decreased</li> </ul>	<ul style="list-style-type: none"> <li>• role of STAT3 probably accounts for redundancy in the effects of IL-6 or IL-21 deficiency on Tfh formation</li> <li>• mutations in STAT3 results in a deficiency in production of Ag-specific Ab, memory B cells and circulating CD4<sup>+</sup>CXCR5<sup>+</sup> T cells in humans; STAT3 probably acts downstream of IL-6, IL-21 as well as IL-12 in human CD4<sup>+</sup> T cells to induce Tfh cells</li> </ul>	1,25,26
	STAT4	<ul style="list-style-type: none"> <li>• transient decrease</li> </ul>	<ul style="list-style-type: none"> <li>• IL-12/STAT4 signalling induced T-bet which suppresses Bcl-6 function</li> </ul>	27
	STAT5	<ul style="list-style-type: none"> <li>• increased</li> </ul>	<ul style="list-style-type: none"> <li>• directly binds Bcl6 to inhibit expression</li> <li>• activated by IL-2, which induces Blimp-1 to suppress Tfh commitment</li> </ul>	14
	NIK (NFkB inducible kinase)	<ul style="list-style-type: none"> <li>• decreased</li> </ul>	<ul style="list-style-type: none"> <li>• NIK-dependent signaling downstream of BAFF-R on B cells is required for constitutive expression of ICOS-L on these cells, which is required for Tfh formation</li> </ul>	28
	Roquin	<ul style="list-style-type: none"> <li>• increased</li> </ul>	<ul style="list-style-type: none"> <li>• mutation in <i>san roqin</i> gene causes severe lupus-like disease due to aberrant Tfh accumulation and subsequent formation of GCs and autoAb production</li> </ul>	29
<b>Transcription factors</b>	Bcl-6	<ul style="list-style-type: none"> <li>• absent</li> </ul>	<ul style="list-style-type: none"> <li>• required for Tfh formation</li> <li>• inhibits expression/function of T-bet, Gata3, RORγt and generation of Th1/2/17 cells</li> <li>• suppresses expression of clusters of miRNAs that inhibit Tfh formation</li> <li>• considered the “master regulator” of Tfh cells, but likely to require actions of other transcription factors to induce Tfh cells</li> </ul>	4,30-32

## SUPPLEMENTARY INFORMATION

	c-Maf	• decreased	<ul style="list-style-type: none"> <li>• induces expression of IL-21, ICOS</li> <li>• co-operates with Bcl-6 to induce/enhance expression of some key features of Tfh cells</li> </ul>	13,33-35
	BATF	• absent	<ul style="list-style-type: none"> <li>• functions by inducing Bcl-6 and c-Maf</li> <li>• additional BATF-targets also likely to be involved in Tfh formation</li> </ul>	36,37
	IRF4	• decreased/absent	<ul style="list-style-type: none"> <li>• co-operates with STAT3 to mediate IL-21-induce gene expression in CD4<sup>+</sup> T cells, however its exact role in Tfh cells remains unknown</li> </ul>	38
	<i>PRDM1</i> (Blimp1)	• increased	<ul style="list-style-type: none"> <li>• Blimp-1 restricts Tfh formation by antagonising Bcl-6 function</li> </ul>	30
	<i>Tbx21</i> (T-bet)	• increased	<ul style="list-style-type: none"> <li>• T-bet suppresses Bcl-6 function</li> </ul>	27,39
<b>Surface receptors</b>	CD40/CD40L	• decreased/absent	<ul style="list-style-type: none"> <li>• mutations in <i>CD40L</i> or <i>ICOS</i> result in impaired Ab production, memory B cell formation and reduced circulating CD4<sup>+</sup>CXCR5<sup>+</sup> T cells in humans</li> <li>• due to a CD4<sup>+</sup> T cell intrinsic defect in the ability to form GC</li> <li>• ICOS signalling can also induce c-Maf and IL-21 (although it is unknown whether ICOS directly induces IL-21 or indirectly by inducing c-Maf)</li> </ul>	40-42
	ICOS/ICOS-L	• decreased/absent		
	CD28	• decreased/absent	<ul style="list-style-type: none"> <li>• Tfh deficiency in CD28 ko phenocopied by CD86, but not CD80, deficiency; CD86 dominant over CD80 for Tfh formation. CD80 may have role in Tfh formation, but it is modest</li> </ul>	41,43,44
	CD80	• minimal/no effect		
	CD86	• decreased/absent		
	CD84	• decreased/no effect	<ul style="list-style-type: none"> <li>• Tfh cells were reduced in <i>Cd84</i><sup>-/-</sup> mice in response to immunization with protein Ag, but was less severe than SAP deficiency; however there was no defect in Tfh formation following viral infection. Thus additional SLAM receptors are also involved in SAP-dependent development of Tfh cells</li> </ul>	21,45
	Ly108	• no effect of single gene knockout	<ul style="list-style-type: none"> <li>• although Ly108 deficiency had no effect on Tfh formation, the absence of both Ly108 and SAP restored the Tfh defect observed in SAP-deficient mice. This revealed the ability of Ly108 to deliver inhibitory signals via SHP-1 which countered the positive signals delivered through other receptors (including SLAM family/SAP-associating receptors) on Tfh formation</li> </ul>	45
	OX40/OX40L	• decreased/ no effect	<ul style="list-style-type: none"> <li>• OX40 signalling is required for induction of CXCR5 expression on activated CD4<sup>+</sup> T cells and their subsequent relocation from the T zone to the B-cell follicle</li> <li>• OX40 expression on CD4<sup>+</sup> T cells depends on CD28 signalling, while OX40L expression on DCs requires engagement of CD40; this places OX40/OX40L interactions downstream of CD28/B7 and CD40L/CD40 interactions between CD4<sup>+</sup> T cells and APCs</li> </ul>	41,46-49
	TACI	• increased	<ul style="list-style-type: none"> <li>• BAFF and APRIL-mediated signals through TACI function to intrinsically suppress ICOS-L expression on B cells</li> </ul>	50
	PD-1	• increased	<ul style="list-style-type: none"> <li>• PD-1 negatively regulates Tfh formation; PD-L1 on B cells is the predominant ligand providing the inhibitory signal to constrain Tfh development via PD-1</li> <li>• depending on the experimental model, the increase in Tfh cells in the absence of PD-1 signalling can result in either enhanced or diminished GC and Ab responses</li> <li>• PD-1 may preferentially suppress formation of follicular Treg, rather than Tfh, cells</li> </ul>	51-55

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